# Leaf Photosynthesis and Conductance of Selected Triticum Species at Different Water Potentials'

Received for publication July 28, 1986 and in revised form November 21, 1986

RICHARD C. JOHNSON\*, DOLORES W. MORNHINWEG, DAVID M. FERRIS, AND JAMES J. HEITHOLT Department ofAgronomy, Oklahoma State University, Stillwater, Oklahoma 74078

## ABSTRACT

Leaf gas exchange characteristics of a desert annual (Triticum kotschyi [Boiss.j Bowden) and the wheat cultivar TAM W-101 (Triticum aestivum L. em Thell) were compared over a range of leaf water potentials from  $-0.50$  to  $-2.9$  megapascals. At an ambient  $[CO<sub>2</sub>]$  of 330 microliters per liter, T. kotschyi had higher conductance and  $CO<sub>2</sub>$  assimilation (A) at a given water potential than T. aestivum. Under well watered conditions, A versus internal  $CO<sub>2</sub>$  concentration  $(C<sub>i</sub>)$  response curves for both species were similar in shape and magnitude, and the higher A of T. kotschyi at an ambient  $[CO<sub>2</sub>]$  of 330 microliters per liter was mostly related to the higher stomatal conductance of T. kotschyi. The higher conductance of T. kotschyi than T. aestivum under well watered conditions was associated with higher  $C_i$  and lower water use efficiency. Under water deficits, however,  $C_i$  at 330 microliters per liter ambient  $[CO_2]$  did not differ significantly between species. T. kotschyi had higher A under water deficits than T. *aestivum* primarily because its A *versus*  $C_i$  response curves had higher A at C, values above about 150 microliters per liter. The results show that conductance played an important role in the high A of T. kotschyi under well watered conditions, but under water deficits the high A of T. kotschyi was related more to the maintenance of <sup>a</sup> higher capacity for mesophyll photosynthesis.

For some time it was thought that reduced photosynthesis under water deficits was to a significant extent mediated by stomatal closure, causing reduced  $CO<sub>2</sub>$  concentrations inside the leaf. Recently, however, it has been shown that reductions in mesophyll photosynthetic capacity and reductions in stomatal conductance under water deficits may, to a significant degree, be coupled (7). As a result, leaf  $C_i^2$  is relatively stable and reduced stomatal conductance is not considered the major cause of reduced photosynthesis under water deficits (7, 14). Stomata may still, however, play an important role in determining and explaining differences in the photosynthetic rate and water use efficiency between given plant species.

Many wild diploid and tetraploid Triticum species have photosynthetic rates that are considerably higher than domestic hexaploid wheats (Triticum aestivum), yet the physiological basis for these differences is not well understood (1). Triticum kotschyi is a tetraploid desert annual with reportedly higher photosynthesis both under well watered conditions and at reduced water potentials compared to  $T$ . aestivum (13). Although  $T$ . kotschyi

<sup>2</sup> Abbreviations:  $C_i$ , internal  $CO_2$  concentration; A,  $CO_2$  assimilation.

appears to maintain a higher degree of relative stomatal opening at reduced water potentials than T. aestivum, it is not clear if this is the only factor involved in the control of photosynthesis (13). In this study, we report that stomata had an important role in causing the relatively high photosynthetic rates of T. kotschyi under well watered conditions, but under water deficits the mesophyll capacity for photosynthesis was more important.

## MATERIALS AND METHODS

Plant Material. Triticum aestivum L. em Thell. (cv TAM W-101) and Triticum kotschyi (Boiss) Bowden are both herbaceous annuals. The tetraploid T. kotschyi grows in the Negev desert of Israel (13) and the T. aestivum cultivar TAM W-<sup>101</sup> is <sup>a</sup> hexaploid winter wheat grown widely in the southern Great Plains of the United States.

Unvernalized plants were grown in growth chambers at 2O°C, 60% RH, and 14 h light (600  $\mu$ mol m<sup>-2</sup> s<sup>-1</sup> PAR at pot level). Three seeds were planted per pot about 1.5 cm deep in pots holding <sup>1</sup> L of sterilized sand. Emerged seedlings were thinned to <sup>1</sup> per pot after a week and the remaining plants watered daily with 25% Hoagland solution until the stress treatments were imposed. In the first experiment plants of T. kotschyi and T. aestivum were grown under well watered conditions for 42 d after which gas exchange measurements were made as described below. The experiment was randomized in complete blocks with five replications. In a second experiment plants of the two species were grown for 35 d under well-watered conditions before water deficits were imposed by withholding water from pots. The experiment was randomized in complete blocks with four replications with genotype and water stress level (days without water) as experimental factors. Gas exchange measurement for the four water levels were made over a 5-d period with a given stress level completed each day. Plants of each species were measured at 0, 2,3, and 4 d after withholding water from pots. The measurement order for stress level and species within a stress level was randomized. In a third experiment water deficits were applied to T. kotschyi and T. aestivum over a more prolonged period. The water-holding capacity of the sand was determined and the weight of each pot, filled with equal volumes of dry sand, was recorded. The pots were weighed daily and their percent water calculated. Plants were watered to the maximum water holding capacity each day before the start of the stress treatment. For the stress treatment, plants were grown under well watered conditions for 24 d and then allowed to dehydrate to just below 20% of pot water-holding capacity, which took 4 d, and then were watered each day thereafter up to 20% of pot water-holding capacity for a 10-day period. Thus, the stress treatment consisted of a 4-d dehydration period followed by 10 d of prolonged stress. Pots from well watered treatments were watered daily to their pot water holding capacity. The experiment was completely randomized with three replications.

Gas Exchange Measurements. Simultaneous measurements of

<sup>&#</sup>x27;Journal article 5046 of the Oklahoma Agricultural Experiment Station. Financial support was provided by the Oklahoma Water Research Institute and by United States Department of Agriculture-Competetive Research Grants Office grants 84-CRCR-1-1461 and 83-CRCR-1-12 15.

A and transpiration (E) per unit leaf area were made on intact leaves using a stirred, temperature and humidity controlled reaction chamber described by Bingham et al. (2) and Coyne et al. (4). Humidity was measured inside the chamber with a condensation dew-point hygrometer (General Eastern 1 100DP, Watertown MA) and  $[CO<sub>2</sub>]$  by passing chamber exhaust through a differential  $CO<sub>2</sub>$  gas analyzer (Horiba PIR 2000 R, Irvine CA). Leaves were measured at 1800  $\mu$ mol PAR m<sup>-2</sup> s<sup>-1</sup>, 20°C, 21%  $O<sub>2</sub>$ , and 1 KPa vapor pressure (for some of the water stressed treatments with low E, vapor pressure was somewhat lower than <sup>1</sup> KPa). For each plant, gas exchange was measured on a single fully emerged leaf of a given tiller. In the first experiment all measurements were made at an ambient  $CO<sub>2</sub>$  concentration of 330  $\mu$ l/L. In the second experiment A versus C<sub>i</sub> curves were developed by measuring A at different ambient  $CO<sub>2</sub>$  concentrations of 0, 75, 160, 315, 330, 380, 480, and 573  $\mu$ l/L. Values of A, E, diffusive conductance, and  $C_i$  were calculated according to von Caemmerer and Farquhar (3).

Water Potential. Immediately after gas exchange measurements were completed on a given leaf, leaf water potential was measured as described by Johnson *et al.* (8). Two  $0.24 \text{ cm}^2$  leaf discs were removed using leaf cutter psychrometers, sealed in a psychrometer chamber, and allowed to come to temperature and vapor equilibrium in a 30°C water bath. Wet bulb depression was then determined using a microvolt meter (Wescor HP-115, Logan, UT). The resulting readings were used to calculate water potential for the individually calibrated psychrometers.

Stomatal Frequency. In experiment 2, adaxial and abaxial leaf impressions were made by applying clear fingernail polish to the midportions of fully emerged leaves. After removing the dried transparent images, the number of stomata in a  $6.5 \text{ mm}^2$  grid were counted under a microscope. For each species, four different leaves were counted on each plant from each of the four blocks.

#### RESULTS

Under well watered conditions and at an ambient  $[CO<sub>2</sub>]$  of 330  $\mu$ l/L, T. kotschyi had a higher rate of A than T. aestivum (Table I). The higher A of T. kotschyi was associated with higher conductance, lower water use efficiency, and higher values of  $C_i$ compared to  $T$ . *aestivum* (Table I). The lower water use efficiency and higher  $C_i$  of  $T$ . kotschyi was again observed under well watered conditions in experiment 2 (data not shown).

Reduction in water potential significantly decreased A without a significant interaction with species-that is, the average difference in A between  $T$ . *aestivum* and  $T$ . *kotschyi* was, within experimental error, maintained as water potential declined (Fig. 1). As expected, the reduction in leaf water potential led to a reduction in leaf conductance (Fig. 1). The higher conductance of T. kotschyi than T. aestivum observed under well watered conditions (Table I) was also observed at lower water potentials and, as with A, there was no significant water potential  $\times$  species interaction. Although the difference was smaller as water potential declined, the higher A and conductance of T. kotschyi than T. aestivum was maintained over the entire range of water potential values from  $-0.5$  to  $-2.9$  MPa. The stomatal frequency for leaves of T. kotschyi  $(34 \text{ per mm}^2 \text{ a}$  daxial and 42 abaxial) and T. aestivum (32 per mm<sup>2</sup> adaxial and 43 abaxial) were nearly identical, so the difference between species in conductance was not directly related to stomatal frequency. The higher C<sub>i</sub> observed in T. kotschyi than T. aestivum under well watered conditions (Table I) was not observed under water deficits. There was, however, a significant effect of water potential on  $C_i$ . The average Ci of both species first declined as water potential declined but then increased sharply past  $-1.9$  MPa (Fig. 2).

In a separate experiment, A versus  $C_i$  response curves were developed to determine more precisely the role of conductance as compared to the capacity for mesophyll photosynthesis under well watered and stressed conditions. As in previous experiments, T. kotschyi had significantly higher A at an ambient  $[CO<sub>2</sub>]$  of  $330 \mu l/L$  under both well watered and lower water potentials than did T. aestivum (Fig. 3). Under well watered conditions the A versus  $C_i$  response curves of the two species were quite similar. T. kotschyi did have somewhat higher A values than T. aestivum at  $C_i$  values above about 150  $\mu$ l/L. Nevertheless, most of the higher A of T. kotschyi under well watered conditions measured at an ambient  $[CO_2]$  of 330  $\mu$ l/L (Fig. 3) was associated with the higher conductance of T. kotschyi (0.46 mol H<sub>2</sub>O m<sup>-2</sup> s<sup>-1</sup>) than T. aestivum (0.24 mol  $H_2O$  m<sup>-2</sup> s<sup>-1</sup>). This higher conductance apparently led to the higher  $C_i$  in T. kotschyi compared to T. *aestivum* at an ambient  $[CO_2]$  of 330  $\mu$ l/L (Fig. 3). Thus, under well watered conditions, the higher stomatal conductance of T. kotschyi than T. aestivum, which was also observed in previous experiments, apparently led to most of the higher A in T. kotschyi. In the stressed treatments, however, the higher A of T. *kotschyi* than T. *aestivum* at an ambient  $[CO_2]$  of 330  $\mu$ l/L was mostly the result of differences in the A versus  $C_i$  response curves rather than conductance. Under water deficits, T. kotschyi had <sup>a</sup> higher A mostly because its response curve had <sup>a</sup> higher A plateau at a given  $C_i$  than that of T. *aestivum* (Fig. 3). As in the second experiment (Fig. 2) there was a shift upward in the  $C_i$  at an ambient  $[CO_2]$  of 330  $\mu$ l/L under stress. Water deficits also caused an increase in the  $CO<sub>2</sub>$  compensation concentration from about 25 to 65  $\mu$ l/L (Fig. 3).

# **DISCUSSION**

Our results showing Triticum kotschyi to have higher A than Triticum aestivum at near current atmospheric  $CO<sub>2</sub>$  concentrations and over a range of water potentials is consistent with the results of Shimshi et al. (13). In that study, however, they could not attribute the higher A to stomatal control or to <sup>a</sup> higher cellular or subcellular capacity for photosynthesis. Since the A versus C<sub>i</sub> response curves under well watered conditions were quite similar between species (Fig. 3), the higher conductance of T. kotschyi resulted in a higher C<sub>i</sub> at 330  $\mu$ l/L ambient  $[CO_2]$ 

Table I. Leaf Gas Exchange Characteristics for the Leaves of Two Triticum Species

The measurements were under well watered conditions at 20°C, 1800  $\mu$ mol quanta m<sup>-2</sup> s<sup>-1</sup>, 21% O<sub>2</sub>, and an ambient vapor pressure of <sup>1</sup> KPa.



<sup>a</sup> Water use efficiency as the ratio of A to leaf transpiration. b Least significant difference at 5% probability level.



FIG. 1. Assimilation of  $CO<sub>2</sub>(A)$  and conductance to water vapor as a function of leaf water potential for T. kotschyi and T. aestivum. Measurements were made at 330  $\mu$ I/L ambient  $[CO_2]$  concentration, a leaf temperature of 20°C, 1800  $\mu$ mol PAR m<sup>-2</sup> s<sup>-1</sup>, 21% O<sub>2</sub>, and an average vapor pressure of <sup>I</sup> KPa. There was no significant effect of species on leaf water potential at a given water potential level  $(P = 0.93)$  so water potentials were averaged within each water potential level. Both species and dehydration effects were highly significant for A and conductance  $(P < 0.01)$  but there was no significant species  $\times$  dehydration level interaction for either A ( $P = 0.81$ ) or conductance ( $P = 0.64$ ). Each point is the mean of measurements of four recently emerged leaves from different plants.

than in T. aestivum. T. kotschyi had more  $CO<sub>2</sub>$  available to mesophyll tissue than T. aestivum, which lead to a higher rate of A. Thus, leaf conductance was more important in controlling A than the mesophyll capacity for photosynthesis under well watered conditions.

Under water deficits, however, there was little difference in the  $C_i$  at an ambient  $[CO_2]$  of 330  $\mu$ l/L between species (Figs. 2 and 3) and the higher A of T. kotschyi appeared to be associated with its ability to maintain a higher mesophyll capacity for photosynthetic expressed as a higher plateau in the A versus  $C_i$  response curve (Fig. 3). Interpretation of the higher A plateau using the model of Farquhar et al. (6) suggests a maintenance of ribulose bisphosphate regeneration capacity in T. kotschyi under water deficits, which could in turn be related to a number of factors. Mayoral et al. (11) report that ribulose bisphosphate carboxylase activity, electron transport rates, and photophosphorylation of isolated chloroplasts of T. kotschyi were maintained to lower water potential levels than in T. aestivum.

With moderate water deficits C<sub>i</sub> declined as water potential declined (Fig. 2) indicating that reductions in conductance were relatively greater than reductions in the mesophyll capacity for photosynthesis. This could result in an increased stomatal limi-



FIG. 2. Average internal  $[CO_2]$  ( $C_1$ ) as a function of leaf water potential for T. kotschyi and T. aestivum. The species effect was not significant (P  $= 0.80$ ) so the average C<sub>i</sub> for the species is presented for each water potential level. Water potential level had a significant effect on  $C_i$  (P < 0.01) without a significant species  $\times$  C<sub>i</sub> interaction (P = 0.64). Measurement conditions were those given in Figure 1. Each point is the mean of eight recently emerged leaves from different plants.



FIG. 3. Assimilation of  $CO<sub>2</sub>$  (A) as a function of internal  $[CO<sub>2</sub>]$  (C<sub>i</sub>) in leaves of two Triticum species under well-watered  $(-0.75 \text{ MPa}, \text{SE} =$ 0.11) and water stressed  $(-2.20 \text{ MPa}, \text{SE} = 0.18)$  conditions. The arrows show the  $C_i$  value corresponding to an ambient  $[CO_2]$  of 330  $\mu$ l/L. Other measurement conditions were those given in Figure 1. Each point is the mean of three determinations on recently emerged leaves of different plants for the well watered treatment and two determinations for water stress treatment.

tation to A. Nevertheless, the reduction in  $C_i$  from an average of 242 to 196  $\mu$ l/L at -1.9 MPa was only 19%, whereas the average decline in A associated with that decline in water potential was 79% (Figs. <sup>1</sup> and 2). Thus, nonstomatal inhibition-damage to the plant's internal photosynthetic capability-was apparently the dominant factor reducing A under water deficits in both species. A faster relative decline in the mesophyll photosynthetic capacity than in conductance apparently occurred below  $-1.9$  $\widehat{MPa}$ , resulting in an increase in  $C_i$  values (Fig. 2). Even though  $C_i$  was not constant, there was still a close linear relationship between A and conductance  $(r = 0.96)$  observed in the second experiment (Fig. 1). The mechanism by which A and conductance are coupled is not known, but given the lack of coupling between  $C_i$  and conductance,  $C_i$  does not appear to be the principal controlling factor.

There was a shift in the  $CO<sub>2</sub>$  compensation concentration resulting from water deficits (Fig. 3). This has been previously reported for T. aestivum when the compensation concentration was measured by placing a leaf in an illuminated closed chamber until steady state  $CO<sub>2</sub>$  concentration was reached (9). A similar



FIG. 4. Relationship between transpiration  $(E)$  and  $CO<sub>2</sub>$  assimilation (A) in leaves of two Triticum species calculated as described by Dubbe et al. (5). Curves were developed using simultaneously measured values of A, E, and conductance associated with data from Figure 3. Arrows indicate the values for leaves undergoing gas exchange at an ambient [CO<sub>2</sub>] of 330  $\mu$ l/L and from species at a water potential of -0.75 MPa (well watered) and at  $-2.20$  MPa (stressed). Measurement conditions were those given in Figure 1.

increase was observed by Mawson and Colman (10) in osmotically stressed isolated mesophyll cells by Phaseolus and Lycopersicon. The increase in the  $CO<sub>2</sub>$  compensation concentration under water deficits could stem from an increase in dark respiration (9) or a decrease in the rate of carboxylation relative to oxygenation (10, 12).

If the partial differential  $\partial E/\partial A$ —the ratio of environmental gains of transpiration and assimilation-is large but constant, leaf carbon assimilation is maximal and the amount of water loss is high (5). When  $\partial E/\partial A$  is small, carbon assimilation is reduced, but the water loss required to take up carbon is minimized. Under well watered conditions the maintenance of higher conductance in T. kotschyi results in an  $\partial E/\partial A$  that is higher than that of  $T$ . *aestivum* (Fig. 4). Under water deficits there was an increase in  $\partial E/\partial A$  for both species, but T. aestivum increased more then T. kotschyi, and there was little difference in this gain ratio between species at reduced water potential levels. The ecological significance of this could be related to the adaptation of T. kotschyi to its desert environment. When water is available T. kotschyi apparently maximizes carbon gain at the expense of water conservation. But when water becomes limiting it has the ability to maintain more carbon assimilation than  $T$ . aestivum at a similar gain ratio.

Even though the gain ratio was not constant-it increased in association with water deficits- $T$ . kotschyi was as conservative with water for a given amount of A as was  $T$ . *aestivum* under

water deficits (Fig. 4). In an agricultural setting, provided that the boundary layer conductance in a crop canopy was large enough so that stomata had a significant influence over transpiration, the popular idea of developing wheat with higher water conservation potential-that is, higher water use efficiencymust be considered carefully. High water use efficiency, especially if it resulted from a greater capacity for mesophyll photosynthesis so that <sup>a</sup> high rate of A was maintained, may benefit productivity under water limited conditions. But as exemplified by T. kotschyi, lower water use efficiency results in maximum carbon assimilation when water is adequate and can be coupled with the maintenance of higher assimilation when water is limited. In certain drought prone environments, especially when the time for growth is limited, this may be a more effective way to maximize crop production than strict water conservation.

#### LITERATURE CITED

- 1. AUSTIN RB, CL MORGAN, MA FORD, SC BHAGWAT <sup>1982</sup> Flag leaf photosynthesis of Triticum aestivum and related diploid and tetraploid species. Ann Bot49: 177-189
- 2. BINGHAM GE, PI COYNE, RB KENNEDY, WL JACKSON <sup>1980</sup> Design and fabrication of a portable minicuvette system for measuring leaf photosynthesis and stomatal conductance under controlled conditions. Lawrence Livermore Laboratory UCRL-52895
- 3. CAEMMERER VON S, GD FARQUHAR <sup>1981</sup> Some relations between the biochemistry of photosynthesis and the gas exchange of leaves. Planta 153: 376- 387
- 4. COYNE PI, JA BRADFORD, CL DEWALD 1982 Leaf water relations and gas exchange in relation to forage production in four asiatic bluestems. Crop Sci 22: 1036-1040
- 5. DUBBE DR, GD FARQUHAR, KLAUS RASCHKE <sup>1978</sup> Effect of abscisic acid on the gain of the feedback loop involving carbon dioxide and stomata. Plant Physiol 62: 413-417
- 6. FARQUHAR GD, S VON CAEMMERER, JA BERRY 1980 A biochemical model of photosynthetic  $CO<sub>2</sub>$  assimilation in leaves of  $C<sub>3</sub>$  species. Planta 149: 78-90
- 7. HUTMACHER RB, DR KRIEG <sup>1983</sup> Photosynthetic rate control in cotton. Stomatal and nonstomatal factors. Plant Physiol 73: 658-661 JOHNSON RC, HT NGUYEN, RW MCNEW, DM FERRIS 1986 Sampling error
- for leaf water potential measurements in wheat. Crop Sci 26: 380-383
- LAWLOR DW 1976 Water stress induced changes in photosynthesis, photorespiration, respiration and CO<sub>2</sub> compensation concentration of wheat. Photosynthetica 10: 378-387
- 10. MAWSON BT, B COLMAN 1983 The inhibition of photosynthesis and photorespiration in isolated mesophyll cells of Phaseolus and Lycopersicon by reduced osmotic potentials. Physiol Plant 57: 21-27
- <sup>1</sup> 1. MAYORAL ML, D ATSMON, Z GRoMET-ELHANAN, D SHIMSHI <sup>1981</sup> The effect of water stress on various enzymatic activities in wheat and related wild species, carboxylase activity, electron transport, and photophosphorylation in isolated chloroplasts. Aust J Plant Physiol 8: 385-394
- 12. OGREN WL <sup>1984</sup> Photorespiration: pathways, regulation, and modification. Annu Rev Plant Physiol 35: 415-442
- 13. SHIMSHI D, ML MAYORAL, D ATSMON <sup>1982</sup> Responses to water stress in wheat and related wild species. Crop Sci 22: 123-128
- 14. WONG SC, IR COWAN, GD FARQUHAR <sup>1979</sup> Stomatal conductance correlates with photosynthetic capacity. Nature 282: 424-426