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Brain $\alpha 2\delta$ -1–Bound NMDA Receptors Drive Calcineurin Inhibitor–Induced Hypertension

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Abstract

Background: Calcineurin is highly enriched in immune T cells and in the nervous system. Calcineurin inhibitors, including cyclosporine and tacrolimus (FK506), are the cornerstone of immunosuppressive regimens for preserving transplanted organs and tissues. However, these drugs often cause persistent hypertension owing to excess sympathetic outflow, which is maintained by N-methyl-D-aspartate receptor (NMDAR)-mediated excitatory input to the hypothalamic paraventricular nucleus (PVN). It is unclear how calcineurin inhibitors increase NMDAR activity in the PVN to augment sympathetic vasomotor activity. α28–1 (encoded by the *Cacna2d1* gene), known colloquially as a calcium channel subunit, is a newly discovered NMDAR-interacting protein. Here, we determined whether α28–1 plays a role in calcineurin inhibitor–induced synaptic NMDAR hyperactivity in the PVN and hypertension development.

Methods and Results: Immunoblotting and coimmunoprecipitation assays revealed that prolonged treatment with FK506 in rats significantly increased protein levels of α 28–1, GluN1 (the obligatory NMDAR subunit), and the α 28–1–GluN1 complex in PVN synaptosomes. These effects were blocked by inhibiting α 28–1 with gabapentin or interrupting the α 28–1–NMDAR interaction with an α 28–1 C-terminus peptide. Whole-cell recordings in brain slices showed that treatment with FK506 potentiated the activity of presynaptic and postsynaptic NMDARs in spinally projecting PVN neurons; such effects were abolished by gabapentin, *Cacna2d1* knockout, or α 28–1 C-terminus peptide. Furthermore, microinjection of α 28–1 C-terminus peptide into the PVN diminished renal sympathetic nerve discharges and arterial blood pressure that had been increased by FK506 treatment. Remarkably, telemetry recording showed that concurrent administration of gabapentin prevented the development of FK506-induced hypertension in rats.

Disclosures

None.

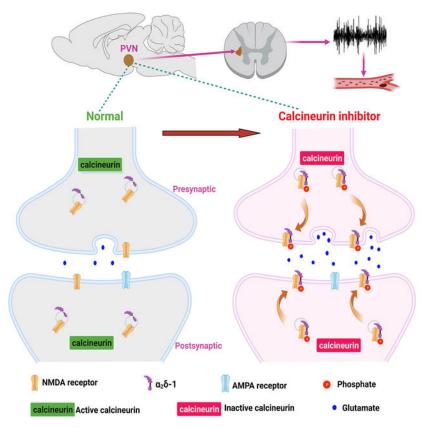
Supplemental Materials Expanded Materials & Methods Online Figures S1–S7 Online Tables S1–S10

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Additionally, FK506 treatment induced sustained hypertension in wild-type mice but not in *Cacna2d1* knockout mice.

Conclusions: These findings indicate that $\alpha 2\delta - 1$ is essential for calcineurin inhibitor–induced increases in synaptic NMDAR activity in PVN presympathetic neurons and sympathetic outflow. Thus, $\alpha 2\delta - 1$ and $\alpha 2\delta - 1$ -bound NMDARs represent new targets for treating calcineurin inhibitor–induced hypertension.

Graphical Abstract



Keywords

autonomic nervous system; immunosuppressant; gabapentinoid; protein phosphatase; synaptic plasticity; sympathetic nervous system

Subject Terms:

Animal Models of Human Disease; Basic Science Research; Hypertension; Physiology; Translational Studies

Introduction

Drug-induced hypertension can occur as a result of either the unintended consequences of a drug or its antagonistic effect on antihypertensive medications. Calcineurin inhibitors,

such as cyclosporine and tacrolimus (FK506), are standard immunosuppressants used for minimizing rejection of transplanted organs and cells/tissues (i.e., bone marrow and hematopoietic stem cells) and for treating autoimmune diseases. However, long-term use of these drugs often causes uncontrolled, persistent hypertension. ¹⁻⁴ Sympathetic nerve discharges are substantially increased in calcineurin inhibitor–induced hypertension (CIH) in patients and in animal models. ^{1,5-7} Widespread vasoconstriction caused by augmented sympathetic nerve activity impairs blood flow and perfusion to transplanted organs and tissues, which can severely limit the viability of grafts and increase recipient mortality. ^{2-4,8} CIH is blunted by ganglionic blockade, ^{7,9,10} a1-adrenergic receptor antagonists, ^{9,11} surgical adrenalectomy, ⁷ or chemical sympathectomy. ¹¹ Although a causal role for the sympathetic nervous system in the pathogenesis of CIH has been documented, it remains uncertain how excess central sympathetic outflow is generated at the molecular and cellular levels in CIH.

Calcineurin, also known as protein phosphatase-2B, is expressed abundantly not only in immune T cells but also in many brain regions, including the hypothalamic paraventricular nucleus (PVN), which plays a key role in generating elevated sympathetic output in hypertensive conditions. 12-14 The presympathetic neurons in the PVN regulate vasomotor tone mainly via their projection to the rostral ventrolateral medulla and intermediolateral cell column in the spinal cord. 15,16 Calcineurin is a Ca²⁺/calmodulin-dependent serinethreonine phosphatase that actively controls the phosphorylation status of N-methyl-Daspartate receptors (NMDARs). 5,17–19 Normal calcineurin activity in the brain is important for maintaining physiological neuronal activity and synaptic NMDAR plasticity. 17,20,21 Systemic treatment with FK506 diminishes calcineurin activity in the PVN, which augments phosphorylation and activity of synaptic NMDARs in the PVN, leading to increased glutamatergic excitatory synaptic input and firing activity of presympathetic neurons and sympathetic outflow. ⁵ Correspondingly, NMDAR antagonists are effective in reducing CIH in the animal model.⁵ However, it is unclear how synaptic NMDAR activity in the PVN is augmented in CIH. Furthermore, NMDARs in the central nervous system are involved in many physiological functions, including learning and memory.²² Because nonselectively blocking all NMDARs is associated with some serious adverse effects, such as dizziness, confusion, ataxia, and hallucination, ^{23,24} long-term use of NMDAR antagonists is challenging. Therefore, there is a pressing need to identify a better therapeutic target for managing CIH.

 $\alpha 2\delta - 1$, encoded by the *Cacna2d1* gene, previously known as a subunit of voltage-gated calcium channels (VGCCs), is a newly discovered regulatory protein that preferentially interacts with phosphorylated NMDARs to promote synaptic trafficking of NMDARs in the spinal cord. ^{25,26} In fact, NMDAR phosphorylation alone does not increase synaptic trafficking and activity of NMDARs unless $\alpha 2\delta - 1$ is present. ²⁶ Given the critical role of $\alpha 2\delta - 1$ in regulating NMDARs, we tested the hypothesis that calcineurin inhibition promotes $\alpha 2\delta - 1$ -NMDAR interactions and their synaptic trafficking and activity in the PVN, thereby augmenting sympathetic outflow. Our study identifies for the first time the indispensable role of the $\alpha 2\delta - 1$ protein in calcineurin inhibitor–augmented synaptic NMDAR activity in the PVN and sympathetic vasomotor activity. Furthermore, our findings suggest that $\alpha 2\delta - 1$ or $\alpha 2\delta - 1$ -bound NMDARs could be targeted for the prevention and treatment of CIH.

Materials and Methods

Data Availability

The data that support the findings of this study are available from the corresponding author upon reasonable request. Detailed methods can be found in the Supplemental Material.

Animal Models

All experiments were approved by Institutional Animal Care and Use Committee of MD Anderson Cancer Center. Sprague-Dawley rats were purchased from Envigo. *Cacna2d1* knockout (KO, *Cacna2d1*^{-/-}) mice and wild-type (WT, *Cacna2d1*^{+/+}) littermates were obtained by breeding *Cacna2d1*^{+/-} mice.²⁵ For induction of CIH, FK506 was injected intraperitoneally at a dose of 3 mg/kg once daily for 14 consecutive days in rats and mice.⁵ Final experiments were conducted 5–7 days after the last FK506 injection. Animals received a same amount of dimethyl sulfoxide as the vehicle group.

PVN Synaptosome Preparation, Coimmunoprecipitation, and Immunoblotting

Synaptosomes from PVN tissues were prepared, and coimmunoprecipitation and immunoblotting were performed as described previously 13,27 . The protein samples were incubated with Protein G bead prebound to a rabbit anti-GluN1 antibody. Protein samples on the beads were separated by electrophoresis and then transferred to a polyvinylidene difluoride membrane. The membrane was incubated with a mouse anti- $\alpha 2\delta -1$ antibody, a mouse anti-PSD-95 antibody, or a mouse anti-GluN1 antibody.

Arterial Blood Pressure Measurement with Telemetry

Arterial blood pressure (ABP) in rats and mice was measured using a Millar telemetry system and a DSI telemetry system, respectively.⁵ Heart rate (HR) values were derived from ABP pulse signals.

Electrophysiological Recording in Brain Slices

Spinally projecting PVN neurons were retrogradely labeled and recorded as described previously. $^{34-36}$ Whole-cell configurations were used to record miniature excitatory postsynaptic currents (mEPSCs) and puff NMDAR currents in labeled PVN neurons. The mEPSCs were recorded in the presence of 1 μ M tetrodotoxin and 20 μ M bicuculline at a holding potential of -60 mV. To record postsynaptic NMDAR activity, we puffed NMDA (100 μ M) directly to labeled PVN neurons at a holding potential of -60 mV in the presence of tetrodotoxin. 5,36

Recording of Renal Sympathetic Nerve Activity and PVN Microinjection

Renal sympathetic nerve activity (RSNA) was recorded as descried previously.^{5,27} Rats were anesthetized with intraperitoneal injection of a mixture of α-chloralose (60–75 mg/kg) and urethane (800 mg/kg), and RSNA was recorded from a branch of the left renal nerve. PVN microinjections were performed as reported previously.^{12,29} A glass pipette was advanced into the PVN, and the drug solution was pressure-ejected via a calibrated microinjector.

Statistical Analysis

All data are expressed as mean \pm SEM. Student t test was used to determine difference between 2 groups, and one-way or two-way ANOVA followed by Dunnett's or Bonferroni's *post hoc* test was used to determine differences among 3 or more groups.

Results

Calcineurin inhibition promotes the $\alpha 28-1-NMDAR$ interaction and synaptic trafficking of $\alpha 28-1-bound\ NMDARs$ in the PVN

Systemic treatment with the calcineurin inhibitor FK506 diminishes calcineurin activity in the forebrain and increases NMDAR phosphorylation in the PVN. 5 $\alpha 2\delta - 1$ is a phosphobinding protein and preferentially interacts with phosphorylated NMDARs independently of VGCCs in the cell line and spinal cord. 26 We first determined whether calcineurin inhibition affects the synaptic protein levels of $\alpha 2\delta - 1$ and NMDARs in the PVN. For induction of CIH, rats were systemically treated with FK506 (3 mg/kg per day) for consecutive 14 days, which gradually and persistently increased mean arterial blood pressure (MAP) and heart rate (HR), lasting more than 7 days after FK506 treatment was discontinued. 5 Because treatment with FK506 for 14 days caused a similar degree of increases in MAP and HR in male and female rats (n = 6 rats per group; Figure S1), we mainly used male rats for the following experiments.

Treatment with FK506 increases phosphorylation of GluN1, but not GluN2A or GluN2B, in the PVN.⁵ GluN1 is the obligatory subunit of NMDARs, and other NMDAR subunits require GluN1 to function and traffic together with GluN1.²² Immunoblotting showed that treatment with FK506 markedly increased the protein levels of α 28–1 and GluN1 in PVN synaptosomes (n = 6 samples per group, each sample contained PVN tissues from 2 rats; α 28–1: P= 1.0e-04, F(2,15) = 17.36; GluN1: P= 5.0e-04, F(2,15) = 11.97; Figure 1A and 1B). However, treatment with FK506 had no significant effect on the total protein level of α 28–1 in the PVN lysate (Figure S2). The total protein levels of NMDARs in the PVN do not differ significantly between vehicle-treated and FK506-treated rats.⁵

In the hypothalamus from rats and humans, $\alpha 28-1$ physically interacts with NMDARs.³⁴ We thus determined whether treatment with FK506 alters the $\alpha 28-1$ -NMDAR interaction in the PVN. Coimmunoprecipitation analysis showed that a specific GluN1 antibody, but not the irrelevant IgG, precipitated $\alpha 28-1$ proteins in PVN synaptosomes. Treatment with FK506 substantially increased the amount of $\alpha 28-1$ -GluN1 protein complexes in PVN synaptosomes (n = 6 samples per group, each sample contained PVN tissues from 2 rats; P=2.0e-04, $F_{(2,15)}=14.92$; Figure 1A and 1B). These results suggest that calcineurin inhibition enhances the $\alpha 28-1$ -NMDAR interaction and their synaptic trafficking in the PVN.

We next determined whether $\alpha 28-1$ is required for the FK506-induced increase in the level of NMDARs in PVN synaptosomes. Gabapentin is an $\alpha 28-1$ inhibitory ligand ^{42,43} and used for treating patients with chronic neuropathic pain and epilepsy. Hypothalamic brain slices from FK506-treated rats were incubated with gabapentin (100 μ M) for 30 minutes, and synaptosomes in the PVN tissue were then isolated. Immunoblotting of PVN synaptosomes showed that gabapentin treatment fully reversed the increased protein levels

of $\alpha.28-1$, GluN1, and $\alpha.28-1$ -GluN1 complexes by FK506 treatment (n = 6 samples per group; $\alpha.28-1$: P=9.0e-04, $F_{(2,15)}=17.36$; GluN1: P=0.0055, $F_{(2,15)}=11.97$; $\alpha.28-1$ -GluN1 complex: P=0.0037, $F_{(2,15)}=14.92$; Figure 1A and 1B). By contrast, gabapentin treatment had no statistically significant effect on the protein levels of $\alpha.28-1$, GluN1, or $\alpha.28-1$ -GluN1 complexes in PVN synaptosomes obtained from vehicle-treated rats (n = 6 samples per group, each sample contained PVN tissues from 2 rats; Figure 1C and 1D). These findings suggest that $\alpha.28-1$ is essential for calcineurin inhibitor–induced increases in synaptic trafficking of NMDARs in the PVN.

α2δ–1 physically interacts with NMDARs via its C-terminus, which is an intrinsically disordered protein region. ^{25,26} A cell-penetrating, Tat-fused α2δ–1 C terminus peptide (termed the $\alpha 2\delta$ -1CT peptide) that mimics the C-terminal domain of $\alpha 2\delta$ -1 effectively disrupts the $\alpha 2\delta - 1$ -NMDAR interaction. ^{25,44} We thus used $\alpha 2\delta - 1$ CT peptide to determine whether calcineur in inhibition increases $\alpha 2\delta - 1$ -dependent NMDAR trafficking in the PVN. Hypothalamic brain slices from FK506-treated rats were first incubated with Tat-fused α2δ–1CT peptide (1 μM) or Tat-fused scrambled control peptide (1 μM) for 30 minutes before synaptosomes in the PVN tissues were isolated. Compared with the control peptide, treatment with $\alpha 2\delta - 1$ CT peptide significantly decreased protein levels of $\alpha 2\delta - 1$, GluN1, and the $\alpha 2\delta$ -1-GluN1 complex in PVN synaptosomes from FK506-treated rats (n = 6 samples per group; $\alpha 2\delta - 1$: P = 1.9e-05, $t_{(10)} = 7.57$; GluN1: P = 6.3e-05, $t_{(10)} = 6.56$; α 28–1–GluN1 complex: P = 8.8e-05, t(10) = 6.30; Figure 1E and 1F). However, treatment with a28-1CT peptide had no significant effect on the protein levels of a28-1, GluN1, or a28–1–GluN1 complexes in PVN synaptosomes from vehicle-treated rats (n = 6 samples per group, each sample contained PVN tissues from 2 rats; Figure 1G and 1H). These data support the notion that calcineurin inhibition enhances the $\alpha 2\delta - 1$ -NMDAR interaction, which promotes synaptic expression of NMDARs in the PVN.

Calcineurin inhibition potentiates synaptic NMDAR activity in PVN presympathetic neurons via $\alpha 2\delta$ -1 in rats

Tonic activation of presynaptic NMDARs increases synaptic glutamate release to PVN presympathetic neurons in CIH. Because calcineurin inhibition increased synaptic expression of NMDARs via $\alpha 2\delta - 1$, we next determined whether $\alpha 2\delta - 1$ mediates the increased presynaptic NMDAR activity in PVN presympathetic neurons in CIH. Hypothalamic brain slices were obtained from vehicle- and FK506-treated rats. To quantify NMDAR-mediated spontaneous quantal release of glutamate from presynaptic terminals, we recorded mEPSCs of retrogradely labeled, spinally projecting PVN neurons. The baseline frequency, but not the amplitude, of mEPSCs was significantly greater in FK506-treated than in control rats (n = 10 neurons per group; Figure 2A–2D). Bath application of AP5 (50 μ M), a specific NMDAR antagonist, ^{36,45} had no significant effect on mEPSCs in vehicle-treated rats but normalized the increased frequency of mEPSCs in FK506-treated rats, indicating increased glutamate release from presynaptic terminals via NMDARs by FK506 treatment (Figure 2A–2D). Incubation with gabapentin (100 μ M) for 30 minutes significantly attenuated the higher baseline frequency of mEPSCs in labeled PVN neurons in FK506-treated rats. Subsequent bath application of AP5 had no further effect on the already

decreased frequency of mEPSCs in the same neurons (n = 10 neurons per group; Figure 2C and 2D).

Postsynaptic NMDAR activity is also increased in the PVN and plays a major role in augmenting glutamatergic excitatory input and firing activity of presympathetic neurons in CIH. We thus determined whether inhibiting $\alpha 2\delta - 1$ with gabapentin affects the increased postsynaptic NMDAR activity in CIH. The amplitude of puff-elicited NMDAR currents in labeled PVN neurons was much larger in FK506-treated rats than in control rats (n = 10 neurons per group; Figure 2E). Treatment with gabapentin (100 μ M) for 30 minutes reversed the FK506 treatment–increased amplitude of puff NMDAR currents in labeled PVN neurons (Figure 2E). By contrast, gabapentin had no statistically significant effect on puff-elicited NMDAR currents in labeled PVN neurons in control rats (Figure S3). Together, these findings suggest an essential role of $\alpha 2\delta - 1$ in the increased presynaptic and postsynaptic NMDAR activity of PVN presympathetic neurons induced by the calcineurin inhibitor.

Genetic ablation of a28–1 abolishes the calcineurin inhibitor–induced potentiation of synaptic NMDAR activity in PVN presympathetic neurons in mice

Because gabapentin binds to both $\alpha 28-1$ and $\alpha 28-2$ proteins, 42,43 we then attempted to use Cacna2d1 knockout (KO) mice to validate the role of $\alpha 28-1$ in synaptic NMDAR activity increased by calcineurin inhibition. We treated WT mice and Cacna2d1 KO mice with FK506 (3 mg/kg per day) for 14 days and then examined synaptic NMDAR activity in spinally projecting PVN neurons in brain slices. The frequency and amplitude of mEPSCs and the amplitude of puff-elicited NMDAR currents in labeled PVN neurons were similar in WT and Cacna2d1 KO mice (Figure 3A–3D). As expected, treatment with FK506 profoundly increased the baseline frequency of mEPSCs in labeled PVN neurons in WT mice but not in Cacna2d1 KO mice (n = 12 neurons in both vehicle-treated WT and FK506-treated Cacna2d1 KO groups, n = 13 neurons in FK506-treated WT group; Figure 3A–3D). Subsequent bath application of AP5 (50 μ M) reversed the increased mEPSCs frequency in labeled PVN neurons in FK506-treated WT mice but had no such effect in FK506-treated Cacna2d1 KO mice (Figure 3A–3D).

In addition, FK506 treatment significantly increased the amplitude of puff-elicited NMDAR currents in labeled PVN neurons in WT mice but had no significant effect on puff-elicited NMDAR currents in labeled PVN neurons in *Cacna2d1* KO mice (n = 11 neurons per group; Figure 3E). These data provide unambiguous evidence that α 2 δ -1 is indispensable for calcineurin inhibitor–induced potentiation in presynaptic and postsynaptic NMDAR activity in PVN presympathetic neurons.

Calcineurin inhibitor potentiates glutamatergic input to PVN presympathetic neurons via $\alpha 2\delta$ -1-bound NMDARs

To determine directly whether $\alpha 28-1-$ coupled NMDARs are responsible for calcineurin inhibitor–induced glutamatergic excitatory input to PVN presympathetic neurons, we incubated hypothalamic brain slices from FK506-treated rats with 1 μM Tat-fused $\alpha 28-1CT$ peptide or 1 μM Tat-fused control peptide for 30 minutes 25,37 and then examined presynaptic and postsynaptic NMDAR activity in spinally projecting PVN neurons.

Treatment with $\alpha 2\delta$ -1CT peptide, but not the control peptide, largely attenuated the increased baseline frequency of mEPSCs in labeled PVN neurons in FK506-treated rats (n = 10 neurons per group; Figure 4A–4C). Subsequent bath application of AP5 (50 μ M) lowered the mEPSCs frequency of labeled PVN neurons treated with the control peptide (Figure 4A–4C). However, AP5 had no further effect on the mEPSCs frequency of labeled PVN neurons in brain slices treated with $\alpha 2\delta$ -1CT peptide (Figure 4A–4C).

Moreover, in labeled PVN neurons from brain slices of FK506-treated rats, the amplitude of puff-elicited NMDAR currents was much smaller in slices treated with $\alpha 2\delta - 1CT$ peptide than in those treated with the control peptide (n = 10 neurons per group; Figure 4D). Additionally, neither the control peptide nor $\alpha 2\delta - 1CT$ peptide had any statistically significant effect on the amplitude of puff-elicited NMDAR currents in labeled PVN neurons in control rats (Figure S3). These data suggest that $\alpha 2\delta - 1$ -coupled NMDARs are responsible for calcineurin inhibitor–augmented glutamatergic synaptic input to PVN presympathetic neurons.

α2δ-1-bound NMDARs in the PVN mediate calcineurin inhibitor-potentiated sympathetic vasomotor activity

NMDAR hyperactivity in the PVN plays a critical role in the increased sympathetic output in CIH.⁵ Having demonstrated the essential role of $\alpha 2\delta - 1$ in synaptic NMDAR activity of PVN presympathetic neurons potentiated by the calcineurin inhibitor in brain slices, we sought to determine whether α2δ-1-bound NMDARs in the PVN mediate calcineurin inhibitor-elevated sympathetic vasomotor activity in vivo. The absolute baseline voltage level of RSNA was significantly higher in FK506-treated rats than in vehicle-treated rats $(0.19 \pm 0.07 \,\mu\text{V} \text{ vs. } 0.09 \pm 0.05 \,\mu\text{V}, P = 0.0017, t_{(18)} = 3.68)$. In vehicle-treated rats, bilateral microinjection of the control peptide (50 pmol, 50 nL), α2δ–1CT peptide (50 pmol, 50 nL), or AP5 (1.0 nmol, 50 nL)^{13,27} into the PVN had no significant effect on RSNA, MAP, or HR (n = 9 rats, Figure S4). In FK506-treated rats, initial microinjection of the control peptide into the PVN had no significant effect on RSNA, MAP, or HR, but subsequent microinjection of AP5 significantly reduced RSNA, MAP, and HR (n = 6 rats, Figure 5A, 5C, 5E, and 5F). By contrast, microinjection of $\alpha 2\delta$ -1CT peptide alone into the PVN of FK506-treated rats markedly decreased RSNA, MAP, and HR (n = 9 rats, Figure 5B and 5D). In these rats receiving prior microinjection of α 28–1CT peptide, subsequent microinjection of AP5 into the PVN had no further effect on RSNA, MAP, or HR (Figure 5B and 5D). These results suggest that $\alpha 2\delta - 1$ -bound NMDARs in the PVN are required to support the heightened sympathetic outflow caused by the calcineurin inhibitor.

Systemic administration of gabapentin is effective for treating CIH in rats

To substantiate the clinical relevance of our findings, we next determined whether systemic treatment with gabapentin is effective against CIH. The gabapentin binding in the brain and other tissues is diminished in Cacna2d1 KO mice, 43 and the $in\ vivo$ doses of gabapentin required for effective $\alpha 2\delta - 1$ inhibition have been well documented. 25,26,34,46,47 Radiotelemetry recording showed that a single intraperitoneal injection of gabapentin (60 or 100 mg/kg), which is effective in reducing pain hypersensitivity in rodent models, 46,48 had no significant effect on MAP or HR in vehicle-treated male rats (n = 6 rats, Figure 6A and

6B). By contrast, a single intraperitoneal injection of gabapentin (60 mg/kg) rapidly reduced MAP and HR in FK506-treated male rats (n = 6 rats; Figure 6A and 6B). The inhibitory effect occurred within 30 minutes after gabapentin injection and lasted about 150 minutes. Intraperitoneal injection of 100 mg/kg similarly attenuated MAP and HR in FK506-treated male rats (n = 6 rats, Figure 6A and 6B). Similarly, intraperitoneal injection of 60 mg/kg or 100 mg/kg gabapentin rapidly reduced MAP and HR in FK506-treated female rats but had no such effect in vehicle-treated female rats (n = 6 rats per group, Figure S5).

Gabapentinoids (i.e., gabapentin and pregabalin) are orally active drugs for treating patients with epilepsy and neuropathic pain. We next determined whether concurrent treatment with gabapentin and FK506 is effective in attenuating the development of CIH. Male rats were given gabapentin, at a dose of 60 mg/kg/day⁴⁹, in the drinking water during daily systemic injection of FK506 (3 mg/kg/day) for 14 days. Radiotelemetry recording showed that, compared with vehicle treatment, concurrent treatment with gabapentin diminished the increase in MAP and HR in both light and dark cycles in FK506-treated rats (n = 6 rats per group, Figure 6C and 6D).

We also performed power spectrum analysis of systolic ABP variability to determine whether it could be a useful index of sympathetic outflow in conscious animals. Prolonged treatment with FK506 significantly increased the low-frequency power in light and dark cycles when hypertension was developed (n = 6 rats per group, Figure 6E and 6F). Remarkably, concurrent treatment with gabapentin reversed the increased low-frequency power in both light and dark cycles caused by FK506 treatment (Figure 6E and 6F). Treatment with FK506 alone or with gabapentin did not significantly change the high-frequency power of systolic ABP variability. Together, these data provide strong evidence that inhibiting α 28–1 with gabapentinoids is highly effective in treating CIH by attenuating elevated sympathetic output.

a28-1 is integral to the development of CIH in mice

In addition, we took the advantage of available *Cacna2d1* KO mice to ascertain the role of α2δ–1 in the development of CIH. After implanting radiotelemetry, WT and *Cacna2d1* KO mice were injected intraperitoneally with FK506 (3 mg/kg/day) for consecutive 14 days. The baseline MAP and HR in light and dark cycles were similar between WT and *Cacna2d1* KO mice before FK506 treatment. Similar to rats, treatment with FK506 gradually and profoundly increased MAP and HR in light and dark cycles in WT mice only (Figure 7A and 7B). The increase in MAP and HR persisted at least 7 days after discontinuation of FK506 treatment in WT mice. Strikingly, prolonged treatment with FK506 failed to significantly increase MAP or HR in light and dark cycles in *Cacna2d1* KO mice (Figure 7A and 7B). As expected, treatment with vehicle for 14 days had no statistically significant effect on MAP or HR in WT and *Cacna2d1* KO mice (n = 6 mice per group; Figure S6).

The low- and high-frequency power of systolic ABP variability did not differ significantly between WT and *Cacna2d1* KO mice at the baseline before FK506 treatment (n = 6 mice per group). Prolonged treatment with FK506 in WT mice significantly increased the low-frequency power in light and dark cycles when hypertension developed (Figure 7C and 7D). By contrast, treatment with FK506 had no such effects in *Cacna2d1* KO mice. FK506

treatment did not significantly change high-frequency power of systolic ABP variability in either WT or *Cacna2d1* KO mice (Figure 7C and 7D). These findings indicate that $\alpha 2\delta - 1$ is indispensable for the development of CIH and is a promising therapeutic target for CIH.

Discussion

Our study reveals that calcineur in inhibition increases NMDAR interactions with $\alpha 2\delta - 1$ and their synaptic trafficking in the PVN, extending our recent findings that the "ontarget" effect of calcineurin inhibitors in the brain is the major cause of CIH. Calcineurin inhibitors have revolutionized transplant medicine by significantly prolonging graft survival and minimizing rates of acute rejection. However, persistent hypertension is a major adverse effect associated with long-term use of calcineurin inhibitors. Both cyclosporine and FK506 can readily cross the blood-brain barrier and impair normal calcineurin activity in the brain. 50,51 Systemically administered FK506 profoundly inhibits calcineurin activity in the forebrain but not brainstem.⁵ NMDARs are highly mobile at synapses, and their synaptic levels are critically controlled by trafficking from the intracellular pool.⁵² NMDAR phosphorylation in the hypothalamus is balanced by relative activities of protein phosphatases and kinases, including PKC, casein kinase II, and calcineurin. 5,29,45 Inhibition of the phosphatase activity by calcineurin inhibitors increases phosphorylation and synaptic expression levels of NMDARs in the PVN.⁵ $\alpha 2\delta - 1$ is a highly glycosylated protein that promotes synaptic and surface expression of its interacting proteins.⁵³ α2δ–1 preferentially interacts with phosphorylated NMDARs to promote NMDAR surface trafficking in the cell line and spinal cord independently of VGCCs. ²⁶ Because α2δ–1 predominantly binds to phosphorylated NMDARs, it primarily mediates neuronal activity-dependent NMDAR hyperactivity. In this study, we demonstrated that inhibiting $\alpha 2\delta - 1$ with gabapentin or disrupting the $\alpha 2\delta - 1$ -NMDAR interaction with $\alpha 2\delta - 1$ CT peptide normalized synaptic levels of NMDAR in the PVN that had been increased by calcineurin inhibition, suggesting that α2δ-1 is required for NMDAR synaptic trafficking in the PVN associated with CIH. It is likely that calcineurin inhibition potentiates NMDAR phosphorylation and subsequently increases the α26–1–NMDAR interaction, augmenting NMDAR trafficking at the synapses in the PVN.

Another new finding of our study is that $\alpha 2\delta - 1$ is integral to the potentiation of synaptic NMDAR activity in PVN presympathetic neurons in CIH. Normal endogenous calcineurin activity in the nervous system constitutively restricts synaptic activity of NMDARs by limiting their phosphorylation levels. ^{17,28} We showed recently that calcineurin inhibition augments the activity of NMDAR at presynaptic and postsynaptic sites in PVN presympathetic neurons. ⁵ In this study, by recording AP5-sensitive presynaptic release of glutamate, we showed that presynaptic NMDARs in the PVN are functionally quiescent under normal conditions, which is similar to findings in other brain regions such as the nucleus accumbens ³⁷ and striatum. ⁵⁴ We found that treatment with gabapentin, $\alpha 2\delta - 1CT$ peptide, or genetic *Cacna2d1* KO fully reversed the calcineurin inhibitor–potentiated presynaptic and postsynaptic NMDAR activity in spinally projecting PVN neurons. Previous work indicates that FK506 treatment has no effect on calcineurin activity and NMDAR phosphorylation in the rostral ventrolateral medulla (RVLM) ⁵ and that RVLM projecting and spinally projecting neurons in the PVN have the same functional properties. ^{14,27,36,55,56}

Also, RVLM projecting PVN neurons ultimately synapse with spinal cord neurons to control sympathetic outflow. $^{14-16}$ Because FK506 treatment similarly increased NMDAR activity in RVLM projecting and spinally projecting neurons in the PVN, 5 α 28–1 likely has a comparable role in calcineurin inhibitor–induced NMDAR hyperactivity in both RVLM projecting and spinally projecting neurons in the PVN.

Although $\alpha 2\delta - 1$ has long been considered a VGCC subunit, blocking $\alpha 2\delta - 1$ by gabapentin or *Cacna2d1* KO has little effect on overall VGCC activity^{25,57,58} or VGCC-mediated neurotransmitter release at presynaptic terminals.^{59,60} $\alpha 2\delta - 1$ predominantly interacts with NMDARs via its C-terminal domain,²⁵ whereas $\alpha 2\delta - 1$ associates with VGCCs via the von Willebrand factor type A domain near the N terminus.⁶¹ We showed that $\alpha 2\delta - 1$ CT peptide fully reversed FK506-induced synaptic NMDAR hyperactivity in presympathetic PVN neurons. Thus, it is unlikely that VGCCs are involved in glutamatergic synaptic plasticity in the PVN in CIH. Because $\alpha 2\delta - 1$ is extensively expressed in the central nervous system,^{62,63} calcineurin inhibitors may likewise increase the $\alpha 2\delta - 1$ -NMDAR interaction to augment NMDAR activity in other forebrain regions, such as circumventricular organs,^{64,65} which could also contribute to augmented sympathetic outflow in CIH.

We also provide new *in vivo* evidence that $\alpha 2\delta - 1$ —coupled NMDAR in the PVN is the key substrate required to maintain elevated sympathetic vasomotor activity caused by FK506 treatment. The PVN is the interface between the nervous and endocrine systems and plays a crucial role in coordinating sympathetic output.¹⁴ NMDAR-driven glutamatergic input in the PVN contributes predominantly to increased sympathetic outflow in the rat model of CIH.⁵ In this study, we revealed that disrupting the $\alpha 2\delta - 1$ -NMDAR interaction with $\alpha 2\delta - 1$ CT peptide in the PVN markedly reduced the elevated renal sympathetic nerve discharges and ABP by FK506 treatment. By contrast, microinjection of $\alpha 2\delta - 1$ CT peptide into the PVN had no effect on the baseline RSNA or ABP in vehicle-treated rats. Together, our findings suggest that diminished phosphatase activity by calcineurin inhibitors increases NMDAR phosphorylation in the PVN, which leads to enhanced physical interaction of phosphorylated NMDARs with $\alpha 2\delta - 1$. This increased $\alpha 2\delta - 1$ and NMDAR association can subsequently augment synaptic trafficking and activity of NMDARs, thereby augmenting excitatory glutamatergic input to PVN presympathetic neurons and sympathetic vasomotor activity (Figure S7).

Our findings have clear clinical implications, because targeting $\alpha 2\delta - 1$ with gabapentinoids or $\alpha 2\delta - 1$ -bound NMDARs with $\alpha 2\delta - 1$ CT peptides could represent a better option for treating CIH. We showed in this study that systemic administration of gabapentin rapidly attenuated ABP and HR elevated in the animal model of CIH but had no such effect in vehicle-treated rats. Strikingly, concurrent treatment with gabapentin in rats or $\alpha 2\delta - 1$ genetic KO in mice prevented the development of hypertension caused by prolonged FK506 treatment, indicating the functional significance of $\alpha 2\delta - 1$ proteins in maintaining high sympathetic vasomotor tone in CIH. Compared with clinically used NMDAR antagonists, such as ketamine and memantine, gabapentin and pregabalin have far fewer adverse effects because they target mainly $\alpha 2\delta - 1$ -bound NMDARs but do not affect basal, $\alpha 2\delta - 1$ -free NMDARs. 25,54

Some sex differences in ABP regulation and brain NMDAR activity have been reported. 66,67 However, we found that FK506 treatment caused a similar increase in ABP and HR in male and female rats. Furthermore, gabapentin equally reduced ABP in FK506-treated male and female rats, suggesting that α26-1 likely has a similar role in CIH in both sexes. Previous studies on the mechanisms of CIH have focused largely on the kidney and peripheral blood vessels. 6,68,69 The reported effects of calcineurin inhibitors on the kidney and blood vessels may be secondary to increased sympathetic vasomotor tone in CIH. In this regard, calcineurin inhibitor-induced excess sympathetic outflow and augmented renal sympathetic nerve activity could increase renin secretion and activation of the renin-angiotensin system, which can cause vasoconstriction and sodium retention. ^{70,71} Remarkably, renal denervation abrogates sodium retention by calcineurin inhibition, suggesting that increased sympathetic nerve activity plays a key role in the renal effect of calcineurin inhibitors.⁶ A single injection of calcineurin inhibitors acutely stimulates renal afferent nerves.⁷² However, patients with kidney transplantation still develop CIH,²⁻⁴ suggesting that renal innervation is not essential for CIH. Nonetheless, renal afferent nerve stimulation by calcineurin inhibitors may indirectly impact NMDAR activity in the hypothalamus via activating the renin-angiotensin system.29,34

In summary, our study uncovers a new mechanism in which $\alpha 28-1$, particularly $\alpha 28-1-$ bound NMDARs in the PVN, plays an essential role in calcineurin inhibitor–induced excess sympathetic outflow and hypertension. Our findings provide strong evidence linking $\alpha 28-1$ to increased synaptic NMDAR activity in the PVN and elevated sympathetic vasomotor activity in CIH. These findings not only offer new mechanistic insight into the pathogenesis of CIH but also suggest alternative, mechanism-based therapeutic targets for preventing and treating CIH. Owing to the adverse effects of the withdrawal of immunosuppressants in patients with transplants and autoimmune diseases, calcineurin inhibitors are rarely discontinued because of hypertension. Although NMDAR antagonists effectively reduce CIH,⁵ these drugs produce serious CNS adverse effects. Alternatively, gabapentinoids and $\alpha 28-1$ CT interfering peptides have no effect on physiological, $\alpha 28-1$ –free NMDARs, and thus might circumvent the adverse effects caused by general NMDAR antagonists. Because gabapentinoids are orally bioavailable, FDA-approved drugs, these agents could be readily repurposed for treating patients with CIH. Further research is warranted to validate the efficacy of gabapentinoids in patients with CIH.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

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Nonstandard Abbreviations and Acronyms:

ABP arterial blood pressure

AP5 2-amino-5-phosphonopentanoic acid

CIH calcineurin inhibitor—induced hypertension

HR heart rate

mEPSC miniature excitatory postsynaptic current

NMDAR N-methyl-D-aspartate receptor

PVN paraventricular nuclear

RSNA renal sympathetic nerve activity

RVLM rostral ventrolateral medulla

VGCC voltage-gated calcium channel

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NOVELTY AND SIGNIFICANCE

What Is Known?

 Clinically used calcineurin inhibitors including immunosuppressants for transplant rejection and for treating autoimmune diseases may cause persistent hypertension.

 Calcineurin inhibitors elevate sympathetic vasomotor activity by diminishing calcineurin activity and potentiating NMDA receptor activity in the hypothalamus.

What New Information Does This Article Contribute?

- Calcineurin inhibition increases α2δ-1-NMDA receptor interactions and induces α2δ-1-dependent NMDA receptor synaptic expression and hyperactivity in the hypothalamus.
- Disrupting α2δ-1–NMDA receptor interactions in the hypothalamus diminishes calcineurin inhibitor–potentiated sympathetic vasomotor activity.
- Systemic treatment with gabapentin or $\alpha 28-1$ genetic knockout prevents the development of calcineurin inhibitor–induced hypertension.

Calcineurin inhibitors are immunosuppressants used clinically to treat autoimmune disorders and transplant rejection. However, their prolonged use often leads to sustained hypertension, which adversely affects allograft and patient survival. Calcineurin inhibitors augment sympathetic vasomotor activity by diminishing calcineurin activity and potentiating NMDA receptor activity in the hypothalamus. Our study demonstrates the essential role of $\alpha 2\delta - 1$ (previously known as a calcium channel subunit) in the hypothalamus in the development of elevated sympathetic outflow and hypertension caused by calcineurin inhibitors. We demonstrated that systemic treatment with tacrolimus augments α2δ-1-NMDA receptor interactions and their synaptic trafficking and activity in the hypothalamus. Inhibition of $\alpha 2\delta - 1$ with gabapentin or $\alpha 2\delta - 1$ genetic knockout abolishes tacrolimus-induced NMDA receptor hyperactivity of sympatheticrelated neurons in the hypothalamus. Disrupting α2δ–1–NMDA receptor interactions in the hypothalamus reverses the tacrolimus-induced sympathetic outflow. Strikingly, concurrent treatment with gabapentin or using a $\alpha 2\delta - 1$ genetic knockout prevents the development of tacrolimus-induced hypertension. These findings uncover novel molecular mechanisms underlying the heightened sympathetic nervous system response caused by calcineurin inhibitors. Gabapentinoids (gabapentin and pregabalin), commonly used for chronic pain and epilepsy management, hold potential for repurposing in the treatment of calcineurin inhibitor-induced neurogenic hypertension.

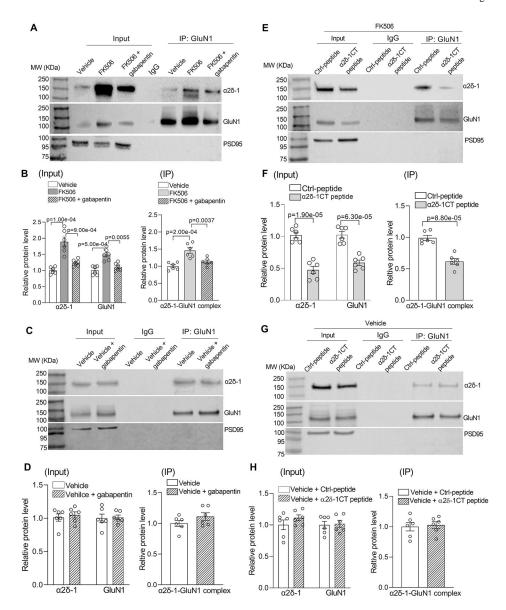


Figure 1. Calcineurin inhibition increases $\alpha 2\delta - 1 - NMDAR$ interactions and synaptic levels of NMDARs in the PVN.

A and B, Representative gel images (A) and qualification (B) show the effects of FK506 treatment with and without 100 μ M gabapentin on protein levels of $\alpha 2\delta - 1$, GluN1, and $\alpha 2\delta - 1$ –GuN1 complexes in PVN synaptosomes (n = 6 samples per group; each sample included PVN tissues from 2 male rats). C and D, Original gel images (C) and qualification (D) show the lack of effect of 100 μ M gabapentin on protein levels of $\alpha 2\delta - 1$, GluN1, or $\alpha 2\delta - 1$ –GluN1 complexes in PVN synaptosomes from vehicle control rats (n = 6 samples per group; each sample included PVN tissues from 2 male rats). E and F, Representative gel images (E) and qualification (F) show effects of pretreatment with 1 μ M control peptide (Ctrl-peptide) or 1 μ M $\alpha 2\delta - 1$ C terminus peptide ($\alpha 2\delta - 1$ CT peptide) on protein levels of $\alpha 2\delta - 1$, GluN1, and $\alpha 2\delta - 1$ –GluN1 complexes in PVN synaptosomes from FK506-treated rats (n = 6 samples per group; each sample included PVN tissues from 2 male rats). G and H, Original gel images (G) and qualification (H) show the lack of effect of

 $\alpha 28-1$ CT peptide on protein levels of $\alpha 28-1$, GluN1, or $\alpha 28-1$ -GluN1 complexes in PVN synaptosomes from vehicle control rats (n = 6 samples per group; each sample included PVN tissues from 2 male rats). MW, molecular weight. PSD95, a synaptic protein marker, was used as a loading control. One-way ANOVA with Bonferroni's *post hoc* test was used in **B**; two-tailed Student's *t* test was used in **D**, **F**, and **H**.

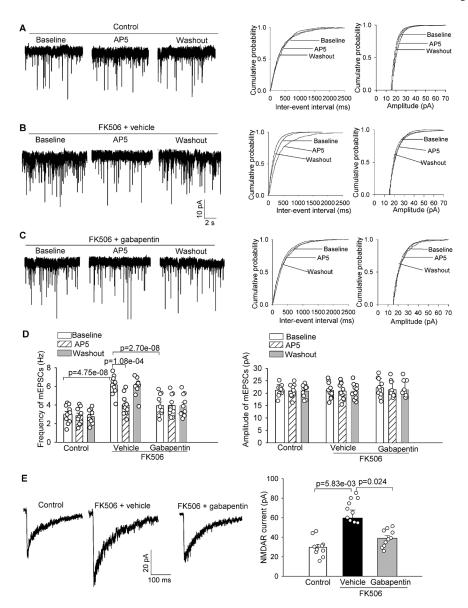


Figure 2. α 28–1 inhibition with gabapentin diminishes FK506-induced increases in presynaptic and postsynaptic NMDAR activity in spinally projecting PVN neurons.

A-C, Representative recording traces and cumulative probability plots show the effects of bath application of $50~\mu M$ AP5 on the frequency and amplitude of miniature of excitatory postsynaptic currents (mEPSCs) of spinally projecting PVN neurons in brain slices pretreated with $100~\mu M$ gabapentin in FK506-treated rats. **D**, Summary data show the effects of gabapentin and AP5 on the frequency and amplitude of mEPSCs of labeled PVN neurons in vehicle-treated control rats and FK506-treated rats (n = 10 neurons from 4 male rats per group). **E**, Representative recording traces and quantification show the effect of gabapentin ($100~\mu M$) on the amplitude of puff-elicited NMDAR currents in labeled PVN neurons in brain slices from FK506-treated rats (n = 10 neurons from 4 male rats per group). The repeated measures models were fitted for statistical analysis.

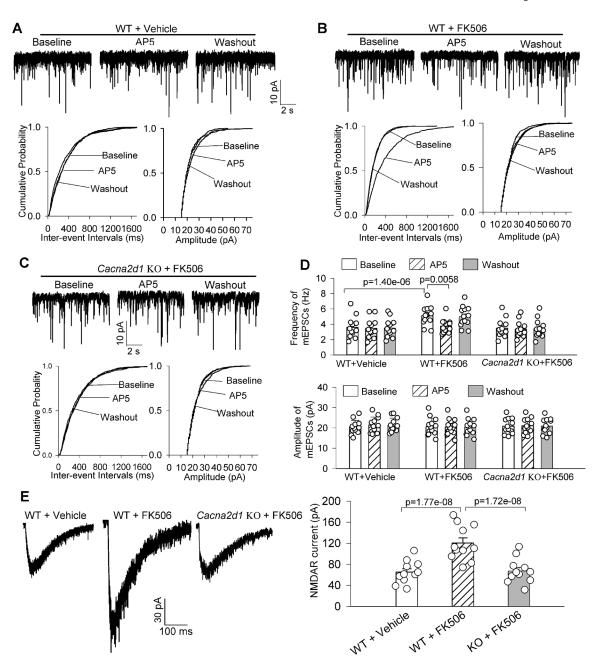


Figure 3. α .28–1 is essential for the increased synaptic NMDAR activity in spinally projecting PVN neurons by calcineurin inhibition.

A-C, Representative recording traces and cumulative probability plots show the effects of bath application of AP5 (50 μ M) on the frequency and amplitude of miniature of excitatory postsynaptic currents (mEPSCs) in labeled PVN neurons in brain slices from vehicle- treated or FK506-treated wild-type (WT) mice (**A** and **B**) and FK506-treated *Cacna2d1* knockout (KO) mice (**C**). **D**, Summary data show the effects of AP5 on mEPSCs of labeled PVN neurons in brain slices from WT and *Cacna2d1* KO mice treated with vehicle or FK506 (n = 12 neurons from 4 male mice in vehicle-treated WT mice and FK506-treated *Cacna2d1* KO mice, n = 13 neurons from 4 male mice in FK506-treated WT group). **E**, Representative

recording traces and quantification show the amplitude of puff-elicited NMDAR currents in labeled PVN neurons in WT and *Cacna2d1* KO mice treated with vehicle or FK506 (n = 11 neurons from 4 male mice per group). The repeated measures models were fitted for statistical analysis.

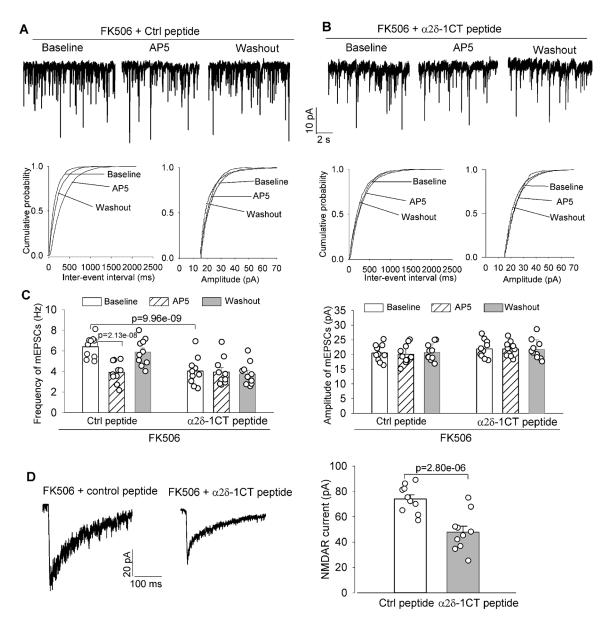


Figure 4. α 28–1-bound NMDARs are responsible for the increased glutamatergic input to PVN presympathetic neurons by calcineurin inhibition.

A-C, Original recording traces and cumulative probability plots (**A** and **B**) and quantification (**C**) show the effects of bath application of AP5 (50 μM) on miniature of excitatory postsynaptic currents (mEPSCs) in labeled PVN neurons in brain slices pretreated with 1 μM control (Ctrl) peptide or 1 μM α 2δ–1 C terminus peptide (α 2δ–1CT peptide) in FK506-treated rats (n = 10 neurons from 4 male rats per group). **D**, Representative recording traces and quantification show effects of pretreatment with 1 μM Ctrl peptide or 1 μM α 2δ–1CT peptide on the amplitude of puff-elicited NMDAR currents in labeled PVN neurons in brain slices from FK506-treated rats (n = 10 neurons from 4 male rats per group). The repeated measures models were fitted for statistical analysis.

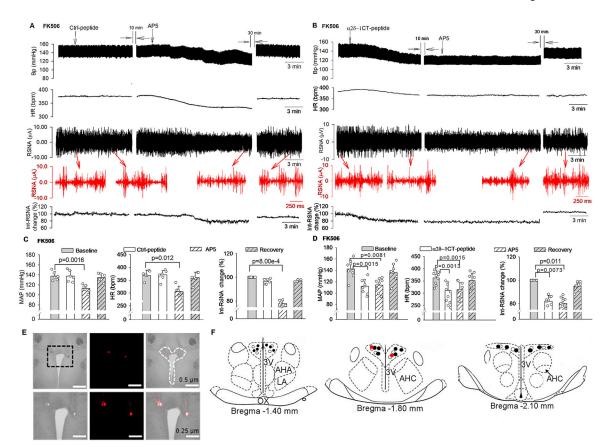
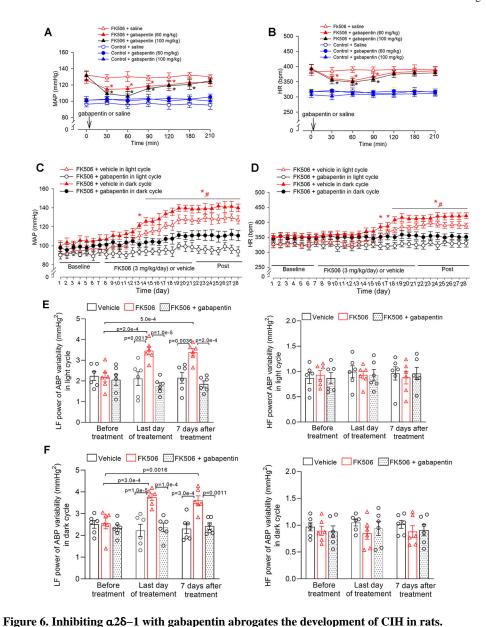


Figure 5. α 28–1-bound NMDARs in the PVN maintain elevated sympathetic vasomotor activity in CIH.

A and **B**, Original recording traces show the effects of bilateral microinjection of control (Ctrl) peptide (50 pmol in 50 nL, **A**) or α2δ−1CT peptide (50 pmol in 50 nL, **B**) followed by AP5 (1.0 nmol in 50 nL) into the PVN on arterial blood pressure (Bp), heart rate (HR), and integrated renal sympathetic activity (Int-RSNA) in FK506-treated rats. Insets (in red): expanded raw recording traces of RSNA. **C** and **D**, Mean data show changes in mean arterial blood pressure (MAP), HR, and Int-RSNA in FK506-treated rats after microinjection of Ctrl peptide (**C**, n = 6 male rats) or α2δ−1CT peptide (**D**, n = 9 male rats) followed by AP5 into the PVN. Repeated measures ANOVA with Dunnett's *post hoc* test was used in **C** and **D**. **E** and **F**, Representative low- and high-magnification brightfield and fluorescence images (**E**) and schematic drawings (**F**, the injection sites corresponding to **E** are indicated in red dots) show microinjection sites in the PVN in FK506-treated rats. O, microinjection sites in FK506-treated rats in **D**. 3V, third ventricle; AHA, anterior hypothalamic area; AHC, central division of the anterior hypothalamus; LA, latero-anterior hypothalamus; OX, optic chiasm.



A and B, Radiotelemetry recording data show acute effects of intraperitoneal injection of gabapentin (60 mg/kg or 100 mg/kg) on mean arterial blood pressure (MAP, A) and heart rate (HR, \mathbf{B}) in conscious rats treated with FK506 or vehicle (n = 6 male rats per group). C and D, Radiotelemetry recording shows the effect of concurrent treatment with vehicle or gabapentin (60 mg/kg/day in drinking water) on MAP (C) and HR (D) in FK506-treated male rats during light and dark cycles (n = 6 male rats per group). E and F, Power spectral analysis of systolic ABP variability shows changes in low-frequency (LF) and high-frequency (HF) power in FK506-treated male rats concurrently treated with vehicle or gabapentin during light (**E**) and dark (**F**) cycles; n = 6 rats per group). *P < 0.05, compared with respective baseline values within the same group, ${}^{\#}P < 0.05$, compared with respective values in the FK506 + gabapentin group at the same time point during light/dark cycles. Repeated measures ANOVA with Dunnett's post hoc test was used in A and B; two-way

ANOVA with Bonferroni's *post hoc* test was used in C, D, E, and F. Exact P values are shown in Tables S1–S4 in Supplemental Materials.

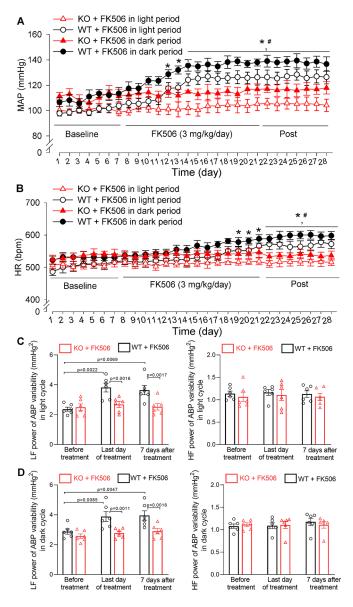


Figure 7. Genetic ablation of α2δ-1 prevents the development of CIH in mice.

A and **B**, Radiotelemetry recording data show the time course of changes in mean arterial blood pressure (MAP, **A**) and heart rate (HR, **B**) in FK506-treated wild-type mice (WT) and FK506-treated *Cacna2d1* knockout mice (KO) during light and dark cycles (n = 3 male and 3 female mice per group). **C** and **D**, Power spectral analysis of systolic ABP variability shows the changes in low-frequency (LF) and high-frequency (HF) power in FK506-treated WT and FK506-treated *Cacna2d1* KO mice during light cycle (**C**) and dark cycle (**D**) (n = 6 mice per group). *P< 0.05, compared with respective baseline values in FK506-treated WT mice during light/dark cycles (repeated measures ANOVA with Dunnett's *post hoc* test). *P< 0.05, compared with respective values in FK506-treated *Cacna2d1* KO mice at the same time point during light/dark cycles (two-way ANOVA with Bonferroni's *post hoc* test). Exact P values are shown in Tables S5 and S6 in Supplemental Materials.