

Umbilical vessel wall fatty acids after normal and retarded fetal growth

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Abstract

In a prospective observational study, the fatty acid content of human umbilical artery and vein wall phospholipids was determined in fetuses classified according to their change in abdominal circumference during the third trimester. Three groups were identified: appropriate for gestational age (AGA; 24 infants) and small for gestational age (SGA; 38 infants) with normal antenatal growth rate, and SGA with fetal growth retardation (22 infants). The venous linoleic acid (18:2 ω 6) content (expressed as a percentage of the total fatty acids identified) was greater in growth retarded SGA fetuses (3.5 (0.6)%) than in SGA fetuses with a normal growth rate (3.1 (0.5)%) and AGA fetuses (3.0 (0.5)%), whereas the venous contents of eicosatrienoic (20:3 ω 6) and docosa-hexaenoic acid (22:6 ω 3) were lower. In growth retarded SGA fetuses, the venous and arterial 20:3 ω 6 content correlated with the change in abdominal circumference. In SGA fetuses with a normal growth rate, lower contents of arterial 18:2 ω 6 and 22:6 ω 3 were associated with a smaller change in abdominal circumference and birth weight. Different metabolic derangements appear to underly normal and subnormal growth rate in SGA fetuses, suggesting that different strategies of dietary intervention may be required to aid fetal growth and reduce the sequelae of fetal growth retardation.

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The umbilical artery and vein walls contain fatty acids derived from the blood flowing through them. Differences in their fatty acid composition reflect nutritional supply and use during pregnancy.¹ A study of fatty acids in cord blood from appropriate for gestational age (AGA), small for gestational age (SGA), and preterm SGA fetuses showed a higher linoleic acid (18:2 ω 6) and a lower eicosatrienoic acid (20:3 ω 6) content in the SGA fetuses.² This study was limited, however, because birth weight was used to define nutritional status. Birth weight is a poor predictor of perinatal outcome associated with fetal growth retardation as some SGA infants are preceded by a normal growth rate, as judged by ultrasound, and are clinically well nourished.³⁻⁵ Serial ultrasound measurements of abdominal circumference have been shown to predict neonatal wasting associated with fetal growth retardation.⁶

Differences in the placental supply of essential fatty acids may influence fetal growth.

To investigate this, we measured the umbilical artery and vein wall phospholipid fatty acid contents in AGA and SGA infants who had normal and subnormal growth rates.

Methods

SUBJECTS

Eighty four women were recruited after referral to the ultrasound department because of a suspected small fetus during the third trimester of pregnancy. In 60 subjects a SGA fetus was confirmed by an abdominal circumference less than the 10th centile for gestational age.⁷ The remaining 24 fetuses had an abdominal circumference larger than the 10th centile for gestational age and were used as AGA controls. All fetuses were subsequently scanned at intervals of one to two weeks until delivery. All women had at least three scans and delivered after 36 weeks' gestation. All were certain of the date of their last menstrual period and had fetal size confirmed as appropriate by biparietal diameter and femur length at the 18-20 week anomaly scan. Fetal growth rate was quantified by calculating the change in SD score of abdominal circumference between the first scan after recruitment and the last before delivery. This methodology has been described in detail and been shown to identify neonatal morphometry consistent with fetal growth retardation.⁶ Individual SD scores were calculated as the measured abdominal circumference minus the mean abdominal circumference from a reference population at the same gestational age, divided by the SD of abdominal circumference from the reference population. We used our own reference standards.⁸ A change in SD greater than -1.5 was used to classify fetal growth retardation. Informed consent was obtained and the study was approved by the hospital ethics committee.

ANALYSIS OF UMBILICAL ARTERY AND VEIN FATTY ACIDS

Lengths of umbilical cord (5-10 cm) were stored at -20°C for up to six months. Before analysis a 1 cm length of umbilical vein and artery was dissected out, cut lengthwise, and rinsed in ice cold isotonic saline. To extract the lipids, the vessels were homogenised in redistilled chloroform/methanol (2:1, v/v) containing 0.01% w/v butylated hydroxytoluene as antioxidant. After a Folch wash⁹ the phospholipid fraction was separated by thin layer chromatography. Phospholipid fatty acids were transmethylated using 0.5 M sodium methoxide in dry methanol (50°C for 10 min-

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Table 1 Demographic and other details of subjects

	AGA (n=24)	SGA normal growth rate (n=38)	SGA growth retarded (n=22)
No (%) primigravidae	12 (50)	20 (53)	12 (55)
No (%) smokers	9 (38)	5 (13)	10 (45)*
Birth weight (g)	3570 (3200-4200)	2569 (1701-3120)	2277 (1360-2900)
Gestational age at delivery (days)	274 (254-289)	272 (257-292)	269 (253-290)
No (%) caesarean sections for fetal distress	3 (13)	4 (11)	3 (14)
Admissions to NICU†	0	1 (3)	5 (23)*‡
Ponderal index	2.60 (2.38-2.98)	2.45 (2.13-3.42)	2.28 (1.90-2.93)*‡
MAC/HC ratio†	0.31 (0.28-0.35)	0.28 (0.22-0.33)	0.26 (0.23-0.30)*‡
Subscapular skinfold	4.6 (2.9-6.1)	3.1 (2.2-4.4)	2.7 (2.0-4.3)*‡
Triceps skinfold	4.7 (3.1-6.2)	3.1 (2.2-4.4)	2.7 (1.8-4.2)*‡

*p<0.05 versus SGA fetuses with normal growth rate.
 †NICU=neonatal intensive care unit; MAC/HC=mid-arm circumference/head circumference.
 ‡p<0.05 versus AGA fetuses.
 Categorical variables tested by Fisher's exact test or χ^2 test where appropriate.
 Continuous variables assessed by Mann-Whitney U test. Values are median (range).

utes). After extraction of the methyl esters into hexane, their fatty acid composition was determined using a PU4400 gas liquid chromatograph (Phillips Scientific, Cambridge) fitted with a 1.5 m x 4.0 mm ID glass column packed with 10% SP2330 on 100/120 Chromosorb (Supelchem, Essex). Retention times were compared with those of high purity (>99%) standards (Sigma Chemical, Poole, Dorset). Peaks were analysed using an IBM PS/2 microcomputer and Nelson 2600 software (Perkin-Elmer Nelson Systems, Buckinghamshire). Precision was assessed using replicate aliquots of serum extracts. At compositions of 1.9, 7.5, and 36.7% fatty acid the coefficients of variation were 3.9, 0.9, and 0.8% respectively. Individual fatty acids were expressed as a percentage of the total fatty acids identified.

STATISTICAL ANALYSIS

Fatty acid data were analysed using Student's paired (umbilical artery and vein comparisons) and unpaired (fetal groups) *t* test (two tailed). Similar results were obtained when the data were analysed using non-parametric tests. Linear regression and Pearson's correlation coefficients were used to examine relations between the fatty acid content and fetal growth in the SGA groups. Demographic data were analysed using the Mann-Whitney U test and either Fisher's exact test or the χ^2 test.

Results

All 60 SGA fetuses had a birth weight less than the 10th centile for gestational age, whereas all 24 AGA fetuses showed normal fetal growth and had a birth weight greater than the 10th centile for gestational age. Of the SGA fetuses, 22 showed a change in SD score of abdominal circumference greater than -1.5 and were considered to be growth retarded. Thirty eight had a normal growth rate. There was no significant difference in mean gestational age at delivery between the three groups (table 1). The median birth weight was significantly lower in the growth retarded SGA group (1360-2900 g) than for SGA fetuses with normal growth rate (1701-3120 g; p<0.05) and AGA controls (3200-4200 g; p<0.05). The prevalence of maternal smoking was similar in the growth retarded SGA (45%) and AGA (38%) groups, but was significantly lower in the SGA group with normal growth rate (13%; p<0.05) compared with the growth retarded SGA group. In the growth retarded SGA group, the ponderal index, subscapular skinfold, triceps skinfold, and mid-arm circumference/head circumference ratio were significantly lower than the SGA group with a normal growth rate (p<0.05) and AGA controls (p<0.05).

In all three fetal groups, the venous wall content of series 6 polyunsaturated fatty acids was higher and the content of monounsaturated fatty acids and docosahexaenoic acid (22:6 ω 3) were lower than in the arterial wall (table 2). The venous wall 18:2 ω 6 content was greater (3.5 (0.6)%) in the growth retarded SGA group than in the SGA group with normal growth rate (3.1 (0.5)%; p<0.05) and AGA (3.0 (0.5)%; p<0.01) group, and also in the SGA group with normal growth rate compared with the AGA (p<0.01) group. No between group difference in the arterial wall content of 18:2 ω 6 was observed. In growth retarded SGA fetuses, the venous and arterial wall contents of 20:3 ω 6 (2.3 (0.6) and 1.5 (0.3)%) respectively were lower than in the SGA fetuses with normal growth rate (2.6 (0.4)%; p<0.01 and 1.9 (0.3)%; p<0.01) and the AGA fetuses (2.6 (0.5)%; p<0.05 and 2.0 (2.0)%; p<0.001) (table 2).

Table 2 Comparison of fatty acid content of umbilical artery and vein wall phospholipids. Values are mean (SD) percentages

Fatty acid	Umbilical artery			Umbilical vein		
	AGA (n=24)	SGA normal growth rate (n=38)	SGA growth retarded (n=22)	AGA (n=24)	SGA normal growth rate (n=38)	SGA growth retarded (n=22)
14:0 (myristic)	1.1 (0.2)	1.2 (0.3)	1.2 (0.3)	0.9 (0.2)	1.0 (0.2)	0.9 (0.3)
16:0 (palmitic)	20.5 (1.0)	20.9 (2.0)	20.5 (2.2)	22.2 (1.3)	22.8 (2.4)	22.7 (2.9)
16:1 ω 7 (palmitoleic)	3.0 (0.7)	3.2 (0.9)	3.2 (0.5)	2.0 (0.5)	2.2 (0.6)	2.3 (0.8)
18:0 (stearic)	20.8 (0.9)	20.5 (0.9)	20.8 (1.4)	18.8 (1.1)	18.8 (1.2)	19.0 (1.1)
18:1 ω 9 (oleic)	18.3 (2.8)	18.5 (2.8)	19.5 (2.4)	13.7 (1.5)	13.7 (1.5)	14.1 (1.6)
18:2 ω 6 (linoleic)	2.2 (2.2)	2.2 (0.4)	2.1 (0.4)	3.0 (0.5)	3.1 (0.5)**	3.5 (0.6)**†
20:1 ω 9 (eicosenoic)	1.0 (0.3)	1.0 (0.2)	1.1 (0.3)	0.5 (0.2)	0.5 (0.1)	0.6 (0.1)
20:3 ω 6 (eicosatrienoic)	2.0 (2.0)	1.9 (0.3)	1.5 (0.3)***††	2.6 (0.5)	2.6 (0.4)	2.3 (0.6)*††
20:4 ω 6 (arachidonic)	14.7 (1.9)	15.0 (2.1)	14.3 (2.5)	19.1 (1.5)	19.1 (1.7)	19.2 (2.3)
22:4 ω 6 (docosatetraenoic)	3.7 (0.8)	3.5 (0.8)	3.4 (0.7)	5.9 (1.1)	5.5 (1.0)	5.4 (1.2)
24:1 ω 9 (tetracosenoic)	3.9 (0.9)	3.9 (0.8)	4.1 (1.1)	3.1 (1.1)	3.0 (0.8)	2.9 (0.8)
22:6 ω 3 (docosahexaenoic)	6.9 (1.5)	6.9 (1.2)	6.4 (1.2)	6.5 (1.1)	6.1 (1.1)	5.7 (1.3)*
Other‡	1.9 (1.6)	1.5 (0.4)	2.1 (1.4)	1.5 (0.4)	1.6 (0.3)	1.7 (0.3)

*p<0.05 v control; **p<0.01 v control; ***p<0.001 v control.
 †p<0.05 SGA growth retarded v SGA normal growth; ††p<0.01 v SGA growth retarded v SGA normal growth.
 ‡Trace fatty acids, each representing <1% of total.

Table 3 Correlations between fatty acid content and growth indices in the SGA groups

Growth index	Fatty acid							
	18:2 ω 6		20:3 ω 6		22:6 ω 3		20:3 ω 6/18:2 ω 6 ratio	
	Vein	Artery	Vein	Artery	Vein	Artery	Vein	Artery
Growth rate (ASD of AC)								
Growth retarded	0.06	0.18	0.63**	0.68***	0.34	0.20	0.41	0.48*
Normal growth	0.20	0.62***	0.14	0.11	0.26	0.16	0.10	0.34*
Birth weight								
Growth retarded	0.11	0.07	0.52*	0.52*	0.60**	0.44*	0.46*	0.41
Normal growth	0.02	0.31	0.24	0.14	0.27	0.32*	0.13	0.15
Gestational age								
Growth retarded	0.29	0.10	0.11	0.08	0.34	0.20	0.32	0.04
Normal growth	0.11	0.20	0.11	0.18	0.22	0.37*	0.14	0.16
Head circumference								
Growth retarded	0.15	0.19	0.44*	0.35	0.66**	0.53*	0.43*	0.56**
Normal growth	0.02	0.14	0.04	0.02	0.12	0.18	0.30	0.19

Figures refer to Pearson correlation coefficients determined between growth indices and measures of percentage composition.

* $p < 0.05$; ** $p < 0.01$; *** $p < 0.001$.

ASD of AC = change in SD score of abdominal circumference.

There was no apparent relation between the venous wall 18:2 ω 6 content and change in abdominal circumference SD score, birth weight, gestation, or head circumference (table 3). In SGA fetuses with normal growth rate, change in abdominal circumference SD score was strongly associated with the arterial wall 18:2 ω 6 content. In the growth retarded SGA group, but not the SGA group with normal growth rate, the venous wall content of 20:3 ω 6 was positively associated with a change in the abdominal circumference SD score, birth weight, and head circumference. In addition, the arterial wall content of 20:3 ω 6 was positively associated with a change in the abdominal circumference SD score and birth weight in growth retarded SGA fetuses.

The arterial wall 20:3 ω 6/18:2 ω 6 ratio was positively associated with change in abdominal circumference SD score ($r = 0.48$, slope = 2.1, $p < 0.05$), birth weight, and head circumference in the growth retarded SGA group, whereas in the SGA group with normal growth rate this ratio was negatively associated with change in abdominal circumference SD score ($r = 0.34$, slope = -2.1, $p < 0.05$).

The venous wall 20:3 ω 6/18:2 ω 6 ratio was weakly associated with the change in abdominal circumference SD score ($r = 0.41$, $p = 0.057$), but positively associated with birth weight ($r = 0.46$, $p < 0.05$) and head circumference ($r = 0.43$, $p < 0.05$) in growth retarded SGA fetuses. No associations were found with the venous wall ratio in the SGA fetuses with normal growth rate.

The venous and arterial wall 22:6 ω 3 content was positively associated with birth weight and head circumference in the growth retarded SGA group. In SGA fetuses with normal growth rate, the arterial 22:6 ω 3 content was positively associated with birth weight and gestation. No associations were apparent with the venous wall fatty acid content.

Discussion

This study has shown differences in the umbilical vessel wall fatty acid content of SGA infants with growth retardation compared with SGA and AGA infants with normal fetal growth. This suggests a possible impairment of the placental supply of essential fatty acid

derivatives in fetal growth retardation and implies that intervention would necessitate a specific nutritional strategy.

Fetal growth retardation increases the risk of perinatal morbidity and mortality.^{3,4} Although inadequate maternal nutrition is an important factor in the aetiology of fetal growth retardation worldwide,^{10,11} this is not always so.^{2,12} During pregnancy metabolic adjustments occur that can protect the fetus from dietary inadequacy.¹³ Fetal growth retardation may also be caused by an inadequate supply of nutrients or an inability to use them.¹⁴ This is especially pertinent in the third trimester of pregnancy where fetal growth is particularly dependent on the supply of long chain polyunsaturated derivatives of essential fatty acids such as arachidonic and docosahexaenoic acid² and prostaglandins. A reduced supply or use of essential fatty acids may be due to impaired placental perfusion, impaired placental processing of parent essential fatty acids,¹⁵ reduced fetal uptake, or impaired fetal lipolysis.¹⁴

Differences in fatty acid content within and between groups were apparent in both venous and arterial samples. Concentrations of ω 6 fatty acids were higher in veins than arteries whereas levels of monounsaturates were higher in arteries. This is in agreement with earlier observations from normal pregnancies,¹ perhaps reflecting preferential use of polyunsaturated fatty acids by the fetus.

The placenta converts linoleic to arachidonic acid using a series of enzyme catalysed desaturation and elongation reactions.¹⁶ The activity of these enzymes may be compromised by impaired placental development or inhibited by metabolic derangements, such as hypoglycaemia, hypoinsulinaemia, and increased adrenalin and glucocorticoid concentrations, typical of growth retarded SGA fetuses.¹⁷ An impaired ability of the placenta to supply derivatives of 18:2 ω 6, such as long chain polyunsaturated fatty acids, prostaglandins, and leukotrienes, may influence fetal growth. In the growth retarded SGA group, impaired placental processing of linoleic acid may account for the increased venous content of 18:2 ω 6 and decreased content of 20:3 ω 6. The positive correlation in this group between the venous and arterial wall

20:3 ω 6 content and change in abdominal circumference SD score suggests that the impairment may be of graded severity. As the generation of 22:6 ω 3 from α -linolenic acid (18:3 ω 3) involves the same enzyme systems, the positive correlation observed in the same group between the venous wall 22:6 ω 3 content and both birth weight and head circumference further supports an impaired and graded placental enzyme activity in this group. As 22:6 ω 3 is an important lipid of neural tissue,¹⁸ this may influence the incidence of later neurological sequelae in severely growth retarded fetuses.¹⁹

Eicosatrienoic acid is the first desaturation and elongation product during the placental metabolism of linoleic acid to arachidonic acid (20:4 ω 6). Therefore, in umbilical arteries and veins, the ratio of 20:3 ω 6/18:2 ω 6 may provide an index of placental ability to supply metabolically important derivatives of linoleic acid to the fetus. The arterial ratios of 20:3 ω 6/18:2 ω 6 in SGA fetuses with growth retardation and normal growth rate were positively and negatively associated with change in abdominal circumference SD score respectively. In growth retarded SGA fetuses this may be attributed to the positive correlation between the arterial wall 20:3 ω 6 content and change in abdominal circumference SD score. In SGA fetuses with normal growth rate a lower content of arterial wall (but not venous wall) 18:2 ω 6 was associated with a reduced change in abdominal circumference SD score and may reflect a marginal deficiency in essential fatty acids in this group. This observation in SGA fetuses with normal growth rate is further supported by an association between lower arterial wall (but not venous wall) 22:6 ω 3 content and lower birth weight.

Maternal essential fatty acids enter the placental circulation along a concentration gradient.²⁰ Fetal growth may then be dependent on adequate placental synthesis of long chain derivatives from these essential fatty acids. During the third trimester the major fatty acids that accrue in fetal tissue are long chain derivatives of essential fatty acids, the accumulation of linoleic acid being minimal (2–3%).²¹ Postnatal tissue is characterised by a substantial accretion of linoleic acid, increases in long chain derivatives not occurring for several weeks. Different metabolic derangements underlying the nutritional status associated with normal growth and growth retardation in SGA fetuses may have marked implications for any potential dietary intervention. Even though the maternal essential fatty acid status may be adequate in cases of growth retardation, direct supplementation of long chain derivatives of

essential fatty acids may circumvent any impairment in the placental supply of these fatty acids, thereby assist fetal growth, and prevent some of the associated physiological and neurological sequelae.

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- Hornstra G, van Houwelingen AC, Simonis M, Gerrard JM. Fatty acid composition of umbilical arteries and veins; possible implications for the fetal EFA-status. *Lipids* 1989; 24: 511–7.
- Vilbergsson G, Samsoie G, Wennergren M, Karlsson K. Essential fatty acids in pregnancies complicated by intra-uterine growth retardation. *Int J Gynecol Obstet* 1991; 36: 277–86.
- Patterson RM, Pouliot RN. Neonatal morphometrics and perinatal outcome: who is growth retarded? *Am J Obstet Gynecol* 1987; 157: 691–3.
- Villar J, de Onis M, Kestler E, Bolanos E, Cerezo R, Bernedes H. The differential neonatal morbidity of the intra-uterine growth retardation syndrome. *Am J Obstet Gynecol* 1990; 163: 151–7.
- Chang TC, Robson SC, Boys RJ, Spencer JAD. Prediction of the small for gestational age infant: which ultrasonic measurement is best? *Obstet Gynecol* 1992; 80: 1030–8.
- Chang TC, Robson SC, Spencer JAD, Gallivan S. Identification of fetal growth retardation: comparison of Doppler waveform indices and serial ultrasound measurements of abdominal circumference and fetal weight. *Obstet Gynecol* 1993; 82: 230–6.
- Deter RL, HARRIST RB, Hadlock FP, Carpenter RJ. Fetal head and abdominal circumferences: II. A critical re-evaluation of the relationship to menstrual age. *Journal of Clinical Ultrasound* 1982; 10: 365–72.
- Gallivan S, Robson SC, Chang TC, Vaughan J, Spencer JAD. An investigation of fetal growth using serial ultrasound data. *Ultrasound in Obstetrics and Gynecology* 1993; 3: 109–14.
- Folch J, Lees M, Sloane-Stanley GH. A simple method for the isolation and purification of total lipids from animal tissues. *J Biol Chem* 1957; 226: 497–509.
- Doyle W, Crawford MA, Laurance BM, Drury P. Dietary survey during pregnancy in a low socio-economic group. *Human Nutrition: Applied Nutrition* 1982; 36A: 95–106.
- Hornstra G, van der Schouw YT, Bulstra-Ramakers MT, Huisjes HJ. Biochemical EFA status of mothers and their neonates after normal pregnancy. *Early Hum Dev* 1990; 24: 239–48.
- Ministry of Agriculture, Fisheries and Foods. *Household food consumption and expenditure. Annual report of the National Food Survey Committee*. London: HMSO, 1979.
- Keirse MJNC. Epidemiology and aetiology of the growth retarded baby. *Clin Obstet Gynecol* 1984; 11: 415–36.
- Olegard R, Gustafsson A, Kjellmer I, Victorin L. Nutrition in low birth-weight infants. II. Lipolysis and free fatty acid elimination after intravenous administration of fat emulsion. *Acta Paediatr Scand* 1975; 64: 745–51.
- Diaz M, Leal C, Cajal JR, et al. Cord blood lipoprotein-cholesterol: relationship to birth-weight and gestational age of newborns. *Metabolism* 1989; 38: 435–8.
- Zimmerman T, Winkler L, Moller U, Schubert H, Goetze E. Synthesis of arachidonic acid in human placenta in vitro. *Biol Neonate* 1979; 35: 209–12.
- Brenner RR. Nutritional and hormonal factors influencing desaturation of essential fatty acids. In: Holman RT, ed. *Progress in lipid research*. New York: Pergamon Press, 1981: 41–7.
- Crawford MA, Costeloe K, Doyle W, Leighfield MJ, Lennon EA, Meadows N. Potential diagnostic value of the umbilical artery as a definition of neural fatty acid status of the fetus during its growth: the umbilical artery as a diagnostic tool. *Biochem Soc Trans* 1990; 18: 761–6.
- Soothill PW, Ajayi RA, Campbell S, et al. Relationship between fetal acidemia at cordocentesis and subsequent neurodevelopment. *Ultrasound in Obstetrics and Gynecology* 1992; 2: 80–3.
- Thomas CR. Placental transfer of non-esterified fatty acids in normal and diabetic pregnancy. *Biol Neonate* 1987; 51: 94–101.
- Clandinin MT, Chappell JE, Heim T. Do low weight infants require nutrition with long chain elongation-desaturation products of essential fatty acids? *Prog Lipid Res* 1981; 20: 901–4.