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Bubble cloud characteristics and ablation efficiency in dualfrequency intrinsic threshold histotripsy

Connor Edsall^{1,*} \bullet , Laura Huynh², Timothy L Hall³ \bullet and Eli Vlaisavlievich^{1,4}

- ¹ Department of Biomedical Engineering and Mechanics, Virginia Polytechnic Institute and State University, 325 Stanger St., Blacksburg, VA, 24061, United States of America
- ² Department of Materials Science and Engineering, Virginia Polytechnic Institute and State University, 445 Old Turner St., Blacksburg, VA 24061, United States of America
- ³ Biomedical Engineering, University of Michigan, Carl A. Gerstacker Building, 2200 Bonisteel Blvd, Ann Arbor, MI 48109-2133, United States of America
- ⁴ ICTAS Center for Engineered Health, Virginia Polytechnic Institute and State University, 325 Stanger St., Blacksburg, VA, 24061, United States of America
- Author to whom any correspondence should be addressed.

E-mail: cwedsall@vt.edu

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Abstract

Histotripsy is a non-thermal focused ultrasound ablation method that destroys tissue through the generation and activity of acoustic cavitation bubble clouds. Intrinsic threshold histotripsy uses single-cycle pulses to generate bubble clouds when the dominant negative pressure phase exceeds an intrinsic threshold of∼25–30 MPa. The ablation efficiency is dependent upon the size and density of bubbles within the bubble cloud. This work investigates the effects of dual-frequency pulsing schemes on the bubble cloud behavior and ablation efficiency in intrinsic threshold histotripsy. A modular 500 kHz:3 MHz histotripsy transducer treated agarose phantoms using dual-frequency histotripsy pulses with a 1:1 pressure ratio from 500 kHz and 3 MHz frequency elements and varying arrival times for the 3 MHz pulse relative to the arrival of the 500 kHz pulse (-100 ns, 0 ns, and $+100$ ns). Highspeed optical imaging captured cavitation effects to characterize bubble cloud and individual bubble dynamics. The effects of dual-frequency pulsing on lesion formation and ablation efficiency were also investigated in red blood cell (RBC) phantoms. Results showed that the single bubble and bubble cloud size for dual-frequency cases were intermediate to published results for the component singlefrequencies of 500 kHz and 3 MHz. Additionally, bubble cloud size and dynamics were shown to be altered by the arrival time of the 3 MHz pulse with respect to the 500 kHz pulse, with more uniform cloud expansion and collapse observed for early (-100 ns) arrival. Finally, RBC phantom experiments showed that dual-frequency exposures were capable of generating precise lesions with smaller areas and higher ablation efficiencies than previously published results for 500 kHz or 3 MHz. Overall, results demonstrate dual-frequency histotripsy's ability to modulate bubble cloud size and dynamics can be leveraged to produce precise lesions at higher ablation efficiencies than previously observed for single-frequency pulsing.

Introduction

Histotripsy is a non-invasive and non-thermal focused ultrasound ablation method currently being developed for the treatment of cancer and other clinical applications (Bader *et al* [2019,](#page-12-0) Xu *et al* [2021](#page-13-0), Vidal-Jove *et al* [2022](#page-13-0)). Histotripsy mechanically destroys tissue through the precise control of acoustic cavitation (Vlaisavljevich et al [2013b](#page-13-0), Vlaisavljevich et al [2016,](#page-13-0) , Bader et al [2019](#page-12-0)). High-pressure (>10 MPa), short-duration (<20 μ s) pulses applied at very low duty cycles (<1%) produce histotripsy 'bubble clouds' (Xu et al [2004,](#page-13-0) Parsons et al [2006](#page-12-0),

Roberts et al [2006](#page-13-0)). The high stress and strain induced in the target tissue by the rapid expansion and collapse of the individual bubbles comprising the bubble cloud result in the destruction of cells and the surrounding extracellular matrix (Hall et al [2009](#page-12-0), Vlaisavljevich et al [2013b](#page-13-0), Vlaisavljevich et al [2016](#page-13-0)). Complete ablation of the focal volume following repeated pulsing is characterized by well-defined lesions closely matching the size and location of the bubble cloud (Roberts et al [2014,](#page-13-0) Zhang et al [2015,](#page-13-0) Vlaisavljevich et al [2017](#page-13-0)), with the necessary number of pulses dependent on the tissue's mechanical properties and bubble cloud characteristics (Vlaisavljevich et al [2015c,](#page-13-0) Mancia et al [2017,](#page-12-0) Vlaisavljevich et al [2017](#page-13-0), Edsall et al [2021a](#page-12-0)).

Intrinsic threshold histotripsy is one of three currently established histotripsy methods, along with shock scattering histotripsy and boiling histotripsy (Bader et al [2019](#page-12-0)). Intrinsic threshold histotripsy distinctly forms acoustic cavitation directly from the dominant, high negative-pressure phase of a single pulse (\leq 2 cycles) with generation dependent upon the duration and amplitude of the negative pressure as well as the target medium's material properties (Maxwell et al [2013,](#page-12-0) Vlaisavljevich et al [2015b](#page-13-0)). The intrinsic threshold for cavitation generation has consistently been shown to be∼25–30 MPa for water-based soft tissues for histotripsy pulse frequencies ranging from 345 kHz to 3 MHz (Maxwell et al [2013](#page-12-0), Vlaisavljevich et al [2015b,](#page-13-0) Edsall et al [2021a](#page-12-0)). Acoustic cavitation near the threshold presents as individual bubbles with the characteristic histotripsy bubble cloud forming with increasing pressure (Vlaisavljevich et al [2015b](#page-13-0), Vlaisavljevich et al [2017](#page-13-0)). The bubble cloud's dimensions closely match the volume predicted by the focal region above the intrinsic threshold (Lin et al [2014b](#page-12-0)).

Pulsing parameters have experimentally been shown to affect bubble cloud characteristics in intrinsic threshold histotripsy. For instance, prior work has shown that decreasing transducer f-number increases the bubble density within the bubble cloud, with a corresponding increase in ablation efficiency (Vlaisavljevich et al [2017](#page-13-0)). Studies have also shown that decreasing pulse frequency leads to increased individual bubble expansion and bubble cloud size due to the longer duration of the suprathreshold negative pressure (Vlaisavljevich et al [2015b](#page-13-0), Mancia et al [2017](#page-12-0)). As a result of these changes in bubble cloud characteristics, lower frequency correlates with an increased ablation efficiency, which was hypothesized to be due to increased bubble expansion inducing higher strain on the targeted tissue (Edsall et al [2021a](#page-12-0)). However, lower-frequency pulsing also produces clouds with reduced bubble density (Edsall *et al* [2021a](#page-12-0)), suggesting that further increases in ablation efficiency could be achieved using methods capable of simultaneously increasing bubble density as well as individual bubble expansion.

Dual-frequency histotripsy pulsing strategies have previously been developed to modulate bubble cloud size and individual bubble expansion (Lin et al [2014a](#page-12-0), Vlaisavljevich et al [2015a](#page-13-0)). These prior studies have shown that the size of the bubble cloud and individual bubble size can be modulated using dual-frequency pulsing strategies by changing the percentage of pressure coming from the respective frequencies (Lin et al [2014a,](#page-12-0) Lin et al [2015](#page-12-0), Vlaisavljevich et al [2015c](#page-13-0)). These prior works have also shown that the size of a single histotripsy bubble and the corresponding region of tissue ablation were not affected by the arrival times of the respective frequencies as long as the peak negative pressures $(p-)$ remained above the intrinsic threshold, which was shown to occur for cases in which the high frequency 3 MHz pulse arrived within 0.15 μ s relative to the low frequency 500 kHz pulse (Lin et al [2014a](#page-12-0)). However, these prior studies did not characterize the bubble cloud characteristics and behavior in dual-frequency histotripsy, nor did they assess the effects of the pulse arrival time on the bubble cloud density and ablation efficiency for higher peak negative pressures that are commonly used in histotripsy to generate bubble clouds instead of individual bubbles, which is critical to understanding and optimizing dual-frequency histotripsy pulsing methods for specific clinical applications.

Using the same modular transducer and similar methods as our prior investigation of single-frequency (Edsall et al [2021a](#page-12-0)), this study investigates the effects of dual-frequency histotripsy pulsing on the characteristics, behavior, and ablation efficiency of histotripsy bubble clouds generated inside agarose tissue phantoms. This study also compares the effects of 3 MHz pulse arrival modulation relative to the 500 kHz pulse arrival on the bubble cloud characteristics, behavior, and ablation efficiency. Cavitation activity was captured using two modes of high-speed optical imaging to compare the single bubble and bubble cloud dimensions, behavior, and the resulting bubble cloud ablation efficiency for dual-frequency (500 kHz:3 MHz) cases. This study's primary hypothesis is that dual-frequency pulsing can modulate the bubble cloud characteristics and ablation efficiency in intrinsic threshold histotripsy distinctly from its component frequencies with changes in high frequency (3 MHz) arrival time imparting additional degrees of frequency-specific characteristics on cavitation nucleation and dynamics. Based on our prior investigation of the effects of single-frequencies, we hypothesized that bubble cloud size and resulting lesion dimensions would be intermediate to the single-frequency cases. We further hypothesized that dual-frequency histotripsy would result in an increased ablation efficiency (evaluated as the number of pulses required to remove the area of the bubble cloud) as these bubble clouds should be denser bubble clouds(due to higher frequency nucleation) comprised of bubbles with larger expansion (due to the elongated tensile phase of lower frequency component). Overall, the findings of this study will increase our

Figure 1. Dual-frequency histotripsy experimental schematic. (A) Optical imaging using a FLIR camera and high-speed strobe captured cavitation nucleation and red blood cell (RBC) ablation inside agarose tissue phantoms exposed to histotripsy pulses applied by the 32-element modular transducer(B)with 500 kHz (blue) and 3 MHz (yellow) arranged in an alternating pattern. High-speed imaging using a Phantom high-speed camera captured single bubble and bubble cloud dynamics. (C) Single bubbles were captured in greyscale videos and traced in binary to measure the bubble expansion and collapse. (D) For bubble cloud behavior, each bubble cloud was captured in greyscale, converted to binary, and the centroid of the cloud located. (E) The cloud area of growth and collapse was then measured in total as well as before (front) and after (rear) the centroid of the cloud in the axial dimension (green dotted line).

understanding of how dual-frequency pulsing techniques can be used to modulate bubble cloud characteristics and resulting ablative effects in histotripsy.

Methods

Ultrasound pulse generation

To directly compare the results of this study with our prior work investigating the effects of frequency on histotripsy cavitation and behavior (Edsall et al [2021a](#page-12-0)), all experiments used the same custom-designed, modular histotripsy transducer with a geometric focus of 75 mm, an aperture size of 120.5 mm, and a f-number of 0.62 (figure 1(B)). Sixteen 500 kHz and sixteen 3 MHz elements(alternating every other element) populated the transducer scaffold which was comprised of three concentric rings of six, twelve, and fourteen ports (figure 1(B)). The transducer was positioned horizontally in a tank of DI water which was degassed to an oxygen concentration <28% O2. A custom, high-voltage amplifier populated with interchangeable amplifier boards matched to the frequency (500 kHz or 3 MHz) of the respective elements produced short therapy pulses of <2 cycles. Afield-programmable gate array (FPGA) board (Altera DE0-Nano Terasic Technology, Dover, DE, USA), programmed by a custom MATLAB (The MathWorks, Natick, MA, USA) code, synchronized the pulse arrivals by controlling the phase-delay and the charge-times of each individual element for each frequency. For each of these experiments, a MATLAB code coordinated the motion of a three-axis positioning system to orient agarose tissue phantoms or hydrophones with the pulsing of the transducer for the respective experiments (figure $1(A)$).

Hydrophone focal pressure calibration

A fiberoptic probe hydrophone (FOPH), cross-calibrated with a high-sensitivity reference rod hydrophone (HNR-0500, Onda Corporation, Sunnyvale, CA, USA), measured the dual-frequency pulsing, which consisted of a 1:1 pressure amplitude of 500 kHz:3 MHz elements. The 500 kHz and 3 MHz peaks were first aligned such that the p− of each frequency arrived at the focal point concurrently (0 ns offset) with the p− resulting from the linear addition of the p− of the individual 500 kHz and 3 MHz waveforms(figure [2](#page-3-0)). The FOPH was then also used to determine the necessary phase delay for each element to have the 3 MHz waveform arrive ± 100 ns

relative to the peak of the 500 kHz (figure 2). The FOPH directly measured the waveforms in degassed water up to peak negative pressures (p−) of $p = 18$ MPa for each arrival time. Higher focal pressures, up to ~36 MPa, were estimated by summing the outputs from the transducer sub-apertures of sixteen elements (eight 500 kHz and eight 3 MHz) to avoid cavitation damage to the hydrophone. An oscilloscope (TBS2000 series, Tektronix, Beaverton, OR, USA) collected waveforms at a sample rate of 500 MS s⁻¹. The waveforms were averaged over 512 pulses to minimize signal noise and recorded to MATLAB.

Agarose gel phantom preparation

Agarose preparation followed previously published methods (Vlaisavljevich et al [2014](#page-13-0), Edsall et al [2021a,](#page-12-0) [2021b](#page-12-0)). 1% (w/v) agarose tissue phantoms, shown in prior work to have a Young's modulus of∼21.7 kPa (Vlaisavljevich et al [2015c](#page-13-0)), were used to replicate soft human tissue's mechanical and viscoelastic properties(Yamada [1973](#page-13-0), Duck [1990](#page-12-0)). To make the tissue phantom, the agarose gel was prepared by mixing 0.5% agarose powder(Type VII-A, Sigma Aldrich Corporation, St. Louis, MO, USA) with 99.5% of 0.45% saline at room temperature. Saline was prepared from a stock 4.5% saline and DI water degassed using a Portable Water Degasser (DS-50, FUS Instruments, Toronto, ON, CAN). The mixture was heated in a microwave until boiling and then stirred to fully dissolve the agarose. Next, the sample was repeatedly heated to boiling and stirred to produce flash boiling to release dissolved gas from the mixture until the solution was fully degassed and 50% of the volume remained, leaving a degassed 1% (w/v) agarose gel mixture. The mixture was placed under a partial vacuum (∼33.62 kPa, absolute) for twenty minutes to remove the remaining gas and minimize regassing as the agarose solution cooled. The pressure was decreased to∼16.75 kPa for an additional five minutes to force any remaining gas from solution. Once the temperature of the agarose dropped to 40 °C, a serological pipette was used to slowly inject 120 ml of the gel down the wall of a rectangular silicone mold into a custom-designed polylactide (PLA) phantom holder frame. The silicone mold containing the gel was then stored in a refrigerator for one hour to solidify. Each experiment was performed within two hours of the gel creation to ensure a consistent agarose concentration and the degassed state of the gel during testing.

Optical imaging for characterization of bubble cloud size

Optical imaging using a high-speed camera (FLIR Blackfly S monochrome, BFS-U3-32S4M-C 3.2 MP, 118 FPS, Sony IMX252, Mono, FLIR Integrated Imaging Solutions, Richmond, BC, Canada) and a 100 mm F2.8 Macro lens(Tokina AT-X Pro, Kenko Tokina Co., LTD, Tokyo, Japan) monitored cavitation events and characterized the bubble clouds. Captured images using this combination had a resolution of 3.25 μ m per pixel. A custom high-speed LED strobe light backlit each sample with the exposure duration kept as low as possible (1 μ s) to minimize the motion blur of the expanding bubbles. Timing accuracy of the pulse delivery, camera and strobe timings, and all delays were accurate to the 10 ns clock speed of the FPGA. The FPGA in the amplifier box triggered the strobe and camera such that all exposures were centered at a 3 μ s delay after the arrival of the peak negative pulse. This delay was optimized to detect clearly visible bubbles while minimizing bubble overlap within the bubble cloud. A 1 Hz pulse repetition frequency (PRF) was used for bubble cloud characterization experiments to limit the memory effect on subsequent pulses (Maxwell et al [2013](#page-12-0)). Images captured very early in the life of the bubble cloud at a low PRF minimized memory effect between pulses. The optical images for each sample were analyzed using a custom MATLAB script that identified bubble clouds for dimensional measurement by converting the captured grayscale image into and negative binary image based on an intensity

threshold determined by the difference from the background intensity following previously published methods (Maxwell et al [2013](#page-12-0)).

The effect of the 3 MHz arrival in dual-frequency histotripsy on bubble cloud size was compared by capturing bubble clouds for each condition (−100, 0, +100 ns). For each dual-frequency condition, 100 histotripsy pulses were applied to agarose phantoms for each frequency and pressure level combination. The bubble cloud length was detected as the distance from the closest edge of the nearest bubble to the transducer to the furthest edge of the furthest bubble from the transducer. Similarly, the bubble cloud height was defined as the distance from the topmost edge of the bubble highest in the positive elevational direction to the bottom edge of the lowest bubble in the negative elevational direction. These dimensions were then used to calculate the mean cloud area using $A = \left(\frac{\text{AxialDimension}}{2}\right) \left(\frac{\text{ElevationalDimension}}{2}\right) \pi$. ElevationalDimension 2

Single bubble and bubble cloud dynamics

A high-speed camera (Phantom High-Speed TMX-6410, Mono, 18.5 μpixel, Vision Research Inc., Wayne, NJ, USA) coupled to an 85 mm f/2.8 1–5x super-macro lens(Creator, Mitakon Zhongyi, Liaoning Province, CHN) monitored the growth and collapse behavior of single bubbles and bubble clouds in 1% agarose phantoms. The phantoms were backlit by steady LED-array light source (LT-V9-15, GSVITEC, Bad Soden-Salmünster, DEU). The Phantom high-speed camera was set to image at 200 000 frames per second capturing 1 frame every 5 μ s from −1 μ s before until 274 μ s after p − arrival. The maximum resolution at this speed was 512 × 256 pixels giving a per pixel resolution of \sim 5 µm.

For single bubble dynamics, 50 pulses were applied to the phantom at 1 Hz PRF at pressures just above their respective threshold each 3 MHz arrival time (−100, 0, +100 ns). These ranges were selected for analysis as they fell within a narrow range of delays(−150 to 150 ns) previously reported to have minimal effect on cavitation threshold and respective lesion size from single bubbles (Lin *et al* [2014a](#page-12-0)). The high-speed camera collected and saved 56 frames per pulse. These frames were converted to an AVI file tracking the single bubble growth and collapse. A custom MATLAB script traced the boundary of the bubble through each frame of the AVI file (figure [1](#page-2-0)(C)), marked the diameter in the axial and elevation directions, and reported the mean bubble radius for the bubble that would approximate a circular area at each $5 \mu s$ interval for each generated bubble. These dimensions were averaged for each 5 μ s time point across for the first 25 single bubble (excluding instances of 0 bubbles and >1 bubble) producing the mean radius and standard deviation. The mean maximum radius, R_{max} , and the mean time, t_{max} , to reach that maximum bubble size were recorded. Mean collapse time the bubble before rebounding was also recorded (t_{c1}). Lastly, the final time point of collapse was recorded, t_{c2} , was recorded after a small rebound of each bubble was observed.

For bubble cloud dynamics, 30 pulses were applied to the phantom at 1 Hz PRF at 36 MPa for each 3 MHz arrival time (−100, 0, +100 ns). The high-speed camera collected and saved 56 frames per pulse. These frames were converted to greyscale JPEG images tracking the growth and collapse of the bubble clouds generated by each pulse (figure [1](#page-2-0)(D)). A custom MATLAB program created binary images of the greyscale cloud using thresholding to identify bubbles using prior methods (Edsall et al [2021a](#page-12-0)) similar to those listed in the characterization of bubble cloud size in Optical Imaging for Characterization of Bubble Cloud Size above. The centroid of the cloud captured in frame 2 (4 μ s) was located by MATLAB and designated as the center point to divide all subsequent clouds into front and back regions(figure [1](#page-2-0)(E)). The code then recorded the total, front, and rear areas of each cloud for each 5 μ s time point through growth and collapse. The area was averaged for each 5 μ s time point across for all 30 bubble clouds producing the mean total, front, and rear areas, and the respective standard deviations for each area at each time point.

Red blood cell phantom creation

Red blood cell (RBC) phantoms were created consisting of three layers of agarose, with the middle layer containing 5% (v/v) red blood cells following previously published methods (Maxwell et al [2010](#page-12-0), Vlaisavljevich et al [2013a](#page-13-0), Edsall et al [2021a](#page-12-0)). A 1% agarose mixture was generated using the agarose preparation above. Fresh porcine blood was obtained from subjects in an unrelated study and added to an anticoagulant solution of Citrate Phosphate Dextrose Anticoagulant (CPD, Sigma Aldrich Corporation, St. Louis, MO, USA), with a CPDto-blood ratio of 1:9 ml. Whole blood was separated by centrifugation at 3000 rpm for 10 min. The plasma and white buffy coat were removed, and the RBCs were saved for addition to the phantom. The RBC phantom was created using an initial layer of agarose poured into the tissue phantom holders at 45 °C. The housing was placed in a refrigerator at 4 °C to allow the base agarose layer to cool and solidify. The remaining solution was kept in the vacuum chamber until it had cooled to 38 °C. 9.5 ml of the respective agarose solutions was combined with the RBCs (5% v/v) by gentle inversion and poured on top of the chilled solidified agarose layer. The liquid RBC-agarose coated the entire surface before the excess solution was poured out leaving behind a thin layer of the RBC-agarose solution. The phantom was replaced in the refrigerator for five minutes. Once the thin

RBC-agarose layer was solidified, the remaining agarose solution without RBCs was poured on top of the first two layers to fill the silicone mold and replaced in the refrigerator for one hour to fully solidify producing a thin layer of RBCs suspended in the center of the clear 1% agarose phantom.

Red blood cell ablation

The histotripsy ablation efficiency was examined and compared using agarose tissue phantoms with embedded RBC layers. RBC ablation can be directly visualized as successive pulses lyse the red blood cells and turn the embedded layer translucent (Maxwell et al [2010,](#page-12-0) Lin et al [2014b](#page-12-0)). Lesion visualized in RBC phantoms have been shown to closely resemble histology of lesions generated in tissue in previous studies (Maxwell et al [2010](#page-12-0), Lin et al [2014b](#page-12-0)). The RBC phantom was oriented at the transducer's focus with the embedded RBC layer parallel to the direction of ultrasound propagation. 2000 histotripsy pulses were applied at $p = 31$ MPa to a single point in the RBC layer at 1 Hz PRF. High-speed optical imaging using the FLIR camera and high-speed strobe captured the bubble cloud formed from each pulse and an image of the resulting ablated region between pulses. The bubble cloud images were separated from the ablation images and the ablated areas resulting from each pulse were measured using image thresholding by a custom MATLAB script. Finally, to normalize this data, the ablated area of each image for each frame was divided by the respective mean area of the bubble cloud formed in the RBC phantom for each 3 MHz arrival time.

Results

Single bubble dynamics

High-speed optical imaging captured the growth and collapse of the bubble cloud at 5 μ s intervals from 1 μ s before to 109 μ s after pulse arrival (figure 3). Bubbles are seen to grow and collapse nearly symmetrically with little translation. 20 individual bubbles' growth and collapse events were captured for each 3 MHz arrival time resulting in individual radius versus time curves for -100 ns, 0 ns, and $+100$ ns (figures $4(A)$ $4(A)$ –(C)). All individual bubble's ($n = 20$) growths and collapses were averaged at each time point to produce the mean radius versus time plot in figure [4](#page-6-0)(D) showing each bubble as it reached its maximum radius (R_{max}) at a time (t_{max}) and tracking its size until it reached total collapse (t_{c1}, t_{c2}). The mean R_{max} for each 3 MHz arrival time was found to be equal to 260.2 \pm 16.3 μ m, 289.5 \pm 19.9 μ m, and 241.87 \pm 25.2 μ m for -100 ns, 0 ns, and $+100$ ns respectively. The respective t_{max} to reach R_{max} was found to be equal to 24.5 \pm 3.6 μs, 27.6 \pm 4.3 μs, 24.0 \pm 11.2 μs for −100 ns, 0 ns, and +100 ns 3 MHz arrival times. The bubbles all collapsed from these maximum points to the time of first collapse (t_{c1}) at 46.5 \pm 4.1 μ s, 55 \pm 4.5 μ s, and 42.8 \pm 4.8 μ s for -100 ns, 0 ns, and +100 ns before showing a rebound in diameter for all and a second collapse at time (t_{c2}) of 70.3 \pm 7.4 μ s, 85.25 \pm 10.0 μ s, 63.8 \pm 8.2 μ s for -100 ns, 0 ns, and $+100$ ns cases.

Bubble cloud size

Optical imaging was used to visualize histotripsy bubble clouds generated at p− ranging from 22 to 31 MPa at 1 Hz PRF inside agarose tissue phantoms early just after nucleation at 3 μ s following p− pulse arrival (figure [5](#page-6-0)). For all cases, the bubble cloud dimensions were measured in the axial and elevational directions for pressures

Figure 4. Single bubble R_t curves. Plots shows all ($n = 20$) single bubble radius versus time for (A) −100 ns, (B) 0 ns, and (C) +100 ns 3 MHz pulse arrivals as well as (D) the mean radius (Rt) for single bubble cloud dimensions at 5 μs increments from −1 to 109 μs after peak negative pulse arrival. The mean maximum radius (R_{max}) was found to be equal to 260.2 \pm 16.3 μ m, 289.5 \pm 19.9 μ m, and 241.87 ± 25.2 μm at times to maximum radius (t_{max}) of 24.5 ± 3.6 μs, 27.6 ± 4.3 μs, 24.0 ± 11.2 μs for -100 ns, 0 ns, and +100 ns cases. The respective times to total collapse (t_{c2})as determined from optical imaging were 70.3 \pm 7.4 μ s, 85.25 \pm 10.0 μ s, 63.8 \pm 8.2 μ s for -100 ns, 0 ns, and $+100$ ns cases.

Figure 5. Bubble cloud images. Optical images captured by the FLIR camera and high-speed strobe of cavitation bubble clouds generated by 1 Hz PRF histotripsy pulses inside 1% agarose phantoms shown at the same actual scale. Ultrasound propagating left to right.

above the respective cavitation thresholds up to a maximum p − of 31 MPa (figure [6](#page-7-0)). As hypothesized, cavitation bubbles were observed for all dual-frequency cases when the focal pressure exceeded∼24–26 MPa. For all cases, sharply delineated bubble clouds were observed, with bubble clouds of increasing size containing more bubbles seen with increasing pressure levels beyond the cavitation threshold, matching previous intrinsic threshold histotripsy studies (figure 5) (Maxwell et al [2013](#page-12-0), Lin et al [2014b](#page-12-0), Vlaisavljevich et al [2017](#page-13-0)). The measured bubble cloud areas at p – ranging from 2[6](#page-7-0) to 31 MPa are shown in figure 6 with the minimum and maximum mean cloud dimensions and areas for each 3 MHz arrival time summarized in table [1](#page-7-0).

Bubble cloud nucleation and dynamics

High-speed optical imaging of the bubble cloud nucleation growth and collapse using the Phantom high-speed camera can be seen in figure [7](#page-7-0) and is quantified in figure [8.](#page-8-0) Figure [7](#page-7-0) shows selected clouds generated from a single

Table 1. Bubble cloud dimensional summary. Mean bubble cloud lengths, heights and areas plus or minus the respective standard deviations for 26 and 31 MPa for each dual-frequency case.

dual-frequency histotripsy pulse taken at 5 μ s intervals from just before peak negative pulse arrival through collapse. Examining these images(figure 7) along with figure [8,](#page-8-0) the overall effect of the 3 MHz pulse arrival timing on the bubble clouds dynamics can be assessed. At 4 μ s following p− arrival, the bubble clouds have nucleated very similar clouds for all three cases, with only some slight differences in bubble size and bubble cloud area. For example, the 0 ns and +100 ns cases showed similarly sized bubble clouds at this timepoint with mean areas of 0.96 \pm 0.04 mm² and 0.92 \pm 0.04 mm², whereas the -100 ns case produced a slightly smaller cloud of 0.69 ± 0.04 mm² at this timepoint. Beginning at 9 μ s, significant differences in cloud dynamics were observed. Bubble clouds produced by −100 ns and 0 ns continued to grow and covered a greater area that was uniformly

aligned over the entire location where the bubbles were initially nucleated. In contrast, the rear portion of the +100 ns cloud (post focal region) had already begun to collapse despite the front portion of the cloud continuing to expand. The total area covered by +100 ns grew to a mean maximum area of 1.03 \pm 0.05 mm² at ∼19 μ s and began to collapse from this time, while 0 ns and −100 ns continued to grow to 1.47 \pm 0.078 mm² at ~29 μ s and 1.2 ± 0.12 mm² at ∼34 μs, respectively, before beginning to collapse. These differences were primarily due to the reduced expansion and more rapid collapse of bubbles in the rear portion of the cloud for the later arrival time of the 3 MHz pulse (+100 ns). The collapse time for the entire bubble cloud produced for the 0 ns and −100 ns cases were similar, with the $+100$ ns cloud showing a slightly earlier collapse time.

The observation that the bubble cloud dynamics varied in the front and rear regions of the clouds(figure [7](#page-7-0)) led to individual analysis comparing the effects of 3 MHz pulse arrival on the bubble cloud dynamics in the front and rear halves of the cloud (figures $8(B)/C$). In general, results showed similar behavior in the front regions of the bubble clouds for all cases, with significant changes to the expansion and collapse of bubbles in the rear portion of the clouds(figures [7,](#page-7-0) 3.8). The front regions of the bubble clouds were observed to nucleate, grow, and collapse similarly for all cases with each of the arrival times reaching their maximum mean areas of 0.78 \pm 0.12 mm² at ~39 μ s, 0.89 ± 0.07 mm² at ~44 μ s, and 0.69 ± 0.06 mm² at ~34 μ s, respectively (figure 8(B)). These front regions reached their respective collapse times at ~94 μs, ~89 μs, and ~84 μs for −100 ns, 0 ns, and +100 ns arrival times(figure 8(B)). In examining the rear portions of the bubble clouds, results showed significant differences in the bubble growth and collapse dynamics (figure $8(C)$). For instance, bubble clouds for the −100 ns case achieved a maximum mean rear area of 0.49 \pm 0.08 mm² at a similar time to the front half of the bubble cloud (∼39 μs) before collapsing at∼69 μs. In contrast, clouds produced at 0 ns expanded until \sim 24 μs to reach a maximum mean rear area of 0.65 \pm 0.08 mm² before collapsing at \sim 59 μs but at a more accelerated rate. This trend was even more apparent in the rapid collapse of rear portion of the bubble cloud for the +100 ns case, which reached a mean maximum area of only 0.44 \pm 0.06 mm² at ∼4 μ s before collapse at \sim 44 μ s (figure 8(C)). In addition, supplemental figure 1 shows cloud differences in bubble cloud growth between 1 and 9 μ s in greater detail for different clouds captured at 0.5 μ s interval by the machine vision FLIR camera and 1 μ s strobe. As a note, the number of bubbles in each cloud and the corresponding bubble density analysis could not be effectively collected in this study and compared to our prior single-frequency study due to the extensive bubble overlap resulting from very dense bubble clouds formed with larger bubbles in all dualfrequency cases.

Ablation in RBC phantoms

RBC ablation experiments demonstrating the clinically translatable impact of the results presented in the prior sections showed that dual-frequency histotripsy predictably and reproducibly created lesions with clear demarcations between treated and untreated RBC regions(figure [9](#page-9-0)). The lesion area increased with increasing pulse number across all cases resulting in well-defined, complete ablations localized to the focal region (figure [9](#page-9-0), figure $10(A)$ $10(A)$). Figure $10(C)$ shows ablation from the first 100 pulses to better visualize the lesion formation during these initial pulses when the lesion is rapidly forming. The ablation areas were then normalized to the focal area (measured as the mean area of the bubble cloud in the RBC phantom) to quantitatively assess and compare the ablation efficiencies(table [2](#page-9-0)) (figures [10](#page-9-0)(B)/(D). All tested dual-frequency cases produced rapid ablation, achieving >75% ablation after an average of 37.7 \pm 17.5, 69.3 \pm 48.4, and 49 \pm 11.5 pulses for −100 ns, 0 ns, and $+100$ ns respectively. The final lesions' mean areas were A_{-100} $=$ 0.68 \pm 0.04 mm², A_0 $=$ 1.09 \pm 0.21 mm², and $A_{+100} = 1.26 \pm 0.11$ $A_{+100} = 1.26 \pm 0.11$ $A_{+100} = 1.26 \pm 0.11$ mm² (figure 10(A)) reflecting 156 \pm 5.7, 148 \pm 28.3%, and 149 \pm 13.5% (figure [10](#page-9-0)(C)) of the focal area after the complete treatment of 2000 pulses. Closely examining the lesion

Figure 9. RBC ablation. Images captured by optical imaging using the FLIR camera show the cavitation bubble cloud (dark) and histotripsy lesions(white) generated in RBC phantoms(grey) by histotripsy pulsing applied at 31 MPa at 1 Hz PRF propagating left to right for each 3 MHz arrival time.

Figure 10. RBC ablation plots. Plots show the mean and standard deviation of (A) the measured ablation area formed after every 40th pulse in the RBC agarose gel phantoms by 1 Hz PRF histotripsy pulsing at 31 MPa,(B) the measured ablation area for every 40th pulse normalized to the respective mean measured clouds areas formed in the RBCs, and (C) the ablation areas and (D) normalized areas following every other pulse of the first hundred pulses.

Table 2. Dual-frequency ablation efficiency summary. The table shows the mean number of pulses plus or minus the standard deviation required to achieve the specified percent ablation of the respective normalized focal areas from all dual-frequency histotripsy cases applied at p− was 31 MPa.

formation over the first 100 pulses, the lesion areas were A_{-100} $=$ 0.44 ± 0.01 mm², A_0 $=$ 0.67 ± 0.16 mm², $A_{+100} = 0.74 \pm 0.004$ $A_{+100} = 0.74 \pm 0.004$ $A_{+100} = 0.74 \pm 0.004$ mm² (figure 10(B)) reflecting 102 \pm 8.1%, 91.4 \pm 22.3%, and 88 \pm 0.49% (figure 10(D)) of the focal area as predicted by the size of the bubble cloud.

Discussion

This work builds on prior studies that have proposed dual-frequency histotripsy methods for generating more precise ablation of deep targets (Lin et al [2014a,](#page-12-0) Lin et al [2015](#page-12-0)) and providing precise control over bubble size to design more tissue selective treatment methods (Mancia et al [2019](#page-12-0)). In this study, the bubble cloud behavior and ablative efficiency of intrinsic threshold histotripsy generated using dual-frequency pulsing (500 kHz:3 MHz with a 1:1 pressure ratio) were evaluated. Results showed that the bubble cloud dimensions, bubble size, and bubble density observed for the dual-frequency cases were between those previously reported for the individual 500 kHz and 3 MHz cases(Vlaisavljevich et al [2015c](#page-13-0)). Bubble clouds across dual-frequency cases proved intermediate to the previously published dimensions for single-frequency 500 kHz and 3 MHz cases, with the bubble cloud dimensions being closer to previously published dimensions for 3 MHz (area $= 0.01 \pm 0.03$ mm² at 26 MPa to 0.62 ± 0.03 mm² at 40 MPa) (Edsall *et al* [2021a](#page-12-0)). The 3 MHz arrival time was also shown to slightly modulate cloud size with the late arrival (+100 ns) of the 3 MHz producing a larger cloud and the early arrival (−100 ns) producing a smaller cloud than the aligned arrival (0 ns). This difference in cloud size demonstrates some capability for dual-frequency pulsing to further alter cloud size by altering the arrival times of the contributing pulses.

Single bubble results showed that the bubble radii and collapse times were intermediate to those of previously published results for 3 MHz and 500 kHz single-frequencies, but more closely approached the those of 500 kHz, $(R_{max} = 297.4 \pm 50.5 \mu m$, $t_{max} = 28 \mu s$ and $t_c = 72 \mu s$) (Vlaisavljevich *et al* [2015c](#page-13-0)). These findings suggest that the individual bubbles are more strongly influenced by the low-frequency pulse resulting in larger bubbles with only slightly reduced R_{max} and collapse time results. This also confirms the results of prior work (Lin et al [2014a](#page-12-0)) by showing insignificant effects of 3 MHz pulse arrival time on individual bubble size. At higher pressures, the results from this study showed the significant benefits of using dual-frequency histotripsy for generating dense, well-confined bubble clouds that can harness the respective features of low(larger bubble expansion) and high (higher bubble density, more confined bubble cloud) frequency pulsing to achieve more efficient tissue ablation. Optical imaging results of cloud formation show delineated, densely populated bubble clouds of a size intermediate to previously published results for the contributing single-frequencies of 500 kHz and 3 MHz, but more closely aligned with the higher-frequency's cloud size (Vlaisavljevich et al [2015c,](#page-13-0) Edsall et al [2021a](#page-12-0)). These results confirm findings of previous studies showing bubble cloud dimensions predictably increase in size and number of bubbles with increasing pressure (Maxwell et al [2013,](#page-12-0) Lin et al [2014b,](#page-12-0) Vlaisavljevich et al [2017](#page-13-0)) and further support the predictability and reproducibility of using intrinsic threshold histotripsy for precise ablative applications. Although bubble density could not be quantified in this study due to the very high density cloud that consisted of rapidly expanding bubbles observed in this dual-frequency case, the qualitative observations of this study highlight how dual-frequency pulsing can be utilized to optimize the tradeoffs in bubble density and bubble expansion that were observed in our prior frequency study (Edsall *et al* [2021a](#page-12-0)). These findings suggest that dual-frequency histotripsy can be used to generate more precise ablation without the normal trade-offs of using higher frequency, such as reduced bubble size (Lin et al [2014a,](#page-12-0) Vlaisavljevich et al [2015c](#page-13-0)), increased tissue attenuation (Vlaisavljevich et al [2015b](#page-13-0)), and reduced ablation efficiency (Lin et al [2014a,](#page-12-0) Edsall et al [2021a](#page-12-0)).

Results from the bubble cloud analysis showed dual-frequency clouds exhibited typical intrinsic threshold nucleation, with well-defined bubble clouds covering the region of the focus above the cavitation threshold. The rates presented in the results show an increased ablation efficiency for all dual-frequency cases compared to previously published results for 500 kHz (>250 pulses), 1 MHz (>800 pulses), and 3 MHz (>1000 pulses) single-frequency histotripsy pulsing when removing the respective bubble cloud areas (Edsall *et al* [2021a](#page-12-0)). Another interesting finding of this study showed that the timing of the 3 MHz pulse significantly affected the dynamics of the bubble clouds, particularly the rear portion of the bubble clouds. While the front portions of the bubble clouds showed similar rates of growth and collapse, the rear portions of the bubble cloud showed distinctly different dynamics depending upon the arrival time of the 3 MHz pulse. For instance, the early arrival of the 3 MHz pulses produced a bubble cloud with bubbles expanding to similar sizes with similar collapse times in the front and rear portions of the cloud. In contrast, the aligned (0 ns) and delayed $(+100 \text{ ns})$ arrival of the 3 MHz resulted in decreasing growth of the rear half of the bubble cloud relative to the front of the bubble cloud. This change in bubble expansion due to pulse arrival indicates the sensitivity of pulse alignment in determining the ablative capacity of dual-frequency histotripsy. These findings suggested that an earlier arrival time for the higher frequency pulse allows for more uniform ablation of the entire focal area when using dual-frequency

pulsing methods. We hypothesize that aligning the 3 MHz pulse with the initial arrival of the longer 500 kHz pulse allows for the bubbles within the cloud to be nucleated at an earlier point of the 500 kHz pulse, allowing for the extended tensile phase of the pulse to grow a higher percentage of the nucleated bubbles to a larger size. This hypothesis is based upon prior experimental (Lin et al [2014a](#page-12-0), Vlaisavljevich et al [2015c](#page-13-0)) and theoretical (Mancia et al [2017](#page-12-0)) findings that show low frequencies produce greater bubble expansion due to the longer duration of the applied p−. In contrast, results from the aligned and late arrival cases for the 3 MHz pulse show that only the bubbles in the front of the cloud appear to be expanded due to the 500 kHz pulse, whereas bubbles in the rear portion of the cloud appear to be experiencing more limited expansion behavior that is typical of the higher frequency 3 MHz pulse.

Ablation experiments showed that dual-frequency pulsing resulted in precise ablation of the focal areas with even greater efficiency than what was previously observed for any single-frequency ablation in our prior study, including the most efficient ablation that was observed for 500 kHz pulsing. The lesions produced in the RBC phantoms in this study were generated in fewer pulses than what was previously reported for 500 kHz or 3 MHz, regardless of 3 MHz arrival time. The arrival time for the high frequency 3 MHz pulse was shown to impart a greater degree of lesion control with an earlier arrival(−100 ns)resulting in a smaller, more confined lesion that was produced at a faster rate than what was observed for the late (+100 ns) arrival time that created a larger lesion at a slower rate. This finding aligns well with the observed differences in bubble cloud nucleation and dynamics described in the previous section. For instance, comparing the lesion formation in figure [9](#page-9-0) to the data from the bubble cloud dynamics, it can be seen that the late arrival $(+100 \text{ ns})$ lesions were formed more rapidly toward the fronts of the clouds while the rears of the lesions were slower to form, likely due to the reduced growth of bubbles toward the back of the cloud (figure [7](#page-7-0)). In both the aligned and late arrival $(+100 \text{ ns})$ data, ablation clearly favors the regions of the cloud with the greater degree of bubble expansion, supporting prior work demonstrating the significance of bubble expansion in histotripsy ablative capacity (Vlaisavljevich et al [2016](#page-13-0), Mancia et al [2019](#page-12-0), Mancia et al [2020](#page-12-0), Edsall et al [2021b](#page-12-0)). Altogether, the ablation experiments show the ability of dual-frequency to rapidly ablate tissue with high precision and efficiency, with the ability to modulate these properties by changing the pulse arrival times of the respective frequencies, thereby impacting the uniformity and efficiency of the lesion formation. To completely unlock the potential benefits of dual-frequency pulsing, future work should explore this concept further by investigating the ability to modulate bubble cloud size, bubble cloud density, and bubble expansion across a much wider range using larger dual-frequency transducers that change more parameters including array geometry, pulse arrival times, and the relative pressure ratio produced by the respective high and low frequency components.

Based on all of the prior studies investigating intrinsic threshold histotripsy, our working hypothesis is that histotripsy the portion of tissues ablated by the formation of the bubble cloud is determined by the dimensions of the bubble cloud while the ablation efficiency is determined from the integrated effects of the size and rate of bubble expansion and the density of the bubbles within the bubble cloud. The results in this study suggest that it may be possible to independently manipulate cloud size, bubble size, and bubble density using dual-frequency pulsing methods by selectively altering the frequency components, the transducer geometry, pressure ratios, and other properties. Although further work is needed to fully test this possibility, the results from this study suggest that the bubble cloud dimensions are influenced more strongly by the higher frequency, which provides a constraint on the region of the focus above the intrinsic threshold. In contrast, bubble size appears to be influenced more strongly by the lower frequency components of the waveform that enable enhanced bubble growth, particularly when the bubbles are nucleated at an early time point with respect to the lower frequency pulse. While these results provide significant insight into dual-frequency histotripsy, it is important to note that this study investigates single bubble and bubble cloud dynamics, and biomechanical effects inside a bulk medium and may present different findings than work investigating cavitation formed near boundaries(Brujan et al [2005](#page-12-0), Brujan et al [2012](#page-12-0)). Future work will more closely examine the cavitation behavior in different tissue conditions and will explore a more comprehensive parametric assessment of transducer geometry, array configuration, pulse timing, and p − ratios to provide a more complete understanding of the effects of each parameter on bubble cloud characteristics and the resulting ablative efficiency.

Conclusion

The results of this study show the potential to modulate bubble cloud and ablation characteristics through dualfrequency histotripsy pulsing. These findings support prior research showing that bubble cloud characteristics in intrinsic threshold histotripsy, including the bubble cloud size, density, and bubble expansion, can be precisely tailored by modulating the single-cycle pulsing parameters. These finding suggest the potential for altering the rate and extent of ablation through the application of dual-frequency pulse modulation. Overall, this study demonstrates the potential of dual-frequency to increase control over histotripsy bubble cloud

characteristics, behavior, and ablation efficiency and thereby potentially improve the efficiency, safety, and efficacy of histotripsy-based therapies.

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All of the authors contributed significantly to the development and execution of this study. EV and CE conceived the initial ideas for this work. CE and TH contributed to the focused ultrasound transducer design and construction and the development of the experimental setups. CE and LH collected and analyzed all cavitation and red blood cell lesion data captured by optical imaging. CE also collected, conceived, developed, and performed methods for analyzing single bubble and bubble cloud growth and collapse data collected by highspeed imaging. CE led the writing of this manuscript. All authors contributed critically to the interpretation of results, drafting of the manuscript and gave final approval for this submission.

Data availability statement

The data cannot be made publicly available upon publication because they are not available in a format that is sufficiently accessible or reusable by other researchers. The data that support the findings of this study are available upon reasonable request from the authors.

ORCID iDs

Connor Edsa[l](https://orcid.org/0000-0001-6876-1496)l @ [https:](https://orcid.org/0000-0001-6876-1496)//orcid.org/[0000-0001-6876-1496](https://orcid.org/0000-0001-6876-1496) Timothy L Hall @ [https:](https://orcid.org/0000-0002-4867-2856)//orcid.org/[0000-0002-4867-2856](https://orcid.org/0000-0002-4867-2856) Eli Vlaisavljevic[h](https://orcid.org/0000-0002-4097-6257) ® [https:](https://orcid.org/0000-0002-4097-6257)//orcid.org/[0000-0002-4097-6257](https://orcid.org/0000-0002-4097-6257)

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