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SOHO State of the Art Updates and Next Questions | Challenging Cases in Rare T-Cell Lymphomas

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Author manuscript

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Abstract

Mature T- and NK-cell neoplasms (MTNKN) collectively represent a rare disorder, representing less than 15% of all non-Hodgkin lymphoma (NHL) cases and qualifying for orphan disease designation by the U.S. Food and Drug Administration (FDA). These consist of 9 families in the 5th revised World Health Organization (WHO) classification of lymphoid neoplasms, which are made up of over 30 disease subtypes, underscoring the heterogeneity of clinical features, molecular biology, and genetics across this disease group. Moreover, the 5 most common subtypes (peripheral T-cell lymphoma, not otherwise specified; nodal TFH cell lymphoma, angioimmunoblastic type; extranodal NK-cell/T-cell lymphoma; adult T-cell leukemia/ lymphoma; and ALK-positive or -negative anaplastic large cell lymphoma) comprise over 75% of MTNKN cases, so other subtypes are exceedingly rare in the context of all NHL diagnoses and consequently often lack consensus on best practices in diagnosis and management. In this review, we discuss the following entities – enteropathy-associated T-cell lymphoma (EATL), monomorphic epitheliotropic intestinal T-cell lymphoma (MEITL), hepatosplenic T-cell lymphoma (HSTCL), subcutaneous panniculitis-like T-cell lymphoma (SPTCL), and primary cutaneous yδ T-cell lymphoma (PCGD-TCL) – with an emphasis on clinical and diagnostic features and options for management.

Keywords

EATL; MEITL; HSTCL; SPTCL; PCG-DTCL

Introduction

Although non-Hodgkin lymphoma (NHL) is considered relatively common, representing approximately 4% of all new cancer diagnoses in the United States(1), the majority

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RSB and SKB conceived the organization and scope of the review, co-wrote the text, and made the figures. All authors have read and agreed to the published version of the manuscript.

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of these arise from malignant B-cells. Conversely, mature T- and NK-cell neoplasms (MTNKN) comprise less than 15% of all NHL in Western countries and generally have a worse prognosis than B-cell NHL with 5-year overall survival (OS) under 50% for most subtypes(2). Unfortunately, most patients will experience relapsed or refractory disease, after which the median OS is approximately 6 months(3). One of the major challenges in addressing these suboptimal outcomes is the substantial disease heterogeneity across subtypes of MTNKN. The list of MTNKN has been consistently expanding for several decades, informed by an evolving understanding of cells of origin, clinical features of disease, and genomics, and in 2022, over 30 distinct disease entities have been identified in the MTNKN family based on updated classifications of lymphoid neoplasms developed by the World Health Organization (WHO)(4) and the International Consensus Classification (ICC) Clinical Advisory Committee(5).

This raises a unique dilemma in the clinical management of patients with MTNKN because five subtypes – peripheral T-cell lymphoma, not otherwise specified (PTCL, NOS); nodal TFH cell lymphoma, angioimmunoblastic type (NTFH-AITL); extranodal NK-cell/ T-cell lymphoma (ENKTL); adult T-cell leukemia/lymphoma (ATLL); and ALK-positive or -negative anaplastic large cell lymphoma (ALCL) – comprise over 75% of all cases (Fig. 1)(2). As such, the vast majority of MTNKN subtypes are underrepresented or excluded in large clinical trials and registry studies impacting changes in standard of care(6, 7, 8).

In this review, we will discuss clinical and diagnostic features, outcomes, and treatment considerations of several of the more common "rare" MTNKN, accounting for 6–7% of all diagnoses(2). These include enteropathy-associated T-cell lymphoma (EATL), monomorphic epitheliotropic intestinal T-cell lymphoma (MEITL), hepatosplenic T-cell lymphoma (HSTCL), subcutaneous panniculitis-like T-cell lymphoma (SPTCL), and primary cutaneous $\gamma\delta$ T-cell lymphoma (PCGD-TCL).

Enteropathy-associated T-cell lymphoma and monomorphic epitheliotropic intestinal T-cell lymphoma

The association of celiac disease with intestinal lymphoma was first described in 1962(9), though subsequent studies described a wide range of histologies and clinical features of what were termed enteropathy-type T-cell lymphomas (ETL)(10, 11, 12, 13). The pathophysiology of this connection is rooted in the progressive accumulation of intraepithelial lymphocytes (IELs) in the setting of chronic intestinal inflammation(14). Secretion of interleukin 15 (IL-15) promotes an anti-apoptotic effect, ultimately selecting for malignant transformation of clonally expanded IELs(15, 16, 17). More recently, DeLeeuw and colleagues performed whole-genome analysis and HLA-genotyping on 30 patients with ETL and observed that there are two distinct morphologic and genetic subtypes, which they termed EATL type I or type II(18). We will henceforth refer to these as EATL or MEITL, respectively, based on updated classification of nomenclature(4, 5). EATL is the most common type of intestinal T-cell lymphoma, accounting for 60–80% of cases, while MEITL represents about 30%(13, 19). EATL classically arises in the setting of refractory celiac disease (RCD) II, though de novo transformation in patients with newly diagnosed or

uncomplicated celiac disease has also been described, albeit much less frequently(20, 21). In contrast, MEITL is not associated with celiac disease and sporadically arises from aberrant IELs.

Given this association with celiac disease, it is unsurprising that EATL is largely found in patients of Northern European descent in whom celiac disease is more prevalent(11, 19, 21, 22). MEITL, however, is the most common intestinal T-cell lymphoma in Asian populations, in whom EATL is exceedingly rare(19). These studies have generally observed similar frequency across sexes, though a slight male predominance is seen in MEITL. The incidence of both EATL and MEITL increases with age with a median age of diagnosis of 60-65 years(18, 19, 21, 22, 23). Clinically, patients often may present similarly to an exacerbation of celiac disease with symptoms of abdominal pain/discomfort (80–100%), diarrhea (40– 70%), and weight loss (50-80%)(11, 24, 25). Unlike other types of aggressive lymphomas, B symptoms are relatively infrequent, reported in approximately 30% of patients(11, 19, 24, 25, 26), though this should be considered suspicious for clinical progression or higher-risk disease(27). Moreover, signs of bowel perforation or obstruction are frequently seen and are thought, in part, to contribute to the historically poor prognosis for these patients. Laboratory workup for EATL and MEITL is often non-specific and more so may reflect RCD (in the case of EATL), such as hypoalbuminemia or anemia related to nutritional deficiencies; lactate dehydrogenase (LDH) is normal or only mildly elevated in most cases(11, 19, 26). As such, the diagnosis of EATL or MEITL is often found incidentally during endoscopy for gastrointestinal symptoms or during laparotomy for intestinal perforation or obstruction. Tumor involvement presents multifocally in nearly 25% of cases; EATL and MEITL most commonly involve the proximal small bowel, especially the jejunum(28). Although positron emission tomography/computed tomography (PET/CT) or CT imaging is recommended in staging, extra-intestinal involvement at presentation is fairly uncommon(19); furthermore, EATL metabolic activity is usually restricted to sites of active celiac disease, whereas MEITL is more likely to be eumetabolic(29).

Histologically, EATL is characterized by pleomorphic medium to large lymphocytes with increased mitotic index (Ki67 >50%) and an inflammatory milieu of eosinophils, histiocytes, and small lymphocytes. Given its association with celiac disease, surrounding bowel tends to demonstrate crypt hyperplasia and villous atrophy. Angiocentricity and angioinvasion with necrotic tissue is also frequently seen(30). MEITL conversely exhibits transmural infiltration of monomorphic small to medium sized lymphocytes; inflammatory background, as in EATL, is uncommon(4). Immunohistochemistry (IHC), cytogenetics, and genetics furthermore may be helpful in distinguishing these two entities (Table 1). IHC in EATL is typically positive for TIA-1, CD2, and CD3; variably positive for CD8 and CD30; and negative for CD4, CD5, CD7 and CD56, while IHC in MEITL is typically positive for TIA-1, CD2, CD3, CD7, CD8, and CD56, and negative for CD4, CD5, and CD30(13, 18, 19, 30). Although IELs can normally express TCRa β or TCRy δ , aberrantly proliferating cytotoxic IELs are often TCR $\alpha\beta$ negative though harbor clonal TCR γ or TCR β rearrangements (15, 16, 17, 24); accordingly, most cases of EATL lack TCR $\alpha\beta$ or TCR $\gamma\delta$ expression, whereas MEITL typically expresses TCR $y\delta$, or less commonly TCR $\alpha\beta(30)$. Of note, a subset of patients with EATL can also have an anaplastic large cell phenotype(18), which is more frequently associated with CD8 negativity and CD30 positivity.

The most frequent chromosomal abnormalities identified in all types of ETL are 9q gains and 16q losses, which are typically mutually exclusive. EATL specifically is characterized by frequent gains of 1q and 5q while MEITL often harbors 8q24 gains at the MYC locus(18). Additionally, nearly all patients with celiac disease have HLA-DQ2 or HLA-DQ8 positivity(31), which are both also associated with increased risk of EATL(18, 32, 33). While HLA-DQ2/8 positivity should not be considered diagnostic of EATL, negativity of both should bring into question the diagnosis and may favor MEITL. Few studies have specifically compared the somatic mutational landscape between these disease entities (Table 1), though a report by Nicolae et al. found that JAK/STAT pathway and RAS pathway alterations are common in both diseases(23); however, EATL more commonly has STAT3/JAK1 mutations while MEITL more frequently harbors STAT5B/JAK3 mutations. Other groups have also noted that epigenetic regulators (TET2) and DNA damage/apoptotic mediators (TP53, BCL11B) are mutated in approximately 10-15% of patients with either type of lymphoma, though EATL is enriched for SOCS1 and DAPK3 mutations, while MEITL is enriched for alterations in epigenetic modifiers like CREBBP, EP300, EZH2, and SETD2 and MAPK pathway members (33, 34). Moreover, MEITL tends to overexpress MATK, which may aid in appropriate diagnosis(34).

Historically, outcomes for ETL have been poor, owing at least in large part to the frequent presentation of surgical emergencies like bowel perforation or obstruction(11). Many of these patients have poor performance statuses at diagnosis and nearly one-third die prior to any treatment or shortly after completion of a single cycle of chemotherapy. Historical 5-year OS is below 20% with a median OS of 4–11 months(2, 11, 13, 19, 24, 26). Frontline treatment of EATL usually includes multiagent combination chemotherapy. However, in a retrospective analysis of rare T-cell lymphomas, only 30% of patients with ETL achieved a complete response/unconfirmed complete response (CR/CRu) with frontline therapy, which almost universally was anthracycline-based(26). As such, the question arises of whether there would be benefit to intensification of therapy and consolidative hematopoietic cell transplant (HCT) in first remission. At times, this may require resection of involved bowel prior to chemotherapy for debulking, and treatment can be complicated by a high frequency of malnourishment and a significant risk of bowel perforation.

A study from Scotland and Newcastle Lymphoma Group retrospectively analyzed records of patients with EATL (prior to the distinction of EATL vs MEITL) who were treated with a novel regimen consisting of cyclophosphamide, doxorubicin, vincristine, prednisone (CHOP) followed by ifosfamide, epirubicin, etoposide, intermediate-dose methotrexate (IVE/MTX), ultimately culminating in consolidative HCT(35). Most patients had extensive disease and required prior laparotomy for acute abdominal symptoms. When compared to historical CHOP-like regimens, patients who were able to complete the IVE/MTX protocol had better response rates (93% vs 41%) and notably had a nearly 50% reduction in both mortality and lymphoma-related mortality. However, this regimen was associated with higher rates of febrile neutropenia and sepsis compared to CHOP-like therapy alone. A follow up single-arm phase II trial from the same group in 21 patients found that 86% of patients were able to complete the full CHOP plus IVE/MTX regimen with an overall response rate of 71% though with a 1-year OS of 45%(36). A retrospective study from the EBMT Lymphoma Working Party analyzed records of 44 ETL patients who received

HCT between 2000–2010, noting that 57% had celiac disease(37). CHOP-like frontline therapy was used in 43% of patients, and 70% of patients underwent HCT while in first CR or partial response (PR). Relapse occurred in 39% of patients at up to 4 years after transplant with only one relapse after 18 months. Moreover, progression free survival (PFS) and OS were 54% and 59% at 4 years, respectively, with a trend toward better OS in those who underwent HCT in first CR/PR. Given the improved OS and possibility for durable remission, consolidative HCT in first remission should strongly be considered in eligible patients.

As previously mentioned, EATL tends to express CD30, so CD30-targeted therapy may offer a novel opportunity in the upfront management of this disease. The EATL-001 study is evaluating the efficacy of brentuximab vedotin, a CD30-directed antibody-drug conjugate, plus cyclophosphamide, doxorubicin, prednisone (BV-CHP) in patients with newly diagnosed CD30 positive EATL(38). In this study, responding patients received consolidative etoposide/high-dose MTX followed by HCT. Preliminary data found an overall response rate of 79% with 64% of patients achieving CR. Although two patients died of septic shock during HCT, at 2-years, no patients had yet relapsed, possibly suggesting a new standard of care for this population when compared to historical cohorts(35, 36, 37, 39).

Given that it has only recently been established as a diagnostic entity, there is no current standard of care for MEITL, though it has been included (as Type II EATL) in several of the aforementioned studies(35, 37, 39) and therefore is often treated similarly to EATL. One retrospective study found that although CHOP was the most frequently used frontline regimen for these patients, CR rates were significantly higher for those who received more intensive therapy than CHOP (71% vs 37%)(40); similarly, one report observed that intensive L-asparaginase-containing regimens are associated with a higher response rate than CHOP(41). Moreover, HCT has been associated with significant improvement in survival and, consistent with the literature in EATL, upfront HCT may portend better outcomes than salvage HCT(40, 41). Ultimately, both EATL and MEITL are associated with poor outcomes and significant disease-related morbidity and typically require intensified chemotherapy regimens with consolidative HCT, as CHOP alone is usually not considered sufficient for most patients (Table 2). Our understanding of these distinct entities is continually evolving, and future prospective studies will be critical to optimizing care for these patients.

Hepatosplenic T-cell lymphoma

HSTCL is another rare MTNKN, originally reported to arise from $\gamma\delta$ -T-cells(42, 43), although cases with TCR $\alpha\beta$ expression were later described as well. $\gamma\delta$ -T-cells are the first T cells to develop in the embryonic thymus and are infrequently found in secondary lymphoid tissues. They represent less than 5% of peripheral blood T-cells. After leaving the thymus as CD4/CD8 negative (double negative) T-cells, they circulate and reside in peripheral tissues like skin, adipose tissue, intestine, liver, lungs, and most abundantly in the spleen. They have multiple functions ranging from immune surveillance through excess cytokine production to mucosal barrier maintenance (reviewed in (44)). This immunoregulatory role may play a critical role in both the pathophysiology of HSTCL as well as its clinical presentation, as approximately 20% of cases can arise in patients

on immunosuppressive therapy or with immune-dysregulatory disorders(45, 46, 47, 48, 49). Inflammatory bowel disease, particularly Crohn's disease, has been associated with HSTCL, though this often is thought to occur in the setting of immunosuppressive agents, including anti-tumor necrosis factor (TNF) agents, mercaptopurine, or azathioprine(50, 51, 52). Concomitant use of TNF inhibitors with other immunosuppressive agents is the strongest risk factor for HSTCL(47), suggesting that the pathobiology of disease may be multifactorial stemming from chronic inflammatory antigen exposure with persistence of clonally expanded cytotoxic $\gamma \delta$ -T-cells of the splenic pool in an immunosuppressed environment, ultimately leading to accumulation of driving genetic alterations. However, as previously mentioned, cases of $\alpha\beta$ -T-cell HSTCL have been found, which may suggest a distinct mechanism of disease.

HSTCL occurs in younger patients, with a median age of diagnosis reported between 29-38 years; it is also 2–3-fold more common in males than females(48, 49, 53, 54). B symptoms are seen in 70-80% of patients at presentation. Splenomegaly is almost universal in patients, though hepatomegaly is also seen in 40-88% of patients. Most commonly, this is found in the setting of abdominal discomfort, and signs of hepatic dysfunction, such as jaundice, may occur. Laboratory abnormalities include cytopenias (most commonly thrombocytopenia), elevated LDH, and elevated transaminases. This may be multifactorial due to hypersplenism, bone marrow involvement, or inflammatory myelosuppression. Of note, dysplastic changes in other lineages(55) as well as hemophagocytic syndrome (HPS)(48, 56) have also been reported in patients with HSTCL, though these are unlikely to be common drivers of cytopenias. Given that HSTCL can manifest with cytopenias, elevated LDH, B symptoms, and features of HPS, bone marrow biopsy is an important part of baseline evaluation, as it is involved in 60-70% of patients. While lymphocytosis is uncommon, peripheral blood flow cytometry can detect a neoplastic population of lymphocytes in approximately 50% of patients(46), though overt leukemic phenotypes are seen in less than 2% of patients(48). PET/CT may be non-specific with avidity diffusely in the liver, spleen, and bone marrow, and diagnosis is most commonly made with bone marrow and/or liver biopsy.

Neoplastic cells typically appear as atypical, small to intermediate sized lymphocytes densely infiltrating the sinusoids of the liver and spleen, although blastoid changes have been reported as well(57). However, liver involvement can also manifest as periportal infiltration with spillage into the sinusoids or nodular parenchymal infiltration. Hemophagocytosis can be seen with or without overt HPS. HSTCL cells lack granules, which can help distinguish them from T-cell large granular lymphocytic leukemia. IHC is typically positive for CD2, CD3, CD7, and CD56, while negative for CD4, CD5, CD8, and CD57. Perforin and granzyme B are negative, and TIA-1 and granzyme M are positive, suggestive of a nonactivated cytotoxic phenotype. As previously discussed, approximately 80% of HSTCL are derived from $\gamma \delta$ -T-cells, typically of the V δ 1 subset.

Notably, $\alpha\beta$ variants appear more frequently in females and older patients and are associated with a worse prognosis(48, 58). The most frequent chromosomal abnormalities found in HSTCL are isochromosome 7q and trisomy 8, which are seen in 63% and 50% of cases, respectively, and are not mutually exclusive(2, 54). More recently, next generation sequencing has allowed for the identification of other recurrent genetic abnormalities (Table

1), which may aid in the diagnosis of HSTCL(59, 60). McKinney and colleagues(60) performed whole exome sequencing on 68 HSTCL cases and found recurrent alterations in epigenetic modifiers, JAK/STAT pathway, PI3K pathway, and consensus cancer genes (*TP53, UBR5*, and *IDH2*). Mutations seen in other MTNKN such as *RHOA* and *CCR4* are uncommon in HSTCL, while isochromosome 7q and trisomy 8 are rare in other types of MTNKN.

Outcomes for patients with HSTCL are traditionally very poor, with a median OS of 10–12 months and 5-year OS under 10%(2, 26, 46, 48, 60). Certain prognostic factors, like elevated bilirubin, presence of isochromosome 7q and trisomy 8, or $\alpha\beta$ variants are associated with worse outcomes. Although CHOP-like regimens can achieve CR in patients with HSTCL, median OS remains under 1 year due to frequent relapses(26, 46, 48, 49, 53, 58, 61). Voss and colleagues reported that 7 patients in a 14-patient series were alive at a median follow up of 65.6 months, 6 of whom received ifosfamide-based induction(61). All surviving patients proceeded to undergo HCT as well, so it is unclear whether the benefit in this subset of patients was driven by transplant-related outcomes or alternative induction strategies. Nevertheless, HCT is an important consideration for patients with HSTCL, as accumulating evidence from case series and retrospective analyses demonstrates that patients undergoing HCT have a higher chance of long-term responses with improvements in OS, though this may not always be durable(26, 46, 48, 49, 53, 58, 61). A retrospective study from the EBMT suggested that patients undergoing allogeneic HCT were less likely to experience relapse than those undergoing autologous HCT, though these data are limited by small sample size and relatively short follow up so should be interpreted cautiously(62).

Cytopenias, in particular, present a clinical challenge due to prolonged treatment-related myelosuppression, infectious complications, and difficulty mobilizing stem cells in those who are eligible for HCT, so splenectomy can be considered if alleviation of thrombocytopenia may help the patient receive full doses of chemotherapy. Finally, it has previously been reported that MDR-1 and pgp-1 amplification, which is commonly seen in HSTCL(63), can drive chemoresistance, underpinning rationale for the assessment of non-traditional chemotherapeutics or targeted therapies in future studies; several case reports have suggested alemtuzumab, a CD52-targeting monoclonal antibody, and newer agents such as pralatrexate or the histone deacetylase inhibitor romidepsin may have efficacy in patients with HSTCL(64, 65). Overall, treatment of patients with HSTCL remains a significant challenge with a paucity of data; with the available data(61), a reasonable approach would be to consider induction with ifosfamide, carboplatin, etoposide (ICE), ifosfamide, etoposide, high-dose cytarabine (IVAC), or other platinum-containing high dose regimens; if patients are unable to tolerate this, then CHOP with etoposide (CHOEP or EPOCH) should be considered (Table 2). Preferably, patients should undergo consolidative HCT in patients upon first CR. While either autologous or allogeneic HCT can be considered, there is low-quality evidence that allogeneic HCT may be preferable in eligible patients.

Subcutaneous panniculitis-like T-cell lymphoma and primary cutaneous $\gamma \delta$ T-cell lymphoma

SPTCL and PCGD-TCL are rare primary cutaneous T-cell lymphomas, which were previously both considered under the category of "subcutaneous panniculitis-like T-cell lymphoma" until the 2008 WHO classification of lymphoid neoplasms(66). SPTCL was initially described in 1991 in a case series of patients with T-cell lymphoma in the subcutaneous adipose tissue, 75% of whom had HPS(67). Later studies identified that these cutaneous T-cell lymphomas arise from distinct cells of origin, which are associated with a more indolent (TCR $\alpha\beta$ phenotype) or aggressive (TCR $\sqrt{\delta}$ phenotype) disease course(68). Approximately 20% of patients with SPTCL have a concurrent autoimmune disease, most commonly systemic lupus erythematosus(69, 70); this can be a challenging since lupus erythematosus panniculitis (LEP) can appear strikingly similar, and biopsies require expert dermatopathology review for distinction(73, 74). It is possible that SPTCL and LEP may exist on a spectrum, but in SPTCL, malignant CD8 cytotoxic T-cells expressing CCR5 migrate to adipose tissue, which expresses CCR5 ligands(71). There, the cytotoxic T-cells disrupt adipose membranes, sparing the dermis and epidermis, leading to panniculitis. PCGD-TCL, conversely, is more heterogeneous and aggressive. Similar to HSTCL, PCDG-TCL arises from $\sqrt{\delta}$ -T-cells; recent studies have demonstrated that these can either be the V δ l or V δ 2 subtype leading to superficial epitheliotropic lymphomas or panniculitic lymphomas, respectively, based on their tissue tropism; furthermore, the V $\delta 2$ subtype expresses a cytokine profile consistent with what is seen in HPS(72).

Despite both involving the skin, SPTCL and PCGD-TCL are clinically very different diseases. SPTCL is seen in younger patients (median age 36–38), though can present across a wide age range, while the median age of diagnosis for PCGD-TCL is 59–61 years(70, 73, 74). Both have a female predominance and are characterized by multifocal skin nodules; however, nodules in SPTCL are adipotropic, erythematous, and often painless with a waxing and waning course, whereas nearly half of patients with PCGD-TCL have ulcerative lesions, which can be painful and rapidly progressive. Although PCGD-TCL classically has dermal and epidermal tropism, underpinning the ulcerative phenotype, panniculitic subtypes can be seen(72, 75). Moreover, PCGD-TCL has a lower limb predominance, whereas SPTCL can be distributed along upper or lower extremities or truncally. Diagnosis of either entity is usually made with a skin punch biopsy, though it should be noted that older lesions in SPTCL may exhibit granulomatous inflammation, lipomembranous fat necrosis and/or fibrosis rather than lymphoma. B symptoms are reported in 60–65% of patients with either SPTCL or PCGD-TCL, though HPS is significantly more frequently seen in patients with PCGD-TCL (approximately 50% vs less than 20% in SPTCL)(68, 70). In patients with HPS, typical markers of inflammation, such as hypofibrinogenemia, transaminase elevation, elevated LDH, cytopenias, hyperferritinemia, and hypertriglyceridemia can be seen. For diagnosis of hemophagocytic lymphohistiocytosis (HLH), biopsy specimen should ideally be obtained from the bone marrow, spleen, lymph node, or liver rather than the skin(76, 77). While extracutaneous involvement is uncommon in SPTCL, it can be seen with progressive PCGD-TCL and can involve sites including the lung, thyroid, breast, testis, and mucosa(75).

There are histological differences that can help distinguish SPTCL and PCGD-TCL (Table 1). SPTCL is characterized by a lobular panniculitis-like infiltrate on a background of fat necrosis and histiocytes, occasionally exhibiting hemophagocytosis. Neoplastic cytotoxic T-cells rim adipocytes, which is characteristic of SPTCL. Moreover, plasma cells, lymphoid follicles, and hyaline lipomembranous fat necrosis are less obvious, which can be useful to help distinguish SPTCL from LEP(78, 79). PCGD-TCL is characterized by medium to large lymphocytes, rarely with blastoid morphology, which are distributed either epidermally, dermally, or subcutaneously(73, 80, 81). Neoplastic cells in SPTCL typically stain positive for CD3, CD8, granzyme B, TIA-1, and perforin and negative for CD56 and CD30, whereas PCGD-TCL is characterized by cells that are positive for CD3, CD56, granzyme B, TIA-1, and perforin and negative for CD3, CD56, granzyme B, TIA-1, and perforin and negative for CD3, CD56, granzyme B, TIA-1, and perforin and negative for CD3, CD56, granzyme B, TIA-1, and perforin and negative for CD3, CD56, granzyme B, TIA-1, and perforin and negative for CD3, CD56, granzyme B, TIA-1, and perforin and negative for CD3, CD56, granzyme B, TIA-1, and perforin and negative for CD3, CD56, granzyme B, TIA-1, and perforin and negative for CD3, CD56, granzyme B, TIA-1, and perforin and negative for CD3, CD56, granzyme B, TIA-1, and perforin and negative for CD3, CD56, granzyme B, TIA-1, and perforin and negative for CD3, CD56, granzyme B, TIA-1, and perforin and negative for CD3, CD56, granzyme B, TIA-1, and perforin and negative for CD3, CD56, granzyme B, TIA-1, and perforin and negative for CD3, CD56, granzyme B, TIA-1, and perforin and negative for CD3, CD56, granzyme B, TIA-1, and perforin and negative for CD3, CD56, granzyme B, TIA-1, and perforin and negative for CD3, CD56, granzyme B, TIA-1, and perforin and negative for CD3, CD56, granzyme B, TIA-1, and perforin and negative for CD3, CD56, granzyme B, TIA-1, and perforin and

Biallelic *HAVCR2* (TIM-3) germline mutations are frequently seen in sporadic SPTCL(82, 83, 84). Aberrant loss of TIM-3 expression can result in immune activation and excessive cytokine release, possibly driving the HPS phenotype in a subset of patients(83). Other recurrent somatic mutations in immune response genes (*ASXL1, JAK2, PIAS3, PLCG2*) and epigenetic regulators (*KMT2D, KMT2C, NUP98*) have also been identified in SPTCL(84). Notably, SPTCL without *HAVCR2* mutations has an increase in expression of genes associated with lymphocyte homing and immune regulation. This may have prognostic relevance as well, as younger patients and mutated *HAVCR2* are more likely to have HPS and worse outcomes(84). PCGD-TCL often harbors recurrent translocations at breakpoints involving 9p21, 14q11.2, 14q32.1, or 16q23.1, which involve critical T-cell regulatory genes including *BCL11B* and *TCL*(85). Interestingly, although the V δ I or V δ 2 subtypes of PCGD-TCL have clinically distinct phenotypes, their mutational landscape is similar, involving MAPK signaling, JAK/STAT signaling, chromatin modification, and consensus cancer genes (*CDKN2A, IDH2, TP53*)(72).

Generally, SPTCL is considered to have an excellent prognosis with a 5-year OS over 80%, though patients who experience HPS have worse outcomes with a 5-year OS of 46%(70). Conversely, patients with PCGD-TCL have a 5-year OS of only 11%(70). Of note, $\gamma \delta$ mycosis fungoides (MF) may exist on a spectrum with PCGD-TCL. $\gamma \delta$ MF usually behaves indolently, but a subset of patients may experience PCGD-TCL-like progression, which has a clinical, mutational and gene expression profile identical to de novo PCGD-TCL and has equally dismal outcomes after the phenotypic switch(72).

Willemze et al. found that CR/PR could be achieved in 71% and 88% of patients with SPTCL treated with CHOP-like therapy or immunomodulatory therapy (i.e., cyclosporine), respectively(70). This has been more recently corroborated in a retrospective study showing that either chemotherapy or immunomodulatory therapy can achieve high response rates, and although relapse is common, it did not significantly affect prognosis(74). Notably, cyclosporine achieved responses in 94% of patients, and methotrexate showed responses in 100% of patients when used in the first-line, suggesting that these lower-intensity therapies should be considered frontline for patients without severe disease or HLH (Table 2). Nevertheless, patients with more severe disease and/or HLH often still required chemotherapy with consolidative HCT.

Data on appropriate treatment for PCGD-TCL are sparser. In the study by Willemze and colleagues, of 14 patients with PCGD-TCL treated with CHOP-like chemotherapy, CR was only observed in 3 patients, with frequent progression to visceral involvement and HPS, though outcomes were similar whether or not patients had HPS(70). The use of brentuximab vedotin for CD30-expressing PCGD-TCL has only been reported in case series, and though this may have some efficacy, it warrants further investigation (86, 87). Ultimately, in patients with PCGD-TCL, consolidation with allogeneic stem cell transplantation should be considered in eligible patients. In a small retrospective series of 10 patients with PCGD-TCL and 4 with refractory SPTCL, 7 underwent allogeneic HCT after intensive chemotherapy with CHOP with or without etoposide. Of these patients, 4 were alive at up to 5.9 years after transplant, 3 of whom had PCGD-TCL(88). More recently, real-world data was presented on 48 patients with PCGD-TCL(89); although front-line treatment was very heterogeneous, consolidative HCT was performed in 7 patients, of which 6 were allogeneic. Patients who underwent consolidative HCT in first CR had significantly improved OS. There is no consensus or guidance on initial therapy for this rare disease, but most experts would agree that treatment should be commensurate to the acuity and aggressiveness of the initial presentation. Nevertheless, at present, the limited data suggest that regardless of induction chemotherapy, consolidative HCT is critical for improvement in long-term outcomes (Table 2).

Conclusion

Over the last several decades, there have been significant advances in the field of MTNKN, including expanding therapeutic options and updates in disease classifications(4, 5). Unfortunately, data on the utility of many novel agents are limited to case reports in the majority of MTNKN due to their rarity, which often precludes them from being heavily represented in prospective clinical trials. This has posed a significant challenge to clinicians and patients because many of these MTNKN are treated based off anecdotal experience or by extrapolating data from other biologic entities. As such, outcomes for many rare T-cell lymphomas, especially EATL, MEITL, HSPTCL, and PCGD-TCL, are still dismal with 3-year OS under 50%(26). Moreover, relative inexperience in the diagnostic evaluation of different MTNKN can lead to delays in care or sub-optimal treatment strategies; two groups have independently reported that referrals to tertiary or "expert" centers for MTNKN result in diagnostic reclassification in up to approximately one-third of cases(90, 91), often with implications in therapeutic decision-making. Thus, it is critical that patients with MTNKN have their pathology reviewed at a center with expertise in these diseases.

Nevertheless, retrospective studies and small case series have provided some evidence base for current clinical practices, such as the use of brentuximab vedotin in EATL(38) or consolidative allogeneic HCT in patients with PCGD-TCL(89). Moreover, our understanding of disease biology continues to improve through the use of genomic profiling(92). These technologies have allowed for the identification of multiple potentially targetable pathways across these rare lymphomas, including JAK/STAT signaling, PI3K signaling, and epigenetic pathways, all of which are areas under active investigation. Thus, although MTNKN display extensive disease heterogeneity, accumulating molecular data will hopefully refine these entities and help identify more personalized approaches to treatment.

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Conflicts of Interest

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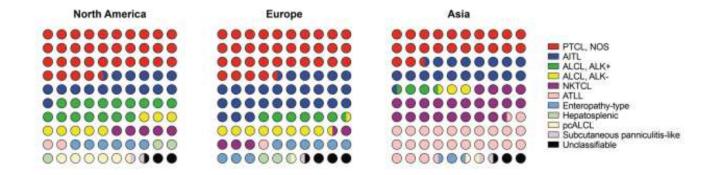


Figure 1. MTNKN display geographic and phenotypic heterogeneity.

Dot plots (10×10) show relative proportions of MTNKN stratified by geographic population; data adapted from the International T-Cell Lymphoma Project(2). Note that nomenclature/abbreviations are based on the previous WHO classification of lymphoid tumors(93). Abbreviations: Peripheral T-cell lymphoma, not otherwise specified, PTCL, NOS; Angioimmunoblastic T-cell lymphoma, AITL, Anaplastic large cell lymphoma, ALCL; NK/T-cell lymphoma, NKTCL; ATLL, adult T-cell leukemia/lymphoma; pcALCL, primary cutaneous ALCL.

Table 1.

Histopathologic and genetic features of rare MTNKN.

MTNKN	Immunophenotype	TCR	Cytogenetics	Recurrent genetic mutations	
EATL	TIA-1+, CD2+, CD3+, CD8+/-, CD30+/-, granzyme B+, perforin+	$\begin{array}{l} \text{TCR-silent} > \\ \text{TCRa\beta} > \\ \text{TCR} \gamma \delta \end{array}$	+1q, +5q, +9q, -16q	SET2D, TET2, STAT3, STAT5B, JAK1, JAK3, NRAS, BRAF, BCL11B, TP53, USP10, TERT, DAPK3, SOCS1	
MEITL	TIA-1+, CD2+, CD3+, CD8+, CD56+, granzyme B+, perforin+	TCRγδ> TCRαβ> TCR-silent	+8q24, +9q, -16q	CREBBP, STAT3, STAT5B, SET2D, GNAI2, JAK1, JAK3, DNAH9, PRR16, ASXL3, MEGF6, EZH2, EP300, TET2, USP10, TERT, KRAS, BRAF, NRAS, BCL11B, TP53	
HSTCL	TIA-1+, CD2+, CD3+, CD4–, CD8+/ –, FasL+, CD56+, granzyme M+	TCRγδ> TCRαβ	i(7q), trisomy 8	SETD2, INO80, TET3, SMARCA2, PIK3CD, STAT5B, STAT3, IDH2, EZH2, ARID1, DNMT3A	
SPLTCL	TIA-1+, CD2+, CD3+, CD4–, CD8+, CD56–, granzyme B+, perforin+	ΤCRαβ	Nonspecific	HAVCR2, ASXL1, JAK3, PIAS3, PLCG2, KMT2D, KMT2C, BAZ2A, NUP98, DDX11, IDH1, BRD2	
PCGD- TCL	TIA-1+, CD2+, CD3+, CD4–, CD8+/ –, CD30+/–, CD56+/–, granzyme B+, perforin+	TCRγδ	Translocations involving breakpoints at 9p21, 14q11, 14q32, or 16q23	KRAS, NRAS, MAPKI, MYC, MYCN, FBXW7, STAT3, STAT5B, JAK3, SOCSI, ARIDIA, TRRAP, TET2, KMT2D, CDKN2A, IDH2, TP53	

Table 2.

Preferred frontline treatment for rare MTNKN.

MTNKN	Preferred treatment modality for fit patients	Role of transplant	Other considerations	Evidence
EATL	Intensified chemotherapy (i.e., CHOP-IVE/ MTX, Hyper-CVAD) preferred. Consider BV-CHP if CD30+.	Consolidative HCT preferred in first remission.	Consider prophylactic bowel resection in select patients at high risk for perforation.	(35, 36, 37, 38)
MEITL	CHOEP/EPOCH preferred. Consider L- asparaginase-based therapy (i.e., SMILE) or intensified chemotherapy (i.e., CHOP- IVE/MTX, Hyper-CVAD), though this is extrapolated from EATL population.	Consolidative HCT preferred in first remission.	Consider prophylactic bowel resection in select patients at high risk for perforation.	(40, 41)
HSTCL	ICE or IVAC preferred. Other platinum- containing high dose regimens, such as GDP may be appropriate. Consider CHOEP/EPOCH if unable to tolerate ifosfamide- or platinum-based therapy.	Consolidative HCT preferred in first remission. Consider allogeneic HCT in eligible patients.	Consider splenectomy if severe thrombocytopenia precludes optimal treatment.	(26, 46, 48, 49, 53, 58, 61, 62)
SPTCL	Immunomodulatory therapy (i.e., CsA, MTX/Prednisone) is preferred unless severe symptoms and/or HLH. In those patients, consider multiagent chemotherapy, such as CHO(E)P or ICE.	Consolidative HCT to be considered for patients with severe symptoms and/or HLH.	Relapse after initial immunomodulatory therapy has not been consistently shown to worsen outcomes, so conservative therapy in mildly symptomatic patients is preferred.	(70, 74)
PCGD- TCL	CHO(E)P preferred.	Consolidative HCT recommended with preference for allogeneic HCT in eligible patients.		(70, 88, 89)

Chemotherapy regimen abbreviations: CHOP (cyclophosphamide, doxorubicin, vincristine, prednisone), IVE/MTX (ifosfamide, epirubicin, etoposide, intermediate-dose methotrexate), BV-CHP (brentuximab vedotin, cyclophosphamide, doxorubicin, prednisone), Hyper-CVAD (hyperfractionated cyclophosphamide, vincristine, doxorubicin, dexamethasone alternating with methotrexate and cytarabine, CHOEP/EPOCH (cyclophosphamide, doxorubicin, vincristine, prednisone, etoposide), SMILE (dexamethasone, methotrexate, ifosfamide, L-asparaginase, etoposide), ICE (ifosfamide, cisplatin, etoposide), IVAC (ifosfamide, etoposide, high-dose cytarabine), GDP (gemcitabine, dexamethasone, cisplatin), CsA (cyclosporine).