

Gut and flesh microbiome sequencing of the Bangladesh national fish hilsa (*Tenualosa ilisha*)

Tofazzal Islam,¹ M. Nazmul Hoque²

AUTHOR AFFILIATIONS See affiliation list on p. 3.

ABSTRACT The gut and flesh microbiome of the national fish of Bangladesh, *Tenualosa ilisha*, were analyzed using 16S rRNA gene sequencing. Our findings revealed a significant microbial disparity between sample categories and the habitat of hilsa fish, which will serve as a valuable foundation for further comprehensive studies on the hilsa microbiome.

KEYWORDS *Tenualosa ilisha*, Hilsa flesh, gut microbiome, metagenomics

Hilsa (*Tenualosa ilisha*) is the national fish of Bangladesh. It contributes ~12% of the total fish production and 1.15% GDP of the country (1). It is widely distributed in Southeast and South Asia (2, 3) and found in coastal areas, estuaries, brackish, and freshwater rivers (3, 4). Hilsa is an anadromous marine fish that comes to freshwater in rivers during spawning. Gut microbiota, which modulate host homeostasis, immunity, and fitness (5, 6), have been analyzed in many fish species (7, 8) but rarely in the hilsa fish (1). Nothing is known about the gut and flesh microbiome of hilsa, which is known to be free from diseases. This study aimed to analyze the gut and flesh microbiome of hilsa fish using 16S rRNA gene amplicon sequencing.

Eighteen adult hilsa fishes were collected and frozen from September to October 2020 from five different sites in Bangladesh (Table 1). Sampling of their intestinal contents ($n = 15$) and flesh ($n = 3$) was performed within 48 h after the collection. Fish scales from the body of all individual fishes were removed by sterile forceps under aseptic conditions, and flesh (100 mg) and lower stomach contents (100 mg) from each fish were collected. Genomic DNA was extracted from each specimen after grinding (flesh) and homogenization (gut content) using DNeasy Blood and Tissue Kit (Qiagen, UK) following the manufacturer's instructions. A NanoDrop 2000c spectrophotometer was used to check the purity and concentration of DNA. The amplicon libraries were prepared using Nextera XT Index Kit (Illumina Inc.) as per the 16S amplicon sequencing library preparation protocol, and V3–V4 regions were amplified through a 40-cycle PCR using 341F and 806R primers (9). Paired-end sequencing (2×150 bp, 300 cycles) was performed on the Illumina MiniSeq sequencer. FastQC v0.11.9 (10) was used to check the quality of the reads, while Trimmomatic v0.39 (11) removed Illumina adapters, known Illumina artifacts, and phiX reads. The demultiplexed sequencing data were processed using QIIME 2 (2023.2.0) and associated plugins (12). Sequences showing $\geq 98\%$ similarity were grouped into operational taxonomic units (OTUs) using the SILVA database v.138 (13). Default parameters were used for bioinformatic analyses, except where otherwise stated.

The 16S rRNA amplicon sequencing of 18 samples generated 3,696,608 raw reads (average: 205,367 reads/sample), of which 390,289 quality reads (10.56%) mapped to 325 OTUs of bacteria. The rest of the reads (90%) were unassigned to any OTUs and might come from host DNA. Among these, 67 and 258 OTUs were identified in flesh and gut samples, respectively, through the SILVA database v.138 (Table 1). These OTUs

Editor Frank J. Stewart, Montana State University, Bozeman, Montana, USA

Address correspondence to Tofazzal Islam, tofazzalislam@bsmrau.edu.bd.

The authors declare no conflict of interest.

Received 25 May 2023

Accepted 15 August 2023

Published 25 September 2023

Copyright © 2023 Islam and Hoque. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

TABLE 1 Summary of metadata and SRA accession numbers of the 16S rRNA amplicon sequences of hilsa fish gut and flesh samples, along with OTUs mapped against bacterial taxa

Sample ID	Collection site	Coordinate	Habitat	Source	No. of raw reads	No. of mapped reads	No. of observed OTUs	SRA accessions
CG1	Confluence of Meghna and Padma River, Chandpur	23.2321° N, 90.6631° E	Freshwater	Gut	325,812	33,295	22	SRR24402593
CG2	Confluence of Meghna and Padma River, Chandpur	23.2321° N, 90.6631° E	Freshwater	Gut	140,124	12,418	22	SRR24402592
CG3	Confluence of Meghna and Padma River, Chandpur	23.2321° N, 90.6631° E	Freshwater	Gut	119,676	4,738	10	SRR24402608
CG4	Confluence of Meghna and Padma River, Chandpur	23.2321° N, 90.6631° E	Freshwater	Gut	697,544	291,731	23	SRR24402607
CG5	Confluence of Meghna and Padma River, Chandpur	23.2321° N, 90.6631° E	Freshwater	Gut	131,276	9,514	20	SRR24402606
RG3	Padma River, Rajshahi	24.3745° N, 88.6042° E	Freshwater	Gut	194,464	2,256	18	SRR24402605
MG1	Meghna River, Munshiganj	23.5422° N, 90.5305° E	Freshwater	Gut	165,096	1,333	15	SRR24402604
MG2	Meghna River, Munshiganj	23.5422° N, 90.5305° E	Freshwater	Gut	126,908	7,353	15	SRR24402602
MG4	Meghna River, Munshiganj	23.5422° N, 90.5305° E	Freshwater	Gut	260,420	3,077	11	SRR24402601
PG1	Payra River, Patuakhali	22.3586° N, 90.3317° E	Brackish water	Gut	149,988	611	23	SRR24402600
PG2	Payra River, Patuakhali	22.3586° N, 90.3317° E	Brackish water	Gut	119,052	1,142	11	SRR24402598
PG3	Payra River, Patuakhali	22.3586° N, 90.3317° E	Brackish water	Gut	361,312	8,111	20	SRR24402599
PG5	Payra River, Patuakhali	22.3586° N, 90.3317° E	Brackish water	Gut	178,372	3,982	18	SRR24402597
XG1	Bay of Bengal, Cox's Bazar	21.4272° N, 92.0058° E	Marine water	Gut	112,684	509	16	SRR24402596
XG3	Bay of Bengal, Cox's Bazar	21.4272° N, 92.0058° E	Marine water	Gut	151,480	260	14	SRR24402595
CF4	Confluence of Meghna and Padma River, Chandpur	23.2321° N, 90.6631° E	Freshwater	Flesh	180,944	4,994	16	SRR24402609
CF5	Confluence of Meghna and Padma River, Chandpur	23.2321° N, 90.6631° E	Freshwater	Flesh	106,604	2,226	14	SRR24402603
PF4	Payra River, Patuakhali	22.3586° N, 90.3317° E	Brackish water	Flesh	174,852	2,739	17	SRR24402594

were represented by six phyla, nine classes, 19 orders, 26 families, and 40 genera of bacteria. Firmicutes constituted >75.0% of the hilsa bacteriome, with 78.17% and 2.86% relative abundances in the gut and flesh, respectively. *Vagococcus* (67.02%), *Morganella* (13.25%), *Enterobacter* (5.74%), *Plesiomonas* (3.51%), *Shigella* (1.75%), *Clostridium* (1.60%), *Klebsiella* (1.02%), and *Serratia* (1.0%) were the top abundant bacterial genera detected in hilsa fish, with distinct variations in their relative abundances according to the sample categories (gut and flesh; $P = 0.0127$; Kruskal Wallis test). The genomic data of the

flesh and gut bacteriome of hilsa fish revealed in this study has laid the foundation for shedding light on the microbiome of this economically important trans-boundary fish.

ACKNOWLEDGMENTS

We are thankful to the "BSMRAU Physical Facility and Research Capacity Strengthening Project" under the Ministry of Education, People's Republic of Bangladesh, for funding this research. The authors sincerely thank A.Q.M. Robiul Kawser, Assistant Professor, Department of Aquaculture, Bangabandhu Sheikh Mujibur Rahman Agricultural University (BSMRAU), Gazipur 1706, Bangladesh, and Nur Uddin Mahmud of the IBGE, BSMRAU, for their support in sampling, technical assistance in genomic DNA extraction, and PCR.

The authors declare no conflicts of interest regarding this paper.

AUTHOR AFFILIATIONS

¹Institute of Biotechnology and Genetic Engineering (IBGE), Bangabandhu Sheikh Mujibur Rahman Agricultural University (BSMRAU), Gazipur, Bangladesh

²Department of Gynecology, Obstetrics and Reproductive Health, BSMRAU, Gazipur, Bangladesh

AUTHOR ORCID*s*

Tofazzal Islam  <http://orcid.org/0000-0002-7613-0261>

M. Nazmul Hoque  <http://orcid.org/0000-0002-4861-0030>

AUTHOR CONTRIBUTIONS

Tofazzal Islam, Conceptualization, Funding acquisition, Project administration, Resources, Supervision, Writing – review and editing, Writing – original draft | M. Nazmul Hoque, Data curation, Formal analysis, Investigation, Methodology, Resources, Software, Validation, Writing – original draft

DATA AVAILABILITY

The 16S rRNA gene amplicon sequencing data are available at the NCBI Sequence Read Archive (SRA) under BioProject accession number [PRJNA964437](https://www.ncbi.nlm.nih.gov/bioproject/PRJNA964437). The accession numbers for all SRA experiments are listed in Table 1.

REFERENCES

- Foysal MJ, Momtaz F, Robiul Kawser AQM, Chaklader MR, Siddik MAB, Lamichhane B, Tay ACY, Rahman MM, Fotedar R. 2019. Microbiome patterns reveal the transmission of pathogenic bacteria in hilsa fish (*Tenualosa ilisha*) marketed for human consumption in Bangladesh. *J Appl Microbiol* 126:1879–1890. <https://doi.org/10.1111/jam.14257>
- Sahoo AK, Wahab M, Phillips M, Rahman A, Padiyar A, Puvanendran V, Bangera R, Belton B, De DK, Meena DK, Behera BK, Sharma AP, Bhaumik U, Mohanty BP, Choudhury SR, Mohan CV. 2018. Breeding and culture status of hilsa (*Tenualosa ilisha*, ham. 1822) in South Asia: a review. *Rev Aquacult* 10:96–110. <https://doi.org/10.1111/raq.12149>
- Mandal S, Lal KK, Singh RK, Sah RS, Jena JK, Singh A, Mohindra V. 2018. Comparative length-weight relationship and condition factor of hilsa shad, *Tenualosa ilisha* (Hamilton, 1822) from freshwater, estuarine and marine environments in India. *Indian J. Fish* 65:33–41. <https://doi.org/10.21077/ijf.2018.65.2.73732-04>
- Nima A, Hossain M, Rahman M, Mawa Z, Hasan M, Islam M, Rahman M, Tanjin S, Sabbir W, Bashar M, Mahmud Y. 2020. Temporal variations of length, weight, and condition of hilsa shad, *Tenualosa ilisha* (Hamilton, 1822) in the meghna river, southeastern Bangladesh. *Egypt J of Aquatic Biolo and Fish* 24:481–494. <https://doi.org/10.21608/ejabf.2020.88776>
- Wu H-J, Wu E. 2012. The role of gut microbiota in immune homeostasis and autoimmunity. *Gut Microbes* 3:4–14. <https://doi.org/10.4161/gmic.19320>
- Hoque MN, Rahman MS, Islam T, Sultana M, Crandall KA, Hossain MA. 2022. Induction of mastitis by cow-to-mouse fecal and milk microbiota transplantation causes microbiome dysbiosis and genomic functional perturbation in mice. *Anim Microbiome* 4:43. <https://doi.org/10.1186/s42523-022-00193-w>
- Ogita T. 2023. 16S rRNA gene amplicon sequencing of the gut microbiota of chimaera phantasma (silver chimaera) captured off Koshimoda in Suruga Bay, Japan. *Microbiol Resour Announc* 12:e0114922. <https://doi.org/10.1128/mra.01149-22>
- Egerton S, Culloty S, Whooley J, Stanton C, Ross RP. 2018. The gut microbiota of marine fish. *Front Microbiol* 9:873. <https://doi.org/10.3389/fmicb.2018.00873>
- Islam T, Fatema, Hoque MN, Gupta DR, Mahmud NU, Sakif TI, Sharpe AG. 2023. Improvement of growth, yield and associated Bacteriome of rice by the application of Probiotic Paraburkholderia and Delftia. *Front Microbiol* 14:1212505. <https://doi.org/10.3389/fmicb.2023.1212505>
- Andrews S. 2017. Fastqc: A quality control tool for high throughput sequence data. <https://github.com/s-andrews/FastQC>.

11. Bolger AM, Lohse M, Usadel B. 2014. Trimmomatic: a flexible trimmer for illumina sequence data. *Bioinformatics* 30:2114–2120. <https://doi.org/10.1093/bioinformatics/btu170>
12. Bolyen E, Rideout JR, Dillon MR, Bokulich NA, Abnet CC, Al-Ghalith GA, Alexander H, Alm EJ, Arumugam M, Asnicar F, Bai Y, Bisanz JE, Bittinger K, Brejnrod A, Brislawn CJ, Brown CT, Callahan BJ, Caraballo-Rodríguez AM, Chase J, Cope EK, Da Silva R, Diener C, Dorrestein PC, Douglas GM, Durall DM, Duvallet C, Edwardson CF, Ernst M, Estaki M, Fouquier J, Gauglitz JM, Gibbons SM, Gibson DL, Gonzalez A, Gorlick K, Guo J, Hillmann B, Holmes S, Holste H, Huttenhower C, Huttley GA, Janssen S, Jarmusch AK, Jiang L, Kaehler BD, Kang KB, Keefe CR, Keim P, Kelley ST, Knights D, Koester I, Kosciulek T, Kreps J, Langille MGI, Lee J, Ley R, Liu Y-X, Loftfield E, Lozupone C, Maher M, Marotz C, Martin BD, McDonald D, McIver LJ, Melnik AV, Metcalf JL, Morgan SC, Morton JT, Naimey AT, Navas-Molina JA, Nothias LF, Orchanian SB, Pearson T, Peoples SL, Petras D, Preuss ML, Pruesse E, Rasmussen LB, Rivers A, Robeson MS, Rosenthal P, Segata N, Shaffer M, Shiffer A, Sinha R, Song SJ, Spear JR, Swafford AD, Thompson LR, Torres PJ, Trinh P, Tripathi A, Turnbaugh PJ, Ul-Hasan S, van der Hoof JJJ, Vargas F, Vázquez-Baeza Y, Vogtmann E, von Hippel M, Walters W, Wan Y, Wang M, Warren J, Weber KC, Williamson CHD, Willis AD, Xu ZZ, Zaneveld JR, Zhang Y, Zhu Q, Knight R, Caporaso JG. 2019. Reproducible, interactive, scalable and extensible microbiome data science using QIIME 2. *Nat Biotechnol* 37:852–857. <https://doi.org/10.1038/s41587-019-0252-6>
13. Quast C, Pruesse E, Yilmaz P, Gerken J, Schweer T, Yarza P, Peplies J, Glöckner FO. 2013. The SILVA ribosomal RNA gene database project: improved data processing and web-based tools. *Nucleic Acids Res* 41:D590–D596. <https://doi.org/10.1093/nar/gks1219>