# The multifaceted roles of activating transcription factor 3 (ATF3) in inflammatory responses – Potential target to regulate neuroinflammation in acute brain injury

Iournal of Cerebral Blood Flow & Metabolism 2023, Vol. 43(2S) 8-17 © The Author(s) 2023 Article reuse guidelines: sagepub.com/journals-permissions DOI: 10.1177/0271678X231171999 journals.sagepub.com/home/jcbfm



Yan Li<sup>1,\*</sup>, Qiuyue Fan<sup>1,\*</sup>, Fengshi Li<sup>2,\*</sup>, Rui Pang<sup>1</sup>, Chen Chen<sup>1</sup>, Peiying Li<sup>1</sup>, Xin Wang<sup>1</sup>, Wei Xuan<sup>1</sup> and Weifeng Yu<sup>1</sup>

### Abstract

Activating transcription factor 3 (ATF3) is one of the most important transcription factors that respond to and exert dual effects on inflammatory responses. Recently, the involvement of ATF3 in the neuroinflammatory response to acute brain injury (ABI) has been highlighted. It functions by regulating neuroimmune activation and the production of neuroinflammatory mediators. Notably, recent clinical evidence suggests that ATF3 may serve as a potential ideal biomarker of the long-term prognosis of ABI patients. This mini-review describes the essential inflammation modulatory roles of ATF3 in different disease contexts and summarizes the regulatory mechanisms of ATF3 in the ABI-induced neuroinflammation.

### **Keywords**

Activating transcription factor 3, inflammatory response, neuroinflammation, acute brain injuries

Received I November 2022; Revised 22 February 2023; Accepted 3 April 2023

### Introduction

Activating transcription factor 3 (ATF3), a transcription factor with a basic leucine zipper, has long been recognized as a member of the ATF/cAMP response element binding (CREB) protein family.<sup>1</sup> The expression of ATF3 is at low levels in quiescent cells, but increases in response to stressors such as oxidative damage and ischemia/reperfusion.<sup>2-4</sup> ATF3 is emerging as a master regulator of inflammation in multiple pathophysiological processes such as cardiovascular disease, dementia, and ischemia/reperfusion-induced damage.1,5-15

Neuroinflammation is one of the key pathological features of a wide range of acute brain injuries (ABIs),<sup>16–19</sup> which mainly include stroke and traumatic brain injury (TBI).<sup>20,21</sup> Neuroinflammation plays a pivotal role in ischemic brain injury during the acute phase of ABI, with excessive immune cell activation and the release of inflammatory mediators.<sup>22-24</sup> A persistent neuroinflammatory response will further lead to secondary neurodegenerative processes such as cognitive <sup>1</sup>Department of Anesthesiology, Clinical Research Center, Renji Hospital, Shanghai Jiao Tong University School of Medicine, Shanghai, China <sup>2</sup>Department of Neurosurgery, Clinical Research Center, Renji Hospital, Shanghai Jiao Tong University School of Medicine, Shanghai, China

\*These authors contributed equally to this work.

#### **Corresponding authors:**

Weifeng Yu, Department of Anesthesiology, State Key Laboratory of Oncogenes and Related Genes, Shanghai Cancer Institute, Renji Hospital, Shanghai Jiao Tong University School of Medicine, 160 Pujian Rd, Shanghai 200127, China.

Email: ywf808@yeah.net

Wei Xuan, Department of Anesthesiology, State Key Laboratory of Oncogenes and Related Genes, Shanghai Cancer Institute, Renji Hospital, Shanghai Jiao Tong University School of Medicine, 160 Pujian Rd, Shanghai 200127, China.

Email: muthaxw@163.com

Xin Wang, Department of Anesthesiology, State Key Laboratory of Oncogenes and Related Genes, Shanghai Cancer Institute, Renji Hospital, School of Medicine Shanghai Jiaotong University, 160 Pujian Rd, Shanghai 200127, China.

Email: wxsarah\_sjtu@163.com

impairment and dementia.<sup>18,25–27</sup> Clinical evidence also supports that unresolved neuroinflammation continuously shapes the evolving brain pathology and affects the long-term neurological outcome of patients.<sup>16,28–30</sup> Therefore, it's urgently needed to understand the putative mechanisms for dysregulated brain inflammatory responses in the pursuit of improving long-term neurological outcomes after ABI. Regarding ABI, recent studies suggested that ATF3 could be an ideal predictor of outcomes in stroke and TBI,<sup>31,32</sup> and ATF3 failure to regulate inflammatory responses account for poorer ABI outcomes by exacerbating neuroinflammation.<sup>33,34</sup>

In this mini review, we describe the regulation of ATF3 in various inflammatory responses, scrutinize the latest research findings, and highlight new insights into the potential involvement of ATF3 in neuroinflammation and several common ABIs.

# ATF3, is a stress-associated transcription factor that regulates local and systemic inflammation

ATF3 is a stress-associated transcription factor and a key regulator of local and systemic inflammation. Taking type 2 diabetes mellitus as an example, ATF3 is necessary for the complete induction of matrix metalloproteinase (MMP12) expression, a negative regulator of glucose metabolism, in macrophages, which connects microbiota-dependent inflammation to insulin resistance.<sup>35–37</sup> In addition to proinflammatory effects, ATF3 deficiency is associated with exacerbated inflammation and inflammation-related disease processes, such as neuronal apoptosis and, cellular metabolic dysfunction of macrophages and endothelial cells (ECs) in ischemia/reperfusion injury and cancer therapy.<sup>2,5,14,38,39</sup> Protein-protein interaction network analvsis further revealed that ATF3 interacts with 48 important stress-relevant proteins including other transcriptional complexes involved in inflammatory responses (Rela, Jun, Junb, Jund, Fos), indicating the core position of ATF3 in cellular adaptive-response networks.40

### ATF3 exerts double-sided effects on inflammatory response

As a multifaceted and controversial transcription factor, ATF3 exerts both protective and deleterious effects on inflammatory responses. Under stress, ATF3 may play a protective role against inflammation-associated damage in diseases as a compensatory factor.<sup>35</sup> It functions as an immunomodulator by interacting with nuclear factor kappa B (NF- $\kappa$ B) and suppressing proinflammatory cytokines such as interleukin(IL)-6-, IL-12, and tolllike receptor (TLR) 4 in mice exposed to lipopolysaccharide (LPS).<sup>41–45</sup> Furthermore, ATF3 interacts with BRG1-Associated Factor 60 A (BAF60a), a regulator of obesity-induced adipose tissue macrophages, to inhibit proinflammatory cytokine production in obese people.<sup>46</sup> In addition to cytokines, ATF3 links to the regulation of chemokine C-C motif chemokine 4 (CCL4) in macrophages and the attenuation of inflammatory responses.<sup>47</sup> In the early stress response, acute hypoxiainduced activation of tumor necrosis factor-alpha (TNF- $\alpha$ ) enhances ATF3 expression in ECs, and ATF3 regulates the activation of the c-Jun NH<sub>2</sub>-terminal protein kinase pathway, which protects cardiomyocytes from doxorubicin-induced death.<sup>48</sup> Thus, ATF3 can engage in a compensatory or adaptive homeostatic response to alleviate inflammatory responses during inflammation-related pathological processes.

By contrast, ATF3 has also proinflammation properties and can detect molecular patterns generated by necrotic cells to induce inflammation.<sup>49</sup> In respiratory infections caused by Staphylococcus aureus (S. aureus), ATF3 promotes inflammation by upregulating IL-6 and IL-12p40.<sup>50</sup> The amplified inflammatory response supports the antimicrobial effects, and ATF3 stimulates the migration of F4/80 macrophages for pathogen elimination in S. aureus-induced pneumonia.<sup>50</sup> Moreover, the ATF3-dependent upregulation of IL-6R and IL-6 may increase drug resistance to sorafenib and regorafenib in hepatocellular carcinoma.<sup>39</sup> In the tumor microenvironment of an immune-competent mouse model, shATF3 significantly decreased the PD-L1 level, thereby boosting the activated CD8<sup>+</sup> T cell population to increase its anti-tumor efficacy.<sup>51</sup> Notably, the ATF3/c-Jun/Lgals3 axis may enhance microglial activation in central diabetes insipidus following hypothalamic damage.52

# ATF3 mediates inflammatory responses by interacting with a complex metabolic-immune response network

Transcription and metabolic regulation have been recognized as powerful principles guiding inflammation.<sup>53–56</sup> As an important transcription factor, ATF3 interacts with a complex metabolic-immune response network in various cells, such as macrophages and ECs.<sup>57</sup> Several studies have suggested that the ATF3 function in metabolic, tumor, and cardiovascular diseases in part originates from its influence on cell metabolism.<sup>1,58–60</sup>

Abnormal metabolism induces stress reactions and subsequently influences ATF3-mediated inflammatory responses.<sup>36,46,53</sup> Guided by symptoms of lipodystrophy and fever among patients with proteasomeassociated autoinflammatory syndromes, scientists discovered that proteasome dysfunction blocks adipogenesis and induces inflammation by activating ATF3.<sup>61</sup> It is of note that ATF3 negatively regulates the acquisition of a pro-inflammatory phenotype induced by high-density lipoprotein (HDL) in macrophages.<sup>20</sup> ChIP-sequencing revealed that several key pro-inflammatory genes were directly targeted by ATF3 following its induction by HDL.20 As an HDL-inducible target gene in macrophages, ATF3 down-regulates the expression of TLR-induced proinflammatory cytokines and alleviates cellular stress.<sup>20</sup> Another disease that involves ATF3 and lipid metabolism is atherosclerosis, a progressive disease characterized by the accumulation of lipids and inflammatory responses in arteries. The synthetic ctype lectin receptor 4e (Clec4e) ligand induces nuclear translocation of ATF3 in a Clec4e-dependent manner in both peritoneal macrophages and bone marrowderived macrophages.<sup>21,62–64</sup> On the other hand, ATF3 can promote cholesterol efflux by regulating Clec4e signaling in macrophages, alleviate Clec4einduced endoplasmic reticulum (ER) stress response, and reduce the production of pro-inflammatory mediators and growth factors.<sup>65</sup> Furthermore, it limits macrophage foam cell formation and inflammation in granuloma remodeling and prevents the progression of atherosclerosis.<sup>65</sup> Another way to prevent macrophage foam cell formation by ATF3 relies on regulating the intracellular neutral fat accumulation by inhibiting the cholesterol 25-hydroxylase in macrophages and, modulating cytokine production and ER stress.<sup>66</sup> Therefore, ATF3 can be initially activated by metabolic stressors, then works as a modulator and inhibitor in response to a variety of inflammatory stimuli.<sup>7,12,50,67</sup>

Apart from the anti-inflammatory properties mentioned above, persistent ATF3 expression induced by excessive reactive oxygen species (ROS) or ER stress may have detrimental effects, overwhelming its initial compensatory property. For instance, triglyceride-rich lipoprotein (TL) lipolysis products promote nuclear ATF3 accumulation in carotid artery ECs of mice and increase vascular apoptosis,<sup>68</sup> whereas ATF3 promotes lipolysis products-induced transcription of E-selectin and IL-8 in human aortic Ecs.<sup>68</sup> Hence, ATF3 deletion can prevent vascular apoptosis by decreasing triglyceride-rich lipoproteins and lipolysis productsinduced inflammation.<sup>68</sup> Additionally, decreased ATF3 levels were necessary for HDL-induced inhibition of saturated fatty acid-mediated enhancement of LPS-induced inflammatory responses in ECs.<sup>69</sup> In response to lipotoxic brain microvascular damage, ATF3 mainly governs TL-induced inflammation and TNF signaling in the cerebrovascular system and increases EC apoptosis.<sup>67</sup> The ATF3 effects on inflammation depends on the various cellular environments under stress (Figure 1).

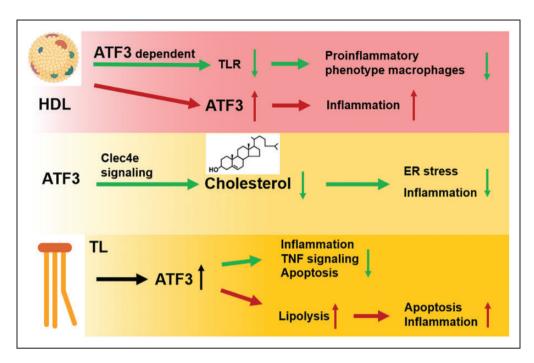
In summary, ATF3 may be therapeutically targeted to combat excessive inflammatory responses, or conversely, ATF3 can be repressed to strengthen an insufficient immune system.

# Neuroinflammation is an emerging therapeutic target for improving neurological function following ABI

ABI triggers robust inflammatory- responses,<sup>28,70–72</sup> then peripheral and brain immune dysregulation initiate a cascade of inflammatory signaling events, ultimately resulting in neuroinflammation.<sup>73</sup> The resulting intense neuroinflammation can last for years and trigger secondary neurodegeneration due to the excessive neuroimmune reaction induced by both resident brain cells and infiltrating peripheral inflammatory cells.<sup>26,74–77</sup> Thus, controlling disproportionate neuroinflammatory responses represents a promising therapeutic strategy to improve neurological outcomes after ABIs.<sup>25,78,79</sup>

Both innate and adaptive immune cells are highly involved in ABI-induced neuroinflammation. Neutrophils are the first leukocyte subtype to infiltrate the lesion site within 30 min after stroke and release excessive ROS, proinflammatory cytokines (IL-1β, IL-6, TNF-α) and chemokines (MCP-1, MIP-1α, IL-8), MMPs and adhesion molecules (PSGL-1, L-selectin), which may exacerbate brain injury.<sup>80</sup> Microglia are primary innate immune cells within the central nervous system (CNS) and can produce pro-inflammatory factors in response to noxious stimuli, such as IL-18. interferon  $\gamma$ , TNF- $\alpha$ , nitric oxide, and ROS.<sup>81,82</sup> Representative adaptive immune cells T and B lymphocytes migrate into the brain at a later stage than neutrophils after brain injury, they can significantly influence microglial phenotype and function.75,83 Neuroinflammation is also driven by neuroimmunerelated inflammatory mediators, for instance, complement activation promotes neuroinflammation and secondary brain injury after ABI, such as stroke, TBI, and intracerebral hemorrhage.<sup>84-86</sup> Inhibition of complement can reverse neuroinflammatory transcriptome changes and significantly improve cognitive performance after TBI.<sup>76,84,87,88</sup> TLR represent another recently emerging target for modulation of neuroinflammation after ABI<sup>89-92</sup> Inhibition of TLR4 signaling pathways has been suggested to reduce microglia/ macrophages activation and neutrophil infiltration, thereby ameliorating neuroinflammation and improving neurological function in animal models of ABI.93,94

In addition to basic research, clinical studies also emphasize the significance of neuroinflammation in patients with ABI.<sup>95</sup> Imitating the pathogenesis of



**Figure 1.** The interplay of ATF3 with metabolism on inflammation. HDL mediates anti-inflammatory reprogramming of macrophages via ATF3 in macrophages, but also induced ATF3 activity in endothelial cells then promote inflammation. ATF3 regulates Clec4e signaling to increase cholesterol efflux and reduce ER stress and inflammation. TL increases nuclear ATF3 accumulation. The accumulation of ATF3 can down-regulate TNF signaling and alleviate inflammation and apoptosis. ATF3 can also increases lipolysis, which exacerbates apoptosis and inflammation in carotid artery endothelial cells in the cerebrovascular system. ATF3: activating transcription factor 3; HDL: high-density lipoprotein; TLR: toll-like receptor; Clec4e: c-type lectin receptor 4e; ER: endoplasmic reticulum; TL: Triglyceride; TNF: tumor necrosis factor.

ischemic stroke, neonates receiving cardiac surgery with deep hypothermic circulatory arrest showed immediate inflammatory responses based on the analysis of their vena cava superior blood samples, showing increased immune cell counts and cytokine concentrations.<sup>96</sup> The neuroinflammatory biomarker MMP-9 was highly correlated with poor prognosis of patients with ischemic stroke.<sup>97,98</sup> Moreover, atorvastatin and rosuvastatin exhibited in TBI patients neuroprotective effects with better functional outcomes due to the antiinflammatory properties of statins.<sup>99,100</sup>

The efforts of limiting neuroinflammation after ABI are rewarded by improved neurological recovery, therefore, attracting increasing attention and interest in the exploration of novel neuroinflammatory modulators.

# The emerging role of ATF3 in ABI associated neuroinflammation

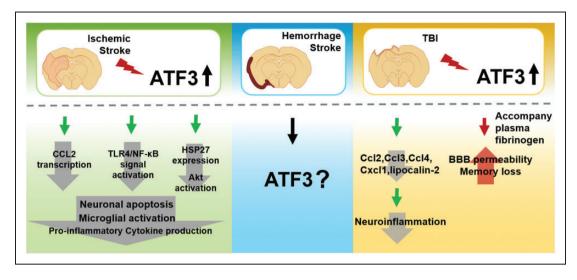
Stroke, divided into cerebral ischemic and hemorrhagic strokes, is, together with TBI the most typical ABI which is characterized byneuroinflammation.<sup>101,102</sup> Subsequently, the activation of cerebral resident neural and peripheral immune cells amplifies neuroinflammatory responses due to the compromised

permeability of the blood brain barrier (BBB) and the proinflammatory cytokine storm.<sup>103,104</sup>

Evidence shows that ATF3 can be expressed in microglia,<sup>105,106</sup> brain ECs,<sup>67,107</sup> and neurons<sup>9,108</sup> in the CNS and can exert effects on neuroinflammatory modulators to affect brain physiologic function or development.<sup>31,32</sup> Similarly, the expression of ATF3 in peripheral immune cells, such as macrophages, is involved in peripheral inflammation.<sup>57</sup> Thus, the underlying regulatory mechanism of ATF3 in inflammation is highly correlated with its participation in neuroinflammation.

# ATF3 inhibits neuronal apoptosis following ischemic stroke

ATF3 has been implicated in the cellular response to ischemia/reperfusion of the brain,<sup>5</sup> which features by inflammatory responses<sup>109–111</sup> (Figure 2). Accumulating evidence suggests that ATF3 is activated in the ischemic brain.<sup>32,112,113</sup> Recently, ATF3 as an immediate early gene was proposed as a potential biomarker for diagnosis and prognosis in elderly women with focal ischemia.<sup>1,32,112,113</sup> Studies discovered that gene transcript and protein levels of ATF3 were increased at the acute stage after ischemia/reperfusion, inhibiting glutamate-induced



**Figure 2.** The function of ATF3 on acute brain injury. In the ischemic stroke, ATF3 overexpression attenuated neuronal caspasedependent apoptosis, microglial activation, and pro-inflammatory cytokine production to alleviate brain injury. ATF3 can block CCL2 transcription and TLR4/NF- $\kappa$ B signal activation to reduce the activation of microglia and prevent neuronal apoptosis. Another is to prevent neuronal apoptosis induced by c-Jun N-terminal kinase by promoting the expression of the anti-apoptotic neuronal survival factor HSP27 and activation of Akt. There is a lack of knowledge regarding the regulation of ATF3 in hemorrhagic stroke. In TBI, upregulation of ATF3 in the brain negatively regulates CCL and CXCL chemokines (Ccl2, Ccl3, Ccl4, and Cxcl1) and lipocalin-2 expression, which then alleviates neuroinflammation after TBI. But increased BBB permeability and loss of memory after TBI are both caused by Fg-induced overexpression of ATF3.

TLR: toll like receptor; HSP: heat shock protein; TBI: traumatic brain injury; CcI: C-C motif ligand; CxcI: CXC chemokine ligand; Fg: plasma fibrinogen; BBB: blood brain barrier.

apoptosis.<sup>114</sup> Reasonably, knockout of ATF3 significantly increased the infarct volume, worsening neurological function, as well as eliciting the cellular inflammatory response 3 days after transient middle cerebral artery occlusion (MCAO).<sup>33</sup> Although it was reported that ATF3 expression was decreased 3 days after MCAO,<sup>32</sup> ATF3 overexpression attenuated neuronal caspasedependent apoptosis, microglial activation, and proinflammatory cytokine production to alleviate brain injury.<sup>32,113</sup> For example, ATF3 can block CCL2 transcription and TLR4/NF-kB signal activation to reduce the activation of microglia and prevent neuronal apoptosis. Another is to prevent neuronal apoptosis induced by c-Jun N-terminal kinase by promoting the expression of the anti-apoptotic neuronal survival factor HSP27 and activation of Akt.<sup>32,113</sup> As mentioned above, ATF3 can be therapeutically targeted to modulate ischemic stroke. However, existing evidence of ATF3 regulation is currently described only for ischemic, not hemorrhagic stroke. No research has investigated the role of ATF3 during the chronic phase of inflammation. Thus, further research is required in the future.

# TBI induces upregulation of ATF3 and exerts dualistic roles in neuroinflammation and BBB damage

TBI triggers an inflammatory response in the CNS and poses a substantial risk for neurodegenerative

diseases.<sup>115,116</sup> Previous studies shown that ATF3 is induced 2 hours after TBI<sup>31,117,118</sup> and rapidly upregulated in neuronal and immune cells, thereby regulating neuroinflammation<sup>34</sup> (Figure 2). Similar to ischemic stroke models, the upregulation of ATF3 occurs within 1-2 hours after TBI and decreases 2-3 days later.<sup>34</sup> Functionally, ATF3 is necessary to inhibit the production of CCL and CXCL chemokines (CCL2, CCL3, CCL4, and CXCL1), as well as lipocalin-2, subsequently reducing neuroinflammation in the subacute phase of TBI.<sup>34</sup> However, ATF3 overexpression following TBI caused by plasma fibrinogen is associated with increased BBB permeability<sup>119</sup> and memory loss. The fibrinogen inhibitor fibrinogen antisense oligonucleotide can decrease inflammatory responses and BBB permeability, thereby ameliorating neurological diseases.<sup>120</sup> Further studies are warranted to illustrate more detailed regulatory mechanisms of ATF3 in TBI owing to its dual effects in inflammatory responses.

## Conclusion

Neuroinflammation is one of the key pathological features of a wide range of ABIs. Unresolved neuroinflammation continuously shapes the evolving brain pathology and affects the long-term neurological outcomes of affected patients.<sup>16,30</sup> Neuroinflammation is an emerging therapeutic target for improving neurological function following ABI.

As an intermediate modulator in various pathological contexts, ATF3 plays a pivotal role in the regulation of inflammatory responses. In this mini review, we described that ATF3, a stress-associated transcription factor, can be rapidly induced in the brain after ABI and detail multiple ways by which ATF3 can participate in neuroimmune reactions after ABI, e.g., modulating the production of inflammatory mediators and altering cerebral vascular integrity. ATF3 might be therapeutically targeted to combat excessive inflammatory responses to modulate brain damage. However, there is a lack of knowledge regarding the regulation of ATF3 in hemorrhagic stroke and subarachnoid hemorrhage, in which neuroinflammation plays a critical role in determining neurological functions. Further studies are highly warranted to expand our knowledge of ATF3 regulation following hemorrhagic stroke and subarachnoid hemorrhage. Moreover, the differential role of ATF3 in acute versus chronic inflammation after brain injury should be explored in the pursuit of providing a framework to develop novel therapies for ABI.

### Funding

The author(s) disclosed receipt of the following financial support for the research, authorship, and/or publication of this article: W.Y is supported by the National Natural Science Foundation of China (32030043,81971223), Shanghai Municipal Key Clinical Specialty (shslczdzk03601), Shanghai Engineering Research Center of Peri-operative Organ Support and Function Preservation (20DZ2254200), Area Municipal Commission of Health and Family Planning Funding (PWZxq2017-06), National Key R&D Program (2018YFA0108204), the Shanghai Municipal Education Commission (Grant number 2019-01-07-00-01-E00074). P.L. is supported by the National Natural Science Foundation of China (NSFC 91957111, 81971096, 8206 1130224. M-0671), New Frontier Technology Joint Research sponsored by Shanghai Shenkang Hospital Development Center (SHDC12019102), Shanghai Municipal Education Commission-Gaofeng Clinical Medical Grant Support (20181805), Shuguang Program supported by Shanghai Education Development Foundation and Shanghai Municipal Education Commission (20SG17), and Shanghai Outstanding Academic Leaders' Program Shanghai Municipal Science and Technology from Committee (20XD1422400), Newton Advanced Fellowship grant provided by the UK Academy of Medical Sciences (NAF\R11\1010) P.L. is also supported by SHSMU-ZLCX20211602, Innovative Research Team of High-level Local Universities in Shanghai. W.X. is supported by the National Natural Science Foundation of China (NSFC, 81901985). X. W. is supported by the National Natural Science Foundation of China (NSFC, 82001381) and Shanghai Sailing Program (20YF1425400). This work is also supported by Clinical Research Plan of SHDC.

### Acknowledgements

Thank the authors of the references

### **Declaration of conflicting interests**

The author(s) declared no potential conflicts of interest with respect to the research, authorship, and/or publication of this article.

### **ORCID** iD

Peiying Li (D) https://orcid.org/0000-0002-5721-9914

### References

- Zhou H, Li N, Yuan Y, et al. Activating transcription factor 3 in cardiovascular diseases: a potential therapeutic target. *Basic Res Cardiol* 2018; 113: 37.
- 2. Hai T, Wolfgang CD, Marsee DK, et al. ATF3 and stress responses. *Gene Expr* 1999; 7: 321–335.
- 3. Liu H, Mo H, Yang C, et al. A novel function of ATF3 in suppression of ferroptosis in mouse heart suffered ischemia/reperfusion. *Free Radic Biol Med* 2022; 189: 122–135.
- 4. Wei X, Deng W, Dong Z, et al. Identification of subtypes and a delayed graft function predictive signature based on ferroptosis in renal ischemia-reperfusion injury. *Front Cell Dev Biol* 2022; 10: 800650.
- Lin H and Cheng CF. Activating transcription factor 3, an early cellular adaptive responder in ischemia/ reperfusion-induced injury. *Ci Ji Yi Xue Za Zhi* 2018; 30: 61–65.
- Borgesius NZ, de Waard MC, van der Pluijm I, et al. Accelerated age-related cognitive decline and neurodegeneration, caused by deficient DNA repair. *J Neurosci* 2011; 31: 12543–12553.
- Zhou YW and Wu Y. Substrate viscoelasticity amplifies distinctions between transient and persistent LPSinduced signals. Adv Healthc Mater 2022; 11: e2102271.
- Paul D, Dixit AB, Srivastava A, et al. Altered expression of activating transcription factor 3 in the hippocampus of patients with mesial temporal lobe epilepsy-hippocampal sclerosis (MTLE-HS). *Int J Neurosci* 2022; 7: 1–7.
- Tian F, Cheng Y, Zhou S, et al. Core transcription programs controlling injury-induced neurodegeneration of retinal ganglion cells. *Neuron* 2022; 110: 2607–2624.e8.
- Tang Y, Liu C, Zhu T, et al. Transcriptome profiles of IncRNA and mRNA highlight the role of ferroptosis in chronic neuropathic pain with memory impairment. *Front Cell Dev Biol* 2022; 10: 843297.
- Wu X, Qiu F, Jin X, et al. ATF3: a novel biomarker for the diagnosis of acute kidney injury after cardiac surgery. *Ann Transl Med* 2021; 9: 1655.
- 12. Hu H, Zhang F, Li L, et al. Identification and validation of ATF3 serving as a potential biomarker and correlating with pharmacotherapy response and immune infiltration characteristics in rheumatoid arthritis. *Front Mol Biosci* 2021; 8: 761841.

- Wang K, Wang S, Chen Y, et al. Single-cell transcriptomic analysis of somatosensory neurons uncovers temporal development of neuropathic pain. *Cell Res* 2021; 31: 904–918.
- Renthal W, Tochitsky I, Yang L, et al. Transcriptional reprogramming of distinct peripheral sensory neuron subtypes after axonal injury. *Neuron* 2020; 108: 128–144 e129.
- Eleftheriadis T, Pissas G, Golfinopoulos S, et al. Role of indoleamine 2,3-dioxygenase in ischemia-reperfusion injury of renal tubular epithelial cells. *Mol Med Rep* 2021; 23: 472.
- Endres M, Moro MA, Nolte CH, et al. Immune pathways in etiology, acute phase, and chronic sequelae of ischemic stroke. *Circ Res* 2022; 130: 1167–1186.
- Mastorakos P, Russo MV, Zhou T, et al. Antimicrobial immunity impedes CNS vascular repair following brain injury. *Nat Immunol* 2021; 22: 1280–1293.
- Drieu A, Lanquetin A, Prunotto P, et al. Persistent neuroinflammation and behavioural deficits after single mild traumatic brain injury. J Cereb Blood Flow Metab 2022; 42: 2216–2229.
- Xue M and Yong VW. Neuroinflammation in intracerebral haemorrhage: immunotherapies with potential for translation. *Lancet Neurol* 2020; 19: 1023–1032.
- De Nardo D, Labzin LI, Kono H, et al. High-density lipoprotein mediates anti-inflammatory reprogramming of macrophages via the transcriptional regulator ATF3. *Nat Immunol* 2014; 15: 152–160.
- Ntaios G, Wintermark M and Michel P. Supracardiac atherosclerosis in embolic stroke of undetermined source: the underestimated source. *Eur Heart J* 2021; 42: 1789–1796.
- Zheng RZ, Lee KY, Qi ZX, et al. Neuroinflammation following traumatic brain injury: take it seriously or not. *Front Immunol* 2022; 13: 855701.
- Candelario-Jalil E, Dijkhuizen RM and Magnus T. Neuroinflammation, stroke, blood-brain barrier dysfunction, and imaging modalities. *Stroke* 2022; 53: 1473–1486.
- Ding P, Chen W, Yan X, et al. BMPER alleviates ischemic brain injury by protecting neurons and inhibiting neuroinflammation via Smad3-Akt-Nrf2 pathway. CNS Neurosci Ther 2022; 28: 593–607.
- Prabhakar NK, Khan H, Grewal AK, et al. Intervention of neuroinflammation in the traumatic brain injury trajectory: in vivo and clinical approaches. *Int Immunopharmacol* 2022; 108: 108902.
- Stuckey SM, Ong LK, Collins-Praino LE, et al. Neuroinflammation as a key driver of secondary neurodegeneration following stroke? *IJMS* 2021; 22: 13101.
- Sanchez-Bezanilla S, Hood RJ, Collins-Praino LE, et al. More than motor impairment: a spatiotemporal analysis of cognitive impairment and associated neuropathological changes following cortical photothrombotic stroke. *J Cereb Blood Flow Metab* 2021; 41: 2439–2455.
- Stoll G and Nieswandt B. Thrombo-inflammation in acute ischaemic stroke – implications for treatment. *Nat Rev Neurol* 2019; 15: 473–481.

- 29. Daglas M, Truong P, Miles LQ, et al. Deferiprone attenuates neuropathology and improves outcome following traumatic brain injury. *Br J Pharmacol* 2023; 180: 214–234.
- Levard D, Buendia I, Lanquetin A, et al. Filling the gaps on stroke research: focus on inflammation and immunity. *Brain Behav Immun* 2021; 91: 649–667.
- Forstner P and Knoll B. Interference of neuronal activity-mediated gene expression through serum response factor deletion enhances mortality and hyperactivity after traumatic brain injury. *Faseb J* 2020; 34: 3855–3873.
- 32. Song DY, Oh KM, Yu HN, et al. Role of activating transcription factor 3 in ischemic penumbra region following transient middle cerebral artery occlusion and reperfusion injury. *Neurosci Res* 2011; 70: 428–434.
- Wang L, Deng S, Lu Y, et al. Increased inflammation and brain injury after transient focal cerebral ischemia in activating transcription factor 3 knockout mice. *Neuroscience* 2012; 220: 100–108.
- 34. Forstner P, Rehman R, Anastasiadou S, et al. Neuroinflammation after traumatic brain injury is enhanced in activating transcription factor 3 mutant mice. *J Neurotrauma* 2018; 35: 2317–2329.
- 35. Hai T, Wolford C and Chang Y. ATF3, a hub of the cellular adaptive-response network, in the pathogenesis of diseases: is modulation of inflammation a unifying component? *Gene Expr* 2010; 15: 1–11.
- Li Z, Gurung M, Rodrigues RR, et al. Microbiota and adipocyte mitochondrial damage in type 2 diabetes are linked by Mmp12<sup>+</sup> macrophages. *J Exp Med* 2022; 219: 20220603.
- Chen M, Liu Y, Yang Y, et al. Emerging roles of activating transcription factor (ATF) family members in tumourigenesis and immunity: Implications in cancer immunotherapy. *Genes Dis* 2022; 9: 981–999.
- Guo W, Wu Z, Chen J, et al. Nanoparticle delivery of miR-21-3p sensitizes melanoma to anti-PD-1 immunotherapy by promoting ferroptosis. *J Immunother Cancer* 2022; 10: e004381.
- Dai Z, Wang X, Peng R, et al. Induction of IL-6Ralpha by ATF3 enhances IL-6 mediated sorafenib and regorafenib resistance in hepatocellular carcinoma. *Cancer Lett* 2022; 524: 161–171.
- Li Y, Li Z, Zhang C, et al. Cardiac fibroblast-specific activating transcription factor 3 protects against heart failure by suppressing MAP2K3-p38 signaling. *Circulation* 2017; 135: 2041–2057.
- Rohini M, Haritha Menon A and Selvamurugan N. Role of activating transcription factor 3 and its interacting proteins under physiological and pathological conditions. *Int J Biol Macromol* 2018; 120: 310–317.
- Gilchrist M, Thorsson V, Li B, et al. Systems biology approaches identify ATF3 as a negative regulator of toll-like receptor 4. *Nature* 2006; 441: 173–178.
- Hoetzenecker W, Echtenacher B, Guenova E, et al. ROS-induced ATF3 causes susceptibility to secondary infections during sepsis-associated immunosuppression. *Nat Med* 2011; 18: 128–134.

- 44. Jadhav K and Zhang Y. Activating transcription factor 3 in immune response and metabolic regulation. *Liver Res* 2017; 1: 96–102.
- 45. Thompson MR, Xu D and Williams BR. ATF3 transcription factor and its emerging roles in immunity and cancer. J Mol Med (Berl) 2009; 87: 1053–1060.
- 46. Kong Q, Zou J, Zhang Z, et al. BAF60a deficiency in macrophage promotes diet-induced obesity and metabolic inflammation. *Diabetes* 2022; 71: 2136–2152.
- Khuu CH, Barrozo RM, Hai T, et al. Activating transcription factor 3 (ATF3) represses the expression of CCL4 in murine macrophages. *Mol Immunol* 2007; 44: 1598–1605.
- Nobori K, Ito H, Tamamori-Adachi M, et al. ATF3 inhibits doxorubicin-induced apoptosis in cardiac myocytes: a novel cardioprotective role of ATF3. *J Mol Cell Cardiol* 2002; 34: 1387–1397.
- Bhattacharya B, Xiao S, Chatterjee S, et al. The integrated stress response mediates necrosis in murine *Mycobacterium tuberculosis* granulomas. J Clin Invest 2021; 131: e130319.
- Du Y, Ma Z, Zheng J, et al. ATF3 positively regulates antibacterial immunity by modulating macrophage killing and migration functions. *Front Immunol* 2022; 13: 839502.
- Liu H, Kuang X, Zhang Y, et al. ADORA1 inhibition promotes tumor immune evasion by regulating the ATF3-PD-L1 axis. *Cancer Cell* 2020; 37: 324–339 e328.
- Zhou M, Ou Y, Wu G, et al. Transcriptomic analysis reveals that Atf3/c-Jun/Lgals3 axis is associated with central diabetes insipidus after hypothalamic injury. *Neuroendocrinology* 2022; 112: 874–893.
- Bambouskova M, Gorvel L, Lampropoulou V, et al. Electrophilic properties of itaconate and derivatives regulate the IkappaBzeta-ATF3 inflammatory axis. *Nature* 2018; 556: 501–504.
- 54. More VR, Campos CR, Evans RA, et al. PPAR-alpha, a lipid-sensing transcription factor, regulates bloodbrain barrier efflux transporter expression. J Cereb Blood Flow Metab 2017; 37: 1199–1212.
- Zhang Y, Li F, Chen C, et al. RAGE-mediated T cell metabolic reprogramming shapes T cell inflammatory response after stroke. *J Cereb Blood Flow Metab* 2022; 42: 952–965.
- Kurita N, Yamashiro K, Kuroki T, et al. Metabolic endotoxemia promotes neuroinflammation after focal cerebral ischemia. J Cereb Blood Flow Metab 2020; 40: 2505–2520.
- Wang B, Yang X, Sun X, et al. ATF3 in atherosclerosis: a controversial transcription factor. J Mol Med 2022; 100: 1557–1568.
- Luo X, Ge J, Liu J, et al. TFCP2, a binding protein of ATF3, promotes the progression of glioma by activating the synthesis of serine. *Exp Cell Res* 2022; 416: 113136.
- Peace CG and O'Neill LAJ. The role of itaconate in host defense and inflammation. J Clin Invest 2022; 132: e148548.
- Zuo Z, Li L, Yan X, et al. Glucose starvation causes ptau(S409) increase in N2a cells through ATF3/

PKAcalpha signaling pathway. *Neurochem Res* 2022; 47: 3298–3308.

- Willemsen N, Arigoni I, Studencka-Turski M, et al. Proteasome dysfunction disrupts adipogenesis and induces inflammation via ATF3. *Mol Metab* 2022; 62: 101518.
- Maheshwari A, Norby FL, Roetker NS, et al. Refining prediction of atrial fibrillation-related stroke using the P2-CHA2DS2-VASc score. *Circulation* 2019; 139: 180–191.
- 63. Bjorkegren JLM and Lusis AJ. Atherosclerosis: recent developments. *Cell* 2022; 185: 1630–1645.
- 64. Georgakis MK, Bernhagen J, Heitman LH, et al. Targeting the CCL2-CCR2 axis for atheroprotection. *Eur Heart J* 2022; 43: 1799–1808.
- Clement M, Basatemur G, Masters L, et al. Necrotic cell sensor Clec4e promotes a proatherogenic macrophage phenotype through activation of the unfolded protein response. *Circulation* 2016; 134: 1039–1051.
- Gold E, Ramsey S, Sartain M, et al. ATF3 protects against atherosclerosis by suppressing 25-hydroxycholesterol-induced lipid body formation. *J Exp Med* 2012; 209: 807–817.
- Nyunt T, Britton M, Wanichthanarak K, et al. Mitochondrial oxidative stress-induced transcript variants of ATF3 mediate lipotoxic brain microvascular injury. *Free Radic Biol Med* 2019; 143: 25–46.
- Aung HH, Lame MW, Gohil K, et al. Induction of ATF3 gene network by triglyceride-rich lipoprotein lipolysis products increases vascular apoptosis and inflammation. *Arterioscler Thromb Vasc Biol* 2013; 33: 2088–2096.
- Zhang WY, Franco DA, Schwartz E, et al. HDL inhibits saturated fatty acid mediated augmentation of innate immune responses in endothelial cells by a novel pathway. *Atherosclerosis* 2017; 259: 83–96.
- Shichita T, Hasegawa E, Kimura A, et al. Peroxiredoxin family proteins are key initiators of post-ischemic inflammation in the brain. *Nat Med* 2012; 18: 911–917.
- Caso JR, Pradillo JM, Hurtado O, et al. Toll-like receptor 4 is involved in brain damage and inflammation after experimental stroke. *Circulation* 2007; 115: 1599–1608.
- 72. Askenase MH, Goods BA, Beatty HE, MISTIE III Consortium, et al. Longitudinal transcriptomics define the stages of myeloid activation in the living human brain after intracerebral hemorrhage. *Sci Immunol* 2021; 6: eabd6279.
- 73. Risbrough VB, Vaughn MN and Friend SF. Role of inflammation in traumatic brain Injury-Associated risk for neuropsychiatric disorders: State of the evidence and where do We go from here. *Biol Psychiatry* 2022; 91: 438–448.
- 74. Bergold PJ. Treatment of traumatic brain injury with anti-inflammatory drugs. *Exp Neurol* 2016; 275 Pt 3: 367–380.
- Simon DW, McGeachy MJ, Bayir H, et al. The farreaching scope of neuroinflammation after traumatic brain injury. *Nat Rev Neurol* 2017; 13: 171–191.

- 76. Toutonji A, Mandava M, Guglietta S, et al. Chronic complement dysregulation drives neuroinflammation after traumatic brain injury: a transcriptomic study. *Acta Neuropathol Commun* 2021; 9: 126.
- 77. Thelin EP, Tajsic T, Zeiler FA, et al. Monitoring the neuroinflammatory response following acute brain injury. *Front Neurol* 2017; 8: 351.
- Yu F, Wang Y, Stetler AR, et al. Phagocytic microglia and macrophages in brain injury and repair. CNS Neurosci Ther 2022; 28: 1279–1293.
- Liu J, Gu Y, Guo M, et al. Neuroprotective effects and mechanisms of ischemic/hypoxic preconditioning on neurological diseases. *CNS Neurosci Ther* 2021; 27: 869–882.
- Jin R, Yang G and Li G. Inflammatory mechanisms in ischemic stroke: role of inflammatory cells. *J Leukoc Biol* 2010; 87: 779–789.
- Aungst SL, Kabadi SV, Thompson SM, et al. Repeated mild traumatic brain injury causes chronic neuroinflammation, changes in hippocampal synaptic plasticity, and associated cognitive deficits. *J Cereb Blood Flow Metab* 2014; 34: 1223–1232.
- Kumar A and Loane DJ. Neuroinflammation after traumatic brain injury: opportunities for therapeutic intervention. *Brain Behav Immun* 2012; 26: 1191–1201.
- Stevens SL, Bao J, Hollis J, et al. The use of flow cytometry to evaluate temporal changes in inflammatory cells following focal cerebral ischemia in mice. *Brain Res* 2002; 932: 110–119.
- Alawieh A, Langley EF, Weber S, et al. Identifying the role of complement in triggering neuroinflammation after traumatic brain injury. *J Neurosci* 2018; 38: 2519–2532.
- Yang S, Nakamura T, Hua Y, et al. The role of complement C3 in intracerebral hemorrhage-induced brain injury. J Cereb Blood Flow Metab 2006; 26: 1490–1495.
- Wang M, Xia F, Wan S, et al. Role of complement component 3 in early erythrolysis in the hematoma after experimental intracerebral hemorrhage. *Stroke* 2021; 52: 2649–2660.
- Pu H, Ma C, Zhao Y, et al. Intranasal delivery of interleukin-4 attenuates chronic cognitive deficits via beneficial microglial responses in experimental traumatic brain injury. *J Cereb Blood Flow Metab* 2021; 41: 2870–2886.
- Chen J, Lin X, Yao C, et al. Transplantation of roxadustat-preconditioned bone marrow stromal cells improves neurological function recovery through enhancing grafted cell survival in ischemic stroke rats. *CNS Neurosci Ther* 2022; 28: 1519–1531.
- Laird MD, Shields JS, Sukumari-Ramesh S, et al. High mobility group box protein-1 promotes cerebral edema after traumatic brain injury via activation of toll-like receptor 4. *Glia* 2014; 62: 26–38.
- Hyakkoku K, Hamanaka J, Tsuruma K, et al. Toll-like receptor 4 (TLR4), but not TLR3 or TLR9, knock-out mice have neuroprotective effects against focal cerebral ischemia. *Neuroscience* 2010; 171: 258–267.
- 91. Khan ZA, Sumsuzzman DM, Choi J, et al. Pre- and post-conditioning with poly I:C exerts neuroprotective effect against cerebral ischemia injury in animal models:

a systematic review and meta-analysis. CNS Neurosci Ther 2022; 28: 1168–1182.

- Lyu J, Xie D, Bhatia TN, et al. Microglial/macrophage polarization and function in brain injury and repair after stroke. *CNS Neurosci Ther* 2021; 27: 515–527.
- 93. Liu FY, Cai J, Wang C, et al. Fluoxetine attenuates neuroinflammation in early brain injury after subarachnoid hemorrhage: a possible role for the regulation of TLR4/MyD88/NF-kappaB signaling pathway. *J Neuroinflamm* 2018; 15: 347.
- 94. Zhu HT, Bian C, Yuan JC, et al. Curcumin attenuates acute inflammatory injury by inhibiting the TLR4/ MyD88/NF-kappaB signaling pathway in experimental traumatic brain injury. J Neuroinflamm 2014; 11: 59.
- Liu Y, Fu H and Wang T. Neuroinflammation in perioperative neurocognitive disorders: from bench to the bedside. *CNS Neurosci Ther* 2022; 28: 484–496.
- 96. Algra SO, Groeneveld KM, Schadenberg AW, et al. Cerebral ischemia initiates an immediate innate immune response in neonates during cardiac surgery. *J Neuroinflamm* 2013; 10: 24.
- 97. Gori AM, Giusti B, Piccardi B, et al. Inflammatory and metalloproteinases profiles predict three-month poor outcomes in ischemic stroke treated with thrombolysis. *J Cereb Blood Flow Metab* 2017; 37: 3253–3261.
- Li Y, Han X, Luo S, et al. Predictive value of longitudinal changes of serum matrix metalloproteinase-9 and brain-derived neurotrophic factor in acute ischemic stroke. *Front Aging Neurosci* 2022; 14: 952038. 20220825. DOI: 10.3389/fnagi.2022.952038.
- Govindarajan KA, Narayana PA, Hasan KM, et al. Cortical thickness in mild traumatic brain injury. *J Neurotrauma* 2016; 33: 1809–1817.
- Sanchez-Aguilar M, Tapia-Perez JH, Sanchez-Rodriguez JJ, et al. Effect of rosuvastatin on cytokines after traumatic head injury. JNS 2013; 118: 669–675.
- 101. Feigin VL, Stark BA, Johnson CO, et al. Global, regional, and national burden of stroke and its risk factors, 1990-2019: a systematic analysis for the global burden of disease study 2019. *Lancet Neurol* 2021; 20: 795–820.
- 102. James SL, Theadom A, Ellenbogen RG, et al. Global, regional, and national burden of traumatic brain injury and spinal cord injury, 1990-2016: a systematic analysis for the global burden of disease study 2016. *Lancet Neurol* 2019; 18: 56–87.
- Rebelo AL, Chevalier MT, Russo L, et al. Role and therapeutic implications of protein glycosylation in neuroinflammation. *Trends Mol Med* 2022; 28: 270–289.
- 104. Cucullo L, Marchi N, Hossain M, et al. A dynamic in vitro BBB model for the study of immune cell trafficking into the central nervous system. J Cereb Blood Flow Metab 2011; 31: 767–777.
- 105. Zhou M, Ou Y, Wu G, et al. Transcriptomic analysis reveals that activating transcription factor 3/c-Jun/ Lgals3 axis is associated with Central diabetes insipidus after hypothalamic injury. *Neuroendocrinology* 2022; 112: 874–893.
- McGill MM, Richman AR, Boyd JR, et al. p38 MAP kinase signaling in microglia plays a Sex-Specific protective

role in CNS autoimmunity and regulates microglial transcriptional states. *Front Immunol* 2021; 12: 715311.

- 107. Fan Z, Turiel G, Ardicoglu R, et al. Exercise-induced angiogenesis is dependent on metabolically primed ATF3/4(+) endothelial cells. *Cell Metab* 2021; 33: 1793–1807 e1799.
- Loganathan N, McIlwraith EK and Belsham DD. Bisphenol a induces agrp gene expression in hypothalamic neurons through a mechanism involving ATF3. *Neuroendocrinology* 2021; 111: 678–695.
- 109. Hu X, Leak RK, Shi Y, et al. Microglial and macrophage polarization-new prospects for brain repair. *Nat Rev Neurol* 2015; 11: 56–64.
- 110. Zhang Y, Lian L, Fu R, et al. Microglia: the hub of intercellular communication in ischemic stroke. *Front Cell Neurosci* 2022; 16: 889442.
- 111. Li Y, Zhu ZY, Huang TT, et al. The peripheral immune response after stroke-A double edge sword for blood-brain barrier integrity. *CNS Neurosci Ther* 2018; 24: 1115–1128.
- 112. van der Weerd L, Tariq Akbar M, Aron Badin R, et al. Overexpression of heat shock protein 27 reduces cortical damage after cerebral ischemia. J Cereb Blood Flow Metab 2010; 30: 849–856.
- 113. Ma N, Li G and Fu X. Protective role of activating transcription factor 3 against neuronal damage in rats with cerebral ischemia. *Brain Behav* 2022; 12: e2522.

- 114. Takarada T, Kou M, Hida M, et al. Protective upregulation of activating transcription factor-3 against glutamate neurotoxicity in neuronal cells under ischemia. *J Neurosci Res* 2016; 94: 378–388.
- Russo MV and McGavern DB. Inflammatory neuroprotection following traumatic brain injury. *Science* 2016; 353: 783–785.
- 116. Brett BL, Gardner RC, Godbout J, et al. Traumatic brain injury and risk of neurodegenerative disorder. *Biol Psychiatry* 2022; 91: 498–507.
- 117. Natale JE, Ahmed F, Cernak I, et al. Gene expression profile changes are commonly modulated across models and species after traumatic brain injury. *J Neurotrauma* 2003; 20: 907–927.
- 118. Losurdo M, Davidsson J and Skold MK. Diffuse axonal injury in the rat brain: Axonal injury and oligodendrocyte activity following rotational injury. *Brain Sci* 2020; 10: 229.
- Petersen MA, Ryu JK and Akassoglou K. Fibrinogen in neurological diseases: mechanisms, imaging and therapeutics. *Nat Rev Neurosci* 2018; 19: 283-301.
- 120. Muradashvili N, Charkviani M, Sulimai N, et al. Effects of fibrinogen synthesis inhibition on vascular cognitive impairment during traumatic brain injury in mice. *Brain Res* 2021; 1751: 147208.