

# Photosynthesis and Growth of Water Hyacinth under CO<sub>2</sub> Enrichment<sup>1</sup>

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## ABSTRACT

Water hyacinth (*Eichhornia crassipes* [Mart.] Solms) plants were grown in environmental chambers at ambient and enriched CO<sub>2</sub> levels (330 and 600 microliters CO<sub>2</sub> per liter). Daughter plants (ramets) produced in the enriched CO<sub>2</sub> gained 39% greater dry weight than those at ambient CO<sub>2</sub>, but the original mother plants did not. The CO<sub>2</sub> enrichment increased the number of leaves per ramet and leaf area index, but did not significantly increase leaf size or the number of ramets formed. Flower production was increased 147%. The elevated CO<sub>2</sub> increased the net photosynthetic rate of the mother plants by 40%, but this was not maintained as the plants acclimated to the higher CO<sub>2</sub> level. After 14 days at the elevated CO<sub>2</sub>, leaf resistance increased and transpiration decreased, especially from the adaxial leaf surface. After 4 weeks in elevated as compared to ambient CO<sub>2</sub>, ribulose biphosphate carboxylase activity was 40% less, soluble protein content 49% less, and chlorophyll content 26% less; whereas starch content was 40% greater. Although at a given CO<sub>2</sub> level the enriched CO<sub>2</sub> plants had only half the net photosynthetic rate of their counterparts grown at ambient CO<sub>2</sub>, they showed similar internal CO<sub>2</sub> concentrations. This suggested that the decreased supply of CO<sub>2</sub> to the mesophyll, as a result of the increased stomatal resistance, was counterbalanced by a decreased utilization of CO<sub>2</sub>. Photorespiration and dark respiration were lower, such that the CO<sub>2</sub> compensation point was not altered. The photosynthetic light and CO<sub>2</sub> saturation points were not greatly changed, nor was the O<sub>2</sub> inhibition of photosynthesis (measured at 330 microliters CO<sub>2</sub> per liter). It appears that with CO<sub>2</sub> enrichment the temporary increase in net photosynthesis produced larger ramets. After acclimation, the greater total ramet leaf area more than compensated for the lower net photosynthetic rate on a unit leaf area basis, and resulted in a sustained improvement in dry weight gain.

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For many plants, especially those with C<sub>3</sub> photosynthesis, an increase in CO<sub>2</sub> produces an increase in net photosynthetic rate (9). This is largely attributable to the elevated CO<sub>2</sub> competing with O<sub>2</sub> to promote the activity of RuBP carboxylase but inhibit that of RuBP oxygenase (30). Although there are notable examples where these short-term increases in photosynthetic efficiency are directly extrapolated into long-term growth improvements (6, 22, 28, 33), this is not always the case (20, 21, 24). Many

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plants exhibit some form of acclimation to high CO<sub>2</sub>, such that the initial increase in net photosynthesis is moderated or even lost, and the expected gains in productivity are not realized (10, 16, 25). The nature of this acclimation has not been fully elucidated, and it differs among species and developmental stages (11, 13, 21). It does not seem to affect directly the competitive interaction between CO<sub>2</sub> and O<sub>2</sub> for fixation by RuBPCase,<sup>2</sup> but may be mediated by more indirect effects, such as increased stomatal resistance or starch production (7, 13, 17).

This study was designed to evaluate the photosynthetic and growth responses of water hyacinth to CO<sub>2</sub> enrichment and to characterize the nature of any acclimations. Water hyacinth is a fresh-water aquatic angiosperm usually occurring as a floating plant, with its roots submersed but its leaves completely aerial. It is most likely a C<sub>3</sub> plant (23). Unlike submersed macrophytes which are shade plants (3), water hyacinth leaves growing in exposed locations can utilize full sunlight for photosynthesis (23). Despite C<sub>3</sub> characteristics, water hyacinth exhibits prolific growth in tropical and subtropical regions, with an annual biomass production of up to 250 tonnes dry weight ha<sup>-1</sup> (2), and it is now regarded as one of the foremost aquatic weed problems in the world (14). However, because of its ability to scavenge nutrients from water and its high biomass production rate, it has been used experimentally in secondary waste-water treatment facilities, and also as a biomass source for methane generation (18). For use as an energy crop, besides its high productivity, water-hyacinth has the advantages that it is not a food commodity and it does not compete for land space with agricultural food and fiber production.

Given that methane digesters produce CO<sub>2</sub> as a by-product, together with the predicted doubling of atmospheric CO<sub>2</sub> levels in the next century, it was of interest to determine both short- and long-term effects of a relatively small CO<sub>2</sub> enhancement (doubling of atmospheric levels) on the growth of this plant. The data presented here indicate that significant and sustained dry weight increases are attainable, even though the plant exhibits several acclimations to CO<sub>2</sub> enrichment which result in a reduced net photosynthetic rate on a unit leaf area basis.

## MATERIALS AND METHODS

**Plant Material.** Four plants of *Eichhornia crassipes* (Mart.) Solms, each approximately 215 g fresh weight, were placed into a 20 L plastic tub filled with one-third strength macronutrient and full strength micronutrient Hoagland solution (12), which was replaced weekly. Two Conviron E-15 growth chambers (Controlled Environments Inc.) each received four replicate tubs. The quantum irradiance at the plant level was 650 μmol m<sup>-2</sup> s<sup>-1</sup> (400–700 nm) and the plants were exposed to a 12-h, 25°C photoperiod and a 20°C scotoperiod. Growth levels of CO<sub>2</sub> in

<sup>2</sup> Abbreviations: RuBPCase, ribulose biphosphate carboxylase-oxygenase; PEPCase, phosphoenolpyruvate carboxylase.

the ambient CO<sub>2</sub> chamber were maintained at approximately 330  $\mu\text{L CO}_2 \text{ L}^{-1}$  by a flow-through system open to outside air. For the enriched-CO<sub>2</sub> chamber, an IR gas analyzer and an input solenoid system connected to a cylinder of ethylene-free liquid CO<sub>2</sub> maintained the CO<sub>2</sub> level in the air at  $600 \pm 20 \mu\text{L CO}_2 \text{ L}^{-1}$  both day and night. The fresh weight, number of daughter plants (ramets), and flowers produced were recorded weekly. After 4 weeks, the plants were separated into mother plants and ramets, weighed, their leaf area and number of leaves determined, and they were dried to constant weight at 60°C. The complete experiment was performed twice, and the data are expressed as the means of the two experiments (8 replicates)  $\pm$  the standard deviations. Statistical analysis was performed using a paired *t* test.

**Gas Exchange Measurements.** Transpiration rates and resistances for the adaxial and abaxial leaf surfaces were determined *in situ* with a portable LI-COR porometer model LI-1600 at weeks, 0, 1, 2, and 3 after the start of the experiment. At the start and end of the 4 week growth period, gas exchange measurements were made on excised leaves in a Plexiglas leaf chamber. Ancillary experiments showed that leaf excision had little effect on the net photosynthetic rate, apart from a transitory reduction of up to 16%. Photorespiratory CO<sub>2</sub> release rates into CO<sub>2</sub>-free air and CO<sub>2</sub> compensation points were determined in a closed system similar to that described by Van *et al.* (31), using an ADC (Analytical Development Company Ltd.) series 225-Mk3 IR gas analyzer. Net photosynthetic and dark respiration rates were determined with the system in an open mode. Transpiration rates were determined with an in-line EG&G Dew-All humidity analyzer model 911, from dew point measurements of the gas at the inlet and outlet of the leaf chamber. Leaf internal CO<sub>2</sub> concentrations were calculated as in Farquhar and Sharkey (8). Unless otherwise stated, all measurements were made at 30°C, and a quantum irradiance of 600  $\mu\text{mol m}^{-2} \text{ s}^{-1}$  (400–700 nm).

**Leaf Extraction and Analyses.** At the end of the experimental period, leaf samples were harvested in the light and stored in liquid N<sub>2</sub> until analysis as described previously (32). Leaf extracts were prepared by grinding to a powder approximately 1 g of frozen material in a mortar with liquid N<sub>2</sub>. The frozen powder was then extracted at 4°C in a Ten Broeck homogenizer containing 10 ml of 50 mM Tris-HCl, 10 mM MgCl<sub>2</sub>, 0.1 mM EDTA, 5 mM isoascorbate, and 1% w/v PVP-40 at pH 8.0. Aliquots were removed for Chl (1), soluble protein (4), and starch (5) determinations. The glucose released from starch (5) was measured using a Yellow Springs Instruments Company glucose analyzer model 27. RuBPCase and PEPCase assays were performed at 30°C. RuBPCase activity was assayed in the active form as described by Vu *et al.* (32), and PEPCase activity as in Van *et al.* (31).

## RESULTS

Water hyacinth grown under enriched-CO<sub>2</sub> conditions (600  $\mu\text{L CO}_2 \text{ L}^{-1}$ ) showed a 32% increase in dry matter production as

compared to the plants grown at ambient CO<sub>2</sub> levels (Table I). Most of this increase was due to an increase in ramets. The dry matter production of the ramets from the enriched-CO<sub>2</sub> treatments was 39% greater than that of the ambient-grown ramets, but the mother plants showed little difference (Table I). The dry weight of the individual ramets was significantly greater in the enriched-CO<sub>2</sub> treatment, but the number of ramets produced per m<sup>2</sup> did not change (Table I). There was also a large increase in the number of flowers formed in the enriched-CO<sub>2</sub> treatment (Table I).

As shown in Table II, the number of leaves and the leaf area per plant increased under the elevated CO<sub>2</sub>, resulting in a 46% increase in the total leaf area index. However, the area of individual leaves exhibited only a slight increase. The dry weight per g fresh weight was higher in ramets from the enriched-CO<sub>2</sub> treatment, as was the starch content of the leaves (Table III); but both soluble protein and Chl were substantially decreased by the CO<sub>2</sub> enrichment (Table III).

Net photosynthetic rates of the enriched-CO<sub>2</sub> and ambient grown leaves, measured at 330  $\mu\text{L CO}_2 \text{ L}^{-1}$ , saturated at a quantum irradiance of approximately 900 and 1100  $\mu\text{mol m}^{-2} \text{ s}^{-1}$  (400–700 nm), respectively. Light compensation points of 16 and 14  $\mu\text{mol m}^{-2} \text{ s}^{-1}$  (400–700 nm), respectively, also were comparable (Fig. 1). Net photosynthetic rates of the enriched-CO<sub>2</sub> grown leaves, when measured at 330  $\mu\text{L CO}_2 \text{ L}^{-1}$ , were considerably less at all irradiances than those of the ambient grown plants (Fig. 1).

With respect to external CO<sub>2</sub> concentration, the light-saturated net photosynthetic rate of the mother plants at the start of the experiment was saturated at approximately 1100  $\mu\text{L CO}_2 \text{ L}^{-1}$ . Increasing the CO<sub>2</sub> level from ambient to 600  $\mu\text{L CO}_2 \text{ L}^{-1}$  resulted in a 42% increase in net photosynthetic rate (data not shown). Similar results were obtained at the conclusion of the experiment for the ramets produced in the growth chamber at ambient levels of CO<sub>2</sub> (Fig. 2). The leaves of the enriched-CO<sub>2</sub> grown ramets required less CO<sub>2</sub> to achieve saturation (about 800  $\mu\text{L CO}_2 \text{ L}^{-1}$ ). Also, they had a lower net photosynthetic rate (expressed on a Chl basis) at each CO<sub>2</sub> concentration than the leaves of ambient grown plants (Fig. 2). However, when measured at the CO<sub>2</sub> level under which they were grown, net photosynthetic rates of leaves from the enriched-CO<sub>2</sub> and ambient growth treatments were similar (68 versus 64  $\mu\text{mol mg}^{-1} \text{ Chl h}^{-1}$ , respectively; Fig. 2). When net photosynthetic rates were expressed on a per unit leaf area basis (Table IV) the enriched plants showed a rate that was about half that of the ambient grown plants, measured at either growth CO<sub>2</sub> level.

Leaves of the enriched-CO<sub>2</sub> grown plants had lower net photosynthetic rates for a given internal CO<sub>2</sub> concentration than their ambient grown counterparts (Fig. 3). Calculated leaf internal CO<sub>2</sub> levels were almost linearly related to the external CO<sub>2</sub> level between 100 and 900  $\mu\text{L CO}_2 \text{ L}^{-1}$  (Fig. 4). However, for a given external CO<sub>2</sub> concentration, leaves grown under the two

Table I. Increase in Dry Weight, Number of Flowers, and Ramet Production of Water Hyacinth Grown under 330 and 600  $\mu\text{L CO}_2 \text{ L}^{-1}$  over a 4 Week Period

	Increase in Wt			Individual Ramet Wt	Ramet Production	Flower Production
	Total Plants	Mother Plants	Ramets			
		<i>g dry wt · m<sup>-2</sup></i>		<i>g dry wt</i>	<i>number · m<sup>-2</sup></i>	
Grown at 330 $\mu\text{L CO}_2 \text{ L}^{-1}$	122.2 <sup>a</sup> $\pm$ 14.2	44.4 $\pm$ 14.5	108.1 $\pm$ 7.1	2.3 $\pm$ 0.4	46.6 $\pm$ 5.4	1.9 $\pm$ 1.6
Grown at 600 $\mu\text{L CO}_2 \text{ L}^{-1}$	161.0 $\pm$ 14.6	47.0 $\pm$ 9.2	150.0 $\pm$ 11.4	3.1 $\pm$ 0.3	48.1 $\pm$ 6.1	4.7 $\pm$ 1.1
% change relative to 330 $\mu\text{L CO}_2 \text{ L}^{-1}$	32 <sup>b</sup>	6	39 <sup>b</sup>	35 <sup>b</sup>	3	147 <sup>b</sup>

<sup>a</sup> Each value is the mean of 8 replications  $\pm$  SD.

<sup>b</sup> Difference between the means significant at the  $\alpha = 0.05$  level as determined by a paired *t* test.

Table II. Number of Leaves and Leaf Area of Ramets Produced by Water Hyacinth Grown under 330 and 600  $\mu\text{L CO}_2 \text{ L}^{-1}$  over a 4 Week Period

	Leaves Per Plant	Leaf Area Per Plant	Area Per Leaf	Leaf Area Index
	number	$\text{dm}^2$		$\text{dm}^2 \cdot \text{dm}^{-2}$
Grown at 330 $\mu\text{L CO}_2 \text{ L}^{-1}$	5.9 <sup>a</sup> $\pm$ 0.4	6.0 $\pm$ 0.5	1.0 $\pm$ 0.4	2.8 $\pm$ 0.2
Grown at 600 $\mu\text{L CO}_2 \text{ L}^{-1}$	7.5 $\pm$ 0.8	8.4 $\pm$ 0.9	1.1 $\pm$ 0.9	4.1 $\pm$ 0.5
% change relative to 330 $\mu\text{L CO}_2 \text{ L}^{-1}$	27 <sup>b</sup>	40 <sup>b</sup>	10	46 <sup>b</sup>

<sup>a</sup> Each value is the mean of 7 replications  $\pm$  SD. <sup>b</sup> Difference between the means significant at the  $\alpha = 0.05$  level as determined by a paired *t* test.

Table III. Dry:Fresh Weight Ratio, and Starch, Soluble Protein, and Chlorophyll Content of Ramets Produced by Water Hyacinth Grown under 330 and 600  $\mu\text{L CO}_2 \text{ L}^{-1}$  over a 4 Week Period

	Dry Wt:Fresh Wt Ratio	Starch	Protein	Chlorophyll
	$\text{g} \cdot \text{g}^{-1}$		$\text{mg} \cdot \text{g}^{-1}$ fresh wt	
Grown at 330 $\mu\text{L CO}_2 \text{ L}^{-1}$	0.039 <sup>a</sup> $\pm$ 0.004	7.0 $\pm$ 0.9	7.8 $\pm$ 0.6	1.9 $\pm$ 0.01
Grown at 600 $\mu\text{L CO}_2 \text{ L}^{-1}$	0.049 $\pm$ 0.001	9.8 $\pm$ 1.3	4.0 $\pm$ 0.3	1.4 $\pm$ 0.05
% change relative to 330 $\mu\text{L CO}_2 \text{ L}^{-1}$	26	40 <sup>b</sup>	-49 <sup>b</sup>	-26 <sup>b</sup>

<sup>a</sup> Each value is the mean of 3 replications  $\pm$  SD. <sup>b</sup> Difference between the means significant at the  $\alpha = 0.05$  level as determined by a paired *t* test.

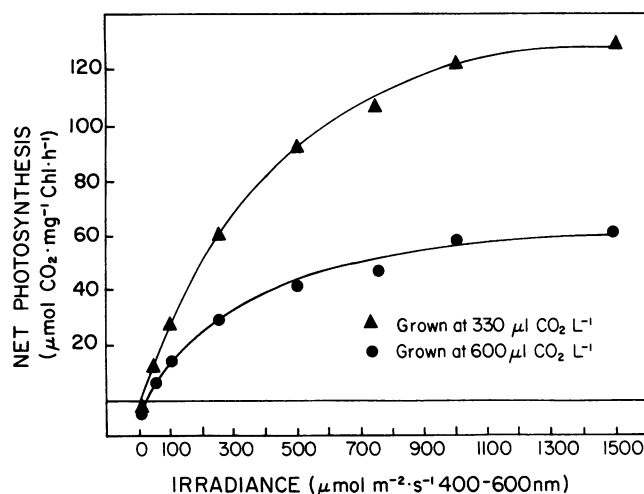


FIG. 1. Photosynthetic response to irradiance of water hyacinth grown under 330 and 600  $\mu\text{L CO}_2 \text{ L}^{-1}$  for a 4 week period. Measurements were made at 330  $\mu\text{L CO}_2 \text{ L}^{-1}$ .

different  $\text{CO}_2$  regimes exhibited almost identical internal  $\text{CO}_2$  levels.

The  $\text{CO}_2$  compensation points for leaves of the enriched and ambient grown plants were similar (Table V), and did not change from the initially measured values. The degree of inhibition of photosynthesis by 21%  $\text{O}_2$ , although slightly less for the enriched as compared to the ambient grown ramets, was not statistically different. In contrast, estimated photorespiratory rates, measured as  $\text{CO}_2$  release into  $\text{CO}_2$ -free air in the light, were almost halved following growth at the enriched- $\text{CO}_2$  level (Table V). Dark respiration rates (measured at 330  $\mu\text{L CO}_2 \text{ L}^{-1}$ ) of the leaves were reduced by about one-third in the enriched plants, as compared with those grown at ambient  $\text{CO}_2$  levels (Table V).

Transpiration and total leaf resistance were measured on leaves in the growth chambers throughout the experiment (Table VI). The transpiration rate of enriched- $\text{CO}_2$  plants declined over the

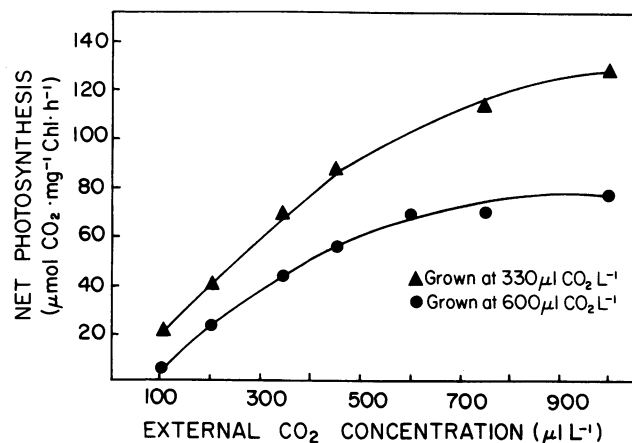


FIG. 2. Photosynthetic response to external  $\text{CO}_2$  concentration of water hyacinth grown under 330 and 600  $\mu\text{L CO}_2 \text{ L}^{-1}$  for a 4 week period.

Table IV. Comparison of the Photosynthetic Rates of Air- and Enriched- $\text{CO}_2$  Grown Water Hyacinth Ramets on a Unit Leaf Area Basis

$\text{CO}_2$ Concentration Used for Plant Growth	Net Photosynthetic Rate	
	Measured at 330 $\mu\text{L CO}_2 \text{ L}^{-1}$	Measured at 600 $\mu\text{L CO}_2 \text{ L}^{-1}$
$\mu\text{L L}^{-1}$	$\mu\text{mol CO}_2 \cdot \text{m}^{-2} \cdot \text{s}^{-1}$	
330	11.4 <sup>a</sup> $\pm$ 1.7	16.7 $\pm$ 1.9
600	5.8 $\pm$ 2.7	8.9 $\pm$ 4.3

<sup>a</sup> Each value is the mean of three replications  $\pm$  SD.

course of the experiment. This was especially evident for the adaxial leaf surfaces. After 21 d, adaxial transpiration from the enriched- $\text{CO}_2$  leaves was only 15% of that of the ambient grown plants, while for the abaxial surfaces it was 67%. Concomitantly, total leaf resistance increased during the same time period in the enriched- $\text{CO}_2$  plants (Table VI). When enriched- $\text{CO}_2$  plants were

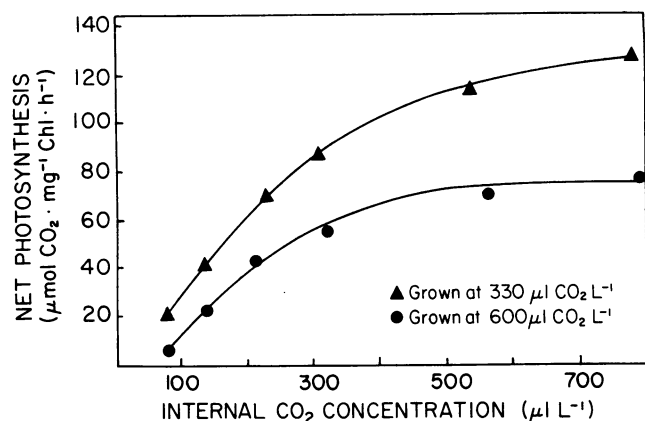


FIG. 3. Photosynthetic response to internal CO<sub>2</sub> concentration of water hyacinth grown under 330 and 600 µL CO<sub>2</sub> L<sup>-1</sup> for a 4 week period.

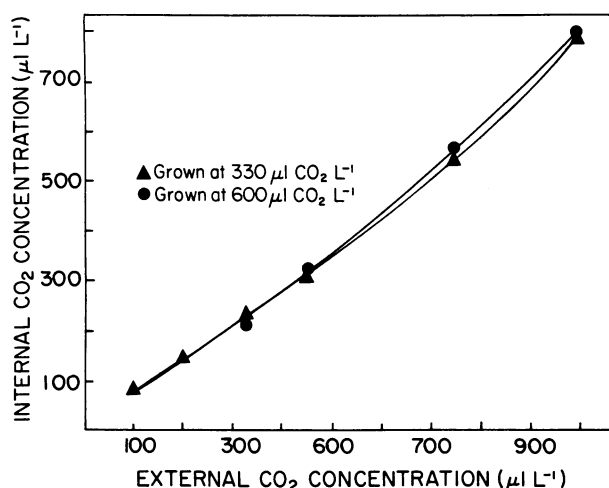


FIG. 4. Internal CO<sub>2</sub> concentration versus external CO<sub>2</sub> concentration of water hyacinth grown under 330 and 600 µL CO<sub>2</sub> L<sup>-1</sup> for a 4 week period.

returned to ambient air, the leaf resistances and adaxial transpiration rates did not return to initial levels, even after several days (data not shown).

Maximum PEPCase activities measured in water hyacinth leaves were about 6 to 7% of the maximum RuBPCase values (Table VII). RuBPCase was the primary carboxylation enzyme and its activity on a Chl basis declined by approximately 20% during growth at enriched as compared to ambient levels (Table VII). This decline was even more evident when the activity was

expressed on a fresh weight basis. PEPCase activity showed less difference between the ambient- and enriched-CO<sub>2</sub> grown plants.

## DISCUSSION

The growth enhancement of water hyacinth due to CO<sub>2</sub> enrichment was largely restricted to the ramets. A number of workers have reported that immature tissue is more responsive to CO<sub>2</sub> (11, 21, 28) but this cannot fully explain the current findings, since the mother plants did show a large, temporary, enhancement of photosynthetic rate when initially exposed to high CO<sub>2</sub>. Presumably, much of the increase in assimilate formed by the mother plants during the temporary photosynthetic rate enhancement was allocated to the ramets. The temporary rate enhancement for the mother plants in enriched CO<sub>2</sub> did not increase the number of ramets produced, but did increase the dry weight, number of leaves, leaf area per ramet, and leaf area index.

Water hyacinth growth is indeterminate, and the major method of reproduction is vegetative by ramet production (26). The results presented here indicate that CO<sub>2</sub> enrichment led to an enhancement of leaf meristem initiation, rather than an enhancement of ramet-producing meristems whose numbers may be more limited. This contrasts with observations for wheat, in which tiller production is greater under elevated CO<sub>2</sub> (29) and soybean, where dry weight increases have been attributed to greater specific leaf weights and not the initiation of new leaves or greater leaf area (6). For some species, CO<sub>2</sub> enrichment has been reported to delay flower production (19). This is not the case for water hyacinth which showed a substantial increase in flower production at 600 µL CO<sub>2</sub> L<sup>-1</sup>, a response similar to that reported for greenhouse-grown tomato and cucumber (33). Thus, CO<sub>2</sub> enrichment could increase the potential for sexual reproduction in water hyacinth, but this is still an uncertain conclusion because seed production was not determined. Seed production is not necessary for the maintenance of water hyacinth in a nonfluctuating water-body (26), as regrowth is usually initiated vegetatively by plants that have over-wintered. However, in ephemeral water bodies where vegetative material desiccates, regeneration from seed becomes important (26).

As is commonly found with C<sub>3</sub> plants, doubling the ambient CO<sub>2</sub> concentration during short-term photosynthesis measurements resulted in an immediate 40 to 60% increase in the net photosynthetic rate of water hyacinth. However, the initial increase per unit leaf area of water hyacinth was not maintained. After 4 weeks, the net photosynthetic rate of the enriched plants was approximately half that of ambient grown plants when both were measured at the same external CO<sub>2</sub> concentration. As might be anticipated from the lack of any major change in the CO<sub>2</sub> compensation point and O<sub>2</sub> inhibition of photosynthesis, the reduced net photosynthetic rate of the enriched-CO<sub>2</sub> plants was accompanied by a concomitant reduction in the estimated rate

Table V. CO<sub>2</sub> Compensation Point, O<sub>2</sub> Inhibition of Photosynthesis, Photorespiratory CO<sub>2</sub> Release, and Dark Respiration of Ramets Produced by Water Hyacinth Grown under 330 and 600 µL CO<sub>2</sub> L<sup>-1</sup> for 4 Weeks

	CO <sub>2</sub> Compensation Point	O <sub>2</sub> Inhibition <sup>a</sup>	CO <sub>2</sub> Release into CO <sub>2</sub> Free Air	Dark Respiration
	µL L <sup>-1</sup>	%	µmol CO <sub>2</sub> · mg <sup>-1</sup> Chl · h <sup>-1</sup>	
Grown at 330 µL CO <sub>2</sub> L <sup>-1</sup>	51.7 <sup>b</sup> ± 3.2	41.0 ± 4.8	11.0 ± 3.5	5.0 ± 0.3
Grown at 600 µL CO <sub>2</sub> L <sup>-1</sup>	55.5 ± 2.6	33.7 ± 7.4	5.6 ± 0.6	3.2 ± 1.7
% change relative to 330 µL CO <sub>2</sub> L <sup>-1</sup>	7	-18	-49 <sup>c</sup>	-36

<sup>a</sup> Measured at 21% versus 1% O<sub>2</sub> and at 330 µL CO<sub>2</sub> L<sup>-1</sup>. <sup>b</sup> Each value is the mean of 3 replications ± SD. <sup>c</sup> Difference between the means significant at the α = 0.05 level as determined by a paired *t* test.

Table VI. *Transpiration Rates and Leaf Resistances of Ramets Produced by Water Hyacinth Grown in Growth Chambers at 330 and 600  $\mu\text{L CO}_2 \text{ L}^{-1}$  over a 3 Week Period*

Time	Grown at 330 $\mu\text{L CO}_2 \text{ L}^{-1}$			Grown at 600 $\mu\text{L CO}_2 \text{ L}^{-1}$		
	Transpiration		Total Leaf Resistance	Transpiration		Total Leaf Resistance
	Adaxial	Abaxial		Adaxial	Abaxial	
<i>d</i>	<i>mmol H<sub>2</sub>O m<sup>-2</sup> s<sup>-1</sup></i>		<i>s m<sup>-1</sup></i>	<i>mmol H<sub>2</sub>O m<sup>-2</sup> s<sup>-1</sup></i>		<i>s m<sup>-1</sup></i>
2	8.1 <sup>a</sup>	6.3	48	6.7	7.1	49
7	9.3	8.4	19	6.2	7.5	60
14	6.2	3.4	72	0.6	4.1	378
21	7.6	8.2	66	1.2	5.5	115

<sup>a</sup> Each value is the mean of 3 measurements.

Table VII. *RuBPCase and PEPCase Activities of Ramets Produced by Water Hyacinth Grown under 330 and 600  $\mu\text{L CO}_2 \text{ L}^{-1}$  over a 4 Week Period*

	RuBPCase	PEPCase	RuBPCase	PEPCase
	$\mu\text{mol} \cdot \text{mg}^{-1} \text{ Chl} \cdot \text{h}^{-1}$		$\mu\text{mol} \cdot \text{g}^{-1} \text{ fresh wt} \cdot \text{h}^{-1}$	
Grown at 330 $\mu\text{L CO}_2 \text{ L}^{-1}$	380 ± 11 <sup>a</sup>	23 ± 0.3	703 ± 20.3	43 ± 0.6
Grown at 600 $\mu\text{L CO}_2 \text{ L}^{-1}$	301 ± 12	22 ± 1.2	424 ± 16.9	31 ± 1.7
% change relative to 330 $\mu\text{L CO}_2 \text{ L}^{-1}$	-21 <sup>b</sup>	-4	-40 <sup>b</sup>	-28 <sup>b</sup>

<sup>a</sup> Each value is the mean of 3 replications ± the SD.  
= 0.05 level as determined by a paired *t* test.

<sup>b</sup> Difference between the means significant at the  $\alpha$

of photorespiration. The dark respiration rate also decreased. Although the lower dark respiration rate was probably a factor in achieving the higher dry weight of the enriched- $\text{CO}_2$  plants, by itself it was of insufficient magnitude to account for the phenomenon.

The increase in leaf resistance and decrease in transpiration, together with the decreased photosynthetic rate, all point to a leaf acclimation to elevated  $\text{CO}_2$ , which occurred for water hyacinth after 7 to 14 d of growth at 600  $\mu\text{L CO}_2 \text{ L}^{-1}$ . The adaxial and abaxial stomates responded differently to the increased  $\text{CO}_2$ , such that stomatal closure on the adaxial side appeared to be the major cause of the increased leaf resistance measurements. This stomatal acclimation was not a transitory effect, as it did not disappear when the plants were returned to ambient  $\text{CO}_2$  levels. These data, together with the increase in starch content, and decreases in levels of soluble protein, RuBPCase activity, and Chl indicate that a variety of adaptations occur during growth at elevated  $\text{CO}_2$  levels. Thus, the observed growth response cannot be attributed simply to a lessening of photorespiration, as is the case for short-term photosynthesis measurements at high  $\text{CO}_2$  levels.

From the viewpoint of using water hyacinth as an energy crop (27), the reallocation of fixed carbon at elevated  $\text{CO}_2$  levels to favor carbohydrates, rather than protein, would be beneficial; conversely this effect would be detrimental if the plant were to be used as an animal feed supplement (18). A similar redistribution of carbon to favor starch formation is a common phenomenon among  $\text{CO}_2$ -enriched plants (15, 25).

Plants whose growth responds positively to  $\text{CO}_2$  enrichment seem to exhibit two general photosynthetic patterns. In some the net photosynthetic rate per unit leaf area is increased at the elevated  $\text{CO}_2$  and usually remains so throughout the growth cycle, although the magnitude of the response may be modified at different developmental stages (11, 15). Thus, the improvement in total dry weight is largely due to a permanent increase in photosynthetic efficiency, as a result of the reduced  $\text{O}_2$  effects on RuBP carboxylase-oxygenase (30). In these plants, RuBPCase shows little reduction in total activity by growth at up to 1000  $\mu\text{L CO}_2 \text{ L}^{-1}$  (11, 32).

The more usual pattern of response to elevated  $\text{CO}_2$  is that

shown by water hyacinth in this study. There is an initial, but temporary, enhancement in net photosynthesis per unit leaf area, which leads to an increased production of biomass. In the case of water hyacinth, the increase in biomass was most apparent as an increase in the number of leaves, which resulted in an increase in leaf area index. Following the rate enhancement, the plants acclimate to the high  $\text{CO}_2$  and the photosynthetic rate diminishes, sometimes even below that of the plants at ambient levels (7, 25). For water hyacinth the sustained total dry weight increase appeared to be due to the greater leaf area index, rather than an improved photosynthetic efficiency per unit leaf area. Although  $\text{CO}_2$  enrichment reduced the competitive inhibition of RuBPCase by  $\text{O}_2$ , this advantage was more than offset by the eventual large decrease in RuBPCase activity, which must have been a factor in the reduced overall photosynthetic efficiency per unit leaf area (25). Crowding of water hyacinth ramets and thus maximizing the leaf area index, could be expected to reduce the  $\text{CO}_2$ -enrichment effects on dry weight, though the yield increase should be self-sustaining if the plants were harvested or thinned-out at appropriate intervals to prevent overcrowding.

The cause of the acclimation and eventual diminution of photosynthetic rate seen in many plants grown at elevated  $\text{CO}_2$  levels has been the subject of debate (11, 16, 25). For  $\text{CO}_2$ -enriched water hyacinth, the increase in leaf resistance after 14 d, due to the closure of adaxial stomates, could not by itself explain the reduced photosynthetic rate. This is because at any given external  $\text{CO}_2$  level the internal  $\text{CO}_2$  concentrations of the ambient grown and enriched plants were similar, despite the high leaf resistances of the enriched plants. This indicates that not only was the supply of  $\text{CO}_2$  to the mesophyll cells restricted in the enriched plants, but that utilization of the internal  $\text{CO}_2$  was also reduced, counterbalancing the restricted supply.

Starch accumulation, leading to some form of feedback inhibition or chloroplast disruption, has been most frequently postulated as responsible for the reduced photosynthetic rates in  $\text{CO}_2$  enriched plants (13). Certainly in water hyacinth the carbon allocated to starch was much greater in the enriched- $\text{CO}_2$  grown plants. However, the redistribution of carbon was paralleled by a substantial reduction in extractable RuBPCase activity and Chl. All of these factors potentially impinge on the photosyn-

thetic rate, and at this stage it would be premature to single out any one as being the primary cause of the reduced rate in water hyacinth. Until a more detailed kinetic analysis of these parameters is undertaken for water hyacinth during the acclimation period, it can only be speculated as to whether the increased leaf resistance precedes, or is subsequent to, the long-term changes in the mesophyll cells. The data are suggestive, however, that long-term stomatal behavior and the photosynthetic metabolism of the underlying mesophyll cells may be closely regulated in concert.

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#### LITERATURE CITED

- ARNON DI 1949 Copper enzymes in isolated chloroplasts. Polyphenoloxidase in *Beta vulgaris*. *Plant Physiol* 24: 1–15
- BEADLE CL, SP LONG 1985 Photosynthesis—is it limiting to biomass production? *Biomass* 8: 119–168
- BOWES G 1985 Pathways of CO<sub>2</sub> fixation by aquatic organisms. In WJ Lucas, JA Berry, eds. *Inorganic Carbon Uptake by Aquatic Photosynthetic Organisms*. American Society of Plant Physiologists, Rockville, MD, pp 187–210
- BRADFORD MM 1976 A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein-dye binding. *Anal Biochem* 72: 248–254
- BUDKE CC 1984 Determination of total available glucose in corn base materials. *J Agric Food Chem* 32: 34–37
- CLOUGH JM, MM PEET 1981 Effects of intermittent exposure to high atmospheric CO<sub>2</sub> on vegetative growth in soybean. *Physiol Plant* 53: 565–569
- DOWNTON WJS, O BJÖRKMÄN, CS PIKE 1980 Consequences of increased atmospheric concentrations of carbon dioxide for growth and photosynthesis of higher plants. In GI Pearman, ed. *Carbon Dioxide and Climate*. Australian Academy of Science, Canberra, pp 143–151
- FARQUHAR GH, TD SHARKEY 1982 Stomatal conductance and photosynthesis. *Annu Rev Plant Physiol* 33: 317–345
- FOCK H, K KLUG, DT CANVIN 1979 Effect of carbon dioxide and temperature on photosynthetic CO<sub>2</sub> uptake and photorespiratory CO<sub>2</sub> evolution in sunflower leaves. *Planta* 145: 219–223
- FRYDRYCH J 1976 Photosynthetic characteristics of cucumber seedlings grown under two levels of carbon dioxide. *Photosynthetica* 10: 335–338
- HICKLENTON PR, PA JOLLIFFE 1980 Alterations in the physiology of CO<sub>2</sub> exchange in tomato plants grown in CO<sub>2</sub>-enriched atmospheres. *Can J Bot* 58: 2181–2189
- HOAGLAND DR, DI ARNON 1950 The water culture method of growing plants without soil. *Calif Agric Exp Stn Circ* 347: 1–32
- HOFSTRA G, JD HESKETH 1975 The effects of temperature and CO<sub>2</sub> enrichment on photosynthesis in soybean. In R Marcelle, ed. *Environmental and Biological Control of Photosynthesis*, Dr W Junk, The Hague, pp 71–80
- HOLM LG, DL PLUCKNETT, JV PANCHO, JP HERBERGER 1977 *The World's Worst Weeds: Distribution and Biology*. University Press of Hawaii, Honolulu, pp 72–77
- HUBER SC, H ROGERS, DW ISRAEL 1984 Effects of CO<sub>2</sub> enrichment on photosynthesis and photosynthate partitioning in soybean (*Glycine max*) leaves. *Physiol Plant* 62: 95–101
- IMAI K, Y MURATA 1978 Effect of carbon dioxide concentration on growth and dry matter production of crop plants. V. Analysis of after-effect of carbon dioxide-treatment on apparent photosynthesis. *Jpn J Crop Sci* 47: 587–595
- JURIK TW, JA WEBER, DM GATES 1984 Short-term effects of CO<sub>2</sub> on gas exchange of leaves of Bigtooth Aspen (*Populus grandidentata*) in the field. *Plant Physiol* 75: 1022–1026
- KOBAYASHI T, K Ueki 1981 Cultivation and utilization of new biomass resources (An aquatic weed, water hyacinth). *Energy Dev Jpn* 3: 285–300
- MARC J, RM GIFFORD 1984 Floral initiation in wheat, sunflower, and sorghum under carbon dioxide enrichment. *Can J Bot* 62: 9–14
- MAUNEY JR, KE FRY, G GUINN 1978 Relationship of photosynthetic rate to growth and fruiting of cotton, soybean, sorghum, and sunflower. *Crop Sci* 18: 259–263
- NEALES TF, AO NICHOLLS 1978 Growth responses of young wheat plants to a range of ambient CO<sub>2</sub> levels. *Aust J Plant Physiol* 5: 45–59
- NILSEN S, K HOVLAND, C DONS, SP SLETTEN 1983 Effect of CO<sub>2</sub> enrichment on photosynthesis, growth, and yield of tomato. *Sci Hortic* 20: 1–14
- PATTERSON DT, SO DUKE 1979 Effect of growth irradiance on the maximum photosynthetic capacity of water hyacinth (*Eichhornia crassipes* (Mart.) Solms). *Plant Cell Physiol* 20: 177–184
- PEET MM 1986 Acclimation to high CO<sub>2</sub> in monoecious cucumbers. I. Vegetative and reproductive growth. *Plant Physiol* 80: 59–62
- PEET MM, SC HUBER, DT PATTERSON 1986 Acclimation to high CO<sub>2</sub> in monoecious cucumbers. II. Carbon exchange rates, enzyme activities, and starch and nutrient concentrations. *Plant Physiol* 80: 63–67
- PENFOUND WT, TT EARLE 1948 The biology of the water hyacinth. *Ecol Monogr* 18: 447–472
- REDDY KR, DL SUTTON, G BOWES 1983 Freshwater aquatic plant biomass production in Florida. *Soil Crop Sci Soc Fl Proc* 42: 28–40
- SIONIT N, H HELLMERS, BR STRAIN 1982 Interaction of atmospheric CO<sub>2</sub> enrichment and irradiance on plant growth. *Agron J* 74: 721–725
- SIONIT N, BR STRAIN, H HELLMERS 1981 Effects of different concentrations of atmospheric CO<sub>2</sub> on growth and yield components of wheat. *J Agric Sci* 79: 335–339
- TOLBERT NE 1980 Photorespiration. In PK Stumpf, EE Conn, eds. *The Biochemistry of Plants*, Vol 2. DD Davies, ed. *Metabolism and Respiration*, Academic Press, New York
- VAN TK, WT HALLER, G BOWES 1976 Comparison of the photosynthetic characteristics of three submersed aquatic plants. *Plant Physiol* 58: 761–768
- VU CV, LH ALLEN JR, G BOWES 1983 Effects of light and elevated atmospheric CO<sub>2</sub> on the ribulose biphosphate carboxylase activity and ribulose biphosphate level of soybean leaves. *Plant Physiol* 73: 729–734
- WITTWER SH, W ROBB 1964 Carbon dioxide enrichment of greenhouse atmospheres for food crop production. *Econ Bot* 18: 34–56