



# *Review* **Benefits of Combining** *Sonchus brachyotus DC.* **Extracts and Synbiotics in Alleviating Non‑Alcoholic Fatty Liver Disease**

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Abstract: Non-alcoholic fatty liver disease, commonly abbreviated to NAFLD, is a pervasive ailment within the digestive system, exhibiting a rising prevalence, and impacting individuals at increasingly younger ages. Those afflicted by NAFLD face a heightened vulnerability to the onset of profound liver fibrosis, cardiovascular complications, and malignancies. Currently, NAFLD poses a significant threat to human health, and there is no approved therapeutic treatment for it. Recent studies have shown that synbiotics, which regulate intestinal microecology, can positively impact glucolipid metabolism, and improve NAFLD‑related indicators. *Sonchus brachyotus DC.*, a Chinese herb, exhibits hepatoprotective and potent antioxidant properties, suggesting its potential therapeutic use in NAFLD. Our preclinical animal model investigation suggests that the synergy between *Sonchus brachyotus DC.* extracts and synbiotics is significantly more effective in preventing and treat– ing NAFLD, compared to the isolated use of either component. As a result, this combination holds the potential to introduce a fresh and encouraging therapeutic approach to addressing NAFLD.

**Keywords:** NAFLD; synbiotics; gut microbiota; *Sonchus brachyotus DC.* extracts



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# **1. Introduction**

NAFLD is a clinicopathological condition characterized by hepatic steatosis and lipid accumulation. It serves as a prominent underlying factor for chronic liver diseases, carrying a notable risk of liver fibrosis, which can occur in up to 25% of cases[[1\]](#page-12-0). NAFLD, often correlated with metabolic syndrome and type 2 diabetes mellitus (T2DM), fosters the progression of cirrhosis, hepatocellular carcinoma, cardiovascular disease, and extrahepatic cancers. Consequently, cardiovascular ailments, malignant tumors, and advanced liver conditions are the chief contributors to mortality in individuals afflicted by NAFLD.

In recent years, the prevalence of NAFLD has continued to rise, due to improved material living standards and dietary changes. Presently, non‑alcoholic fatty liver disease (NAFLD) stands as one of the most widespread chronic liver conditions globally. It affects roughly 30% of adults, constituting a substantial fraction of the hepatic-related morbidity and mortality[[2\]](#page-12-1). In 2018, the global prevalence of NAFLD was estimated at 25%, with 27.37% of cases in Asia. Modeling studies project a continuous 30% increase in fatty liver disease worldwide by 2030 compared to previous years in eight countries, including China, the UK, France, and Germany[[3,](#page-13-0)[4](#page-13-1)]. This alarming trend, with the NAFLD prevalence in Chinarising from 13% to 43% over the last 20 years [[5\]](#page-13-2), signifies a growing burden on individuals and society. Currently, there are no approved therapies for NAFLD, except for weight loss through various interventions to slow the disease progression. Current treatments focus on improving insulin sensitivity, and reducing liver enzyme levels through lifestyle changes and medications. However, these interventions lack comprehensiveness and effectiveness in alleviating NAFLD. The side effects and long-term administration of insulin sensitizers and vitamin E may limit their widespread acceptance[[6](#page-13-3)]. Although

dietary intervention and exercise are recommended as first-line therapies, they are often required in combination with pharmacological or surgical treatments for disease improvement, which are not entirely effective. Consequently, the lack of an efficient therapy for NAFLD continues to be a significant problem in healthcare. The current NAFLD research aims to develop single‑agent and effective therapies to improve patient compliance.

As the research on intestinal microecology advances rapidly, probiotics and prebiotics have attracted increasing interest among researchers, due to their regulatory functions in the gut. Synbiotics, a synergistic blend of probiotics and prebiotics, offer an indirect yet profound advantage to human wellbeing. They operate by fostering the growth of advantageous gut microbiota, and overseeing their metabolic processes. This intricate interplay culminates in a fortified immunity, and refined metabolic equilibrium. Moreover, synbiotics have been found to improve blood glucose and insulin levels[[7\]](#page-13-4), which aligns with the prevailing view that NAFLD has a connection with metabolic levels and insulin resistance. This suggests the potential efficacy of synbiotics in treating NAFLD and related disorders.

This article presents a systematic review of the research on synbiotics development, intestinal microecology, NAFLD treatment, and the application of synbiotics in NAFLD treatment. Additionally, it proposes an improved synbiotic combination, comprising lactulose, arabinose, and *Lactobacillus plantarum*, which enhances the therapeutic efficacy of synbiotics for NAFLD when paired with traditional Chinese herbal *Sonchus brachyotus DC.* extracts.

#### **2. NAFLD and Its Treatment**

#### *2.1. Nonalcoholic Fatty Liver Disease (NAFLD)*

Nonalcoholic fatty liver disease (NAFLD) is a prevalent liver metabolic syndrome characterized by an abnormal metabolism and excessive lipid accumulation, resulting in steatosis in over 5% of liver cells[[8\]](#page-13-5). While its pathological alterations bear a resemblance to alcoholic liver disease, NAFLD manifests in individuals who do not have a history of significant alcohol intake. It is frequently associated with metabolically abnormal obesity, featuring dyslipidemia and hyperglycemia. NAFLD is a leading cause of chronic liver dis‑ eases, progressing to nonalcoholic steatohepatitis (NASH), cirrhosis, and complications such as an abnormal glucose tolerance, hypertension, hyperviscosity, and coronary heart disease. Additionally, NAFLD has a close connection with the development of hepato– cellular carcinoma [\[9](#page-13-6)]. Recent studies have introduced the term metabolic dysfunctionassociated fatty liver disease (MAFLD) to encompass NAFLD, expanding its pathology classification, and explicitly associating it with type 2 diabetes and metabolic dysfunction. This redefinition opens up new avenues for future NAFLD treatment [\[10](#page-13-7),[11\]](#page-13-8).

#### *2.2. The Pathogenesis of NAFLD*

The pathogenesis of NAFLD is still not understood. Petersen et al.[[12\]](#page-13-9) suggest that NAFLD results from an overabundance of triglyceride accumulation in the liver due to an energy surplus, which is consistent with the higher prevalence of NAFLD in obese individuals. The liver accumulates lipids by absorbing fatty acids released from peripheral adipose tissue and ingested orally. Skeletal muscle insulin resistance leads to excessive hepatic fat accumulation via the shifting of glucose from skeletal muscle glycogen synthe– sis to de novo lipid synthesis [\[13](#page-13-10)]. Consequently, hepatic insulin resistance hinders the function of glycogen synthase, redirecting glucose into lipogenic processes, and promoting NAFLD. Studies using mice lacking hepatic glycogen synthase have shown increased hepatic adipogenesis and NAFLD development due to hepatic insulin resistance[[14\]](#page-13-11). In conclusion, insulin resistance promotes excessive fatty acid intake and de novo lipogenesis in the liver, ultimately leading to hepatocyte dysfunction and NAFLD development.

The "second hit" theory, first proposed internationally in 1998, has gained wide acceptance  $[14]$ . It suggests that unhealthy lifestyles and dietary habits lead to lipid accumulation in the liver, causing hepatocyte steatosis and sustained cellular damage ("first hit"). This damage triggers the secretion of inflammatory cytokines, contributing to mitochondrial dysfunction, oxidative stress, and massive liver cell death, ultimately resulting in hepatocyte damage and steatohepatitis ("second hit") (Figure [1\)](#page-2-0). The emergence of NAFLD is intricately linked to heightened oxidative stress, an undue accumulation of lipids within the liver, and inflammatory processes. Research has elucidated that disruptions in lipid metabolism lead to the buildup of hepatic lipids, thereby exerting a profound impact on the generation of various reactive oxygen species (ROS). These sources of ROS span from mitochondria and the endoplasmic reticulum (ER) to  $\mathrm{NADPH}$  oxidase, contributing to the oxidative milieu within the liver. NAFLD results in excessive mitochondrial ROS production in the liver, and increased ROS generation is involved in the regulation of insulin signaling and lipid-metabolism-related enzyme expression and activity, further promot-ingNAFLD development [[15\]](#page-13-12). The redox signaling pathway interacts with the immune signaling network to regulate the inflammatory response. Consequently, the pathogenic progression of NAFLD is extremely intricate, prompting the formulation of the "multiple" hit"theory to elucidate its developmental mechanisms [[16\]](#page-13-13).

<span id="page-2-0"></span>

**Figure 1.** The second-hit pathogenesis of NAFLD. A genetic predisposition coupled with an unhealthy diet leads to an excessive accumulation of the liver accumulation of the liver. Concurrent liver accumulation of the heighthealthy diet leads to an excessive accumulation of fatty acids in the liver. Concurrently, the heightened insulin resistance in the adipose tissue initiates an inflammatory response, prompting the re‐ lease of adipocytokines. These cytokines activate hepatic stellate cells, initiating a cascade that culminates in hepatic fibrosis and fatty liver, constituting the "first hit". Subsequently, this sets the stage for persistent hepatocyte damage, stress responses, and hepatocyte apoptosis, collectively manifesting as liver hepatitis, or the "second hit".

Although the exact pathogenesis of NAFLD remains enigmatic, there is a consensus regarding its substantial correlation with metabolic disorders and insulin resistance. This connection is substantiated by the "second hit" and "multiple hit" theories, along with the corresponding empirical findings  $[17,18]$  $[17,18]$ . Amongst the two theories, the former garners greater acceptance. This theory posits that, subsequent to hepatocytes undergoing the ini‐ tial impact, insulin resistance and leptin resistance ensue, culminating in hepatic steatosis. This process, in turn, triggers inflammation, necrosis, and fibrosis as a response to oxidative stress. This highlights the role of an abnormal endocrine axis function in NAFLD development. These findings may provide new directions for the development of drugs or functional foods to treat NAFLD, with the goal of improving insulin resistance, and preventing the second blow to hepatocytes from oxidative stress, thus preventing the development of hepatocellular carcinoma, cirrhosis, and complications.

## *2.3. The Treatment for NAFLD*

Ganesh et al. [\[6](#page-13-3)] summarized the current global treatment options and recommendations for NAFLD in their review. Currently, there are no approved treatments for NAFLD, and weight loss interventions may be the most effective approach. Pharmacological interventions for NAFLD primarily seek to enhance insulin sensitivity, while reducing hepatic inflammation and fibrosis biomarkers. An ongoing study in the United States for NASH treatment has shown that omega-3 fatty acid esters (eicosapentaenoic acid) may be a potential candidate for the first‑line treatment of hypertriglyceridemia in NAFLD patients. Certain studies propose that polyunsaturated fatty acids (PUFAs) play a pivotal role in enhancing hepatic steatosis and the biochemical markers associated with non-alcoholic fatty liver disease (NAFLD), while also improving insulin sensitivity, and mitigating inflamma-tion [\[19](#page-13-16),[20\]](#page-13-17). However, another study indicated that PUFA did not have disease-modifying effects in NASH patients with diabetes[[21\]](#page-13-18), suggesting certain limitations in its use for these patients. Further research is required to determine the optimal dosage of omega-3 PUFA supplementation, and its effects on hepatic lipids.

Insulin sensitizers, extensively evaluated and recognized in previous treatment studies for NASH, have shown promising results. For instance, pioglitazone, a trial drug, has demon-strated an improvement in steatohepatitis, compared to vitamin E and the placebo [\[22](#page-13-19)]. Long– term pioglitazone treatment resulted in significant improvements in liver damage. However, long-term administration may carry the risks of cardiovascular disease and bone loss [\[23](#page-13-20)[–28](#page-14-0)]. Metformin, commonly used to treat type 2 diabetes, also improves NAFLD, by increasing glucose utilization in the peripheral tissues. Studies have demonstrated that metformin improves insulin sensitivity, liver histology, and serum alanine transaminase (ALT) levels, particularly in obese NASH patients [\[29](#page-14-1)]. Vitamin E, a lipophilic antioxidant, has been shown to slow down the progression of NAFLD, by effectively reducing oxida-tive damage, and inhibiting inflammatory cytokine production in the liver [\[22](#page-13-19),[30\]](#page-14-2). Clinical trials have revealed that, after 96 weeks of treatment, a notable 43% of all participants exhibited marked histological enhancements. These improvements were particularly pronounced in relation to the amelioration of inflammation in both the hepatocyte ballooning and lobular areas. These positive changes can be largely attributed to a reduction in oxidative stress‑induced damage[[31\]](#page-14-3).

Probiotics have also been mentioned in treatment protocols, and have shown potential benefits in NAFLD treatment. In a review published in 2021, Yao et al.[[32](#page-14-4)] analyzed the application of intestinal flora in NAFLD treatment, specifically highlighting the positive effects of *Lactobacillus plantarum* in reducing ALT and AST levels in patients. The use of active ingredients from Chinese herbal medicine, following the discovery of artemisinin for the treatment of malaria, has gained increasing attention. Traditional Chinese medicine often refers to fatty liver as "liver accumulation" and "liver gangrene", and several herbs, such as *Rhizoma Coptidis* (Huang Lian), *Radix Salvia Miltiorrhizae* (Dan Shen), *Rhei Rhizoma* (Da Huang), *Fructus Gardeniae* (Zhi Zi), and *Semen Cassiae* (Jue Ming Zi), are commonly used in NAFLD treatment, due to their natural ingredients that can reverse steatosis, regulate blood lipids, inhibit inflammation, and resist oxidative stress. Studies have shown the therapeutic effects of *Radix Salvia Miltiorrhizae* extract salvia phenolic acid B, *Semen Cassiae*, and *Fructus Gardeniae* extracts in NAFLD via the inhibition of inflammation, and anti‑oxidative stress [\[33](#page-14-5)[–35](#page-14-6)]. These findings provide valuable insights into, and theoretical support for, combining synbiotics with active ingredients from herbal medicines in NAFLD treatment.

#### **3. Definition and Function of, and Research into, Synbiotics**

# *3.1. Definition*

Synbiotics are a combination of probiotics and prebiotics that promote the physiological activity of probiotics, selectively adjust the distribution of intestinal flora, and enhance the effectiveness and longevity of beneficial bacteria. Probiotics are beneficial intestinal bacteria that colonize the human body, and positively impact the body's microecology. They achieve this goal by regulating the host mucosa and systemic immune function, or

by balancing the intestinal flora, promoting nutrient absorption, and maintaining intestinal and overall health. Common examples of intestinal probiotics include *Bifidobacterium* and *Lactobacillus*. On the contrary, prebiotics are organic compounds that cannot be digested or absorbed. They are specifically designed to nourish and promote growth in the beneficial microbes essential for metabolic functions and overall development. Their profound impact on the host's wellbeing stems from their ability to modulate changes within the intestinal microbe community. These changes, in turn, exert influence over the endocrine, barrier, and immune functions, effectively restoring a harmonious balance to the intestinal microecology. The concept of synbiotics emerged in 1995, and they were defined as a synergistic blend of probiotics and prebiotics. This unique combination serves to enhance the host's health, by fostering the survival and successful integration of microorganism food supplements in the digestive tract. Additionally, synbiotics selectively stimulate growth in specific health-promoting bacteria, and activate their metabolism, thereby yielding a targeted and amplified positive impact [\[36](#page-14-7)]. This definition is consistent with those of pre‑ biotics and probiotics, confirming their complementary and synergistic relationship. In August 2020, the International Scientific Association of Probiotics and Prebiotics (ISAPP) updated the term "synbiotics", and redefined it as "a mixture comprising live microorganisms and substrate (s) selectively utilized by host microorganisms that confers a health benefit on the host" [\[37](#page-14-8)], emphasizing the valuable effects and potential of synbiotics for human health, and providing insights and guidance for future synbiotics development.

## *3.2. Functions of Synbiotics*

Probiotics, functioning as beneficial microorganisms within the intestinal tract, play a pivotal role in mitigating the host's weight, and ameliorating metabolic irregularities. This is achieved through the regulation of distinct metabolic pathways and immune responses, via their metabolites. Furthermore, they contribute to weight reduction in the host, by facilitating the absorption of fats and cholesterol. Probiotics play a role in improvements inthe digestion of lactose  $[38]$  $[38]$ , the treatment of antibiotic-associated diarrhea  $[39]$  $[39]$ , and the prevention of necrotizing small intestinal colitis in preterm neonates, as demonstrated in systematic reviews and meta-analyses  $[40]$ , remission induction in inflammatory bowel disease [\[41](#page-14-12)],the prevention and control of hyperglycemia [[7\]](#page-13-4), the improvement of lipid levels and inflammation, including a reduction in total cholesterol, the level of high-density lipoprotein (HDL), and the inflammatory marker tumor necrosis factor (TNF)- $\alpha$  [\[42](#page-14-13)], and bloodglucose and insulin resistance in diabetic patients [[43\]](#page-14-14). Probiotics additionally generate short-chain fatty acids. These compounds help reduce the local pH, encourage the production of immunomodulatory cytokines, and stimulate mucin generation. As a result, they foster a more robust and beneficial intestinal microenvironment[[44\]](#page-14-15).

Synbiotics represent a synergy between probiotics and prebiotics, endowing the host with augmented advantages. This synergy manifests through the selective enhancement of favorable bacterial growth within the intestinal tract, as well as the activation of path‑ ways that bolster the metabolism of bacteria conducive to human wellbeing. Furthermore, they amplify the viability in, and integration of, probiotics within the gut, consequently fostering a more propitious microbial milieu within the intestines, thus promoting human health. These benefits include enhancing immunity, preventing respiratory and gastrointestinal infections, promoting lactose digestion, alleviating symptoms such as bloating and abdominal pain due to lactose intolerance, enhancing nutrient absorption, and facilitating the degradation of indigestible plant fibers into short-chain fatty acids. Overall, synbiotics significantly improve insulin secretion and sensitivity, by modulating the intestinal flora's composition. This therapeutic effect has been demonstrated in NAFLD, via the alleviation of insulin resistance and the promotion of the organism's metabolism. Furthermore, synbiotics enhance the absorption of antioxidant components in the combination, leading to hepatoprotective effects.

### *3.3. Gut Microecology and NAFLD*

The human gut microbiota comprises 10–100 trillion microorganisms, surpassing the number of human cells. Its distribution and composition are closely related to the host's life, and play essential roles in the host's immune response, food digestion, regulation of the intestinal secretory function, neural signaling, drug metabolism, and catabolism [\[45](#page-14-16)]. In recent years, the advancements in intestinal microecology have garnered considerable interest. Studies have identified a correlation between the distribution of intestinal microflora and various diseases, leading to research on the potential treatment of human diseases through regulation of the intestinal microecology.

The "hepatic–intestinal axis" is a critical component of the human metabolic system. It entails intricate biliary, portal, and systemic interconnections, linking the liver and the intestine. This axis is regulated by various signals from the intestinal flora and its metabolites, environmental toxins, and food antigens[[46\]](#page-14-17). Long‑term studies have demonstrated that a healthy intestinal flora helps regulate the function of crucial metabolic organs such as the liver, promoting host metabolic stability  $[45]$ . Irregularities within the intestinal microbiota, including an excessive proliferation of pathogenic bacteria, bacterial translocation, the generation of harmful metabolites, and perturbation in the signaling for substance and energy metabolism along the gut–liver axis, have the potential to modify the immune landscape of the host. These alterations, in turn, play a pivotal role in fostering the emergence of liver-related disorders, notably non-alcoholic fatty liver disease (NAFLD) and alcoholic liver disease (ALD) [\[47](#page-14-18)] (Figure [2\)](#page-5-0). Studies with NAFLD case studies have shown statistically significant differences in the levels of *Lactobacillus*, *Bifidobacterium*, and *Escherichia coli* in patients, compared to the control group [\[48\]](#page-14-19). Multiple case studies focusing on NAFLD (non‑alcoholic fatty liver disease) and NASH (non‑alcoholic steatohepatitis) have unveiled noteworthy distinctions in the intestinal flora, within both animal models and patients. These disparities underscore the paramount importance of the intestinal microecology in the progression of NAFLD[[49,](#page-14-20)[50](#page-14-21)]. Recent research indicates that interventions aimed at manipulating the gut microbiota could present a promising therapeutic avenue for liver diseases [\[51](#page-14-22),[52\]](#page-14-23).

<span id="page-5-0"></span>

**Figure 2.** Gut microbiota and NAFLD. Obesity or a high-fat diet can trigger an imbalance in the **Figure 2.** Gut microbiota and NAFLD. Obesity or a high‑fat diet can trigger an imbalance in the interesting the function microecology, and dispute the function of the interesting of the intestinal microecology, and disrupt the function of the intestinal barrier. This disruption allows pathogenic bacteria and harmful metabolites to migrate to the liver. Consequently, this can lead to apoptosis in the liver parenchymal cells, and the activation of non-parenchymal immune cells within the liver. These cascading effects ultimately give rise to liver lesions and the progression of NAFLD.

Moreover, NAFLD is not a singular fatty liver disorder but, rather, a spectrum of metabolism-related diseases closely associated with diabetes mellitus and metabolic syndrome. T2DM is a common complication of NAFLD, and shares similarities with NAFLD, such as insulin resistance and a chronic low‑grade inflammatory state. Research has shown a close association between the intestinal microecology and T2DM, revealing potential mechanisms through which regulating the intestinal microecology benefits T2DM patients. Furthermore, patients with NAFLD and T2DM exhibit similar patterns in microflora dis‑ tribution, suggesting the possibility of simultaneously improving NAFLD and T2DM by modulating intestinal microflora [\[53](#page-15-0)]. In light of these findings, a new synbiotic combination consisting of lactulose, arabinose, and *Lactobacillus plantarum* has been developed, showing promising results in treating T2DM[[54\]](#page-15-1). Its efficacy on NAFLD model mice has been validated, and further formulation modification is underway, to obtain a new synbiotic mixture.

#### **4.** *Sonchus brachyotus DC.*

According to Flora of China (FRPS), both *Sonchus brachyotus DC.* and *Sonchus arven‑ sis L.* are annual herbs belonging to the genus *Sonchus*, in the family *Compositae*. *Sonchus brachyotus DC.* is also known as *Ixeris sonchifolia*, wild bitter cabbage, and *Ixeris polycephala Cass*. It has a lengthy history of medicinal use in Chinese medicine, where it is referred to as bitter cabbage. Early Chinese medical texts, such as the classic *Shennong Threshes the Hundred Grasses* and *Compendium of Materia Medica*, describe its bitter and cold taste, and highlight its effects in clearing heat, eliminating toxins, reducing swelling and pus, promoting blood circulation and removing blood stasis, clearing the lungs and relieving cough, invigorating the liver and promoting diuresis, and aiding digestion and harmonizing the stomach. Additionally, as well as its medicinal uses, *Sonchus brachyotus DC.* is commonly consumed as a wild vegetable in folk traditions, due to its high nutritional value, as it contains protein, various amino acids, carotene, and rich trace elements [\[55](#page-15-2)].

Several studies have showcased that disruptions in the ecosystem of the gastrointestinal tract can hinder the organism's typical metabolic functions. This can trigger a range of pathological damages, and contribute to the advancement of chronic diseases [\[56](#page-15-3)[–59](#page-15-4)]. Hence, the precise manipulation of the intestinal microbiota emerges as indispensable in addressing metabolic irregularities. Over recent years, bioactive compounds derived from botanical sources have assumed a pivotal role in fostering diverse functionalities, particularly in governing the dynamics of intestinal microecology, a domain where investigations have borne remarkable outcomes. Abundant research underscores the fact that *Sonchus* species, apart from their culinary use, boast a plethora of medicinal properties [\[60](#page-15-5)[–64](#page-15-6)]. *Sonchus* species boast an abundance of vitamin C and omega-3 PUFA [\[65](#page-15-7)], potentially imbuing them with noteworthy anti-inflammatory and antioxidant properties. Omega-3 PUFA has been employed in numerous clinical trials for the therapeutic intervention of NAFLD, according to reports [\[66](#page-15-8)[–68](#page-15-9)]. Significantly, species of *Sonchus* have been documented for their potential therapeutic effects in diabetes management [\[65](#page-15-7)]. The findings showcased in these studies underscore the potential of *Sonchus* species in contributing to the management of metabolic syndromes. With *Sonchus brachyotus DC*. serving as a traditional vernacular herb, renowned for its efficacy in dispelling heat-related pollutants, arresting hemorrhages, and addressing ailments such as acute dysentery and enteritis, its significant anti-inflammatory and antioxidant attributes have been documented. These has been established through the identification of compounds that exhibit robust antioxidant and antiinflammatory properties[[61,](#page-15-10)[69](#page-15-11)]. The components derived from *Sonchus brachyotus DC*. exhibit the ability to effectively restrain the production of free radicals, as well as the process of lipid peroxidation [\[61](#page-15-10)]. Furthermore, recent research has unveiled the finding that *Sonchus brachyotus DC*. harbors a plethora of bioactive constituents, notably flavonoids [\[70](#page-15-12)]. These compounds have demonstrated a remarkable capability to impede the proliferation of various microbes, such as *Escherichia coli*, *Enterobacter cloacae*, and *Klebsiella pneumoniae*, while concurrently fostering the induction of apoptosis[[71\]](#page-15-13). Apart from its demonstration

of antioxidant and antibacterial activities in vitro, there are reports suggesting that *Sonchus brachyotus DC*. holds the capability to mitigate drug‑induced oxidative stress damage, by regulating the abundance of the intestinal microflora [\[72](#page-15-14)]. This further substantiates the favorable contribution of *Sonchus brachyotus DC*. in the treatment of NAFLD.

*Sonchus brachyotus DC*. has identified several chemical constituents in the plant, including β‑sitosterol, lignan, apigenin, and quercetin, which possess biological activities, such as antibacterial, hepatoprotective, anti-inflammatory, antioxidant, and insulin-resista nce‑improving properties. For example, studies conducted at the University of Science and Technology of Tianjin have identified potent antioxidant active components in an extract from *Sonchus brachyotus DC*., suggesting its potential application in the treatment of NAFLD, due to its remarkable hepatoprotective and antioxidant effects (unpublished data from our collaborators). Additionally, *Sonchus* species are abundant in fatty acids, such as Omega‑3 PUFA [\[73](#page-15-15)], which is the main active ingredient in drugs under development for NAFLD treatment, further supporting its potential use in NAFLD treatment.

Utilizing these data, the article suggests a therapeutic approach for NAFLD, involving a synergistic utilization of extracts from *Sonchus brachyotus DC*. (SBE) and a synbiotics compound. An appropriate amount of SBE is added to the previously developed synbiotics, consisting of lactulose, arabinose, and *Lactobacillus plantarum*, for compounding. The efficacy of this combination is then tested in a mouse model of NAFLD, to assess the improvement in their pathology. As a result, the addition of herbal active ingredients significantly improves the liver condition of NAFLD‑model mice.

# **5. Synbiotic Compounds for NAFLD Treatment**

#### *5.1. Synbiotic Compounds*

The synbiotic compound is composed of functional sugars and *Lactobacillus plantarum*, with a cell count of live bacteria ranging from  $10^8$  to  $10^{10}$  CFU/g under deposit number CGMCC 8198. By weight percentage, the synbiotic compound consists of 30% lactulose, 30% arabinose, and 40% *Lactobacillus plantarum*. To obtain SBE, an aqueous solution of ethanol is mixed with the crude powder of *Sonchus brachyotus DC.*, followed by the ultrasonic extraction, rotary evaporation, and freeze-drying of the supernatant. The resulting SBE is used at a dosage of 2.0 g/kg per serving.

L-arabinose is beneficial for controlling glycolipid metabolism, by inhibiting the metabolic conversion of sucrose, effectively reducing blood glucose levels. It does so by inhibiting the activity of disaccharide-hydrolyzing enzymes, which leads to an increase in sucrose metabolism[[74,](#page-15-16)[75](#page-15-17)]. The remaining undigested sucrose is then catabolized by microorganisms in the colon, producing organic acids that inhibit fat synthesis in the liver. L-arabinose has been demonstrated to alter the abundance and diversity of the intestinal flora as a prebiotic, resulting in improvements in the body lipid rate, blood lipid levels, fasting blood glucose, glucose tolerance status, and the extent of liver damage in an animal model of metabolic syndrome [\[76](#page-15-18),[77\]](#page-15-19).

Lactulose, a disaccharide composed of galactose and fructose, is not digested after ingestion, but is catabolized by the intestinal flora in the colon, leading to changes in the pHvalue in the intestinal tract [[78](#page-15-20)]. This change stimulates colonic motility, relieves constipation, and regulates the physiological rhythm of the colon. The breakdown products of lactulose also inhibit the proliferation of harmful bacteria, and alter the number of dominant intestinal flora. Studies have shown that lactulose supplementation in mice fed a high-salt diet resulted in changes in the structure of the intestinal flora, and an improve in intestinal permeability[[79\]](#page-15-21).

*Lactobacillus plantarum* belongs to the *Lactobacillus* genus of the *phylum Firmicutes*, and is commonly used in fermented foods, due to its numerous health benefits. Experimental studies have confirmed the beneficial effects of *Lactobacillus plantarum* in decreasing bloodlipid levels, and regulating the intestinal flora [[80–](#page-16-0)[87\]](#page-16-1). These effects include immunomodulatory properties, lower levels of ALT and AST, the inhibition of pathogenic bacteria proliferation, decreased serum cholesterol levels, and the prevention of cardiovascular diseases. *Lactobacillus plantarum* also helps maintain the balance of the intestinal microflora, promotes nutrient absorption, alleviates lactose intolerance, and inhibits the growth of tumor cells. In this experiment, the strain used is *Lactobacillus plantarum* (L. plan‑ tarum) CGMCC 8198, which has demonstrated the abilities to decrease serum cholesterol, and exhibit antioxidant activity [\[88](#page-16-2)].

The synbiotic compounds composed of L‑arabinose, lactulose, and *Lactobacillus plan‑ tarum* have demonstrated hypoglycemic and hypolipidemic effects, as well as control of the body weight. A study by Jiang et al.[[54\]](#page-15-1) used this synbiotic compound in a T2DM mouse model, and observed a significant decrease in body weight and the levels of blood glucose and blood lipids. These findings provide a basis for the further application of this combination in NAFLD. Additionally, SBE is added to the list of compounds to enhance liver antioxidative capacities and further improve the body's metabolic capacity. The combination effectively controls blood glucose and blood lipid levels, reduces body weight, decreases ALT and AST levels, and improves liver damage in NAFLD patients.

# *5.2. Animal Experiment*

In preclinical experiments, an increasing body of research underscores the therapeutic potential inherent in extracts from various traditional Chinese medicines (TCMs) for ad‑ dressing NAFLD. Ye et al.[[89\]](#page-16-3) discovered that administering luteolin to db/db mice over an 8-week period yielded notable outcomes. Through the mediation of LXR-SREBP-1c signaling, luteolin exhibited a remarkable capacity to mitigate both liver cholesterol accumulation, and the initiation of novel lipid synthesis, in the context of NAFLD. This, in turn, contributed to the promotion of hepatic steatosis attenuation. Further investigations per‑ taining to NAFLD demonstrated that rutin, a natural flavonoid replete with diverse biological effects, holds promise. Rutin exhibited the potential to reinstate PPAR enzyme activity, while concurrently suppressing the expression of fatty acid synthases, including FAS and ACC, in murine models of NAFLD. This dual action effectively hindered the escalation of oxidative stress levels, and facilitated an amelioration of liver disease-associated damage[[90](#page-16-4)[,91](#page-16-5)]. A comparable outcome was observed in another flavonoid study conducted by Panchita et al. [\[92](#page-16-6)], who discovered that administering quercetin-3-O- $\beta$ -glucoside (Q3G) and fructooligosaccharide (FOS) to rats on a dextrin diet for 45 days resulted in a reduction in both oral glucose tolerance tests (OGTTs) and total cholesterol levels. Additionally, there was an increase in GLP-1, indicating that the combination of Q3G and FOS holds therapeutic potential for NAFLD. *Sonchus brachyotus DC*. has been found to contain multiple identifiable flavonoids. These discoveries strongly indicate that *Sonchus brachyotus DC*. may possess predictable therapeutic effects on NAFLD. In order to substantiate the efficacy of the *Sonchus brachyotus DC*. extract and synbiotics combination for NAFLD treatment, this blend was tested on an established murine NAFLD model, to evaluate its capacity for ameliorating NAFLD pathology. Upon the successful induction of the NAFLD model, groups of seven mice were analyzed for NAFLD-related markers (Table [1\)](#page-8-0), and diverse interventions were implemented during the designated period. The potential therapeutic impact of *Sonchus brachyotus DC*. on NAFLD was evaluated via an assessment of its phenotypic influence, and the liver histology and plasma levels of the pertinent cytokines in the mice.

<span id="page-8-0"></span>**Table 1.** The proportion of symbiotics components and supplementation.



# *5.3. Efficacy of SBE in Combination of Synbiotics in Alleviating NAFLD*

Following drug interventions, the mice in the model group showed elevated blood sugar and insulin levels, while the treatment groups exhibited varying degrees of control over their blood glucose and lipids, leading to significant reductions in FBG and INS levels (Figure [3B](#page-9-0),C). This indicates that the synbiotics combination has a significant positive effect on NAFLD progression in mice, with the experimental group (NAFLD+Synb+SBE) showing the most significant improvement. Therefore, the synbiotics combination demonstrates an effective control of blood glucose and insulin levels.

<span id="page-9-0"></span>

**Figure 3.** Effects of a synbiotic and SBE compound on NAFLD. After administration in mice, the **Figure 3.** Effects of a synbiotic and SBE compound on NAFLD. After administration in mice, the changes in  $(A)$  body weight,  $(B)$  fasting blood glucose (FBG),  $(C)$  serum insulin concentration (INS), (D) total cholesterol (TC), (E) triglycerides (TG), and (F) alanine aminotransferase are shown. The representative results from three independent experiments are included. NS, no significance;  $* p < 0.05$ ; 0.05; \*\* *p* < 0.01. n = 3 biological replicates/group; one-way ANOVA. All analyses were performed \*\* *p* < 0.01. n = 3 biological replicates/group; one‑way ANOVA. All analyses were performed using the GraphPad Prism 8 software (Version 8.0.2; GraphPad Software, Inc., San Diego, CA, USA).

The levels of total cholesterol and triglyceride content in the mice under intervention intervention groups, compared to the model group (NAFLD), and the experimental group (NAFLD+SBE+Synb) showed the most significant decrease. Although the total cholesterol (TC) levels were not significantly different, each group still demonstrated some control ability compared to the model group (NAFLD). This suggests that a combination of synbiotics and SBE plays a more effective role in regulating hyperlipidemia. were assessed (Figures [3](#page-9-0)D and [3](#page-9-0)E, respectively). The TG level was reduced in the drug

Furthermore, the alanine aminotransferase (ALT) levels in the serum of the NAFLD and synbiotics, compared to the model control group and the control groups (Figure [3F](#page-9-0)). This reduction indicates a reversal in the progression of chronic liver damage due to oxida-<br>.. the superstance of the integration of the integration of  $\mathcal{L}$  and  $\mathcal{L}$  focus for  $\mathcal{L}$ mice were significantly reduced by the combination of extract from *Sonchus brachyotus DC*. tive stress.

Histological analysis of the liver further confirmed the beneficial effects of the treat–<br>Histological analysis of the liver further confirmed the beneficial effects of the treat– ment. The liver wet weight and the liver coefficient in the embodiment and proportional proves were reduced compared to the twin ED model group. The reduction is abnormalities was more significant in the embodiment (SBE+Synb) and proportion 2 (SBE)<br>answers assumed the proportion 1 (Sanb), supporting a formalle relatif SBE in linearmates that a computerion of *Lactobacial* computerion of *Lactobacillus reuterinium* in the computerion of  $\frac{1}{2}$ tion (Table [2\)](#page-10-0). All these data are from our accepted Chinese patent (no. CN2022110780786). groups were reduced compared to the NAFLD model group. The reduction in liver tissue groups, compared to proportion 1 (Synb), suggesting a favorable role of SBE in liver protec‑



<span id="page-10-0"></span>**Table 2.** Results of the liver histological analysis.

#### *5.4. Clinical Research*

The gut microbiome is considered a key factor in the development of NAFLD[[59\]](#page-15-4), so the regulation of the intestinal microecology is a crucial focus for future NAFLD treat– ments. In recent years, the clinical research on synbiotics for NAFLD treatment has shown promising results[[93–](#page-16-7)[102\]](#page-17-0). The selection of appropriate probiotics and prebiotics to im‑ prove NAFLD progression is becoming increasingly feasible and predictable.

One randomized controlled trial (RCT) involving 50 NAFLD patients demonstrated that a combination of *Lactobacillus reuteri* and inulin led to reduced hepatic steatosis, along with im‑ provements in body weight, BMI, and serum uric acid levels[[95](#page-16-8)]. Another study by Behrouz et al. [\[103\]](#page-17-1), which treated 89 NAFLD patients with various probiotics, also showed significant effects on blood glucose levels, and other related biochemical markers of NAFLD[[103](#page-17-1)]. Similarly, in patients with NASH, the administration of *Bifidobacterium longum* along with fructooligosaccharides resulted in significant reductions in inflammation-related factors, AST levels, and steatosis [\[104\]](#page-17-2). Combining probiotics with metformin in a NASH trial showed improved outcomes for liver injury [\[105\]](#page-17-3). A separate RCT involving *Lactobacillus acidophilus* supplementation in NAFLD patients demonstrated reduced ALT and AST levels[[106](#page-17-4)]. Aller et al. [\[107\]](#page-17-5) utilized *Lactobacillus bulgaricus* and *Streptococcus thermophiles*, observing similar ben‑ eficial effects in NAFLD patients.

These clinical studies provide strong evidence of the positive effects of synbiotics in NAFLD treatment. Building on these developments, the positive impact of prebiotics in NAFLD treatment is further confirmed. Our experiments demonstrated that administering appropriate prebiotic combinations to model mice improved liver damage and overall condition. The intestinal microecology's application in treating various diseases has become a significant research area, with ongoing investigations in experimental and clinical research. Compared to traditional drug therapies, synbiotics offer milder, safer, and more cost‑effective options that are suitable for most patients, and have a more noticeable effect. Introducing synbiotics formally into NAFLD treatment, and enhancing their therapeutic effect through new synbiotic combinations, will offer great hope to patients.

Based on our results and models of previous clinical studies, we propose a clinical re‑ search plan for new synbiotic combinations. The plan involves several main areas of focus, including the effects of an oral intake of synbiotic combinations with SBE on the control of body weight, blood glucose levels, and other physiological indexes in NAFLD patients, and whether this treatment would effectively improve liver damage in these patients. Only systematic medical examinations would be able to reveal the clinical significance of this synbiotic and SBE combination.

# **6. Possible Mechanisms of Synbiotics in NAFLD**

Numerous studies have shown that the dysregulation of the gut microbiome is strongly linked to NAFLD severity [\[108](#page-17-6)[–112](#page-17-7)]. Animal experiments by Roy et al.[[113\]](#page-17-8) demonstrated that gut microbial transplantation (FMT) from different mouse models led to germ‑free mice exhibiting distinct phenotypes of liposynthesis and steatosis. Another experiment byChiu et al. [[93](#page-16-7)] involved transferring gut microbes from NASH patients to germ-free mice on a high‑fat diet, resulting in mice showing biochemical changes similar to those of NASH patients, including increased serum ALT and AST levels[[114\]](#page-17-9). This suggests that the gut microbiome plays a decisive role in NAFLD development.

Further research indicates that the gut microbiome mainly influences the human body through its metabolites, with changes in bile acid metabolism signals, intestinal permeability, and shortchain fatty acid production being the main mechanisms involved[[115–](#page-17-10)[117\]](#page-17-11). Primary bile acids support fat-soluble substance digestion and absorption in the intestine, maintain the intestinal barrier, and regulate lipid and sugar metabolism, by activating receptors such as the farnesoid X receptor (FXR) and G protein-coupled bile acid receptor 1 (GPBAR1/TGR5) [\[118](#page-17-12)[–120\]](#page-17-13). The activation of FXR promotes the secretion of fibroblast growth factor 15/19 (FGF15/19), which targets the liver, and inhibits hepatic accumulation and degeneration[[97](#page-16-9)[–99\]](#page-16-10). TGR5 activation increases the secretion of glucagon-like peptide-1 (GLP-1), which, together with FXR, reduces blood lipid levels, and improves fatty liver symptoms [\[121](#page-17-14),[122](#page-17-15)]. Intestinal flora regulation also impacts in‑ testinal permeability, with changes in the intestinal bacteria affecting the synthesis and secretion of angiopoietin-like 4 protein (ANGPTL4) in the small intestine, thereby regulating liver fat storage [\[123](#page-17-16),[124](#page-17-17)].

Studies by Okubo et al. [\[125](#page-18-0)], using methionine-choline-deficient NASH mice, demonstrated that administering the *Lactobacillus casei* strain Shirota (Lcs) increased the number of other lactic acid bacteria. This led to decreased intestinal inflammation and serum LPS concentration in NASH mice, improved liver damage, and proved the effectiveness of the *Lactobacillus casei* strain Shirota (Lcs) in NAFLD. Similar results were obtained by Nguyen et al. [\[126](#page-18-1)] using *Lactobacillus plantarum* PH04, which showed significant cholesterol and triglyceride‑lowering effects. A report by Zhao et al. [\[127\]](#page-18-2) indicated that administering *Lactobacillus plantarum* NA136 to a high-fat diet-induced NAFLD mouse model effectively reduced NAFLD severity, and reversed insulin resistance (Figure [4\)](#page-11-0).

<span id="page-11-0"></span>

**Figure 4.** The potential mechanisms of synbiotics in NAFLD. The therapeutic impacts of synbiotics **Figure 4.** The potential mechanisms of synbiotics in NAFLD. The therapeutic impacts of synbiotics on NAFLD encompass three key aspects. Firstly, the catabolism of prebiotics results in the generation tion of SCFAs. These SCFAs enter hepatocytes via the MCP1 transporter and, subsequently, bind to of SCFAs. These SCFAs enter hepatocytes via the MCP1 transporter and, subsequently, bind to PPAR,  $\mathcal{P}(t)$  in the catabolism signaling leptin signaling. Secondly, the catabolism of prediction  $\mathcal{P}(t)$  is yields actively in the catabolism of prediction  $\mathcal{P}(t)$  is a conditional of prediction  $\mathcal{P}(t)$  is a condi effectively inhibiting leptin signaling. Secondly, the catabolism of prebiotics yields active acidic sub-<br>effectively inhibiting leptin signaling. Secondly, the catabolism of prebiotics yields active acidic substances (such as  $\rm H_2O_2$  and SCFAs), along with antibacterial agents (including bacteriocin and bacteriophages). This dual action serves to impede the proliferation of detrimental bacteria. Lastly, the influence of probiotics is manifested in their ability to enhance the maintenance of the intestinal barrier integrity. This is achieved through mechanisms involving the TLR- or DN-SIGN-mediated regulation of tight junctions. DC‑SIGN, dendritic cell‑specific intercellular adhesion molecule‑3‑grabbing non-integrin; IECs, intestinal epithelial cells; MCP1, monocyte chemoattractant protein 1; SCFAs, **7. Future Prospective**  short-chain fatty acids; TLR, toll-like receptor.

## **7. Future Prospective**

NAFLD is a widespread chronic liver disease affecting nearly one-quarter of the world's population, and its prevalence is projected to keep increasing at a high rate, imposing a significant burden on healthcare systems. Currently, there are no approved treatments for NAFLD, leading to considerable distress among patients. Lifestyle modifications are considered the most reliable approach to managing the condition. However, with the deepening research on the intestinal microbiome and its connection to metabolic diseases, synbiotics, as food items capable of regulating the intestinal flora, offer great promise in alleviating NAFLD.

Various probiotics and prebiotics possess distinct characteristics, and their combination as a synbiotic provides a dietary option that benefits human health, and shows promise in regulating and alleviating diseases. This approach is expected to become one of the essential methods for preventing and treating metabolism-related diseases in the future. The synbiotic combination used in this experiment effectively regulates the intestinal flora through L‑arabinose, lactulose, and *Lactobacillus plantarum*. It leads to reduced blood glucose and blood lipids, improved insulin resistance and liver damage in patients, and enhanced protection and improvement of liver function, through the addition of active ingredient extracts from traditional Chinese medicinal herbs. Animal experiments have demonstrated the positive effects of this synbiotic combination in treating NAFLD.

We anticipate that this synbiotic combination will perform similarly well in clinical trials, serving as a dietary aid to control patients' weight, reduce blood glucose, lipid, ALT, and AST levels, improve liver damage, and promote overall health in NAFLD patients, and individuals with metabolism-related diseases. Furthermore, it may enhance the defenses of healthy individuals, and serve as a preventive measure against metabolic syndrome, NAFLD, T2DM, and other diseases, when incorporated into their diets.

Given the ongoing rise in NAFLD incidence, effective preventive or early intervention methods will significantly alleviate the burden of NAFLD on the general population. We are optimistic about the potential of the synbiotics combination research to eventually provide a low‑cost, safe, and highly effective therapeutic approach to the public, bringing good news to NAFLD patients, and contributing to improved overall health.

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**Institutional Review Board Statement:** All mice were bred and kept under specific pathogen-free conditions, following the National Institute of Health Guide for Care and Use of Laboratory An‑ imals. The present study and its associated experimental procedures received approval from the Institutional Animal Care and Use Committee of Wuhan University of Science and Technology (Ap‑ proved No.: SYXK (鄂) 2018-0045). Every aspect of animal housing and experimentation strictly adhered to the guidelines outlined in the National Institute of Health's Guide for the Care and Use of Laboratory Animals.

**Data Availability Statement:** The data are available from the corresponding author.

**Conflicts of Interest:** The authors declare no conflict of interest.

# **References**

- <span id="page-12-0"></span>1. Younossi, Z.M.; Koenig, A.B.; Abdelatif, D.; Fazel, Y.; Henry, L.; Wymer, M. Global epidemiology of nonalcoholic fatty liver disease‑Meta‑analytic assessment of prevalence, incidence, and outcomes. *Hepatology* **2016**, *64*, 73–84.[[CrossRef](https://doi.org/10.1002/hep.28431)]
- <span id="page-12-1"></span>2. Fernando, D.H.; Forbes, J.M.; Angus, P.W.; Herath, C.B. Development and Progression of Non-Alcoholic Fatty Liver Disease: The Role of Advanced Glycation End Products. *Int. J. Mol. Sci.* **2019**, *20*, 5037. [\[CrossRef](https://doi.org/10.3390/ijms20205037)]
- <span id="page-13-0"></span>3. Estes, C.; Anstee, Q.M.; Arias-Loste, M.T.; Bantel, H.; Bellentani, S.; Caballeria, J.; Colombo, M.; Craxi, A.; Crespo, J.; Day, C.P.; et al. Modeling NAFLD disease burden in China, France, Germany, Italy, Japan, Spain, United Kingdom, and United States for the period 2016–2030. *J. Hepatol.* **2018**, *69*, 896–904.[[CrossRef](https://doi.org/10.1016/j.jhep.2018.05.036)]
- <span id="page-13-1"></span>4. Younossi, Z.; Tacke, F.; Arrese, M.; Chander Sharma, B.; Mostafa, I.; Bugianesi, E.; Wai-Sun Wong, V.; Yilmaz, Y.; George, J.; Fan, J.; et al. Global Perspectives on Nonalcoholic Fatty Liver Disease and Nonalcoholic Steatohepatitis. *Hepatology* **2019**, *69*, 2672–2682.[[CrossRef](https://doi.org/10.1002/hep.30251)]
- <span id="page-13-2"></span>5. Wu, T.; Liao, X.; Zhong, B. Epidemiology of nonalcoholic fatty liver disease in some regions of China. *J. Clin. Hepatol.* **2020**, *36*, 1370–1373. [\[CrossRef](https://doi.org/10.3969/j.issn.1001-5256.2020.06.039)]
- <span id="page-13-3"></span>6. Ganesh, S.; Rustgi, V.K. Current Pharmacologic Therapy for Nonalcoholic Fatty Liver Disease. *Clin. Liver Dis.* **2016**, *20*, 351–364. [\[CrossRef\]](https://doi.org/10.1016/j.cld.2015.10.009) [\[PubMed](https://www.ncbi.nlm.nih.gov/pubmed/27063274)]
- <span id="page-13-4"></span>7. Ruan, Y.; Sun, J.; He, J.; Chen, F.; Chen, R.; Chen, H. Effect of Probiotics on Glycemic Control: A Systematic Review and Meta-Analysis of Randomized, Controlled Trials. *PLoS ONE* **2015**, *10*, e0132121.[[CrossRef\]](https://doi.org/10.1371/journal.pone.0132121)
- <span id="page-13-5"></span>8. Chalasani, N.; Younossi, Z.; Lavine, J.E.; Charlton, M.; Cusi, K.; Rinella, M.; Harrison, S.A.; Brunt, E.M.; Sanyal, A.J. The diagnosis and management of nonalcoholic fatty liver disease: Practice guidance from the American Association for the Study of Liver Diseases. *Hepatology* **2018**, *67*, 328–357.[[CrossRef](https://doi.org/10.1002/hep.29367)]
- <span id="page-13-6"></span>9. Marengo, A.; Rosso, C.; Bugianesi, E. Liver Cancer: Connections with Obesity, Fatty Liver, and Cirrhosis. *Annu. Rev. Med.* **2016**, *67*, 103–117. [\[CrossRef\]](https://doi.org/10.1146/annurev-med-090514-013832) [\[PubMed](https://www.ncbi.nlm.nih.gov/pubmed/26473416)]
- <span id="page-13-7"></span>10. Eslam, M.; Newsome, P.N.; Sarin, S.K.; Anstee, Q.M.; Targher, G.; Romero-Gomez, M.; Zelber-Sagi, S.; Wai-Sun Wong, V.; Dufour, J.F.; Schattenberg, J.M.; et al. A new definition for metabolic dysfunction-associated fatty liver disease: An international expert consensus statement. *J. Hepatol.* **2020**, *73*, 202–209.[[CrossRef](https://doi.org/10.1016/j.jhep.2020.03.039)]
- <span id="page-13-8"></span>11. Eslam, M.; Sanyal, A.J.; George, J. MAFLD: A Consensus‑Driven Proposed Nomenclature for Metabolic Associated Fatty Liver Disease. *Gastroenterology* **2020**, *158*, 1999–2014.e1. [\[CrossRef\]](https://doi.org/10.1053/j.gastro.2019.11.312)
- <span id="page-13-9"></span>12. Petersen, K.F.; Oral, E.A.; Dufour, S.; Befroy, D.; Ariyan, C.; Yu, C.; Cline, G.W.; DePaoli, A.M.; Taylor, S.I.; Gorden, P.; et al. Leptin reverses insulin resistance and hepatic steatosis in patients with severe lipodystrophy. *J. Clin. Investig.* **2002**, *109*, 1345–1350. [\[CrossRef\]](https://doi.org/10.1172/JCI0215001)
- <span id="page-13-10"></span>13. Irimia, J.M.; Meyer, C.M.; Segvich, D.M.; Surendran, S.; DePaoli‑Roach, A.A.; Morral, N.; Roach, P.J. Lack of liver glycogen causes hepatic insulin resistance and steatosis in mice. *J. Biol. Chem.* **2017**, *292*, 10455–10464.[[CrossRef](https://doi.org/10.1074/jbc.M117.786525)][[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/28483921)
- <span id="page-13-11"></span>14. Day, C.P.; James, O.F. Steatohepatitis: A tale of two "hits"? *Gastroenterology* **1998**, *114*, 842–845.[[CrossRef](https://doi.org/10.1016/S0016-5085(98)70599-2)][[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/9547102)
- <span id="page-13-12"></span>15. Chen, Z.; Tian, R.; She, Z.; Cai, J.; Li, H. Role of oxidative stress in the pathogenesis of nonalcoholic fatty liver disease. *Free Radic. Biol. Med.* **2020**, *152*, 116–141. [\[CrossRef\]](https://doi.org/10.1016/j.freeradbiomed.2020.02.025)
- <span id="page-13-13"></span>16. Buzzetti, E.; Pinzani, M.; Tsochatzis, E.A. The multiple‑hit pathogenesis of non‑alcoholic fatty liver disease (NAFLD). *Metab. Clin. Exp.* **2016**, *65*, 1038–1048. [\[CrossRef\]](https://doi.org/10.1016/j.metabol.2015.12.012)[[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/26823198)
- <span id="page-13-14"></span>17. Almeda‑Valdés, P.; Cuevas‑Ramos, D.; Alberto Aguilar‑Salinas, C. Metabolic syndrome and non‑alcoholic fatty liver disease. *Ann. Hepatol.* **2009**, *8*, S18–S24.[[CrossRef](https://doi.org/10.1016/S1665-2681(19)31822-8)][[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/19381120)
- <span id="page-13-15"></span>18. Rector, R.S.; Thyfault, J.P.; Wei, Y.; Ibdah, J.A. Non‑alcoholic fatty liver disease and the metabolic syndrome: An update. *World J. Gastroenterol.* **2008**, *14*, 185–192.[[CrossRef](https://doi.org/10.3748/wjg.14.185)]
- <span id="page-13-16"></span>19. Capanni, M.; Calella, F.; Biagini, M.R.; Genise, S.; Raimondi, L.; Bedogni, G.; Svegliati‑Baroni, G.; Sofi, F.; Milani, S.; Abbate, R.; et al. Prolonged n-3 polyunsaturated fatty acid supplementation ameliorates hepatic steatosis in patients with non-alcoholic fatty liver disease: A pilot study. *Aliment. Pharmacol. Ther.* **2006**, *23*, 1143–1151.[[CrossRef\]](https://doi.org/10.1111/j.1365-2036.2006.02885.x)
- <span id="page-13-17"></span>20. Masterton, G.S.; Plevris, J.N.; Hayes, P.C. Review article: Omega-3 fatty acids—A promising novel therapy for non-alcoholic fatty liver disease. *Aliment. Pharmacol. Ther.* **2010**, *31*, 679–692. [\[CrossRef\]](https://doi.org/10.1111/j.1365-2036.2010.04230.x)
- <span id="page-13-18"></span>21. Dasarathy, S.; Dasarathy, J.; Khiyami, A.; Yerian, L.; Hawkins, C.; Sargent, R.; McCullough, A.J. Double-blind randomized placebo-controlled clinical trial of omega 3 fatty acids for the treatment of diabetic patients with nonalcoholic steatohepatitis. *J. Clin. Gastroenterol.* **2015**, *49*, 137–144.[[CrossRef\]](https://doi.org/10.1097/MCG.0000000000000099)[[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/24583757)
- <span id="page-13-19"></span>22. Sanyal, A.J.; Chalasani, N.; Kowdley, K.V.; McCullough, A.; Diehl, A.M.; Bass, N.M.; Neuschwander‑Tetri, B.A.; Lavine, J.E.; Tonascia, J.; Unalp, A.; et al. Pioglitazone, vitamin E, or placebo for nonalcoholic steatohepatitis. *N. Engl. J. Med.* **2010**, *363*, 1185–1186. [\[CrossRef](https://doi.org/10.1056/NEJMoa0907929)]
- <span id="page-13-20"></span>23. Pearlman, M.; Loomba, R. State of the art: Treatment of nonalcoholic steatohepatitis. *Curr. Opin. Gastroenterol.* **2014**, *30*, 223–237. [\[CrossRef\]](https://doi.org/10.1097/mog.0000000000000060)
- 24. Hajiaghamohammadi, A.A.; Ziaee, A.; Oveisi, S.; Masroor, H. Effects of metformin, pioglitazone, and silymarin treatment on non‑alcoholic Fatty liver disease: A randomized controlled pilot study. *Hepat. Mon.* **2012**, *12*, e6099.[[CrossRef](https://doi.org/10.5812/hepatmon.6099)]
- 25. Bell, L.N.; Wang, J.; Muralidharan, S.; Chalasani, S.; Fullenkamp, A.M.; Wilson, L.A.; Sanyal, A.J.; Kowdley, K.V.; Neuschwander-Tetri, B.A.; Brunt, E.M.; et al. Relationship between adipose tissue insulin resistance and liver histology in nonalcoholic steatohepatitis: A pioglitazone versus vitamin E versus placebo for the treatment of nondiabetic patients with nonalcoholic steatohepatitis trial follow‑up study. *Hepatology* **2012**, *56*, 1311–1318.[[CrossRef\]](https://doi.org/10.1002/hep.25805)[[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/22532269)
- 26. Takahashi, Y.; Sugimoto, K.; Inui, H.; Fukusato, T. Current pharmacological therapies for nonalcoholic fatty liver dis‑ ease/nonalcoholic steatohepatitis. *World J. Gastroenterol.* **2015**, *21*, 3777–3785. [\[CrossRef\]](https://doi.org/10.3748/wjg.v21.i13.3777)
- 27. Gastaldelli, A.; Harrison, S.A.; Belfort‑Aguilar, R.; Hardies, L.J.; Balas, B.; Schenker, S.; Cusi, K. Importance of changes in adipose tissue insulin resistance to histological response during thiazolidinedione treatment of patients with nonalcoholic steatohepatitis. *Hepatology* **2009**, *50*, 1087–1093.[[CrossRef\]](https://doi.org/10.1002/hep.23116)
- <span id="page-14-0"></span>28. Promrat, K.; Lutchman, G.; Uwaifo, G.I.; Freedman, R.J.; Soza, A.; Heller, T.; Doo, E.; Ghany, M.; Premkumar, A.; Park, Y.; et al. A pilot study of pioglitazone treatment for nonalcoholic steatohepatitis. *Hepatology* **2004**, *39*, 188–196. [\[CrossRef](https://doi.org/10.1002/hep.20012)]
- <span id="page-14-1"></span>29. Cusi, K. Role of obesity and lipotoxicity in the development of nonalcoholic steatohepatitis: Pathophysiology and clinical impli‑ cations. *Gastroenterology* **2012**, *142*, 711–725.e6.[[CrossRef\]](https://doi.org/10.1053/j.gastro.2012.02.003)[[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/22326434)
- <span id="page-14-2"></span>30. Lavine, J.E.; Schwimmer, J.B.; Van Natta, M.L.; Molleston, J.P.; Murray, K.F.; Rosenthal, P.; Abrams, S.H.; Scheimann, A.O.; Sanyal, A.J.; Chalasani, N.; et al. Effect of vitamin E or metformin for treatment of nonalcoholic fatty liver disease in children and adolescents: The TONIC randomized controlled trial. *JAMA* **2011**, *305*, 1659–1668.[[CrossRef](https://doi.org/10.1001/jama.2011.520)][[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/21521847)
- <span id="page-14-3"></span>31. Sato, K.; Gosho, M.; Yamamoto, T.; Kobayashi, Y.; Ishii, N.; Ohashi, T.; Nakade, Y.; Ito, K.; Fukuzawa, Y.; Yoneda, M. Vitamin E has a beneficial effect on nonalcoholic fatty liver disease: A meta‑analysis of randomized controlled trials. *Nutrition* **2015**, *31*, 923–930.[[CrossRef](https://doi.org/10.1016/j.nut.2014.11.018)][[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/26059365)
- <span id="page-14-4"></span>32. Yao, M.; Qv, L.; Lu, Y.; Wang, B.; Berglund, B.; Li, L. An Update on the Efficacy and Functionality of Probiotics for the Treatment of Non‑Alcoholic Fatty Liver Disease. *Engineering* **2021**, *7*, 679–686.[[CrossRef](https://doi.org/10.1016/j.eng.2020.01.017)]
- <span id="page-14-5"></span>33. Li, B.; Chen, Y.; Pan, J.J.G.T.C.M. The hepatoprotective effect of semen cassiae extract by antagonizing insulin resistance and inhibiting oxidative‑glycation in rats with nonalcoholic fatty liver disease. *Glob. Tradit. Chin. Med.* **2015**, *8*, 1171–1175.
- 34. Meng, L.‑C.; Zheng, J.‑Y.; Qiu, Y.‑H.; Zheng, L.; Zheng, J.‑Y.; Liu, Y.‑Q.; Miao, X.‑L.; Lu, X.‑Y. Salvianolic acid B ameliorates non‑alcoholic fatty liver disease by inhibiting hepatic lipid accumulation and NLRP3 inflammasome in ob/ob mice. *Int. Im‑ munopharmacol.* **2022**, *111*, 109099. [\[CrossRef](https://doi.org/10.1016/j.intimp.2022.109099)]
- <span id="page-14-6"></span>35. Zhang, Y.; Zhou, F. Effects of geniposide on inflammation and oxidative stress of ApoE knockout mice with atherosclerosis and none‑alcoholic fatty liver disease. *Tradit. Chin. Drug Res. Clin. Pharmacol.* **2015**, *26*, 581–586.
- <span id="page-14-7"></span>36. Gibson, G.R.; Roberfroid, M.B. Dietary modulation of the human colonic microbiota: Introducing the concept of prebiotics. *J. Nutr.* **1995**, *125*, 1401–1412. [\[CrossRef\]](https://doi.org/10.1093/jn/125.6.1401)
- <span id="page-14-8"></span>37. Marco, M.L.; Sanders, M.E.; Gänzle, M.; Arrieta, M.C.; Cotter, P.D.; De Vuyst, L.; Hill, C.; Holzapfel, W.; Lebeer, S.; Merenstein, D.; et al. The International Scientific Association for Probiotics and Prebiotics (ISAPP) consensus statement on fermented foods. *Nat. Reviews. Gastroenterol. Hepatol.* **2021**, *18*, 196–208.[[CrossRef\]](https://doi.org/10.1038/s41575-020-00390-5)
- <span id="page-14-9"></span>38. Sanders, M.E. Probiotics in 2015: Their Scope and Use. *J. Clin. Gastroenterol.* **2015**, *49*, S2–S6.[[CrossRef](https://doi.org/10.1097/MCG.0000000000000350)]
- <span id="page-14-10"></span>39. D'Souza, A.L.; Rajkumar, C.; Cooke, J.; Bulpitt, C.J. Probiotics in prevention of antibiotic associated diarrhoea: Meta-analysis. *BMJ* **2002**, *324*, 1361.[[CrossRef](https://doi.org/10.1136/bmj.324.7350.1361)]
- <span id="page-14-11"></span>40. Deshpande, G.; Rao, S.; Patole, S.; Bulsara, M. Updated meta‑analysis of probiotics for preventing necrotizing enterocolitis in preterm neonates. *Pediatrics* **2010**, *125*, 921–930.[[CrossRef](https://doi.org/10.1542/peds.2009-1301)]
- <span id="page-14-12"></span>41. Shen, J.; Zuo, Z.X.; Mao, A.P. Effect of probiotics on inducing remission and maintaining therapy in ulcerative colitis, Crohn's disease, and pouchitis: Meta‑analysis of randomized controlled trials. *Inflamm. Bowel Dis.* **2014**, *20*, 21–35.[[CrossRef](https://doi.org/10.1097/01.MIB.0000437495.30052.be)]
- <span id="page-14-13"></span>42. Gao, X.; Zhu, Y.; Wen, Y.; Liu, G.; Wan, C. Efficacy of probiotics in non‑alcoholic fatty liver disease in adult and children: A meta‑analysis of randomized controlled trials. *Hepatol. Res. Off. J. Jpn. Soc. Hepatol.* **2016**, *46*, 1226–1233.[[CrossRef](https://doi.org/10.1111/hepr.12671)][[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/26866817)
- <span id="page-14-14"></span>43. Sun, J.; Buys, N.J. Glucose‑ and glycaemic factor‑lowering effects of probiotics on diabetes: A meta‑analysis of randomised placebo‑controlled trials. *Br. J. Nutr.* **2016**, *115*, 1167–1177.[[CrossRef\]](https://doi.org/10.1017/S0007114516000076)
- <span id="page-14-15"></span>44. Preidis, G.A.; Versalovic, J. Targeting the human microbiome with antibiotics, probiotics, and prebiotics: Gastroenterology enters the metagenomics era. *Gastroenterology* **2009**, *136*, 2015–2031. [\[CrossRef](https://doi.org/10.1053/j.gastro.2009.01.072)]
- <span id="page-14-16"></span>45. Lynch, S.V.; Pedersen, O. The Human Intestinal Microbiome in Health and Disease. *N. Engl. J. Med.* **2016**, *375*, 2369–2379. [\[CrossRef\]](https://doi.org/10.1056/NEJMra1600266)
- <span id="page-14-17"></span>46. Fan, Y.; Pedersen, O. Gut microbiota in human metabolic health and disease. *Nat. Rev. Microbiol.* **2021**, *19*, 55–71.[[CrossRef](https://doi.org/10.1038/s41579-020-0433-9)] [\[PubMed](https://www.ncbi.nlm.nih.gov/pubmed/32887946)]
- <span id="page-14-18"></span>47. Wang, R.; Tang, R.; Li, B.; Ma, X.; Schnabl, B.; Tilg, H. Gut microbiome, liver immunology, and liver diseases. *Cell. Mol. Immunol.* **2021**, *18*, 4–17. [\[CrossRef\]](https://doi.org/10.1038/s41423-020-00592-6)[[PubMed](https://www.ncbi.nlm.nih.gov/pubmed/33318628)]
- <span id="page-14-19"></span>48. Wei, Z.; Song, Z.; Xi‑xiang, A. Study on the relationship between immune system stability of intestinal microecological flora and insulin resistance in nonalcoholic fatty liver disease. *J. Community Med.* **2022**, *20*, 277–280. [\[CrossRef\]](https://doi.org/10.19790/j.cnki.JCM.2022.05.09)
- <span id="page-14-20"></span>49. Zhang, C.; Zhang, M.; Wang, S.; Han, R.; Cao, Y.; Hua, W.; Mao, Y.; Zhang, X.; Pang, X.; Wei, C.; et al. Interactions between gut microbiota, host genetics and diet relevant to development of metabolic syndromes in mice. *ISME J.* **2010**, *4*, 232–241.[[CrossRef](https://doi.org/10.1038/ismej.2009.112)]
- <span id="page-14-21"></span>50. Zhu, L.; Baker, S.S.; Gill, C.; Liu, W.; Alkhouri, R.; Baker, R.D.; Gill, S.R. Characterization of gut microbiomes in nonalcoholic steatohepatitis (NASH) patients: A connection between endogenous alcohol and NASH. *Hepatology* **2013**, *57*, 601–609.[[CrossRef](https://doi.org/10.1002/hep.26093)] [\[PubMed](https://www.ncbi.nlm.nih.gov/pubmed/23055155)]
- <span id="page-14-22"></span>51. Sharpton, S.R.; Maraj, B.; Harding-Theobald, E.; Vittinghoff, E.; Terrault, N.A. Gut microbiome-targeted therapies in nonalcoholic fatty liver disease: A systematic review, meta‑analysis, and meta‑regression. *Am. J. Clin. Nutr.* **2019**, *110*, 139–149. [\[CrossRef\]](https://doi.org/10.1093/ajcn/nqz042)
- <span id="page-14-23"></span>52. Nobili, V.; Mosca, A.; Alterio, T.; Cardile, S.; Putignani, L. Fighting Fatty Liver Diseases with Nutritional Interventions, Probiotics, Symbiotics, and Fecal Microbiota Transplantation (FMT). *Adv. Exp. Med. Biol.* **2019**, *1125*, 85–100. [\[CrossRef\]](https://doi.org/10.1007/5584_2018_318) [\[PubMed](https://www.ncbi.nlm.nih.gov/pubmed/30578461)]
- <span id="page-15-0"></span>53. Scorletti, E.; Byrne, C.D. Extrahepatic Diseases and NAFLD: The Triangular Relationship between NAFLD, Type 2‑Diabetes and Dysbiosis. *Dig. Dis.* **2016**, *34* (Suppl. S1), 11–18.[[CrossRef\]](https://doi.org/10.1159/000447276)
- <span id="page-15-1"></span>54. Jiang, H.; Cai, M.; Shen, B.; Wang, Q.; Zhang, T.; Zhou, X. Synbiotics and Gut Microbiota: New Perspectives in the Treatment of Type 2 Diabetes Mellitus. *Foods* **2022**, *11*, 2438.[[CrossRef](https://doi.org/10.3390/foods11162438)]
- <span id="page-15-2"></span>55. Wang, C.; Liu, J.; Su, Y.; Li, M.; Xie, X.; Su, J. Complete Chloroplast Genome Sequence of Sonchus brachyotus Helps to Elucidate Evolutionary Relationships with Related Species of Asteraceae. *BioMed Res. Int.* **2021**, *2021*, 9410496.[[CrossRef](https://doi.org/10.1155/2021/9410496)][[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/34901281)
- <span id="page-15-3"></span>56. Albuquerque‑Souza, E.; Sahingur, S.E. Periodontitis, chronic liver diseases, and the emerging oral‑gut‑liver axis. *Periodontology 2000* **2022**, *89*, 125–141.[[CrossRef](https://doi.org/10.1111/prd.12427)]
- 57. Hou, K.; Wu, Z.X.; Chen, X.Y.; Wang, J.Q.; Zhang, D.; Xiao, C.; Zhu, D.; Koya, J.B.; Wei, L.; Li, J.; et al. Microbiota in health and diseases. *Signal Transduct. Target. Ther.* **2022**, *7*, 135.[[CrossRef\]](https://doi.org/10.1038/s41392-022-00974-4)
- 58. Martel, J.; Chang, S.H.; Ko, Y.F.; Hwang, T.L.; Young, J.D.; Ojcius, D.M. Gut barrier disruption and chronic disease. *Trends Endocrinol. Metab. TEM* **2022**, *33*, 247–265.[[CrossRef](https://doi.org/10.1016/j.tem.2022.01.002)] [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/35151560)
- <span id="page-15-4"></span>59. Safari, Z.; Gérard, P. The links between the gut microbiome and non‑alcoholic fatty liver disease (NAFLD). *Cell. Mol. Life Sci. CMLS* **2019**, *76*, 1541–1558. [\[CrossRef\]](https://doi.org/10.1007/s00018-019-03011-w)
- <span id="page-15-5"></span>60. Wang, L.; Xu, M.L.; Liu, J.; Wang, Y.; Hu, J.H.; Wang, M.H. Sonchus asper extract inhibits LPS‑induced oxidative stress and pro‑inflammatory cytokine production in RAW264.7 macrophages. *Nutr. Res. Pract.* **2015**, *9*, 579–585. [\[CrossRef\]](https://doi.org/10.4162/nrp.2015.9.6.579)[[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/26634045)
- <span id="page-15-10"></span>61. Xia, D.Z.; Yu, X.F.; Zhu, Z.Y.; Zou, Z.D. Antioxidant and antibacterial activity of six edible wild plants (*Sonchus* spp.) in China. *Nat. Prod. Res.* **2011**, *25*, 1893–1901.[[CrossRef](https://doi.org/10.1080/14786419.2010.534093)][[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/21793765)
- 62. Hong, Y.Y.; Ma, Y.C.; Zhou, Y.G.; Gao, F.; Liu, H.C.; Chen, S.F. Paenibacillus sonchi sp. nov., a nitrogen-fixing species isolated from the rhizosphere of Sonchus oleraceus. *Int. J. Syst. Evol. Microbiol.* **2009**, *59*, 2656–2661. [\[CrossRef](https://doi.org/10.1099/ijs.0.009308-0)] [\[PubMed](https://www.ncbi.nlm.nih.gov/pubmed/19625436)]
- 63. De Paula Filho, G.X.; Barreira, T.F.; Pinheiro‑Sant'Ana, H.M. Chemical Composition and Nutritional Value of Three Sonchus Species. *Int. J. Food Sci.* **2022**, *2022*, 4181656.[[CrossRef](https://doi.org/10.1155/2022/4181656)]
- <span id="page-15-6"></span>64. Cardoso Vilela, F.; Soncini, R.; Giusti‑Paiva, A. Anxiolytic‑like effect of *Sonchus oleraceus* L. in mice. *J. Ethnopharmacol.* **2009**, *124*, 325–327.[[CrossRef](https://doi.org/10.1016/j.jep.2009.04.029)]
- <span id="page-15-7"></span>65. Li, X.‑M.; Yang, P.‑L. Research progress of Sonchus species. *Int. J. Food Prop.* **2018**, *21*, 147–157.[[CrossRef\]](https://doi.org/10.1080/10942912.2017.1415931)
- <span id="page-15-8"></span>66. Kobyliak, N.; Abenavoli, L.; Falalyeyeva, T.; Mykhalchyshyn, G.; Boccuto, L.; Kononenko, L.; Kyriienko, D.; Komisarenko, I.; Dynnyk, O. Beneficial effects of probiotic combination with omega‑3 fatty acids in NAFLD: A randomized clinical study. *Minerva Medica* **2018**, *109*, 418–428.[[CrossRef\]](https://doi.org/10.23736/S0026-4806.18.05845-7)[[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/30221912)
- 67. Oscarsson, J.; Önnerhag, K.; Risérus, U.; Sundén, M.; Johansson, L.; Jansson, P.A.; Moris, L.; Nilsson, P.M.; Eriksson, J.W.; Lind, L. Effects of free omega-3 carboxylic acids and fenofibrate on liver fat content in patients with hypertriglyceridemia and non‑alcoholic fatty liver disease: A double‑blind, randomized, placebo‑controlled study. *J. Clin. Lipidol.* **2018**, *12*, 1390–1403.e4. [\[CrossRef\]](https://doi.org/10.1016/j.jacl.2018.08.003)
- <span id="page-15-9"></span>68. Šmíd, V.; Dvořák, K.; Šedivý, P.; Kosek, V.; Leníček, M.; Dezortová, M.; Hajšlová, J.; Hájek, M.; Vítek, L.; Bechyňská, K.; et al. Effect of Omega‑3 Polyunsaturated Fatty Acids on Lipid Metabolism in Patients With Metabolic Syndrome and NAFLD. *Hepatol. Commun.* **2022**, *6*, 1336–1349.[[CrossRef\]](https://doi.org/10.1002/hep4.1906)[[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/35147302)
- <span id="page-15-11"></span>69. Lee, Y.K.; Lee, H.; Kim, Y.N.; Kang, J.; Jeong, E.J.; Rho, J.R. Sesquiterpene Lactones with Anti‑Inflammatory Activity from the Halophyte Sonchus brachyotus DC. *Molecules* **2023**, *28*, 1518. [\[CrossRef\]](https://doi.org/10.3390/molecules28041518) [\[PubMed](https://www.ncbi.nlm.nih.gov/pubmed/36838505)]
- <span id="page-15-12"></span>70. Qie, P.; Duan, X.; Wang, M.; Li, X. Antioxidant activity of each polar composition from methanol extracts of Sonchus brachyotus DC. *Sci. Technol. Food Ind.* **2016**, *37*, 146–156. [\[CrossRef\]](https://doi.org/10.13386/j.issn1002-0306.2016.16.021)
- <span id="page-15-13"></span>71. Sun, Y.; Wang, Y.; Tan, B.‑L.; Deng, Y.‑R. Comparison of Six Flavonoid Components of Closely‑Related Plants Agrimonia pilosa, Potentilla chinensis and Potentilla discolor. *Zhong Yao Cai = Zhongyaocai = J. Chin. Med. Mater.* **2016**, *39*, 991–995.
- <span id="page-15-14"></span>72. Yang, J.; Zhou, W.W.; Shi, D.D.; Pan, F.F.; Sun, W.W.; Yang, P.L.; Li, X.M. The Interaction between Oxidative Stress Biomarkers and Gut Microbiota in the Antioxidant Effects of Extracts from Sonchus brachyotus DC. in Oxazolone-Induced Intestinal Oxidative Stress in Adult Zebrafish. *Antioxidants* **2023**, *12*, 192.[[CrossRef](https://doi.org/10.3390/antiox12010192)][[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/36671053)
- <span id="page-15-15"></span>73. Guil‑Guerrero, J.L.; Giménez‑Giménez, A.; Rodríguez‑García, I.; Torija‑Isasa, M.E. Nutritional composition of *Sonchus* species (*S asper*L, *S oleraceus*L and *S tenerrimus*L). *J. Sci. Food Agric.* **1998**, *76*, 628–632.[[CrossRef](https://doi.org/10.1002/(SICI)1097-0010(199804)76:4%3C628::AID-JSFA997%3E3.0.CO;2-U)]
- <span id="page-15-16"></span>74. Krog‑Mikkelsen, I.; Hels, O.; Tetens, I.; Holst, J.J.; Andersen, J.R.; Bukhave, K. The effects of L‑arabinose on intestinal sucrase activity: Dose‑response studies in vitro and in humans. *Am. J. Clin. Nutr.* **2011**, *94*, 472–478.[[CrossRef](https://doi.org/10.3945/ajcn.111.014225)]
- <span id="page-15-17"></span>75. Shibanuma, K.; Degawa, Y.; Houda, K. Determination of the transient period of the EIS complex and investigation of the suppression of blood glucose levels by L‑arabinose in healthy adults. *Eur. J. Nutr.* **2011**, *50*, 447–453. [\[CrossRef\]](https://doi.org/10.1007/s00394-010-0154-3) [\[PubMed](https://www.ncbi.nlm.nih.gov/pubmed/21165628)]
- <span id="page-15-18"></span>76. Zhao, L.; Wang, Y.; Zhang, G.; Zhang, T.; Lou, J.; Liu, J. L‑Arabinose Elicits Gut‑Derived Hydrogen Production and Ameliorates Metabolic Syndrome in C57BL/6J Mice on High‑Fat‑Diet. *Nutrients* **2019**, *11*, 3054.[[CrossRef](https://doi.org/10.3390/nu11123054)][[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/31847305)
- <span id="page-15-19"></span>77. Li, Y.; Pan, H.; Liu, J.X.; Li, T.; Liu, S.; Shi, W.; Sun, C.; Fan, M.; Xue, L.; Wang, Y.; et al. L‑Arabinose Inhibits Colitis by Modulating Gut Microbiota in Mice. *J. Agric. Food Chem.* **2019**, *67*, 13299–13306. [\[CrossRef](https://doi.org/10.1021/acs.jafc.9b05829)] [\[PubMed](https://www.ncbi.nlm.nih.gov/pubmed/31674784)]
- <span id="page-15-20"></span>78. Chen, X.; Zuo, Q.; Hai, Y.; Sun, X.J. Lactulose: An indirect antioxidant ameliorating inflammatory bowel disease by increasing hydrogen production. *Med. Hypotheses* **2011**, *76*, 325–327. [\[CrossRef](https://doi.org/10.1016/j.mehy.2010.09.026)]
- <span id="page-15-21"></span>79. Zhang, Z.; Zhao, J.; Tian, C.; Chen, X.; Li, H.; Wei, X.; Lin, W.; Zheng, N.; Jiang, A.; Feng, R.; et al. Targeting the Gut Microbiota to Investigate the Mechanism of Lactulose in Negating the Effects of a High-Salt Diet on Hypertension. *Mol. Nutr. Food Res.* **2019**, *63*, e1800941.[[CrossRef\]](https://doi.org/10.1002/mnfr.201800941)[[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/30825362)
- <span id="page-16-0"></span>80. Yin, Y.; Liao, Y.; Li, J.; Pei, Z.; Wang, L.; Shi, Y.; Peng, H.; Tan, Y.; Li, C.; Bai, H.; et al. Lactobacillus plantarum GX17 benefits growth performance and improves functions of intestinal barrier/intestinal flora among yellow‑feathered broilers. *Front. Immunol.* **2023**, *14*, 1195382. [\[CrossRef\]](https://doi.org/10.3389/fimmu.2023.1195382)
- 81. Wu, Y.; Jha, R.; Li, A.; Liu, H.; Zhang, Z.; Zhang, C.; Zhai, Q.; Zhang, J. Probiotics (*Lactobacillus plantarum* HNU082) Supplemen‑ tation Relieves Ulcerative Colitis by Affecting Intestinal Barrier Functions, Immunity-Related Gene Expression, Gut Microbiota, and Metabolic Pathways in Mice. *Microbiol. Spectr.* **2022**, *10*, e0165122.[[CrossRef](https://doi.org/10.1128/spectrum.01651-22)]
- 82. Hao, H.; Zhang, X.; Tong, L.; Liu, Q.; Liang, X.; Bu, Y.; Gong, P.; Liu, T.; Zhang, L.; Xia, Y.; et al. Effect of Extracellular Vesicles Derived From Lactobacillus plantarum Q7 on Gut Microbiota and Ulcerative Colitis in Mice. *Front. Immunol.* **2021**, *12*, 777147. [\[CrossRef\]](https://doi.org/10.3389/fimmu.2021.777147)
- 83. Ding, Q.; Cao, F.; Lai, S.; Zhuge, H.; Chang, K.; Valencak, T.G.; Liu, J.; Li, S.; Ren, D. Lactobacillus plantarum ZY08 relieves chronic alcohol‑induced hepatic steatosis and liver injury in mice via restoring intestinal flora homeostasis. *Food Res. Int.* **2022**, *157*, 111259.[[CrossRef](https://doi.org/10.1016/j.foodres.2022.111259)][[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/35761571)
- 84. Yao, C.; Tian, W.; Song, J.; Wang, J. Antihyperlipidaemic effect of microencapsulated Lactobacillus plantarum LIP-1 on hyperlipidaemic rats. *J. Sci. Food Agric.* **2020**, *100*, 2007–2017.[[CrossRef](https://doi.org/10.1002/jsfa.10218)][[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/31849068)
- 85. Wang, L.X.; Liu, K.; Gao, D.W.; Hao, J.K. Protective effects of two Lactobacillus plantarum strains in hyperlipidemic mice. *World J. Gastroenterol.* **2013**, *19*, 3150–3156.[[CrossRef](https://doi.org/10.3748/wjg.v19.i20.3150)][[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/23716997)
- 86. Huang, C.H.; Lin, C.H.; Huang, H.H.; Tsai, G.J. Development of Fermented Shrimp Shell Product with Hypoglycemic and Hypolipidemic Effects on Diabetic Rats. *Metabolites* **2022**, *12*, 695.[[CrossRef\]](https://doi.org/10.3390/metabo12080695)[[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/35893262)
- <span id="page-16-1"></span>87. Fortes, P.M.; Teles Filho, R.V.; Azevêdo, L.H.S.; Queiroz, V.C.J.; Costa, P. Inflammatory cytokines and lipid profile in children and adolescents with nephrotic syndrome receiving L. Plantarum: A randomized, controlled feasibility trial. *Rev. Assoc. Medica Bras. (1992)* **2020**, *66*, 1487–1492. [\[CrossRef\]](https://doi.org/10.1590/1806-9282.66.11.1487) [\[PubMed](https://www.ncbi.nlm.nih.gov/pubmed/33295397)]
- <span id="page-16-2"></span>88. Gu, X.C.; Luo, X.G.; Wang, C.X.; Ma, D.Y.; Wang, Y.; He, Y.Y.; Li, W.; Zhou, H.; Zhang, T.C. Cloning and analysis of bile salt hydrolase genes from Lactobacillus plantarum CGMCC No. 8198. *Biotechnol. Lett.* **2014**, *36*, 975–983. [\[CrossRef\]](https://doi.org/10.1007/s10529-013-1434-9)
- <span id="page-16-3"></span>89. Yin, Y.; Gao, L.; Lin, H.; Wu, Y.; Han, X.; Zhu, Y.; Li, J. Luteolin improves non-alcoholic fatty liver disease in db/db mice by inhibition of liver X receptor activation to down‑regulate expression of sterol regulatory element binding protein 1c. *Biochem. Biophys. Res. Commun.* **2017**, *482*, 720–726. [\[CrossRef\]](https://doi.org/10.1016/j.bbrc.2016.11.101)
- <span id="page-16-4"></span>90. Liu, Q.; Pan, R.; Ding, L.; Zhang, F.; Hu, L.; Ding, B.; Zhu, L.; Xia, Y.; Dou, X. Rutin exhibits hepatoprotective effects in a mouse model of non‑alcoholic fatty liver disease by reducing hepatic lipid levels and mitigating lipid‑induced oxidative injuries. *Int. Immunopharmacol.* **2017**, *49*, 132–141. [\[CrossRef\]](https://doi.org/10.1016/j.intimp.2017.05.026)
- <span id="page-16-5"></span>91. Wu, C.H.; Lin, M.C.; Wang, H.C.; Yang, M.Y.; Jou, M.J.; Wang, C.J. Rutin inhibits oleic acid induced lipid accumulation via reducing lipogenesis and oxidative stress in hepatocarcinoma cells. *J. Food Sci.* **2011**, *76*, T65–T72.[[CrossRef](https://doi.org/10.1111/j.1750-3841.2010.02033.x)][[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/21535797)
- <span id="page-16-6"></span>92. Phuwamongkolwiwat, P.; Suzuki, T.; Hira, T.; Hara, H. Fructooligosaccharide augments benefits of quercetin-3-O-β-glucoside on insulin sensitivity and plasma total cholesterol with promotion of flavonoid absorption in sucrose‑fed rats. *Eur. J. Nutr.* **2014**, *53*, 457–468. [\[CrossRef\]](https://doi.org/10.1007/s00394-013-0546-2) [\[PubMed](https://www.ncbi.nlm.nih.gov/pubmed/23784089)]
- <span id="page-16-7"></span>93. Mofidi, F.; Yari, Z.; Poustchi, H.; Merat, S.; Nourinayyer, B.; Malekzadeh, R.; Hekmatdoost, A. Effects of Synbiotics Supplementation in Lean Patients with Nonalcoholic Fatty Liver Disease: Study Protocol of a Pilot Randomized Double-blind Clinical Trial. *Arch. Iran. Med.* **2016**, *19*, 282–284. [\[PubMed](https://www.ncbi.nlm.nih.gov/pubmed/27041524)]
- 94. Laue, C.; Papazova, E.; Pannenbeckers, A.; Schrezenmeir, J. Effect of a Probiotic and a Synbiotic on Body Fat Mass, Body Weight and Traits of Metabolic Syndrome in Individuals with Abdominal Overweight: A Human, Double-Blind, Randomised, Controlled Clinical Study. *Nutrients* **2023**, *15*, 3039. [\[CrossRef](https://doi.org/10.3390/nu15133039)]
- <span id="page-16-8"></span>95. Ferolla, S.M.; Couto, C.A.; Costa-Silva, L.; Armiliato, G.N.; Pereira, C.A.; Martins, F.S.; Ferrari Mde, L.; Vilela, E.G.; Torres, H.O.; Cunha, A.S.; et al. Beneficial Effect of Synbiotic Supplementation on Hepatic Steatosis and Anthropometric Parameters, But Not on Gut Permeability in a Population with Nonalcoholic Steatohepatitis. *Nutrients* **2016**, *8*, 397.[[CrossRef\]](https://doi.org/10.3390/nu8070397)[[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/27367724)
- 96. Eslamparast, T.; Poustchi, H.; Zamani, F.; Sharafkhah, M.; Malekzadeh, R.; Hekmatdoost, A. Synbiotic supplementation in nonalcoholic fatty liver disease: A randomized, double‑blind, placebo‑controlled pilot study. *Am. J. Clin. Nutr.* **2014**, *99*, 535– 542.[[CrossRef\]](https://doi.org/10.3945/ajcn.113.068890)[[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/24401715)
- <span id="page-16-9"></span>97. Scorletti, E.; Afolabi, P.R.; Miles, E.A.; Smith, D.E.; Almehmadi, A.; Alshathry, A.; Moyses, H.E.; Clough, G.F.; Wright, M.; Patel, J.; et al. Design and rationale of the INSYTE study: A randomised, placebo controlled study to test the efficacy of a synbiotic on liver fat, disease biomarkers and intestinal microbiota in non‑alcoholic fatty liver disease. *Contemp. Clin. Trials* **2018**, *71*, 113–123. [\[CrossRef\]](https://doi.org/10.1016/j.cct.2018.05.010) [\[PubMed](https://www.ncbi.nlm.nih.gov/pubmed/29787859)]
- 98. Scorletti, E.; Afolabi, P.R.; Miles, E.A.; Smith, D.E.; Almehmadi, A.; Alshathry, A.; Childs, C.E.; Del Fabbro, S.; Bilson, J.; Moyses, H.E.; et al. Synbiotics Alter Fecal Microbiomes, But Not Liver Fat or Fibrosis, in a Randomized Trial of Patients With Nonalcoholic Fatty Liver Disease. *Gastroenterology* **2020**, *158*, 1597–1610.e7. [\[CrossRef](https://doi.org/10.1053/j.gastro.2020.01.031)] [\[PubMed](https://www.ncbi.nlm.nih.gov/pubmed/31987796)]
- <span id="page-16-10"></span>99. Sayari, S.; Neishaboori, H.; Jameshorani, M. Combined effects of synbiotic and sitagliptin versus sitagliptin alone in patients with nonalcoholic fatty liver disease. *Clin. Mol. Hepatol.* **2018**, *24*, 331–338. [\[CrossRef\]](https://doi.org/10.3350/cmh.2018.0006)[[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/29890569)
- 100. Mofidi, F.; Poustchi, H.; Yari, Z.; Nourinayyer, B.; Merat, S.; Sharafkhah, M.; Malekzadeh, R.; Hekmatdoost, A. Synbiotic supple‑ mentation in lean patients with non-alcoholic fatty liver disease: A pilot, randomised, double-blind, placebo-controlled, clinical trial. *Br. J. Nutr.* **2017**, *117*, 662–668.[[CrossRef\]](https://doi.org/10.1017/S0007114517000204)
- 101. Bakhshimoghaddam, F.; Shateri, K.; Sina, M.; Hashemian, M.; Alizadeh, M. Daily Consumption of Synbiotic Yogurt Decreases Liver Steatosis in Patients with Nonalcoholic Fatty Liver Disease: A Randomized Controlled Clinical Trial. *J. Nutr.* **2018**, *148*, 1276–1284. [\[CrossRef](https://doi.org/10.1093/jn/nxy088)]
- <span id="page-17-0"></span>102. Abhari, K.; Saadati, S.; Yari, Z.; Hosseini, H.; Hedayati, M.; Abhari, S.; Alavian, S.M.; Hekmatdoost, A. The effects of Bacillus coagulans supplementation in patients with non‑alcoholic fatty liver disease: A randomized, placebo‑controlled, clinical trial. *Clin. Nutr. ESPEN* **2020**, *39*, 53–60. [\[CrossRef\]](https://doi.org/10.1016/j.clnesp.2020.06.020)[[PubMed](https://www.ncbi.nlm.nih.gov/pubmed/32859329)]
- <span id="page-17-1"></span>103. Behrouz, V.; Jazayeri, S.; Aryaeian, N.; Zahedi, M.J.; Hosseini, F. Effects of Probiotic and Prebiotic Supplementation on Leptin, Adiponectin, and Glycemic Parameters in Non‑alcoholic Fatty Liver Disease: A Randomized Clinical Trial. *Middle East J. Dig. Dis.* **2017**, *9*, 150–157. [\[CrossRef\]](https://doi.org/10.15171/mejdd.2017.66)
- <span id="page-17-2"></span>104. Malaguarnera, M.; Vacante, M.; Antic, T.; Giordano, M.; Chisari, G.; Acquaviva, R.; Mastrojeni, S.; Malaguarnera, G.; Mistretta, A.; Li Volti, G.; et al. Bifidobacterium longum with fructo-oligosaccharides in patients with non alcoholic steatohepatitis. *Dig. Dis. Sci.* **2012**, *57*, 545–553.[[CrossRef\]](https://doi.org/10.1007/s10620-011-1887-4)
- <span id="page-17-3"></span>105. Shavakhi, A.; Minakari, M.; Firouzian, H.; Assali, R.; Hekmatdoost, A.; Ferns, G. Effect of a Probiotic and Metformin on Liver Aminotransferases in Non‑alcoholic Steatohepatitis: A Double Blind Randomized Clinical Trial. *Int. J. Prev. Med.* **2013**, *4*, 531–537.
- <span id="page-17-4"></span>106. Abdel Monem, S.M. Probiotic Therapy in Patients with Nonalcoholic Steatohepatitis in Zagazig University Hospitals. *Euroasian J. Hepato‑Gastroenterol.* **2017**, *7*, 101–106. [\[CrossRef\]](https://doi.org/10.5005/jp-journals-10018-1226)
- <span id="page-17-5"></span>107. Aller, R.; De Luis, D.A.; Izaola, O.; Conde, R.; Gonzalez Sagrado, M.; Primo, D.; De La Fuente, B.; Gonzalez, J. Effect of a probiotic on liver aminotransferases in nonalcoholic fatty liver disease patients: A double blind randomized clinical trial. *Eur. Rev. Med. Pharmacol. Sci.* **2011**, *15*, 1090–1095. [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/22013734)
- <span id="page-17-6"></span>108. Aron‑Wisnewsky, J.; Vigliotti, C.; Witjes, J.; Le, P.; Holleboom, A.G.; Verheij, J.; Nieuwdorp, M.; Clément, K. Gut microbiota and human NAFLD: Disentangling microbial signatures from metabolic disorders. *Nat. Reviews. Gastroenterol. Hepatol.* **2020**, *17*, 279–297.[[CrossRef](https://doi.org/10.1038/s41575-020-0269-9)][[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/32152478)
- 109. Chen, J.; Vitetta, L.J.H. Could butyrate be incorporated with Farnesoid X receptor agonist cilofexor to enhance primary sclerosing cholangitis treatment? *Hepatology* **2020**, *72*, 1497–1498. [\[CrossRef\]](https://doi.org/10.1002/hep.31269)
- 110. Chen, J.; Vitetta, L. Butyrate in Inflammatory Bowel Disease Therapy. *Gastroenterology* **2020**, *158*, 1511. [\[CrossRef](https://doi.org/10.1053/j.gastro.2019.08.064)]
- 111. Boursier, J.; Mueller, O.; Barret, M.; Machado, M.; Fizanne, L.; Araujo-Perez, F.; Guy, C.D.; Seed, P.C.; Rawls, J.F.; David, L.A.; et al. The severity of nonalcoholic fatty liver disease is associated with gut dysbiosis and shift in the metabolic function of the gut microbiota. *Hepatology* **2016**, *63*, 764–775. [\[CrossRef\]](https://doi.org/10.1002/hep.28356) [\[PubMed](https://www.ncbi.nlm.nih.gov/pubmed/26600078)]
- <span id="page-17-7"></span>112. Saltzman, E.T.; Palacios, T.; Thomsen, M.; Vitetta, L. Intestinal Microbiome Shifts, Dysbiosis, Inflammation, and Non‑alcoholic Fatty Liver Disease. *Front. Microbiol.* **2018**, *9*, 61.[[CrossRef](https://doi.org/10.3389/fmicb.2018.00061)][[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/29441049)
- <span id="page-17-8"></span>113. Le Roy, T.; Llopis, M.; Lepage, P.; Bruneau, A.; Rabot, S.; Bevilacqua, C.; Martin, P.; Philippe, C.; Walker, F.; Bado, A.; et al. Intestinal microbiota determines development of non‑alcoholic fatty liver disease in mice. *Gut* **2013**, *62*, 1787–1794. [\[CrossRef](https://doi.org/10.1136/gutjnl-2012-303816)]
- <span id="page-17-9"></span>114. Chiu, C.C.; Ching, Y.H.; Li, Y.P.; Liu, J.Y.; Huang, Y.T.; Huang, Y.W.; Yang, S.S.; Huang, W.C.; Chuang, H.L. Nonalcoholic Fatty Liver Disease Is Exacerbated in High‑Fat Diet‑Fed Gnotobiotic Mice by Colonization with the Gut Microbiota from Patients with Nonalcoholic Steatohepatitis. *Nutrients* **2017**, *9*, 1220.[[CrossRef](https://doi.org/10.3390/nu9111220)]
- <span id="page-17-10"></span>115. Lorenzo‑Zúñiga, V.; Bartolí, R.; Planas, R.; Hofmann, A.F.; Viñado, B.; Hagey, L.R.; Hernández, J.M.; Mañé, J.; Alvarez, M.A.; Ausina, V.; et al. Oral bile acids reduce bacterial overgrowth, bacterial translocation, and endotoxemia in cirrhotic rats. *Hepatology* **2003**, *37*, 551–557.[[CrossRef](https://doi.org/10.1053/jhep.2003.50116)] [\[PubMed](https://www.ncbi.nlm.nih.gov/pubmed/12601352)]
- 116. Ogata, Y.; Nishi, M.; Nakayama, H.; Kuwahara, T.; Ohnishi, Y.; Tashiro, S. Role of bile in intestinal barrier function and its inhibitory effect on bacterial translocation in obstructive jaundice in rats. *J. Surg. Res.* **2003**, *115*, 18–23. [\[CrossRef\]](https://doi.org/10.1016/S0022-4804(03)00308-1) [\[PubMed](https://www.ncbi.nlm.nih.gov/pubmed/14572768)]
- <span id="page-17-11"></span>117. Wang, X.; Xia, J.; Jiang, C.J.L.R. Role of gut microbiota in the development of non‑alcoholic fatty liver disease. *Liver Res.* **2019**, *3*, 25–30.[[CrossRef](https://doi.org/10.1016/j.livres.2019.01.006)]
- <span id="page-17-12"></span>118. Schumacher, J.D.; Guo, G.L. Pharmacologic Modulation of Bile Acid-FXR-FGF15/FGF19 Pathway for the Treatment of Nonalcoholic Steatohepatitis. *Handb. Exp. Pharmacol.* **2019**, *256*, 325–357.[[CrossRef](https://doi.org/10.1007/164_2019_228)][[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/31201553)
- 119. Tomlinson, E.; Fu, L.; John, L.; Hultgren, B.; Huang, X.; Renz, M.; Stephan, J.P.; Tsai, S.P.; Powell‑Braxton, L.; French, D.; et al. Transgenic mice expressing human fibroblast growth factor-19 display increased metabolic rate and decreased adiposity. *Endocrinology* **2002**, *143*, 1741–1747.[[CrossRef\]](https://doi.org/10.1210/endo.143.5.8850)
- <span id="page-17-13"></span>120. Alvarez‑Sola, G.; Uriarte, I.; Latasa, M.U.; Fernandez‑Barrena, M.G.; Urtasun, R.; Elizalde, M.; Barcena‑Varela, M.; Jiménez, M.; Chang, H.C.; Barbero, R.; et al. Fibroblast growth factor 15/19 (FGF15/19) protects from diet-induced hepatic steatosis: Development of an FGF19‑based chimeric molecule to promote fatty liver regeneration. *Gut* **2017**, *66*, 1818–1828.[[CrossRef\]](https://doi.org/10.1136/gutjnl-2016-312975)
- <span id="page-17-14"></span>121. Chiang, J.Y.; Pathak, P.; Liu, H.; Donepudi, A.; Ferrell, J.; Boehme, S. Intestinal Farnesoid X Receptor and Takeda G Protein Couple Receptor 5 Signaling in Metabolic Regulation. *Dig. Dis.* **2017**, *35*, 241–245. [\[CrossRef](https://doi.org/10.1159/000450981)]
- <span id="page-17-15"></span>122. Onoviran, O.F.; Li, D.; Toombs Smith, S.; Raji, M.A. Effects of glucagon-like peptide 1 receptor agonists on comorbidities in older patients with diabetes mellitus. *Ther. Adv. Chronic Dis.* **2019**, *10*, 2040622319862691. [\[CrossRef\]](https://doi.org/10.1177/2040622319862691)
- <span id="page-17-16"></span>123. Bäckhed, F.; Manchester, J.K.; Semenkovich, C.F.; Gordon, J.I. Mechanisms underlying the resistance to diet-induced obesity in germ‑free mice. *Proc. Natl. Acad. Sci. USA* **2007**, *104*, 979–984.[[CrossRef](https://doi.org/10.1073/pnas.0605374104)] [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/17210919)
- <span id="page-17-17"></span>124. Bäckhed, F.; Ding, H.; Wang, T.; Hooper, L.V.; Koh, G.Y.; Nagy, A.; Semenkovich, C.F.; Gordon, J.I. The gut microbiota as an environmental factor that regulates fat storage. *Proc. Natl. Acad. Sci. USA* **2004**, *101*, 15718–15723. [\[CrossRef\]](https://doi.org/10.1073/pnas.0407076101) [\[PubMed](https://www.ncbi.nlm.nih.gov/pubmed/15505215)]
- <span id="page-18-0"></span>125. Okubo, H.; Sakoda, H.; Kushiyama, A.; Fujishiro, M.; Nakatsu, Y.; Fukushima, T.; Matsunaga, Y.; Kamata, H.; Asahara, T.; Yoshida, Y.; et al. Lactobacillus casei strain Shirota protects against nonalcoholic steatohepatitis development in a rodent model. *Am. J. Physiol. Gastrointest. Liver Physiol.* **2013**, *305*, G911–G918. [\[CrossRef\]](https://doi.org/10.1152/ajpgi.00225.2013)[[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/24113768)
- <span id="page-18-1"></span>126. Nguyen, T.D.; Kang, J.H.; Lee, M.S. Characterization of Lactobacillus plantarum PH04, a potential probiotic bacterium with cholesterol‑lowering effects. *Int. J. Food Microbiol.* **2007**, *113*, 358–361. [\[CrossRef\]](https://doi.org/10.1016/j.ijfoodmicro.2006.08.015)
- <span id="page-18-2"></span>127. Zhao, Z.; Chen, L.; Zhao, Y.; Wang, C.; Duan, C.; Yang, G.; Niu, C.; Li, S. Lactobacillus plantarum NA136 ameliorates nonalcoholic fatty liver disease by modulating gut microbiota, improving intestinal barrier integrity, and attenuating inflammation. *Appl. Microbiol. Biotechnol.* **2020**, *104*, 5273–5282.[[CrossRef](https://doi.org/10.1007/s00253-020-10633-9)]

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