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Nano transdermal system combining mitochondria-targeting cerium oxide nanoparticles with all-trans retinoic acid for psoriasis



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ABSTRACT

Psoriasis is an inflammatory skin disease that is intricately linked to oxidative stress. Antioxidation and inhibition of abnormal proliferation of keratinocytes are pivotal strategies for psoriasis. Delivering drugs with these effects to the site of skin lesions is a challenge that needs to be solved. Herein, we reported a nanotransdermal delivery system composed of all-trans retinoic acid (TRA), triphenylphosphine (TPP)-modified cerium oxide (CeO₂) nanoparticles, flexible nanoliposomes and gels (TCeO₂-TRA-FNL-Gel). The results revealed that TCeO₂ synthesized by the anti-micelle method, with a size of approximately 5 nm, possessed excellent mitochondrial targeting ability and valence conversion capability related to scavenging reactive oxygen species (ROS). TCeO₂-TRA-FNL prepared by the film dispersion method, with a size of approximately 70 nm, showed high drug encapsulation efficiency (>96%). TCeO₂-TRA-FNL-Gel further showed sustained drug release behaviors, great transdermal permeation ability, and greater skin retention than the free TRA. The results of *in vitro* EGF-induced and H₂O₂-induced models suggested that TCeO₂-TRA-FNL effectively reduced the level of inflammation and alleviated oxidative stress in HaCat cells. The results of *in vivo* imiquimod (IMQ)-induced model indicated that TCeO₂-TRA-FNL-Gel could greatly alleviate the psoriasis symptoms. In summary, the transdermal drug delivery system designed in this study has shown excellent therapeutic effects on psoriasis and is prospective for the safe and accurate therapy of psoriasis.

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1. Introduction

Psoriasis is an inflammatory skin disease mainly mediated by immunity and it affects millions of people around the world [1–2]. It is reported that topical therapy, phototherapy, and immunotherapy have been used for the treatment [3–5]. Topical therapy mainly utilizes drugs such as retinoids, vitamin D derivatives to treat psoriasis [6]. These drugs are effective to some extent, but their demerits, including skin irritation and instability, are still stumbling blocks for better clinical efficacy [7]. Hence, the exploration of an appropriate drug delivery system is of vital necessity and urgency for patients.

Retinoic acid has been used to treat skin disease for nearly 40 years [8]. All-trans retinoic acid (TRA) is regarded as the first-generation therapeutic drug used for skin diseases, such as wound healing, acne, and psoriasis [9–11]. TRA can bind with retinoic acid receptors such as RAR- α and RAR- γ and then regulate the abnormal differentiation of keratinocytes, which is beneficial for relieving the proliferation of keratinocytes and reducing the level of inflammation [12,13]. Although TRA is useful for psoriasis, its application is hindered because of its sensitivity to light and oxygen, poor water solubility, and skin irritation [14].

In addition to conventional therapy, the mitigation of oxidative stress is also important for psoriasis [15]. When the skin is exposed to oxidative stress, a large amount of reactive oxygen species (ROS) is generated that can participate in promoting cell proliferation, differentiation, and inflammation by regulating signal pathways such as NF- κ B, MAPK and JAK-STAT pathways [16]. Natural compounds such as epigallocatechin-3-gallate, glycyrrhizic isoflavones and proanthocyanidins have been used to alleviate the oxidative stress state of psoriasis [17–19]. However, the use of these active components is still limited because their efficient extraction and separation still need further study. Recently, nanomaterials with enzyme-like activity have attracted research interest. Among nanomaterial-based antioxidants, cerium oxide (CeO₂) nanoparticles exhibit tremendous potential as superoxide dismutase (SOD)/catalase (CAT)-mimetics by scavenging ROS through shuttling between the redox state (Ce³⁺) and oxidation state (Ce⁴⁺) [20,21]. They have also been applied in major diseases, including Alzheimer's disease, ischemic stroke, and cancer [22–24]. The activity of eradicating ROS makes CeO₂ a promising candidate for psoriasis therapy. Meanwhile, there is a close connection between mitochondria and ROS. Both endogenous and exogenous ROS can easily damage mitochondrial DNA (mtDNA) because mtDNA has the limited repair capacity and a lack of histone barrier [25,26]. mtDNA damage causes mitochondrial dysfunction and is mainly involved in psoriasis through two pathways. On the one hand, mtDNA is released into the cytoplasm as mitochondrial damage-associated molecular patterns, activating Toll-like receptor (TLR)–9 [27,28]. The activation of TLR-9 can take part in subsequent inflammatory processes by inducing several signaling pathways [29–31]. On the other hand, mtDNA damage can lead to loss of mitochondrial polypeptide expression to cause defective functioning of the electron

transport chain (ETC), further increasing ROS [32–34]. ROS can participate in promoting cell proliferation, differentiation, and inflammation by regulating some signaling pathways [16]. Hence, a mitochondria-targeting strategy is beneficial for psoriasis therapy. Triphenylphosphine (TPP) is a lipophilic cation whose lipid solubility and positive charge endow it with mitochondria-targeting ability [35,36]. The combination of CeO₂ and TPP can anchor the mitochondria of keratinocytes and accurately remove ROS. However, the poor skin absorption of CeO₂ is the main obstacle remaining to be solved.

Transdermal delivery is attractive for the treatment of psoriasis because of the imperfect skin barrier, but it is challenging to realize efficient and safe delivery [37]. On the one hand, the stratum corneum can extremely block the penetration of most drugs due to its highly organized structure [38]. On the other hand, some properties, such as high molecular weight and poor water solubility, affect drug delivery [39]. In particular, the broken barrier also limits the delivery of drugs with skin irradiation in some skin diseases, such as psoriasis. Currently, many novel drug delivery systems have been explored such as dendrimers, ethosomes and transfersomes [40–42]. Flexible nanoliposomes are prospective carriers for transdermal delivery. It can increase the stability of encapsulated drugs and increase the permeability of drugs into the skin [43]. Distinct from the rigid structure of liposomes, they contain membrane softeners such as ethanol, Tween-80, and cholate, which contribute substantially to their high deformability, flexibility, and skin permeability [44,45]. Flexible nanoliposomes are commonly used in skin diseases, such as wound healing and psoriasis. Topical gels aiming to realize the efficient delivery of drugs to skin are also used in skin diseases to improve skin adhesion and prolong skin retention time [45].

In particular, we reported a nanotransdermal delivery system composed of TRA, TPP-modified CeO₂ nanoparticles, flexible nanoliposomes, and gels (TCeO₂-TRA-FNL-Gel). TRA inhibited the excessive proliferation of keratinocytes, and TCeO₂ effectively scavenged ROS by targeting mitochondria, thus alleviating oxidative stress in psoriatic skin. Using flexible nanoliposomes as carriers could improve drug stability and penetration efficiency by virtue of their high deformability and excellent permeability as well as significantly increase drug accumulation in the dermis. The uniform and stable network structure of gels could improve the adhesion of flexible nanoliposomes to skin and finally achieve the safe and accurate treatment of psoriasis.

2. Materials and methods

2.1. Materials

Cerium (III) acetate hydrate, soybean lecithin, TRA and carbomer 940 were obtained from Dalian Meilun Biotechnology Co., Ltd. (Dalian, China). Oleylamine, xylene, triphenylphosphine and Tween-80 were obtained from Macklin Biochemical Co., Ltd. (Shanghai, China). Fluorescein isothiocyanate was obtained from Aladdin Biochemical Technology Co., Ltd. (Shanghai, China). Fetal bovine serum

(FBS) was obtained from Hangzhou Tianhang Biotechnology Co., Ltd. (Hangzhou, China). Dulbecco's minimum essential medium (DMEM) was obtained from Senrui Biotechnology Co., Ltd. (Zhejiang, China). IMQ (5%) was obtained from Sichuan Med-shine Pharmaceutical Co., Ltd. (Sichuan, China). ELISA kits for IL-6 and TNF- α were obtained from Jiangsu Meimian Industrial Co., Ltd. (Jiangsu, China). SOD kits and ROS kits were obtained from Beyotime Biotechnology Co., Ltd. (Shanghai, China). HaCat cells were obtained from Procell Life Science & Technology Co., Ltd. (Wuhan, China) and cultured in DMEM with 10% FBS in a 37 °C cell incubator with 5% CO₂. BALB/c mice (female, 6 weeks old, 15 to 20 g) were obtained from Hangzhou Medical College (Hangzhou, China). All animal experiments were conducted according to the rules of the animal care and use committee of Zhejiang University.

2.2. Synthesis of TCeO₂

TPP-modified CeO₂ (TCeO₂) nanoparticles were synthesized by the reverse micelle method [46,47]. A total of 0.43 g cerium (III) acetate hydrate, 2.14 g oleylamine, and 1.05 g TPP were added into 15 ml xylene and then heated at 90 °C under an argon atmosphere after stirring for 18 h. When the liquid became clear, 1 ml deionized water was quickly added, then continuously heated at 90 °C for 3 h until the liquid turned yellow. After cooling to room temperature, 100 ml ethanol and centrifugation (4,000 rpm, 15 min) were used to collect TCeO₂, which was finally stored in chloroform.

2.3. Characterization of TCeO₂

The particle size and morphology were observed by dynamic light scattering (DLS) instrument (Litesizer 500, Anton-Paar, Austria) and transmission electron microscopy (TEM) (JEM-1400, JEOL, Japan). Spectra of samples were analyzed by Fourier transform infrared spectrometry (FTIR) (NICOLET iS50FT-IR, Thermo Scientific, USA) and nuclear magnetic resonance spectroscopy (NMR) (Avance III 500 M, Bruker Biospin, Switzerland). The cerium ion valence was obtained by X-ray photoelectron spectroscopy (XPS) (ESCALAB 250Xi, Thermo Scientific, USA), and the valence cycling ability was evaluated by stimulating H₂O₂. The cerium concentration was analyzed by inductively coupled plasma-optical emission spectrometry (ICP-MS) (Agilent 7800, Agilent Technologies, USA).

2.4. Preparation and characterization of flexible nanoliposomes

Flexible nanoliposomes coloaded with TCeO₂ and TRA (TCeO₂-TRA-FNL) were obtained by film dispersion method. First, soybean lecithin, Tween 80, TRA and TCeO₂ were added into 10 ml absolute ethanol according to Table 1. Second, the mixture was evaporated in the dark until the film was completely formed, and then PBS (4 ml, pH 6.8) was added for hydration. Finally, TCeO₂-TRA-FNL was obtained after probe sonication.

The particle size and morphology were observed by DLS and TEM. Encapsulation efficiency (EE) and drug loading (DL) were measured by ultrafiltration centrifugation. In brief, 0.5 ml

Table 1 – Weight of added substances in liposomes.

Substance	Added weight (mg)
Soybean lecithin	400
Tween 80	80
TRA	4
Cerium in TCeO ₂	4

flexible nanoliposome suspension was ultrafiltered (MWCO: 100 kD) and centrifuged (5,000 rpm, 10 min) to separate the free and encapsulated TRA. The concentrations of TRA were detected by HPLC shown in Table S1. The concentrations of unencapsulated cerium and total cerium were detected by ICP-MS. The following equations were used to calculate the EE and DL:

$$EE (\%) = \frac{\text{total mass of drug} - \text{mass of unencapsulated drug}}{\text{total mass of drug}} \times 100\%$$

$$DL (\%) = \frac{\text{total mass of drug} - \text{mass of unencapsulated drug}}{\text{total mass of liposomes}} \times 100\%$$

2.5. Preparation of gel loaded with flexible nanoliposomes

The blank gel was first prepared according to the prescription, in which carbomer 940, glycerin, EDTA and ethylparaben accounted for 1%, 10%, 0.1%, 0.1% (w/w), respectively. Next, the pH was adjusted to 6.5 by adding triethanolamine. Finally, the nanoliposomes were mixed with blank gel, and the TRA content was controlled to 0.05% to obtain the final gel.

2.6. In vitro TRA release of gel

The TRA release behavior of the gel was investigated by the dialysis method, and a mixture of ethanol and PBS (pH 6.8) (1:1, v/v) was used as the release medium. Next, 0.5 g of the gel was added into a dialysis bag (MWCO: 3.5 kD) and then put in 10 ml release medium with sustained oscillation (60 rpm, 37 °C). The whole release medium was collected at preset time points and TRA concentration was determined by HPLC.

2.7. In vitro skin retention and permeability of gel

The skin retention and permeability of gel was investigated by the diffusion cell method with skin obtained from an experimental pig. A mixture of ethanol and PBS (pH 6.8) (1:1, v/v) was the receiving medium. The skin was fixed flat on the junction of the diffusion cell. 1 g of gel was put into the donor chamber and 15 ml the medium was added into the receptor chamber (37 °C, 300 rpm). Then, 200 μ l the receiving medium was collected at preset time points and TRA concentration was determined by HPLC.

Next, the skin was collected and cleaned with PBS (pH 6.8), and TRA was extracted with 1 ml methanol. The TRA concentration that remained in the skin was determined by HPLC after centrifugation (5,000 rpm, 10 min). Skin treated with FITC-labeled TCeO₂-TRA-FNL-Gel for 24 h was also taken as the sample to evaluate the time-dependent skin permeability.

2.8. Rheology characterization of gel

The kinetics of gel were studied by a rheometer (DHR 2, TA instrument, America). Gels with a thickness of 1 mm were placed between the parallel plate (25 mm in diameter) and the rheometer. A strain sweep test was performed from 0.01% to 1,000% (frequency: 1 Hz) to measure the modulus of the gels (G' : storage modulus, G'' : loss modulus) [48]. The complex viscosity of gels was also obtained in this process.

2.9. In vitro cytotoxicity

HaCat cells were incubated for 24 h (1×10^4 cells/well, 96-well plates). T_{CeO₂}-TRA-FNL (0, 250, 500, 750, 1,000, 1,250, 1,500 and 2,000 µg/ml) was added and then cultured for 24 h. Every concentration had six repetitions. The viability of HaCat cells was studied by CCK-8 assay.

2.10. Cellular uptake

HaCat cells were incubated for 24 h (5×10^4 cells/well, 24-well plates). FITC-labeled T_{CeO₂}-TRA-FNL (400 µg/ml) was added and then cultured for preset time points. The process of cellular uptake was observed through confocal laser scanning microscopy (CLSM).

2.11. Mitochondria-targeting ability

First, ODA-FITC and T_{CeO₂}/CeO₂ were mixed (1:2, w/w) and stirred continuously for 8 h to obtain FITC-labeled T_{CeO₂}/CeO₂. Then, FITC-labeled T_{CeO₂}-FNL/CeO₂-FNL was prepared following the method mentioned previously.

HaCat cells were incubated for 24 h (5×10^4 cells/well, 24-well plates). FITC-labeled T_{CeO₂}-FNL/CeO₂-FNL (400 µg/ml) was added and then cultured for preset time points. Finally, MitoTracker Deep Red (100 nM) was coincubated with HaCat cells for 40 min to stain the mitochondria. The colocalization between T_{CeO₂}-FNL/CeO₂-FNL and mitochondria was observed through CLSM and ImageJ.

2.12. In vitro antioxidant ability

Intracellular ROS detection: HaCat cells were incubated for 24 h (1×10^5 cells/well, 12-well plates). After preincubation with different preparations (T_{CeO₂}-TRA-FNL, T_{CeO₂}-FNL, CeO₂-FNL, TRA-FNL, and Free-TRA) for 4 h, HaCat cells were treated with H₂O₂ (750 µM) for 24 h. Then, a DCFH-DA probe (10 µM) was coincubated with HaCat cells for 30 min. Finally, the ROS detection were observed through CLSM.

SOD level determination: HaCat cells were incubated for 24 h (1×10^5 cells/well, 12-well plates). After preincubation with different preparations (T_{CeO₂}-TRA-FNL, T_{CeO₂}-FNL, CeO₂-FNL, TRA-FNL, and Free-TRA) for 4 h, HaCat cells were treated with H₂O₂ (750 µM) for 24 h. Then, the supernatant collected by lysis and centrifugation (12,000 rpm, 10 min) was used to determine the level of SOD.

2.13. In vitro anti-inflammatory ability

HaCat cells were incubated for 24 h (1×10^5 cells/well, 12-well plates). After preincubation with EGF (80 ng/ml) for 1 h, HaCat cells were treated with different preparations (T_{CeO₂}-

TRA-FNL, T_{CeO₂}-FNL, CeO₂-FNL, TRA-FNL, and Free-TRA) for 24 h. Then, the supernatant after centrifugation (12,000 rpm, 10 min) was collected to determine the level of interleukin-6 (IL-6)/tumor necrosis factor- α (TNF- α).

2.14. Establishment of the psoriasis model

The psoriasis model can be established by applying 5% IMQ at a daily dosage of 62.5 mg to the depilated back skin (2 cm \times 1.5 cm) for 7 d.

2.15. Psoriasis treatment

BALB/c mice were randomly divided into seven groups: (1) control group, (2) IMQ group, (3) T_{CeO₂}-TRA-FNL-Gel group, (4) T_{CeO₂}-FNL-Gel group, (5) CeO₂-FNL-Gel group, (6) TRA-FNL-Gel group, and (7) Free-TRA-Gel group (for each group, $n = 5$). From Day 3, gels were applied at a daily dosage of 100 mg. The whole process, including model establishment and treatment, lasted for 7 d.

2.16. Weight observation and skin PASI score

The weight of the mice was recorded, and the skin condition was evaluated according to the psoriasis area and severity index (PASI).

2.17. Skin compliance study

A skin compliance study of gel was conducted on the skin of mice [49,50]. Gels were applied on the skin at 100 mg/d for 7 d. The skin condition was recorded in the treatment (1, 3, 7, 10 d).

2.18. In vivo antioxidant ability

ROS detection in diseased skin: The skin tissue was stained with ROS dye, and the expression of ROS was detected by immunofluorescence. **SOD level determination:** PBS was utilized to grind the skin tissue into 10% homogenate. Then, centrifugation (12,000 rpm, 10 min) was used to collect the supernatant, which was utilized to determine the level of SOD.

2.19. In vitro anti-inflammatory ability

PBS was utilized to grind the skin tissue into a 10% homogenate. Then, centrifugation (12,000 rpm, 10 min) was used to collect the supernatant, which was utilized to measure the levels of IL-6/TNF- α .

2.20. Skin histopathology and immunohistochemistry

The skin tissue was removed from back for H&E staining. At the same time, the expression levels of CD3, CD31 and Ki67 were examined by immunohistochemistry.

2.21. Statistical analysis

We utilize T test to determine differences between two groups. $P < 0.05$ was considered significant.

3. Results and discussion

3.1. Synthesis and characterization of TCeO₂

TCeO₂ was obtained by the reverse micelle method [46,47]. The DLS and TEM results were shown in Fig. 1A and 1B and Table S2. The particle size of TCeO₂ was 5.82 ± 0.24 nm, while that of CeO₂ was 4.54 ± 0.39 nm. Clearly, the sizes of both particles were small, and the TEM image also showed a small size and fine morphology. It has been reported that the ratio of surface area to volume was high due to the small size, which is beneficial for providing a large surface area for catalysis [51]. Hence, the TCeO₂ we synthesized might have potential catalytic activity.

The FTIR result in Fig. 1C showed that compared with CeO₂, new peaks (1120–1050 cm⁻¹) appeared in the spectrum of TCeO₂, which was consistent with that in the spectrum of TPP, indicating that TPP was successfully modified. The XPS analysis in Fig. 1D proved the existence of phosphorus, and the ¹H/¹³C NMR results in Figs. 1F and S2 also implied

that the same absorption belonging to the phenyl ring of TPP existed in TPP and TCeO₂. All the results further verified the modification of TPP, which is rational to target mitochondria for ROS elimination since TPP has been used to enhance the antioxidant ability of mitochondria in some studies [52,53]. In addition, utilizing the SOD/CAT-mimetic activity of TCeO₂ (SOD: Ce³⁺, CAT: Ce⁴⁺) to remove ROS has been reported to be reasonable and effective. Hence, the valence conversion of TCeO₂ also needs to be considered. Fig. 1E likewise showed that both Ce³⁺ and Ce⁴⁺ existed on the surface of TCeO₂ (Ce³⁺: u₁, v₁; Ce⁴⁺: u₀, u₂, u₃, v₀, v₂, v₃), which was of vital significance for the redox ability of TCeO₂. In addition, we could directly see the color cycling of the TCeO₂ dispersion within 20 d in Fig. 1G. When Ce³⁺ was oxidized to Ce⁴⁺ by H₂O₂, the color turned orange, while when Ce⁴⁺ was converted to Ce³⁺, the color gradually faded back to light yellow. The results both illustrated that TCeO₂ had excellent valence conversion ability and was the basis for ROS eradication, thus playing a prominent role in improving the antioxidant ability of mitochondria.

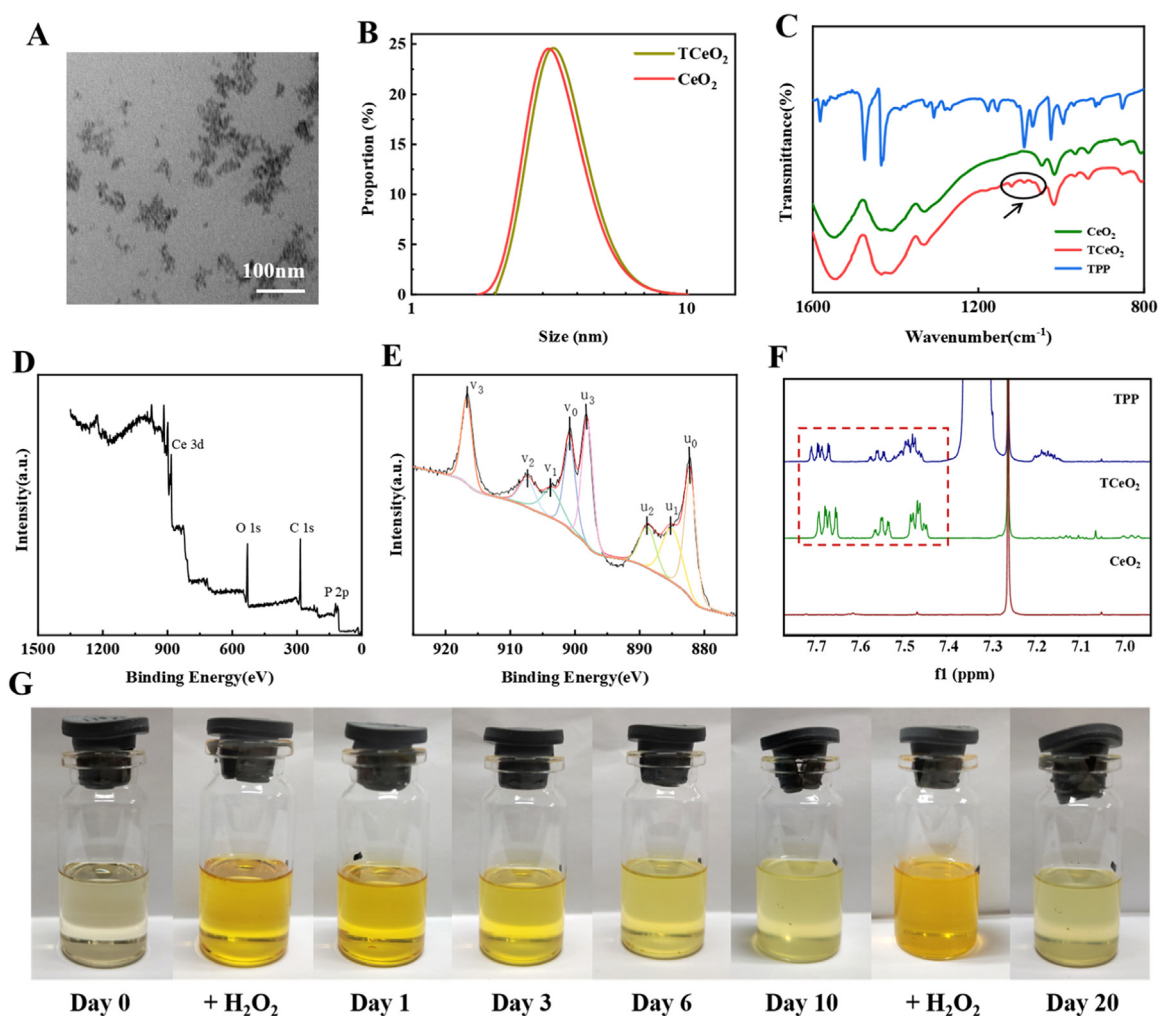


Fig. 1 – Characterization of TCeO₂. (A–B) TEM image of TCeO₂, DLS results of TCeO₂ and CeO₂; (C) Infrared spectra of CeO₂, TCeO₂ and TPP; (D) XPS spectrum of TCeO₂; (E) Ce 3d XPS spectrum of TCeO₂ (u₀=882.20 eV, u₁=885.03 eV, u₂=888.78 eV, u₃=898.11 eV; v₀=900.80 eV, v₁=903.63 eV, v₂=907.38 eV, v₃=916.71 eV); (F) ¹H NMR spectra of TPP, TCeO₂, and CeO₂ (in CDCl₃); (G) The valence cycling ability of TCeO₂.

3.2. Preparation and characterization of gel loaded with flexible nanoliposomes

The nanotransdermal delivery system we reported contained flexible nanoliposomes and gels. Therefore, flexible nanoliposomes and gels were prepared and then evaluated. We first determined the viability of HaCat cells treated with TRA (0–40 $\mu\text{g/ml}$) and cerium (0–60 $\mu\text{g/ml}$) for 24 h. As was shown in Fig. 2A and 2B, we ultimately chose the appropriate ratio of TRA to cerium (w/w, 1:1) to prepare flexible nanoliposomes. Fig. 2D and Table S3 revealed that the average sizes of different flexible nanoliposomes were nearly 60–70 nm, which means that they were promising for transdermal delivery [54]. TEM observation results in Fig. 2C and S1 also indicated that the liposomes had an obvious phospholipid bilayer and were relatively uniform in size. The smaller sizes of flexible liposomes than conventional liposomes were mainly due to the incorporation of Tween 80, which could achieve higher curvature [55]. In addition, the high encapsulation efficiency (TRA > 96%, cerium > 99%) shown in Table S3 might also be attributed to the properties of Tween 80 as a surfactant and solubilizer [56].

Next, gels were prepared, and TRA release behavior as well as skin retention and permeability were studied. Fig. 2E clearly showed that the release rate of TRA was rapid and nearly

all the TRA was released within 8 h in Free-TRA-Gel. In contrast, TRA displayed a slow sustained release for 72 h when encapsulated into liposomes. The sustained release due to the lipid bilayer was helpful in the treatment because more drugs could accumulate in therapeutic sites for durable therapy and side effects were reduced [57]. Fig. 2F described the retention results, and it could be seen that the drug skin retention of TCeO₂-TRA-FNL-Gel was higher than that of Free-TRA-Gel ($P < 0.01$). Specifically, TCeO₂-TRA-FNL-Gel and TRA-FNL-Gel were 1.81 and 1.45 times the drug skin retention of Free-TRA-Gel.

The kinetics of gels were studied by a rheometer (G' : solid property, G'' : liquid property) [48]. It was evident that G' and G'' values were relatively stable in the strain range from 0.01% to 1%, indicating the linear viscoelastic region of TCeO₂-TRA-FNL-Gel. Then, G' values decreased while G'' values increased until intersecting at a strain of approximately 167.8%, which represented the breakdown of the gel network (Fig. 2G). A similar phenomenon was observed in other gels (Fig. S3). The results exhibited the viscoelasticity behavior of the gels we prepared, which was consistent with what a previous study reported [58]. The result in Fig. 2H implied that the decreasing viscosity of the gels was related to the increasing strain, which might be because the internal network of the gels was gradually destroyed [59].

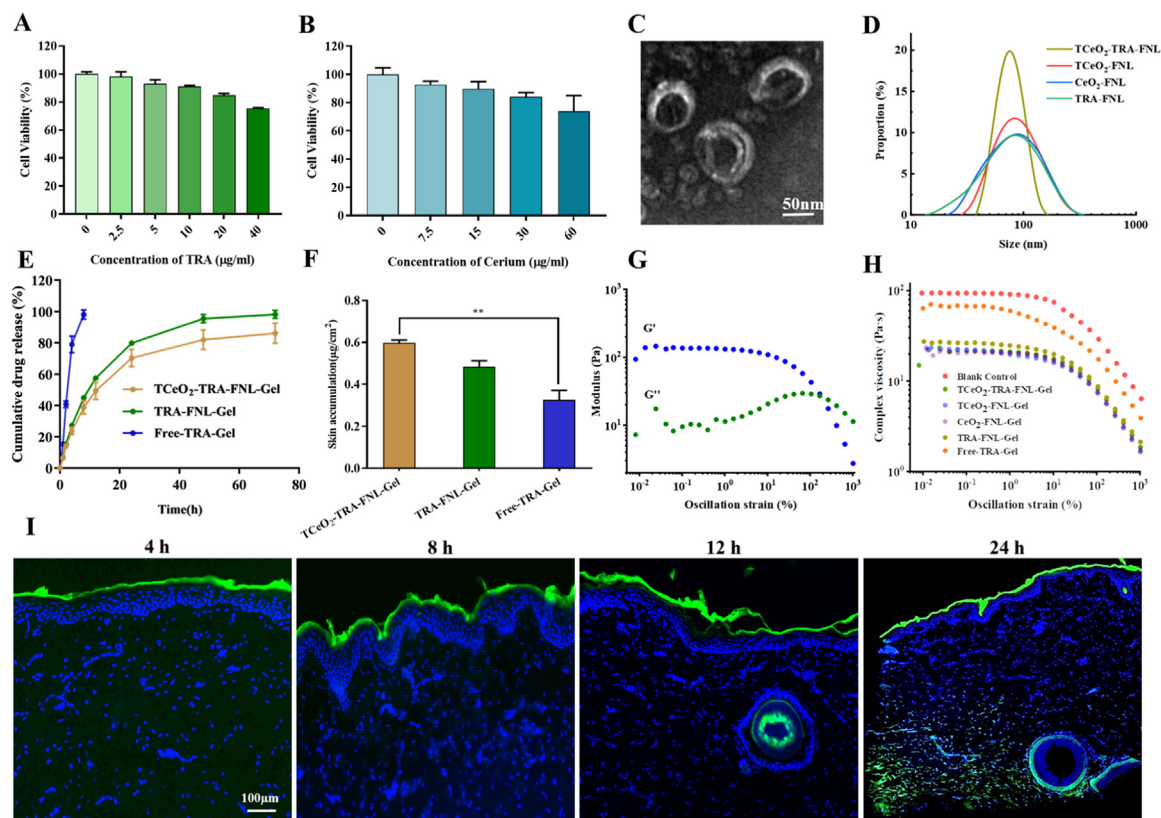


Fig. 2 – Characterization of TCeO₂-TRA-FNL-Gel. (A–B) Viability of HaCat cells after treated with TRA (0–40 $\mu\text{g/ml}$)/Cerium (0–60 $\mu\text{g/ml}$) for 24 h; (C–D) TEM image of TCeO₂-TRA-FNL and DLS results of TCeO₂-TRA-FNL, TCeO₂-FNL, CeO₂-FNL and TRA-FNL; (E) *In vitro* TRA release behaviors of TCeO₂-TRA-FNL-Gel, TRA-FNL-Gel and Free-TRA-Gel; (F) *In vitro* skin retention of TCeO₂-TRA-FNL-Gel, TRA-FNL-Gel and Free-TRA-Gel; (G) The variation of G' and G'' values with strain for TCeO₂-TRA-FNL-Gel; (H) The complex viscosity of gels; (I) CLSM images of pig skin after treatment with TCeO₂-TRA-FNL-Gel for preset time points. Data are expressed as the mean \pm SD, $n = 3$, * $P < 0.05$, ** $P < 0.01$, *** $P < 0.001$.

Furthermore, from the skin permeation result shown in Fig. 2I, we found that the liposomes were mainly distributed on the surface of skin within 12 h due to the blocking effect of the stratum corneum. However, they penetrated downward into the dermis after 24 h, which might be attributed to the extended retention, prolonging the penetration time. The results above demonstrated the sustained release behavior and high skin retention and permeability of flexible nanoliposomes, which were beneficial to the treatment of local diseases such as psoriasis [60].

3.3. Cytotoxicity and cellular uptake

HaCat cells were treated with T_{CeO₂}-TRA-FNL for 24 h and FITC-labeled T_{CeO₂}-TRA-FNL for predetermined time points (2, 4, 8 and 12 h) to evaluate cytotoxicity and cellular uptake ability, respectively. As shown in Fig. 3A, T_{CeO₂}-TRA-FNL (0–2000 µg/ml) showed no obvious cytotoxicity. We also observed that weak green fluorescence appeared after 2 h, and the fluorescence intensity gradually increased with the time of coincubation (Fig. 3B). In brief, T_{CeO₂}-TRA-FNL prepared in this study had excellent biocompatibility and could be successfully absorbed by HaCat cells.

3.4. Mitochondria-targeting ability

The red probe for mitochondria was used to verify the mitochondria-targeting ability of T_{CeO₂}. CLSM was used to observe fluorescence, and the results were shown in Fig. 3C and 3D. We observed that the fluorescence of CeO₂-FNL partially overlapped with that of mitochondria, and the possible reason might be the special properties of CeO₂, such as its small size and hydrophobicity [61]. Nonetheless, we found that the fluorescence overlap between T_{CeO₂}-FNL and mitochondria was significantly higher. In addition, fluorescence semiquantitative analysis was obtained by ImageJ, and the results in Fig. 3E and 3F showed that the colocalization ratio between T_{CeO₂}-FNL and mitochondria was higher than that of CeO₂-FNL, suggesting that T_{CeO₂}-FNL had better mitochondria-targeting ability with the help of TPP.

3.5. In vitro antioxidant and anti-inflammatory ability

Inflammatory cytokines and oxidative stress-related factors are rich in psoriasis lesions; therefore, reducing the levels of these factors is critical for treating psoriasis [62,63].

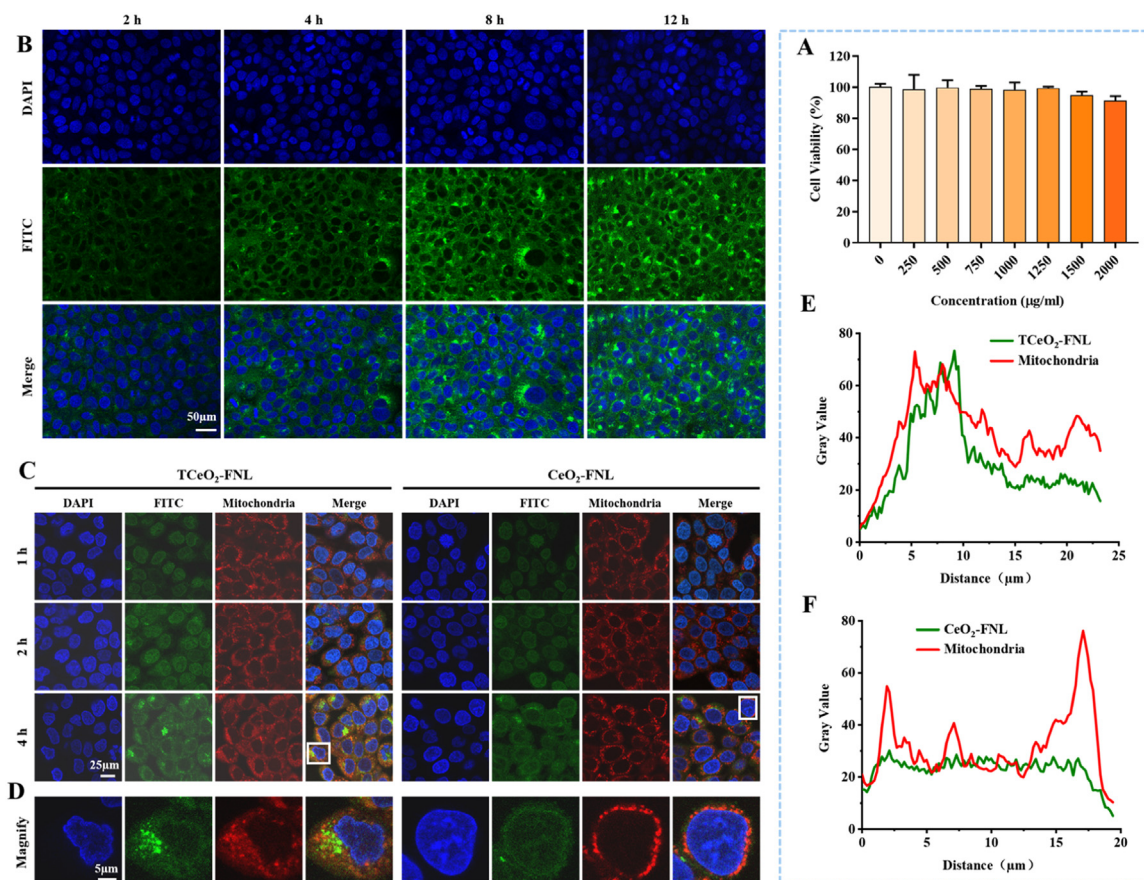


Fig. 3 – Cytotoxicity and cellular uptake of T_{CeO₂}-TRA-FNL and the mitochondria-targeting ability of T_{CeO₂}-FNL/CeO₂-FNL. (A) Viability of HaCat cells after treatment with T_{CeO₂}-TRA-FNL (0–2,000 µg/ml) for 24 h; (B) CLSM images of HaCat cells after treatment with T_{CeO₂}-TRA-FNL for preset time points; (C–D) CLSM images of HaCat cells after treatment with T_{CeO₂}-FNL/CeO₂-FNL for preset time points (red: mitochondria; green: T_{CeO₂}-FNL/CeO₂-FNL); (E–F) Fluorescence intensity analysis of images from Fig. 3D, which shows the colocalization between T_{CeO₂}-FNL/CeO₂-FNL and mitochondria. Data are expressed as the mean ± SD, n = 3, *P < 0.05, **P < 0.01, *P < 0.001.**

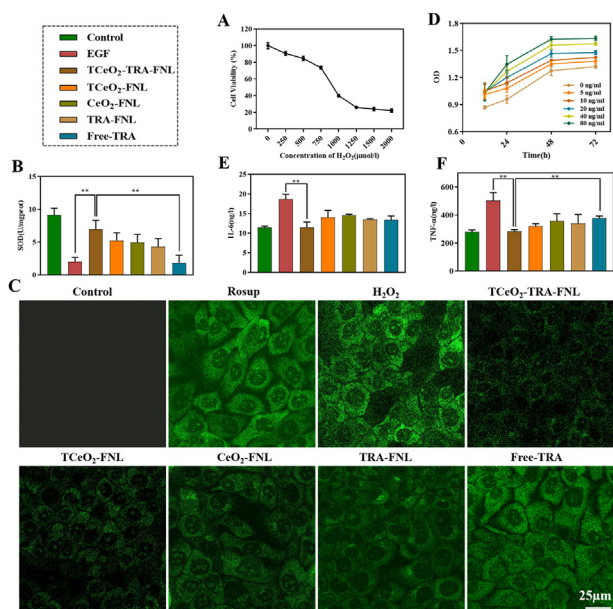


Fig. 4 – In vitro antioxidant and anti-inflammatory ability of TCeO₂-TRA-FNL (the concentrations of TRA and cerium were both 10 µg/ml). (A) Cell viability of HaCat cells after incubation with H₂O₂ (0–2,000 µg/ml) for 24 h. (B) SOD activity of HaCat cells preincubated with different preparations for 4 h before treatment with H₂O₂ for 24 h. (C) Fluorescence images of HaCat cells preincubated with different preparations for 4 h before treatment with H₂O₂ for 24 h, which intuitively show the intracellular ROS. (D) Optical density of HaCat cells after incubation with EGF (0–80 µg/ml) for 12, 24, 48 and 72 h. (E) IL-6 level and (F) TNF-α level of HaCat cells preincubated with EGF (80 ng/ml) for 1 h before treatment with different preparations for 24 h. Data are expressed as the mean ± SD, n = 3, *P < 0.05, **P < 0.01, *P < 0.001.**

We adopted the H₂O₂-induced model to evaluate the antioxidant ability of TCeO₂-TRA-FNL. We found that cell viability was 73.57% ± 1.86% when the concentration of H₂O₂ reached 750 µM, which was appropriate in the following research (Fig. 4A). Acting as an important antioxidant in the body, the enzyme activity of SOD is always used to evaluate antioxidant capacity [47]. The result shown in Fig. 4B implied that compared with H₂O₂-stimulated cells, TCeO₂-TRA-FNL could dramatically increase SOD level and was more efficient than free TRA (P < 0.01). This was mainly because liposomes promoted better intracellular drug delivery and TCeO₂ achieved better clearance of intracellular ROS. It was found that TCeO₂ still effectively scavenged ROS despite being encapsulated into liposomes, which was consistent with a previous study [64]. The 2',7'-dichlorodihydrofluorescein diacetate (DCFH-DA) can be oxidized from a nonfluorescent module to a fluorescent agent, so we use it to detect intracellular ROS [65]. According to the result in Fig. 4C, the green fluorescence intensity increased after treatment with H₂O₂. Nevertheless, the green fluorescence intensity decreased after incubation with different preparations, especially TCeO₂-TRA-FNL, indicating

that the ability of TCeO₂-TRA-FNL to scavenge ROS was better than that of the other preparations. According to the SOD and ROS results, we believed that TCeO₂-TRA-FNL had superior antioxidant capacity *in vitro*.

Epidermal growth factor (EGF) is involved in cell growth, differentiation, and proliferation [66]. So we utilized EGF to imitate the state of psoriasis. First, we examined the incubation concentration and time of EGF by measuring optical density. As shown in Fig. 4D, optical density was positively correlated with concentration and time. We finally chose to incubate with 80 ng/ml EGF for 24 h as the modeling condition. Then, IL-6 and TNF-α levels were estimated. Fig. 4E and 4F showed that after incubation with TCeO₂-TRA-FNL, the two levels decreased compared with that in the EGF treatment group (P < 0.01). The results demonstrated that TCeO₂-TRA-FNL had good anti-inflammatory capability, possibly because TRA exerted an anti-inflammatory effect. In addition, TCeO₂ removed intracellular ROS, which might inhibit some signal pathways to alleviate inflammation [16]. The reason why there was no significant difference between the other groups and the TCeO₂-TRA-FNL group might be that the growth state and density of cells were difficult to control completely consistently, although we assigned the same initial cell count to each group.

3.6. In vivo efficiency against psoriasis

IMQ can serve as an agonist of TLR-7/8 to promote the occurrence of psoriasis [67]. Hence, the ability of TCeO₂-TRA-FNL-Gel to alleviate psoriasis was further investigated in an IMQ-induced model [49]. BALB/c mice were randomly divided into seven groups: (1) control group, (2) IMQ group, (3) TCeO₂-TRA-FNL-Gel group, (4) TCeO₂-FNL-Gel group, (5) CeO₂-FNL-Gel group, (6) TRA-FNL-Gel group, and (7) Free-TRA-Gel group (for each group, n = 5).

The results of weight changes were shown in Fig. 5A. Obviously, no significant changes were observed during the experiment. The skin compliance study was conducted on the skin of mice to evaluate the safety of gels we prepared [49,50]. We found that the application of Free-TRA-Gel for 7 d could cause skin irritation, which confirmed the skin side effect of TRA. In contrast, other groups didn't observe skin irritation (Fig. 5B). This result confirmed that the skin irritation could be reduced by encapsulating TRA into liposomes. The possible reason was that the liposomes could hydrate the epidermis simply by providing lipids to the stratum corneum [68]. The results demonstrated that TCeO₂-TRA-FNL-Gel can increase the safety during application.

Erythema, scaling and skin thickness are prominent features of psoriasis [69]. Hence, we evaluated these indexes in mice during the experiment. Clearly, the mice in the IMQ group had features mentioned above, accompanied by symptoms of redness and swelling and after treatment with different preparations, especially TCeO₂-TRA-FNL-Gel, these symptoms were alleviated to varying degrees, with the reduction in scale and skin thickness being the most evident (Fig. 5F). Meanwhile, the scores of erythema and scale and skin thickness of mice in each treatment group were lower (Fig. 5C and 5E). Compared with the Free-TRA-Gel group, the scores of erythema and scale and skin thickness in the TCeO₂-TRA-

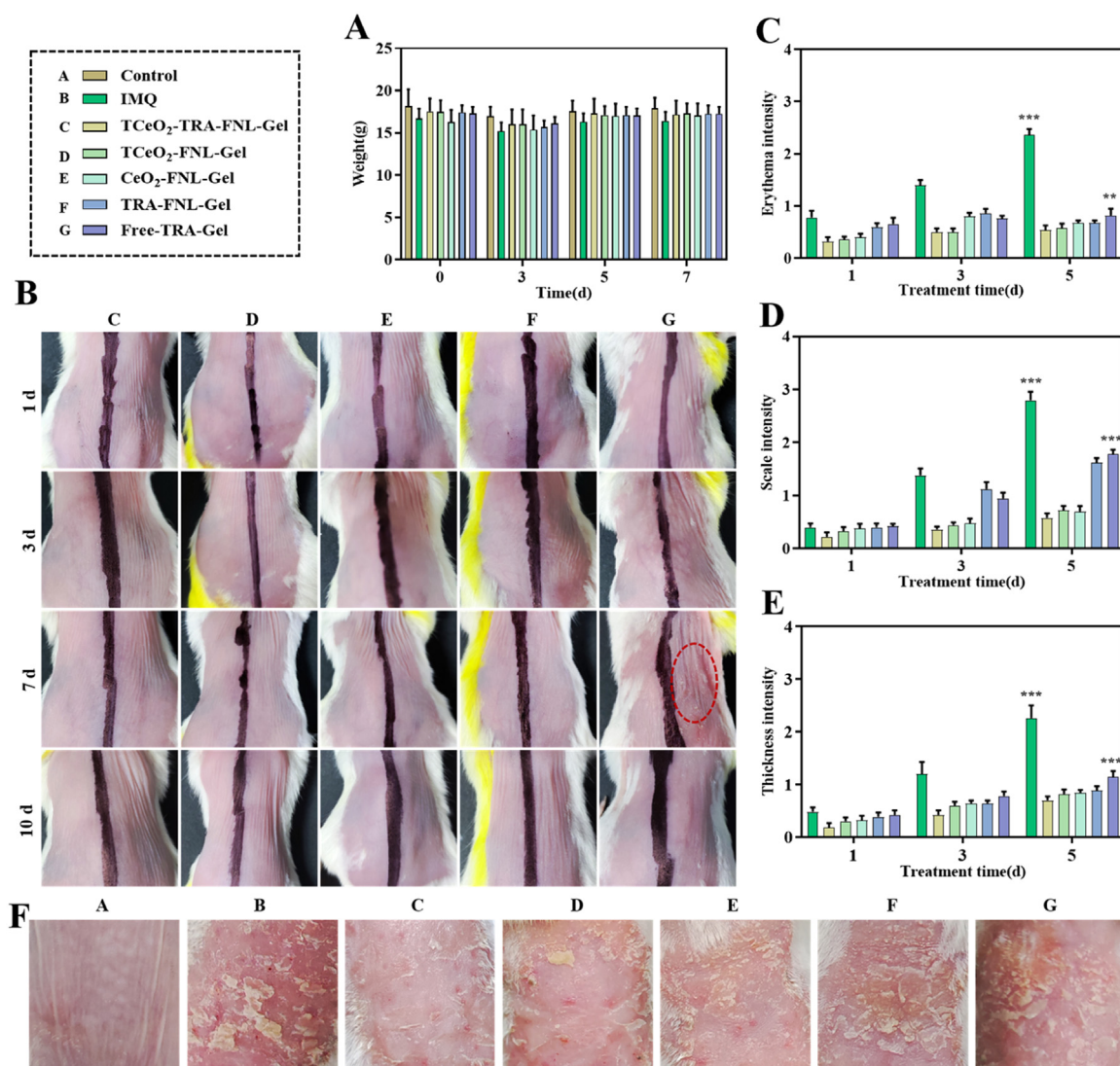


Fig. 5 – In vivo antipsoriasis effect. (A) Weight on Day 0, 3, 5, and 7. (B) Skin irritation result (left: blank Gel, right: different formulations). (C-E) PASI scores of mice. (F) Final images of diseased skin. Data are expressed as the mean \pm SD, $n \geq 3$, * $P < 0.05$, ** $P < 0.01$, * $P < 0.001$, compared with the TCeO₂-TRA-FNL-Gel group.**

FNL-Gel group were significantly reduced (erythema: $P < 0.01$; scale and skin thickness: $P < 0.001$). The limited effect of Free-TRA-Gel might be attributed to the difficult skin delivery and skin irradiation of TRA. The results proved that TCeO₂-TRA-FNL-Gel was beneficial for reducing erythema and scaling and inhibiting skin thickness, indicating that using liposomes to encapsulate TRA could reduce side effects and improve antipsoriasis effects.

It is widely reported that both IL-6 and TNF- α participate in the pathological process of psoriasis, and they are considered indications for disease severity [70,71]. We used ELISA kits to determine the two levels. From the results in Fig. 6A and 6B, we observed that the two levels were downregulated after treatment with TCeO₂-TRA-FNL-Gel, which was significantly different from the IMQ group (IL-6: $P < 0.01$; TNF- α : $P < 0.001$). Meanwhile, a similar tendency appeared when compared with the Free-TRA-Gel group (IL-6: $P < 0.001$; TNF- α : $P < 0.05$). The

results explained that TCeO₂-TRA-FNL-Gel had an enormous capacity to relieve inflammation in psoriatic skin.

ROS levels in skin tissue were detected by immunofluorescence. Fig. 6D showed that the ROS level was significantly higher in IMQ group. After incubation with different gels, the red fluorescence intensity decreased. The red fluorescence intensity in the TCeO₂-TRA-FNL-Gel group was the lowest, which illustrated that ROS nearly recovered to normal levels. Furthermore, Fig. 6C showed that SOD levels were upregulated after incubation with different preparations. The SOD level of the TCeO₂-TRA-FNL-Gel group was higher compared with the IMQ and Free-TRA-Gel groups (IMQ: $P < 0.01$; Free-TRA-Gel: $P < 0.05$). The results above demonstrated that TCeO₂-TRA-FNL-Gel could effectively relieve the oxidative stress of skin.

Excessive proliferation of keratinocytes, infiltration of inflammatory cells, and angiogenesis commonly exist in

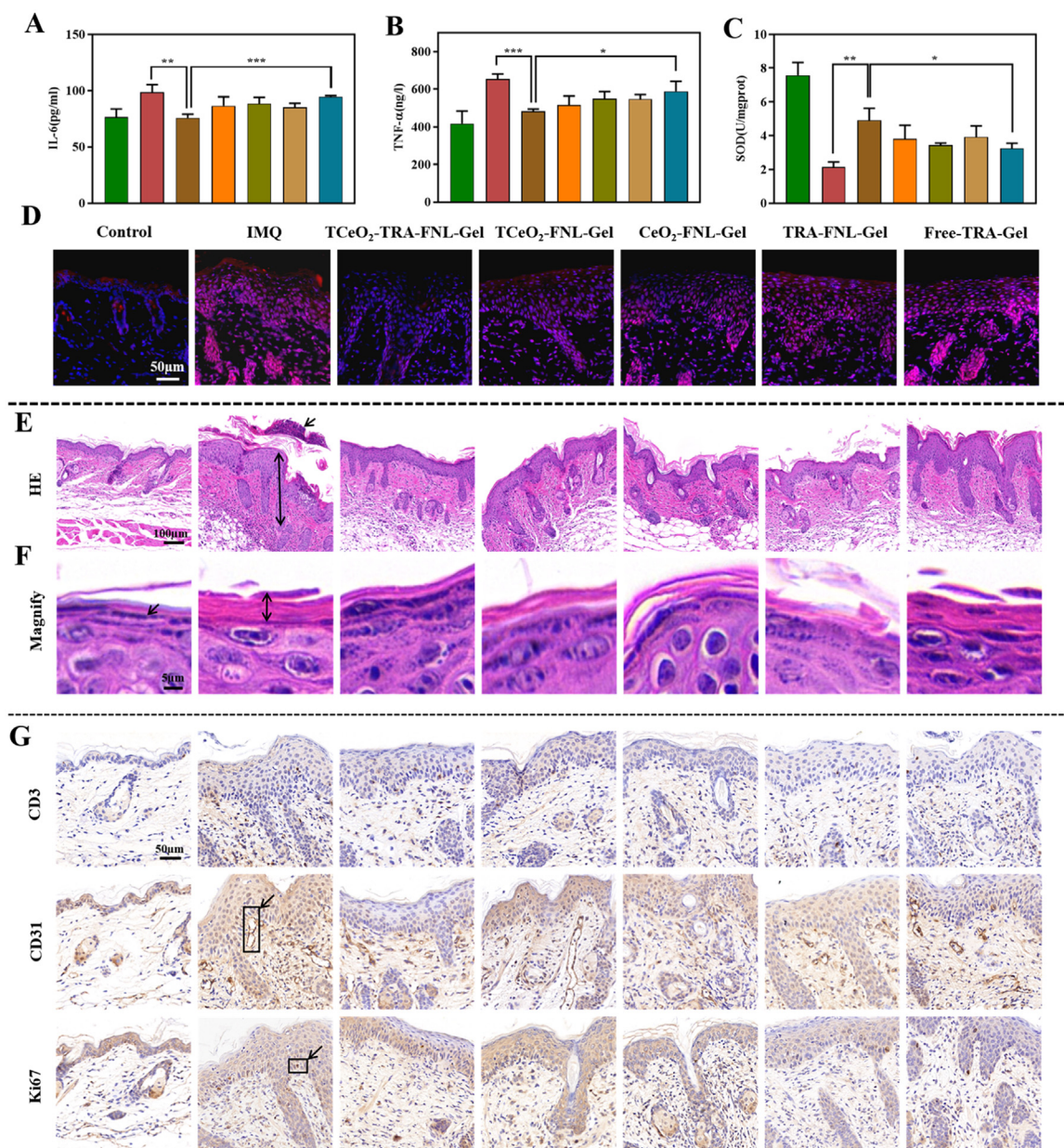


Fig. 6 - In vivo efficiency against psoriasis. (A) IL-6 level and (B) TNF- α level of diseased skin after different treatment. (C) SOD activity of diseased skin after different treatment. (D) Immunofluorescence images of diseased skin (red represents ROS). (E-F) H&E-stained diseased skin with different scale bars. (G) Expression of CD3, CD31 and Ki67. Data are expressed as the mean \pm SD, $n = 3$, * $P < 0.05$, ** $P < 0.01$, * $P < 0.001$, compared with the TcCeO₂-TRA-FNL-Gel group.**

psoriasis [72,73]. They were evaluated by H&E staining and immunohistochemistry.

Skin thickening, elongation of the epidermal crest and Munro microabscess are regarded as the main histopathological features of psoriasis [74]. As shown in Fig. 6E and 6F, we observed similar features in the IMQ group. Furthermore, the stratum corneum thickened and the granular layer disappeared after the application of IMQ. After treatment with Free-TRA-Gel, the thickening of the skin showed little reduction. In contrast, the TcCeO₂-TRA-FNL-Gel group showed an obvious reduction, suggesting that TcCeO₂-TRA-FNL-Gel was

capable of inhibiting the epidermal thickening of psoriatic skin.

CD3, CD31, and Ki67 are vital markers in psoriatic skin, and they are always used to evaluate the severity of psoriasis [49,75,76]. The results in Fig. 6G suggested that the levels of CD3, CD31, and Ki67 in the IMQ group were noticeably increased. However, the three levels were decreased after treatment and the TcCeO₂-TRA-FNL-Gel group had the lowest levels. The downregulation of CD3, CD31, and Ki67 might be attributed to the fact that TcCeO₂-TRA-FNL-Gel relieved inflammation related to excessive proliferation of keratinocytes and angiogenesis.

According to the results mentioned above, we could see that T_{CeO₂}-TRA-FNL-Gel had great antipsoriasis capacity *in vivo*, which could be attributed to the high drug retention and deep penetration of the liposome-gel system, thus maximizing the drug effect. Despite the satisfying results we obtained, some limitations in the *in vivo* study were also demonstrated later. The sample size was insufficient in comparison with clinical trials, and individual differences might affect the statistical analysis.

4. Conclusion

In summary, a nanotransdermal delivery system composed of TRA and TPP-modified CeO₂ nanoparticles, flexible nanoliposomes, and gels (T_{CeO₂}-TRA-FNL-Gel) was successfully constructed, and its effect was evaluated. T_{CeO₂}-TRA-FNL with an appropriate size can excellently enter the keratinocytes after overcoming obstacles from the stratum corneum. The effect is apparent with the release of TRA and T_{CeO₂}. TRA can inhibit the excessive proliferation of keratinocytes, and T_{CeO₂} can target mitochondria to eradicate excessive ROS. The synergistic effect endows the system with satisfying anti-inflammatory and antioxidant capacities. Overall, this nanotransdermal delivery system is a prospective strategy for the treatment of psoriasis. Unfortunately, the insufficient sample size in our study makes it difficult to provide practical guidance for clinical research. In the future, the sample size can be expanded, and other strains of animals can also be used *in vivo* to verify the universality of our system.

Conflicts of Interest

The authors report no conflicts of interest. The authors alone are responsible for the content and writing of this article.

Acknowledgments

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Supplementary materials

Supplementary material associated with this article can be found, in the online version, at doi:10.1016/j.ajps.2023.100846.

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