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The emerging Janus face of SVEP1 in development and disease

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Abstract

Sushi, von Willebrand factor type A, epidermal growth factor, and pentraxin domain containing 1 (SVEP1) is a large extracellular matrix protein that is also detected in the circulation. Recent plasma proteomics and genomics studies have revealed a large number of associations between SVEP1 and human traits, particularly chronic disease. These include associations with cardiac death and disease, diabetes, platelet traits, glaucoma, dementia, and aging; many of which are causal. Animal models demonstrate that SVEP1 is critical in vascular development and disease, but its molecular and cellular mechanisms remain poorly defined. Future studies should aim to characterize these mechanisms and determine the diagnostic, prognostic, and therapeutic value of measuring or intervening on this enigmatic protein.

A brief history of SVEP1

SVEP1 is a poorly understood **extracellular matrix (ECM)** (see Glossary) protein [1,2]. The sudden rise of population plasma proteomics has revealed a profound number

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JSE and I-HJ reviewed the primary literature. JSE and NOS wrote the manuscript. ACR illustrated the figures and advised on aging themes. PCL generated the table and evaluated the data from the human studies. AA advised on structure and function. NOS acquired funds. All authors reviewed, provided critical editing, and approved the manuscript.

Declaration of interests

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of associations between SVEP1 and human chronic disease. For example, among all proteins tested within each respective study, plasma SVEP1 concentration shows the highest acceleration during aging [3], among the strongest associations with heart failure hospitalization or death [4], and the strongest causal relationship with dementia [5]. In addition, **Mendelian randomization (MR)** analyses (Box 1) suggest SVEP1 causally relates to coronary artery disease (CAD) [6,7], hypertension [6], type 2 diabetes [6,8], platelet traits [7], and longevity [9]. Human genetic or proteomic studies identified additional SVEP1 associations, including glaucoma [10,11], pulmonary arterial hypertension [12], cardiovascular risk [13], atrial fibrillation [14], platelet reactivity [15], and aging [16,17]. Many of these diseases share certain attributes, including vascular dysfunction, chronicity, and aging, suggesting a common pathogenic mechanism may underly their association with SVEP1.

SVEP1 was initially cloned and analyzed in 2000 by Gilgès et al., who named it Polydom for its many domains [1]. It is now referred to as SVEP1, a name derived from its constituent domains. The protein is predominately synthesized by cells of mesenchymal origin and is believed to be integrated within the matrix [2,6,7]. It also leaks from its tissues of origin [18] and circulates in plasma [19,20], likely in association with extracellular vesicles [21–23]. SVEP1 is thought to interact with several proteins, including **integrins** [2,24,25], angiopoietins (ANGs) [26], and **platelet endothelial aggregation receptor 1 (PEAR1)**, a receptor tyrosine kinase-like protein [7,27].

Animal models demonstrate that SVEP1 is critical for vascular development [11,26,28,29] and mouse models support its role in vascular disease [6,24,30], consistent with its disease associations in humans. SVEP1 resides within the ECM and plasma, making it a priority candidate for disease diagnostics, prognostics, and therapeutics. Here, we provide an overview of the existing data on SVEP1, focusing on its structure, function, and role in development and disease. We also highlight key unanswered questions that, once addressed, will enhance our understanding of the protein and its impact on human biology.

Molecular composition of SVEP1

SVEP1 was first cloned using degenerative reverse transcriptase polymerase chain reaction for sequences that code for epidermal growth factor (EGF) domains [1] (Figure 1). Such domains often govern interaction with adjacent proteins and are found in many ECM proteins and transmembrane receptors [31]. SVEP1 contains a single von Willebrand factor type A (VWA) domain, a domain named after von Willebrand factor [32] and found in many ECM proteins. Many VWA domains contain metal ion-dependent adhesions sites [33] and mediate protein complex formation [32,34] or receptor–ligand interaction. Approximately half of SVEP1 is comprised of repeat complement control protein (CCP) domains, also known as Sushi domains. These domains often govern protein interactions, including with the complement family [35]. Hyalin repeat (HYR) domains are frequently found in association with CCPs [36] and are also contained within SVEP1 [37]. HYR domains are found within hyalin and are thought to govern cell adhesion and protein-protein interaction [36]. SVEP1 also contains a pentraxin domain, which is shared with the pentraxin family of pentameric proteins, including C-reactive protein and serum amyloid P component [38].

Glycoproteomic studies [20,39] identify SVEP1 as a likely glycoprotein that contains a number of potential glycosylation sites [40]. Recombinant murine SVEP1 is efficiently secreted from cells and is cleaved in the process [2]. This generates fragments of approximately 125 and 300 kDa under reducing and denaturing conditions. We have confirmed this cleavage event and its approximate end products in a similar expression system [6]. SVEP1 may exist as a multimer, like many other ECM proteins, though this has not been formally tested.

Tissue expression and localization of SVEP1

SVEP1 is expressed in vasculature [6,7,26,28], adipose [7,30,41], and bone marrow $[1,42,43]$ tissues. The vascular human placenta expresses extremely high levels of $SVEPI$ [44] and maternal plasma levels of the protein increase ≥20-fold over the course of gestation [45]. Lung, stomach, and intestinal tissue also express the transcript [2]. The cells that express SVEP1 include bone-marrow-derived mesenchymal stem cells [46], preosteoblastic cells [47], activated skeletal muscle satellite cells [48], and vascular cells including adventitial fibroblasts [49] and vascular smooth muscle cells [6,24].

Precisely where SVEP1 exists within the ECM remains unclear. Previous studies reporting in situ SVEP1 staining relied on unvalidated antibodies and did not include necessary negative controls, such as genetic knockouts [50]. This is of particular importance when using antibodies against SVEP1 since it shares many domains with other ECM proteins, resulting in a high potential for undiscernible crossreactivity. Poor antibody validation has led to significant confusion about where SVEP1 is localized within tissue, though SVEP1 pulldown assays suggest it may integrate with the basement membrane after secretion [7]. The generation and rigorous validation of anti-SVEP1 antibodies, or similar reagents, is necessary to characterize the location of SVEP1 during development and pathology.

SVEP1 binding partners

Several SVEP1 binding partners have been proposed, but it is unclear which of these, if any, are responsible for mediating the developmental and disease phenotypes associated with SVEP1. The first protein reported to bind with SVEP1 was integrin α9β1 [2]. Integrin α9 heterodimerizes only with integrin β1 to form integrin α9β1 [51], which binds with an estimated affinity of 32.4 nM to residues 2636–2644, which are conserved in humans and contained within a CCP domain in the C-terminus of murine SVEP1 [2]. Integrin α9β1 is expressed on a wide variety of cell types, particularly muscle and epithelia [52] and is known for its role in various developmental processes, including **lymphangiogenesis** [53] and hematopoiesis [54] through the regulation of cell growth, migration, differentiation, and other behaviors.

PEAR1, a receptor tyrosine kinase-like protein [27], also interacts with SVEP1 [7]. Plasma SVEP1 levels are influenced by PEAR1 levels in humans [7], suggesting endothelial PEAR1 binds and sequesters SVEP1 from the plasma. Indeed, the recombinant proteins bind to each other with an affinity of 0.7–8.8 nM in **label-free binding assays**. PEAR1 tissue expression is highly correlated with $SVEPI$ [7] and the two genes share numerous disease

associations, including platelet reactivity [15,55] and cardiovascular disease [56,57]. PEAR1 is also thought to play a role in neoangiogenesis [58] and hematopoiesis [59].

SVEP1 binds to tyrosine-protein kinase receptor Tie-1 (TIE) [60,61], and potentially ANG1 and ANG2 [26]. These data suggest that SVEP1 modulates **ANG–TIE** signaling. Such regulation of growth factors is a common function of ECM proteins [62]. The ANG–TIE signaling pathway is involved in many of the same developmental and disease processes as SVEP1, including angiogenesis [63], atherosclerosis [64], and glaucoma [11].

Additional binding partners to SVEP1 likely exist. Several partners have been proposed, but their characterization remains limited. These include integrin α4 [24,65] and members of the complement [66] and Notch family [6]. Further investigation is necessary to validate these interactions and determine how they contribute to the biological functions of SVEP1. Given its large size and interaction with multiple signaling proteins, SVEP1 may serve as a signaling nidus and coordinate its effects through multiple interactions.

Cellular response to SVEP1

Various cell types adhere to SVEP1 in a dose-dependent manner [2,6,7,67], consistent with the functions of the domains that comprise the protein. SVEP1-induced cell adhesion depends on integrin α9β1 in rhabdomyosarcoma cells [2], and integrin signaling is activated in various cells exposed to SVEP1 [6]. Certain cells also proliferate rapidly [6] and migrate [61] in response to SVEP1. SVEP1-induced cell proliferation has been shown to depend on integrin α9β1 and Notch signaling [6]; however, the specificity of these effects is unclear. Consistent with its effects on proliferation, exposure of endothelial cells and human coronary artery smooth muscle cells to SVEP1 results in robust **AKT**/**mTOR** activation [7,61]. This signaling is dependent on PEAR1 [7] and is of particular interest, since AKT/ mTOR signaling plays a key role in the development and disease processes associated with SVEP1 [68–76].

Descriptions of several additional cell responses to SVEP1 have been published, but reproduction of these effects is lacking. Many of these studies utilized unpurified recombinant SVEP1 in solution, which is not a physiological approach to interrogating an ECM protein that is immobilized in vivo. In fact, circulating SVEP1 may also be functionally immobilized given its association with extracellular vesicles [21–23]. The generation of force between a ligand in the ECM and its receptor is often critical for signal transduction; soluble ligands are free in solution and may not be able to transmit sufficient force for receptor activation [77]. We believe this biology suggests SVEP1 immobilization should be the standard approach for cellular assays. The use of purified recombinant protein also reduces the risk of confounded data. Modified cellular expression of SVEP1 in vitro is also not ideal, since SVEP1 produced by cultured cells (which typically lack a robust matrix) is readily excreted into the media as a soluble protein. Rigorous and physiologic assay design is necessary to improve reproducibility and dissect the precise mechanisms of the effects of SVEP1 on cell behavior.

SVEP1 and vascular development

Murine embryos lacking SVEP1 exhibit marked edema by mid-gestation and die immediately after birth [26,28]; this is attributed to severe disruptions in lymphatic vessels and capillaries [11,26,28]. Zebrafish lacking Svep1 also have lymphatic defects, suggesting that the role of SVEP1 in lymphatic development is evolutionarily conserved [28,60]. Zebrafish studies also support a role of Svep1 in blood vessel anastomosis [29]. Similarly, mice lacking SVEP1 in neural crest-derived cells develop a hypomorphic **Schlemm's canal** and disrupted vasculature within the eye [11]. Consistent with these developmental findings, analysis of genetic variation within SVEP1 in humans strongly suggests that loss of SVEP1 is not developmentally tolerated [78].

The specific mechanisms by which SVEP1 contributes to development are not well understood, but current understanding is that adjacent endothelial cells respond to SVEP1 produced by nearby mesenchymal cells [6,26,28]. Integrin α9β1 is involved in lymphangiogenesis [53], but the phenotype of I tga $9^{-/-}$ mice is notably milder than Svep1^{-/-} mice [28,79]. Surprisingly, the zebrafish Svep1 lacks the integrin α 9 β 1 binding domain altogether [28], signifying that other proteins likely have a more prominent role. Mice with constitutive loss of *Pear1* also fail to phenocopy the developmental defects observed in $Svep1^{-/-}$ mice, despite the role of PEAR1 in neoangiogenesis [29,58]. The potential regulation of ANG-TIE signaling by SVEP1 is a promising explanation for these developmental effects. Interaction with ANG1 and ANG2 has been proposed as a mechanism [26], but in vivo evidence is lacking. Direct interaction between SVEP1 and the tyrosine-protein kinase receptor TIE1 [60,61] has also recently been proposed by independent groups. Further research is needed to characterize how these pathways mediate the critical role of SVEP1 in vascular development.

SVEP1 and human disease

Our understanding of SVEP1's role in disease has been greatly influenced by its human genomic and proteomic associations. **Genome-wide association studies (GWASs)** have revealed several associations between SVEP1 and human traits and diseases. **Aptamerbased proteomics** have rapidly expanded these associations, with many of these studies identifying SVEP1 as the protein most strongly associated with the outcome of interest. Analyses of these data by MR has yielded additional causal associations. Table 1 outlines the SVEP1 disease and trait associations with the strongest supporting data (Table 1, Figure 2), and categories of SVEP1 disease associations are discussed in more detail below. A summary of their supporting data and proposed mechanisms is included; however, the mechanisms underlying the associations remain poorly understood.

Cardiometabolic disease

In 2016, we reported a robust association between a coding variant in $SVEPI$ (rs111245230, encoding p.D2702G) and coronary artery disease (CAD) [80]. This variant also associated with blood pressure and type 2 diabetes, but not with lipids, suggesting it may influence vascular disease pathogenesis in a lipid-independent manner. Individuals with increased plasma levels of SVEP1 were subsequently found to be at greater risk for incident CAD

and type 2 diabetes [8]. Increased plasma SVEP1 also associates with increased systolic blood pressure and poor survival after incident CAD [8]. MR analyses suggest SVEP1 is deleterious, since genetic variation which associate with elevated levels of plasma SVEP1 also positively associate with CAD [6,7], hypertension [6], and type 2 diabetes [6,8].

A recent study of patients with heart failure found that plasma SVEP1 was as strongly associated with heart-failure-associated hospitalization or cardiovascular death as N-terminal pro-B-type natriuretic peptide (NT-proBNP) [4]. Increased plasma SVEP1 was also shown to portend a poor prognosis in patients with pulmonary arterial hypertension [12]. A machine learning proteomic surrogate for cardiovascular outcomes identified SVEP1 as one of the top plasma proteins predictive of myocardial infarction, stroke, heart failure, or death [13].

Consistent with these observations, SVEP1-deficient mouse models were found to have lower plaque burden and complexity when compared to controls [6]. Winkler *et al.* reported the opposite phenotype [30] in a murine study confounded by differing proportions of male and female animals [81,82]. Integrin signaling [25], Notch signaling [25], and differential CXC motif chemokine ligand (CXCL)1 expression [30] have all been proposed as the molecular mechanisms behind these animal phenotypes; however, $Itga9^{-/-}$ mouse models fail to phenocopy the atherogenesis phenotypes of $Svep1^{-/-}$ mice [25] and changes in CXCL1 expression failed to replicate in other models of SVEP1 depletion [6,7]. Interaction with PEAR1 is an intriguing but untested hypothesis related to how SVEP1 influences atherosclerosis, since PEAR1 also appears to causally relate to CAD [7,56,57,83] through unclear mechanisms.

Additional cardiometabolic phenotypes have also been studied in mice with experimentally reduced SVEP1. Ex vivo aortic contraction studies from $Svep1^{+/-}$ mice suggest SVEP1 may regulate vascular contraction through integrins α 9 and α 4 [24]. Despite this finding, mice lacking SVEP1 post development do not appear to have a clear vascular reactivity phenotype when assessed by *in vivo* catheterization [7]. These mice similarly lack a glucose metabolism phenotype, as assessed by glucose and insulin tolerance tests, as well as indirect calorimetry [7]. Future studies are needed to clarify the role and mechanisms of SVEP1 in murine **cardiometabolic disease**, given the inconsistencies in vascular and atherosclerotic phenotypes reported for murine SVEP1-depletion models. The association between increased plasma SVEP1 and poor cardiovascular outcomes in humans is nonetheless stunning. Additional plasma proteomic studies are ongoing and are likely to reveal additional associations between SVEP1 cardiometabolic disease.

Platelet biology and hematopoiesis

Recent data points to an intriguing role of SVEP1 in platelet biology and hematopoiesis. A missense polymorphism (rs61751937 encoding SVEP1 p.R229G) strongly associates with platelet reactivity in response to ADP [15]; SVEP1 R229G, along with other SVEP1 variants, also associate with additional platelet traits [84], and MR suggests these SVEP1 trait associations are causal [7]. Mice lacking SVEP1 postdevelopment had similar platelet counts to controls, but their platelets had fewer platelet preactivation markers in response to ADP. Exogenous SVEP1 induced adhesion, activation, and agglutination of platelets ex vivo,

Variation within the locus containing SVEP1 is associated with numerous additional hematological parameters, including red blood cell traits [85] and white blood cell counts [84]. Mice lacking SVEP1 had increased lymphocytes and white blood cell counts, in addition to increased red blood cell counts [7]. These findings overlap with the hematological associations of *SVEP1* in humans [84] and suggest the protein may regulate hematopoiesis, as has been described in animal models [86] and ex vivo [87].

platelets by SVEP1 was shown to depend on PEAR1 [7].

These findings support the human disease associations of SVEP1 and implicate the protein in platelet biology and hematopoiesis. SVEP1 also associates with death from septic shock [88], perhaps reflecting its association with vascular and hematological traits. The mechanisms by which SVEP1 impacts hematopoiesis remain unclear; however, integrin α9β1 [89] and PEAR1 [59] are also thought to regulate hematopoiesis. How SVEP1 influences platelet biology and hematopoiesis is an important area of future study that may relate to the other SVEP1 disease associations.

Glaucoma

Glaucoma is also associated with variation within the $SVEPI$ locus. The same coding variant that associates with platelet reactivity (p.R229G) was found to associate with open angle glaucoma in a GWAS of adults [10]. Glycine at 229 correlates with increased plasma SVEP1 levels (through unclear mechanisms), suggesting elevated SVEP1 may contribute to open angle glaucoma. Another missense allele within SVEP1 (rs761025824 encoding p.R997C) was found in four of five affected members of a family of patients with primary congenital glaucoma and who also harbored a mutation in TEK (the gene that encodes TIE2 which is a receptor for ANG1) [90]. Exposure of human umbilical vein endothelial cells to SVEP1 R997C resulted in less TEK expression than wild-type SVEP1, suggesting that SVEP1 modulates TEK-related primary congenital glaucoma [90]. Deletion of Svep1 in neural crest cells of mice resulted in defects within Schlemm's canal and increased intraocular pressure, suggesting insufficient SVEP1 may lead to primary congenital glaucoma [11].

Future studies are necessary to determine if chronic excess SVEP1 is also sufficient to disrupt Schlemm's canal and promote glaucoma, as one might hypothesize from the human GWAS. It is unclear how SVEP1 governs the development of Schlemm's canal; however, its potential interaction with the ANG–TIE signaling pathway [26,60,90] is a leading hypothesis. Both Pear1 and Itga9 are also expressed within Schlemm's canal and abundant PEAR1 is observed in trabecular meshwork cell–ECM adhesion complexes [91], raising questions about whether these receptors also interact with SVEP1 to govern development or homeostasis of Schlemm's canal.

Aging and cognitive function

The disease associations of SVEP1 are broad, but themes related to the pathophysiology of these diseases provide clues to the function of the protein. Relation to aging and chronicity are characteristics of many of its associated diseases. Additional studies have identified

genetic and plasma protein associations between SVEP1 and measures of aging or cognitive decline [13,16,92]. One such analysis found SVEP1 to be the plasma protein with the strongest causal association with dementia among 4877 proteins analyzed [5]. A causal association between plasma SVEP1 and decreased parental lifespan has also been reported [9].

Although increased SVEP1 appears to constitute a health risk during aging, it is also necessary for embryonic viability and vascular development (Figure 3). This biological phenomenon, termed **antagonistic pleiotropy**, has long been described [93]. According to this hypothesis, selective pressure across evolution heavily favors alleles necessary for survival up to the peak reproductive capacity of an organism, even if such alleles have a negative effect on survivorship after reproductive capacity wanes [94,95]. The effects of SVEP1 on human biology appear to closely follow this paradigm, given its causal association with age-related disease.

The ECM is thought to play a critical role in aging [96], but the specific effects of SVEP1 remain unclear. The robust activation of mTOR signaling in endothelial and smooth muscle cells [7] provides a reasonable hypothesis, since mTOR is among the most recognized regulators of longevity [72,74]. Further addressing this intriguing question may shed light on the biology underlying the disease associations of SVEP1 and will be an important direction for future investigation.

Concluding remarks

SVEP1 is a complex and intriguing ECM protein. Recent data implicate the protein in vascular development, chronic disease, and aging. Despite this, there is no consensus on how SVEP1 regulates cells, tissues, or disease. Future studies are necessary to characterize the location of SVEP1 within tissue and its molecular interactions. Elucidating these mechanisms will broaden our understanding of vascular development and disease pathogenesis. Rigorous antibody validation and the use of more physiologic cellular assays are important steps to accomplishing these goals.

Measurements of plasma SVEP1 concentration may have diagnostic and prognostic value across many fields in medicine. The development of a targeted assay to measure plasma SVEP1 concentration is a critical next step to evaluating its clinical utility, since the disease associations of SVEP1 were established using untargeted proteomic techniques. Such an assay can also address important questions related to the protein's apparent ingress into plasma, stability, and other variables that influence its concentration (see Outstanding questions).

Variation of plasma SVEP1 concentration is, in part, genetically determined, suggesting a therapeutic window exists to safely reduce SVEP1 activity (see Clinician's corner). Mouse models of chronic SVEP1 depletion and the theory of antagonistic pleiotropy also hint that functional levels of plasma SVEP1 could be safely reduced in adults. These characteristics, combined with its existence within the extracellular space, make it a priority candidate for intervention. The development of therapies related to SVEP1 require a deeper understanding

of the molecular mechanisms by which it contributes to disease, emphasizing the need for continued research on this fascinating protein.

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Glossary

AKT

serine/threonine kinase that plays a crucial role in cell survival, growth, and metabolism. It is involved in various cellular processes, including cell proliferation, apoptosis, and protein synthesis.

ANG–TIE

The signaling pathway involving the interaction between ANGs and TIE receptors. This pathway plays a significant role in angiogenesis, the formation of new vessels.

Antagonistic pleiotropy

description of a situation where a single gene or genetic variant has multiple effects on different traits or functions. These effects can be beneficial for certain traits at one stage of life but detrimental to other traits at a different stage, leading to trade-offs in evolutionary fitness.

Aptamer-based proteomics

A technique that uses DNA or RNA oligomers called aptamers that bind to known target proteins. This method enables the identification and quantification of proteins in complex biological samples at a greater scale than traditional methods.

Cardiometabolic disease

cluster of common chronic medical conditions which share interrelated risk factors such as high blood pressure, elevated blood sugar, and elevated lipids. This term encompasses cardiovascular conditions such as heart disease and stroke along with metabolic disorders such as type 2 diabetes and obesity.

Extracellular matrix (ECM)

complex network of proteins and carbohydrates that provides structural support and biochemical cues to cells in tissues and organs. It influences various cellular processes, including cell adhesion, migration, and differentiation.

Genome-wide association studies (GWASs)

large studies that investigate the association between genetic variants across the genome and the prevalence of specific traits or diseases. GWASs help identify genetic markers associated with various conditions and provide insights into the genetic basis of complex traits.

Integrins

family of cell surface receptors that mediate cell–cell and cell–ECM interactions. They play a crucial role in cell adhesion, migration, signaling, and tissue organization.

Label-free binding assay

technique used to measure the interaction between molecules without the need for labeling or modification of the molecules. It enables the quantitative analysis of binding affinities and kinetics between proteins, small molecules, or other biomolecules.

Lymphangiogenesis

process of formation and remodeling of lymphatic vessels. It involves the growth and branching of lymphatic endothelial cells and plays a critical role in immune responses and tissue fluid homeostasis.

Mendelian Randomization

statistical method that uses genetic variants as instrumental variables to assess causal relationships between exposures and disease outcomes. It leverages the principles of Mendelian inheritance to infer causal effects in observational studies.

mTOR

mechanistic target of rapamycin (mTOR) is a protein kinase that regulates cell growth, metabolism, and survival in response to various environmental signals. It integrates inputs from growth factors, nutrients, and cellular energy status to control protein synthesis, aging, and other important processes.

Platelet endothelial aggregation receptor 1 (PEAR1)

receptor tyrosine kinase-like protein that strongly associates with platelet reactivity in humans and is activated by SVEP1.

Schlemm's canal

lymphatic-like vessel located near the anterior chamber angle of the eye. It plays a crucial role in regulating intraocular pressure by maintaining the proper drainage of aqueous humor.

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Box 1.

Basics of Mendelian Randomization

Mendelian randomization (MR) uses an experiment of nature to test for causal associations. It utilizes alleles (which are stochastically distributed across a population according to the law of independent assortment) that affect a measured variable, such as the plasma concentration of its gene product. An outcome of interest can then be regressed on the genetically encoded changes of such a variable to test causality.

This approach can estimate the causal relationship of SVEP1 to disease through regression of genetically encoded changes in plasma SVEP1 concentration with outcomes data from GWASs. One limitation in applying this approach to plasma SVEP1 is that its concentration is thought to be regulated, in part, by leakage from origin tissues [18]. This biology obscures the anatomical location of the effects of SVEP1 and may bias the magnitude of the causal estimate, though likely toward the null. Nonetheless, MR remains a powerful tool to test if the observed correlations between SVEP1 and disease reflect causal relationships.

Highlights

Human genomic and proteomic data implicate Sushi, von Willebrand factor type A, epidermal growth factor, and pentraxin domain containing 1 (SVEP1) in the pathogenesis of human chronic disease, including cardiovascular disease, glaucoma, and dementia.

SVEP1 is critical for vascular development and appears to promote aging, consistent with the antagonistic pleiotropy theory of aging.

Many cell types adhere and/or proliferate in response to SVEP1, but the mechanisms underlying its role in development and disease are unclear.

SVEP1 is contained within the extracellular matrix and circulates in plasma, making it a priority candidate for disease diagnostics, prognostics, and therapeutics.

Clinician's corner

SVEP1 is an ECM protein with important roles in vascular development, chronic disease, and aging.

Measuring plasma SVEP1 concentration may have significant diagnostic and prognostic value across various fields of medicine.

SVEP1 appears to associate as strongly as NT-proBNP with heart failure hospitalization or cardiovascular death in patients with heart failure.

Investigating the molecular mechanisms of SVEP1 could lead to new therapeutic approaches to treat or prevent disease.

Outstanding questions

What is the precise location of SVEP1 within the ECM? How does SVEP1 enter circulation and in what form does it circulate?

Which proteins interact with SVEP1 in vivo, and how do these interactions contribute to its role in development and disease?

What is the diagnostic or prognostic value of measuring circulating SVEP1? Does this approximate gestational age, cardiovascular risk, platelet reactivity, risk of developing glaucoma, and/or biological age?

Is it possible to safely target SVEP1 for the treatment or prevention of human chronic disease? What are the most effective strategies to modulate its activity?

Figure 1.

Sushi, von Willebrand factor type A, epidermal growth factor and pentraxin domain containing 1 (SVEP1) protein schematic. Simple Modular Architecture Research Tool and InterPro were used to identify and map domains [97,98]. Coding variants that associate with human disease are indicated at their corresponding peptide. Adapted, with permission, from Elenbaas et al. [7].

Trends in Molecular Medicine

Figure 2.

Sushi, von Willebrand factor type A, epidermal growth factor and pentraxin domain containing 1 (SVEP1) human disease associations. A list of the SVEP1 human disease and trait associations with the strongest supporting data. The traits and diseases with a causal association with SVEP1 are indicated with solid lines. Dashed lines represented correlative associations. Pointers indicate organs affected by the associated trait or disease, not necessarily the physiological location of the effects of SVEP1. Of note, many of the associated diseases are chronic conditions that relate to cardiometabolic or vascular dysfunction. See Table 1 for the associated references.

selected without regard for negative consequences in late life, when the force of natural selection is negligible Antagonistic Pleiotropy: one allele has a positive effect on one trait but a negative effect on another Pleiotropy: one allele affects multiple different traits

Trends in Molecular Medicine

Figure 3.

Sushi, von Willebrand factor type A, epidermal growth factor and pentraxin domain containing 1 (SVEP1) throughout life. SVEP1 and the antagonistic pleiotropy theory of aging. SVEP1 is essential for embryonic viability and appears to promote age-related disease later in life. This can be explained by the theory of antagonistic pleiotropy, which states that alleles necessary for development and reproduction are favored without regard for their effects in later life. The loss of selective pressure on alleles during the period proceeding max reproductive capability is referred to as the selection shadow.

Table 1.

SVEP1 human disease and trait associations

