https://doi.org/10.1093/humrep/dead172 Advance Access Publication Date: August 26, 2023

Original Article

Reproductive epidemiology

A prospective preconception cohort study of the association between Mycoplasma genitalium and fecundability in Kenyan women trying to conceive

Erica M. Lokken 1.*, Emmanuel Kabare², Brenda Oyaro², Maureen Nyaigero², John Kinuthia¹,³,⁴, Walter Jaoko², Kishor Mandaliya⁵, R. Scott McClelland¹,²,6,7, and Lisa E. Manhart¹,6

*Correspondence address. Department of Global Health, University of Washington, Hans Rosling Center, 3980 15th Ave NE, Seattle, WA 98105, USA. E-mail: elokken@uw.edu (6) https://orcid.org/0000-0002-3285-4751

ABSTRACT

STUDY QUESTION: Is Mycoplasma genitalium-infection associated with reduced fecundability?

SUMMARY ANSWER: Preconception M. *genitalium*-infection was associated with 27% lower fecundability though confidence intervals were wide, and the association between M. *genitalium* and fecundability may be dependent on concurrent bacterial vaginosis (BV).

WHAT IS KNOWN ALREADY: *M. genitalium* has been associated with cervicitis, pelvic inflammatory disease, infertility, and preterm birth, but the extent to which *M. genitalium* is causally related to adverse reproductive sequelae in women is debated.

STUDY DESIGN, SIZE, DURATION: Kenyan women enrolled in a prospective preconception cohort provided vaginal fluid specimens and underwent monthly pregnancy testing. Stored samples from 407 women who had been trying to conceive for \leq 6 months were tested for M. genitalium using a nucleic acid amplification test.

PARTICIPANTS/MATERIALS, SETTING, METHODS: Data on first day of last menstrual period, sexual behavior, pregnancy status, and vaginal specimens were collected at monthly preconception visits. The association between *M. genitalium* detected at the visit prior to each pregnancy test and fecundability was estimated using discrete time proportional probabilities models. Secondary analyses explored the influence of concurrent BV on the association between *M. genitalium* and fecundability.

MAIN RESULTS AND THE ROLE OF CHANCE: The 407 participants experienced 1220 menstrual cycles and 213 pregnancies. The prevalence of M. genitalium at enrollment was 7.7%. After adjustment for age, frequency of condomless sex in the last 4 weeks, and study site, M. genitalium was associated with a 27% lower fecundability, but confidence intervals were wide (adjusted fecundability ratio (aFR) 0.73, 95% CI 0.44, 1.23). In secondary analyses, when compared to cycles without M. genitalium or BV at the visit prior, women with both M. genitalium and BV at the visit prior had a 51% lower fecundability (aFR = 0.49, 95% CI 0.22, 1.09) whereas there was no association of M. genitalium alone (aFR = 0.98 (95% CI 0.54, 1.76)), and a smaller reduction in fecundability for women with BV only (aFR = 0.80 (95% CI 0.60, 1.07)).

LIMITATIONS, REASONS FOR CAUTION: Results should be interpreted cautiously given the relatively low prevalence of M. genitalium and wide confidence intervals.

WIDER IMPLICATIONS OF THE FINDINGS: In this cohort of Kenyan women trying to conceive, the association between *M. genitalium* and fecundability was influenced by concurrent BV status, suggesting there may be a synergistic effect of *M. genitalium* and BV on fecundability.

STUDY FUNDING/COMPETING INTEREST(S): This work was supported by a National Institutes of Health grant (NICHD R01 HD087346-RSM). R.S.M. received additional support for mentoring (NICHD K24 HD88229). E.M.L. was supported by pre- and post-doctoral fellowships (NIAID T32 AI07140, NICHD F32 HD100202). Data collection and management were completed using REDCap electronic data capture tools hosted at the University of Washington's Institute of Translational Health Science supported by grants from NCATS/NIH (UL1 TR002319). The content of this paper is solely the responsibility of the authors and does not necessarily represent the official views of the National Institutes of Health. R.S.M. receives research funding, paid to the University of Washington, from Hologic Corporation and consulting fees from Lupin Pharmaceuticals. L.E.M. receives research funding and material for research studies, paid to the University of Washington, from Hologic Corporation and Nabriva Therapeutics, travel support from

¹Department of Global Health, University of Washington, Seattle, WA, USA

²Department of Medical Microbiology, University of Nairobi, Nairobi, Kenya

³Department of Obstetrics & Gynaecology, Kenyatta National Hospital, Nairobi, Kenya

⁴Research and Programs, Kenyatta National Hospital, Nairobi, Kenya

⁵Pathcare, Mombasa, Kenya

⁶Department of Epidemiology, University of Washington, Seattle, WA, USA

⁷Department of Medicine, University of Washington, Seattle, WA, USA

Hologic, and consulting fees from Health Advances. E.M.L.'s contributions to this study primarily occurred while affiliated with the University of Washington; at the time of submission, E.M.L. was an employee of and holds stock or stock grants for AbbVie, Inc. The other authors have no conflicts of interest.

TRIAL REGISTRATION NUMBER: N/A.

Keywords: Mycoplasma genitalium / fecundability / conception / time-to-pregnancy / bacterial vaginosis / infertility / vaginal microhiota

Introduction

Infertility affects ~15% of couples and can lead to significant financial and emotional burden (Evers, 2002). Public health approaches to preventing infertility emphasize preconception health, including screening and treating women for Chlamydia trachomatis and Neisseria gonorrhoeae (US Preventative Services Task Force, 2021). National C. trachomatis and N. gonorrhoeae screening programs were implemented in part due to evidence that these infections reduce reproductive potential (Paavonen et al., 2008).

Mycoplasma genitalium, a sexually transmitted bacterium, causes male urethritis (Taylor-Robinson and Jensen, 2011). Whether M. genitalium causes adverse reproductive sequelae in women is debated, despite studies demonstrating associations between M. genitalium and cervicitis, PID, infertility, and preterm birth (Lis et al., 2015; Workowski et al., 2021). A 2015 metaanalysis demonstrated >2-fold increased odds of PID (OR 2.14, 95% CI 1.31-3.49) and infertility (OR 2.43, 95% CI 0.93-6.24) among women with M. genitalium (Lis et al., 2015). However, few of the infertility studies were prospective and the effect size was imprecise. More recently, in a prospective cohort study of women discontinuing intrauterine devices (IUD) to conceive, lifetime exposure to M. genitalium detected via serology at enrollment was associated with a longer time-to-pregnancy (Hazard Ratio 0.76, 95% CI 0.58-0.99) (Peipert et al., 2021).

The objective of this study was to assess the association between M. genitalium infection and fecundability, the per-menstrual cycle probability of pregnancy, using data from a prospective preconception cohort of Kenyan women.

Materials and methods

Study design and population

Data from the Microbiota and Preterm Birth (MPTB) Study, which enrolled women planning pregnancies in Nairobi and Mombasa, Kenya between 18 April 2017 and 18 March 2020, were used. The MPTB study was designed to assess the association between periconceptual vaginal microbiota and spontaneous preterm birth (Lokken et al., 2020). Women enrolled in the MPTB study were trying to conceive, HIV-seronegative, 18-45 years old, had menstruated in the 3 months before enrollment or recently discontinued hormonal contraception that can influence menstruation. Women who reported medical history potentially increasing preterm birth risk, had used antibiotics in the last 4 weeks, or had ever sought care for infertility were excluded. Participants recently using depot-medroxyprogesterone acetate (DMPA) injection were not eligible until 3 months after their last injection.

Women eligible for the fecundability sub-cohort were participants contributing ≥1 menstrual cycle, without reproductive co-morbidities (ectopic pregnancy, polycystic ovary syndrome, endometriosis, in-hospital treatment of PID), and trying to conceive for ≤6 menstrual cycles prior to enrollment. For this analysis, women who conceived or were lost to follow-up prior to the initiation of M. genitalium sampling, which started in 2018 (1 year after study launch), were excluded as they did not have

M. genitalium results for analysis. Women whose first M. genitalium sample was available at >6 menstrual cycles of total conception attempt time were also excluded to reduce potential bias associated with unobserved cycles (Weinberg et al., 1994; Schisterman et al., 2013).

Ethical approval

The study procedures were approved by the institutional review boards of Kenyatta National Hospital-University of Nairobi and the University of Washington. Participants provided written informed consent.

Clinical and laboratory procedures

Study procedures have been described (Lokken et al., 2020). At enrollment, participants completed an interview on demographics, medical history, sexual behavior, and reproductive history including recent contraceptive use, first day of last menstrual period (LMP), and the number of menstrual cycles attempting to conceive before enrollment ('pre-enrollment conception attempt time'). Clinicians collected vaginal swabs during a pelvic examination. Participants returned for up to 6 monthly preconception visits, where they completed an interim interview, underwent urine pregnancy testing (One Step Urine Pregnancy Test, EROVITA; Eros Ventures Ltd, Nairobi, Kenya), self-collected vaginal swabs, and reported the first day of their most recent LMP. Beginning in July 2018, participants who missed a preconception visit reported all first days of menstrual cycles occurring between their prior and current attended preconception visit ('reported interim menstrual cycles'). Due to a potential for delayed return to fertility with DMPA use, participants who discontinued DMPA 3-6 months prior to enrollment were eligible for an additional 3 months of preconception follow-up (9 months total) (Yland et al., 2020). If participants had genital symptoms suggestive of vaginal infections, including BV, they were treated per Kenyan syndromic management guidelines (National AIDS & STI Control Programme of Kenya, 2018).

Enrollment specimens were tested for N. gonorrhoeae, C. trachomatis, and T. vaginalis (Aptima Combo-2 CT/NG Detection System, Aptima T. vaginalis assay; Hologic, Inc). Treatment for these infections was provided at the participant's next visit if they were not covered by syndromic treatment at enrollment. Bacterial vaginosis (BV) was assessed at enrollment and preconception visits by Gram staining and scoring of vaginal fluid samples using the criteria of Nugent and Hillier (Nugent et al., 1991).

Beginning in May 2018, an additional vaginal specimen was collected at enrollment and self-collected at each monthly preconception visit for M. genitalium testing. Because antimicrobial resistance is selected for during treatment and the link between M. genitalium and adverse reproductive health outcomes in women remains debated, screening for and treating asymptomatic M. genitalium infections is not recommended (Australian Sexual Health Alliance, 2018; Soni et al., 2019; Workowski et al., 2021; Jensen et al., 2022). Therefore, vaginal samples were stored at -80°C after collection and tested for M. genitalium in 2020 and 2021 (Aptima M. genitalium assay, Hologic, Inc). Women whose

last sample in the study was positive were offered immediate retesting and treatment if the re-test was positive.

Data analysis

First days of LMP reported at preconception visits and urine pregnancy test results were used to calculate the number of menstrual cycles for participants (Lokken et al., 2021). Because pregnancy testing occurred at different times in the menstrual cycle based on participants' visit schedules, cycles were considered negative for pregnancy only if the pregnancy test was negative and the participant reported a subsequent LMP at their next visit. Median time-to-pregnancy and cumulative incidence of pregnancy were estimated using Kaplan-Meier methods.

To estimate the association between M. genitalium-infection and fecundability, we used proportional probabilities models (log-binomial generalized linear models with robust standard errors) to generate fecundability ratios (FR) and 95% CI (Weinberg et al., 1989, 1994). The models included an indicator variable for menstrual cycle at risk and delayed entry for prior trying time (Weinberg et al., 1994; Schisterman et al., 2013), defined for this analysis as the total number of menstrual cycles of trying time at the first M. genitalium result for this analysis (number of preenrollment cycles trying to conceive + prospective cycles during follow-up prior to first sample collected for M. genitalium testing). Censoring criteria included self-reported initiation of biomedical infertility treatment (n=3), participant withdrawal or loss to follow-up (n = 76), no conception by end of eligible preconception follow-up (n = 79), and end of in-person study visits due to the COVID-19 pandemic (n = 36). We imputed missing M. genitalium and time-varying covariate data (i.e. frequency of condomless sex, BV) due to missed preconception visits (n = 103) and missingness for time-varying covariates at attended preconception visits (<0.03% missing) using data from the last visit carried forward.

The primary exposure was M. genitalium detected at the visit prior to each pregnancy test, modeled as a time-varying exposure. In secondary analysis, we assessed M. genitalium status at enrollment as a time-independent measure to evaluate whether a single measure of M. genitalium during preconception was associated with reduced fecundability. We also evaluated the potential synergistic effect of M. genitalium and BV at the visit prior to pregnancy with a four-level categorical variable: neither M. genitalium nor BV, M. genitalium positive only, BV positive only, positive for both M. genitalium and BV. To accompany this analysis, we estimated the relative excess risk due to interaction (RERI). Because FRs <1 are interpreted as reduced fecundability and >1 as increased fecundability, we have interpreted the RERI similarly with an RERI <1 demonstrating positive interaction/more than additivity.

We adjusted a priori for participants' age (<25, 25-29, 30-34, 35–39, 40–45), frequency of condomless sex in the prior 4 weeks (continuous), and study site based on existing literature, sitespecific differences in demographic characteristics, and consideration of potential causal associations between variables. Because only a small proportion of participants were prescribed antibiotics with activity against M. genitalium or BV (doxycycline: 2.6% of visits, azithromycin: 1.3% of visits, metronidazole: 2.1% of visits), we did not adjust for antibiotic use.

We conducted sensitivity analyses excluding participants with reproductive co-morbidities or history that may be suggestive of potential sub-fecundity. First, we conducted the primary analysis excluding women with N. gonorrhoeae or C. trachomatis at enrollment. Second, we additionally excluded those with T. vaginalis or PID diagnosis at enrollment, any history of treatment for N.

gonorrhoeae, C. trachomatis, T. vaginalis, syphilis, or PID, self-report of fibroids or unknown uterine abnormality, any DMPA use within 6 months of enrollment, or a partner living with HIV. We conducted two additional sensitivity analyses of the primary analysis including (i) additional adjustment for BV and (ii) excluding the participant-reported interim menstrual cycles to assess whether their inclusion influenced results.

Results

Of the 701 participants enrolled, 98 (14.0%) were excluded from this fecundability analysis due to reproductive co-morbidities or more than six cycles of pre-enrollment conception attempt time and 92 (13%) were excluded solely for contributing no follow-up time. An additional 104 (14.8%) were excluded because they conceived or were lost to follow-up before M. genitalium sampling was initiated, leaving 407 (58.1%) eligible participants (Supplementary Fig. S1). While there were some minor differences in characteristics comparing those eligible to those excluded for missing M. genitalium results (Supplementary Table S1), time-to-pregnancy was similar (logrank P=0.51). Eligible participants attended 1117 study visits and experienced 1220 menstrual cycles, of which 103 (8.4%) were reported after missed preconception visits. There were 213 incident first pregnancies. The median time-topregnancy was four cycles (IQR 2, 10) with a cumulative six-cycle pregnancy rate of 67.4% (95% CI 61.8-72.9) and a 12-cycle pregnancy rate of 82.8% (95% CI 73.7-90.1).

Participants' median age was 29 years (IQR 25, 34). Most had finished secondary school (57.7%, n=235), experienced a prior pregnancy (92.9%, n = 378), and reported zero pre-enrollment cycles trying to conceive (71.0%, n = 289) (Table 1). At enrollment, 36.4% (148/407) had BV, 8.7% (35/407) had C. trachomatis infection, 1.0% (4/403) had N. gonorrhoeae infection, 1.2% (5/403) had T. vaginalis infection, and 7.7% (30/389) had M. genitalium infection. During follow-up, M. genitalium was detected at 7.8% (87/1114) of

Eight percent (98/1220) of menstrual cycles were considered exposed to M. genitalium. Mycoplasma genitalium infection at the visit prior to pregnancy testing was associated with a 19% lower fecundability in unadjusted analysis (FR 0.81, 95% CI 0.48-1.36) (Table 2). After adjustment for age, frequency of condomless sex in the last 4 weeks, and study site, M. genitalium was associated with a 27% lower fecundability, but confidence intervals remained wide (adjusted FR (aFR) 0.73, 95% CI 0.44-1.23). Results were similar for sensitivity analyses that additionally adjusted for BV; excluded participant-reported interim menstrual cycles; and excluded additional participants with potential subfecundity (Supplementary Table S2). When considering M. genitalium at the first visit only, M. genitalium was associated with a 24% lower fecundability ((aFR) 0.76, 95% CI 0.47-1.22).

In the analysis assessing potential synergy between M. genitalium and BV at the visit prior to pregnancy, compared to cycles without M. genitalium or BV, the aFR was 0.98 (95% CI 0.54, 1.76) for women with M. genitalium only, 0.80 (95% CI 0.60, 1.07) for women with BV only, and 0.49 (95% CI 0.22, 1.09) for women with both M. genitalium and BV (Table 2). The RERI estimate was -0.29(95% CI -1.05, 0.47).

Discussion

In this cohort of Kenyan women trying to conceive, M. genitalium infection at the visit prior to pregnancy testing was associated with a non-significant 27% lower per-menstrual cycle probability

Table 1. Enrollment characteristics for 407 Kenyan women planning pregnancies and eligible for the *M. genitalium* and fecundability analysis.

Characteristic	N	Total (N = 407)	
Demographic and partnership characteris			
Cohort	407	200	(70.00/)
Nairobi Mombasa		286 121	(70.3%) (29.7%)
Age (years)	407	121	(23.7 70)
<25	107	84	(20.6%)
25–29		136	(33.4%)
30–34		105	(25.8%)
35–39		69	(17.0%)
40–45		13	(3.2%)
Education level	407	0.4	(7.60/)
<8 years		31	(7.6%)
8–11 years 12–15 years		141 173	(34.6%) (42.5%)
≥16 years		62	(15.2%)
Monthly household income (KSh)	404	02	(13.270)
<2500	101	17	(4.2%)
2500–10 000		138	(34.2%)
10 000–30 000		180	(44.6%)
30 000–75 000		45	(11.1%)
>75 000		24	(5.9%)
Partner's HIV-serostatus	406		, ,
HIV-seronegative		317	(78.1%)
HIV-seropositive		18	(4.4%)
Unknown		71	(17.5%)
Reproductive history	407		
Most recent contraceptive method ^a	407	110	(20.00/)
None		118	(29.0%)
Condoms Oral contracentive pill		19 7	(4.7%) (1.7%)
Oral contraceptive pill DMPA injectable		16	(3.9%)
Copper intrauterine device		97	(23.8%)
Implant		147	(36.1%)
Other		3	(0.7%)
Ever pregnant	407	378	(92.9%)
History of miscarriage	378	76	(20.1%)
Parous	407	366	(89.9%)
Number of menstrual cycles of prior	407		
conception attempt time at enrollmentb			
0		289	(71.0%)
1–3		70	(17.2%)
4–6	4 4	48	(11.8%)
Sexual behavior and vaginal washing in la			(2.0.00/)
Any vaginal washing	407	149	(36.6%)
Frequency of condomless sex No condomless sex	407	29	(7 1%)
1–4		132	(7.1%) (32.4%)
5–8		97	(23.8%)
>9		149	(36.6%)
Median number of condomless		6	(3–12)
sex acts (Q1–Q3)			(- /
STI, BV, and vaginal symptoms			
History of STI ^c	407	4	(1.0%)
N. gonorrhoeae	403	4	(1.0%)
C. trachomatis	403	35	(8.7%)
T. vaginalis	403	5	(1.2%)
M. genitalium ^d	389	30	(7.7%)
BV (Nugent ≥7)	407	148	(36.4%)
Abnormal vaginal dischargee		07	10 4011
Vaginal/vulvar itching ^e	407 407	37 37	(9.1%) (9.1%)

^a Women reporting OCP, contraceptive implant, or copper IUD discontinuation >2 months before enrollment or a last DMPA injection >6 months before enrollment were included in the 'none' category.

of pregnancy and this association appeared to be influenced by concurrent BV status. In exploratory analysis, M. genitalium alone showed no association with fecundability and BV alone showed a modest association with fecundability, but women with concurrent BV and M. genitalium had 51% lower fecundability compared to cycles with neither condition. While this suggests a synergistic relationship between M. genitalium and BV, results should be interpreted cautiously given the wide confidence intervals (aFR 0.49, 95% CI 0.22–1.09).

There are limited data on M. genitalium or BV and fecundability. A recent prospective cohort study of women discontinuing contraception to conceive in the US assessed lifetime exposure to M. genitalium and time-to-pregnancy (Peipert et al., 2021). In a multivariable piecewise exponential proportional hazards model assessing IUD use and time-to-pregnancy that included M. genitalium seropositivity and sociodemographics as covariates, lifetime exposure to M. genitalium was associated with a 24% longer timeto-pregnancy (adjusted Hazard Ratio 0.76, 95% CI 0.58-1.00). Chlamydia trachomatis, in comparison, was not associated with time-to-pregnancy. This effect size was similar to the 27% reduced fecundability for M. genitalium-infection measured by NAAT in our primary analysis. Prior analyses of this Kenyan cohort demonstrated 17% lower fecundability among women with BV at the visit prior to pregnancy testing and 43% lower fecundability associated with persistent BV (Lokken et al., 2021). Given the delay in launching sample collection for M. genitalium testing, this analysis includes a slightly different sub-cohort. Nonetheless, the 20% lower fecundability associated with recent BV alone here is consistent with the original result (Lokken et al., 2021).

Studies of M. genitalium and reproductive outcomes in females have focused on PID and infertility. Although most of these were limited by cross-sectional designs, inclusion of potentially subfertile women, and unclear timing of M. genitalium infection (Taylor et al., 2013; Lis et al., 2015), the limited prospective data suggest that both BV and M. genitalium infection are linked to female infertility. Sero-epidemiologic studies that adjusted for antibody to C. trachomatis observed 4- to 5-fold increased risk of infertility associated with prior M. genitalium infection (Clausen et al., 2001; Svenstrup et al., 2008). In a prospective study of women with clinically suspected PID, cervical or endometrial M. genitalium infection was associated with infertility (adjusted relative risk 1.4, 95% CI 0.6-2.9), though confidence intervals were wide (Haggerty et al., 2008). In the same study, women without N. gonorrhoeae or C. trachomatis who had four BV-associated bacteria (Sneathia sanguinegens, Sneathia amnii, Fannyhessea vaginae (previously known as Atopobium vaginae), and BVAB1) at the cervix or endometrium at baseline had a 3.4-fold increased risk of infertility compared to participants without these bacteria (Haggerty et al., 2016). As the effects of infections on fertility likely represent a continuum ranging from reduced fecundability to clinically defined infertility (e.g. tubal factor infertility) these earlier studies of infertility provide useful background for consideration when interpreting our fecundability results.

Our observation that the greatest reduction in fecundability occurred in women with both *M. genitalium* and BV is consistent with studies suggesting potential synergy. BV is often present in women with *M. genitalium* infection (Nye et al., 2020), BV may enhance susceptibility to *M. genitalium* (Lokken et al., 2017), and periodic presumptive therapy active against BV was associated with decreased incidence of *M. genitalium* (Balkus et al., 2016). These data suggest that *M. genitalium* and BV bacteria may enhance one another's growth, survival, or pathogenicity. Furthermore,

^b Women reporting >6 cycles of conception attempt time prior to enrollment were excluded for this analysis.

^c Self-reported history of syphilis, chlamydia, gonorrhea, and/or trichomoniasis.

 $^{^{}m d}$ N = 18 participants had their first M. genitalium sample collected at a subsequent preconception follow-up visit.

e Self-report.

KSh, Kenyan shillings; DMPA, depot-medroxyprogesterone acetate; (Q1–Q3) Quartile 1–Quartile 3; STI, sexually transmitted infection; BV, bacterial vaginosis.

Table 2. Unadjusted and adjusted associations between M. genitalium (MG) and fecundability among 407 Kenyan women trying to conceive.

Exposure	Menstrual cycles			Pregnancies			Unadjusted		Adjusted ^a	
	N	n (%)	exposed	N	n (%)	exposed	FI	R (95% CI)	FI	R (95% CI)
Primary analysis										
MG at visit prior ^b	1220	98	(8.0)	213	14	(6.6)	0.81	(0.48, 1.36)	0.73	(0.44, 1.23)
Secondary analyses			,			,		, ,		,
MG status at enrollment ^c	1193	101	(8.5)	205	14	(6.8)	0.81	(0.50, 1.30)	0.76	(0.47, 1.22)
MG and BV status at visit priorb	1220		, ,	213		. ,		,		,
MG-, BV-		768	(63.0)		143	(67.1)	REF		REF	
MG+, BV-		43	(3.5)		8	(3.8)	1.04	(0.57, 1.92)	0.98	(0.54, 1.76)
MG-, BV+		354	(29.0)		56	(26.3)	0.84	(0.63, 1.11)	0.80	(0.60, 1.07)
MG+, BV+		55	(4.5)		6	(2.8)	0.57	(0.25, 1.27)	0.49	(0.22, 1.09)

^a A priori inclusion of age, study site, and frequency of condomless sex.

BV, bacterial vaginosis.

successful metronidazole-mediated treatment of BV may facilitate M. genitalium clearance. A recent randomized trial assessed the effect of adding metronidazole, which targets anaerobes commonly present in women with BV, to a standard PID treatment regimen. After 30 days, the proportion of cervical M. genitalium infections was lower in the treatment arm with metronidazole versus without (4.4% versus 14.4%, P=0.04) and endometrial M. genitalium was less common in the metronidazole arm compared to the standard therapy arm (1.2% versus 4.4%, P=0.4) (Wiesenfeld et al., 2021).

There are several potential mechanisms for a synergistic effect of concurrent BV and M. genitalium on fecundity. First, inflammation occurs with both BV and M. genitalium and their cooccurrence may have an additive effect. Two studies have demonstrated increased concentrations of inflammatory mediators associated with M. genitalium in the lower reproductive tract, although a third study did not (Dehon et al., 2016; Garza et al., 2021; Wang et al., 2022). In vitro, an endocervical epithelial cell-model demonstrated host defense activation and pro-inflammatory cytokine secretion with M. genitalium infection (McGowin et al., 2013). While BV is not associated with vaginal neutrophils and does not often present with clinically apparent inflammation, BV has been associated with elevated levels of the pro-inflammatory cytokines IL-1β, IL-8, IL-6, and CXCL10 (Mitchell and Marrazzo, 2014; Sabo et al., 2020; Dabee et al., 2021). It is possible that when both BV and M. genitalium occur together, inflammation induced by one condition may enhance inflammation from the other. The inflammatory response to these infections could reduce fecundability by damaging ciliated cells in Fallopian tubes, disrupting fertilization, reducing endometrial receptivity and implantation, and disrupting embryonic/fetal-maternal immune interactions (Granot et al., 2012; Mor et al., 2017; D'Ippolito et al., 2018).

Given that M. genitalium can induce salpingitis and damage Fallopian tube epithelial and ciliated cells as soon as 6 days of infection (Møller et al., 1985; Cohen et al., 2005; Baczynska et al., 2007), the absence of an association between M. genitalium infection alone and reduced fecundability in sub-analysis was surprising. This supports a second possible mechanism, whereby BV may facilitate the ascension of M. genitalium to the upper reproductive tract. Many BV-associated bacteria produce sialidases and mucinases, which can degrade cervical mucus, reducing the effectiveness of this barrier to ascending pathogens (Wiggins et al., 2001). In the absence of BV, it may be more difficult for M. genitalium to ascend to the upper reproductive tract. We did not

sample the endometrium, so were unable to evaluate whether women with BV had M. genitalium in the upper reproductive tract more often than women without BV. A third potential mechanism for reduced fecundability is a direct influence of M. genitalium on sperm. In vitro, M. genitalium can attach to spermatozoa and may make them less efficient at ascending to the uterus (Svenstrup et al., 2003).

The major strengths of this analysis were its prospective design, which is the gold standard of fecundability studies, and monthly sampling for M. genitalium and BV, enabling cyclespecific estimates. In addition, this study included general population women trying to conceive early in their conception attempt and most had been pregnant before, reducing biases associated with underlying primary fertility problems. Lastly, M. genitalium was assessed using a highly sensitive and specific diagnostic test and BV was evaluated using a research gold standard, the Nugent score

There were also limitations. First, this study had limited statistical power leading to reduced precision of the results. However, the direction and magnitude of the results point to biologically plausible associations between M. genitalium and fecundability that may be driven by concurrent BV. This question deserves further attention. Second, the lack of molecular testing of the vaginal microbiota means we have limited understanding of the underlying microbiota and their relationship to M. genitalium and how that may be driving the observed association. Third, 13% of women enrolled in the parent study were excluded for this analysis solely because they did not contribute any follow-up time and another 15% were excluded because they lacked M. genitalium data due to a 1-year delay in sample collection. For the former, while most enrollment characteristics were similar between those eligible and those excluded because they lacked follow-up data, the women excluded for having no follow-up time may have had different underlying fertility potential as they were less likely have ever been pregnant, were marginally younger, and had a slightly higher M. genitalium prevalence at baseline. Characteristics were also similar when comparing those eligible versus those excluded for only lacking M. genitalium data. Most notably rates of other STIs and BV were similar suggesting that M. genitalium test positivity would not be differential. However, it is possible these exclusions could have biased effect estimates.

Among Kenyan women attempting to conceive, there was a non-significant 27% reduction in fecundability associated with M. genitalium detected at the visit prior to each pregnancy test.

b Mycoplasma genitalium at the visit prior and BV at the visit prior were time-varying.

 $^{^{\}circ}$ N = 389 participants with MG sample at enrollment.

Fecundability was further reduced (~50%) among women with both BV and M. genitalium. While these results should be interpreted cautiously given the small number of M. genitalium cases and reduced precision, they raise important questions about how M. genitalium and BV-associated bacteria may interact to influence fecundability. Further exploration of the extent to which M. genitalium causes a pro-inflammatory response in vivo, whether and how BV-associated bacteria influence M. genitalium acquisition, persistence, and ascension to the upper reproductive tract, and whether M. genitalium may itself enhance the role of nonoptimal reproductive tract microbiota on reproductive function is warranted. Ultimately, the emotional and financial burden associated with delayed time-to-pregnancy and infertility are immense (Ethics Committee of the ASRM, 2015; Luk and Loke, 2015). BV is present in approximately one-third of reproductiveage women (Koumans et al., 2007), and M. genitalium is as, or more, common as C. trachomatis infection (Seña et al., 2018; Torrone et al., 2021). Both conditions can persist for long periods, yet both are treatable. If larger studies confirm these findings, preconception screening for and treatment of BV and M. genitalium may be a low-cost intervention to reduce impaired fertility.

Supplementary data

Supplementary data are available at Human Reproduction online.

Data availability

This study was conducted with approval from the Kenyatta National Hospital—University of Nairobi Ethics and Research Committee (KNH-UON ERC), which requires their written approval for additional analyses and release of data (including de-identified data) from Kenyan studies. Therefore, data for this study will be available from the authors upon request, with written approval for the proposed analysis from the KNH/UON ERC. Application forms and guidelines can be accessed at http:// erc.uonbi.ac. To request these data, please contact KRTC Administrator at kenyares@uw.edu.

Acknowledgements

First, we offer our immense gratitude to the study participants who enrolled into this study as they tried to conceive. Their commitment during such a special and emotional time in their lives was remarkable. Second, we would like to thank the clinic, laboratory, and administrative study staff in Nairobi, Mombasa, and Seattle for their tireless efforts. Third, we are also grateful to Kenyatta National Hospital, Coast General Teaching and Referral Hospital, and the Mombasa County Department of Health for supporting this research and providing clinical and laboratory space. Lastly, we would like to thank Dr Barbra Richardson and Ken Tapia for statistical consultation.

Authors' roles

R.S.M. is the principal investigator of the parent study and supervised study protocol development and implementation. J.K. and W.J. were site principal investigators and oversaw study staff and implementation at Kenyatta National Hospital and Ganjoni Health Center. R.S.M., E.M.L., J.K., W.J., and K.M. participated in designing the parent study, protocol, data collection tools, and staff training. W.J. and K.M. oversaw laboratory protocols and methods. E.K. conducted all APTIMA testing and managed laboratory quality control. B.O. and M.N. oversaw clinical data collection and interpreted data. E.M.L., R.S.M., and L.E.M. designed this analysis and developed the statistical analysis plan for the study. E.M.L. directed the study, conceived of the analysis, conducted the statistical analyses, and wrote the first draft of the manuscript. All authors reviewed and approved the final manuscript.

Funding

This work was supported by a National Institutes of Health grant (NICHD R01 HD087346-RSM). R.S.M. received additional support for mentoring (NICHD K24 HD88229). E.M.L. was supported by pre- and post-doctoral fellowships (NIAID T32 AI07140, NICHD F32 HD100202). Data collection and management were completed using REDCap electronic data capture tools hosted at the University of Washington's Institute of Translational Health Science supported by grants from NCATS/NIH (UL1 TR002319). The content of this paper is solely the responsibility of the authors and does not necessarily represent the official views of the National Institutes of Health.

Conflict of interest

R.S.M. receives research funding, paid to the University of Washington, from Hologic Corporation and consulting fees from Lupin Pharmaceuticals. L.E.M. receives research funding and material for research studies, paid to the University of Washington, from Hologic Corporation and Nabriva Therapeutics, travel support from Hologic, and consulting fees from Health Advances. E.M.L.'s contributions to this study primarily occurred while affiliated with the University of Washington; at the time of submission, E.M.L. was an employee of and holds stock or stock grants for AbbVie, Inc. The other authors have no conflicts of Interest.

References

Australian Sexual Health Alliance. Australian STI Management Guidelines For Use in Primary Care: Mycoplasma genitalium. 2018. http://www.sti.guidelines.org.au/sexually-transmissible-infections/ mycoplasma-genitalium (5 April 2022, date last accessed).

Baczynska A, Funch P, Fedder J, Knudsen HJ, Birkelund S, Christiansen G. Morphology of human Fallopian tubes after infection with Mycoplasma genitalium and Mycoplasma hominis in vitro organ culture study. Hum Reprod 2007;22:968-979.

Balkus JE, Manhart LE, Lee J, Anzala O, Kimani J, Schwebke J, Shafi J, Rivers C, Kabare E, Scott McClelland R. Periodic presumptive treatment for vaginal infections may reduce the incidence of sexually transmitted bacterial infections. J Infect Dis 2016;213: 1932-1937.

Clausen HF, Fedder J, Drasbek M, Nielsen PK, Toft B, Ingerslev HJ, Birkelund S, Christiansen G. Serological investigation of Mycoplasma genitalium in infertile women. Hum Reprod 2001;16: 1866-1874.

Cohen CR, Mugo NR, Astete SG, Odondo R, Manhart LE, Kiehlbauch J A, Stamm WE, Waiyaki PG, Totten P A Detection of Mycoplasma genitalium in women with laparoscopically diagnosed acute salpingitis. Sex Transm Infect 2005;81:463-466.

D'Ippolito S, Di Nicuolo F, Pontecorvi A, Gratta M, Scambia G, Di Simone N. Endometrial microbes and microbiome: recent insights on the inflammatory and immune "players" of the human endometrium. Am J Reprod Immunol 2018;80:e13065.

- Dabee S, Passmore J-AS, Heffron R, Jaspan HB. The complex link between the female genital microbiota, genital infections, and inflammation. Infect Immun 2021;89:1-19.
- Dehon P, Hagensee M, Sutton K, Oddo H, Nelson N, McGowin C. Histological evidence of chronic Mycoplasma genitalium-induced cervicitis in HIV-infected women: a retrospective cohort study. J Infect Dis 2016;213:1828-1835.
- Ethics Committee of the ASRM. Disparities in access to effective treatment for infertility in the United States: an ethics committee opinion. Fertil Steril 2015;104:1104-1110.
- Evers JL. Female subfertility. Lancet 2002;360:151-159.
- Garza J, Gandhi K, Choi S, Sanchez A, Ventolini G. Cytokine profiles and Lactobacillus species presence in pre-menopausal subjects with genital Mycoplasma genitalium or Ureaplasma urealyticum colonization. Womens Health (Lond) 2021;17:17455065211009181.
- Granot I, Gnainsky Y, Dekel N. Endometrial inflammation and effect on implantation improvement and pregnancy outcome. Reproduction 2012;144:661-668.
- Haggerty CL, Totten PA, Astete SG, Lee S, Hoferka SL, Kelsey SF, Ness RB. Failure of cefoxitin and doxycycline to eradicate endometrial Mycoplasma genitalium and the consequence for clinical cure of pelvic inflammatory disease. Sex Transm Infect 2008; 84:338-342
- Haggerty CL, Totten PA, Tang G, Astete SG, Ferris MJ, Norori J, Bass DC, Martin DH, Taylor BD, Ness RB. Identification of novel microbes associated with pelvic inflammatory disease and infertility. Sex Transm Infect 2016;92:441-446.
- Jensen J, Cusini M, Gomberg M, Moi H, Wilson J, Unemo M. 2021 European guideline on the management of Mycoplasma genitalium infections. J Eur Acad Dermatol Venereol 2022;36:641-650.
- Koumans EH, Sternberg M, Bruce C, McQuillan G, Kendrick J, Sutton M, Markowitz LE. The prevalence of bacterial vaginosis in the United States, 2001-2004: associations with symptoms, sexual behaviors, and reproductive health. Sex Transm Dis 2007;34:864-869.
- Lis R, Rowhani-Rahbar A, Manhart LE. Mycoplasma genitalium infection and female reproductive tract disease: a meta-analysis. Clin Infect Dis 2015;61:418-426.
- Lokken EM, Balkus JE, Kiarie J, Hughes JP, Jaoko W, Totten PA, McClelland RS, Manhart LE. Association of recent bacterial vaginosis with acquisition of Mycopasma genitalium. Am J Epidemiol 2017;186:194-201.
- Lokken EM, Mandaliya K, Srinivasan S, Richardson BA, Kinuthia J, Lannon S, Jaoko W, Alumera H, Kemoli A, Fay E et al. Impact of preconception vaginal microbiota on women's risk of spontaneous preterm birth: protocol for a prospective case-cohort study. BMJ Open 2020;10:e035186.
- Lokken EM, Manhart LE, Kinuthia J, Hughes JP, Jisuvei C, Mwinyikai K, Muller CH, Mandaliya K, Jaoko W, Mcclelland RS. Association between bacterial vaginosis and fecundability in Kenyan women planning pregnancies: a prospective preconception cohort study. Hum Reprod 2021;36:1279-1287.
- Luk BHK, Loke AY. The impact of infertility on the psychological well-being, marital relationships, sexual relationships, and quality of life of couples: a systematic review. J Sex Marital Ther 2015; **41**:610-625.
- McGowin CL, Radtke AL, Abraham K, Martin DH, Herbst-Kralovetz M. Mycoplasma genitalium infection activates cellular host defense and inflammation pathways in a 3-dimensional human endocervical epithelial cell model. J Infect Dis 2013;207: 1857-1868.
- Mitchell C, Marrazzo J. Bacterial vaginosis and the cervicovaginal immune response. Am J Reprod Immunol 2014;71:555-563.

- Møller BR, Taylor-Robinson D, Furr PM, Toft B, Allen J. Serological evidence that Chlamydiae and mycoplasmas are involved in infertility of women. J Reprod Fertil 1985;73:237-240.
- Mor G, Aldo P, Alvero AB. The unique immunological and microbial aspects of pregnancy. Nat Rev Immunol 2017;17:469-482. Nature Publishing Group.
- National AIDS & STI Control Programme of Kenya. Kenya National Guidelines for Prevention, Management and Control of Sexually Transmitted Infections. Nairobi, Kenya: National AIDS & STI Control Programme of Kenya, 2018.
- Nugent RP, Krohn MA, Hillier SL. Reliability of diagnosing bacterial vaginosis is improved by a standardized method of Gram stain interpretation. J Clin Microbiol 1991;29:297-301.
- Nye M, Harris A, Pherson A, Cartwright C. Prevalence of Mycoplasma genitalium infection in women with bacterial vaginosis. BMC Womens Health 2020;20:62.
- Paavonen J, Westrom L, Eschenbach D. Pelvic inflammatory disease. In: Holmes K, Sparling P, Stamm W, Piot P, Wasserheit J, Corey L, Cohen M (eds). Sex Transm Dis. New York, NY: McGraw Hill, 2008,
- Peipert JF, Zhao Q, Schreiber C, Teal S, Turok DK, Natavio M, Cordon S, Daggy J. Intrauterine device use, sexually transmitted infections, and fertility: a prospective cohort study. Am J Obstet Gynecol 2021:**225**:e1-157 e9
- Sabo MC, Lehman DA, Pintye JC, Wang B, Drake AL, Kinuthia J, Osborn L, Matemo D, Richardson BA, Overbaugh J et al. Elevation of cervical C-X-C motif chemokine ligand 10 levels is associated with HIV-1 acquisition in pregnant and postpartum women. AIDS 2020;34:1725-1733.
- Schisterman EF, Cole SR, Ye A, Platt RW. Accuracy loss due to selection bias in cohort studies with left truncation. Paediatr Perinat Epidemiol 2013;27:491-502.
- Seña AC, Lee JY, Schwebke J, Philip SS, Wiesenfeld HC, Rompalo AM, Cook RL, Hobbs MM. A silent epidemic: the prevalence, incidence and persistence of Mycoplasma genitalium among young, asymptomatic high-risk women in the United States. Clin Infect Dis 2018; **67**:73-79.
- Soni S, Horner P, Rayment M, Pinto-Sander N, Naous N, Parkhouse A, Bancroft D, Patterson C, Fifer H. British association for sexual health and HIV national guideline for the management of infection with Mycoplasma genitalium (2018). Int J STD AIDS 2019;30:
- Svenstrup HF, Fedder J, Abraham-Peskir J, Birkelund S, Christiansen G. Mycoplasma genitalium attaches to human spermatozoa. Hum Reprod 2003;18:2103-2109.
- Svenstrup HF, Fedder J, Kristoffersen SE, Trolle B, Birkelund S, Christiansen G. Mycoplasma genitalium, Chlamydia trachomatis, and tubal factor infertility-a prospective study. Fertil Steril 2008;90: 513-520.
- Taylor-Robinson D, Jensen JS. Mycoplasma genitalium: from Chrysalis to multicolored butterfly. Clin Microbiol Rev 2011;24: 498-514.
- Taylor BD, Darville T, Haggerty CL. Does bacterial vaginosis cause pelvic inflammatory disease? Sex Transm Dis 2013;40:
- Torrone E, Kruszon-Moran D, Philips C, Morris M, Bowden K, Papp J, Bachmann LH, Weinstock H, Kersh EN. Prevalence of urogenital Mycoplasma genitalium infection, United States, 2017-2018. Sex Transm Dis 2021;48:e160-e162.
- US Preventative Services Task Force. Screening for Chlamydia and Gonorrhea: US preventive services task force recommendation statement. JAMA 2021;326:949-956.

- Wang R, Trent ME, Bream JH, Nilles TL, Gaydos CA, Carson KA, Coleman JS. Mycoplasma genitalium infection is not associated with genital tract inflammation among adolescent and young adult women in Baltimore, Maryland. Sex Transm Dis 2022;49:139-144.
- Weinberg C, Wilcox A, Baird DD. Reduced fecundability in women with prenatal exposure to cigarette smoking. Am J Epidemiol 1989; **129**:1072-1078.
- Weinberg CR, Baird DD, Wilcox AJ. Sources of bias in studies of time to pregnancy. Stat Med 1994;13:671-681.
- Wiesenfeld HC, Meyn LA, Darville T, Macio IS, Hillier SL. A randomized controlled trial of ceftriaxone and doxycycline, with or without metronidazole, for the treatment of acute pelvic inflammatory disease. Clin Infect Dis 2021;72:1181-1189.
- Wiggins R, Hicks SJ, Soothill PW, Millar MR, Corfield AP. Mucinases and sialidases: their role in the pathogenesis of sexually transmitted infections in the female genital tract. Sex Transm Infect 2001;77:402-408.
- Workowski KA, Bachmann LH, Chan PA, Johnston CM, Muzny CA, Park I, Reno H, Zenilman JM, Bolan GA. Sexually transmitted infections treatment guidelines, 2021. MMWR Recomm Rep 2021; **70**:1.
- Yland JJ, Bresnick KA, Hatch EE, Wesselink AK, Mikkelsen EM, Rothman KJ, Sørensen HT, Huybrechts KF, Wise LA. Pregravid contraceptive use and fecundability: prospective -cohort study. BMJ 2020;371:m3966.