Carbon Dioxide Effects on Glucose Catabolism by Mixed Microbial Cultures¹

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ABSTRACT

GAFFNEY, PETER E. (Georgia Institute of Technology, Atlanta). Carbon dioxide effects on glucose catabolism by mixed microbial cultures. Appl. Microbiol. 13:507-510. 1965.—Results have shown that, with mixed culture (sewage) inocula, the lag period in aerobic catabolism of glucose can be reduced by increased CO_2 tension. Conversely, removal of CO_2 from the air supply to the growth flasks and Warburg vessels may increase the lag period.

It has long been known that CO_2 is necessary for the nutrition of autotrophic bacteria (Bonazzi, 1921; Gowda, 1924). That many heterotrophic organisms are capable of fixing CO_2 is generally known, but possible benefit of fixation is little understood.

Valley and Rettger (1926) presented a list of 82 organisms for which CO_2 appeared to be necessary for development. In many instances, they found that growth could be prevented by removving the CO_2 from the atmosphere with which the organism was supplied. After studying more than 100 different organisms representing various families and genera, they concluded that CO_2 is necessary for the growth and development of the bacterial cell (Valley and Rettger, 1927).

Walker (1932) confirmed the work of Valley and Rettger and demonstrated that CO_2 had its greatest effect during the lag phase. He suggested that the phenomenon of lag was due to the time needed for the culture to build up necessary CO_2 content. This focused attention on the possibility of a definite CO_2 requirement for heterotrophs. In an attempt to explain this requirement, Werkman and Wood (1942) postulated a condensation of pyruvate and CO_2 to form oxaloacetic acid, indicating its critical importance with respect to the citric acid cycle. Krampitz and Werkman (1941) showed some support for this, and more definite evidence on an enzymatic basis was later shown by Kaltenbach and Kalnitsky (1951).

During a previous investigation (Gaffney and Heukelekian, 1961) dealing with comparison of oxidation rates of the lower fatty acids under various conditions in a Warburg respirometer,

¹ A portion of this paper was presented to the Southeastern Branch of the American Society for Microbiology, Jacksonville, Fla., 18 October 1963. long lag periods were encountered, and it was shown that these could not be eliminated or significantly reduced by altering the quality ("adapting") or the quantity of inocula.

It was suggested (Pardee, 1949; Krebs, 1951) that the results obtained with the Warburg technique are often in error due to the elimination of CO_2 with the direct method.

The objective of this investigation was to determine the effect of the presence or absence of CO_2 on the initiation and rate of catabolism in a dispersed aeration system, and also the effect on oxygen-uptake rates, as measured by use of the Warburg respirometer.

MATERIALS AND METHODS

The medium consisted of basal salts, phosphate buffer (pH 7.0), and 1,000 mg per liter of glucose, and was inoculated (1.0% by volume) with various settled domestic sewage samples. The mixture was divided into three equal samples, each receiving different air supplies. Fritted-glass bubblers were inserted into the system between the main air supply and the growth flasks (Fig. 1). The bubblers contained water, a supersaturated solution of sodium bicarbonate, or 20% potassium hydroxide, so that the air delivered to the growth flasks was normal, CO₂-supplemented, or CO₂-free, respectively. CO₂-free air was demonstrated by precipitation tests with barium hydroxide. In three of the tests, a CO₂ gas tank instead of the bicarbonate solution was used for supplementation.

Glucose catabolism was measured daily by analysis of chemical oxygen demand (COD) according to the 11th edition of *Standard Methods for Analysis of Water and Wastewater* (American Public Health Association, 1960). The COD analysis of a substrate consists of complete chemical combustion of organic substances by refluxing in acid dichromate, and the value is expressed as the maximal amount of oxygen consumed in destruction of the organic material.

In the respirometric experiments, Dickens-Simer flasks were used with the routine potassium hydroxide CO_2 trap and compared with similar systems containing diethanolamine as a CO_2 buffer, as suggested by Krebs (1951). Use of the diethanolamine under these conditions results in constant maintenance of a 1 to 2% CO_2 atmosphere in the Warburg vessels.

RESULTS

The results with five different sewage inocula are shown in Table 1. The COD values are averages of triplicate analyses, and the experimental error is $\pm 4.0\%$. During test IV, after aeration for 1 day, total bacterial plate counts (on glucoseagar) were made on the solutions from the three growth flasks. The buffer held the *p*H in each test between 6.9 and 7.2. Only the results after 1 day

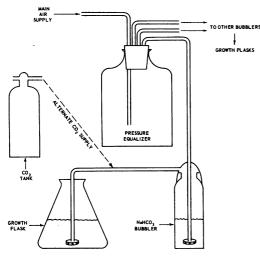


FIG. 1. Gas supply system.

 TABLE 1. Amount of glucose COD dissimilated after

 aeration for 1 day*

Test no.	Water		Bubbler NaHCO3		CO2 gas		КОН	
	Amt†	Per cent	Amt	Per cent	Amt	Per cent	Amt	Per cent
I II III IV V	390 50 40 60 12	34.0 4.4 3.5 5.2 1.0	490 670	43 58	140 380 13	12 33 2	210 0 30 10 25	18 0 2.6 0.9 3.0

* Initial glucose, 1,000 mg per liter; initial COD, 1,150 mg per liter.

† All amounts are expressed as milligrams per liter.

of aeration are given, because the early period is of most interest.

The data from the first four tests consistently show that a greater percentage of glucose COD was dissimilated after 1 day when CO_2 was added or left in the air supply than when CO_2 -free air was supplied. Also, the growth activity (in terms of numbers of organisms) followed the same pattern. The percentage of glucose COD dissimilation for water, CO_2 , and KOH was 5.2, 33, and 0.9%, respectively; the corresponding bacterial counts were 34,000,000, 80,000,000, and 15,000,000 organisms per milliliter. The COD re-

TABLE 2. Increased glucose COD dissimilated with normal and CO₂-enriched air as compared with CO_2 -free air

Test no.	Normal air	Co2-enriched air	
I	+180*	+280	
II	+50	+670	
III	+10	+110	
IV	+50	+370	
V	-13	-12	
Mean increase	+55	+284	

1200 1100 BUBBLERS NoHCO₃ 1000 O WATER ∆ кон 900 800 700 D (mg A) 600 0 U 500 400 300 200 100 0 4 DAYS

FIG. 2. Dissimilation of 1,000 mg per liter of glucose.

* Results expressed as milligrams per liter.

Substrate mixture	Oxygen uptake in 20 hr		
Substrate mixture	No CO2*	With CO ₂ †	
	uliters	µliters	
100% domestic sewage	100	210	
1% sewage + 50 mg per liter of glucose	44	80	
of glucose		100	

* Routine procedure with KOH trap.

† Diethanolamine "buffer" was used (1 to 2% CO₂ atmosphere).

sults of test V do not follow that pattern, and, in this test, CO_2 had no effect on the 1-day values. The data in Table 2 are derived from those in Table 1. Here it is shown that, compared with CO_2 -free air, normal air allowed for an additional 55 mg per liter of glucose to be dissimilated in the first day, and CO_2 -enriched air resulted in an increase of 284 mg per liter on the average.

The complete 7-day curves during test II are presented in Fig. 2. The bulk of catabolic activity occurred during the first day with CO_2 -supplemented air, during the second day with normal air, and not until the third day with CO_2 -free air.

Oxygen uptake was measured in the Warburg flasks for 20 hr with several substrate mixtures; the data in Table 3 are the result of measurement on three replicates of the mixture with and without CO₂. In each case, the oxygen uptake in the first 20 hr is much greater in the presence of CO_2 than in its absence, especially when the ratio of substrate to cells is high.

DISCUSSION

In the past few years, there have been numerous reports of the beneficial nature of CO_2 to the heterotrophic bacteria. Harris (1954) found, by using constant concentrations of CO_2 (0.8%), that the oxygen uptake by dilute suspensions of 38 common bacterial species utilizing glucose as substrate was greater than oxygen uptake in flasks in which the CO_2 had been removed by concentrated alkali. In dilute suspensions, the stimulation seemed to be inversely related to the amount of CO_2 produced by the cells; when many cells were present, this stimulation was not observed.

Farghaly (1950) reported that carbon dioxide was an essential factor for the growth of luminous bacteria. Holm (1954) found that *Actinobacillus actinomycetemcomitans*, which does not normally grow on media under aerobic or anaerobic conditions, would grow on the surface of some solid media under both conditions if a concentration (0.5% or more) of CO₂ was present.

Lui (1954) reported that the growth of hemolytic streptococci can be markedly improved by increased CO₂ concentrations. Maximal growth was achieved by most strains when the concentrations reached 5% and could not be improved by higher concentrations.

Mattman (1954), in comparing the growth of 869 cultures on blood-agar plates, showed that a significant percentage of strains grew only when CO_2 was added. The cultures included especially β -hemolytic streptococci, pneumococci, and micrococci.

Szulmajster (1958) reported that an organism belonging to the genus *Clostridium*, which was isolated from sewage sludge, was able to grow on creatinine (which is found in large percentages in sewage) only in the presence of CO_2 and yeast extract. It was also shown with labeled CO_2 that the CO_2 was incorporated into the cell material.

Field and Lichstein (1958) reported that autoclaved glucose medium produced an unidentified factor which satisfied the CO_2 requirements of propionibacteria for early initiation of growth.

In the early development of the standard-dilution, 5-day biochemical oxygen demand (BOD) test for the strength of sewage, Theriault (1931) reported somewhat higher results when phosphate buffer dilution water was used in place of a suggested bicarbonate water containing 300 mg per liter of sodium bicarbonate. Although this may have been indicative that CO₂ had no particular effect on the lag period, it is also likely that the sewage samples contained enough CO₂ to fill the requirement. Furthermore, the differences in Theriault's results could have been due to pHvariations, as he stated that "it does not appear advisable to recommend the use of readily prepared bicarbonate solution for dilution purposes until more data from different laboratories have been accumulated regarding the effect of pH on the rate of deoxygenation in natural waters."

It was shown (Gaffney, *unpublished data*) that, in standard-dilution BOD tests on glucose and sewage, CO_2 supplementation of the phosphate buffer dilution water presently used had no effect (of reducing lag periods) on short-term BOD values (1 to 5 days). It was concluded that, with the relatively low concentrations of organic material used (10 mg per liter), the CO_2 requirement was not critical, since pronounced lag periods were absent. Thus, it is likely that the differences in the lag-reducing effects of CO_2 between dilution BOD tests and the investigations reported herein can be explained by the relative food-toorganisms ratios.

 CO_2 supplementation of phosphate dilution

water in standard-dilution BOD tests may merit some investigation in cases of relatively resistant industrial wastes in which long lag periods may be observed. Also, since the Warburg respirometer is frequently used to obtain BOD values on numerous types of concentrated wastes, and since a low concentration of organisms is used, it would seem that the use of the common KOH absorbent in the reaction vessels would affect the validity of the data, particularly if the results are evaluated in terms of natural conditions in a stream.

The experiments described in this report were performed with dilute suspensions and sewage inocula containing a large variety of microorganisms. The beneficial effect of CO_2 supplementation has been demonstrated on the basis of the ability of mixed cultures to catabolize the soluble organic nutrient, specifically to reduce the COD at a greater initial rate. Corollary data indicated that the early phase of oxygen uptake in the Warburg flasks is similarly enhanced in the presence of increased CO_2 tension. On the basis of this and previous work, it would appear that CO_2 supplementation is important with respect to lag periods in catabolic measurements with heterotrophic organisms.

ACKNOWLEDGMENTS

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LITERATURE CITED

- AMERICAN PUBLIC HEALTH ASSOCIATION. 1960. Standard methods for the examination of water and wastewater, 11th ed. American Public Health Association, Inc., New York.
- BONAZZI, A. 1921. On nitrification. IV. The carbon and nitrogen relations of the nitrite ferment. J. Bacteriol. 6:479-499.
- FARGHALY, A. 1950. Factors influencing the growth and light production of luminous bacteria. J. Cellular Comp. Physiol. 36:165-183.
- FIELD, M. F., AND H. C. LICHSTEIN. 1958. Growth stimulating effect of autoclaved glucose media and its relationship to the CO₂ requirement of propionibacteria. J. Bacteriol. **76**:485–590.

- GAFFNEY, P. E., AND H. HEUKELEKIAN. 1961. Biochemical oxidation of the lower fatty acids. J. Water Pollution Control Federation 33:1169– 1184.
- GOWDA, R. N. 1924. Nitrification and the nitrifying organisms. J. Bacteriol. 9:251-272.
- HARRIS, J. O. 1954. The influence of carbon dioxide on oxygen uptake by "resting cells" of bacteria. J. Bacteriol. 67:476-479.
- HOLM, P. 1954. The influence of carbon dioxide on the growth of *Actinobacillus actinomycetemcomitans*, (Bacterium actinomycetem comitans). Acta Pathol. Microbiol. Scand. **34**:235-248.
- KALTENBACH, J. P., AND G. KALNITSKY. 1951. The enzymatic formation of oxalacetate from pyruvate and carbon dioxide. J. Biol. Chem. 192:629.
- KRAMPITZ, L. O., AND C. H. WERKMAN. 1941. The enzymic decarboxylation of oxalacetate. Biochem. J. 35:595-602.
- KREBS, H. A. 1951. The use of CO_2 -buffers in manometric measurements of cell metabolism. Biochem. J. 48:349-359.
- LUI, P. 1954. Carbon dioxide requirement of group F and minute colony G hemolytic streptococci. J. Bacteriol. 68:282-288.
- MATTMAN, L. H., T. SAYLOR, G. OLIVER, J. FRAL-ING, P. KICE, M. BUCKLEY, and B. TOLONIN. 1954. Carbon dioxide and routine cultures in the diagnostic laboratory. J. Lab. Clin. Med. 42: 485-488.
- PARDEE, A. B. 1949. Measurement of oxygen uptake under controlled pressures of carbon dioxide. J. Biol. Chem. 179:1085-1091.
- SZULMAJSTER, J. 1958. Bacterial fermentation of creatinine. I. Isolation of N-methyl-hydantoin. J. Bacteriol. 75:633-639.
- THERIAULT. E. J. 1931. Detailed instructions for the determination of biochemical oxygen demand by the excess oxygen (dilution) method. Public Health Rept. U.S. Suppl. 90, p. 18-34.
- VALLEY, G., AND L. F. RETTGER. 1926. Carbon dioxide requirements of bacteria. J. Bacteriol. 11:78-79.
- VALLEY, G., AND L. F. RETTGER. 1927. The influence of carbon dioxide on bacteria. J. Bacteriol. 14:101-135.
- WALKER, H. H. 1932. Carbon dioxide as a factor affecting lag in bacterial growth. Science 76: 602-604.
- WERKMAN, C. H., AND H. G. WOOD. 1942. Heterotrophic assimilation of carbon dioxide. Advan. Enzymol. 2:135.