

# **HHS Public Access**

Neurobiol Learn Mem. Author manuscript; available in PMC 2024 September 01.

Published in final edited form as:

Author manuscript

Neurobiol Learn Mem. 2023 September ; 203: 107792. doi:10.1016/j.nlm.2023.107792.

# **The functional heterogeneity of PACAP: Stress, learning, and pathology**

**Abha K. Rajbhandari**1, **Jessica R. Barson**2, **Marieke R. Gilmartin**3, **Sayamwong E. Hammack**4, **Briana K. Chen, Ph.D.**5,6

1 Icahn School of Medicine at Mount Sinai, New York, NY, USA

<sup>2</sup>Department of Neurobiology and Anatomy, Drexel University College of Medicine, Philadelphia, PA, United States

<sup>3</sup>Department of Biomedical Sciences, Marquette University, Milwaukee, WI, United States

<sup>4</sup>Department of Psychological Science, University of Vermont, 2 Colchester Avenue, Burlington, VT, United States

<sup>5</sup>Division of Systems Neuroscience, Research Foundation for Mental Hygiene, Inc. (RFMH) / New York State Psychiatric Institute (NYSPI), New York, NY, United States

<sup>6</sup>Department of Psychiatry, Columbia University Irving Medical Center (CUIMC), New York, NY, United States

# **Abstract**

Pituitary adenylate cyclase-activating peptide (PACAP) is a highly conserved and widely expressed neuropeptide that has emerged as a key regulator of multiple neural and behavioral processes. PACAP systems, including the various PACAP receptor subtypes, have been implicated in neural circuits of learning and memory, stress, emotion, feeding, and pain. Dysregulation within these PACAP systems may play key roles in the etiology of pathological states associated with these circuits, and PACAP function has been implicated in stress-related psychopathology, feeding and metabolic disorders, and migraine. Accordingly, central PACAP systems may represent important therapeutic targets; however, substantial heterogeneity in PACAP systems related to the distribution of multiple PACAP isoforms across multiple brain regions, as well as multiple receptor subtypes with several isoforms, signaling pathways, and brain distributions, provides both challenges and opportunities for the development of new clinically-relevant strategies to target the PACAP system in health and disease. Here we review the heterogeneity of central

**Corresponding Author:** Briana K. Chen, Ph.D., Address: Columbia University Irving Medical Center, NYSPI Kolb Research Annex, Room 736, 1051 Riverside Drive, Unit 87, New York, NY 100032, Phone: (646) 774-7318, Fax: (646) 774-7102, bkc2120@cumc.columbia.edu.

**Publisher's Disclaimer:** This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

CRediT Author Statement Abha K. Rajbhandari: Conceptualization, Writing – Original draft preparation, Writing – Review & Editing. Jessica R. Barson: Conceptualization, Writing – Original draft preparation, Writing – Review & Editing. Marieke R. Gilmartin: Conceptualization, Writing – Original draft preparation, Writing – Review & Editing. Sayamwong E. Hammack: Conceptualization, Writing – Original draft preparation, Writing – Review & Editing. Briana K. Chen: Conceptualization, Writing – Original draft preparation, Writing – Review & Editing

PACAP systems, as well as the data implicating PACAP systems in clinically-relevant behavioral processes, with a particular focus on the considerable evidence implicating a role of PACAP in stress responding and learning and memory. We also review data suggesting that there are sex differences in PACAP function and its interactions with sex hormones. Finally, we discuss both the challenges and promise of harnessing the PACAP system in the development of new therapeutic avenues and highlight PACAP systems for their critical role in health and disease.

#### **Keywords**

PACAP; learning; memory; stress; PAC1R

#### **Introduction**

Since its original isolation in 1989 from ovine hypothalamus, based on the ability to stimulate adenylate cyclase in cells of the anterior pituitary, pituitary adenylate cyclaseactivating peptide (PACAP) has emerged as a critical regulator of neuronal activity in a variety of cell types and tissues. This neuropeptide exists either in a 27 or 38 amino acid form, based on alternate processing of the precursor protein prepro-PACAP (Arimura, 1998; Li et al., 1999). While both PACAP-27 and PACAP-38 are widely distributed in the brain and body, PACAP-38 is the more abundant peptide in the central nervous system (CNS) (Arimura et al., 1991; Hannibal et al., 1995; Miyata et al., 1990) although, as discussed below, both PACAP isoforms may regulate CNS activity, depending on the region (Gargiulo et al., 2021).

A highly conserved neuropeptide, PACAP is recognized as a member of the glucagon/ secretin/vasoactive intestinal peptide (VIP) family of peptides (Sherwood *et al.*, 2000; Vaudry et al., 2009). Both PACAP and VIP bind with similar affinities to two common G-protein coupled receptors, VPAC1 and VPAC2, but PACAP also binds with high affinity to PACAP-specific PAC1 receptors. Multiple PAC1 isoforms may be expressed based on alternate splicing in the N-terminal extracellular region or in third cytoplasmic loop, and differences in these variants can potentially lead to differences both in ligand binding and the consequent signaling pathways activated, as discussed in more detail below. These high levels of heterogeneity in PACAP systems contribute to a diversity of functions and mechanisms by which PACAP may regulate CNS activity and behavior.

Due to the ubiquitous nature of the peptide, PACAP systems have been implicated in numerous physiological and behavioral functions. Hence, the development of multiple PACAP or PAC1 receptor null mice lines has demonstrated that they have hyperactive psychomotor behavior, poor fertility, circadian rhythm irregularities, and abnormalities in glucose/lipid homeostasis (Colwell et al., 2003; Gray et al., 2001; Hashimoto et al., 2001; Jamen et al., 2000a). Furthermore, PACAP null mice appear to have high early postnatal mortality, which may be related to acute thermosensitivity and apnea (Gray et al., 2002). Importantly, PACAP, its related peptides, and its receptors are widely distributed throughout the nervous system, where high expression levels of PACAP and/or PACAPassociated receptors are observed in many brain regions associated with learning and

memory, addiction, stress responding, and emotional processing (Hashimoto *et al.*, 1996; Jaworski and Proctor, 2000; Zhang et al., 2021). Expression levels of PACAP and the PAC1 receptor in the brain also suggest that the neuropeptide plays a role in regulating glutamatergic and GABAergic neurotransmission throughout the brain (Zhang et al., 2021). Accordingly, PACAP null mice also demonstrate deficits in some learning tasks (Otto *et al.*, 2001a), reduced anxiety-like behavior (Ishihama et al., 2010), and blunted stress responses (Lehmann et al., 2013). Conversely, activation of central PACAP systems in mice and rats has been shown to produce the opposite effects on many of these same tasks (Donahue et al., 2016; Hammack *et al.*, 2009; Iemolo *et al.*, 2016). Following these initial observations in PACAP null mice, over the next two decades several groups have clarified discrete central PACAP circuits that regulate these complex behavioral processes.

Convergence between the animal models and animal models and human clinical data has suggested that dysregulation in PACAP systems may play key roles across many behavioral processes that relate to several clinical disorders, suggesting that PACAP may represent an important target in their treatment. Moreover, substantial heterogeneity in PACAP systems may allow for the development of strategies that better target mechanisms of pathology while reducing off-target actions and subsequent side effects. In this review, we discuss the functional heterogeneity in PACAP systems, with a focus on psychopathology, and discuss how PACAP systems may be clinically targeted in the treatment of disorders related to these specific behavioral and physiological functions.

### **Heterogeneity of PACAP**

The PACAP precursor, prepro-PACAP, is well-conserved across species, having been characterized in flies (Feany and Quinn, 1995), teleost fish (Parker et al., 1993), mice (Okazaki *et al.*, 1995), rats (Ogi *et al.*, 1990), sheep (Kimura *et al.*, 1990), and humans (Ohkubo et al., 1992), among others. Soma in the brain that contain PACAP are relatively restricted to select regions of the limbic system and brainstem, with the most prominent clustering of PACAP<sup>+</sup> cells being within the hypothalamus (Das *et al.*, 2007; Kivipelto *et al.*, 1992; Köves *et al.*, 1990; Nagashima *et al.*, 2022; Piggins *et al.*, 1996). Other PACAP<sup>+</sup> cells can be found in the midline thalamus (Gupta et al., 2018; Skoglösa et al., 1999), bed nucleus of the stria terminalis (BNST) (Köves et al., 1991), hippocampal formation (Köves et al., 1991; Murase et al., 1995), prefrontal cortex (Köves et al., 1991), areas of the brainstem (Das et al., 2007; Légrádi et al., 1994), and the cerebellum (Nielsen et al., 1998). In addition to the brain regions that contain PACAP<sup>+</sup> soma (Hannibal, 2002; Köves *et al.*, 1991; Köves et al., 1990; Légrádi et al., 1994), PACAP<sup>+</sup> fibers and peptide levels have also been detected in the amygdala, nucleus accumbens, ventral tegmental area, locus coeruleus, dorsal raphe, lateral and medial septal nucleus, diagonal band, and central gray (Kozicz and Arimura, 2002; Masuo et al., 1993; Piggins et al., 1996). Thus, the PACAP system appears to be similar across species and to be particularly active in the limbic system.

One point of heterogeneity in the PACAP system is the diversity of the PACAP peptides. Through the different prohormone processing enzymes in the brain, SPC3 (also called PC1, PC3, or BDP) or SPC2 (also called PC2 or RPC2), prepro-PACAP can be processed into mature, bioactive PACAP-38 or PACAP-27, respectively (Li et al., 1999). The prohormone

processing enzymes themselves have complementary distributions in the brain. For example, in the rat, SPC3 is found at high levels in the hippocampus, while SPC2 is at high levels in the paraventricular nucleus of the thalamus, and both SPC3 and SPC2 are at high levels in the paraventricular nucleus of the hypothalamus (Birch et al., 1994). Notably, across the brain, the concentration of PACAP-38 is approximately eight times greater than that of PACAP-27 (Arimura et al., 1991; Hannibal et al., 1995; Miyata et al., 1990). The existence of two peptide isoforms becomes meaningful considering their different binding affinities for their various receptors (discussed below), which is likely responsible for their divergent behavioral and physiological effects. For example, although PACAP-38 and PACAP-27 both suppress standard chow intake following systemic injection (Vu et al., 2015). PACAP-38 but not PACAP-27 reduces time in the light chamber of a light-dark box following accumbal injection (Gargiulo *et al.*, 2021). PACAP-27 enhances the preovulatory luteinizing hormone surge while PACAP-38 instead prevents it following intracerebroventricular injection (Kántora et al., 2000). This evidence further indicates the additional need to understand the roles of different subtypes of PACAP in behavioral functions.

While only recently gaining significant attention, sex-related differences in PACAP levels are another point of heterogeneity in the PACAP system (see Sex Differences in PACAP Function). Levels of PACAP in the hypothalamus of the rat have not been found to differ between males and females (Iwasa et al., 2016; Kiss et al., 2007; Mosca et al., 2015); however, female mice and rats have been found to have greater gene expression of PACAP and more PACAP<sup>+</sup> cells than males in the paraventricular nucleus of the thalamus (Curtis et al., 2023). Female rats also have higher mRNA levels of the PACAP type 1 receptor than males in the prefrontal cortex, and this fluctuates across the estrous cycle (Kirry et al., 2018). These findings present a novel avenue for exploration of an already complex peptide system.

Taken together, PACAP has heterogeneity in distribution and function via the CNS. Understanding the precise mechanisms through which this neuropeptide regulates these functions could be key to unlocking its potential as a therapeutic target. Given that PACAP acts through various receptors subtypes, it is first important to understand its functions through these receptors.

# **Heterogeneity of the PACAP Receptors**

Receptors for PACAP are class B1 G protein-coupled receptors (Lu et al., 2022), and include not only the PACAP type 1 receptor (PAC1R), but also the PACAP type II receptors, VIP type 1 (VPAC1R) and VIP type 2 (VPAC2R) (Ishihara et al., 1992; Lutz et al., 1993; Pisegna and Wank, 1993). The binding affinity of PAC1 and VIP receptors for PACAP is roughly comparable ( $K_d \approx 0.5$  nM vs  $K_d \approx 1.0$  nM), as is the binding affinity of the VIP receptors for PACAP and VIP ( $K_d \approx 1.0$  nM); however, the PAC1R binds PACAP with one thousand-fold greater affinity than VIP ( $K_d \approx 0.5$  nM vs  $K_d > 500$  nM) (Harmar *et* al., 2012; Vaudry et al., 2009; Zhou et al., 2000). While PACAP-38 binds to PAC1R and VPAC2R with slightly higher affinity than PACAP-27, PACAP-27 binds to VPAC1R with slightly higher affinity than PACAP-38 (Ramos-Álvarez *et al.*, 2015). For all three receptors, stimulation is known to regulate the production of cyclic adenosine monophosphate (cAMP)

through  $G_s$  proteins, but it has also been shown to activate phospholipase C (PLC) and intracellular calcium mobilization through  $G_q$  and  $G_i$  proteins (Holighaus *et al.*, 2011; Lu et al., 2022; MacKenzie et al., 2001; Pisegna and Wank, 1993; Van Rampelbergh et al., 1997). In addition, PACAP can promote PAC1 internalization independent of G-protein mechanisms (Parsons and May, 2019), and endosomal signaling can mediate some of the physiological (Johnson et al., 2020b; Parsons and May, 2019) and behavioral (Missig et al., 2014) consequences of PAC1 activation, particularly those that may be long-lasting. PAC1 signaling can be associated with the long-lasting activation of extracellular-signal regulated kinase (ERK)1/2 signaling, which is particularly dependent on PAC1 interactions with β-arrestin2, although interactions between multiple β-arrestin isoforms and PAC1 receptors may help to regulate PAC1 signaling (Shintani et al., 2018). Moreover, these effects of PAC1 signaling may also regulate surface expression of other receptors subtypes, such as 5-HT2A receptors (Hayata-Takano et al., 2021). This means that PACAP receptor binding can induce several different types of intracellular signaling, which may be differentiated in a multitude of cellular actions and with different temporal profiles.

Compared to VPAC1R and VPAC2R, PAC1R is more widely and densely expressed in the brain (Basille et al., 2000). Gene expression for the PAC1 receptor can be found in the hypothalamus, midline thalamus, BNST, hippocampus, cingulate cortex, cerebellum, amygdala, nucleus accumbens, locus coeruleus, dorsal raphe, lateral septal nucleus, and central gray (Hashimoto *et al.*, 2006; Shioda *et al.*, 1997), largely the same regions where PACAP fibers have been detected. These PAC1Rs are predominantly, but not exclusively, postsynaptic (Vaudry et al., 2009). The VPAC1Rs and VPAC2Rs overlap in their distribution with the PAC1Rs; gene expression and receptor binding for VPAC1R is found in the hypothalamus, BNST, hippocampus, cortex, cerebellum, and amygdala, and VPAC2R is additionally found in the thalamus, nucleus accumbens, caudate-putamen, and lateral septal nucleus (Joo et al., 2004; Vertongen et al., 1997). PAC1R binding may induce opposing behavioral effects to those of VPAC1R and VPAC2R binding (Rudecki and Gray, 2016). The overlap in expression between the different receptor types may allow for tighter regulation of behavioral effects, based on the timing and level of PACAP release.

In addition to the three receptors for PACAP, another point of heterogeneity is variants of these receptors, which arise from alternative splicing of the genes. In the rat, most of the differences in the PAC1R occur in the third intracellular loop, and are characterized by the absence (null variant) or presence of either one or two different insertions at the C-terminal end of the loop, which are 28 amino acid cassettes (hip or hop1 variant) or a 27 amino acid cassette (hop2 variant), that can be included together to form hiphop (Holighaus et al., 2011; Vaudry et al., 2009). In the mouse, both the null (short) and hop variants of the PAC1R have been identified [\(https://www.ncbi.nlm.nih.gov/gene/11517\)](https://www.ncbi.nlm.nih.gov/gene/11517). While all variants of the PAC1R are  $G_s$ -coupled, the null and hop1 variants are also strongly  $G_q$ coupled (Holighaus *et al.*, 2011), and the presence of the hip cassette impairs stimulation of cAMP and abolishes stimulation of phospholipase C (Zhou et al., 2000). Thus, PACAP can also act at any of several receptor variants, contributing to the heterogeneity of signaling and possible functional consequences of PACAP receptor binding. Next, we explore the mechanisms through which PACAP regulates functional outcomes associated with stress and aversive learning.

# **PACAP Regulates Stress Signaling Pathways**

Circulating throughout the brain and periphery, PACAP is well positioned to regulate both central and peripheral responses to stress. Following exposure to a stressor, the hypothalamic-pituitary-adrenal (HPA) axis initiates a cascade of neuroendocrine pathways that alter several physiological processes, including autonomic function and immune processes, to maintain homeostasis. Critically, PACAP exerts a major regulatory role in coordinating the acute and sustained activity of this HPA circuit; these functions have been thoroughly explored in a number of publications (Boucher et al., 2021; Denes et al., 2019; Hammack and May, 2015; Hashimoto *et al.*, 2011; King *et al.*, 2017; Lutfy and Shankar, 2019; Mustafa, 2013; Nussdorfer and Malendowicz, 1998; Stroth et al., 2011a). Briefly, PACAP is necessary for the release of adrenocorticotropic hormone (ACTH), corticosterone (CORT), and corticotropic-releasing hormone (CRH) (Agarwal et al., 2005; Lehmann et al., 2013; Stroth and Eiden, 2010; Stroth et al., 2011b; Tsukiyama et al., 2011). Additionally, in PACAP-deficient mice, stress-induced activation of limbic brain regions is impaired, including the amygdala, BNST, hippocampus, paraventricular nucleus of the hypothalamus, dorsal raphe nucleus, and periaqueductal gray (Gaszner *et al.*, 2012; Kormos *et al.*, 2016). Thus, by controlling the release of stress hormones as well as the activation of stress-related brain regions, PACAP plays a major role in stimulating the stress response.

In addition to its role in initiating responses to stress and activating stress-related brain regions, PACAP critically modulates stress-induced behaviors. In the BNST, PACAP is necessary and sufficient for the induction of anxiety-related exploratory behavior and may also facilitate reward seeking behaviors, particularly in response to stress (Ferragud et al., 2021; Hammack et al., 2009; Jovanovic et al., 2013; Maita et al., 2022; Miles et al., 2019; Miles et al., 2018; Roman et al., 2014). PACAP may also alter stress-related reward learning and drug seeking by altering activity of the nucleus accumbens and paraventricular nucleus of the thalamus (Curtis et al., 2020; Gargiulo et al., 2021; Pirino and Barson, 2021). In the amygdala, PACAP may have distinct functions when acting in different subnuclei. In the central amygdala, PACAP facilitates the encoding and consolidation of fear memories and enhances anxiety-like behaviors induced by footshock, restraint, pain, and social defeat stress (Iemolo *et al.*, 2016; Legradi *et al.*, 2007; Meloni *et al.*, 2019; Missig et al., 2014; Seiglie et al., 2022; Varodayan et al., 2020). Conversely, while PACAP does not alter neuronal excitability of central amygdala neurons, PACAP potentiates neuronal signaling in the basolateral amygdala, where it also acts to modulate different aspects of fear memories, such as fear learning, fear memory consolidation, and fear extinction, in a PAC1-dependent manner (Andero et al., 2014; Cho et al., 2012; Rajbhandari et al., 2021; Velasco et al., 2022). In the hippocampus, PACAP is also vital for regulating neuronal activity in response to stress. PACAP projections from hilar mossy cells to dentate gyrus granule cells potentiate excitatory activity and initiate secondary messenger signaling, and this modulation of hippocampal dentate gyrus activity may contribute to altered fear extinction (Ciranna and Cavallaro, 2003; Condro et al., 2016; Johnson et al., 2020a; Johnson et al., 2020b; Kondo et al., 1997; Macdonald et al., 2005; Pecoraro et al., 2017; Roberto and Brunelli, 2000). Finally, in the prefrontal cortex, PACAP, which is mainly expressed in glutamatergic pyramidal neurons, is necessary for cued fear learning, facilitates behavioral

despair and anxiety-like behaviors, and contributes to the sustained response to social defeat stress (Ago et al., 2017; Kirry et al., 2018; Lehmann et al., 2013; Martelle et al., 2021). Overall, PACAP is widely expressed throughout the brain and demonstrably regulates numerous behaviors in response to stress. These actions may functionally overlap in the brain, and reciprocal PACAPergic connections between these and other brain regions may strongly regulate stress-induced behaviors.

Finally, PACAP is a vital regulatory neuropeptide for stimulating the sympathetic nervous system. For instance, PACAP is widely expressed throughout the sympathoadrenomedullary system, such as in neurons along the brainstem, spinal cord, and in the adrenal glands (Das et al., 2007; Farnham et al., 2008; Holgert et al., 1996; Kumar et al., 2010; Nogi et al., 1997). Expression of PAC1 is high in the locus coeruleus, a brain region critical for the fight-flight-freeze response and modulation of metabolic functions in the periphery in response to stress (Duesman et al., 2022). Following psychological stress, PACAP plays a key role in the synthesis of adrenaline and the release of catecholamines from the adrenal glands (Stroth and Eiden, 2010; Tönshoff et al., 1997). When PACAP is released into the sympathetic nervous system, it suppresses the parasympathetic nervous system, indicating the importance of the neuropeptide in activating an active stress response (Tanida *et al.*, 2010). PACAP can also stimulate sympathetic nervous system activity following exposure to several other stressors, including hypoglycemic stress and cold stress (Gray et al., 2002; Hamelink et al., 2002); additionally, it plays neuroprotective and anti-inflammatory roles in the brain and body in response to oxidative stress (see Horvath et al., 2019 and Sadanandan *et al.*, 2021 for a more thorough review). Moreover, PACAP expression is high in parasympathetic pathways, including the vagal ganglion (Kupari et al., 2019). To our knowledge, understanding the role of this neuropeptide in modulating stress pathways via parasympathetic modulation has not yet been conducted but warrants further studies. Thus, by coordinating signaling in the HPA axis, activating key brain regions, and stimulating the sympathetic nervous system, PACAP plays a critical role in activating and sustaining central and peripheral responses to stress and therefore may be a key target for clinical treatments of stress-related psychopathology.

## **PACAP Promotes Aversive Learning**

In addition to regulating the behavioral and physiological responses to stressors, PACAP contributes to the long-term encoding of aversive experience in memory. Genetic deletion of PAC1R either globally or in the forebrain produces mild to severe impairments in contextual fear conditioning, an associative paradigm in which subjects learn to associate the spatial configuration of environmental cues with a footshock. In contrast, navigational learning in the Morris water maze and social transmission of food preference is unaffected by loss of PAC1R (Otto *et al.*, 2001a; Otto *et al.*, 2001b; Sauvage *et al.*, 2000). Mice with genetic deletion of the PACAP peptide also showed impaired contextual fear memory as well as deficits in novel object recognition (Takuma et al., 2014). These studies implicate PACAP in aversive learning, and while the memory deficits in genetically modified mice are confounded somewhat by the role of PACAP in neural development and HPA axis regulation (Ishihama et al., 2010; Takuma et al., 2014; Hashimoto, 2006), direct manipulation of PACAP signaling in adult rats also supports a role for PACAP in aversive learning

and memory. Consolidation of contextual fear memory in males is impaired when the PAC1R antagonist, PACAP6–38, is injected i.c.v. or directly into the dorsal hippocampus or amygdala after training (Sacchetti et al., 2001; Schmidt et al., 2015). Conversely, consolidation is enhanced, or at high doses even impaired, when PACAP is injected in these regions (Meloni et al., 2018; Meloni et al., 2016; Schmidt et al., 2015). In the prefrontal cortex, blocking endogenous PACAP signaling impairs the formation of trace fear memory, a form of cued fear learning dependent on episodic memory systems, while sparing working memory in a spatial alternation task (Kirry et al., 2018). These effects are observed in females, but not males. In the amygdala, viral-mediated deletion of PAC1R in the medial intercalated cells, a region involved in the suppression of fear following extinction (Duvarci and Pare, 2014), results in increased fear generalization and impaired extinction in males but not females (Rajbhandari *et al.*, 2021). Females show a reduced asymptotic level of fear during acquisition. Collectively, these studies reveal that PACAP exerts sex- and regionspecific modulation of fear-related memories, and may provide insight into the genetic link between PAC1R and post-traumatic stress disorder (PTSD) in women (Ressler et al., 2011). The risk allele is associated with enhanced startle to threat-related cues in adults (Jovanovic et al., 2013; Ressler et al., 2011) and in children one year after conditioning Jovanovic et al., 2020; Velasco et al., 2022). Impaired fear and safety discrimination and altered hippocampal and amygdala reactivity in fear conditioning are also observed in women with the risk allele (Ressler et al., 2011; Stevens et al., 2014).

The mechanisms by which PACAP promotes synaptic plasticity and long-term memory for aversive events may be region-specific. CREB-mediated gene transcription downstream of adenylate cyclase/cAMP signaling is necessary for the formation of long-term memories (Asok *et al.*, 2019; Kandel *et al.*, 2014), and PACAP is a potent activator of these signaling cascades. In the dentate gyrus, for example, PACAP-PAC1R signaling drives CREB-mediated transcription and promotes excitability of dentate gyrus granule cells (Johnson et al., 2019; Johnson et al., 2020a; Johnson et al., 2020b). Mossy fiber long-term potentiation (LTP), a cellular correlate of learning and memory, is impaired in mice lacking PACAP or PAC1R (Matsuyama *et al.*, 2003; Otto *et al.*, 2001b). In the central nucleus of the amygdala, the expression of the plasticity-related protein Arc following fear conditioning is enhanced by central administration of PACAP (Meloni et al., 2018). Arc is a key regulator of synaptic plasticity and PACAP may modulate its expression via PKA signaling (Nikolaienko et al., 2018; Bloomer et al., 2008; Shepherd and Bear, 2011). Moreover, PACAP-mediated changes in AMPA and NMDA signaling in the hippocampus and amygdala promote longlasting effects onsynaptic efficacy that may support the acquisition and extinction of fear memories (Ciranna and Cavallaro, 2003; Cho et al., 2012; Varodayan et al., 2019; Costa et al., 2009; Condro et al., 2016; Johnson et al., 2020a; Johnson et al., 2020b; Kondo et al., 1997; Macdonald et al., 2005; Pecoraro et al., 2017; Roberto and Brunelli, 2000; Roberto et al., 2001; Yaka et al., 2003). The mechanisms by which PACAP contributes to plasticity and memory in cortex and other systems remains to be determined.

In summary, PACAP not only facilitates the central and peripheral response to environmental stressors, but also influences the long-lasting memory of stressful situations. Vulnerabilities in PACAP signaling may thus contribute to pathological fear in PTSD

# **Sex Differences in PACAP Function**

events.

While many of the initial studies of PACAP expression and function were conducted in male animals, increasing awareness has been paid to the distinct biological activities of PACAP in females, as evidenced by a growing number of studies documenting sex-specific aspects of PACAP signaling and function. Perhaps one of the most well-known sex differences in PACAP function is its role in mediating susceptibility to PTSD in women, but not men. In 2011, Ressler and colleagues found that the PACAP pathway is implicated in human psychological stress responses in a sex-specific manner (Ressler et al., 2011). In female but not male subjects, high PACAP-38 peptide blood levels significantly predicted PTSD symptoms. Moreover, a single nucleotide polymorphism (SNP) (rs2267735), which replaces a G allele with a C allele in a putative estrogen response element within the PAC1R gene, was significantly associated with PTSD symptoms and heightened physiological fear responses in females, but not males. These results have since been replicated in numerous other studies (Almli et al., 2013; Lind et al., 2017; Stevens et al., 2014; Uddin et al., 2013; Wang et al., 2013). Follow-up investigation suggests that this SNP may alter the reactivity of PACAP-PAC1R signaling in response to circulating levels of estrogen, leading to impaired fear and stress responses in female, but not male individuals (Ramikie and Ressler, 2016; Velasco et al., 2022).

The interactions of PACAP with estrogen have also been documented in independent investigations. For instance, in rodent models, estrogen may modulate the pro-anorectic effect of PACAP in the BNST (Kocho-Schellenberg et al., 2014). Interestingly, when PACAP is infused into the BNST, prior exposure to chronic variable stress enhances the startle response in intact female but not male rats, indicating that gonadal hormone levels may play a role in altering PACAP-mediated responses to stress, particularly in the BNST (King et al., 2017). Beyond the BNST, PACAP expression has been shown to differ in a number of stress-related brain and peripheral regions, such as the adrenal glands and superior cervical ganglia, and PACAP expression in the hypothalamus and brainstem is differentially altered by exposure to stress in a sexually dimorphic manner (Kiss et al., 2007; Mosca et al., 2015). Moreover, PACAP expression in the middle and posterior subregions of the PVT are at baseline higher in females than in males; this sexually dimorphic PACAP expression may have further implications for sex differences in addictive and affective behaviors (Curtis et al., 2023). Thus, sex-specific differences in the PACAP system, both at baseline and in response to stress, likely have functional consequences that contribute to sex differences in behavior and disease.

Finally, PACAP has also been implicated in the regulation of the female reproductive system and during pregnancy (Koppan et al., 2022). In the CNS, PACAP is strongly expressed in various hypothalamic nuclei, where it can influence the release of a number of hormones, including oxytocin, vasopressin, luteinizing hormone (LH), and follicle stimulating hormone (FSH) (Dow et al., 1994; Jolivel et al., 2009; Lutz-Bucher et al., 1996; Murase et al., 1995; Murase et al., 1993; Nomura et al., 1999; Palkovits et al., 1995; Petersen et al., 1996; Yon

et al., 2001). Interestingly, the effects of PACAP on gonadotropin release in female mice may vary depending on age, time of day, and phase of the estrous cycle (Heinzlmann *et al.*, 2008; Köves et al., 1998; Moore et al., 2005). In the periphery, PACAP is strongly expressed in the ovaries, uterus, and placenta where it has been suggested to contribute to follicular development, oogenesis, and vasodilation in the uterus and placenta (Barberi *et al.*, 2007; Gräs et al., 1996; Gräs et al., 2005; Lee et al., 1999; Park et al., 2001; Scaldaferri et al., 2000; Steenstrup *et al.*, 1996; Vaccari *et al.*, 2006). When PACAP or PAC1R is genetically deleted, female mice exhibit reduced fertility, disrupted maternal behaviors, and, in some studies, delayed estrous cycles (Hashimoto et al., 2006; Isaac and Sherwood, 2008; Jamen et al., 2000b; Lajko et al., 2018; Shintani et al., 2002). In women, levels of PACAP in human follicular fluid may influence the success of egg retrieval during in vitro fertilization (IVF), further suggesting that PACAP contributes to the female reproductive system (Brubel *et al.*, 2011; Koppan et al., 2012). Further investigation is necessary to elucidate the sex-specific expression, signaling, and functions of PACAP, particularly in clinical populations.

#### **Clinical Relevance of PACAP/PAC1 Systems in Health and Disease**

Due to its neuromodulatory role in regulating the stress response and aversive learning, vulnerabilities in PACAP signaling are implicated in a variety of disorders. The behavioral, autonomic, and immune functions of PACAP and its receptor systems provide numerous opportunities to develop novel therapeutic treatments for such disorders. Consistent with evidence implicating PACAP dysregulation in stress responding and emotional processing, in 2011 Ressler et al. reported that high blood levels of PACAP and a single nucleotide polymorphism (SNP) in the PAC1R gene are correlated with PTSD symptom severity, especially in women with PTSD (Ressler et al., 2011). This is especially relevant, as women with the PTSD PAC1R risk genotype show an impairment in fear extinction (Velasco et al., 2022). Methylation of the PAC1R gene is also associated with symptoms of PTSD in both men and women (Ressler et al., 2011). These associations in the 2011 study were subsequently tested again in several populations with mixed results (Almli *et al.*, 2013; Chang et al., 2012; Jovanovic et al., 2013; Uddin et al., 2013; Wang et al., 2013), and the SNP was located to a predicted estrogen response element (ERE) where it may regulate the binding of estradiol (E2) (Mercer et al., 2016). Other reports have suggested that PACAP contributes to the pathology of schizophrenia, substance abuse disorder, major depressive disorder, and generalized anxiety disorder (Cooper et al., 2013; Hashimoto et al., 2007; Hashimoto et al., 2010; Ishiguro et al., 2001; Marquez et al., 2009; Miles et al., 2018; Ross et al., 2020; Tseng et al., 2019). Thus, the demonstrated role of PACAP in a number of psychiatric disorders suggests that targeting this neuropeptide may provide novel, effective interventions for both the treatment and prevention of such diseases.

As a member of the glucagon family of neuropeptides, PACAP has also been studied for its central control of digestive processes and feeding behaviors. Specifically, the role of PACAP in anorectic functions indicates that this neuropeptide could indirectly modulate processes in the CNS, by influencing the gut, and directly in the brain (Karpiesiuk and Palus, 2021; Reglodi et al., 2018; Sekar et al., 2017). Expression of PACAP and its receptors has previously been identified in the gastrointestinal tract (Sekar *et al.*, 2017), where the neuropeptide has been shown to: 1) regulate gastric secretion and emptying through

interactions with neuroendocrine cells (Håkanson et al., 2001; Sandvik et al., 2001), 2) modulate blood flow via PAC1 receptors (Naruse *et al.*, 1996), 3) stimulate cell proliferation and differentiation (Läuffer *et al.*, 2001; Oh *et al.*, 2005), and 4) regulate intestinal motility (Chang et al., 1996; Chang et al., 1998; Costa et al., 2021; Cox, 1992; Fuchs et al., 1996; Racké et al., 1996). Centrally, PACAP administration has been shown to induce hypophagia and increase production of other hormones such as leptin, suggesting that it has a role in modulating energy homeostasis, appetite, and satiety (Iemolo *et al.*, 2015; Kocho-Schellenberg et al., 2014; Nagashima et al., 2022; Sekar et al., 2017, Tomimoto et al., 2008). Thus, novel therapies targeting PACAP may have a beneficial effect in treating disorders related to feeding or metabolism, such as anorexia, binge eating disorder, and diabetes (Nakata and Yada, 2004; Yada et al., 2000).

In addition to anxiety and eating disorders, PACAP has been implicated in the pathogenesis of migraine and headache (Christensen et al., 2022; Vécsei et al., 2014). Infusion of PACAP-38 in human volunteers has been shown in several studies to induce migraine attacks (Amin et al., 2014a; Ghanizada et al., 2020; Vollesen et al., 2019; Wienholtz et al., 2021). While the mechanism of this effect is not fully determined, it is hypothesized that the role of PACAP in mediating vasodilation contributes to the onset of pain. Indeed, PACAP receptors are expressed throughout meningeal and arteries (Chan *et al.*, 2011; Knutsson and Edvinsson, 2002), and administration of a vasoconstrictor prior to PACAP-38 infusion reduces the probability of migraine onset (Amin et al., 2013). Additionally, magnetic resonance angiography imaging indicates that there is significant vasodilation of the medial meningeal artery shortly after infusion and prior to the onset of migraine attack (Amin et al., 2012; Amin et al., 2014b; Ghanizada et al., 2019). Therefore, because of its proposed role in headache pathophysiology, therapies targeting PACAP or its receptors may have a profoundly therapeutic role for preventing or treating headache and migraine pathologies. Regarding biomarker development targeting the PACAPergic system, a phase 2 randomized controlled trial for migraine using an antibody against the PAC1 R did not yield positive benefits (Ashina et al., 2021). For neuropathic pain, a PAC1R antagonist has also been developed but is still undergoing further testing (Martin *et al.*, 2005; Takasaki *et al.*, 2020; Takasaki et al., 2018).

Despite the strong therapeutic potential for targeting the PACAP system to better treat and prevent a variety of diseases, there are currently no available PACAP-based therapies. Although the parallel behavioral and physiological functions of PACAP make it an ideal candidate for modulating a range of diseases, it can also complicate the development of PACAPergic systemic therapeutics. Thus, developing a more complete and nuanced understanding of the structures and functions of PACAP as well as its receptors is critical for advancing drug development.

### **Challenges of Harnessing the PACAP System for Developing Therapeutics**

Given the heterogeneity of the PACAP system, both in terms of its functional effects and its signaling profile, there are significant challenges for harnessing this system with pharmacotherapeutics. Thanks to recent advances in cryo-electron microscopy, however, there are now models of the PAC1R, VPAC1R, and VPAC2R that are of sufficient quality

to allow for drug discovery (Kobayashi *et al.*, 2020; Langer *et al.*, 2022; Liang *et al.*, 2020; Wang et al., 2020, Xu et al., 2022). Ligands can now be developed that are receptor-selective. Biased ligands can be developed that favor specific confirmations of the receptors, to activate only some of the signaling pathways (Langer et al., 2022; Martin et al., 2005; Takasaki et al., 2020; Takasaki et al., 2018). Ligands can also be developed to favor specific receptor splice variants, which each have their own signaling properties. One challenge will be to target specific receptors or splice variants in specific brain regions or networks, to accomplish the therapeutic goal with minimal off-target effects. This will require comprehensive mapping of the receptors and receptor variants, which can now be accomplished through whole brain tissue mapping. Although significant resources across multiple laboratories are needed to accomplish this, the field now has the capabilities to finely delineate these aspects of the PACAP system, allowing for incredibly precise manipulation of a highly pleiotropic system.

As consistently demonstrated, PACAP is an important regulatory neuropeptide for modulating the brain and body at the molecular, cellular, systems, and behavioral level. However, important questions remain that will need careful study and experimentation. While PACAP is highly conserved, it remains to be seen whether the expression and functions of PACAP are similar or divergent in different species; this consideration will be necessary for developing PACAP-based therapeutics to treat stress-related disorders. Determining the main points of synthesis and release of PACAP in the central and peripheral nervous system will also be important for investigating how PACAP may be targeted for disease intervention in a global or local manner. Further investigation is also needed to examine the temporal resolution of PACAP circulation and degradation. For instance, pharmacological agents either prolonging or accelerating the synthesis and/or degradation of PACAP could be viable lead targets for developing PACAP-related therapeutic agents. Finally, although PACAP is certainly a critical neuropeptide in the brain and body, its interactions with other neuropeptides and neurotransmitters must also be considered. Thus, determining how PACAP interacts with these other neuroactive agents will provide a more complete and nuanced understanding of the neuropeptide.

### **Conclusion**

In sum, PACAP exerts diverse and critical functions in mediating a variety of central and peripheral processes. The versatility of the neuropeptide creates promising avenues for developing novel therapeutics for a number of disease conditions, including affective, fear, eating, energy metabolism, and pain disorders. However, the pleiotropic nature of PACAP signaling also introduces challenges for developing specific and effective therapeutics. Therefore, further study and investment in the basic and translational potential of PACAP and PACAPergic interventions is necessary to enhance our understanding of the neuropeptide and improve ongoing efforts to improve health outcomes.

### **Funding Sources**

This work was supported by the Brain and Behavior Research Foundation (29227 to AJR); the NIDDK (R21DK129908 to AJR); the NIAA (R01AA028218 to JRB); the NIMH (R15MH118601 to MRG, R01MH097988 to SEH, R21MH080935 to SEH); and the Akira Arimura Foundation (to BKC).

# **References**

- Agarwal A, Halvorson LM, & Legradi G (2005). Pituitary adenylate cyclase-activating polypeptide (PACAP) mimics neuroendocrine and behavioral manifestations of stress: Evidence for PKAmediated expression of the corticotropin-releasing hormone (CRH) gene. Brain Res Mol Brain Res, 138, 45–57. [PubMed: 15882914]
- Ago Y, Hayata-Takano A, Kawanai T, Yamauchi R, Takeuchi S, Cushman JD, Rajbhandari AK, Fanselow MS, Hashimoto H, & Waschek JA (2017). Impaired extinction of cued fear memory and abnormal dendritic morphology in the prelimbic and infralimbic cortices in VPAC2 receptor (VIPR2)-deficient mice. Neurobiol Learn Mem, 145, 222–231. [PubMed: 29030297]
- Almli LM, Mercer KB, Kerley K, Feng H, Bradley B, Conneely KN, & Ressler KJ (2013). ADCYAP1R1 genotype associates with post-traumatic stress symptoms in highly traumatized African-American females. Am J Med Genet B Neuropsychiatr Genet, 162b, 262–272. [PubMed: 23505260]
- Amin FM, Asghar MS, Guo S, Hougaard A, Hansen AE, Schytz HW, van der Geest RJ, de Koning PJ, Larsson HB, Olesen J, & Ashina M (2012). Headache and prolonged dilatation of the middle meningeal artery by PACAP38 in healthy volunteers. Cephalalgia, 32, 140–149. [PubMed: 22174350]
- Amin FM, Asghar MS, Ravneberg JW, de Koning PJ, Larsson HB, Olesen J, & Ashina M (2013). The effect of sumatriptan on cephalic arteries: A 3T MR-angiography study in healthy volunteers. Cephalalgia, 33, 1009–1016. [PubMed: 23588794]
- Amin FM, Hougaard A, Schytz HW, Asghar MS, Lundholm E, Parvaiz AI, de Koning PJ, Andersen MR, Larsson HB, Fahrenkrug J, Olesen J, & Ashina M (2014a). Investigation of the pathophysiological mechanisms of migraine attacks induced by pituitary adenylate cyclaseactivating polypeptide-38. Brain, 137, 779–794. [PubMed: 24501094]
- Amin FM, Lundholm E, Hougaard A, Arngrim N, Wiinberg L, de Koning PJ, Larsson HB, & Ashina M (2014b). Measurement precision and biological variation of cranial arteries using automated analysis of 3 T magnetic resonance angiography. J Headache Pain, 15, 25. [PubMed: 24886137]
- Andero R, Dias BG, & Ressler KJ (2014). A role for Tac2, NkB, and Nk3 receptor in normal and dysregulated fear memory consolidation. Neuron, 83, 444–454. [PubMed: 24976214]
- Arimura A (1998). Perspectives on pituitary adenylate cyclase activating polypeptide (PACAP) in the neuroendocrine, endocrine, and nervous systems. Jpn J Physiol, 48, 301–331. [PubMed: 9852340]
- Arimura A, Somogyvári-Vigh A, Miyata A, Mizuno K, Coy DH, & Kitada C (1991). Tissue distribution of PACAP as determined by RIA: highly abundant in the rat brain and testes. Endocrinology, 129, 2787–2789. [PubMed: 1935809]
- Ashina M, Doležil D, Bonner JH, Zhou L, Klatt J, Picard H, & Mikol DD (2021). A phase 2, randomized, double-blind, placebo-controlled trial of AMG 301, a pituitary adenylate cyclaseactivating polypeptide PAC1 receptor monoclonal antibody for migraine prevention. Cephalalgia, 41, 33–44. [PubMed: 33231489]
- Asok A, Leroy F, Rayman JB, & Kandel ER (2019). Molecular Mechanisms of the Memory Trace. Trends Neurosci, 42, 14–22. [PubMed: 30391015]
- Barberi M, Muciaccia B, Morelli MB, Stefanini M, Cecconi S, & Canipari R (2007). Expression localisation and functional activity of pituitary adenylate cyclase-activating polypeptide, vasoactive intestinal polypeptide and their receptors in mouse ovary. Reproduction, 134, 281–292. [PubMed: 17660238]
- Basille M, Vaudry D, Coulouarn Y, Jegou S, Lihrmann I, Fournier A, Vaudry H, & Gonzalez B (2000). Comparative distribution of pituitary adenylate cyclase-activating polypeptide (PACAP) binding sites and PACAP receptor mRNAs in the rat brain during development. J Comp Neurol, 425, 495–509. [PubMed: 10975876]
- Birch NP, Hakes DJ, Dixon JE, & Mezey E (1994). Distribution and regulation of the candidate prohormone processing enzymes SPC2 and SPC3 in adult rat brain. Neuropeptides, 27, 307–322. [PubMed: 7898639]
- Boucher MN, May V, Braas KM, & Hammack SE (2021). PACAP orchestration of stress-related responses in neural circuits. Peptides, 142, 170554. [PubMed: 33865930]

- Brubel R, Reglodi D, Jambor E, Koppan M, Varnagy A, Biro Z, Kiss P, Gaal V, Matkovits A, Farkas J, Lubics A, Bodis J, Bay C, Veszpremi B, Tamas A, Nemeth J, & Mark L (2011). Investigation of pituitary adenylate cyclase activating polypeptide in human gynecological and other biological fluids by using MALDI TOF mass spectrometry. J Mass Spectrom, 46, 189–194. [PubMed: 21259400]
- Chan KY, Baun M, de Vries R, van den Bogaerdt AJ, Dirven CM, Danser AH, Jansen-Olesen I, Olesen J, Villalón CM, MaassenVanDenBrink A, & Gupta S (2011). Pharmacological characterization of VIP and PACAP receptors in the human meningeal and coronary artery. Cephalalgia, 31, 181–189. [PubMed: 20974589]
- Chang CH, Chey WY, Braggins L, Coy DH, & Chang TM (1996). Pituitary adenylate cyclaseactivating polypeptide stimulates cholecystokinin secretion in STC-1 cells. Am J Physiol, 271, G516–523. [PubMed: 8843778]
- Chang CH, Chey WY, Erway B, Coy DH, & Chang TM (1998). Modulation of secretin release by neuropeptides in secretin-producing cells. Am J Physiol, 275, G192–202. [PubMed: 9688645]
- Chang SC, Xie P, Anton RF, De Vivo I, Farrer LA, Kranzler HR, Oslin D, Purcell SM, Roberts AL, Smoller JW, Uddin M, Gelernter J, & Koenen KC (2012). No association between ADCYAP1R1 and post-traumatic stress disorder in two independent samples. Mol Psychiatry, 17, 239–241. [PubMed: 21912390]
- Cho JH, Zushida K, Shumyatsky GP, Carlezon WA Jr., Meloni EG, & Bolshakov VY (2012). Pituitary adenylate cyclase-activating polypeptide induces postsynaptically expressed potentiation in the intra-amygdala circuit. J Neurosci, 32, 14165–14177. [PubMed: 23055486]
- Christensen CE, Ashina M, & Amin FM (2022). Calcitonin Gene-Related Peptide (CGRP) and Pituitary Adenylate Cyclase-Activating Polypeptide (PACAP) in Migraine Pathogenesis. Pharmaceuticals (Basel), 15.
- Ciranna L, & Cavallaro S (2003). Opposing effects by pituitary adenylate cyclase-activating polypeptide and vasoactive intestinal peptide on hippocampal synaptic transmission. Exp Neurol, 184, 778–784. [PubMed: 14769370]
- Colwell CS, Michel S, Itri J, Rodriguez W, Tam J, Lelievre V, Hu Z, Liu X, & Waschek JA (2003). Disrupted circadian rhythms in VIP- and PHI-deficient mice. Am J Physiol Regul Integr Comp Physiol, 285, R939–949. [PubMed: 12855416]
- Condro MC, Matynia A, Foster NN, Ago Y, Rajbhandari AK, Van C, Jayaram B, Parikh S, Diep AL, Nguyen E, May V, Dong HW, & Waschek JA (2016). High-resolution characterization of a PACAP-EGFP transgenic mouse model for mapping PACAP-expressing neurons. J Comp Neurol, 524, 3827–3848. [PubMed: 27197019]
- Cooper AJ, Narasimhan S, Rickels K, & Lohoff FW (2013). Genetic polymorphisms in the PACAP and PAC1 receptor genes and treatment response to venlafaxine XR in generalized anxiety disorder. Psychiatry Res, 210, 1299–1300. [PubMed: 23972788]
- Costa L, Santangelo F, Li Volsi G, & Ciranna L (2009). Modulation of AMPA receptor-mediated ion current by pituitary adenylate cyclase-activating polypeptide (PACAP) in CA1 pyramidal neurons from rat hippocampus. Hippocampus, 19, 99–109. [PubMed: 18727050]
- Costa M, Spencer NJ, & Brookes SJH (2021). The role of enteric inhibitory neurons in intestinal motility. Auton Neurosci, 235, 102854. [PubMed: 34329834]
- Cox HM (1992). Pituitary adenylate cyclase activating polypeptides, PACAP-27 and PACAP-38: stimulators of electrogenic ion secretion in the rat small intestine. Br J Pharmacol, 106, 498–502. [PubMed: 1393275]
- Curtis GR, Gargiulo AT, Carpenter BA, Pirino BE, Hawks A, Coleman SA, Syed NA, Gupta A, & Barson JR (2023). Sex-related differences in endogenous pituitary adenylate cyclaseactivating polypeptide (PACAP) in the thalamic paraventricular nucleus: Implications for addiction neuroscience. Addict Neurosci, 5.
- Curtis GR, Oakes K, & Barson JR (2020). Expression and Distribution of Neuropeptide-Expressing Cells Throughout the Rodent Paraventricular Nucleus of the Thalamus. Front Behav Neurosci, 14, 634163. [PubMed: 33584216]

- Das M, Vihlen CS, & Legradi G (2007). Hypothalamic and brainstem sources of pituitary adenylate cyclase-activating polypeptide nerve fibers innervating the hypothalamic paraventricular nucleus in the rat. J Comp Neurol, 500, 761–776. [PubMed: 17154257]
- Denes V, Geck P, Mester A, & Gabriel R (2019). Pituitary Adenylate Cyclase-Activating Polypeptide: 30 Years in Research Spotlight and 600 Million Years in Service. J Clin Med, 8.
- Donahue RJ, Venkataraman A, Carroll FI, Meloni EG, & Carlezon WA Jr. (2016). Pituitary Adenylate Cyclase-Activating Polypeptide Disrupts Motivation, Social Interaction, and Attention in Male Sprague Dawley Rats. Biol Psychiatry, 80, 955–964. [PubMed: 26229039]
- Dow RC, Bennie J, & Fink G (1994). Pituitary adenylate cyclase-activating peptide-38 (PACAP)-38 is released into hypophysial portal blood in the normal male and female rat. J Endocrinol, 142, R1–4. [PubMed: 7964266]
- Duesman SJ, Shetty S, Patel S, Ogale N, Mohamed F, Sparman N, Rajbhandari P, & Rajbhandari AK (2022). Sexually dimorphic role of the locus coeruleus PAC1 receptors in regulating acute stress-associated energy metabolism. Front Behav Neurosci, 16, 995573. [PubMed: 36275856]
- Duvarci S, & Pare D (2014). Amygdala microcircuits controlling learned fear. Neuron, 82, 966–980. [PubMed: 24908482]
- Farnham MM, Li Q, Goodchild AK, & Pilowsky PM (2008). PACAP is expressed in sympathoexcitatory bulbospinal C1 neurons of the brain stem and increases sympathetic nerve activity in vivo. Am J Physiol Regul Integr Comp Physiol, 294, R1304–1311. [PubMed: 18272663]
- Feany MB, & Quinn WG (1995). A neuropeptide gene defined by the Drosophila memory mutant amnesiac. Science, 268, 869–873. [PubMed: 7754370]
- Ferragud A, Velazquez-Sanchez C, Minnig MA, Sabino V, & Cottone P (2021). Pituitary adenylate cyclase-activating polypeptide (PACAP) modulates dependence-induced alcohol drinking and anxiety-like behavior in male rats. Neuropsychopharmacology, 46, 509–518. [PubMed: 33191400]
- Fuchs M, Adermann K, Raab HR, Forssmann WG, & Kuhn M (1996). Pituitary adenylate cyclaseactivating polypeptide: a potent activator of human intestinal ion transport. Ann N Y Acad Sci, 805, 640–647. [PubMed: 8993454]
- Gargiulo AT, Pirino BE, Curtis GR, & Barson JR (2021). Effects of pituitary adenylate cyclaseactivating polypeptide isoforms in nucleus accumbens subregions on ethanol drinking. Addict Biol, 26, e12972. [PubMed: 33020973]
- Gaszner B, Kormos V, Kozicz T, Hashimoto H, Reglodi D, & Helyes Z (2012). The behavioral phenotype of pituitary adenylate-cyclase activating polypeptide-deficient mice in anxiety and depression tests is accompanied by blunted c-Fos expression in the bed nucleus of the stria terminalis, central projecting Edinger-Westphal nucleus, ventral lateral septum, and dorsal raphe nucleus. Neuroscience, 202, 283–299. [PubMed: 22178610]
- Ghanizada H, Al-Karagholi MA, Arngrim N, Ghanizada M, Larsson HBW, Amin FM, & Ashina M (2019). Effect of pituitary adenylate cyclase-activating polypeptide-27 on cerebral hemodynamics in healthy volunteers: A 3T MRI study. Peptides, 121, 170134. [PubMed: 31449829]
- Ghanizada H, Al-Karagholi MA, Arngrim N, Mørch-Rasmussen M, Metcalf-Clausen M, Larsson HBW, Amin FM, & Ashina M (2020). Investigation of sumatriptan and ketorolac trometamol in the human experimental model of headache. J Headache Pain, 21, 19. [PubMed: 32093617]
- Gilmartin MR, & Helmstetter FJ (2010). Trace and contextual fear conditioning require neural activity and NMDA receptor-dependent transmission in the medial prefrontal cortex. Learning & Memory, 17, 289–296. [PubMed: 20504949]
- Gilmartin MR, Kwapis JL, & Helmstetter FJ (2013). NR2A-and NR2B-containing NMDA receptors in the prelimbic medial prefrontal cortex differentially mediate trace, delay, and contextual fear conditioning. Learning & Memory, 20, 290–294. [PubMed: 23676200]
- Gräs S, Hannibal J, Georg B, & Fahrenkrug J (1996). Transient periovulatory expression of pituitary adenylate cyclase activating peptide in rat ovarian cells. Endocrinology, 137, 4779–4785. [PubMed: 8895347]
- Gräs S, Høst E, & Fahrenkrug J (2005). Role of pituitary adenylate cyclase-activating peptide (PACAP) in the cyclic recruitment of immature follicles in the rat ovary. Regul Pept, 128, 69–74. [PubMed: 15721490]

- Gray SL, Cummings KJ, Jirik FR, & Sherwood NM (2001). Targeted disruption of the pituitary adenylate cyclase-activating polypeptide gene results in early postnatal death associated with dysfunction of lipid and carbohydrate metabolism. Mol Endocrinol, 15, 1739–1747. [PubMed: 11579206]
- Gray SL, Yamaguchi N, Vencová P, & Sherwood NM (2002). Temperature-sensitive phenotype in mice lacking pituitary adenylate cyclase-activating polypeptide. Endocrinology, 143, 3946–3954. [PubMed: 12239106]
- Gupta A, Gargiulo AT, Curtis GR, Badve PS, Pandey S, & Barson JR (2018). Pituitary Adenylate Cyclase-Activating Polypeptide-27 (PACAP-27) in the Thalamic Paraventricular Nucleus Is Stimulated by Ethanol Drinking. Alcohol Clin Exp Res, 42, 1650–1660. [PubMed: 29969146]
- Håkanson R, Chen D, Lindström E, Bernsand M, & Norlén P (2001). Control of secretion from rat stomach ECL cells in situ and in primary culture. Scand J Clin Lab Invest Suppl, 234, 53–60. [PubMed: 11713981]
- Hamelink C, Tjurmina O, Damadzic R, Young WS, Weihe E, Lee HW, & Eiden LE (2002). Pituitary adenylate cyclase-activating polypeptide is a sympathoadrenal neurotransmitter involved in catecholamine regulation and glucohomeostasis. Proc Natl Acad Sci U S A, 99, 461–466. [PubMed: 11756684]
- Hammack SE, Cheung J, Rhodes KM, Schutz KC, Falls WA, Braas KM, & May V (2009). Chronic stress increases pituitary adenylate cyclase-activating peptide (PACAP) and brain-derived neurotrophic factor (BDNF) mRNA expression in the bed nucleus of the stria terminalis (BNST): roles for PACAP in anxiety-like behavior. Psychoneuroendocrinology, 34, 833–843. [PubMed: 19181454]
- Hammack SE, & May V (2015). Pituitary adenylate cyclase activating polypeptide in stress-related disorders: data convergence from animal and human studies. Biol Psychiatry, 78, 167–177. [PubMed: 25636177]
- Hannibal J (2002). Pituitary adenylate cyclase-activating peptide in the rat central nervous system: an immunohistochemical and in situ hybridization study. The Journal of comparative neurology, 453.
- Hannibal J, Mikkelsen JD, Clausen H, Holst JJ, Wulff BS, & Fahrenkrug J (1995). Gene expression of pituitary adenylate cyclase activating polypeptide (PACAP) in the rat hypothalamus. Regul Pept, 55, 133–148. [PubMed: 7754101]
- Harmar AJ, Fahrenkrug J, Gozes I, Laburthe M, May V, Pisegna JR, Vaudry D, Vaudry H, Waschek JA, & Said SI (2012). Pharmacology and functions of receptors for vasoactive intestinal peptide and pituitary adenylate cyclase-activating polypeptide: IUPHAR review 1. Br J Pharmacol, 166, 4–17. [PubMed: 22289055]
- Hashimoto H, Nogi H, Mori K, Ohishi H, Shigemoto R, Yamamoto K, Matsuda T, Mizuno N, Nagata S, & Baba A (1996). Distribution of the mRNA for a pituitary adenylate cyclase-activating polypeptide receptor in the rat brain: an in situ hybridization study. J Comp Neurol, 371, 567–577. [PubMed: 8841910]
- Hashimoto H, Shintani N, & Baba A (2006). New insights into the central PACAPergic system from the phenotypes in PACAP- and PACAP receptor-knockout mice. Ann N Y Acad Sci, 1070, 75–89. [PubMed: 16888150]
- Hashimoto H, Shintani N, Tanaka K, Mori W, Hirose M, Matsuda T, Sakaue M, Miyazaki J, Niwa H, Tashiro F, Yamamoto K, Koga K, Tomimoto S, Kunugi A, Suetake S, & Baba A (2001). Altered psychomotor behaviors in mice lacking pituitary adenylate cyclase-activating polypeptide (PACAP). Proc Natl Acad Sci U S A, 98, 13355–13360. [PubMed: 11687615]
- Hashimoto H, Shintani N, Tanida M, Hayata A, Hashimoto R, & Baba A (2011). PACAP is implicated in the stress axes. Curr Pharm Des, 17, 985–989. [PubMed: 21524255]
- Hashimoto R, Hashimoto H, Shintani N, Chiba S, Hattori S, Okada T, Nakajima M, Tanaka K, Kawagishi N, Nemoto K, Mori T, Ohnishi T, Noguchi H, Hori H, Suzuki T, Iwata N, Ozaki N, Nakabayashi T, Saitoh O, Kosuga A, Tatsumi M, Kamijima K, Weinberger DR, Kunugi H, & Baba A (2007). Pituitary adenylate cyclase-activating polypeptide is associated with schizophrenia. Mol Psychiatry, 12, 1026–1032. [PubMed: 17387318]
- Hashimoto R, Hashimoto H, Shintani N, Ohi K, Hori H, Saitoh O, Kosuga A, Tatsumi M, Iwata N, Ozaki N, Kamijima K, Baba A, Takeda M, & Kunugi H (2010). Possible association between the

pituitary adenylate cyclase-activating polypeptide (PACAP) gene and major depressive disorder. Neurosci Lett, 468, 300–302. [PubMed: 19914336]

- Hayata-Takano A, Shintani Y, Moriguchi K, Encho N, Kitagawa K, Nakazawa T, & Hashimoto H (2021). PACAP-PAC1 Signaling Regulates Serotonin 2A Receptor Internalization. Front Endocrinol (Lausanne), 12, 732456. [PubMed: 34759890]
- Heinzlmann A, Kirilly E, Meltzer K, Szabó E, Baba A, Hashimoto H, & Köves K (2008). PACAP is transiently expressed in anterior pituitary gland of rats: in situ hybridization and cell immunoblot assay studies. Peptides, 29, 571–577. [PubMed: 18243417]
- Heroux NA, Robinson-Drummer PA, Sanders HR, Rosen JB, & Stanton ME (2017). Differential involvement of the medial prefrontal cortex across variants of contextual fear conditioning. Learn Mem, 24, 322–330. [PubMed: 28716952]
- Holgert H, Holmberg K, Hannibal J, Fahrenkrug J, Brimijoin S, Hartman BK, & Hökfelt T (1996). PACAP in the adrenal gland--relationship with choline acetyltransferase, enkephalin and chromaffin cells and effects of immunological sympathectomy. Neuroreport, 8, 297–301. [PubMed: 9051799]
- Holighaus Y, Mustafa T, & Eiden LE (2011). PAC1hop, null and hip receptors mediate differential signaling through cyclic AMP and calcium leading to splice variant-specific gene induction in neural cells. Peptides, 32, 1647–1655. [PubMed: 21693142]
- Horvath G, Opper B, & Reglodi D (2019). The Neuropeptide Pituitary Adenylate Cyclase-Activating Polypeptide (PACAP) is Protective in Inflammation and Oxidative Stress-Induced Damage in the Kidney. Int J Mol Sci, 20.
- Iemolo A, Ferragud A, Cottone P, & Sabino V (2015). Pituitary Adenylate Cyclase-Activating Peptide in the Central Amygdala Causes Anorexia and Body Weight Loss via the Melanocortin and the TrkB Systems. Neuropsychopharmacology, 40, 1846–1855. [PubMed: 25649277]
- Iemolo A, Seiglie M, Blasio A, Cottone P, & Sabino V (2016). Pituitary adenylate cyclase-activating polypeptide (PACAP) in the central nucleus of the amygdala induces anxiety via melanocortin receptors. Psychopharmacology (Berl), 233, 3269–3277. [PubMed: 27376948]
- Isaac ER, & Sherwood NM (2008). Pituitary adenylate cyclase-activating polypeptide (PACAP) is important for embryo implantation in mice. Mol Cell Endocrinol, 280, 13–19. [PubMed: 17945412]
- Ishiguro H, Ohtsuki T, Okubo Y, Kurumaji A, & Arinami T (2001). Association analysis of the pituitary adenyl cyclase activating peptide gene (PACAP) on chromosome 18p11 with schizophrenia and bipolar disorders. J Neural Transm (Vienna), 108, 849–854. [PubMed: 11515750]
- Ishihama T, Ago Y, Shintani N, Hashimoto H, Baba A, Takuma K, & Matsuda T (2010). Environmental factors during early developmental period influence psychobehavioral abnormalities in adult PACAP-deficient mice. Behavioural brain research, 209.
- Ishihara T, Shigemoto R, Mori K, Takahashi K, & Nagata S (1992). Functional expression and tissue distribution of a novel receptor for vasoactive intestinal polypeptide. Neuron, 8, 811–819. [PubMed: 1314625]
- Iwasa T, Matsuzaki T, Tungalagsuvd A, Munkhzaya M, Yiliyasi M, Kato T, Kuwahara A, & Irahara M (2016). Developmental changes in the hypothalamic mRNA expression levels of PACAP and its receptor PAC1 and their sensitivity to fasting in male and female rats. Int J Dev Neurosci, 52, 33–37. [PubMed: 27181029]
- Jamen F, Persson K, Bertrand G, Rodriguez-Henche N, Puech R, Bockaert J, Ahrén B, & Brabet P (2000a). PAC1 receptor-deficient mice display impaired insulinotropic response to glucose and reduced glucose tolerance. J Clin Invest, 105, 1307–1315. [PubMed: 10792006]
- Jamen F, Rodriguez-Henche N, Pralong F, Jegou B, Gaillard R, Bockaert J, & Brabet P (2000b). PAC1 null females display decreased fertility. Ann N Y Acad Sci, 921, 400–404. [PubMed: 11193864]
- Jaworski DM, & Proctor MD (2000). Developmental regulation of pituitary adenylate cyclaseactivating polypeptide and PAC(1) receptor mRNA expression in the rat central nervous system. Brain research. Developmental brain research, 120.

- Johnson GC, May V, Parsons RL, & Hammack SE (2019). Parallel signaling pathways of pituitary adenylate cyclase activating polypeptide (PACAP) regulate several intrinsic ion channels. Ann N Y Acad Sci.
- Johnson GC, Parsons R, May V, & Hammack SE (2020a). The Role of Pituitary Adenylate Cyclase-Activating Polypeptide (PACAP) Signaling in the Hippocampal Dentate Gyrus. Front Cell Neurosci, 14, 111. [PubMed: 32425759]
- Johnson GC, Parsons RL, May V, & Hammack SE (2020b). Pituitary adenylate cyclase-activating polypeptide-induced PAC1 receptor internalization and recruitment of MEK/ERK signaling enhance excitability of dentate gyrus granule cells. Am J Physiol Cell Physiol, 318, C870–c878. [PubMed: 32186931]
- Jolivel V, Basille M, Aubert N, de Jouffrey S, Ancian P, Le Bigot JF, Noack P, Massonneau M, Fournier A, Vaudry H, Gonzalez BJ, & Vaudry D (2009). Distribution and functional characterization of pituitary adenylate cyclase-activating polypeptide receptors in the brain of non-human primates. Neuroscience, 160, 434–451. [PubMed: 19236905]
- Joo KM, Chung YH, Kim MK, Nam RH, Lee BL, Lee KH, & Cha CI (2004). Distribution of vasoactive intestinal peptide and pituitary adenylate cyclase-activating polypeptide receptors (VPAC1, VPAC2, and PAC1 receptor) in the rat brain. J Comp Neurol, 476, 388–413. [PubMed: 15282712]
- Jovanovic T, Norrholm SD, Davis J, Mercer KB, Almli L, Nelson A, Cross D, Smith A, Ressler KJ, & Bradley B (2013). PAC1 receptor (ADCYAP1R1) genotype is associated with dark-enhanced startle in children, Mol Psychiatry (pp. 742–743). England.
- Jovanovic T, Stenson AF, Thompson N, Clifford A, Compton A, Minton S, van Rooij SJF, Stevens JS, Lori A, Nugent N, Gillespie CF, Bradley B, & Ressler KJ (2020). Impact of ADCYAP1R1 genotype on longitudinal fear conditioning in children: interaction with trauma and sex. Neuropsychopharmacology : official publication of the American College of Neuropsychopharmacology, 45.
- Kandel ER, Dudai Y, & Mayford MR (2014). The molecular and systems biology of memory. Cell, 157, 163–186. [PubMed: 24679534]
- Kántora O, Molnár J, Arimura A, & Köves K (2000). PACAP38 and PACAP27 administered intracerebroventricularly have an opposite effect on LH secretion. Peptides, 21, 817–820. [PubMed: 10959003]
- Karpiesiuk A, & Palus K (2021). Pituitary Adenylate Cyclase-Activating Polypeptide (PACAP) in Physiological and Pathological Processes within the Gastrointestinal Tract: A Review. Int J Mol Sci, 22.
- Kimura C, Ohkubo S, Ogi K, Hosoya M, Itoh Y, Onda H, Miyata A, Jiang L, Dahl RR, Stibbs HH, & et al. (1990). A novel peptide which stimulates adenylate cyclase: molecular cloning and characterization of the ovine and human cDNAs. Biochem Biophys Res Commun, 166, 81–89. [PubMed: 2302217]
- King SB, Lezak KR, O'Reilly M, Toufexis DJ, Falls WA, Braas K, May V, & Hammack SE (2017). The Effects of Prior Stress on Anxiety-Like Responding to Intra-BNST Pituitary Adenylate Cyclase Activating Polypeptide in Male and Female Rats. Neuropsychopharmacology, 42, 1679– 1687. [PubMed: 28106040]
- Kirry AJ, Herbst MR, Poirier SE, Maskeri MM, Rothwell AC, Twining RC, & Gilmartin MR (2018). Pituitary adenylate cyclase-activating polypeptide (PACAP) signaling in the prefrontal cortex modulates cued fear learning, but not spatial working memory, in female rats. Neuropharmacology, 133, 145–154. [PubMed: 29353055]
- Kiss P, Reglodi D, Tamás A, Lubics A, Lengvári I, Józsa R, Somogyvári-Vigh A, Szilvássy Z, & Németh J (2007). Changes of PACAP levels in the brain show gender differences following shortterm water and food deprivation. Gen Comp Endocrinol, 152, 225–230. [PubMed: 17286974]
- Kivipelto L, Absood A, Arimura A, Sundler F, Håkanson R, & Panula P (1992). The distribution of pituitary adenylate cyclase-activating polypeptide-like immunoreactivity is distinct from helodermin- and helospectin-like immunoreactivities in the rat brain. J Chem Neuroanat, 5, 85–94. [PubMed: 1605915]
- Knutsson M, & Edvinsson L (2002). Distribution of mRNA for VIP and PACAP receptors in human cerebral arteries and cranial ganglia. Neuroreport, 13, 507–509. [PubMed: 11930171]

- Kobayashi K, Shihoya W, Nishizawa T, Kadji FMN, Aoki J, Inoue A, & Nureki O (2020). Cryo-EM structure of the human PAC1 receptor coupled to an engineered heterotrimeric G protein. Nat Struct Mol Biol, 27, 274–280. [PubMed: 32157248]
- Kocho-Schellenberg M, Lezak KR, Harris OM, Roelke E, Gick N, Choi I, Edwards S, Wasserman E, Toufexis DJ, Braas KM, May V, & Hammack SE (2014). PACAP in the BNST produces anorexia and weight loss in male and female rats. Neuropsychopharmacology, 39, 1614–1623. [PubMed: 24434744]
- Kondo T, Tominaga T, Ichikawa M, & Iijima T (1997). Differential alteration of hippocampal synaptic strength induced by pituitary adenylate cyclase activating polypeptide-38 (PACAP-38). Neurosci Lett, 221, 189–192. [PubMed: 9121696]
- Koppan M, Nagy Z, Bosnyak I, & Reglodi D (2022). Female reproductive functions of the neuropeptide PACAP. Front Endocrinol (Lausanne), 13, 982551. [PubMed: 36204113]
- Koppan M, Varnagy A, Reglodi D, Brubel R, Nemeth J, Tamas A, Mark L, & Bodis J (2012). Correlation between oocyte number and follicular fluid concentration of pituitary adenylate cyclaseactivating polypeptide (PACAP) in women after superovulation treatment. J Mol Neurosci, 48, 617–622. [PubMed: 22415357]
- Kormos V, Gáspár L, Kovács L, Farkas J, Gaszner T, Csernus V, Balogh A, Hashimoto H, Regl di D, Helyes Z, & Gaszner B (2016). Reduced response to chronic mild stress in PACAP mutant mice is associated with blunted FosB expression in limbic forebrain and brainstem centers. Neuroscience, 330, 335–358. [PubMed: 27282087]
- Köves K, Arimura A, Görcs TG, & Somogyvári-Vigh A (1991). Comparative distribution of immunoreactive pituitary adenylate cyclase activating polypeptide and vasoactive intestinal polypeptide in rat forebrain. Neuroendocrinology, 54, 159–169. [PubMed: 1766552]
- Köves K, Arimura A, Somogyvári-Vigh A, Vigh S, & Miller J (1990). Immunohistochemical demonstration of a novel hypothalamic peptide, pituitary adenylate cyclase-activating polypeptide, in the ovine hypothalamus. Endocrinology, 127, 264–271. [PubMed: 2193797]
- Köves K, Kántor O, Scammell JG, & Arimura A (1998). PACAP colocalizes with luteinizing and follicle-stimulating hormone immunoreactivities in the anterior lobe of the pituitary gland. Peptides, 19, 1069–1072. [PubMed: 9700757]
- Kozicz T, & Arimura A (2002). Dopamine- and cyclic AMP-regulated phosphoproteinimmunoreactive neurons activated by acute stress are innervated by fiber terminals immunopositive for pituitary adenylate cyclase-activating polypeptide in the extended amygdala in the rat. Regul Pept, 109, 63–70. [PubMed: 12409216]
- Kumar NN, Allen K, Parker L, Damanhuri H, & Goodchild AK (2010). Neuropeptide coding of sympathetic preganglionic neurons; focus on adrenally projecting populations. Neuroscience, 170, 789–799. [PubMed: 20674686]
- Kupari J, Häring M, Agirre E, Castelo-Branco G, & Ernfors P (2019). An Atlas of Vagal Sensory Neurons and Their Molecular Specialization. Cell Rep, 27, 2508–2523.e2504. [PubMed: 31116992]
- Lajko A, Meggyes M, Fulop BD, Gede N, Reglodi D, & Szereday L (2018). Comparative analysis of decidual and peripheral immune cells and immune-checkpoint molecules during pregnancy in wild-type and PACAP-deficient mice. Am J Reprod Immunol, 80, e13035. [PubMed: 30091267]
- Langer I, Jeandriens J, Couvineau A, Sanmukh S, & Latek D (2022). Signal Transduction by VIP and PACAP Receptors. Biomedicines, 10.
- Läuffer JM, Tang LH, Zhang T, Hinoue T, Rahbar S, Odo M, Modlin IM, & Kidd M (2001). PACAP mediates the neural proliferative pathway of Mastomys enterochromaffin-like cell transformation. Regul Pept, 102, 157–164. [PubMed: 11730988]
- Lee J, Park HJ, Choi HS, Kwon HB, Arimura A, Lee BJ, Choi WS, & Chun SY (1999). Gonadotropin stimulation of pituitary adenylate cyclase-activating polypeptide (PACAP) messenger ribonucleic acid in the rat ovary and the role of PACAP as a follicle survival factor. Endocrinology, 140, 818–826. [PubMed: 9927311]
- Legradi G, Das M, Giunta B, Hirani K, Mitchell EA, & Diamond DM (2007). Microinfusion of pituitary adenylate cyclase-activating polypeptide into the central nucleus of amygdala of the

rat produces a shift from an active to passive mode of coping in the shock-probe fear/defensive burying test. Neural Plast, 2007, 79102. [PubMed: 17641738]

- Légrádi G, Shioda S, & Arimura A (1994). Pituitary adenylate cyclase-activating polypeptide-like immunoreactivity in autonomic regulatory areas of the rat medulla oblongata. Neurosci Lett, 176, 193196.
- Lehmann ML, Mustafa T, Eiden AM, Herkenham M, & Eiden LE (2013). PACAP-deficient mice show attenuated corticosterone secretion and fail to develop depressive behavior during chronic social defeat stress. Psychoneuroendocrinology, 38, 702–715. [PubMed: 23062748]
- Li M, Shuto Y, Somogyvári-Vigh A, & Arimura A (1999). Prohormone convertases 1 and 2 process ProPACAP and generate matured, bioactive PACAP38 and PACAP27 in transfected rat pituitary GH4C1 cells. Neuroendocrinology, 69, 217–226. [PubMed: 10087454]
- Liang YL, Belousoff MJ, Zhao P, Koole C, Fletcher MM, Truong TT, Julita V, Christopoulos G, Xu HE, Zhang Y, Khoshouei M, Christopoulos A, Danev R, Sexton PM, & Wootten D (2020). Toward a Structural Understanding of Class B GPCR Peptide Binding and Activation. Mol Cell, 77, 656–668.e655. [PubMed: 32004469]
- Lind MJ, Marraccini ME, Sheerin CM, Bountress K, Bacanu SA, Amstadter AB, & Nugent NR (2017). Association of Posttraumatic Stress Disorder With rs2267735 in the ADCYAP1R1 Gene: A Meta-Analysis. J Trauma Stress, 30, 389–398. [PubMed: 28746747]
- Lu J, Piper SJ, Zhao P, Miller LJ, Wootten D, & Sexton PM (2022). Targeting VIP and PACAP Receptor Signaling: New Insights into Designing Drugs for the PACAP Subfamily of Receptors. Int J Mol Sci, 23.
- Lutfy K, & Shankar G (2019). Emerging evidence for the role of pituitary adenylate cyclase-activating peptide in neuropsychiatric disorders. Prog Mol Biol Transl Sci, 167, 143–157. [PubMed: 31601402]
- Lutz EM, Sheward WJ, West KM, Morrow JA, Fink G, & Harmar AJ (1993). The VIP2 receptor: molecular characterisation of a cDNA encoding a novel receptor for vasoactive intestinal peptide. FEBS Lett, 334, 3–8. [PubMed: 8224221]
- Lutz-Bucher B, Monnier D, & Koch B (1996). Evidence for the presence of receptors for pituitary adenylate cyclase-activating polypeptide in the neurohypophysis that are positively coupled to cyclic AMP formation and neurohypophyseal hormone secretion. Neuroendocrinology, 64, 153– 161. [PubMed: 8857610]
- Macdonald DS, Weerapura M, Beazely MA, Martin L, Czerwinski W, Roder JC, Orser BA, & MacDonald JF (2005). Modulation of NMDA receptors by pituitary adenylate cyclase activating peptide in CA1 neurons requires G alpha q, protein kinase C, and activation of Src. J Neurosci, 25, 11374–11384. [PubMed: 16339032]
- MacKenzie CJ, Lutz EM, Johnson MS, Robertson DN, Holland PJ, & Mitchell R (2001). Mechanisms of phospholipase C activation by the vasoactive intestinal polypeptide/pituitary adenylate cyclase-activating polypeptide type 2 receptor. Endocrinology, 142, 1209–1217. [PubMed: 11181537]
- Maita I, Roepke TA, & Samuels BA (2022). Chronic stress-induced synaptic changes to corticotropinreleasing factor-signaling in the bed nucleus of the stria terminalis. Front Behav Neurosci, 16, 903782. [PubMed: 35983475]
- Marquez P, Bebawy D, Lelièvre V, Coûté AC, Evans CJ, Waschek JA, & Lutfy K (2009). The role of endogenous PACAP in motor stimulation and conditioned place preference induced by morphine in mice. Psychopharmacology (Berl), 204, 457–463. [PubMed: 19199096]
- Martelle SE, Cotella EM, Nawreen N, Chen C, Packard BA, Fitzgerald M, & Herman JP (2021). Prefrontal cortex PACAP signaling: organization and role in stress regulation. Stress, 24, 196– 205. [PubMed: 33726625]
- Martin B, Lopez de Maturana R, Brenneman R, Walent T, Mattson MP, & Maudsley S (2005). Class II G protein-coupled receptors and their ligands in neuronal function and protection. Neuromolecular Med, 7, 3–36. [PubMed: 16052036]
- Masuo Y, Suzuki N, Matsumoto H, Tokito F, Matsumoto Y, Tsuda M, & Fujino M (1993). Regional distribution of pituitary adenylate cyclase activating polypeptide (PACAP) in the rat central

nervous system as determined by sandwich-enzyme immunoassay. Brain Res, 602, 57–63. [PubMed: 8095427]

- Matsuyama S, Matsumoto A, Hashimoto H, Shintani N, & Baba A (2003). Impaired long-term potentiation in vivo in the dentate gyrus of pituitary adenylate cyclase-activating polypeptide (PACAP) or PACAP type 1 receptor-mutant mice. Neuroreport, 14.
- Meloni EG, Kaye KT, Venkataraman A, & Carlezon WA Jr. (2018). PACAP increases Arc/Arg 3.1 expression within the extended amygdala after fear conditioning in rats. Neurobiol Learn Mem, 157, 24–34. [PubMed: 30458282]
- Meloni EG, Kaye KT, Venkataraman A, & Carlezon WA Jr. (2019). PACAP increases Arc/Arg 3.1 expression within the extended amygdala after fear conditioning in rats. Neurobiol Learn Mem, 157, 24–34. [PubMed: 30458282]
- Meloni EG, Venkataraman A, Donahue RJ, & Carlezon WA Jr. (2016). Bi-directional effects of pituitary adenylate cyclase-activating polypeptide (PACAP) on fear-related behavior and c-Fos expression after fear conditioning in rats. Psychoneuroendocrinology, 64, 12–21. [PubMed: 26590791]
- Mercer KB, Dias B, Shafer D, Maddox SA, Mulle JG, Hu P, Walton J, & Ressler KJ (2016). Functional evaluation of a PTSD-associated genetic variant: estradiol regulation and ADCYAP1R1. Transl Psychiatry, 6, e978. [PubMed: 27959335]
- Miles OW, May V, & Hammack SE (2019). Pituitary Adenylate Cyclase-Activating Peptide (PACAP) Signaling and the Dark Side of Addiction. J Mol Neurosci, 68, 453–464. [PubMed: 30074172]
- Miles OW, Thrailkill EA, Linden AK, May V, Bouton ME, & Hammack SE (2018). Pituitary Adenylate Cyclase-Activating Peptide in the Bed Nucleus of the Stria Terminalis Mediates Stress-Induced Reinstatement of Cocaine Seeking in Rats. Neuropsychopharmacology, 43, 978– 986. [PubMed: 28656976]
- Missig G, Roman CW, Vizzard MA, Braas KM, Hammack SE, & May V (2014). Parabrachial nucleus (PBn) pituitary adenylate cyclase activating polypeptide (PACAP) signaling in the amygdala: implication for the sensory and behavioral effects of pain. Neuropharmacology, 86, 38–48. [PubMed: 24998751]
- Miyata A, Jiang L, Dahl RD, Kitada C, Kubo K, Fujino M, Minamino N, & Arimura A (1990). Isolation of a neuropeptide corresponding to the N-terminal 27 residues of the pituitary adenylate cyclase activating polypeptide with 38 residues (PACAP38). Biochem Biophys Res Commun, 170, 643–648. [PubMed: 2383262]
- Moore JP Jr., Burger LL, Dalkin AC, & Winters SJ (2005). Pituitary adenylate cyclase activating polypeptide messenger RNA in the paraventricular nucleus and anterior pituitary during the rat estrous cycle. Biol Reprod, 73, 491–499. [PubMed: 15917345]
- Mosca EV, Rousseau JP, Gulemetova R, Kinkead R, & Wilson RJ (2015). The effects of sex and neonatal stress on pituitary adenylate cyclase-activating peptide expression. Exp Physiol, 100, 203–215. [PubMed: 25398710]
- Murase T, Kondo K, Arima H, Iwasaki Y, Ito M, Miura Y, & Oiso Y (1995). The expression of pituitary adenylate cyclase-activating polypeptide (PACAP) mRNA in rat brain: possible role of endogenous PACAP in vasopressin release. Neurosci Lett, 185, 103–106. [PubMed: 7746497]
- Murase T, Kondo K, Otake K, & Oiso Y (1993). Pituitary adenylate cyclase-activating polypeptide stimulates arginine vasopressin release in conscious rats. Neuroendocrinology, 57, 1092–1096. [PubMed: 7901784]
- Mustafa T (2013). Pituitary adenylate cyclase-activating polypeptide (PACAP): a master regulator in central and peripheral stress responses. Adv Pharmacol, 68, 445–457. [PubMed: 24054157]
- Nagashima T, Tohyama S, Mikami K, Nagase M, Morishima M, Kasai A, Hashimoto H, & Watabe AM (2022). Parabrachial-to-parasubthalamic nucleus pathway mediates fear-induced suppression of feeding in male mice. Nat Commun, 13, 7913. [PubMed: 36585411]
- Nakata M, & Yada T (2004). [Physiological and therapeutic roles of PACAP in glucose metabolism and diabetes]. Nihon Yakurigaku Zasshi, 123, 267–273. [PubMed: 15056942]
- Naruse S, Nakamura T, Wei M, Ando E, Nokihara K, Wray V, Ozaki T, Kitagawa M, & Hayakawa T (1996). Effects of PACAP-VIP hybrid peptides on gastric blood flow in conscious dogs. Ann N Y Acad Sci, 805, 511–515. [PubMed: 8993432]

- Nielsen HS, Hannibal J, & Fahrenkrug J (1998). Expression of pituitary adenylate cyclase activating polypeptide (PACAP) in the postnatal and adult rat cerebellar cortex. Neuroreport, 9, 2639–2642. [PubMed: 9721947]
- Nogi H, Hashimoto H, Hagihara N, Shimada S, Yamamoto K, Matsuda T, Tohyama M, & Baba A (1997). Distribution of mRNAs for pituitary adenylate cyclase-activating polypeptide (PACAP), PACAP receptor, vasoactive intestinal polypeptide (VIP), and VIP receptors in the rat superior cervical ganglion. Neurosci Lett, 227, 37–40. [PubMed: 9178853]
- Nomura M, Ueta Y, Serino R, Yamamoto Y, Shibuya I, & Yamashita H (1999). Effects of centrally administered pituitary adenylate cyclase-activating polypeptide on c-fos gene expression and heteronuclear RNA for vasopressin in rat paraventricular and supraoptic nuclei. Neuroendocrinology, 69, 167–180. [PubMed: 10087449]
- Nussdorfer GG, & Malendowicz LK (1998). Role of VIP, PACAP, and related peptides in the regulation of the hypothalamo-pituitary-adrenal axis. Peptides, 19, 1443–1467. [PubMed: 9809661]
- Ogi K, Kimura C, Onda H, Arimura A, & Fujino M (1990). Molecular cloning and characterization of cDNA for the precursor of rat pituitary adenylate cyclase activating polypeptide (PACAP). Biochem Biophys Res Commun, 173, 1271–1279. [PubMed: 2268329]
- Oh DS, Lieu SN, Yamaguchi DJ, Tachiki K, Lambrecht N, Ohning GV, Sachs G, Germano PM, & Pisegna JR (2005). PACAP regulation of secretion and proliferation of pure populations of gastric ECL cells. J Mol Neurosci, 26, 85–97. [PubMed: 15968088]
- Ohkubo S, Kimura C, Ogi K, Okazaki K, Hosoya M, Onda H, Miyata A, Arimura A, & Fujino M (1992). Primary structure and characterization of the precursor to human pituitary adenylate cyclase activating polypeptide. DNA Cell Biol, 11, 21–30. [PubMed: 1739432]
- Okazaki K, Itoh Y, Ogi K, Ohkubo S, & Onda H (1995). Characterization of murine PACAP mRNA. Peptides, 16, 1295–1299. [PubMed: 8545254]
- Otto C, Kovalchuk Y, Wolfer DP, Gass P, Martin M, Zuschratter W, Grone HJ, Kellendonk C, Tronche F, Maldonado R, Lipp HP, Konnerth A, & Schutz G (2001a). Impairment of mossy fiber longterm potentiation and associative learning in pituitary adenylate cyclase activating polypeptide type I receptor-deficient mice. J Neurosci, 21, 5520–5527. [PubMed: 11466423]
- Otto C, Martin M, Wolfer DP, Lipp HP, Maldonado R, & Schutz G (2001b). Altered emotional behavior in PACAP-type-I-receptor-deficient mice. Brain Res Mol Brain Res, 92, 78–84. [PubMed: 11483244]
- Palkovits M, Somogyvári-Vigh A, & Arimura A (1995). Concentrations of pituitary adenylate cyclase activating polypeptide (PACAP) in human brain nuclei. Brain Res, 699, 116–120. [PubMed: 8616598]
- Park JY, Park JH, Park HJ, Lee JY, Lee YI, Lee K, & Chun SY (2001). Stage-dependent regulation of ovarian pituitary adenylate cyclase-activating polypeptide mRNA levels by GnRH in cultured rat granulosa cells. Endocrinology, 142, 3828–3835. [PubMed: 11517159]
- Parker DB, Coe IR, Dixon GH, & Sherwood NM (1993). Two salmon neuropeptides encoded by one brain cDNA are structurally related to members of the glucagon superfamily. Eur J Biochem, 215, 439–448. [PubMed: 8344311]
- Parsons RL, & May V (2019). PACAP-Induced PAC1 Receptor Internalization and Recruitment of Endosomal Signaling Regulate Cardiac Neuron Excitability. J Mol Neurosci, 68, 340–347. [PubMed: 30054797]
- Pecoraro V, Sardone LM, Chisari M, Licata F, Li Volsi G, Perciavalle V, Ciranna L, & Costa L (2017). A subnanomolar concentration of Pituitary Adenylate Cyclase-Activating Polypeptide (PACAP) pre-synaptically modulates glutamatergic transmission in the rat hippocampus acting through acetylcholine. Neuroscience, 340, 551–562. [PubMed: 27816700]
- Petersen B, Buchfelder M, Fahlbusch R, & Adams EF (1996). Pituitary adenylate cyclase-activating polypeptide directly stimulates LH and FSH secretion by human pituitary gonadotrophinomas. Exp Clin Endocrinol Diabetes, 104, 250–255. [PubMed: 8817243]
- Piggins HD, Stamp JA, Burns J, Rusak B, & Semba K (1996). Distribution of pituitary adenylate cyclase activating polypeptide (PACAP) immunoreactivity in the hypothalamus and extended amygdala of the rat. J Comp Neurol, 376, 278–294. [PubMed: 8951643]

- Pirino BE, & Barson JR (2021). A little night(PA)CAP: pituitary adenylate cyclase-activating polypeptide mediates behavioral effects of alcohol withdrawal. Neuropsychopharmacology, 46, 489–490. [PubMed: 33257754]
- Pisegna JR, & Wank SA (1993). Molecular cloning and functional expression of the pituitary adenylate cyclase-activating polypeptide type I receptor. Proc Natl Acad Sci U S A, 90, 6345–6349. [PubMed: 8392197]
- Racké K, Reimann A, Schwörer H, & Kilbinger H (1996). Regulation of 5-HT release from enterochromaffin cells. Behav Brain Res, 73, 83–87. [PubMed: 8788482]
- Rajbhandari AK, Octeau CJ, Gonzalez S, Pennington ZT, Mohamed F, Trott J, Chavez J, Ngyuen E, Keces N, Hong WZ, Neve RL, Waschek J, Khakh BS, & Fanselow MS (2021). A Basomedial Amygdala to Intercalated Cells Microcircuit Expressing PACAP and Its Receptor PAC1 Regulates Contextual Fear. J Neurosci, 41, 3446–3461. [PubMed: 33637560]
- Ramikie TS, & Ressler KJ (2016). Stress-related disorders, pituitary adenylate cyclase-activating peptide (PACAP)ergic system, and sex differences. Dialogues Clin Neurosci, 18, 403–413. [PubMed: 28179812]
- Ramos-Álvarez I, Mantey SA, Nakamura T, Nuche-Berenguer B, Moreno P, Moody TW, Maderdrut JL, Coy DH, & Jensen RT (2015). A structure-function study of PACAP using conformationally restricted analogs: Identification of PAC1 receptor-selective PACAP agonists. Peptides, 66, 26– 42. [PubMed: 25698233]
- Reglodi D, Illes A, Opper B, Schafer E, Tamas A, & Horvath G (2018). Presence and Effects of Pituitary Adenylate Cyclase Activating Polypeptide Under Physiological and Pathological Conditions in the Stomach. Front Endocrinol (Lausanne), 9, 90. [PubMed: 29615974]
- Ressler KJ, Mercer KB, Bradley B, Jovanovic T, Mahan A, Kerley K, Norrholm SD, Kilaru V, Smith AK, Myers AJ, Ramirez M, Engel A, Hammack SE, Toufexis D, Braas KM, Binder EB, & May V (2011). Post-traumatic stress disorder is associated with PACAP and the PAC1 receptor. Nature, 470, 492–497. [PubMed: 21350482]
- Roberto M, & Brunelli M (2000). PACAP-38 enhances excitatory synaptic transmission in the rat hippocampal CA1 region. Learn Mem, 7, 303–311. [PubMed: 11040262]
- Roberto M, Scuri R, & Brunelli M (2001). Differential effects of PACAP-38 on synaptic responses in rat hippocampal CA1 region. Learn Mem, 8, 265–271. [PubMed: 11584073]
- Roman CW, Lezak KR, Hartsock MJ, Falls WA, Braas KM, Howard AB, Hammack SE, & May V (2014). PAC1 receptor antagonism in the bed nucleus of the stria terminalis (BNST) attenuates the endocrine and behavioral consequences of chronic stress. Psychoneuroendocrinology, 47, 151–165. [PubMed: 25001965]
- Ross RA, Hoeppner SS, Hellberg SN, O'Day EB, Rosencrans PL, Ressler KJ, May V, & Simon NM (2020). Circulating PACAP peptide and PAC1R genotype as possible transdiagnostic biomarkers for anxiety disorders in women: a preliminary study. Neuropsychopharmacology, 45, 1125–1133. [PubMed: 31910434]
- Rozeske RR, Valerio S, Chaudun F, & Herry C (2015). Prefrontal neuronal circuits of contextual fear conditioning. Genes Brain Behav, 14, 22–36. [PubMed: 25287656]
- Rudecki AP, & Gray SL (2016). PACAP in the Defense of Energy Homeostasis. Trends Endocrinol Metab, 27, 620–632. [PubMed: 27166671]
- Sacchetti B, Lorenzini CA, Baldi E, Bucherelli C, Roberto M, Tassoni G, & Brunelli M (2001). Pituitary adenylate cyclase-activating polypeptide hormone (PACAP) at very low dosages improves memory in the rat. Neurobiol Learn Mem, 76, 1–6. [PubMed: 11525248]
- Sadanandan N, Cozene B, Park YJ, Farooq J, Kingsbury C, Wang ZJ, Moscatello A, Saft M, Cho J, Gonzales-Portillo B, & Borlongan CV (2021). Pituitary Adenylate Cyclase-Activating Polypeptide: A Potent Therapeutic Agent in Oxidative Stress. Antioxidants (Basel), 10.
- Sandvik AK, Cui G, Bakke I, Munkvold B, & Waldum HL (2001). PACAP stimulates gastric acid secretion in the rat by inducing histamine release. Am J Physiol Gastrointest Liver Physiol, 281, G997g1003. [PubMed: 11557520]
- Sauvage M, Brabet P, Holsboer F, Bockaert J, & Steckler T (2000). Mild deficits in mice lacking pituitary adenylate cyclase-activating polypeptide receptor type 1 (PAC1) performing on memory tasks. Brain research. Molecular brain research, 84.

- Scaldaferri ML, Modesti A, Palumbo C, Ulisse S, Fabbri A, Piccione E, Frajese G, & Moretti C (2000). Pituitary adenylate cyclase-activating polypeptide (PACAP) and PACAP-receptor type 1 expression in rat and human placenta. Endocrinology, 141, 1158–1167. [PubMed: 10698193]
- Schmidt SD, Myskiw JC, Furini CR, Schmidt BE, Cavalcante LE, & Izquierdo I (2015). PACAP modulates the consolidation and extinction of the contextual fear conditioning through NMDA receptors. Neurobiol Learn Mem, 118c, 120–124.
- Seiglie MP, Lepeak L, Velázquez-Sanchez C, Ferragud A, Le T, Cottone P, & Sabino V (2022). The Pituitary Adenylate Cyclase-Activating Polypeptide (PACAP) System of the Central Amygdala Mediates the Detrimental Effects of Chronic Social Defeat Stress in Rats. eNeuro, 9.
- Sekar R, Wang L, & Chow BK (2017). Central Control of Feeding Behavior by the Secretin, PACAP, and Glucagon Family of Peptides. Front Endocrinol (Lausanne), 8, 18. [PubMed: 28223965]
- Sherwood NM, Krueckl SL, & McRory JE (2000). The origin and function of the pituitary adenylate cyclase-activating polypeptide (PACAP)/glucagon superfamily. Endocr Rev, 21, 619– 670. [PubMed: 11133067]
- Shintani N, Mori W, Hashimoto H, Imai M, Tanaka K, Tomimoto S, Hirose M, Kawaguchi C, & Baba A (2002). Defects in reproductive functions in PACAP-deficient female mice. Regul Pept, 109, 45–48. [PubMed: 12409213]
- Shintani Y, Hayata-Takano A, Moriguchi K, Nakazawa T, Ago Y, Kasai A, Seiriki K, Shintani N, Hashimoto H (2018). β-arrestin1 and 2 differentially regulate PACAP-induced PAC1 receptor signaling and trafficking. PLoS One, 13, e0196946. [PubMed: 29734363]
- Shioda S, Shuto Y, Somogyvari-Vigh A, Legradi G, Onda H, Coy DH, Nakajo S, & Arimura A (1997). Localization and gene expression of the receptor for pituitary adenylate cyclase-activating polypeptide in the rat brain. Neurosci Res, 28, 345–354. [PubMed: 9274830]
- Skoglösa Y, Takei N, & Lindholm D (1999). Distribution of pituitary adenylate cyclase activating polypeptide mRNA in the developing rat brain. Brain Res Mol Brain Res, 65, 1–13. [PubMed: 10036302]
- Steenstrup BR, Jørgensen JC, Alm P, Hannibal J, Junge J, Fahrenkrug J, & Ottesen B (1996). Pituitary adenylate cyclase activating polypeptide (PACAP): occurrence and vasodilatory effect in the human uteroplacental unit. Regul Pept, 61, 197–204. [PubMed: 8701036]
- Stevens JS, Almli LM, Fani N, Gutman DA, Bradley B, Norrholm SD, Reiser E, Ely TD, Dhanani R, Glover EM, Jovanovic T, & Ressler KJ (2014). PACAP receptor gene polymorphism impacts fear responses in the amygdala and hippocampus. Proc Natl Acad Sci U S A, 111, 3158–3163. [PubMed: 24516127]
- Stroth N, & Eiden LE (2010). Stress hormone synthesis in mouse hypothalamus and adrenal gland triggered by restraint is dependent on pituitary adenylate cyclase-activating polypeptide signaling. Neuroscience, 165, 1025–1030. [PubMed: 19931358]
- Stroth N, Holighaus Y, Ait-Ali D, & Eiden LE (2011a). PACAP: a master regulator of neuroendocrine stress circuits and the cellular stress response. Ann N Y Acad Sci, 1220, 49–59. [PubMed: 21388403]
- Stroth N, Liu Y, Aguilera G, & Eiden LE (2011b). Pituitary adenylate cyclase-activating polypeptide controls stimulus-transcription coupling in the hypothalamic-pituitary-adrenal axis to mediate sustained hormone secretion during stress. J Neuroendocrinol, 23, 944–955. [PubMed: 21824204]
- Takasaki I, Ogashi H, Okada T, Shimodaira A, Hayakawa D, Watanabe A, Miyata A, Kurihara T, Gouda H, & Toyooka N (2020). Synthesis of a novel and potent small-molecule antagonist of PAC1 receptor for the treatment of neuropathic pain. Eur J Med Chem, 186, 111902. [PubMed: 31771828]
- Takasaki I, Watanabe A, Yokai M, Watanabe Y, Hayakawa D, Nagashima R, Fukuchi M, Okada T, Toyooka N, Miyata A, Gouda H, & Kurihara T (2018). In Silico Screening Identified Novel Small-molecule Antagonists of PAC1 Receptor. J Pharmacol Exp Ther, 365, 1–8. [PubMed: 29363578]
- Takuma K, Maeda Y, Ago Y, Ishihama T, Takemoto K, Nakagawa A, Shintani N, Hashimoto H, Baba A, & Matsuda T (2014). An enriched environment ameliorates memory impairments in PACAP-deficient mice. Behav Brain Res, 272, 269–278. [PubMed: 25014004]

- Tanida M, Shintani N, Morita Y, Tsukiyama N, Hatanaka M, Hashimoto H, Sawai H, Baba A, & Nagai K (2010). Regulation of autonomic nerve activities by central pituitary adenylate cyclaseactivating polypeptide. Regul Pept, 161, 73–80. [PubMed: 20171991]
- Tomimoto S, Ojika T, Shintani N, Hashimoto H, Hamagami K, Ikeda K, Nakata M, Yada T, Sakurai Y, Shimada T, Morita Y, Ishida C, & Baba A (2008). Markedly reduced white adipose tissue and increased insulin sensitivity in adcyap1-deficient mice. J Pharmacol Sci, 107, 41–48. [PubMed: 18446003]
- Tönshoff C, Hemmick L, & Evinger MJ (1997). Pituitary adenylate cyclase activating polypeptide (PACAP) regulates expression of catecholamine biosynthetic enzyme genes in bovine adrenal chromaffin cells. J Mol Neurosci, 9, 127–140. [PubMed: 9407393]
- Tseng A, Singh P, Marquez P, Hamid A, & Lutfy K (2019). The role of endogenous pituitary adenylyl cyclase activating polypeptide (PACAP) in nicotine self-administration, reward and aversion. Pharmacol Biochem Behav, 181, 46–52. [PubMed: 31028757]
- Tsukiyama N, Saida Y, Kakuda M, Shintani N, Hayata A, Morita Y, Tanida M, Tajiri M, Hazama K, Ogata K, Hashimoto H, & Baba A (2011). PACAP centrally mediates emotional stress-induced corticosterone responses in mice. Stress, 14, 368–375. [PubMed: 21438773]
- Twining RC, Lepak K, Kirry AJ, & Gilmartin MR (2020). Ventral Hippocampal Input to the Prelimbic Cortex Dissociates the Context From the Cue Association in Trace Fear Memory. J Neurosci.
- Uddin M, Chang SC, Zhang C, Ressler K, Mercer KB, Galea S, Keyes KM, McLaughlin KA, Wildman DE, Aiello AE, & Koenen KC (2013). Adcyap1r1 genotype, posttraumatic stress disorder, and depression among women exposed to childhood maltreatment. Depress Anxiety, 30, 251–258. [PubMed: 23280952]
- Vaccari S, Latini S, Barberi M, Teti A, Stefanini M, & Canipari R (2006). Characterization and expression of different pituitary adenylate cyclase-activating polypeptide/vasoactive intestinal polypeptide receptors in rat ovarian follicles. J Endocrinol, 191, 287–299. [PubMed: 17065411]
- Van Rampelbergh J, Poloczek P, Françoys I, Delporte C, Winand J, Robberecht P, & Waelbroeck M (1997). The pituitary adenylate cyclase activating polypeptide (PACAP I) and VIP (PACAP II VIP1) receptors stimulate inositol phosphate synthesis in transfected CHO cells through interaction with different G proteins. Biochim Biophys Acta, 1357, 249–255. [PubMed: 9223629]
- Varodayan FP, Minnig MA, Steinman MQ, Oleata CS, Riley MW, Sabino V, & Roberto M (2020). PACAP regulation of central amygdala GABAergic synapses is altered by restraint stress. Neuropharmacology, 168, 107752. [PubMed: 31476352]
- Vaudry D, Falluel-Morel A, Bourgault S, Basille M, Burel D, Wurtz O, Fournier A, Chow BK, Hashimoto H, Galas L, & Vaudry H (2009). Pituitary adenylate cyclase-activating polypeptide and its receptors: 20 years after the discovery. Pharmacol Rev, 61, 283–357. [PubMed: 19805477]
- Vécsei L, Tuka B, & Tajti J (2014). Role of PACAP in migraine headaches. Brain, 137, 650–651. [PubMed: 24549810]
- Velasco ER, Florido A, Flores Á, Senabre E, Gomez-Gomez A, Torres A, Roca A, Norrholm S, Newman EL, Das P, Ross RA, Lori A, Pozo OJ, Ressler KJ, Garcia-Esteve LL, Jovanovic T, & Andero R (2022). PACAP-PAC1R modulates fear extinction via the ventromedial hypothalamus. Nat Commun, 13, 4374. [PubMed: 35902577]
- Vertongen P, Schiffmann SN, Gourlet P, & Robberecht P (1997). Autoradiographic visualization of the receptor subclasses for vasoactive intestinal polypeptide (VIP) in rat brain. Peptides, 18, 1547–1554. [PubMed: 9437715]
- Vollesen LH, Guo S, Andersen MR, & Ashina M (2019). Effect of the H(1)-antihistamine clemastine on PACAP38 induced migraine. Cephalalgia, 39, 597–607. [PubMed: 30165750]
- Vu JP, Goyal D, Luong L, Oh S, Sandhu R, Norris J, Parsons W, Pisegna JR, & Germano PM (2015). PACAP intraperitoneal treatment suppresses appetite and food intake via PAC1 receptor in mice by inhibiting ghrelin and increasing GLP-1 and leptin. Am J Physiol Gastrointest Liver Physiol, 309, G816–825. [PubMed: 26336928]

- Wang J, Song X, Zhang D, Chen X, Li X, Sun Y, Li C, Song Y, Ding Y, Ren R, Harrington EH, Hu LA, Zhong W, Xu C, Huang X, Wang HW, & Ma Y (2020). Cryo-EM structures of PAC1 receptor reveal ligand binding mechanism. Cell Res, 30, 436–445. [PubMed: 32047270]
- Wang L, Cao C, Wang R, Qing Y, Zhang J, & Zhang XY (2013). PAC1 receptor (ADCYAP1R1) genotype is associated with PTSD's emotional numbing symptoms in Chinese earthquake survivors. J Affect Disord, 150, 156–159. [PubMed: 23394710]
- Wienholtz NKF, Christensen CE, Zhang DG, Coskun H, Ghanizada H, Al-Karagholi MA, Hannibal J, Egeberg A, Thyssen JP, & Ashina M (2021). Early treatment with sumatriptan prevents PACAP38-induced migraine: A randomised clinical trial. Cephalalgia, 41, 731–748. [PubMed: 33567890]
- Xu Y, Feng W, Zhou Q, Liang A, Li J, Dai A, Zhao F, Yan J, Chen CW, Li H, Zhao LH, Xia T, Jiang Y, Xu HE, Yang D, & Wang MW (2022). A distinctive ligand recognition mechanism by the human vasoactive intestinal polypeptide receptor 2. Nat Commun, 13, 2272. [PubMed: 35477937]
- Yada T, Sakurada M, Filipsson K, Kikuchi M, & Ahrén B (2000). Intraperitoneal PACAP administration decreases blood glucose in GK rats, and in normal and high fat diet mice. Ann N Y Acad Sci, 921, 259–263. [PubMed: 11193831]
- Yaka R, He DY, Phamluong K, & Ron D (2003). Pituitary adenylate cyclase-activating polypeptide (PACAP(1–38)) enhances N-methyl-D-aspartate receptor function and brain-derived neurotrophic factor expression via RACK1. J Biol Chem, 278, 9630–9638. [PubMed: 12524444]
- Yon L, Alexandre D, Montéro M, Chartrel N, Jeandel L, Vallarino M, Conlon JM, Kikuyama S, Fournier A, Gracia-Navarro F, Roubos E, Chow B, Arimura A, Anouar Y, & Vaudry H (2001). Pituitary adenylate cyclase-activating polypeptide and its receptors in amphibians. Microsc Res Tech, 54, 137–157. [PubMed: 11458398]
- Zhang L, Hernandez VS, Gerfen CR, Jiang SZ, Zavala L, Barrio RA, & Eiden LE (2021). Behavioral role of PACAP signaling reflects its selective distribution in glutamatergic and GABAergic neuronal subpopulations. Elife, 10.
- Zhou CJ, Kikuyama S, Shibanuma M, Hirabayashi T, Nakajo S, Arimura A, & Shioda S (2000). Cellular distribution of the splice variants of the receptor for pituitary adenylate cyclaseactivating polypeptide (PAC(1)-R) in the rat brain by in situ RT-PCR. Brain Res Mol Brain Res, 75, 150–158. [PubMed: 10648899]

# **Highlights**

**•** The neuropeptide PACAP regulates many biological and behavioral processes

- **•** Targeting PACAP could innovate treatment for a variety of disorders
- **•** Heterogeneity in PACAP signaling poses challenges for therapeutic development
- **•** Nonetheless, harnessing the PACAP system may be a powerful strategy against disease

#### **Table 1.**

Clinical relevance of PACAP/PAC1 in disease.



