The intricate role of CCL5/CCR5 axis in Alzheimer disease

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ABSTRACT

The morbidity and mortality associated with Alzheimer disease (AD), one of the most common neurodegenerative diseases, are increasing each year. Although both amyloid β and tau proteins are known to be involved in AD pathology, their detailed functions in the pathogenesis of the disease are not fully understood. There is increasing evidence that neuroinflammation contributes to the development and progression of AD, with astrocytes, microglia, and the cytokines and chemokines they secrete acting coordinately in these processes. Signaling involving chemokine (C-C motif) ligand 5 (CCLS) and its main receptor C-C chemokine receptor 5 (CCRS) plays an important role in normal physiologic processes as well as pathologic conditions such as neurodegeneration. In recent years, many studies have shown that the CCL5/CCR5 axis plays a major effect in the pathogenesis of AD, but there are also a few studies that contradict this. In short, the role of CCL5/CCR5 axis in the pathogenesis of AD is still intricate. This review summarizes the structure, distribution, physiologic functions of the CCL5/CCR5 axis, and the progress in understanding its involvement in the pathogenesis of AD.

KEYWORDS: Alzheimer disease, Axis, CCL5, CCR5, Neuroinflammation

INTRODUCTION

Alzheimer disease (AD) is a chronic neurodegenerative disease that is the most common cause of dementia. It is characterized by progressive cognitive dysfunction and behavioral disorder; early symptoms such as loss of recent memories and mild cognitive decline are relatively insidious. Agnosia, aphasia, apraxia, memory impairment, and personality and behavioral changes can emerge with disease progression, which may eventually result in death. A cross-sectional survey conducted in China in 2020 reported 15.07 million cases of dementia in the population aged over 60 years, with about two-thirds (9.83 million) being AD (1). It is estimated that the number of cases of dementia will reach 130 million by 2050. Given that it accounts for over 50% of all cases of dementia (2, 3), AD will constitute a substantial healthcare burden in the future.

The pathogenesis of AD is not fully understood, and there are no specific curative treatments; existing drugs only alleviate symptoms and delay disease progression. Clarifying the pathogenesis of AD is important for the identification of novel therapeutic targets. The major histopathologic hallmarks of AD are extracellular senile plaques formed by the deposition of amyloid β (A β) protein and intracellular neurofibrillary tangles (NFTs) composed of tau proteins misfolded as a result of

hyperphosphorylation in neurons (4). The A β cascade and tau protein hyperphosphorylation hypotheses have been proposed to explain AD pathogenesis. A β is a 38–43 amino acid polypeptide that is mainly hydrolyzed from amyloid precursor protein (APP). Under physiologic conditions, A β production and clearance are in dynamic equilibrium; however, under pathologic conditions, there is increased production or reduced clearance of A β , leading to its accumulation. A β monomers assemble into oligomers and polymers, mainly $A\beta_{40}$ and $A\beta_{42}$, and ultimately form insoluble neurotoxic amyloid plaques (5, 6). Tau is important for the assembly of the microtubule network, stabilization of microtubule structure, and regulation of axon transport in neurons. Tau exists in phosphorylated and dephosphorylated states that are normally in dynamic equilibrium. Under pathologic conditions, hyperphosphorylated tau does not bind but instead dissociates from microtubules and forms NFTs (7). However, it is unclear how abnormalities in $A\beta$ and tau contribute to the pathogenesis of AD. For example, although drugs that clear A β reduce A β load in the brain of patients with AD, this does not translate into improvements in cognitive function (8). Various drugs have been developed for AD that prevent tau protein aggregation and stabilize microtubule structure, but clinical trials were terminated due

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to insufficient curative effect or toxicity. Although tau protein immunotherapy has shown therapeutic promise, its efficacy has yet to be demonstrated in clinical trials (9).

Neuroinflammation refers to an inflammatory response to trauma, infection, ischemia, toxins, etc. mediated by glial cells, vascular endothelial cells, and peripheral immune cells in the central nervous system (CNS). It involves cytokines such as interleukin-1 β (IL-1 β), interleukin-6 (IL-6), and tumor necrosis factor (TNF), chemokines such as C-C motif chemokine ligand 1 and C-C motif chemokine ligand 5 (CCL5), complement factors, nitric oxide (NO), and reactive oxygen species (ROS) (10). Acute neuroinflammation is a protective mechanism that helps to eliminate pathogens, resist adverse stimuli, and promote nerve cell repair. However, excessive and chronic neuroinflammation leads to the continuous release of proinflammatory cytokines and oxidative stress, which can cause tissue damage and adversely affect neuron structure and function. Neuroinflammation plays an important role in AD pathogenesis and progression (11). Chemokines are small proteins (70-90 amino acids) with a molecular weight of 8-12 kDa and varied functions, including chemotaxis of leukocytes to sites of inflammation, anti-infective activity, and regulation of the immune response. According to the number and spacing of N-terminal cysteines, chemokines can be divided into 4 subfamilies, namely, CXC, CC, C, and CX3C (12). The CCL5/C-C chemokine receptor 5 (CCR5) axis comprising CCL5 and its main receptor CCR5 is involved in multiple pathologic states such as tumors, infectious diseases, and nervous system diseases. Since CCR5 was found to be a coreceptor of HIV-infected target cells, extensive research has been carried out in HIV infection, and several CCR5 antagonists have been designed to block HIV from entering host cells (13). Although accumulating evidence indicates that CCL5/ CCR5 axis can facilitate the development of AD, some studies have come to the opposite conclusion. At this time, the relationships between this pathway and AD are still unclear. This article analyzes associations between the CCL5/CCR5 axis, neuroinflammation, and AD.

OVERVIEW OF CCL5/CCR5 AXIS

CCL5 and CCR5 structure, distribution, and signal pathway CCL5 (also known as regulated on activation, normal T-cell expressed, and secreted [RANTES]) is a member of the CC chemokine subfamily. The gene encoding CCL5 is located on human chromosome 17q11.2-q12. CCL5 is mainly secreted by T cells and is also expressed in platelets, macrophages, synovial fibroblasts, and hepatocytes. Other CCL5 family members include CCL3 (also known as macrophage inflammatory protein $1-\alpha$ [MIP- 1α]) and CCL4 (MIP- 1β). CCL5 can bind to CCR1, CCR3, and CCR4 but the main receptor is CCR5 (14), a 7-transmembrane G protein-coupled receptor composed of 352 amino acids, with 3 extracellular loops and 3 intracellular loops and a molecular weight of 40.6 kDa (15). There are 4 cysteine residues in the extracellular domain that form disulfide bonds and stabilize the extracellular structure; the second intracellular loop contains a conserved sequence (DRYLAVHA) that interacts with G protein (16). CCR5 is

expressed by a variety of immune cells including T cells, macrophages, and dendritic cells; in the CNS, it is expressed at a high level in microglia but at a relatively low level in neurons and astrocytes (17). Since its identification as a co-receptor that facilitates HIV entry into target cells (18, 19), CCR5 has been widely studied in the context of inflammation, tumors, infection, and diseases of the CNS and other systems.

Following CCL5 binding to CCR5, G protein dissociates into the α_i and $\beta\gamma$ subunits; the former inhibits adenylate cyclase, whereas the latter activates phospholipase $C\beta$ (PLC β) and phosphatidylinositol 3 kinase (20). Activated PLC β mediates the production of diacylglycerol (DAG) and inositol-1,4,5-triphosphate (IP3); IP3 then induces the release of intracellular Ca²⁺, leading to activation of calcineurin, dephosphorylation of nuclear factor of activated T cells, and transcription and activation of mitogen-activated protein kinases (MAPKs) such as extracellular signal regulated kinase 1 and 2, p38, and c-Jun N-terminal kinase (JNK) to regulate cell migration and T-cell proliferation. Additionally, Ca²⁺ and DAG activate protein kinase C, which in turn promotes receptor phosphorylation (21, 22). Activated PI3K can also induce the activation of protein kinase B (PKB/Akt) and Rho GTPases; PI3K/Akt signaling is related to cell survival, whereas Rho GTPases participate in cytoskeleton rearrangement and regulate cell polarization, adhesion, and movement (16, 23, 24). The CCL5/CCR5 signaling pathway is summarized in Figure 1.

Physiologic functions of the CCL5/CCR5 axis in the brain In addition to the immune response, CCR5 is involved in neurotransmission, brain development, and learning and memory, among other functions (25). CCR5 is expressed throughout the CNS at all stages of development from embryo to adult, but the distribution varies across brain areas, which may be related to its different functions in different parts of the brain (26, 27). CCR5 is expressed in glial cells and neurons in the cerebral cortex, hippocampus, basal nuclei, and thalamus, and the expression level was shown to increase from birth to 9 months of age, and it was also observed in adult neural progenitor cells (28), which indicates that CCR5 is involved in neural development. Additionally, in the aspect of neurotransmission, CCL5 modulates the release of glutamate from neurons via CCR5 (29). Learning and memory have always been an attractive research direction in the field of neuroscience. CCR5 has had a great impact on learning and memory, resulting in many related studies. For example, a study has shown that CCR5 may affect normal learning and memory by acting on the cyclic AMP response element-binding (CREB) protein pathway (30). CCR5 was also shown to reduce the plasticity of neurons in the cerebral cortex and hippocampus-dependent learning and memory. Therefore, the effect of activating or inhibiting CCL5/CCR5 axis on memory may be opposite, and some studies have also proved this. Inhibition of CCR5 increased MAPK/CREB signaling and enhanced memory, whereas CCR5 overexpression resulted in memory impairment and decreased plasticity (31). It was also reported that CCL5 and CCR5 overexpression during aging was associated with memory impairment in mice, which was reversed by knocking out the CCR5 gene through gene editing or pharmacologic



Figure 1. CCL5/CCR5 signaling pathway. CCR5 is a 7-transmembrane G protein-coupled receptor with 3 extracellular loops and 3 intracellular loops. The disulfide bonds formed by 4 cysteine residues in the extracellular domain are vital for stabilizing the extracellular structure. Three cysteine residues on the carboxyl terminal domain can anchor the tail to the membrane. DRYLAVHA sequence on the second intracellular loop can interact with G protein. Following CCL5 binding to CCR5, G protein dissociates into the α_i and $\beta\gamma$ subunits; the former inhibits adenylate cyclase (AC), whereas the latter activates phospholipase C β (PLC β) and phosphatidylinositol 3 kinase (PI-3K). The following mediators are then activated in turn: diacylglycol (DAG), protein kinase C (PKC), inositol-1,4,5-triphosphate (IP3), intracellular Ca²⁺, calcineurin (CaN), nuclear factor of activated T cells (NFAT), extracellular signal regulated kinase 1 and 2 (ERK1/2), p38, c-Jun N-terminal kinase (JNK), protein kinase B (PKB/Akt), and Rho GTPases, which eventually affect cell migration, survival, cytoskeleton rearrangement, cell polarization, and T-cell proliferation.

inhibition with maraviroc (32). However, activation of CCR5 by CCL3 impaired synaptic plasticity of hippocampal neurons and memory, which can be alleviated by application of the CCR5 inhibitor maraviroc (33). Thus, the above evidence indicates that CCL5/CCR5 axis plays an important role in neurocognitive function, which also provides potential clues for exploring the pathogenesis of disorders associated with cognitive deficits.

CCL5/CCR5 AXIS IN AD

The neuroinflammation hypothesis of AD posits that microglia and astrocytes are activated by $A\beta$ deposition and undergo phenotypic and functional transformation to remove $A\beta$ and cell fragments; however, the secretion of cytokines, chemokines, ROS, and NO during $A\beta$ removal induces oxidative stress and inflammation, which can cause neuronal damage and promote AD progression (10, 34). For a long time in the past, reactive astrocytes and microglia were thought to have 2 phenotypes and play different functions under different pathological conditions. For reactive astrocytes, the toxic A1 type releases proinflammatory cytokines and promotes neuronal damage, and the A2 type protects neurons by secreting anti-inflammatory cytokines and neurotrophic factors (35). Microglia have processes that sense harmful signals in the environment (36). A β activation of microglia results in changes in microglia morphology and function (37). M1 microglia secrete IL-1 β , TNF- α , interferon- γ (IFN- γ), and CCL2; activate inducible NO synthase (iNOS); and induce the production of ROS to promote neuroinflammation. Meanwhile, M2 microglia release interleukin (IL)-4, IL-10, IL-13, and neurotrophic factors that have anti-inflammatory and antioxidant functions and promote the repair of cell and tissue injury to restore brain homeostasis (38, 39). However, we should emphasize that this superficial dichotomy theory has many limitations, which greatly simplifies the functional diversity of neuroglia. With the popularization of whole-genome transcriptomics, epigenomics, and proteomics in the research of reactive astrocytes and microglia, many new evidences show that their different subtypes may coexist in the CNS, and their biological functions are diverse and rich, involving novel regeneration, neural stem cell potential, regulating overactive neurons, synaptic pruning, promoting neuronal survival, and many other aspects (40, 41).

CCR5 is significantly upregulated in reactive microglia in patients with AD, which was shown to be associated with A β deposition (42). One study found that RANTES level was significantly higher in patients with AD than in control subjects and was positively correlated with IL-6 and TNF- α levels (43),

implying that CCL5 contributes to AD pathogenesis by mediating the inflammatory response and is a potential biomarker for early disease diagnosis. Genome-wide analyses have also identified CCR5 as one of the major hub genes in AD (44). In a rat model of lipopolysaccharide (LPS)-induced neuroinflammation, the CCR5 antagonist D-Ala-peptide T-amide reduced the number of microglia and astrocytes in the hippocampus as well as astrocyte hypertrophy, and microglia had slender processes similar to those observed in the nonactivated state (45). In CCR5^{-/-} mice, A β_{1-40} overexpression in hippocampus decreased the aggregation of astrocytes and microglia and alleviated cognitive impairment and synaptic dysfunction; these effects were associated with the downregulation of cyclooxygenase-2, iNOS, and nuclear factor κB (46). These findings suggest that blocking CCR5 can reduce neuroinflammation, which may be an effective therapeutic strategy for AD. As for the mechanism of CCR5 overexpression in AD, it was found that A β promoted CCR5 expression by enhancing the binding of transcription factor early growth response protein 1 (Egr-1) to the CCR5 promoter; Egr-1 gene silencing inhibited CCR5 expression and abolished chemotaxis mediated by CCR5 (47). Thus, A β may induce CCR5 expression in microglia. As immune cells of the CNS, activated microglia release many soluble factors such as TNF- α , NO, IL-10, and insulinlike growth factor-1 (IGF-1). An in vitro experiment showed that CCL5 increased NO secretion by activated microglia and reduced IL-10 and IGF-1 production, indicating that CCL5 not only exerts a chemotactic effect but also regulates microglia to enhance neuroinflammation (48).

The CCL5/CCR5 axis also mobilizes peripheral immune cells to participate in the immune response in AD. A β deposition in the brain leads to immune activation and production of antibodies that promote the clearance of A β (49). Besides antibody-mediated humoral immunity, T-cell-mediated cellular immunity is also involved in AD pathogenesis. T cells in patients with AD are activated and exist both in the periphery and infiltrate the brain (50). Additionally, a flow cytometry study analyzing the phenotypic profile of circulating immune cells in AD found that the proportion of immune cells expressing CCR5 was higher in AD patients than in age-matched controls and that CCR5 was more highly expressed on CD4⁺ versus $CD8^+$ T cells (51). CCR5 expression was almost undetectable in peripheral blood mononuclear cells (PBMCs) of healthy subjects but was upregulated in patients with AD (52). The amount of CCL5 released by PBMCs was also higher in the AD group than in the control group. Treatment with the acetylcholinesterase inhibitor pyridostigmine decreased CCL5 and CCR5 expression in PBMCs of AD patients; this was reversed by overexpression of $A\beta_{1-42}$, which also increased IFN- γ expression but decreased that of IL-4. There is also evidence of crosstalk between T cells and microglia. Microglia act as antigen-presenting cells to present $A\beta$ to T cells, which in turn induce microglial differentiation (53, 54). However, it is unclear how T cells cross the blood-brain barrier (BBB) to enter the brain in individuals with AD. One study found that T cells in patients with AD overexpress MIP-1, which binds to CCR5 expressed by brain endothelial cells to facilitate the passage of T cells through tight junctions of the BBB (55).

Furthermore, interaction of A β and receptor for advanced glycation endproducts (RAGE) expressed by cells of the BBB can activate ERK, JNK, and PI3K signaling to enhance the binding of Egr-1 to the *CCR5* promoter and expression of *CCR5*, resulting in the penetration of MIP-1-expressing T cells through the BBB (56). Thus, A β deposition acts as a signal that is transmitted via RAGE to peripheral T cells, which then promote AD development. These results indicate that the interaction between A β and the CCL5/CCR5 axis not only perturbs immune homeostasis in the CNS but also modulates systemic immunity to promote pathologic processes in AD (summarized in Fig. 2).

There have been a few studies in both animal models and in human subjects that do not support the view that CCL5/ CCR5 axis can promote the development of AD. CCR5 was shown to be downregulated in the APP/prenilin-1 transgenic mouse model of AD (57); compared with control subjects, CCL5 mRNA level in the peripheral blood was significantly lower in AD patients (58). Meanwhile, there was no difference in CCL5 expression between AD model and wild-type mice (59). These results argue against a significant role for the CCL5/CCR5 axis in AD development. However, the contribution of the CCL5/CCR5 axis to AD pathogenesis cannot be extrapolated based solely on alterations in the expression of pathway components. There is also evidence that the CCL5/ CCR5 axis has a protective role in AD. One study using CCR5^{-/-} mice showed that CCR5 deficiency accelerated LPS-induced astrogliosis activation and AB deposition and impaired memory function (60). Similarly, another study found that astrocyte activation, $A\beta_{1-42}$ deposition, and memory impairment were enhanced in CCR5-deficient mice compared with wild-type mice (61). In vitro experiments have shown that RANTES application improved the survival rate of neurons and increased their tolerance to the toxicity of sodium nitroprusside and thrombin (62), suggesting that RANTES exerts a neuroprotective effect. Additionally, transplantation of bone marrow mesenchymal stem cells into the brain of AD mice improved cognitive function, which was attributed to the action of secreted CCL5 on endogenous microglia (63).

The relationship between CCR5 and AD has been investigated from the perspective of genetic diversity, but most studies have found that the distribution of $CCR5\Delta32$ allele does not differ between AD patients and healthy individuals (64– 67). However, a large-scale cohort study reported that although there was no significant association between $CCR5\Delta32$ polymorphism and the screened neurodegenerative diseases, carriers of the $CCR5\Delta32$ allele had an earlier disease onset (68).

CONCLUSION

There is ample evidence that the immune response is involved in the pathogenesis of AD; the neuroinflammation hypothesis is highly attractive and is a powerful complement to the $A\beta$ and tau protein hypotheses. During neuroinflammation, $A\beta$ deposition activates astrocytes and microglia, with the latter transforming from a protective to a proinflammatory phenotype. Acute inflammation at the early stage is beneficial for



Figure 2. Mechanisms of the CCL5/CCR5 axis in the pathogenesis of AD. In patients with AD, $A\beta$ can upregulate CCL5 and CCR5; the resting microglia and astrocytes are transformed into the A1 and M1 phenotypes, and they secrete pro-inflammatory cytokines. $A\beta$ deposition in the brain leads to immune activation and production of antibodies. Interaction of $A\beta$ and receptor for advanced glycation endproducts (RAGE) expressed by cells of the blood-brain barrier (BBB) can promote *CCR5* expression, resulting in the penetration of MIP-1-expressing T cells through tight junctions of the BBB. There is crosstalk between T cells and microglia, microglia act as antigen-presenting cells to present $A\beta$ to T cells, which in turn induce microglia differentiation. With the development of AD, neurons eventually die.

clearing $A\beta$ and repairing neuronal damage. However, with the progression of AD, microglia and astrocytes secrete proinflammatory cytokines and chemokines, leading to excessive and uncontrolled inflammation and neuron death. The CCL5/ CCR5 axis plays an important role in learning, memory, neuroinflammation, and AD pathogenesis; however, the mechanistic details of CCL5/CCR5 axis in AD have not been fully elucidated and some of the existing evidence is contradictory. Even population-based studies on the distribution of the CCR5 Δ 32 allele have shown no association between CCL5/ CCR5 and AD development. A reason for these conflicting findings may be differences in animal models and populations, but it is also possible that CCR5 has as-yet unidentified functions in the CNS. More studies are needed to explore the specific role of the CCL5/CCR5 signaling axis in AD pathogenesis in order to determine whether it can serve as a therapeutic target in treatment of AD. In addition, we believe that it will also be a potentially valuable idea to expand the scope of exploring CCL5/CCR5 axis to other neurodegenerative diseases.

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CONFLICT OF INTEREST

The authors have no duality or conflicts of interest to declare.

REFERENCES

- Jia L, Du Y, Chu L, et al.; COAST Group. Prevalence, risk factors, and management of dementia and mild cognitive impairment in adults aged 60 years or older in China: A cross-sectional study. Lancet Public Health 2020;5:e661–71
- Watermeyer T, Calia C. Neuropsychological assessment in preclinical and prodromal Alzheimer disease: A global perspective. J Glob Health 2019;9:010317
- 3. Hugo J, Ganguli M. Dementia and cognitive impairment: Epidemiology, diagnosis, and treatment. Clin Geriatr Med 2014;30:421-42
- Braak H, Braak E. Neuropathological stageing of Alzheimer-related changes. Acta Neuropathol 1991;82:239–59
- Webers A, Heneka MT, Gleeson PA. The role of innate immune responses and neuroinflammation in amyloid accumulation and progression of Alzheimer's disease. Immunol Cell Biol 2020;98: 28–41
- Andrieu S, Gillette S, Amouyal K, et al.; EPIDOS study. Association of Alzheimer's disease onset with ginkgo biloba and other symptomatic cognitive treatments in a population of women aged 75 years and older from the EPIDOS study. J Gerontol A Biol Sci Med Sci 2003;58:372–7

- Wang Y, Mandelkow E. Tau in physiology and pathology. Nat Rev Neurosci 2016;17:5–21
- Lannfelt L, Relkin NR, Siemers ER. Amyloid-ß-directed immunotherapy for Alzheimer's disease. J Intern Med 2014;275:284–95
- 9. Congdon EE, Sigurdsson EM. Tau-targeting therapies for Alzheimer disease. Nat Rev Neurol 2018;14:399–415
- 10. Wilson DM III, Cookson MR, Van Den Bosch L, et al. Hallmarks of neurodegenerative diseases. Cell 2023;186:693-714
- Kinney JW, Bemiller SM, Murtishaw AS, et al. Inflammation as a central mechanism in Alzheimer's disease. Alzheimers Dement (N Y) 2018;4:575–90
- 12. Luster AD. Chemokines—Chemotactic cytokines that mediate inflammation. N Engl J Med 1998;338:436–45
- Riviere-Cazaux C, Cornell J, Shen Y, Zhou M. The role of CCR5 in HIV-associated neurocognitive disorders. Heliyon 2022;8:e09950
- 14. Aldinucci D, Borghese C, Casagrande N. The CCL5/CCR5 axis in cancer progression. Cancers (Basel) 2020;12:1765
- Tan Q, Zhu Y, Li J, et al. Structure of the CCR5 chemokine receptor-HIV entry inhibitor maraviroc complex. Science 2013; 341:1387–90
- Oppermann M. Chemokine receptor CCR5: Insights into structure, function, and regulation. Cell Signal 2004;16:1201–10
- Fantuzzi L, Tagliamonte M, Gauzzi MC, et al. Dual CCR5/CCR2 targeting: Opportunities for the cure of complex disorders. Cell Mol Life Sci 2019;76:4869–86
- Alkhatib G, Combadiere C, Broder CC, et al. CC CKR5: A RANTES, MIP-1alpha, MIP-1beta receptor as a fusion cofactor for macrophage-tropic HIV-1. Science 1996;272:1955–58
- Gupta RK, Abdul-Jawad S, McCoy LE, et al. HIV-1 remission following CCR5Δ32/Δ32 haematopoietic stem-cell transplantation. Nature 2019;568:244–8
- Zhao J, Ma L, Wu YL, et al. Chemokine receptor CCR5 functionally couples to inhibitory G proteins and undergoes desensitization. J Cell Biochem 1998;71:36–45
- Ganju RK, Dutt P, Wu L, et al. Beta-chemokine receptor CCR5 signals via the novel tyrosine kinase RAFTK. Blood 1998;91:791–7
- 22. Wong M, Uddin S, Majchrzak B, et al. Rantes activates Jak2 and Jak3 to regulate engagement of multiple signaling pathways in T cells. J Biol Chem 2001;276:11427-31
- Burgering BM, Coffer PJ. Protein kinase B (c-Akt) in phosphatidylinositol-3-OH kinase signal transduction. Nature 1995;376:599–602
- Downward J. PI 3-kinase, Akt and cell survival. Semin Cell Dev Biol 2004;15:177–82
- Westmoreland SV, Alvarez X, deBakker C, et al. Developmental expression patterns of CCR5 and CXCR4 in the rhesus macaque brain. J Neuroimmunol 2002;122:146–58
- Alvarez S, Jiménez JL, Serramía MJ, et al. Lack of association of HIV-1 biological or molecular properties with neurotropism for brain cells. J Mol Neurosci 2006;29:131–44
- Boutet A, Salim H, Leclerc P, et al. Cellular expression of functional chemokine receptor CCR5 and CXCR4 in human embryonic neurons. Neurosci Lett 2001;311:105–8
- Ji JF, He BP, Dheen ST, et al. Expression of chemokine receptors CXCR4, CCR2, CCR5 and CX3CR1 in neural progenitor cells isolated from the subventricular zone of the adult rat brain. Neurosci Lett 2004;355:236–40
- 29. Musante V, Longordo F, Neri E, et al. RANTES modulates the release of glutamate in human neocortex. J Neurosci 2008;28: 12231-40
- Sano Y, Shobe JL, Zhou M, et al. CREB regulates memory allocation in the insular cortex. Curr Biol 2014;24:2833–7
- Zhou M, Greenhill S, Huang S, et al. CCR5 is a suppressor for cortical plasticity and hippocampal learning and memory. Elife 2016;5: e20985
- 32. Shen Y, Zhou M, Cai D, et al. CCR5 closes the temporal window for memory linking. Nature 2022;606:146–52

- Marciniak E, Faivre E, Dutar P, et al. The Chemokine MIP-1α/ CCL3 impairs mouse hippocampal synaptic transmission, plasticity and memory. Sci Rep 2015;5:15862
- Frozza RL, Lourenco MV, De Felice FG. Challenges for Alzheimer's disease therapy: Insights from novel mechanisms beyond memory defects. Front Neurosci 2018;12:37
- Liddelow SA, Barres BA. Reactive astrocytes: Production, function, and therapeutic potential. Immunity 2017;46:957–67
- Kettenmann H, Hanisch UK, Noda M, et al. Physiology of microglia. Physiol Rev 2011;91:461–553
- 37. Yu Y, Ye RD. Microglial A β receptors in Alzheimer's disease. Cell Mol Neurobiol 2015;35:71–83
- Varnum MM, Ikezu T. The classification of microglial activation phenotypes on neurodegeneration and regeneration in Alzheimer's disease brain. Arch Immunol Ther Exp (Warsz) 2012;60:251–66
- Orihuela R, McPherson CA, Harry GJ. Microglial M1/M2 polarization and metabolic states. Br J Pharmacol 2016;173:649–65
- Escartin C, Galea E, Lakatos A, et al. Reactive astrocyte nomenclature, definitions, and future directions. Nat Neurosci 2021;24: 312–25
- Wang J, He W, Zhang J. A richer and more diverse future for microglia phenotypes. Heliyon 2023;9:e14713
- 42. Xia MQ, Qin SX, Wu LJ, et al. Immunohistochemical study of the beta-chemokine receptors CCR3 and CCR5 and their ligands in normal and Alzheimer's disease brains. Am J Pathol 1998;153: 31–7
- 43. Vacinova G, Vejražkova D, Rusina R, et al. Regulated upon activation, normal T cell expressed and secreted (RANTES) levels in the peripheral blood of patients with Alzheimer's disease. Neural Regen Res 2021;16:796–800
- Wee JJ, Kumar S. Prediction of hub genes of Alzheimer's disease using a protein interaction network and functional enrichment analysis. Genomics Inform 2020;18:e39
- 45. Rosi S, Pert CB, Ruff MR, et al. Chemokine receptor 5 antagonist D-Ala-peptide T-amide reduces microglia and astrocyte activation within the hippocampus in a neuroinflammatory rat model of Alzheimer's disease. Neuroscience 2005;134:671–6
- 46. Passos GF, Figueiredo CP, Prediger RD, et al. Role of the macrophage inflammatory protein-1alpha/CC chemokine receptor 5 signaling pathway in the neuroinflammatory response and cognitive deficits induced by beta-amyloid peptide. Am J Pathol 2009;175: 1586–97
- 47. Giri RK, Rajagopal V, Shahi S, et al. Mechanism of amyloid peptide induced CCR5 expression in monocytes and its inhibition by siRNA for Egr-1. Am J Physiol Cell Physiol 2005;289: C264–76
- Skuljec J, Sun H, Pul R, et al. CCL5 induces a pro-inflammatory profile in microglia in vitro. Cell Immunol 2011;270:164–71
- 49. Bard F, Cannon C, Barbour R, et al. Peripherally administered antibodies against amyloid beta-peptide enter the central nervous system and reduce pathology in a mouse model of Alzheimer disease. Nat Med 2000;6:916–9
- 50. Town T, Tan J, Flavell RA, et al. T-cells in Alzheimer's disease. Neuromolecular Med 2005;7:255–64
- Goldeck D, Larbi A, Pellicanó M, et al. Enhanced chemokine receptor expression on leukocytes of patients with Alzheimer's disease. PLoS One 2013;8:e66664
- Reale M, Iarlori C, Feliciani C, et al. Peripheral chemokine receptors, their ligands, cytokines and Alzheimer's disease. J Alzheimers Dis 2008;14:147–59
- Monsonego A, Weiner HL. Immunotherapeutic approaches to Alzheimer's disease. Science 2003;302:834–8
- Séguin R, Biernacki K, Prat A, et al. Differential effects of Th1 and Th2 lymphocyte supernatants on human microglia. Glia 2003;42: 36–45
- Man SM, Ma YR, Shang DS, et al. Peripheral T cells overexpress MIP-1alpha to enhance its transendothelial migration in Alzheimer's disease. Neurobiol Aging 2007;28:485–96

- 56. Li M, Shang DS, Zhao WD, et al. Amyloid beta interaction with receptor for advanced glycation end products up-regulates brain endothelial CCR5 expression and promotes T cells crossing the blood-brain barrier. J Immunol 2009;182:5778–88
- Jorda A, Cauli O, Santonja JM, et al. Changes in chemokines and chemokine receptors expression in a mouse model of Alzheimer's disease. Int J Biol Sci 2019;15:453–63
- Kester MI, van der Flier WM, Visser A, et al. Decreased mRNA expression of CCL5 [RANTES] in Alzheimer's disease blood samples. Clin Chem Lab Med 2011;50:61–5
- Vérité J, Janet T, Chassaing D, et al. Longitudinal chemokine profile expression in a blood-brain barrier model from Alzheimer transgenic versus wild-type mice. J Neuroinflammation 2018;15:182
- Hwang CJ, Park MH, Hwang JY, et al. CCR5 deficiency accelerates lipopolysaccharide-induced astrogliosis, amyloid-beta deposit and impaired memory function. Oncotarget 2016;7:11984–99
- Lee YK, Kwak DH, Oh KW, et al. CCR5 deficiency induces astrocyte activation, Abeta deposit and impaired memory function. Neurobiol Learn Mem 2009;92:356–63
- Tripathy D, Thirumangalakudi L, Grammas P. RANTES upregulation in the Alzheimer's disease brain: A possible neuroprotective role. Neurobiol Aging 2010;31:8–16

- 63. Lee JK, Schuchman EH, Jin HK, et al. Soluble CCL5 derived from bone marrow-derived mesenchymal stem cells and activated by amyloid β ameliorates Alzheimer's disease in mice by recruiting bone marrow-induced microglia immune responses. Stem Cells 2012;30:1544–55
- 64. Balistreri CR, Grimaldi MP, Vasto S, et al. Association between the polymorphism of CCR5 and Alzheimer's disease: Results of a study performed on male and female patients from Northern Italy. Ann N Y Acad Sci 2006;1089:454–61
- 65. Khorram Khorshid HR, Manoochehri M, Nasehi L, et al. Ccr2-64i and Ccr5 Δ 32 polymorphisms in patients with late-onset Alzheimer's disease; A study from Iran (Ccr2-64i And Ccr5 Δ 32 polymorphisms in Alzheimer's disease). Iran J Basic Med Sci 2012;15:937–44
- Galimberti D, Fenoglio C, Lovati C, et al. CCR2-64I polymorphism and CCR5Delta32 deletion in patients with Alzheimer's disease. J Neurol Sci 2004;225:79–83
- 67. Combarros O, Infante J, Llorca J, et al. The chemokine receptor CCR5-Delta32 gene mutation is not protective against Alzheimer's disease. Neurosci Lett 2004;366:312–4
- Wojta KJ, Ayer AH, Ramos EM, et al. Lack of association between the CCR5-delta32 polymorphism and neurodegenerative disorders. Alzheimer Dis Assoc Disord 2020;34:244–7