BRIEF REPORT



HIV-Positive Liver Transplant Does not Alter the Latent Viral Reservoir in Recipients With Antiretroviral Therapy-Suppressed HIV

Sarah E. Benner,¹ Xianming Zhu,¹ Sarah Hussain,² Sander Florman,³ Yolanda Eby,¹ Reinaldo E. Fernandez,² Darin Ostrander,² Meenakshi Rana,⁴ Shane Ottmann,⁵ Jonathan Hand,⁶ Jennifer C. Price,⁷ Marcus R. Pereira,⁸ David Wojciechowski,⁹ Jacques Simkins,¹⁰ Valentina Stosor,¹¹ Sapna A. Mehta,¹² Saima Aslam,¹³ Maricar Malinis,^{14,©} Ghady Haidar,^{15,©} Allan Massie,¹⁶ Melissa L. Smith,¹⁷ Jonah Odim,¹⁸ Megan Morsheimer,¹⁸ Thomas C. Quinn,^{2,19} Gregory M. Laird,²⁰ Robert Siliciano,² Ashwin Balagopal,² Dorry L. Segev,¹⁶ Christine M. Durand,² Andrew D. Redd,^{2,19,a} and Aaron A. R. Tobian^{1,2,a,©}

¹Department of Pathology, Johns Hopkins School of Medicine, Baltimore, Maryland, USA; ²Department of Medicine, Division of Infectious Diseases, Johns Hopkins School of Medicine, Baltimore, Maryland, USA; ³Recanati/Miller Transplantation Institute, Icahn School of Medicine at Mount Sinai, New York, New York, USA; ⁴Department of Medicine, Icahn School of Medicine at Mount Sinai, New York, New York, USA; ⁵Department of Surgery, Johns Hopkins University School of Medicine, Baltimore, Maryland, USA; ⁶Department of Infectious Diseases, Ochsner Health, New Orleans, Louisiana, USA; ⁷Department of Medicine, University of California San Francisco, San Francisco, California, USA; ⁸Department of Medicine, Columbia University Irving Medical Center, New York, New York, USA; ⁹Division of Nephrology, University of Texas Southwestern, Dallas, Texas, USA; ¹⁰Department of Medicine/Division of Infectious Diseases, University of Miami School of Medicine, Miami, Florida, USA; ¹¹Departments of Medicine and Surgery, Divisions of Infectious Diseases and Organ Transplantation, Northwestern University Feinberg School of Medicine, Chicago, Illinois, USA; ¹²Department of Medicine, New York University Grossman School of Medicine, New York University Langone Health, New York, New York, USA; ¹³Department of Medicine, Division of Infectious Diseases and Global Public Health, University of California San Diego, La Jolla, California, USA; ¹⁴Department of Internal Medicine, Yale University School of Medicine, New Haven, Connecticut, USA; ¹⁵Department of Medicine, Division of Infectious Diseases, University of Pittsburgh, Pittsburgh, Pennsylvania, USA; ¹⁶Department of Surgery, New York University Grossman School of Medicine, New York University Langone Health, New York, New York, USA; ¹⁷Department of Biochemistry and Molecular Genetics, University of Louisville, Louisville, Kentucky, USA; ¹⁸Division of Extramural Research, National Institute of Allergy and Infectious Diseases, National Institutes of Health, Bethesda, Maryland, USA; ¹⁹Division of Intramural Research, National Institute of Allergy and Infectious Diseases, National Institutes of Health, Bethesda, Maryland, USA; and ²⁰Accelevir Diagnostics, Baltimore, Maryland, USA

The latent viral reservoir (LVR) remains a major barrier to HIV-1 curative strategies. It is unknown whether receiving a liver transplant from a donor with HIV might lead to an increase in the LVR because the liver is a large lymphoid organ. We

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found no differences in intact provirus, defective provirus, or the ratio of intact to defective provirus between recipients with ART-suppressed HIV who received a liver from a donor with (n = 19) or without HIV (n = 10). All measures remained stable from baseline by 1 year posttransplant. These data demonstrate that the LVR is stable after liver transplantation in people with HIV.

Clinical Trials Registration. NCT02602262 and NCT03734393.

Keywords. HIV; HIV cure; latent viral reservoir; liver; organ transplant.

The latent viral reservoir (LVR) remains one of the major barriers in curative strategies for people with human immunodeficiency virus (PWH) [1]. The LVR is established early in infection, augmented throughout periods of active viral replication, and persists primarily in resting CD4⁺ T cells, even in the presence of suppressive antiretroviral therapies [2]. Several strategies aimed at eliminating the LVR or inducing durable immunologic control of HIV have been evaluated in clinical trials, and while a small number of PWH have achieved durable antiretroviral therapy (ART)-free remission [3, 4], no broadly applicable and effective therapeutic approach targeting the LVR has been identified.

Previous studies have monitored the LVR over time in kidney transplant recipients with HIV who received organs from donors with and without HIV [5]. In these kidney recipients, despite a dramatic initial reduction in the LVR size among those receiving lymphocyte-depleting induction therapy, the reservoir reconstituted within 1 year, and there were no differences in the LVR size among those receiving lymphocyte nondepleting immunosuppression induction therapy or by allograft donor HIV status [5, 6]. However, one as yet unanswered question is whether the size of the LVR changes with liver transplantation from a donor with or without HIV into PWH. The liver is a secondary lymphoid organ that contains a substantial number of CD4⁺ T cells and also contains the largest population of tissue-resident macrophages in the body [7]. Accordingly, the liver may be a tissue reservoir of HIV, because HIV DNA and RNA have been found in human hepatocytes [8-11] and liver macrophages, even in the presence of suppressive ART [8, 10-12]. Consequently, the LVR could decrease among individuals who receive an HIV-negative allograft and increase among individuals who receive an HIV-positive allograft. The objective of this study was to evaluate changes in the LVR using the intact proviral DNA assay (IPDA) longitudinally in liver transplant recipients with ART-suppressed HIV participating in the HOPE in Action multicenter trial who received lymphocyte nondepleting therapies and allografts from donors with or without HIV.

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^aShared senior authors.

Correspondence: Aaron A. R. Tobian, MD, PhD, Department of Pathology, School of Medicine, The Johns Hopkins University, 600 North Wolfe Street, Carnegie Room 437, Baltimore, MD 21287 (atobian1@jhmi.edu). Andrew D. Redd, PhD, NIAID, 851 North Wolfe Street, Rangos Room 540, Baltimore, MD 21287 (reddandrew@niaid.nih.gov).

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METHODS

Study Populations and Procedures

Liver transplant recipients with HIV who received an organ from a deceased donor with or without HIV were longitudinally followed in the HOPE in Action multicenter, national clinical trial (clinicaltrials.gov, NCT02602262 and NCT03734393) [13]. Participant and donor inclusion criteria have been previously described [14]. Recipient CD4 cell count, viral load, and ART regimens at the time of transplantation were obtained by extraction from clinical records. Peripheral blood mononuclear cells (PBMCs) were collected from recipients and processed as previously described [14]. Briefly, PBMCs were collected at the time of transplant, prior to immunosuppression induction therapy (week 0), and at subsequent time points following transplant (approximately at 13, 26, and 52 weeks posttransplant, if applicable). All liver transplant recipients were given lymphocyte nondepleting immunosuppressive therapies. Participants had to have a minimum of 1 follow-up time point after transplant to be included in this study.

Laboratory Testing

IPDA was performed as previously described (Accelevir Diagnostics) [5]. Briefly, total $CD4^+$ T cells were isolated from cryopreserved PBMCs via immunomagnetic selection, and genomic DNA was isolated. IPDA is a droplet digital polymerase chain reaction (ddPCR) assay that targets 2 regions (the packaging signal and envelope) in the HIV genome that are either frequently deleted or hypermutated in defective proviruses [15]. Droplets positive for both fluorescence signals are scored as intact, while 1 fluorescence signal can either indicate (1) a hypermutation and/or 3' deletion, or (2) a 5' defective provirus. Double-negative droplets contain either no provirus or defects in both amplicon regions [15]. Of the 87 samples, 82 amplified and the 5 samples that failed to amplify were thus excluded from the analysis.

Donor viral load was evaluated using the Abbot Realtime viral load assay. The limit of detection was 40 copies per mL. Donor CD4 cell counts were measured using the T-cell subset assay (Becton Dickinson).

Statistics

All baseline and follow-up characteristics of the study population were summarized overall and by donor HIV status using descriptive statistics as previously described [5]. The primary outcomes evaluated included intact provirus, defective provirus, ratio of intact/defective provirus frequencies, and CD4⁺ T-cell counts per μ L of blood stratified by donor allograft HIV status. The intact provirus and defective provirus frequencies were analyzed both by frequency per million CD4⁺ T cells and frequency per mL of blood, as previously described [5]. All values were log₁₀ transformed, except the ratio of intact/ defective provirus frequencies, to approximate normal distributions. The biomarkers were plotted over time with locally estimated scatterplot smoothing (LOESS) and *t*-based approximation of 95% confidence intervals (CIs). All analyses were performed in R 4.1 (R-Core Team) and Stata/MP, version 15.1 (StataCorp).

Study Approval

The trial and current study were approved by the Johns Hopkins University Institutional Review Board (IRB) and also IRBs at each center. All transplant recipients (participants) provided written informed consent.

RESULTS

Demographics of the Study Participants

There were 29 liver transplant recipients with a median age of 58 years old (interquartile range [IQR] = 57-64), and the majority were male (83%, n = 24) and white (59%, n = 17) (Table 1). Most recipients were on integrase inhibitor (INSTI)-containing ART (83%, n = 24), less than one-quarter were on nonnucleoside reverse transcriptase inhibitors (NNRTI)-containing ART (17%, n = 5), and only 1 (3%) was on protease inhibitor-based ART. Five received a simultaneous liver and kidney transplant. No recipients received lymphocyte-depleting induction immunosuppression.

Of the 29 participants, 19 received a transplant from a donor with HIV and 10 from a donor without HIV. Of the 19 donors with HIV, 11 were on ART and 10 of the 11 had suppressed HIV RNA <200 copies/mL. The remaining 8 donors had untreated HIV with median HIV-1 RNA 15 186 (IQR = 760–290 222) copies/mL. There were no significant differences between recipients according to liver donor HIV status, except that a greater proportion of recipients of livers from donors without HIV received prednisone maintenance therapy.

Longitudinal Trajectories in the Latent Viral Reservoir

Total CD4⁺ T-cell counts in participants who received livers from donors with or without HIV declined slightly at weeks 13 and 30, but overall returned to baseline by 1 year posttransplant (Figure 1*A*). However, the ratio of intact to defective provirus did not significantly alter over time, and no differences were observed between those who received livers from donors with or without HIV (Figure 1*B*). Levels of intact provirus per million CD4⁺ T cells or per mL of blood had an initial slight increase in those who received livers from donors without HIV but declined by around week 25. Levels returned to near baseline in both groups by 1 year, and no differences between those who received livers from donors with or without HIV were observed (Figure 1*C* and Supplementary Figure 1A). Levels of defective provirus per million CD4⁺ T

Table 1. Baseline Characteristics of Liver and Liver-Kidney Recipients by Donor HIV Status

Characteristics	Total (n = 29)	HIV D+/R+ (n = 19)	HIV D $-/R+(n = 10)^{a}$	<i>P</i> Value
Age, y, median (IQR)	58 (57–64)	57 (56–64)	58 (57–62)	.82
Male sex	24 (83)	16 (84)	8 (80)	.78
Race				
White	17 (59)	11 (58)	6 (60)	.86
African American	8 (28)	5 (26)	3 (30)	
Asian	1 (3)	1 (5)	O (O)	
American Indian or Alaska Native	1 (3)	1 (5)	0 (0)	
Not specified	2 (7)	1 (5)	1 (10)	
Latino ethnicity	6 (21)	4 (21)	2 (20)	
Duration of HIV infection, y, median (IQR)	28 (15–32)	30 (15–33)	26 (9–29)	.16
CD4 ⁺ T-cell count, median (IQR)	278 (198–393)	290 (198–427)	273 (145–392)	.71
HIV RNA < 200 copies/mL	29 (100)	19 (100)	10 (100)	
Received SLK	5 (17)	5 (26)	0 (0)	.07
ART regimen ^{b,c}				
INSTI containing	24 (83)	16 (84)	8 (80)	.78
NRTI containing	11 (38)	8 (42)	3 (30)	.52
NNRTI containing	5 (17)	4 (21)	1 (10)	.45
PI containing	1 (3)	0 (0)	1 (10)	.16
Immunosuppression ^b				
Induction regimen				
Corticosteroids only	25 (86)	15 (79)	10 (100)	.29
Basiliximab and corticosteroids	3 (10)	3 (16)	0 (0)	
Basiliximab only	1 (3)	1 (5)	0 (0)	
Maintenance ^c				
Tacrolimus	27 (93)	17 (89)	10 (100)	.29
Mycophenolate mofetil (MMF)	22 (76)	14 (74)	8 (80)	.71
Prednisone	23 (79)	13 (68)	10 (100)	.05
Everolimus	1 (3)	1 (5)	0 (0)	.46
Donor characteristics				
CD4 ⁺ T-cell count, median (IQR) ^d		156 (112–272)	ND	
On ART		11 (58)	NA	
HIV load <200 copies/mL ^e		10 (53)	NA	
Not on ART		8 (42)	NA	
HIV load, copies/mL, median (IQR)		15 186 (760–290 222)	NA	

Data are presented as median (IQR) for continuous measures, and No. (%) for categorical/binary measures. Pvalues were calculated by Wilcoxon rank sum test while Pearson χ^2 test was used for categorical and binary variables.

Abbreviations: ART, antiretroviral therapy; D, donor; HIV, human immunodeficiency virus; INSTI, integrase strand transfer inhibitor; IQR, interquartile range; ND, not done; NA, not applicable; NNRTI, nonnucleoside reverse transcriptase inhibitors; PI, protease inhibitor; R, recipient; RNA, ribonucleic acid; SLK, simultaneous liver-kidney transplant.

^aOf 10 D–/R+, 3 donors had a false-positive HIV test.

^bImmunosuppression and ART regimens were reported as active prescriptions at 4 weeks posttransplant.

^cPercentages may add up to more than 100 because an individual can fall under more than 1 category.

^dOf 19 D+/R–, 2 donors had missing information on CD4⁺ T-cell count.

^eOf 19 D+/R–, 1 donor had missing information on HIV load.

cells did not dramatically change from baseline to 1 year posttreatment in either group (Figure 1D and Supplementary Figure 1B). A sensitivity analysis was performed among recipients who received only a liver transplant (ie, removing the 5 participants who received a simultaneous liver-kidney transplant), but no differences were observed between the primary analysis and the sensitivity analysis (Supplementary Figure 2). A second sensitivity analysis was performed comparing recipients who received livers from viremic (viral load \geq 200 copies/mL) and non-viremic donors (viral load < 200 copies/ mL). By 1 year posttransplant, no differences were observed between the viremic and non-viremic donors (Supplementary Figure 3).

DISCUSSION

The persistence of the LVR remains the largest barrier in HIV curative strategies. Because of its size and function as a secondary lymphoid organ and its potential to harbor a significant number of HIV-infected cells, we examined whether transplantation of a liver from a donor with HIV or without HIV affects the size of the LVR in the recipient. Our study was designed to



Figure 1. Longitudinal trajectories of CD4 T⁺ cell count, and intact and defective provirus following liver transplant. *A*, CD4⁺ T-cell counts per µL of blood, (*B*) ratios of intact to defective virus, (*C*) intact provirus per million CD4⁺ T cells, and (*D*) defective provirus per million CD4 T-cells were measured longitudinally from time since transplant. Each line represents an individual, and each dot represents a specific time point. Bolded red and blue lines represent the locally estimated scatterplot smoothing (LOESS) curve. Shaded areas represent the 95% confidence interval of the LOESS curves.

answer this question, because we could compare the size of the LVR longitudinally after liver transplantation from donors with or without HIV. Here, liver transplant recipients with HIV had a slight decline in CD4⁺ T-cell counts, but the ratio of intact to defective provirus did not change dramatically. No differences were observed in recipients who received a liver from a donor with or without HIV. By 1 year posttransplant, CD4⁺ T-cell counts, intact provirus, and defective provirus were all comparable to baseline levels. This study demonstrates that the LVR is stable after transplantation of large lymphoid organs, such as the liver, in people with ART-suppressed HIV.

Studies have shown that kidney transplant patients who received lymphocyte nondepleting therapies had a slight reduction of CD4⁺ T-cell counts, and intact and defective provirus initially, but levels returned to baseline [5]. Liver transplant recipients in this study receiving lymphocyte nondepleting therapies show similar results. The ratio of intact to defective provirus did not change over time, suggesting that latently infected cells are able to be maintained even in larger lymphoid organ transplantations. However, liver macrophages have been shown to have higher infective levels than hepatocytes, and future studies should evaluate whether the LVR changes in these cellular reservoirs [8, 11]. There are a few limitations to this study. All participants were on suppressive ART and therefore the findings cannot be generalized to transplant recipients who interrupt HIV treatment. Moreover, this study only assessed proviruses within $CD4^+T$ cells that are in circulation. Therefore, it is possible that if other cells populations were included in the analysis (eg, liver-resident $CD4^+T$ cells), changes in the size of the LVR posttransplant may have been observed. When using the IPDA, not all samples had amplification, and thus some had to be excluded (n = 5). Participants receiving liver transplants were only given lymphocyte nondepleting therapies, and results could not be compared to those who received lymphocyte depleting therapies.

In conclusion, we found that PWH who received a large lymphoid organ transplantation, such as a liver, have similar intact and proviral trajectories compared to patients who received kidney transplants. Moreover, the ratio of intact/defective provirus did not change overtime in liver patients who received lymphocyte nondepleting therapies and no differences were observed among recipients of livers from donors with or without HIV. It is reassuring that the LVR does not seem to increase with this novel transplantation practice.

Supplementary Data

Supplementary materials are available at *The Journal of Infectious Diseases* online. Consisting of data provided by the authors to benefit the reader, the posted materials are not copyedited and are the sole responsibility of the authors, so questions or comments should be addressed to the corresponding author.

Notes

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All authors have submitted the ICMJE Form for Disclosure of Potential Conflicts of Interest. Conflicts that the editors consider relevant to the content of the manuscript have been disclosed.

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