# **Molecules and Cells**



# 3

# Sarcoma Immunotherapy: Confronting Present Hurdles and Unveiling Upcoming Opportunities

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Sarcomas are rare and heterogeneous mesenchymal neoplasms originating from the bone or soft tissues. which pose significant treatment challenges. The current standard treatment for sarcomas consists of surgical resection, often combined with chemo- and radiotherapy; however, local recurrence and metastasis remain significant concerns. Although immunotherapy has demonstrated promise in improving long-term survival rates for certain cancers, sarcomas are generally considered to be relatively less immunogenic than other tumors, presenting substantial challenges for effective immunotherapy. In this review, we examine the possible opportunities for sarcoma immunotherapy, noting cancer testis antigens expressed in sarcomas. We then cover the current status of immunotherapies in sarcomas, including progress in cancer vaccines, immune checkpoint inhibitors, and adoptive cellular therapy and their potential in combating these tumors. Furthermore, we discuss the limitations of immunotherapies in sarcomas, including a low tumor mutation burden and immunosuppressive tumor microenvironment, and explore potential strategies to tackle the immunosuppressive

barriers in therapeutic interventions, shedding light on the development of effective and personalized treatments for sarcomas. Overall, this review provides a comprehensive overview of the current status and potential of immunotherapies in sarcoma treatment, highlighting the challenges and opportunities for developing effective therapies to improve the outcomes of patients with these rare malignancies.

**Keywords:** bone cancer, immunotherapy, sarcomas, soft tissue sarcoma, tumor microenviorment

### **INTRODUCTION**

Sarcomas represent a heterogeneous group of mesenchymal neoplasms originating from the bone or soft tissues such as the cartilage, muscle, and other connective tissues. Sarcomas account for 15% of childhood and 1% of all adult malignancies. Despite being a rare malignancy, approximately 13,190 patients in the United States were diagnosed with sarcoma in

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2022, with 5,130 fatalities (Siegel et al., 2022). Patients with sarcomas experience a high probability of local recurrence (5-year local recurrence rate of 19%-51%) (Pawlik et al., 2006; Thorkildsen et al., 2021; Xie et al., 2015; Zhao et al., 2018), and repeated relapses eventually accompany distant metastasis (Panagi et al., 2022). Sarcomas are classified into soft tissue sarcomas (STSs), including rhabdomyosarcoma and liposarcoma, and bone sarcomas (BSs), including osteosarcoma and chondrosarcoma, encompassing more than 70 distinct subtypes (Hui, 2016). The main treatment challenge to overcome is the propensity for local recurrence or eventual metastatic spread despite extensive adjuvant therapy (Goorin et al., 1984; Patel et al., 2017; von Konow et al., 2021), which is related to the substantial heterogeneity of these tumors, leading to varied symptoms, treatment responses, and prognoses among patients (Du et al., 2020; Grunewald et al., 2020). Delayed diagnosis is common, with symptoms in the pediatric population often misdiagnosed as growing pains, further narrowing the available treatment options (Lodhia et al., 2022).

Surgical removal is the standard treatment approach for localized, clinically resectable STSs and BSs (Blasius et al., 2022; Gronchi, 2021). Radiotherapy can also be administered in neoadjuvant, adjuvant, or definitive settings (Hoefkens et al., 2016), although the optimal approach varies depending on individual cases. Chemotherapy is recommended for patients with high-grade STSs following resection, but has failed to demonstrate distinct improvements in terms of relapse-free survival or overall survival (OS) (Squires et al., 2020). Although BS subtypes generally do not respond well to chemotherapy, multidrug chemotherapy has shown moderate treatment efficacy for osteosarcoma and Ewing sarcoma in combination with surgery (Ingley et al., 2022). Chondrosarcoma, another common BS subtype, is generally refractory to conventional chemotherapy and radiation therapy, with no supporting efficacy data available (Gelderblom et al., 2008; Monga et al., 2020; Riedel et al., 2009). Metastatic sarcomas exhibit substantial resistance to chemo- and radiotherapy (OIivier et al., 2015). Although the exact mechanisms underlying chemotherapy resistance remain elusive, emerging evidence implicates epigenetic (Rytlewski et al., 2022) and genetic modifications (Kim et al., 2020), and epithelial-mesenchymal transition (EMT) plasticity (Sannino et al., 2017), and the tumor microenvironment (TME) (Tu et al., 2016) as contributory factors. Sarcomas exhibit radioresistance, often attributed to epigenetic changes (de Jong et al., 2019), non-coding RNAs (Chen et al., 2022), and slow cell division rates (Chen et al., 2022; Li et al., 2014). Collectively, current adjuvant chemoand radiotherapies are insufficient in preventing the metastasis and recurrence of sarcomas, and there remains uncertainty regarding the best resection margins and appropriate use of chemo- and radiotherapy (Hoefkens et al., 2016; Zhang et al., 2022).

Immunotherapy, a strategy that modulates the immune system to target cancer cells, has demonstrated promise for several cancer types (Tan et al., 2020; Waldman et al., 2020), with significantly improved long-term survival rates in melanoma and lymphomas, eliciting less cytotoxicity compared to traditional chemotherapies (Abramson et al., 2020; Tan et al., 2020). Notably, immunotherapy can achieve simultaneous local and distant control while adapting to changes in antigen expression over time, which is significant for preventing recurrent and metastatic tumors (Finkelstein et al., 2012). Cancer immunotherapy relies on two main immune mechanisms: immune surveillance and immune editing. Immune surveillance involves the detection and elimination of tumors by the immune system (Zitvogel et al., 2006), whereas immune editing modifies tumor immunogenicity to either strengthen the immune system's capacity to eliminate tumor cells or prevent the emergence of immune-resistant tumor variants (O'Donnell et al., 2019). By selectively manipulating the anti-tumor immune mechanism, immunotherapy has potential to prevent cancer metastasis and recurrence (Edwards et al., 2021).

Several successful cancer immunotherapies have emerged recently utilizing cancer vaccines, adoptive transfer therapy, or cytokine therapy (Zhang and Zhang, 2020). However, sarcomas are considered to be relatively less immunogenic than other cancers (Rytlewski et al., 2021; Weng et al., 2022; Zhu et al., 2020). Specifically, sarcomas lack well-established antigens and often display an immunosuppressive TME. Therefore, understanding the immune escape mechanisms and immunosuppressive TME in sarcoma is essential for developing effective sarcoma immunotherapy. In this review, we provide an update of the current status of sarcoma immunotherapy, focusing on the needs, emerging strategies, and challenges.

# IMMUNOTHERAPIES: ADDRESSING UNMET NEEDS FOR SARCOMA TREATMENT

#### Cancer testis antigen-based vaccine therapy

Sarcomas typically express cancer testis antigens (CTAs), a group of tumor-associated antigens found predominantly in male germ cells in the testis but not in the adult somatic tissue. CTAs are generally quiescent in healthy tissues, but their expression is induced in various malignancies (Whitehurst, 2014). The epitopes of CTAs can be recognized by T cells, facilitating immune activation (Juretic et al., 2003).

Among the several CTAs identified in sarcomas to date, New York esophageal squamous cell carcinoma 1 (NY-ESO-1), melanoma-associated antigen (MAGE), and preferentially expressed antigen of melanoma (PRAME) are the most prevalent (Kakimoto et al., 2019; Wei et al., 2020). NY-ESO-1 is detected in various sarcoma subtypes, including myxoid liposarcomas, osteosarcomas, and synovial sarcomas (Hashimoto et al., 2022; Jungbluth et al., 2001; Kakimoto et al., 2019). MAGE expression has been observed in osteosarcoma, synovial sarcoma, and myxoid/round cell liposarcoma, with distinct MAGE subtypes exhibiting variable expression levels. For instance, osteosarcoma exhibited high expression of MAGE-A1, -A2, and -A3, but low expression of MAGE-A12 (Zou et al., 2012). Synovial sarcoma and myxoid/ round cell liposarcoma displayed elevated levels of MAGE-A4 (Hemminger et al., 2014; lura et al., 2017). PRAME expression is elevated in osteosarcoma, synovial sarcoma, and myxoid/round cell liposarcoma (Epping et al., 2005; Wei et al., 2020).

The potential of CTAs as tumor vaccines has been explored



**Fig. 1. Current immunotherapies used for sarcoma treatment.** (A) Cancer vaccine: Dendritic cells present sarcoma-derived antigen or neoantigen peptides through MHC class II to T cell receptor (TCR), activating T cells with CD28-CD80/86 co-stimulation. MHC class I presents tumor-specific antigens such as MAGE-A1 or NY-ESO-1, leading to cytotoxic T-lymphocyte (CTL)-mediated killing of sarcoma cells. (B) Immune checkpoint blockade: Sarcoma cells evade T cell-mediated cytotoxicity by expressing immune checkpoint ligands such as programmed cell death ligand 1 (PD-L1), which suppresses T cell activation by binding with complementary receptors on T cells such as programmed cell death 1 (PD-1). Immune checkpoint blockade antibodies targeting PD-L1 or PD-1 interfere with this interaction. Similarly, blocking the cytotoxic T lymphocyte associated antigen 4 (CTLA-4) receptor binding with CD80/86 complex prevents CTL suppression. (C) Adoptive cell therapy: Engineered TCR-T and chimeric antigen receptor (CAR)-T cells specifically engage with tumor cells that express the target antigen on their surface. CAR-T cells are designed to bind with sarcoma cells independent of MHC class I complex. MAGE, melanoma-associated antigen; NY-ESO-1, New York esophageal squamous cell carcinoma 1; HER2, human epidermal growth factor receptor 2.

with the aim of enhancing immunological recognition and improving immune responses against tumors (Fig. 1A). Tumor vaccines can be classified into two distinct categories: peptide-based vaccines, which involve the direct delivery of antigens, and dendritic cell (DC)-based vaccines, which employ antigen-presenting DCs for targeted immunotherapy (Saxena et al., 2021; Tagliamonte et al., 2014). NY-ESO-1 peptide was successfully delivered using a Salmonella typhimurium vaccine strain, eliciting NY-ESO-1-specific CD8<sup>+</sup> and CD4<sup>+</sup> T cells in peripheral blood mononuclear cells from patients (Nishikawa et al., 2006). Treatment of another NY-ESO-1 vaccine, CDX-1401, induced both humoral and cellular immunity to NY-ESO-1 in 45 patients with multiple malignant tumors, including sarcomas, 13 of whom experienced disease stabilization and two of whom exhibited tumor regression (Dhodapkar et al., 2014). This first-in-human study of DC-based vaccines showed promise in treating sarcomas. However, the application of CTA-targeting vaccines in sarcomas has yielded inadequate results compared to that for other malignant tumors. Possible explanations for this discrepancy include low T cell functional avidity, high regulatory T cell ( $T_{reg}$ ) activity, and higher expression of immune checkpoints such as programmed cell death 1 (PD-1) in sarcomas (Wei et al., 2020).

### Immune checkpoint inhibitors

The expression of immune checkpoint proteins such as PD-1 and its ligand (PD-L1), as well as cytotoxic T lymphocyte associated antigen 4 (CTLA-4), is a key immune evasion mechanism of tumors (Beatty and Gladney, 2015; Lee et al., 2021b). Large cohort studies demonstrated the presence of PD-1 (43% of sarcoma cases) (Kim et al., 2016) and CTLA-4 (32% of sarcoma cases) on T cells, as well as PD-L1 (23% of cases) on tumor cells in various STSs and BSs (Paydas et al., 2016; Samiei et al., 2022; Zheng et al., 2015). These findings underscore the potential of these checkpoint proteins as emerging therapeutic targets for immunotherapies in sarcoma treatment (Fig. 1B).

To date, clinical trials evaluating the efficacy of immune checkpoint inhibitors (ICIs) in patients with STSs have demon-

strated modest results, with varying responses depending on treatment regimens or histological subtypes. Pembrolizumab, a PD-1 monoclonal antibody, was assessed in a phase II trial involving STSs and BSs, yielding meaningful results in undifferentiated pleomorphic sarcoma (objective response in 40% of patients) and liposarcoma (objective response in 20% of patients), while demonstrating limited efficacy in BS (Tawbi et al., 2017). Additionally, pembrolizumab exhibited high-level, prolonged anti-tumor activity in select sarcoma histotypes such as alveolar soft part sarcoma and chordoma, with 6-month OS rates of 100% and 75%, respectively (NCT03012620). Ipilimumab, a CTLA-4 antibody, and nivolumab, a PD-1 antibody, are two other notable immune checkpoint blockade antibodies promoting anti-tumor immunity in sarcoma (Tawbi et al., 2017; Weber, 2007), Nivolumab resulted in a complete or partial response in 5% of patients with advanced sarcoma when delivered as a monotherapy, whereas ipilimumab monotherapy resulted in a 7% partial response rate; their combination displayed promising efficacy with three times higher confirmed objective responses (16%) in multiple sarcoma subtypes (D'Angelo et al., 2018). A recent clinical trial demonstrated a progression-free survival (PFS) rate of 49% at 12 weeks in patients with various advanced or metastatic sarcomas that received combined therapy with durvalumab (anti-PD-L1) and tremelimumab (anti-CTLA-4) (Somaiah et al., 2022).

#### Adoptive cellular therapy

Immune evasion in cancer can arise from a lack of neo-antigens or defects in antigen presentation. Adoptive cellular therapy (ACT), or cellular immunotherapy, has potential to surmount these challenges by modifying patient-specific T cells to enhance the targeting of tumor-specific antigens. ACT is a personalized treatment approach involving the isolation, *ex vivo* modification, expansion, and reinfusion of a patient's own immune cells, thus bypassing antigen presentation defects (Zhao and Cao, 2019). Various forms of ACT exist, including engineered T cell receptor (TCR) and chimeric antigen receptor (CAR) T cell therapy (June et al., 2018) (Fig. 1C).

TCR therapies using Afamitresgene autoleucel (afami-cel) and Letetresgene autoleucel (lete-cel) are currently under assessment in a phase I/II trial to induce anti-tumor responses in patients with synovial sarcoma and myxoid/round cell liposarcoma. These therapies consist of autologous CD4<sup>+</sup> and CD8<sup>+</sup> T cells that are genetically modified to express a TCR recognizing MAGE-A4 (afami-cel) or NY-ESO1 (lete-cel) bound to human leukocyte antigen A02. Pooled data from 69 patients (phase I, n = 18; phase II, n = 51) showed an investigator-assessed objective response rate of 36.2% (40.7% in synovial sarcoma; 10.0% in myxoid/round cell liposarcoma) (D'Angelo et al., 2022).

Human epidermal growth factor receptor 2 (HER2) is a potent antigen identified in various sarcomas, albeit with low expression (Ahmed et al., 2009; Yan et al., 2015). CAR-T therapy targeting HER2 exhibited tumor regression in both localized and metastatic lesions, with prolonged survival in mice inoculated even with the low-HER2-expressing osteosarcoma cell line (Ahmed et al., 2009). In a clinical phase I/II study of

HER2 CAR-T cells to treat HER2-positive osteosarcoma, Ewing sarcoma, neuroectodermal tumor, and desmoplastic small round cell tumor, 4 of the 17 patients experienced stable disease for 12 weeks to 14 months and three patients had their residual tumors removed without additional therapy, with one specimen achieving ≥90% tumor necrosis (Ahmed et al., 2015). Combination of this therapy with lympho-depleting chemotherapy showed further improvement in outcomes (Navai et al., 2019). The ganglioside antigen GD2 is expressed in Ewing sarcoma and osteosarcoma (Kailayangiri et al., 2012; Roth et al., 2014). GD2-specific CAR-T cells in combination with AMG102, a human growth factor receptor neutralizing antibody, inhibited the primary tumor growth and metastasis of Ewing sarcoma (Charan et al., 2020).

# TACKLING IMMUNOSUPPRESSIVE BARRIERS IN THERAPEUTIC INTERVENTIONS FOR SARCOMA

Immunotherapy has shown beneficial results in treating various cancers, yet its efficacy in STSs and BSs remains restricted to specific subtypes (Ayodele and Razak, 2020; Panagi et al., 2022), necessitating further strategies to improve efficacy (Klemen et al., 2021). Some combined immunotherapies, in conjunction with chemotherapy, radiotherapy, and targeted therapies, have demonstrated improved prognosis in sarcomas (D'Angelo et al., 2018; Husain et al., 2023; Kelly et al., 2020; Tang et al., 2021). However, challenges persist in treating immunologically "cold" sarcomas. In this section, we discuss key mechanisms that might contribute to these challenges and the approaches developed to potentially overcome them (Fig. 2).

#### Low tumor mutation burden

Tumor mutation burden (TMB), reflecting the number of mutations harbored by tumors, is a valuable biomarker positively related to the immunotherapy response (Strickler et al., 2021). Among 151 immunotherapy-treated patients, a higher TMB correlated with better outcomes, including response rate, PFS, and OS (Goodman et al., 2017). This favorable response is attributed to a positive correlation between TMB and immunogenic neoantigens (Lagos et al., 2020).

The U.S. Food and Drug Administration approved pembrolizumab for cancers with a TMB  $\geq$  10 mutations/Mb, indicating therapeutic implications based on gualitative next-generation sequencing; however, no patients with sarcoma were enrolled in this study (Marabelle et al., 2020). According to a comprehensive mutational analysis pipeline that incorporated clinical data, it was found that among 206 cases of STS, patients exhibited an average TMB of 1.06 mutations/Mb (Cancer Genome Atlas Research Network, 2017). In the case of BSs, TMB ranged from 0 to 7 mutations/Mb among 31 osteosarcoma patients. The 3rd quartile of TMB in this patient cohort was determined to be 2.565 mutations/Mb (Xie et al., 2021). Collectively, sarcomas are primarily characterized by low TMB (compared to melanoma and lung cancers, with 20% of patients meeting the TMB cutoff of  $\geq$ 10 mutations/ Mb for pembrolizumab), rendering them immunologically cold tumors (Doig et al., 2022). This poses a considerable challenge for the development of effective immunotherapies



**Fig. 2. Barriers to immunotherapy in sarcomas and strategies for modulating their tumor microenvironment.** Sarcomas are primarily recognized by their low tumor mutation burden, which classifies them as tumors displaying limited immune reactivity. The immunosuppressive features characterizing the tumor microenvironment (TME) of sarcomas are correlated with the existence of regulatory T cells, tumor-associated macrophages (TAM), and myeloid-derived suppressor cells (MDSCs) (left). These components present prospective targets for crafting immunotherapy approaches with enhanced efficacy in tackling sarcomas (right). A personalized cancer vaccine addresses low tumor mutation burden by utilizing neoantigens to activate precise cytotoxic T cells targeting sarcoma tumors. TAMs can be regulated by inhibiting M2-like TAMs or by stimulating M1-like macrophages to amplify anti-tumor immune responses. Approaches that combine imatinib with CTLA-4 blockade or employ anti-CD25 antibodies alongside PD-1 blockade can control regulatory T cells (T<sub>regs</sub>) and improve treatment outcomes. CCR4 inhibitors have shown potential in reducing T<sub>reg</sub> infiltration. Cancer-associated fibroblasts (CAFs) expressing LRRC15 have emerged as promising targets, with the antibody-drug conjugate ABBV-085, which targets LRRC15, displaying notable anti-tumor activity in clinical trials. These strategies seek to overcome the immunologically cold TME in sarcomas, enhancing the efficacy of immunotherapy. MSCs, mesenchymal stem cells; IDO, indoleamine 2,3-dioxygenase; TMB, tumor mutation burden; TCR, T cell receptor.

for sarcoma patients.

To overcome the challenge of low TMB, recent strides in cancer immunotherapy have explored the capacity of neoantigens to stimulate cytotoxic T cells, targeting sarcoma tumor cells with distinct specificity (Gubin et al., 2014; Yang et al., 2019). Originating from somatic mutations or chromosomal rearrangements, neoantigens are frequently present in sarcomas. In synovial sarcomas, peptides from the typical SYT-SSX fusion have been effective in generating specific CTLs for synovial sarcoma (Kawaguchi et al., 2012; Sato et al., 2002). The implementation of a customized immunotherapy strategy, tailored to the distinctive tumor traits of each individual, presents exciting new opportunities for effectively treating sarcomas with low TMB.

#### Immunosuppressive TME

The TME plays a pivotal role in shaping the immune response to cancer cells; thus, understanding its composition is essential for developing effective immunotherapies. Petitprez et al.

(2020) demonstrated that STSs with immune-deprived TMEs displayed low expression of immune checkpoint-related genes (PD-1, PD-L2, CTLA-4, and TIM3). In contrast, B-cellrich tertiary lymphoid structures, predominantly observed in immune-rich classes, were positively correlated with improved survival and higher response rates to PD-1 blockade. The immune cell distribution within the TME critically influences the immune response in sarcomas. Tight-junction protein 1 (TJP1) may play a role in shaping the immunosuppressive landscape of TME in leiomyosarcoma. TJP1 contributes to tumor growth by modulating cell-cell aggregation and cytokine-mediated communication in TME. Notably, TJP1 exhibits a negative correlation with expression of colony stimulating factor 1 (CSF1), which typically fosters an immunosuppressive TME through macrophage regulation (Lee et al., 2021a). Only 22% and 12% of 50 sarcoma specimens exhibited PD-1 and PD-L1 expression, respectively, whereas 75% of the specimens contained FOXP3  $^{\!\!+}$   $T_{\scriptscriptstyle regs}$  in the immunological milieu (D'Angelo et al., 2015). Single-cell transcriptomic pro-

filing of osteosarcomas and their TMEs identified 11 main cell clusters such as osteoblastic cells, myeloid cells, mesenchymal stem cells (MSCs), fibroblasts, and tumor-infiltrating lymphocytes (TILs). Notably, the TILs cluster displayed high expression of T cell exhaustion inhibitory receptors, including TIGIT and LAG3, and blocking TIGIT enhanced the cytotoxic effects of CD3<sup>+</sup> T cells against osteosarcoma cells (Zhou et al., 2020).

Antigen presentation by tumor-associated macrophages (TAMs) contributes to anti-tumor immune responses. M1-like macrophages exhibit anti-tumoral functions, whereas M2like macrophages exhibit pro-tumoral functions. Immunohistochemistry of STS samples demonstrated the widespread presence of macrophages in the TME, which were polarized toward the M2-like phenotype. Concurrent expression of the M2-like macrophage marker CD163, along with IL-10 was also observed. Notably, the favorable prognostic role of MS4A1-expressing CD20-positive B cells was not apparent in tumors with high expression of IL-10 and CD163. This finding underscores the rationale for targeting M2-polarized macrophages in STS (Tsagozis et al., 2019), which can be achieved by either inhibiting M2-like TAMs or activating M1-like macrophages.

The TMEs of multiple murine sarcoma models induced tumors to produce retinoic acid (RA), which promoted the differentiation to M2-like macrophages and suppressed DCs. Reducing tumor RA production and inhibiting RA-receptor signaling enhanced anti-tumor immunity. Human sarcomas such as leiomyosarcoma, dedifferentiated liposarcoma, myxofibrosarcoma, undifferentiated pleomorphic sarcoma, and synovial sarcoma, also exhibited RA-mediated immunosuppression (Devalaraja et al., 2020). Inhibition of TREM2, an oncogenic marker found in tumor-infiltrating macrophages, increased the responsiveness to anti-PD-1 immunotherapy in sarcoma model mice. Deletion of TREM2 or administering anti-TREM2 treatments resulted in decreased tumor infiltration of CX3CR1<sup>+</sup>/MRC1<sup>+</sup> TAMs and expansion of myeloid subsets that express immunostimulatory molecules improving T cell responses (Molgora et al., 2020). Moreover, various strategies investigated in other types of cancer may also hold promise for sarcoma treatment. Dual-targeted nanoparticles containing small interfering RNA against colony-stimulating factor-1 receptor depleted M2-like macrophages, increasing pro-inflammatory cytokines and decreasing immunosuppressive cytokines (Qian et al., 2017). R848, an agonist of Tolllike receptors 7 and 8, was identified as a potent driver of the M1 phenotype, and its nanoparticle-mediated delivery significantly shifted TAMs toward the M1 phenotype, leading to inhibited tumor growth (Rodell et al., 2018).

Myeloid-derived suppressor cells (MDSCs) encompass a diverse range of immature and developed myeloid cells that play crucial roles in cancer-associated immunosuppression (Nakamura and Smyth, 2020). MDSCs are divided into monocytic (M-MDSC) and granulocytic (G-MDSC) subgroups. Highfill et al. (2014) showed that G-MDSCs could suppress T cells in rhabdomyosarcoma. G-MDSCs are recruited to tumors via the CXCR2 signaling pathway, leading to immunosuppressive consequences and unfavorable outcomes for anti-PD-1 therapies in patients with rhabdomyosarcoma.

Stromal regions in the TME contain blood and lymphatic endothelial cells, MSCs, cancer-associated fibroblasts (CAFs), pericytes, and other cell types (Turley et al., 2015). CAFs are predominant in the TME, expressing  $\alpha$ -smooth actin and fibroblast activation protein. The accumulation of CAFs in the TME is associated with a poor prognosis in many cancers; CAFs diminish anti-tumor immune responses by recruiting  $T_{reas}$  (Liu et al., 2019; Quante et al., 2011) and higher  $T_{rea}$ levels have been associated with an increased risk of local recurrence in patients with STS (Smolle et al., 2021). Various strategies have been proposed to control  $T_{regs}$  as a therapy for sarcoma. Combining imatinib with CTLA-4 blockade showed synergistic effects in gastrointestinal stromal tumor (a subtype of STS). The intracellular enzyme indoleamine 2,3-dioxygenase (IDO) participates in immune escape pathways as a major regulator of the tryptophan catabolism pathway, with kynurenine being a crucial metabolite. Kynurenine fosters immune tolerance by selectively expanding T<sub>regs</sub> (Holmgaard et al., 2013), IDO1 expression was observed in 39.1% of human sarcoma cases and its blockade led to a significant decrease in both the plasmatic kynurenine-to-tryptophan ratio and tumoral kynurenine level (Nafia et al., 2020). IDO1-deficient mice showed an enhanced response to immunotherapies utilizing ICIs targeting CTLA4, PD-1, and PD-L1 (Holmgaard et al., 2013). These findings suggest IDO1 as a potential target to increase the immunogenicity of sarcomas and improve the efficacy of immunotherapies. Imatinib inhibits IDO and suppresses the number and activity of  $T_{regs}$  (Balachandran et al., 2011). The use of anti-CD25 antibodies in combination with PD-1 blockade resulted in the synergistic elimination of tumors in various cancer models, including a fibrosarcoma model (Arce Vargas et al., 2017). As the CCR4-CCL22 axis assists in the migration of  $T_{regs}$  to tumor sites, CCR4 inhibitors have been considered to inhibit  $T_{reg}$  infiltration. In a clinical study of mogamulizumab, an anti-CCR4 antibody, sarcoma patients exhibited improved prognosis when treated with durvalumab, an anti-PD-1 antibody (Zamarin et al., 2020).

MSCs, which are sources of fibroblasts and pericytes, are also well-known immune modulators that contribute to anti-tumor immunity suppression (Li et al., 2022). MSCs and CAFs have been identified in the TME of osteosarcoma (Zhou et al., 2020). Based on single-cell RNA sequencing and bioinformatic analysis, CAFs exhibit higher infiltration levels in recurrent osteosarcoma, correlating with enrichment of the EMT pathway (Huang et al., 2022). MSCs secrete factors relevant to osteosarcoma growth and immunosuppression, such as angiogenesis factors (vascular endothelial growth factor and platelet-derived growth factor) and tumor growth factor- $\beta$ , which attenuate anti-tumor immune activity (Ohm et al., 2003; Zheng et al., 2018). Stromal cells have also been investigated as therapeutic targets to overcome immunosuppression. Various tumors, including osteosarcoma and undifferentiated pleomorphic sarcoma, displayed high expression of leucine-rich repeat-containing 15 (LRRC15) on CAFs and stromal cells, impeding CD8<sup>+</sup> T cell function (Krishnamurty et al., 2022; Purcell et al., 2018). A first-in-human phase I clinical trial of the antibody-drug conjugate ABBV-085, which targets LRRC15, demonstrated the ability to treat tumors by targeting stromal cells, exhibiting anti-tumor activity in patients

with osteosarcoma and undifferentiated sarcoma (Demetri et al., 2021).

### SUMMARY AND PROSPECTS

Despite advancements in various chemotherapy and radiotherapy options beyond conventional surgical treatment for sarcomas, significant treatment challenges remain. The promise of immunotherapy in various solid cancers has inspired numerous studies investigating its potential in sarcomas, including cancer vaccines, immune checkpoint blockade, and ACT. However, sarcomas are often immunologically cold due to both intrinsic and extrinsic factors. Intrinsically, sarcomas may lack appropriate antigens for immune cell recognition or may induce T cell exhaustion through continuous expression of inhibitory signal. Extrinsic factors such as the TME can contribute to an immunosuppressive environment, compromising immune infiltration or activation. Despite progress in understanding the components of the immunosuppressive TMEs, many immune-resistance mechanisms remain unknown, and sarcoma TMEs can vary widely across subtypes. Therefore, a deeper understanding of TMEs in sarcomas is essential for the development of advanced immunotherapy strategies that can effectively overcome these barriers and provide more personalized and effective treatments.

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### **AUTHOR CONTRIBUTIONS**

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### **CONFLICT OF INTEREST**

The authors have no potential conflicts of interest to disclose.

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## REFERENCES

Abramson, J.S., Palomba, M.L., Gordon, L.I., Lunning, M.A., Wang, M., Arnason, J., Mehta, A., Purev, E., Maloney, D.G., Andreadis, C., et al. (2020). Lisocabtagene maraleucel for patients with relapsed or refractory large B-cell lymphomas (TRANSCEND NHL 001): a multicentre seamless design study. Lancet *396*, 839-852.

Ahmed, N., Brawley, V.S., Hegde, M., Robertson, C., Ghazi, A., Gerken, C., Liu, E., Dakhova, O., Ashoori, A., Corder, A., et al. (2015). Human epidermal growth factor receptor 2 (HER2)-specific chimeric antigen receptormodified T cells for the immunotherapy of HER2-positive sarcoma. J. Clin. Oncol. *33*, 1688-1696.

Ahmed, N., Salsman, V.S., Yvon, E., Louis, C.U., Perlaky, L., Wels, W.S., Dishop, M.K., Kleinerman, E.E., Pule, M., Rooney, C.M., et al. (2009). Immunotherapy for osteosarcoma: genetic modification of T cells overcomes low levels of tumor antigen expression. Mol. Ther. *17*, 1779-1787.

Arce Vargas, F., Furness, A.J.S., Solomon, I., Joshi, K., Mekkaoui, L., Lesko, M.H., Miranda Rota, E., Dahan, R., Georgiou, A., Sledzinska, A., et al. (2017). Fc-optimized anti-CD25 depletes tumor-infiltrating regulatory T cells and synergizes with PD-1 blockade to eradicate established tumors. Immunity *46*, 577-586.

Ayodele, O. and Razak, A.R.A. (2020). Immunotherapy in soft-tissue sarcoma. Curr. Oncol. *27*(Suppl 1), 17-23.

Balachandran, V.P., Cavnar, M.J., Zeng, S., Bamboat, Z.M., Ocuin, L.M., Obaid, H., Sorenson, E.C., Popow, R., Ariyan, C., Rossi, F., et al. (2011). Imatinib potentiates antitumor T cell responses in gastrointestinal stromal tumor through the inhibition of Ido. Nat. Med. *17*, 1094-1100.

Beatty, G.L. and Gladney, W.L. (2015). Immune escape mechanisms as a guide for cancer immunotherapy. Clin. Cancer Res. 21, 687-692.

Blasius, F., Delbruck, H., Hildebrand, F., and Hofmann, U.K. (2022). Surgical treatment of bone sarcoma. Cancers (Basel) *14*, 2694.

Cancer Genome Atlas Research Network (2017). Comprehensive and integrated genomic characterization of adult soft tissue sarcomas. Cell *171*, 950-965.e28.

Charan, M., Dravid, P., Cam, M., Audino, A., Gross, A.C., Arnold, M.A., Roberts, R.D., Cripe, T.P., Pertsemlidis, A., Houghton, P.J., et al. (2020). GD2-directed CAR-T cells in combination with HGF-targeted neutralizing antibody (AMG102) prevent primary tumor growth and metastasis in Ewing sarcoma. Int. J. Cancer *146*, 3184-3195.

Chen, H.H., Zhang, T.N., Zhang, F.Y., and Zhang, T. (2022). Non-coding RNAs in drug and radiation resistance of bone and soft-tissue sarcoma: a systematic review. Elife *11*, e79655.

D'Angelo, S.P., Attia, S., Blay, J.Y., Strauss, S.J., Morales, C.M.V., Razak, A.R.A., Winkle, E.V., Annareddy, T., Sattigari, C., Diamantopoulos, E., et al. (2022). Identification of response stratification factors from pooled efficacy analyses of afamitresgene autoleucel ("Afami-cel" [Formerly ADP-A2M4]) in metastatic synovial sarcoma and myxoid/round cell liposarcoma phase 1 and phase 2 trials. J. Clin. Oncol. 40(16 Suppl), 11562.

D'Angelo, S.P., Mahoney, M.R., Van Tine, B.A., Atkins, J., Milhem, M.M., Jahagirdar, B.N., Antonescu, C.R., Horvath, E., Tap, W.D., Schwartz, G.K., et al. (2018). Nivolumab with or without ipilimumab treatment for metastatic sarcoma (Alliance A091401): two open-label, non-comparative, randomised, phase 2 trials. Lancet Oncol. *19*, 416-426.

D'Angelo, S.P., Shoushtari, A.N., Agaram, N.P., Kuk, D., Qin, L.X., Carvajal, R.D., Dickson, M.A., Gounder, M., Keohan, M.L., Schwartz, G.K., et al. (2015). Prevalence of tumor-infiltrating lymphocytes and PD-L1 expression in the soft tissue sarcoma microenvironment. Hum. Pathol. *46*, 357-365.

de Jong, Y., Ingola, M., Briaire-de Bruijn, I.H., Kruisselbrink, A.B., Venneker,

S., Palubeckaite, I., Heijs, B., Cleton-Jansen, A.M., Haas, R.L.M., and Bovee, J. (2019). Radiotherapy resistance in chondrosarcoma cells; a possible correlation with alterations in cell cycle related genes. Clin. Sarcoma Res. *9*, 9.

Demetri, G.D., Luke, J.J., Hollebecque, A., Powderly, J.D., 2nd, Spira, A.I., Subbiah, V., Naumovski, L., Chen, C., Fang, H., Lai, D.W., et al. (2021). First-in-human phase I study of ABBV-085, an antibody-drug conjugate targeting LRRC15, in sarcomas and other advanced solid tumors. Clin. Cancer Res. *27*, 3556-3566.

Devalaraja, S., To, T.K.J., Folkert, I.W., Natesan, R., Alam, M.Z., Li, M., Tada, Y., Budagyan, K., Dang, M.T., Zhai, L., et al. (2020). Tumor-derived retinoic acid regulates intratumoral monocyte differentiation to promote immune suppression. Cell *180*, 1098-1114.e16.

Dhodapkar, M.V., Sznol, M., Zhao, B., Wang, D., Carvajal, R.D., Keohan, M.L., Chuang, E., Sanborn, R.E., Lutzky, J., Powderly, J., et al. (2014). Induction of antigen-specific immunity with a vaccine targeting NY-ESO-1 to the dendritic cell receptor DEC-205. Sci. Transl. Med. *6*, 232ra51.

Doig, K.D., Fellowes, A., Scott, P., and Fox, S.B. (2022). Tumour mutational burden: an overview for pathologists. Pathology *54*, 249-253.

Du, X.H., Wei, H., Zhang, P., Yao, W.T., and Cai, Q.Q. (2020). Heterogeneity of soft tissue sarcomas and its implications in targeted therapy. Front. Oncol. *10*, 564852.

Edwards, S.C., Hoevenaar, W.H.M., and Coffelt, S.B. (2021). Emerging immunotherapies for metastasis. Br. J. Cancer *124*, 37-48.

Epping, M.T., Wang, L., Edel, M.J., Carlee, L., Hernandez, M., and Bernards, R. (2005). The human tumor antigen PRAME is a dominant repressor of retinoic acid receptor signaling. Cell *122*, 835-847.

Finkelstein, S.E., Fishman, M., Conley, A.P., Gabrilovich, D., Antonia, S., and Chiappori, A. (2012). Cellular immunotherapy for soft tissue sarcomas. Immunotherapy *4*, 283-290.

Gelderblom, H., Hogendoorn, P.C., Dijkstra, S.D., van Rijswijk, C.S., Krol, A.D., Taminiau, A.H., and Bovee, J.V. (2008). The clinical approach towards chondrosarcoma. Oncologist *13*, 320-329.

Goodman, A.M., Kato, S., Bazhenova, L., Patel, S.P., Frampton, G.M., Miller, V., Stephens, P.J., Daniels, G.A., and Kurzrock, R. (2017). Tumor mutational burden as an independent predictor of response to immunotherapy in diverse cancers. Mol. Cancer Ther. *16*, 2598-2608.

Goorin, A.M., Delorey, M.J., Lack, E.E., Gelber, R.D., Price, K., Cassady, J.R., Levey, R., Tapper, D., Jaffe, N., and Link, M. (1984). Prognostic significance of complete surgical resection of pulmonary metastases in patients with osteogenic sarcoma: analysis of 32 patients. J. Clin. Oncol. *2*, 425-431.

Gronchi, A. (2021). Surgery in soft tissue sarcoma: the thin line between a surgical or more conservative approach. Future Oncol. *17*(21s), 3-6.

Grunewald, T.G., Alonso, M., Avnet, S., Banito, A., Burdach, S., Cidre-Aranaz, F., Di Pompo, G., Distel, M., Dorado-Garcia, H., Garcia-Castro, J., et al. (2020). Sarcoma treatment in the era of molecular medicine. EMBO Mol. Med. *12*, e11131.

Gubin, M.M., Zhang, X., Schuster, H., Caron, E., Ward, J.P., Noguchi, T., Ivanova, Y., Hundal, J., Arthur, C.D., Krebber, WJ., et al. (2014). Checkpoint blockade cancer immunotherapy targets tumour-specific mutant antigens. Nature *515*, 577-581.

Hashimoto, K., Nishimura, S., Ito, T., Oka, N., Kakinoki, R., and Akagi, M. (2022). Clinicopathological assessment of cancer/testis antigens NY-ESO-1 and MAGE-A4 in osteosarcoma. Eur. J. Histochem. *66*, 3377.

Hemminger, J.A., Toland, A.E., Scharschmidt, T.J., Mayerson, J.L., Guttridge, D.C., and Iwenofu, O.H. (2014). Expression of cancer-testis antigens MAGEA1, MAGEA3, ACRBP, PRAME, SSX2, and CTAG2 in myxoid and round cell liposarcoma. Mod. Pathol. *27*, 1238-1245.

Highfill, S.L., Cui, Y., Giles, A.J., Smith, J.P., Zhang, H., Morse, E., Kaplan, R.N., and Mackall, C.L. (2014). Disruption of CXCR2-mediated MDSC tumor trafficking enhances anti-PD1 efficacy. Sci. Transl. Med. *6*, 237ra67. Hoefkens, F., Dehandschutter, C., Somville, J., Meijnders, P., and Van Gestel, D. (2016). Soft tissue sarcoma of the extremities: pending questions on surgery and radiotherapy. Radiat. Oncol. *11*, 136.

Holmgaard, R.B., Zamarin, D., Munn, D.H., Wolchok, J.D., and Allison, J.P. (2013). Indoleamine 2,3-dioxygenase is a critical resistance mechanism in antitumor T cell immunotherapy targeting CTLA-4. J. Exp. Med. *210*, 1389-1402.

Huang, X., Wang, L., Guo, H., Zhang, W., and Shao, Z. (2022). Singlecell transcriptomics reveals the regulative roles of cancer associated fibroblasts in tumor immune microenvironment of recurrent osteosarcoma. Theranostics *12*, 5877-5887.

Hui, J.Y. (2016). Epidemiology and etiology of sarcomas. Surg. Clin. North Am. *96*, 901-914.

Husain, M., Quiroga, D., Kim, H.G., Lenobel, S., Xu, M., Iwenofu, H., Chen, J.L., Verschraegen, C., Liebner, D., and Tinoco, G. (2023). Clinical markers of immunotherapy outcomes in advanced sarcoma. BMC Cancer *23*, 326.

Ingley, K.M., Maleddu, A., Grange, F.L., Gerrand, C., Bleyer, A., Yasmin, E., Whelan, J., and Strauss, S.J. (2022). Current approaches to management of bone sarcoma in adolescent and young adult patients. Pediatr. Blood Cancer *69*, e29442.

lura, K., Kohashi, K., Ishii, T., Maekawa, A., Bekki, H., Otsuka, H., Yamada, Y., Yamamoto, H., Matsumoto, Y., Iwamoto, Y., et al. (2017). MAGEA4 expression in bone and soft tissue tumors: its utility as a target for immunotherapy and diagnostic marker combined with NY-ESO-1. Virchows Arch. *471*, 383-392.

June, C.H., O'Connor, R.S., Kawalekar, O.U., Ghassemi, S., and Milone, M.C. (2018). CAR T cell immunotherapy for human cancer. Science *359*, 1361-1365.

Jungbluth, A.A., Antonescu, C.R., Busam, K.J., Iversen, K., Kolb, D., Coplan, K., Chen, Y.T., Stockert, E., Ladanyi, M., and Old, L.J. (2001). Monophasic and biphasic synovial sarcomas abundantly express cancer/testis antigen NY-ESO-1 but not MAGE-A1 or CT7. Int. J. Cancer *94*, 252-256.

Juretic, A., Spagnoli, G.C., Schultz-Thater, E., and Sarcevic, B. (2003). Cancer/testis tumour-associated antigens: immunohistochemical detection with monoclonal antibodies. Lancet Oncol. *4*, 104-109.

Kailayangiri, S., Altvater, B., Meltzer, J., Pscherer, S., Luecke, A., Dierkes, C., Titze, U., Leuchte, K., Landmeier, S., Hotfilder, M., et al. (2012). The ganglioside antigen G(D2) is surface-expressed in Ewing sarcoma and allows for MHC-independent immune targeting. Br. J. Cancer *106*, 1123-1133.

Kakimoto, T., Matsumine, A., Kageyama, S., Asanuma, K., Matsubara, T., Nakamura, T., Iino, T., Ikeda, H., Shiku, H., and Sudo, A. (2019). Immunohistochemical expression and clinicopathological assessment of the cancer testis antigens NY-ESO-1 and MAGE-A4 in high-grade soft-tissue sarcoma. Oncol. Lett. *17*, 3937-3943.

Kawaguchi, S., Tsukahara, T., Ida, K., Kimura, S., Murase, M., Kano, M., Emori, M., Nagoya, S., Kaya, M., Torigoe, T., et al. (2012). SYT-SSX breakpoint peptide vaccines in patients with synovial sarcoma: a study from the Japanese Musculoskeletal Oncology Group. Cancer Sci. *103*, 1625-1630.

Kelly, C.M., Antonescu, C.R., Bowler, T., Munhoz, R., Chi, P., Dickson, M.A., Gounder, M.M., Keohan, M.L., Movva, S., Dholakia, R., et al. (2020). Objective response rate among patients with locally advanced or metastatic sarcoma treated with talimogene laherparepvec in combination with pembrolizumab: a phase 2 clinical trial. JAMA Oncol. *6*, 402-408.

Kim, C., Kim, E.K., Jung, H., Chon, H.J., Han, J.W., Shin, K.H., Hu, H., Kim, K.S., Choi, Y.D., Kim, S., et al. (2016). Prognostic implications of PD-L1 expression in patients with soft tissue sarcoma. BMC Cancer *16*, 434.

Kim, H., Cho, Y., Kim, H.S., Kang, D., Cheon, D., Kim, Y.J., Chang, M.J., Lee, K.M., Chang, C.B., Kang, S.B., et al. (2020). A system-level approach identifies HIF-2alpha as a critical regulator of chondrosarcoma progression. Nat. Commun. *11*, 5023.

Klemen, N.D., Kelly, C.M., and Bartlett, E.K. (2021). The emerging role of immunotherapy for the treatment of sarcoma. J. Surg. Oncol. *123*, 730-738.

Krishnamurty, A.T., Shyer, J.A., Thai, M., Gandham, V., Buechler, M.B., Yang, Y.A., Pradhan, R.N., Wang, A.W., Sanchez, P.L., Qu, Y., et al. (2022). LRRC15(+) myofibroblasts dictate the stromal setpoint to suppress tumour immunity. Nature *611*, 148-154.

Lagos, G.G., Izar, B., and Rizvi, N.A. (2020). Beyond tumor PD-L1: emerging genomic biomarkers for checkpoint inhibitor immunotherapy. Am. Soc. Clin. Oncol. Educ. Book *40*, 1-11.

Lee, E.Y., Kim, M., Choi, B.K., Kim, D.H., Choi, I., and You, H.J. (2021a). TJP1 contributes to tumor progression through supporting cell-cell aggregation and communicating with tumor microenvironment in leiomyosarcoma. Mol. Cells *44*, 784-794.

Lee, Y.J., Lee, J.B., Ha, S.J., and Kim, H.R. (2021b). Clinical perspectives to overcome acquired resistance to anti-programmed death-1 and anti-programmed death ligand-1 therapy in non-small cell lung cancer. Mol. Cells 44, 363-373.

Li, X., Fan, Q., Peng, X., Yang, S., Wei, S., Liu, J., Yang, L., and Li, H. (2022). Mesenchymal/stromal stem cells: necessary factors in tumour progression. Cell Death Discov. *8*, 333.

Li, Y., Geng, P., Jiang, W., Wang, Y., Yao, J., Lin, X., Liu, J., Huang, L., Su, B., and Chen, H. (2014). Enhancement of radiosensitivity by 5-Aza-CdR through activation of G2/M checkpoint response and apoptosis in osteosarcoma cells. Tumour Biol. *35*, 4831-4839.

Liu, T., Han, C., Wang, S., Fang, P., Ma, Z., Xu, L., and Yin, R. (2019). Cancerassociated fibroblasts: an emerging target of anti-cancer immunotherapy. J. Hematol. Oncol. *12*, 86.

Lodhia, J., Goodluck, G., Tendai, J., Urassa, E., Nkya, G., and Mremi, A. (2022). Case series of high-grade soft tissue sarcoma of the lower limb with delayed diagnosis: experience at a tertiary hospital in northern Tanzania. Int. J. Surg. Case Rep. *97*, 107475.

Marabelle, A., Fakih, M., Lopez, J., Shah, M., Shapira-Frommer, R., Nakagawa, K., Chung, H.C., Kindler, H.L., Lopez-Martin, J.A., Miller, W.H., Jr., et al. (2020). Association of tumour mutational burden with outcomes in patients with advanced solid tumours treated with pembrolizumab: prospective biomarker analysis of the multicohort, open-label, phase 2 KEYNOTE-158 study. Lancet Oncol. *21*, 1353-1365.

Molgora, M., Esaulova, E., Vermi, W., Hou, J., Chen, Y., Luo, J., Brioschi, S., Bugatti, M., Omodei, A.S., Ricci, B., et al. (2020). TREM2 modulation remodels the tumor myeloid landscape enhancing anti-PD-1 immunotherapy. Cell *182*, 886-900.e17.

Monga, V., Mani, H., Hirbe, A., and Milhem, M. (2020). Non-conventional treatments for conventional chondrosarcoma. Cancers (Basel) *12*, 1962.

Nafia, I., Toulmonde, M., Bortolotto, D., Chaibi, A., Bodet, D., Rey, C., Velasco, V., Larmonier, C.B., Cerf, L., Adam, J., et al. (2020). IDO targeting in sarcoma: biological and clinical implications. Front. Immunol. *11*, 274.

Nakamura, K. and Smyth, M.J. (2020). Myeloid immunosuppression and immune checkpoints in the tumor microenvironment. Cell. Mol. Immunol. *17*, 1-12.

Navai, S.A., Derenzo, C., Joseph, S., Sanber, K., Byrd, T., Zhang, H., Mata, M., Gerken, C., Shree, A., Mathew, P.R., et al. (2019). Abstract LB-147: Administration of HER2-CAR T cells after lymphodepletion safely improves T cell expansion and induces clinical responses in patients with advanced sarcomas. Cancer Res. *79*(13 Suppl), LB-147.

Nishikawa, H., Sato, E., Briones, G., Chen, L.M., Matsuo, M., Nagata, Y., Ritter, G., Jager, E., Nomura, H., Kondo, S., et al. (2006). In vivo antigen delivery by a Salmonella typhimurium type III secretion system for therapeutic cancer vaccines. J. Clin. Invest. *116*, 1946-1954.

O'Donnell, J.S., Teng, M.W.L., and Smyth, M.J. (2019). Cancer immunoediting and resistance to T cell-based immunotherapy. Nat. Rev. Clin. Oncol. *16*, 151-167.

Ohm, J.E., Gabrilovich, D.I., Sempowski, G.D., Kisseleva, E., Parman, K.S., Nadaf, S., and Carbone, D.P. (2003). VEGF inhibits T-cell development and may contribute to tumor-induced immune suppression. Blood *101*, 4878-4886.

Olivier, T., Pop, D., Chouiter Djebaili, A., Falk, A.T., Iannessi, A., Saada, E., Nettekoven, W., Blay, J.Y., Baque, P., Cupissol, D., et al. (2015). Treating metastatic sarcomas locally: a paradoxe, a rationale, an evidence? Crit. Rev. Oncol. Hematol. *95*, 62-77.

Panagi, M., Pilavaki, P., Constantinidou, A., and Stylianopoulos, T. (2022). Immunotherapy in soft tissue and bone sarcoma: unraveling the barriers to effectiveness. Theranostics *12*, 6106-6129.

Patel, S.A., Royce, T.J., Barysauskas, C.M., Thornton, K.A., Raut, C.P., and Baldini, E.H. (2017). Surveillance imaging patterns and outcomes following radiation therapy and radical resection for localized extremity and trunk soft tissue sarcoma. Ann. Surg. Oncol. 24, 1588-1595.

Pawlik, T.M., Pisters, P.W., Mikula, L., Feig, B.W., Hunt, K.K., Cormier, J.N., Ballo, M.T., Catton, C.N., Jones, J.J., O'Sullivan, B., et al. (2006). Long-term results of two prospective trials of preoperative external beam radiotherapy for localized intermediate- or high-grade retroperitoneal soft tissue sarcoma. Ann. Surg. Oncol. *13*, 508-517.

Paydas, S., Bagir, E.K., Deveci, M.A., and Gonlusen, G. (2016). Clinical and prognostic significance of PD-1 and PD-L1 expression in sarcomas. Med. Oncol. *33*, 93.

Petitprez, F., de Reynies, A., Keung, E.Z., Chen, T.W., Sun, C.M., Calderaro, J., Jeng, Y.M., Hsiao, L.P., Lacroix, L., Bougouin, A., et al. (2020). B cells are associated with survival and immunotherapy response in sarcoma. Nature *577*, 556-560.

Purcell, J.W., Tanlimco, S.G., Hickson, J., Fox, M., Sho, M., Durkin, L., Uziel, T., Powers, R., Foster, K., McGonigal, T., et al. (2018). LRRC15 is a novel mesenchymal protein and stromal target for antibody-drug conjugates. Cancer Res. *78*, 4059-4072.

Qian, Y., Qiao, S., Dai, Y., Xu, G., Dai, B., Lu, L., Yu, X., Luo, Q., and Zhang, Z. (2017). Molecular-targeted immunotherapeutic strategy for melanoma via dual-targeting nanoparticles delivering small interfering RNA to tumor-associated macrophages. ACS Nano *11*, 9536-9549.

Quante, M., Tu, S.P., Tomita, H., Gonda, T., Wang, S.S., Takashi, S., Baik, G.H., Shibata, W., Diprete, B., Betz, K.S., et al. (2011). Bone marrow-derived myofibroblasts contribute to the mesenchymal stem cell niche and promote tumor growth. Cancer Cell *19*, 257-272.

Riedel, R.F., Larrier, N., Dodd, L., Kirsch, D., Martinez, S., and Brigman, B.E. (2009). The clinical management of chondrosarcoma. Curr. Treat. Options Oncol. *10*, 94-106.

Rodell, C.B., Arlauckas, S.P., Cuccarese, M.F., Garris, C.S., Li, R., Ahmed, M.S., Kohler, R.H., Pittet, M.J., and Weissleder, R. (2018). TLR7/8-agonistloaded nanoparticles promote the polarization of tumour-associated macrophages to enhance cancer immunotherapy. Nat. Biomed. Eng. *2*, 578-588.

Roth, M., Linkowski, M., Tarim, J., Piperdi, S., Sowers, R., Geller, D., Gill, J., and Gorlick, R. (2014). Ganglioside GD2 as a therapeutic target for antibody-mediated therapy in patients with osteosarcoma. Cancer *120*, 548-554.

Rytlewski, J., Brockman, Q.R., Dodd, R.D., Milhem, M., and Monga, V. (2022). Epigenetic modulation in sensitizing metastatic sarcomas to therapies and overcoming resistance. Cancer Drug Resist. *5*, 25-35.

Rytlewski, J., Milhem, M.M., and Monga, V. (2021). Turning 'Cold' tumors 'Hot': immunotherapies in sarcoma. Ann. Transl. Med. *9*, 1039.

Samiei, A., Gjertson, D.W., Memarzadeh, S., Konecny, G.E., and Moatamed, N.A. (2022). Expression of immune checkpoint regulators, programmed death-ligand 1 (PD-L1/PD-1), cytotoxic T lymphocyte antigen 4 (CTLA-4), and indolaimine-2, 3-deoxygenase (IDO) in uterine mesenchymal tumors. Diagn. Pathol. *17*, 70.

Sannino, G., Marchetto, A., Kirchner, T., and Grunewald, T.G.P. (2017).

Epithelial-to-mesenchymal and mesenchymal-to-epithelial transition in mesenchymal tumors: a paradox in sarcomas? Cancer Res. 77, 4556-4561.

Sato, Y., Nabeta, Y., Tsukahara, T., Hirohashi, Y., Syunsui, R., Maeda, A., Sahara, H., Ikeda, H., Torigoe, T., Ichimiya, S., et al. (2002). Detection and induction of CTLs specific for SYT-SSX-derived peptides in HLA-A24(+) patients with synovial sarcoma. J. Immunol. *169*, 1611-1618.

Saxena, M., van der Burg, S.H., Melief, C.J.M., and Bhardwaj, N. (2021). Therapeutic cancer vaccines. Nat. Rev. Cancer *21*, 360-378.

Siegel, R.L., Miller, K.D., Fuchs, H.E., and Jemal, A. (2022). Cancer statistics, 2022. CA Cancer J. Clin. 72, 7-33.

Smolle, M.A., Herbsthofer, L., Granegger, B., Goda, M., Brcic, I., Bergovec, M., Scheipl, S., Prietl, B., Pichler, M., Gerger, A., et al. (2021). T-regulatory cells predict clinical outcome in soft tissue sarcoma patients: a clinico-pathological study. Br. J. Cancer *125*, 717-724.

Somaiah, N., Conley, A.P., Parra, E.R., Lin, H., Amini, B., Solis Soto, L., Salazar, R., Barreto, C., Chen, H., Gite, S., et al. (2022). Durvalumab plus tremelimumab in advanced or metastatic soft tissue and bone sarcomas: a single-centre phase 2 trial. Lancet Oncol. *23*, 1156-1166.

Squires, M.H., Ethun, C.G., Suarez-Kelly, L.P., Yu, P.Y., Hughes, T.M., Shelby, R.D., Tran, T.B., Poultsides, G., Charlson, J., Gamblin, T.C., et al. (2020). Trends in the use of adjuvant chemotherapy for high-grade truncal and extremity soft tissue sarcomas. J. Surg. Res. *245*, 577-586.

Strickler, J.H., Hanks, B.A., and Khasraw, M. (2021). Tumor mutational burden as a predictor of immunotherapy response: is more always better? Clin. Cancer Res. *27*, 1236-1241.

Tagliamonte, M., Petrizzo, A., Tornesello, M.L., Buonaguro, F.M., and Buonaguro, L. (2014). Antigen-specific vaccines for cancer treatment. Hum. Vaccin. Immunother. *10*, 3332-3346.

Tan, S., Li, D., and Zhu, X. (2020). Cancer immunotherapy: pros, cons and beyond. Biomed. Pharmacother. *124*, 109821.

Tang, F., Tie, Y., Wei, Y.Q., Tu, C.Q., and Wei, X.W. (2021). Targeted and immuno-based therapies in sarcoma: mechanisms and advances in clinical trials. Biochim. Biophys. Acta Rev. Cancer *1876*, 188606.

Tawbi, H.A., Burgess, M., Bolejack, V., Van Tine, B.A., Schuetze, S.M., Hu, J., D'Angelo, S., Attia, S., Riedel, R.F., Priebat, D.A., et al. (2017). Pembrolizumab in advanced soft-tissue sarcoma and bone sarcoma (SARC028): a multicentre, two-cohort, single-arm, open-label, phase 2 trial. Lancet Oncol. *18*, 1493-1501.

Thorkildsen, J., Norum, O.J., Myklebust, T.A., and Zaikova, O. (2021). Chondrosarcoma local recurrence in the Cancer Registry of Norway cohort (1990-2013): patterns and impact. J. Surg. Oncol. *123*, 510-520.

Tsagozis, P., Augsten, M., Zhang, Y., Li, T., Hesla, A., Bergh, J., Haglund, F., Tobin, N.P., and Ehnman, M. (2019). An immunosuppressive macrophage profile attenuates the prognostic impact of CD20-positive B cells in human soft tissue sarcoma. Cancer Immunol. Immunother. *68*, 927-936.

Tu, B., Zhu, J., Liu, S., Wang, L., Fan, Q., Hao, Y., Fan, C., and Tang, T.T. (2016). Mesenchymal stem cells promote osteosarcoma cell survival and drug resistance through activation of STAT3. Oncotarget *7*, 48296-48308.

Turley, S.J., Cremasco, V., and Astarita, J.L. (2015). Immunological hallmarks of stromal cells in the tumour microenvironment. Nat. Rev. Immunol. *15*, 669-682.

von Konow, A., Ghanei, I., Styring, E., and Vult von Steyern, F. (2021). Late local recurrence and metastasis in soft tissue sarcoma of the extremities and trunk wall: better outcome after treatment of late events compared with early. Ann. Surg. Oncol. *28*, 7891-7902.

Waldman, A.D., Fritz, J.M., and Lenardo, M.J. (2020). A guide to cancer immunotherapy: from T cell basic science to clinical practice. Nat. Rev. Immunol. *20*, 651-668.

Weber, J. (2007). Review: anti-CTLA-4 antibody ipilimumab: case studies of clinical response and immune-related adverse events. Oncologist *12*, 864-872.

Wei, R., Dean, D.C., Thanindratarn, P., Hornicek, F.J., Guo, W., and Duan, Z. (2020). Cancer testis antigens in sarcoma: expression, function and immunotherapeutic application. Cancer Lett. *479*, 54-60.

Weng, W., Yu, L., Li, Z., Tan, C., Lv, J., Lao, I.W., Hu, W., Deng, Z., Liu, Z., Wang, J., et al. (2022). The immune subtypes and landscape of sarcomas. BMC Immunol. *23*, 46.

Whitehurst, A.W. (2014). Cause and consequence of cancer/testis antigen activation in cancer. Annu. Rev. Pharmacol. Toxicol. 54, 251-272.

Xie, C., Whalley, N., Adasonla, K., Grimer, R., and Jeys, L. (2015). Can local recurrence of a sacral chordoma be treated by further surgery? Bone Joint J. 97-B, 711-715.

Xie, L., Yang, Y., Guo, W., Che, D., Xu, J., Sun, X., Liu, K., Ren, T., Liu, X., Yang, Y., et al. (2021). The clinical implications of tumor mutational burden in osteosarcoma. Front. Oncol. *10*, 595527.

Yan, M., Schwaederle, M., Arguello, D., Millis, S.Z., Gatalica, Z., and Kurzrock, R. (2015). HER2 expression status in diverse cancers: review of results from 37,992 patients. Cancer Metastasis Rev. *34*, 157-164.

Yang, W., Lee, K.W., Srivastava, R.M., Kuo, F., Krishna, C., Chowell, D., Makarov, V., Hoen, D., Dalin, M.G., Wexler, L., et al. (2019). Immunogenic neoantigens derived from gene fusions stimulate T cell responses. Nat. Med. *25*, 767-775.

Zamarin, D., Hamid, O., Nayak-Kapoor, A., Sahebjam, S., Sznol, M., Collaku, A., Fox, F.E., Marshall, M.A., and Hong, D.S. (2020). Mogamulizumab in combination with durvalumab or tremelimumab in patients with advanced solid tumors: a phase I study. Clin. Cancer Res. *26*, 4531-4541.

Zhang, H., Huang, W., Feng, Q., Sun, W., Yan, W., Wang, C., Zhang, J., Huang, K., Yu, L., Qu, X., et al. (2022). Clinical significance and risk factors of local recurrence in synovial sarcoma: a retrospective analysis of 171 cases. Front. Surg. *8*, 736146.

Zhang, Y. and Zhang, Z. (2020). The history and advances in cancer immunotherapy: understanding the characteristics of tumor-infiltrating immune cells and their therapeutic implications. Cell. Mol. Immunol. *17*, 807-821.

Zhao, L. and Cao, Y.J. (2019). Engineered T cell therapy for cancer in the clinic. Front. Immunol. *10*, 2250.

Zhao, R., Yu, X., Feng, Y., Yang, Z., Chen, X., Wand, J., Ma, S., Zhang, Z., and Guo, X. (2018). Local recurrence is correlated with decreased overall survival in patients with intermediate high-grade localized primary soft tissue sarcoma of extremity and abdominothoracic wall. Asia Pac. J. Clin. Oncol. *14*, e109-e115.

Zheng, W., Xiao, H., Liu, H., and Zhou, Y. (2015). Expression of programmed death 1 is correlated with progression of osteosarcoma. APMIS *123*, 102-107.

Zheng, Y., Wang, G., Chen, R., Hua, Y., and Cai, Z. (2018). Mesenchymal stem cells in the osteosarcoma microenvironment: their biological properties, influence on tumor growth, and therapeutic implications. Stem Cell Res. Ther. *9*, 22.

Zhou, Y., Yang, D., Yang, Q., Lv, X., Huang, W., Zhou, Z., Wang, Y., Zhang, Z., Yuan, T., Ding, X., et al. (2020). Single-cell RNA landscape of intratumoral heterogeneity and immunosuppressive microenvironment in advanced osteosarcoma. Nat. Commun. *11*, 6322.

Zhu, M.M.T., Shenasa, E., and Nielsen, T.O. (2020). Sarcomas: immune biomarker expression and checkpoint inhibitor trials. Cancer Treat. Rev. *91*, 102115.

Zitvogel, L., Tesniere, A., and Kroemer, G. (2006). Cancer despite immunosurveillance: immunoselection and immunosubversion. Nat. Rev. Immunol. *6*, 715-727.

Zou, C., Shen, J., Tang, Q., Yang, Z., Yin, J., Li, Z., Xie, X., Huang, G., Lev, D., and Wang, J. (2012). Cancer-testis antigens expressed in osteosarcoma identified by gene microarray correlate with a poor patient prognosis. Cancer *118*, 1845-1855.