Effects of Substerilization Doses of Co⁶⁰ Gamma Radiation on the Cold-Storage Life Extension of Shucked Soft-Shelled Clams and Haddock Fillets

E. B. MASUROVSKY, S. A. GOLDBLITH, AND J. T. R. NICKERSON

Department of Nutrition and Food Science, Massachusetts Institute of Technology, Cambridge, Massachusetts

Received for publication 31 December 1962

ABSTRACT

MASUROVSKY, E. B. (Massachusetts Institute of Technology, Cambridge), S. A. GOLDBLITH, AND J. T. R. NICKERSON. Effects of substerilization doses of Co⁶⁰ gamma radiation on the cold-storage life extension of shucked soft-shelled clams and haddock fillets. Appl. Microbiol. 11:220-228. 1963.-Total aerobic-facultative and anaerobic (clostridia) macrocolony count data are presented, with analyses and interpretation, for both haddock fillets and shucked soft-shelled clams which received doses of from 50,000 to 800,000 rad of Co⁶⁰ gamma rays. These data indicated that haddock fillets may be maintained in good condition at refrigeration temperatures above freezing for about 1 week at 6 C, and approximately 2 weeks at 0 C, when treated with from 50,000 to 150,000 rad of ionizing radiation. In the dose range from 200,000 to 350,000 rad, the storage life may be extended up to some 2 weeks at 6 C, and 3 weeks at 0 C. Treatments in the dose range from 400,000 to 500,000 rad may defer spoilage for about 1 month, and doses of 550,000 to 650,000 rad afford protection against bacterial spoilage up to approximately 1.5 months. At the high substerilization doses of 700,000 to 800,000 rad, haddock fillets may be held for from 2 to 3 months in refrigerated storage before becoming unfit for marketing and consumption. Shucked soft-shelled clams can be held for about 2.5 weeks at 0 C and close to 12 days at 6 C, when given low substerilization doses of from 50,000 to 150,000 rad of ionizing radiation. At doses of from 200,000 to 350,000 rad, the clams may be preserved effectively for periods up to 3 weeks at 0 or 6 C, and some 6 weeks at these temperatures with doses of about 450,000 rad. With treatments of 500,000 to 600,000 rad, the storage life may be extended for some 2 months, and at doses of 650,000 to 800,000 rad the shucked clams remain in a good state of preservation for up to 3 months at temperatures of 0 to 6 C. Thus, it would appear that shucked soft-shelled clams may be maintained for significantly longer periods in refrigerated storage than haddock fillets when the same radiation treatments are applied to each product. Clostridia levels in both products were relatively low initially, and were reduced significantly by the gamma rays at the doses studied. Moreover, those clostridia that survived the radiation treatments were found to remain at safe, low levels during the various periods in refrigerated storage employed for these products, a very encouraging result from the public health, as well as commercial, standpoint.

The fisheries industry has long cherished the hope of supplying inaccessible, inland markets with "fresh" or "freshlike" sea food products, and of balancing the supply and demand of marine products at coastal markets over substantially longer periods of time than present commercial practices permit. In recent years, with the great strides made in harnessing the power of the atom for peaceful purposes, the means for transforming this hope into reality appears to have excellent possibilities.

The pioneering work that demonstrated the feasibility of extending the storage life of fresh sea foods by treatment with substerilization doses of atomic radiation was carried out by Nickerson and Goldblith (1950), Nickerson et al. (1954), and Shewan and Liston (1958). Since the principal cause of sea food spoilage, at storage temperatures above freezing, is microbial in origin, and since potentially pathogenic microbes, such as Clostridium botulinum type E (Dolman, 1957), Staphylococcus aureus (Nickerson et al., 1962), and certain members of the Enterobacteriaceae (Raj, Wiebe, and Liston, 1961), are occasionally encountered in some marine foodstuffs, a major research program has been undertaken at the Department of Nutrition and Food Science, Massachusetts Institute of Technology, to study the effects of ionizing radiation on the inactivation of these microorganisms in situ.

In this communication, data are presented from investigations concerned with the total numbers of aerobicfacultative and obligately anaerobic microorganisms obtained from two commercially important sea foods, shucked soft-shelled clams and haddock fillets, before and at various periods of time after irradiation and storage at 0 and 6 C. The attendant changes in the constitution of the microflora of these marine products, and the deleterious effects (spoilage, etc.) attributable to those changes, are reported in the second paper of this series (Masurovsky, Voss, and Goldblith, 1963). These studies constitute the first phases of an integrated program intended to bring to realization the potentialities for commercial preservation of sea foods by ionizing radiation.

MATERIALS AND METHODS

General. Macrocolony counts of the aerobic-facultative microorganisms associated with these foodstuffs were made from samples plated out on both distilled water and synthetic sea water culture media. Preliminary investigations with over a score of different culture media (Department of Nutrition and Food Science, Massachusetts Institute of Technology, 1961) had shown that, although maximal plate counts often could be obtained from these sea foods on a suitable distilled water culture medium, the generic constitution of the microflora was subject to rather wide fluctuations, thus indicating the prudence of employing a synthetic sea water culture medium as well.

Anaerobic (clostridia) macrocolony counts were made with the medium of Mossel et al. (1955), as modified by Angelotti and Hall (1961).

Irradiation of the haddock and clams was carried out in a Mark I Co⁶⁰ Food Irradiator, designed as a prototype commercial unit by the U.S. Atomic Energy Commission and the Brookhaven National Laboratories, and installed in the facilities of the Department of Nutrition and Food Science of the Massachusetts Institute of Technology. The source contained some 32,000 c of Co⁶⁰ at the time of installation, and had a food-irradiation capacity of about 320 lb per day at an average dose level of 1 Mrad. The studies reported herein were conducted in the substerilization dose range from 50,000 to 800,000 rad.

Irradiated and control samples of both sea foods to be stored for different periods of time were placed in metal containers maintained at the temperature of melting ice (0 C) and at $6.0 \pm 0.5 \text{ C}$. These temperatures were selected as representative of the lower and higher ranges of storage temperature employed in regular commercial food channels.

Microbiological assay of irradiated and stored haddock fillets and shucked soft-shelled clams: sampling procedure. Haddock fillets and shucked soft-shelled clams were taken from the processing lines of a filleting and a shucking plant in the greater Boston area known by a previous survey of fishery establishments (Nickerson, Goldblith, and Masurovsky, J. Milk and Food Technol., in press) to provide quality products. These samples were packed in clean containers surrounded by crushed ice and quickly transported to the laboratory for assaying or radiation processing, or both.

Preparation of samples for irradiation. Samples (50 g), selected at random from a given lot of haddock fillets and shucked soft-shelled clams, were aseptically cut, weighed, and transferred into radiation-sterilized ($\geq 2 \times 10^6$ rad) polyethylene film envelopes. These envelopes were heat-sealed and over-wrapped and sealed with a second layer

of polyethylene film. The finished package unit was approximately 7.5 by 7.5 by 2.0 cm. Five replicate packages of each sea food were prepared in this manner for each irradiation dose, storage temperature, and storage time. Unirradiated control samples were prepared in the same manner as the units which were to be irradiated. All samples were then cooled to about 5 C.

Samples designated for irradiation were secured to a specially designed rack (made entirely of wood to minimize gamma photon attenuation and scatter) which was positioned in the center of the middle insulated irradiation chamber. In this position, the samples received as uniform a dose of radiation as was possible according to the geometry of the Co⁶⁰ rods in the source.

Microbiological assay procedure. The irradiated or control packages were carefully opened, the polyethylene film envelopes containing the haddock or clam sections were aseptically opened in turn, and the 50 g of sea food were aseptically transferred to a sterile, precooled Waring Blendor cup. The envelopes were next rinsed out three times with sterile, refrigerated diluent into the same Waring Blendor cup. A sample plating diluent contained (g/liter of distilled water): Trypticase (BBL), 1.0; KH₂-PO₄, 0.0425; and Antifoam AF (Dow Corning Corp.), 0.01. The final pH of the emulsion was 7.2 ± 0.1 . A total of 450 ml of diluent was put into a given blender cup together with the sea food sample, and blended for 3 min at 17,000 rev/min.

Aerobic-facultative plating procedure. Appropriate decimal dilutions were made from the blended samples by use of sterile, precooled diluent; 0.1 ml from these dilutions was aseptically pipetted onto the surface of previously poured agar culture dishes of medium B(DW) and medium B(SSW).

Medium B(DW) was that medium found in previous investigations (Department of Nutrition and Food Science, Massachusetts Institute of Technology, 1961) to give the highest average macrocolony counts for both haddock fillets and shucked soft-shelled clams. It was prepared with distilled water (DW). Medium B(SSW) contained the same basal complex organic nutrient and energy sources as medium B(DW), but was prepared with a solution of inorganic salts of such a composition and concentration, as recommended by MacLeod and Onofrey (1956), to provide an optimal ionic environment for those microorganisms of marine origin which require some of the elements present in sea water.

The surface of the agar in these culture dishes had been dried at 37 C for 24 hr and cooled to 20 C prior to the above inoculation. The 0.1-ml inoculum was next spread over the surface of the surface-dried agar with silicone (Siliclad)-treated glass rods, by the method of Buck and Cleverdon (1960), to distribute the organisms as homogeneously as possible, and to provide optimal aerobic conditions. Three replicate culture plates were prepared for each dilution and each medium. Separate glass rods were employed for spreading the inoculum in each dish. These petri-dish cultures were incubated for 5 days at 20 C before observation and enumeration of the macrocolonies which developed.

Anaerobic tube procedure. The anaerobic microflora of these same samples were cultured and scored in the sulfite-polymixin-sulfadiazine agar of Angelotti and Hall (1961), using Pyrex no. 9200 oval tubes with a thioglycolate agar overlay. Three replicate tubes were made from the 1:10 dilution of each sample, and incubated at 30 and 37 C for 40 hr. Black zone-forming macrocolonies were analyzed for catalase activity, morphology, and spore formation subsequent to enumeration as a *Clostridium* sp.

RESULTS AND DISCUSSION

Haddock fillets. Total aerobic-facultative and anaerobic (clostridia) macrocolony count data for haddock fillets irradiated at doses of from 50,000 to 350,000 rad are presented in Tables 1 and 2.

Me-

dium

DW^b

SSW¢

DW

SSW

DW

SSW

DW

SSW

DW

SSW

595

199

228

173

149

107

426

278

112

104

50.000

 $\times 10^3$

 $\times 10^3$

 $\times 10^2$

 $\times 10^{2}$

 $\times 10^4$

 $\times 10^4$

 $\times 10^2$

 $\times 10^2$

 $\times 10^{6}$

 $\times 10^{6}$

Stor-

age temp

С

6

n

6

Treatment

Before irradiation

radiation

Immediately after ir-

After 3 days of refrig-

After 6 days of refrig-

erated storage

erated storage

The aerobic-facultative plate counts were less than 25×10^3 viable microorganisms per g of haddock tissue immediately after irradiation for all the aforementioned doses, which was equivalent to the inactivation of approximately 96% of the original commensal and contaminating microflora.

Results from storage studies with the irradiated fillets indicated that aerobic-facultative counts remained below 110×10^4 /g at 6 C and 40 $\times 10^3$ /g at 0 C during the first week after treatments with doses of 200,000 to 350,000 rad of Co⁶⁰ gamma rays. In the dose range from 50,000 to 150,000 rad, the counts rose to over 1 million per g at 6 C and into the tens of thousands per g at 0 C, after about 1 week in storage.

By the 12th day of storage at 6 C, macrocolony counts of well into the millions per g of haddock tissue were obtained from the entire series of doses from 50,000 to 350,000 rad. However, fillets receiving doses of 200,000 to 350,000 rad, when stored at 0 C, did not show aerobic-

250.000

 55.5×10^{1}

 52.5×10^{1}

 41.8×10^{2}

 23.6×10^{2}

 21.3×10^{2}

 13.7×10^{2}

 70.1×10^4

 45.3×10^{4}

336

197

 $\times 10^3$

 $\times 10^3$

300,000

 49.1×10^{1}

 36.5×10^{1}

 45.2×10^{2}

 28.5×10^{2}

 23.7×10^{2}

 15.3×10^{2}

 73.9×10^{4}

 50.1×10^{4}

445

341

 $\times 10^3$

 $\times 10^3$

350.000

 $\times 10^3$

 $\times 10^3$ 30.5×10^{1}

 21.5×10^{1}

 22.9×10^2

 17.9×10^{2}

 14.7×10^{2}

 11.1×10^{2}

_

445

341

TABLE 1. Aerobic-facultative macrocolony counts obtained from haddock fillets before and after treatment with Co⁶⁰ gamma rays and storage at 6 or 0 C for 24 days^a

294

204

150,000

 56.0×10^{2}

 56.5×10^{2}

 18.1×10^{4}

 15.7×10^{4}

 73.5×10^{2}

 56.9×10^{2}

 73.1×10^{5}

 49.9×10^{5}

 $\times 10^3$

 $\times 10^3$

100,000

 $\times 10^3$

 $\times 10^3$

 $\times 10^2$

 $\times 10^2$

 $\times 10^2$

 $\times 10^2$

 42.1×10^{4}

 37.5×10^{4}

 78.8×10^{5}

 63.6×10^{5}

294

204

169

127

115

103

Absorbed dose (rad)

200,000

 12.6×10^{2}

 9.27×10^{2}

 63.6×10^{2}

 51.2×10^{2}

 20.7×10^{2}

 14.1×10^{2}

 85.5×10^{4}

 $\times 10^4$

 $\times 10^3$

 $\times 10^3$

336

197

109

-	0	DW	399	X	104	64.9	×	10 ³	50.7	×	10 ³	182	×	10 ²	38.1	×	10 ³	18.8	Х	10 ³			
		SSW	279	×	104	60.5	Х	10 ³	38.6	×	10 ³	159	х	10 ²	22.0	х	103	12.8	Х	10 ³			
After 9 days of refrig-	6	DW		≫	106	148	Х	106	103	×	106	127	х	105	66.5	×	105	37.0	Х	105	30.	$2 \times$	105
erated storage ^d		SSW				120	х	106	82.3	×	106	119	х	105	59.2	×	105	32.9	Х	105	26.	ЗX	105
	0	DW	55.7	X	106	113	х	104	77.5	×	104	53.5	х	10 ³	39.6	×	10 ³	50.1	Х	10 ³	45.	$5 \times$	10 ³
		SSW	50.1	ιx	106	103	х	104	67.1	×	104	41.3	х	10 ³	26.9	×	10 ³	46.8	Х	10 ³	38.	$5 \times$	10 ³
After 12 days of refrig-	6	DW					≫	106		≫	106		≫	106		≫	106	-			58.	5 X	106
erated storage		SSW																-			46.	ЭX	106
-	0	DW	· ·			119	×	105	74.7	×	105	133	×	10 ³	72.6	×	10 ³	-			30.	$1 \times$	104
		SSW				109	×	105	52.9	×	105	120	×	10 ³	54.0	×	10 ³	-			17.	$\delta \times$	104
After 15 days of refrig-	6						≫	106		\gg	106		≫	106		\gg	106		\gg	106			
erated storage																						—	
	0	DW				83.8	Х	106	553	×	106	205	Х	104	126	х	104	89.1	Х	104			
		SSW				57.5	×	106	484	×	106	182	х	104	110	×	104	56.7	Х	104			
After 18 days of refrig-	6												≫	106		≫	106		\gg	106		≫	106
erated storage	0	DW										30.7	х	106	204	×	105	128	Х	105	413	×	104
		SSW										18.6	Х	106	176	х	105	121	Х	105	396	×	104
After 24 days of refrig-	6												≫	106		\gg	106		\gg	106		≫	106
erated storage	0	DW										189	Х	106	70.5	×	106	116	Х	106	78.	3 ×	106
		SSW										179	×	106	64.5	×	106	97.2	Х	106	63.	$1 \times$	106
	1	1	1						1			1			1								

^a Average (\bar{X}) of five replicate samples plated in triplicate (X = 15) and incubated for 5 days at 20 C. Results expressed as count per g of medium.

^b Distilled water culture medium.

^c Synthetic sea water culture medium.

^d Readings for 100,000, 150,000, 200,000, and 250,000 rad doses were taken after 8 days of storage.

 TABLE 2. Anaerobic (clostridia) macrocolony counts obtained from haddock fillets before and after treatment with Co⁶⁰ gamma rays and storage at 6 or 0 C for 24 days^a

Treatment Before irradiation Emmediately after irradiation After 3 days of refrigerated storage After 6 days of refrigerated storage After 9 days of refrigerated storage After 12 days of refrigerated storage After 15 days of refrigerated storage After 18 days of refrigerated storage After 24 days of refrigerated	Storage			A	bsorbed dose (rad	i)		
1 reatment	temp	50,000	100,000	150,000	200,000	250,000	300,000	350,000
	C							
Before irradiation		1.73	1.54	1.54	4.13	4.13	3.73	3.73
Immediately after irradiation		$\sim \! 1.00^{b}$	~1.00	~1.00	~1.00	~1.00	~1.00	<1.00°
After 3 days of refrigerated	6	~ 1.00	~1.00	~1.00	~1.00	~1.00	~1.00	~1.00
storage	0	~ 1.00	~ 1.00	~ 1.00	~1.00	~ 1.00	~1.00	~1.00
After 6 days of refrigerated	6	~ 1.00	~1.00	~ 1.00	~1.00	~ 1.00	~ 1.00	—
storage	0	~ 1.00	~1.00	~ 1.00	~1.00	~1.00	~ 1.00	
After 9 days of refrigerated	6		~1.00	~1.00	~1.00	~1.00	~1.00	~1.00
storage	0	~ 1.00	~1.00	~ 1.00	~1.00	~1.00	~1.00	~1.00
After 12 days of refrigerated	6			_	_	-		~1.00
storage	0		~1.00	~1.00	~1.00	~ 1.00		~1.00
After 15 days of refrigerated	6			_	_	_	_	
storage	0		~1.00	~1.00	~1.00	~1.00	~1.00	—
After 18 days of refrigerated	6					_		
storage	0				~1.00	~ 1.00	~1.00	~ 1.00
After 24 days of refrigerated	6						_	
storage	0				~1.00	~1.00	~1.00	~1.00

^a Average (\bar{X}) of five replicate samples tubed in triplicate (X · = 15) and incubated at 30 and 37 C for 40 hr. Results expressed as (count \times 10)/g.

^b At \sim 1.00, several, but not all, replicate anaerobic culture tubes displayed at least one *Clostridium* macrocolony.

^c At <1.00, no anaerobic culture tubes displayed a single *Clostridium* macrocolony.

^d Counts at 100,000, 150,000, 200,000, and 250,000 rad levels were taken after 8 days of storage.

TABLE 3.	. Aerobic-facultative	macrocolony	counts ol	btained from	haddock	fillets	before and	l after	treatment	with	C0 ⁶⁰	gamma	rays	and st	orage
				at 6 or	$\cdot 0 C for$	3 mon	thsa								

Treatment	Stor-	Med-				Absorbed	dose (rad)			
Treatment	temp	ium⁵	450,000	500,000	550,000	600,000	650,000	700,000	750,000	800,000
	C									
Before irradia-		DW	572×10^{3}	129×10^{3}	191×10^{3}	91.3×10^{3}	245×10^{3}	118×10^{3}	45.5×10^{3}	118×10^{3}
tion		SSW	325×10^{3}	81.5×10^{3}	88.8×10^{3}	57.7×10^{3}	127×10^{3}	49.9×10^{3}	18.7×10^{3}	49.9×10^{3}
Immediately		DW	19.4×10^{1}	39.5×10^{1}	11.9×10^{1}	21.3×10^{1}	27.3×10^{1}	22.4×10^{1}	11.5×10^{1}	13.5×10^{1}
after irradia- tion		SSW	17.5×10^{1}	24.3×10^{1}	11.2×10^{1}	18.0×10^{1}	14.6×10^{1}	18.6×10^{1}	7.46×10^{1}	9.26×10^{1}
After 1 month	6	DW	$55.7 imes 10^{4c}$	23.4×10^{4c}	44.5×10^{5d}	46.8×10^{5}	283×10^{4}	129×10^{3}	108×10^{3}	155×10^{3}
of refrigerated		SSW	55.6×10^{4}	21.6×10^{4}	41.0×10^{5}	46.1×10^{5}	253×10^{4}	146×10^{3}	97.1×10^{3}	146×10^{3}
storage	0	DW	$74.1 imes 10^2$	67.1×10^2	$32.0 imes 10^3$	67.1×10^{4}	63.8×10^{4}	$22.5 imes 10^3$	$66.9 imes 10^2$	68.3×10^{2}
		SSW	70.4×10^2	65.4×10^2	$29.1 imes 10^3$	60.3×10^{4}	58.5×10^{4}	18.7×10^3	$51.0 imes 10^2$	68.2×10^{2}
After 2 months	6	DW	77.9×10^{6e}	50.3×10^{6e}	218×10^{6}	33.8×10^{6}	203×10^{6}	36.7×10^{5}	50.7×10^{5}	60.3×10^{5}
of refrigerated		SSW	73.2×10^{6}	46.0×10^{6}	210×10^{6}	32.8×10^{6}	184×10^{6}	32.9×10^{5}	43.6×10^{5}	56.8×10^5
storage	0	DW	33.1×10^{5}	27.0×10^{6}	60.3×10^{6}	169×10^{5}	48.5×10^{5}	98.6×10^{3}	$52.5 imes 10^4$	27.2×10^4
		SSW	28.3×10^{5}	25.0×10^{6}	64.5×10^{6}	164×10^{5}	46.0×10^{5}	94.3×10^{3}	47.5×10^{4}	24.1×10^{4}
After 3 months	6	DW					_	198×10^{6}	222×10^{6}	199×10^{6}
of refrigerated		SSW	_			_		182×10^{6}	216×10^{6}	173×10^{6}
storage	0	DW		—				58.3×10^{6}	149×10^6	104×10^{5}
		SSW					-	54.3×10^{6}	137×10^{6}	99.3×10^{5}

^a Average (\hat{X}) of five replicate samples plated in triplicate (X · = 15) and incubated for 5 days at 20 C. Results expressed as count per g of medium.

 b DW = distilled water culture medium; SSW = synthetic sea water culture medium.

^c Readings taken after 2 weeks of storage.

^d Readings taken after 3 weeks of storage.

Readings taken after 1 month of storage.

TABLE 4.	Anaerobic	(clostridia)	macrocolony	counts	obtained	from	haddock	fillets	before	and	after	treatment	with	Co^{60}	gamma	rays d	ınd
				8	storage at	6 or (C for 3	month	as^a								

	Storage				Absorbed de	ose (rad)			
Ireatment	temp	450,000	500,000	550,000	600,000	650,000	700,000	750,000	800,000
	<i>C</i>			-					
Before irradiation		6.40	2.87	2.13	2.33	2.67	$\sim 1.00^{b}$	~1.00	~1.00
Immediately after irradiation		~ 1.00	~ 1.00	<1.00°	<1.00	<1.00	<1.00	<1.00	<1.00
After 1 month of refrigerated stor-	6	1.00 ^d	$\sim 1.00^{d}$	$\sim 1.00^{o}$	~ 1.00	~ 1.00	~1.00	~1.00	~ 1.00
age	0	~ 1.00	~ 1.00	~ 1.00	~ 1.00	~ 1.00	~ 1.00	~1.00	~ 1.00
After 2 months of refrigerated	6	$\sim 1.00'$	1.00	1.00	~ 1.00	~1.00	~1.00	~1.00	~ 1.00
storage	0	~ 1.00	~ 1.00	~1.00	~ 1.00	~ 1.00	~ 1.00	~1.00	~1.00
After 3 months of refrigerated	6			-			1.53	1.53	1.26
storage	0	—					~1.00	~1.00	~ 1.00

^a Average (\bar{X}) of five replicate samples tubed in triplicate (X · = 15) and incubated at 30 and 37 C for 40 hr. Results expressed as (count \times 10) per g.

^b At ~1.00, several but not all replicate anaerobic culture tubes displayed at least one *Clostridium* macrocolony.

^c At <1.00, no anaerobic culture tubes displayed a single *Clostridium* macrocolony.

^d Counts taken after 2 weeks of storage.

• Counts taken after 3 weeks of storage.

¹ Counts taken after 1 month of storage.

TABLE 5. Aerobic-facultative macrocolony counts obtained from haddock fillets treated with a 400,000-rad dose of Co[®] gamma rays and subsequently stored at 0 or 6 C for some 2 weeks*

		At 0 0	C		At 6 C	
Me- dium†	Stor- age	Irradiated samples (4 × 10 ⁵ rad)	Nonirradiated controls	Stor- age	Irradiated samples (4 × 10 ⁵ rad)	Nonirradiated controls
	days			days		
DW	0	35.9×10^{1}	36.7×10^{4}	0	35.9×10^{1}	36.7×10^4
SSW		27.3×10^{1}	$21.5 imes 10^4$		$27.3 imes 10^1$	21.5×10^{4}
DW	5	25.5×10^{2}	46.4×10^{5}	4	$29.2 imes 10^2$	$28.4 imes 10^5$
SSW		15.8×10^2	36.3×10^{5}		$20.3 imes10^2$	$18.3 imes 10^5$
DW	8	75.1×10^{2}	$12.5 imes 10^{6}$	7	$10.6 imes 10^3$	$27.0 imes 10^{6}$
SSW		68.7×10^2	61.8×10^{5}		$98.0 imes10^2$	$16.5 imes 10^{6}$
DW	13	61.7×10^{3}		12	$64.3 imes10^4$	24.3×10^{7}
SSW		$56.0 imes 10^3$			$51.9 imes 10^4$	$17.2 imes 10^7$
DW	15	37.7×10^{4}	10.5×10^{7}	14	$94.3 imes 10^5$	47.9×10^7
SSW		27.7×10^{4}	87.4×10^{6}		$87.3 imes 10^5$	41.3×10^{7}

* Average (\bar{X}) of five replicate samples plated in triplicate $(X \cdot = 15)$ and incubated for 5 days at 20 C. Results expressed as count/g of medium.

 \dagger DW = distilled water culture medium; SSW = synthetic sea water culture medium.

facultative counts in the millions until over 1 week later. Thus, fillets treated at these dose levels, and subsequently stored at the temperature of melting ice, may be kept in a microbiologically satisfactory state of preservation for about 2 to 3 weeks. On the other hand, at doses of 50,000 to 150,000 rad, no more than 9 to 15 days of storage at 0 C may elapse before the microbial populations rise into the millions per g and cause serious spoilage of the product.

Turning to the higher substerilization dose range, from 400,000 to 800,000 rad, one finds significantly longer

	At 0 C			At 6 C	
Storage	Irradiated samples $(4 \times 10^{5} \text{ rad})$	Nonirradiated controls	Storage	Irradiated samples (4 × 10 ⁵ rad)	Nonirradiated controls
days			days		
0	0	~1.00	0	0	~ 1.00
5	~ 1.00	~ 1.00	4	~ 1.00	1.50
8	~ 1.00	1.14	7	~ 1.00	1.70
13	~ 1.00	1.44	12	~ 1.00	1.90
15	~ 1.00	2.46	14	~ 1.00	2.31

TABLE 6. Anaerobic (clostridia) macrocolony counts obtained from

haddock fillets treated with a 400,000-rad dose of Co⁶⁰ gamma

rays and subsequently stored at 0 or 6 C for some 2 weeks*

* Average (\bar{X}) of five replicate samples tubed in triplicate $(X \cdot = 15)$ and incubated at 30 and 37 C for 40 hr. Results expressed as count \times 10 per g of medium.

storage times to have passed before the bacterial growth in the product reached high population levels. Macrocolony count data for this dose range are presented in Tables 3 to 6.

Aerobic-facultative plate counts were generally less than 400 organisms per g shortly after irradiation over this entire dose range, or better than 99% of the original microflora was destroyed by the absorption of these levels of ionizing energy.

Data from storage studies with the fillets irradiated at doses of 700,000 to 800,000 rad showed that aerobicfacultative counts stayed below $1.5 \times 10^5/g$ at 6 C and $<2.5 \times 10^4/g$ at 0 C, up to and including 1 month of storage, but increased to about $6.0 \times 10^6/g$ at 6 C and $<55 \times 10^4/g$ at 0 C after 2 months of storage.

By contrast, the aerobic-facultative macrocolony counts of fillet sections treated with from 550,000 to 650,000 rad of ionizing radiation reached approximately $50 \times 10^5/g$ at 6 C and $\leq 67 \times 10^4/g$ at 0 C after only 1 month of storage. After 2 months in storage, the counts rose to some 20 to $200 \times 10^6/g$ at 6 C, and 50 to $150 \times 10^5/g$ at 0 C.

It is noteworthy that haddock fillets irradiated in the dose range from 450,000 to 500,000 rad had counts of about 50×10^4 /g at 6 C, and $\sim 70 \times 10^2$ /g at 0 C after only 2 weeks in storage, then became unfit for marketing after 1 month in storage, with counts in the millions per g at both 6 and 0 C. Such results are not surprising, however, in light of the microbial population levels of $\sim 90 \times 10^5$ /g at 6 C and $\sim 30 \times 10^4$ /g at 0 C that were reached after about 2 weeks in storage with fillet sections given a dose of 400,000 rad of ionizing radiation. Nevertheless, it does indicate that in the dose range from 400,000 to 500,000 rad, the useful refrigerated-storage life of haddock fillets

may be extended only from about 2 weeks to 1 month. In the dose range from 550,000 to 650,000 rad, the storage life is prolonged up to approximately 1.5 months, and at doses of 700,000 to 800,000 rad the fillets can be well maintained under refrigeration for from 2 to 3 months.

Anaerobic (clostridia) tubes were generally negative for the freshly irradiated samples over the entire dose range studied. The number of positive tubes began to increase after the samples had been stored for at least 1 week at 0 or 6 C. These counts were negligible for all practical purposes, a very promising result from the commercial and public health standpoints.

Shucked soft-shelled clams. In Tables 7 and 8 are presented total aerobic-facultative and anaerobic (clostridia) macrocolony count data for shucked soft-shelled clams

 TABLE 7. Aerobic-facultative macrocolony counts obtained from shucked soft-shelled clams before and after treatment with Co⁶⁰ gamma rays

 and storage at 6 or 0 C for up to 24 days^a

Treatment	Stor-	Med-			1	Absorbed dose (rad)		
Treatment	temp	ium ⁶	50,000	100,000	150,000	200,000	250,000	300,000	350,000
	С								
Before irradiation		DW	37.6×10^{3}	37.6×10^{3}	78.3×10^{3}	193×10^{3}	89.0×10^3	89.0×10^{3}	123×10^{3}
		SSW	$39.5 imes 10^3$	$39.5 imes 10^3$	60.1×10^{3}	152×10^{3}	61.8×10^{3}	61.8×10^{3}	134×10^{3}
Immediately after ir-		DW	78.6×10^{2}	$35.4 imes10^2$	$87.0 imes 10^2$	$38.5 imes 10^2$	$20.9 imes 10^2$	$15.4 imes 10^2$	131×10^{1}
radiation		SSW	$62.2 imes 10^2$	$29.5 imes 10^2$	$79.5 imes 10^2$	31.8×10^2	17.3×10^2	9.13×10^2	113×10^{1}
After 3 days of refrig-	6	DW	$82.5 imes 10^4$	263×10^{3}	478×10^{2}	$69.3 imes 10^2$	$45.6 imes 10^2$	33.0×10^{2}	$21.5 imes10^2$
erated storage		SSW	$52.1 imes 10^4$	147×10^{3}	335×10^{2}	38.6×10^2	$32.9 imes 10^2$	$19.6 imes 10^2$	$19.0 imes 10^2$
	0	DW	$18.9 imes 10^3$	$96.7 imes 10^2$	112×10^{2}	40.9×10^2	30.1×10^{2}	$25.2 imes 10^2$	$14.7 imes 10^2$
		SSW	$12.2 imes 10^3$	$58.4 imes10^2$	105×10^{2}	$36.7 imes 10^2$	$22.9 imes 10^2$	$19.3 imes 10^2$	$13.4 imes 10^2$
After 6 days of refrig-	6	DW	$89.0 imes 10^5$	$57.9 imes 10^5$	$30.7 imes 10^5$	134×10^{4}	$38.5 imes 10^3$	$78.9 imes 10^2$	$72.3 imes10^2$
erated storage ^c		SSW	64.2×10^{5}	$42.3 imes 10^5$	22.0×10^{5}	120×10^{4}	22.4×10^{3}	$40.8 imes 10^2$	$60.2 imes10^2$
	0	DW	$36.1 imes 10^4$	$62.1 imes 10^3$	260×10^{2}	106×10^{2}	57.7×10^{2}	$25.5 imes 10^2$	$26.0 imes 10^2$
		SSW	$22.8 imes10^4$	46.7×10^{3}	239×10^{2}	88.1×10^2	40.1×10^{2}	13.3×10^2	$12.5 imes 10^2$
After 9 days of refrig-	6	DW	537 $\times 10^{6}$	374×10^{6}	129×10^{5}	$80.7 imes 10^5$	$13.8 imes 10^4$	$73.1 imes 10^3$	$93.0 imes 10^3$
erated storage ^d		SSW	510×10^{6}	357×10^{6}	119×10^{5}	$75.8 imes 10^5$	11.5×10^{4}	$53.7 imes 10^3$	$72.5 imes10^{3}$
	0	DW	$26.2 imes 10^5$	19.7×10^{5}	$40.5 imes 10^3$	$45.9 imes 10^3$	16.3×10^{3}	$12.3 imes 10^3$	$37.1 imes 10^3$
		SSW	17.9×10^{5}	$15.0 imes 10^5$	$30.7 imes 10^3$	$44.9 imes 10^3$	11.1×10^{3}	$7.20 imes 10^3$	$34.5 imes 10^3$
After 12 days of re-	6	DW	$\gg 10^{6}$	$\gg 10^{6}$	124×10^{6}		$52.9 imes10^4$	—	$13.0 imes 10^4$
frigerated storage ^e		SSW			115×10^{6}	_	38.3×10^{4}		$12.5 imes 10^4$
	0	DW	$25.1 imes10^6$		$62.7 imes 10^3$	— —	29.5×10^{3}	_	$97.5 imes10^3$
		SSW	$15.1 imes 10^6$		$25.1 imes 10^3$		15.3×10^{3}	_	$94.5 imes 10^3$
After 15 days of re-	6	DW		$\gg 10^{6}$	$\gg 10^{6}$	122×10^{6}	76.3×10^{4}	$97.0 imes 10^4$	
frigerated storage		SSW				107×10^{6}	$62.7 imes 10^4$	81.7×10^4	
	0	DW		66.3×10^{5}	164×10^{4}	106×10^{3}	$32.8 imes 10^3$	63.9×10^3	
		SSW		$53.5 imes 10^5$	124×10^{4}	100×10^{3}	16.3×10^{3}	43.0×10^{3}	
After 18 days of re-	6	DW		$\gg 10^6$	$\gg 10^6$	263×10^{6}	857×10^{6}	23.7×10^{5}	$21.9 imes 10^5$
frigerated storage ^f		SSW				249×10^{6}	796×10^{6}	22.9×10^{5}	$20.5 imes 10^5$
	0	DW		178×10^{6}	$64.5 imes 10^6$	$52.6 imes 10^5$	$24.0 imes10^4$	111×10^{3}	$59.5 imes 10^4$
		SSW		161×10^{6}	$55.2 imes 10^6$	48.6×10^5	18.9×10^4	109×10^{3}	$58.7 imes 10^4$
After 24 days of re-	6	DW				$\gg 10^6$	$\gg 10^{6}$	33.6×10^{6}	42.1×10^{6}
frigerated storage ^g		SSW						$26.8 imes10^6$	$31.8 imes 10^6$
	0	DW				$28.7 imes 10^6$	$19.1 imes 10^6$	$25.1 imes 10^5$	$19.9 imes10^6$
		SSW				$18.5 imes 10^6$	$12.1 imes 10^6$	$18.6 imes10^{5}$	$14.9 imes 10^6$

^a Average (\bar{X}) of five replicate samples plated in triplicate (X · = 15) and incubated for 5 days at 20 C. Results expressed as count per g of medium.

^b DW, distilled water culture medium; SSW, synthetic sea water culture medium.

^c Counts at 250,000 rad ware taken after 5 days of storage.

^d Counts at 200,000 rad were taken after 10 days of storage.

^e Counts at 50,000 rad were taken after 13 days of storage.

¹ Counts at 200,000 rad were taken after 20 days of storage.

^g Counts at 300,000 rad were taken after 22 days of storage.

before and after treatment with Co^{60} gamma rays at doses of 50,000 to 350,000 rad and storage for 24 days at 0 and 6 C.

Approximately 98% of the initial commensal and contaminating aerobic-facultative microflora of this shellfish

TABLE 8. Anaerobic (clostridia) macrocolony counts obtained
from shucked soft-shelled clams before and after treatment
with Co^{60} gamma rays and storage at 6 or 0 C
for up to 24 days ^a

Treatment	Stor-			Absor	bed dose	(rad)		
Treatment	temp	50,000	100,000	150,000	200,000	250,000	300,000	350,000
Before ir-	С	13.2	13.2	15.2	28.7	24.9	24.9	38.1
Immedi- ately after ir- radiation		8.73	6.60	11.0	20.1	18.9	10.8	15.9
After 3 days of refriger- ated storage	6 0	10.3 7.93	8.40 6.60	9.53 7.13	26.3 17.1	12.6 9.20	13.9 8.47	17.6 16.6
After 6 days of refriger- ated storage ^b	6 0	8.93 6.33	13.6 8.73	12.3 8.47	9.20 7.87	14.1 9.06	11.0 6.86	18.0 16.2
After 9 days of refriger- ated storage ^c	6 0	11.7 6.87	8.73 5.93	10.1 8.27	$\begin{array}{c} 12.8 \\ 6.33 \end{array}$	12.3 7.86	11.7 9.40	18.5 15.7
After 12 days of refriger- ated storage ^d	6 0	5.73		6.87 5.87		10.6 7.80		18.5 15.3
After 15 days of refriger- ated storage	6 0		6.20	9.00 6.93	21.9 19.7	8.13 6.80	9.73 6.53	
After 18 days of refriger- ated	6 0		4.47	9.53	16.8 8.80	8.27 4.67	12.7 6.40	25.1 11.3
After 24 days of refriger- ated storage ⁷	6 0				9.00	8.33	10.5 9.47	12.6 8.93

^a Average (\bar{X}) of five replicate samples tubed in triplicate $(X \cdot = 15)$ and incubated at 30 and 37 C for 40 hr. Results expressed as count \times 10 per g.

 $^{\rm c}$ Counts at 200,000 rad were taken after 10 days of storage.

^d Counts at 50,000 rad were taken after 13 days of storage.

was inactivated at these moderate substerilization doses, just as in the case of the haddock fillet microflora. Shucked clams irradiated in the dose range from 200,000 to 350,000 rad had aerobic-facultative macrocolony counts below 150×10^4 /g at 6 C and $< 100 \times 10^2$ /g at 0 C after about 1 week in storage. At doses of 50,000 to 150,000 rad, the counts rose into the hundreds of thousands per g with samples stored at 6 C, and in the tens of thousands per g at 0 C during the first week of storage.

Counts of over 1 million microorganisms per g were obtained from clam meats, treated with doses of 200,000 to 350,000 rad of ionizing radiation, after the samples had been held at 6 C for some 18 days. Comparably high levels of microbial populations were found in clams irradiated at 50,000 to 150,000 rad by the 9th to 12th day of storage at 6 C. At 0 C, counts in the millions per g were not observed until after some 3 weeks in storage for samples irradiated at 150,000 rad of gamma rays, whereas clams treated at 150,000 rad exhibited counts of about 1 million per g after some 2 weeks of storage at 0 C. These storage periods were significantly longer than for haddock fillets irradiated and stored under the same conditions.

Data from the high substerilization dose range from 400,000 to 800,000 rad are presented in Tables 9 and 10.

Once again it is to be noted that more than 99% of the original aerobic-facultative microflora of this mollusk was destroyed at these high substerilization doses, as with the haddock fillet microflora in this high radiation dose range. After 1 month in cold storage, the microbial populations in the clam samples treated at doses of from 650,000 to 800,000 rad remained below $50 \times 10^4/g$ at 6 C and $30 \times 10^4/g$ at 0 C.

After 2 months in storage, the counts on these clams increased to about 4.0 to $20.0 \times 10^5/g$ at 6 C, and between 3 and 90 $\times 10^4/g$ at 0 C; after 3 months of storage, the counts went into the millions of microbes per g of product.

Results from storage studies of clams irradiated at doses of 600,000 rad and lower indicated that concentrations of some hundreds of thousands of microorganisms per g were present after no longer than 1 month in storage at both temperatures. Counts of over 1 million per g were obtained from samples kept in storage at 6 C for some 3 to 6 weeks, and at 0 C for from 1 to 2 months.

The foregoing data suggest that shucked soft-shelled clams may be effectively maintained in refrigerated storage for periods of up to 3 months when treated with from 650,000 to 800,000 rad of Co^{60} gamma rays. In the dose range of 500,000 to 600,000 rad, the storage life of this product may be extended for some 2 months, and with treatments of 400,000 to 450,000 rad the shucked clams might be held under refrigeration for 1 month to 6 weeks.

The high degree of inactivation of the aerobic-facultative microbial populations achieved with these high radiation doses was likewise reflected in the highly significant reduction brought about in the anaerobic (clostridia) microflora of this shellfish. At the moderate radia-

^b Counts at 250,000 rad were taken after 5 days of storage.

<sup>Counts at 200,000 rad were taken after 20 days of storage.
Counts at 300,000 rad were taken after 22 days of storage.</sup>

227

tion doses of 50,000 to 350,000 rad, more of these sporeforming, radiation-resistant microorganisms survived the irradiation treatments.

During storage, the samples irradiated with from 400,000 to 800,000 rad of Co⁶⁰ gamma rays showed variably slight increases in number of clostridia present, and remained

below 100 organisms per g at both 0 and 6 C, even after 3 months in storage. Shucked clams treated with doses of 50,000 to 350,000 rad exhibited a similar fluctuation in clostridia levels during storage, but stayed below 200 per g at 0 or 6 C, or both, up through 24 days in storage. These results are most encouraging from a public health stand-

ABLE 9. Aerobic-facultative macrocolony counts obtained from shucked soft-shelled clams before and after treatment with Co⁶⁰ gamma rays and storage at 6 or 0 C for up to 3 months^a

T	Stor-	Medi-														At	osor	bed	dose	e (rad	I)													
Ireatment	age temp	ium ^b		400,0	00	4	50,00	0		500	,000)		550	0,00	0		600	0,00	0		650	,000			700	,000		7:	50,00	00	8	00,0	00
	C												-																					
Before irra-		DW	49	.7 >	< 10 ^a	112	×	103	15	. 1	×	10	311	.9	×	10 ³	31	.2	×	10 ³	43	.7	×	103	37	.2	×	10 ³	11.	9 X	< 10	¹ 20.5	; >	< 10 ⁴
diation		SSW	20	.9 >	< 10 ³	106	X	103	6	. 93	×	10	³ 10	.7	×	10 ³	20	.6	×	10 ³	23	.4	×	10 ³	26	.6	Х	10 ³	65 .	3 X	< 10	³ 12.3	; >	< 10 ⁴
Immediately		DW	22	.9 >	(10 ¹	24.7	X	10 ¹	5	. 80	×	10	۱ <u>3</u>	.93	×	10 ¹	9	.80	×	10 ¹	4	. 47	\times	10 ¹	3	.87	х	10 ¹	22.	2 ×	< 10	4 6.7	3>	< 10 ¹
after irrad- diation		SSW	41	.9 >	(10 ¹	17.5	×	10 ¹	3	.00	×	10 ¹	2	. 87	×	10 ¹	5	. 33	×	10 ¹	1	. 87	×	10 ¹	2	.35	×	10 ¹	12.	7 ×	< 10	6.5	3>	< 10 ¹
After 1	6	DW	29	.1 >	< 10 ⁴	° 78.1	X	1030	11	.6	×	10	4 65	.1	×	104	15	2	×	104	38	.5	×	10 ²	36	.3	х	10 ³	42.	7 >	< 10	³ 35.7	` >	< 10 ⁴
month of	ĺ	SSW	26	.5 >	(104	76.2	X	103	11	.2	×	10	⁴ 60	.1	×	104	14	1	×	104	30	.6	×	10 ²	28	0	×	10 ³	36.	9 X	< 10	³ 31.2	: >	< 10 ⁴
refrigerated	0	DW	21	.7 >	(10 ³	61.7	X	10 ²	33	.6	×	10	³ 50	.8	×	10 ³	86	.3	×	104	25	.5	×	10²	38	7	х	10²	32.	0 X	< 10	322.3	; >	< 10 ⁴
storage		SSW	19	.8 >	(10 ³	57.7	X	10 ²	33	.7	Х	10	3 47	.8	×	10 ³	81	.3	×	104	22	.6	×	10°	32	.7	х	10 ²	21.	1 >	< 10	³ 21.7	∕≻	< 10 ⁴
After 2	6	DW	16	.6 ×	(106	$^{d}23.5$	X	1040	46	.9	×	10	5 11	.1	×	10 ^{5e}	67	.7	×	1050	39	.6	×	103	30	3	х	104	30 .	1 >	< 10	21.5	i >	< 10 ⁵
months of		SSW	12	.9 🗙	(106	17.7	X	104	43	.1	Х	10	5 10	.5	×	105	61	.1	×	10^5	35	.8	×	10 ³	23	7	Х	104	21.	7 ×	< 10	17.2	: >	< 10 ⁵
refrigerated	0	DW	18	.0 >	(105	9.3	$3 \times$	103	24	.9	×	10	28	.5	×	104	24	.2	×	105	28	.0	×	10 ³	80	.7	×	10 ³	9 9.	8 ×	< 10 ⁻	³ 91.8	\$ >	< 106
storage		SSW	14	.3 >	(105	6.5	$3 \times$	103	21	.4	Х	10	⁵ 24	.4	×	10^{4}	21	.1	×	105	26	.9	×	10 ³	69	.2	Х	10 ³	7 9.	9 X	< 10 ⁻	378.5	5 >	< 10 ⁶
After 3	6	$\mathbf{D}\mathbf{W}$			-	3.4	X	1060		-			1	.9	×	106/	1	.1	×	10%	41	.8	×	105	16	.6	х	105	15.	5 X	< 10	⁵ 64.6	; >	< 106
months of		SSW			-	4.1	X	106		-			0	.5	×	106	1	.5	×	106	41	.8	×	105	13	.1	Х	105	14.	0 X	< 10	^{\$} 65.8	\$ >	< 1 ⁴⁶
refrigerated	0	DW				6.9	X	105		-			1	.3	×	105	4	.6	×	105	30	. 1	×	105	97	.5	Х	104	66.	0 X	< 10	⁴ 11.7	' >	< 10 ⁴
storage		SSW				6.5	×	105		-			2	.1	×	105	1	.4	Х	105	25	.1	×	105	90	.8	Х	104	61.	6 ×	< 10	10.4	>	< 100

"Average (\bar{X}) of five replicate samples plated in triplicate $(X \cdot = 15)$ and incubated for 5 days at 20 C. Results expressed as count per g of medium.

^b DW, distilled water culture medium; SSW, synthetic sea water culture medium.

0

^c Count taken after 2 weeks of storage.

^d Count taken after 3 weeks of storage.

^e Count taken after 6 weeks of storage.

^f Count taken after 8 weeks of storage.

^o Count taken after 10 weeks of storage.

rays and storage at 6 or 0 C for up to 5 months										
Treatment	Storage	Absorbed dose (rad)								
	temp	400,000	450,000	500,000	550,000	600,000	650,000	700,000	750,000	800,000
	С									
Before irradiation		42.1	10.9	15.5	29.1	42.4	11.9	9.07	14.3	34.2
Immediately after irradiation		17.7	4.80	4.80	8.20	15.3	2.00	1.47	2.13	2.53
After 1 month of refrigerated stor-	6	11.1^{b}	8.875	4.07	14.7	13.1	3.07	3.80	10.7	4.60
age	0	9.53	5.87	5.20	9.87	8.07	2.33	3.27	5.33	4.10
After 2 months of refrigerated stor-	6	16.5°	12.3°	7.93	12.2^{d}	10.5^{4}	4.58	4.33	8.00	5.00
age	0	15.3	9.87	7.13	8.27	7.73	3.00	1.73	2.07	1.20
After 3 months of refrigerated stor-	6		11.07 ^d		10.5	10.47	2.33	1.47	3.47	6.73

TABLE 10. Anaerobic (clostridia) macrocolony counts obtained from shucked soft-shelled clams before and after treatment with Co⁶⁰ gamma rays and storage at 6 or 0 C for up to 3 months^a

^a Average (\bar{X}) of five replicate samples tubed in triplicate (X · = 15) and incubated at 30 and 37 C for 40 hr. Results expressed as count \times 10 per g of medium.

10.00

8.43

1.67

1.00

1.80

2.46

7.07

^b Count taken after 2 weeks of storage.

age

^c Count taken after 3 weeks of storage.

^d Count taken after 6 weeks of storage.

• Count taken after 8 weeks of storage.

¹ Count taken after 10 weeks of storage.

ŵ.

point, for they indicate that those clostridia remaining after irradiation can be kept at safe, low levels during refrigerated storage.

Acknowledgments

This investigation was conducted under contract nos. AT(30-1)-2329 and AT(30-1)-3006, Task XII, of the U.S. Atomic Energy Commission.

The authors gratefully acknowledge the technical assistance of Judith S. Voss and Jane M. Sprogis in the preparation and microbiological assay of the sea food samples, and Richard F. Zimpel and René Lagasse in the operation of the Mark I Co⁶⁰ Food Irradiator. They also express their thanks to J. J. Licciardello, who supervised the calibration of source dose levels, and to Gerald Silverman and Norman S. Davis for their helpful suggestions in some phases of the studies.

LITERATURE CITED

- ANGELOTTI, R., AND H. E. HALL. 1961. Rapid procedure for the detection and quantitation of *Clostridium perfringens* in foods. Bacteriol. Proc., p. 68.
- BUCK, J. D., AND R. C. CLEVERDON. 1960. The spread plate as a method for the enumeration of marine bacteria. Limnol. Oceanog. 5:78.
- DEPARTMENT OF NUTRITION, FOOD SCIENCE AND TECHNOLOGY, MASSACHUSETTS INSTITUTE OF TECHNOLOGY. 1961. A study of the effects of sub-sterilization doses of radiation on the storage

life extension of soft-shelled clams and haddock fillets. U.S. At. Energy Comm. Quarterly Progress Rept. May to July NYO-9572.

- DOLMAN, C. E. 1957. Type E (fish-borne) botulism; a review. Japan. J. Med. Sci. Biol. 10:383.
- MACLEOD, R. A., AND E. ONOFREY. 1956. Nutrition and metabolism of marine bacteria. II. Observations on the relation of sea water to the growth of marine bacteria. J. Bacteriol. 71:661-667.
- MASUROVSKY, E. B., J. S. VOSS, AND S. A. GOLDBLITH. 1963. Changes in the microflora of haddock fillets and shucked soft-shelled clams after irradiation with Co⁶⁰ gamma rays and storage at 0 C and 6 C. Appl. Microbiol. **11**:229-234.
- MOSSEL, D. A. A., A. S. DEBRUIN, H. M. J. VANDIEPEN, C. M. A. VENDRIG, AND G. ZONTELWELLE. 1955. The enumeration of anaerobic bacteria, and Clostridium species in particular, in foods. J. Appl. Bacteriol. 19:142–154.
- NICKERSON, J. T. R., AND S. A. GOLDBLITH. 1950. A comparison of chemical changes in mackerel tissues treated by ionizing radiation. Food Technol. 4:84-88.
- NICKERSON, J. T. R., E. E. LOCKHART, B. E. PROCTOR, AND J. J. LICCIARDELLO. 1954. Ionizing radiations for the control of fish spoilage. Food Technol. 8:32-34.
- NICKERSON, J. T. R., G. J. SILVERMAN, M. SOLBERG, D. W. DUNCAN, AND M. M. JOSELOW. 1962. Microbial analysis of commercial frozen fish sticks. J. Milk Food Technol. 25:45–47.
- RAJ, H., W. J. WIEBE, AND J. LISTON. 1961. Detection and enumeration of fecal indicator organisms in frozen sea foods. II. Enterococci. Appl. Microbiol. 9:295-303.
- SHEWAN, J. M., AND J. LISTON. 1958. Experiments on the irradiation of fish with 4 mev cathode rays and Co⁶⁰ gamma rays. Proc. Intern. Conf. Peaceful Uses At. Energy Geneva, vol. 27.