

Gene Cloning and Characterization of a Novel Cellulose-Binding β -Glucosidase from *Phanerochaete chrysosporium*

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Analysis of a 2.4-kb cDNA of the cellulose-binding extracellular β -glucosidase (CBGL) from *Phanerochaete chrysosporium* suggested that CBGL is organized into two domains, an N-terminal cellulose-binding domain and a C-terminal catalytic domain. Genomic sequence analysis suggested that *cbgl* is encoded by 30 exons. Southern analysis of DNA from homokaryotic cultures indicated that CBGL is encoded by two alleles, *cbgl-1* and *cbgl-2*, of a single gene.

Cellulose-degrading cultures of the white rot basidiomycete *Phanerochaete chrysosporium* apparently produce three different β -glucosidases—extracellular, intracellular, and cell wall bound—depending on the carbon source (9, 26). Deshpande et al. (9) reported that cellulose induces intracellular and cell wall-bound enzymes and purified five isozymes of extracellular β -glucosidases from cellulose-degrading cultures of *P. chrysosporium*. Molecular weights of these glucosidases ranged from 165,000 to 182,000. Smith and Gold (26) partially purified an extracellular β -glucosidase from *P. chrysosporium* OGC101 and characterized it as a monomer with a molecular weight of 90,000. Recently, we purified and characterized a cellulose-binding extracellular β -glucosidase (CBGL) with a molecular mass of 114,000 from cellulose-supplemented cultures of *P. chrysosporium* OGC101. When CBGL was treated with papain, its molecular weight decreased to 95,000; it lost the ability to bind to cellulose, but its catalytic activity was unchanged. This suggested that CBGL is organized into two domains—a cellulose-binding domain (CBD) and a catalytic domain (20). The glucosidase isolated previously by Smith and Gold (26) from this strain might be the non-cellulose-binding form. The kinetic properties of the cellulose-binding and nonbinding forms were similar, indicating that the CBD was not involved in catalysis. Here cloning and characterization of a cDNA clone and a genomic clone of CBGL are reported. Sequence analysis confirmed our prediction that this β -glucosidase consists of a catalytic domain and a CBD.

Organism. *P. chrysosporium* OGC101 (a derivative of BKM-F-1767) was obtained from Michael H. Gold (Oregon Graduate Institute) (1). *Escherichia coli* XL1-Blue MRF' and SOLR were obtained from Stratagene (La Jolla, Calif.).

Nucleotides. Oligonucleotides were prepared by the Oregon Regional Primate Research Center (Beaverton, Ore.). The plasmid isolation kit was obtained from Qiagen, Inc. (Chatsworth, Calif.).

Isolation of a cDNA clone of *cbgl*. The cDNA λ ZAP expression library, prepared as described previously (18), was screened with an anti-CBGL antibody and a secondary an-

tibody labeled with alkaline phosphatase. The pBluescript SK(-) plasmid containing a putative β -glucosidase cDNA insert was rescued by in vivo excision with a helper phage. The plasmid was purified with a commercial plasmid isolation kit (Qiagen, Inc.). The cDNA was sequenced by the dideoxy method with the primer walking strategy (25, 27).

Isolation of a genomic clone of *cbgl*. A λ EMBL3 genomic library of *P. chrysosporium* OGC101 was screened at high stringency (4.8 \times SSC [1 \times SSC is 0.15 M NaCl plus 0.015 M sodium citrate]–48% formamide at 50°C) with a 550-bp *Apa*I fragment from the 3' end of the *cbgl* cDNA clone. Based on the restriction mapping of the genomic clones, four overlapping restriction fragments (3.6-kb *Sac*I, 1.7-kb *Sac*I, 4.5-kb *Sal*I, and 1.2-kb *Sal*I) covering the entire region of *cbgl* were subcloned into pBluescript SK (Stratagene) and sequenced by the primer walking method (27). Sequencing was performed with an automatic sequencer (model 377; Applied Biosystems) and *Ampli*Taq DNA polymerase, FS.

Isolation of a full-length cDNA clone of *cbgl*. The cDNA library of *P. chrysosporium* was probed with a restriction fragment (66 to 324 bp) obtained by digesting *cbgl-2* with *Msc*I and *Nde*I. Hybridization was performed at high stringency (4.8 \times SSC, 48% formamide, 50°C). Positive clones were purified by further screening. The pBluescript II SK plasmid containing the putative *cbgl* cDNA insert was rescued by in vivo excision with a helper phage. The plasmid was purified with a plasmid midi kit (Qiagen, Inc.).

Isolation and analysis of homokaryons. Single homokaryotic basidiospores were isolated as described previously (1, 12). DNAs from homokaryotic cultures were isolated by standard procedures and restriction digested with *Sal*I, size fractionated in a 0.7% agarose gel, blotted onto a Magnagraph nylon transfer membrane (Micron Separations, Westboro, Mass.), and probed with a ³²P-labeled 1.4-kb *Sac*I fragment of *cbgl* (nucleotides 1446 to 2866).

Northern (RNA) blot analysis. Total RNA was isolated from 11-day-old mycelia of *P. chrysosporium* cultured with 1% cotton linters, cellobiose, or glucose as the carbon source (2, 8). RNA was electrophoresed in 1.5% agarose gel containing 2.2 M formaldehyde, transferred to Magnagraph nylon membranes (Micron Separations), and probed with cDNA for CBGL at 42°C as described previously (6).

Southern blot analysis of *cbgl*. DNA from *P. chrysosporium* was restriction digested and electrophoresed with a 0.7% agarose gel. The DNA was transferred to Magnagraph nylon mem-

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TCGAAGTTGCCGCTGTTTGTCCGCGCCTTATCCGCTCGACGCGGCTGCTGTGGGCAACGACGTTGGCCAAAGACATTTCCGATCTACATATCGACTTCTTCAAACATGTTTCGAGAAGTAT 120
AAGTGCCTTTGACCATGCGTACGCTGTCTACGCAAGGACGCCCTCTCCCTACGCTGACAAAATGGGACTGACACTGGTCTTATACATCTTGCTTAGGTTGGTCTGGGGTCC 240
M G L T L V V L L H L A L G L L T G V 19
AAGCTCAGTCCGCGCTGTATCAGCAATGTGGCGGTATCCGGATGgttagctggtctctctcctcgtccggatggtgatgctcatatgttctcgcgatgacGACTGGTCAACTGTG 360
Q A Q S G L Y Q Q C G G I G W T G A T T C 0

CGTCAGCGCGCGACTGCACAGTGCAGAACCCATgtacgtgcacacagatcatgtcacagcagcagctcatgacacgtagACTACTCGCAATGTCTCCCTGGCGCCGCCACCAG 480
V S G A T C T V L N P Y Y S Q C L P G A A T T 63

AGTGTTCCTAGTTCATTCGTCAGGTCAGCAGCCTCCAGCAGTGCAGCAGCAGCAGCATAAGTTCTACTAGCACCAGCCCCGGCGCCCTCCAGCCCGTGGCGAAC 600
S V S S S H S S S S S V S S S H S S S A S S S I S S T S T S P P A P S Q T V A N 103
(E)

GTGTCTCCAGAATGGGCGCGCTTACGTCAAAGTgtggaacgagctcttcagatgtatattgtgatgtcactgaaacacgcgcagGCACAAGCTGCTGTAGCAAGCTATCCGGTACTGA 720
V S P E W A A A Y V K A Q A A V A K L S V T D 126

TATGGTCAACTCGCAACAGTgagatgactogtgattaactctctcttaacaactcttgccagGTGTGCAGTGGCAGAAAGTCCATGTTCGGCAACACCCTGCTATCTCGTC 840
M V N L A T G V Q W Q K G P C V G N T P A I S S 150
TATACCCGTTTACTGGACTTTCGCTGCAAGgtatgcttcaacgtccctacagatgtgtgagctgacttattccatggtccacagACAGTCCCGTGGTCCGCTACGCTGATGGA 960
I P G F T G L C L Q D S P V G V R Y A D G 171

ACGTGTCTTCCCACCgtacgtgctgagcagatgctgtaactctactcctggtgaaacagttacctacagTGAATCAACGTGTGCTACATGGAACCGTACGCTTATTCGCCAA 1080
T S V F P P E I N V A A T W N R T L M R Q 192

CGCGAGCTGCTATGGGAGCGGAATTTAAGGGGAAAGGCGTGCACGTCGCTCTCGGTCCCATGATGAACCTCATGCGGTGCCCCGGCTGGCGAAACTGGGAAGGgtgagttctttg 1200
R G A A M T A A F K G K G V H V A L G P M M N L M R V P A A G R N W E G 228

gagtggttacacgctgtgcggcagctacttaattctcctgcttggattgtagTGGCGGTGTGATCCTTCTCTGGCGAAGTGGCGTTCGAGACCATCTGGCATTGCTCT 1320
G G G D P F L S G E V A F E T I T G I Q S 249
(S)

CCGTTGCCAGGCTTGTGCCAAGCATTTTCATCAACAAGTgctgttctctctcctgagcttcgcttactgagctattatgagcGAGCAAGAACAATTTAGGGACTCCAGTTCAT 1440
S G A Q A C A K H F I N N E Q E H F R D S S S 272
CTAACGTCGACGATAGgtgagtttaccgtgttcgacagtcgggaaagtacgtgacctgctctcatagGACGgtgagataatcgtagtttatatgacatgcttctcaaggtgt 1560
S N V D D R T 279
agGACATGAATCTACGGACCCCTGtaaatggttctctctcattcgcctcaccgactctgacatcaatagTTCCTCGTAGCTCCAGGCAAACTAGTTCCTGTAATGTGCAAGC 1680
E H E L Y G H P F L R S V Q A N V A S V M C S 302
TACAgtaagttgtagctcagcctcgtgcagacgcaactaaatcgtctgtagtttactacagATCAAAGtagcaatgcatcagaatgattccagcagacgctaatctatactactc 1800
Y N Q 305

agTCAACGGAACCTTCTTTCGCAAAATGAGAAGACCTTTGTCAGGACTCCTCAAGGAGAGTACGGCTTCCAAGGCTgtgagtgctctttttacggggcaggcaatcaaatctctc 1920
I N G T F S C E N E K T L S G L L K G E Y G F Q G 330
agtatcgtagACGTATCTGTgtaggttttacagtggtccatgtcaagctcggcggtgagctgacagTACTGGTGGCCACTACTCTGGTGCACCCGCTGTGAACCGCGCCCT 2040
Y V M S D W W A T H S G A P A V N A G L 350

GACgtatgtctcgcgcttcccatggcgcctactgctaactgcttttagATGACCATTCCGGCGGACGAGACTCAGCTCTGGAACGACgtacgcagctcctgcccgaagctgtctgt 2160
D M T M P G D E T L S S G T T 365

cggtattctgagctgctgcttccagCTACTTCGGACAGAACCTCGTCAACGCTGTGAACAGCGCCAAAGTCTCGCAGGCCAGGTAAAGgtacgcagaatcgaccatatactactaat 2280
Y F G Q N L V N A V N S G Q V S Q A R V K 386

aggacttgaatgtgtcttgcacagGACATGGCGACTCGCATCCTTGGCGGTGGTACCTCCTTGGCAGAACAAACTTCCCGCGGTCAACTTCAACTCGTGGAACTTGGTCAAGG 2400
D M A T R I L A A W Y L L G Q D Q N F P A V N F N S W N S G Q G 418

CCAGCATGTGAACCTTCCGGCAACCATGCGAGgtactgtaaacgggtatccactactcggcgcgaccgctgacacgctgtagCCTCATCCGCACCATCGCGCCCTCGCAAATC 2520
Q H V N V S G N H A S L I R T I G G C G S C T G I 440

CTGCTCAAGAACGTGAACAGTGGCTTCCGCTCAAAAAGCCCAAGACTATTGGCATCATTGGGAACGCTGGATCAAAACCTAAACGCTCCCAACGCTgtagcagacacaatccagcttta 2640
L L K N V N S A L P L K K P K T I G I I G N G A G S N P N G P N A 473
(A) (G)

cgttcccaagcttgtgagtttcttaccgagTTTCTCCGACCGCGCGGAGACGTCGCTGCTCCGCTTGGTGGGGCAGTGGtaagtcggcgcggcagcttggggtgtgta 2760
F S D R A G D V G V L A L G W G S G 491

caagctcacggtgccacaaaatagCACTGCGAACTTCCCGTACTTGGTCCGAgtaggtctatgctctctgctgcttccgctgtaactgacgcgcttagCCCGTCGACCGGATTACT 2880
T A N F P Y L V A P V D A I T 506

GCTCGTCCGTCAGGAGTGGGACACCTGCTGCTGCTGACTGAGTGCACCCGCTGACGGCGCTGCTAACACCGCGACCGGGAAGGACGTCGCGATGGTTCATCACGGCGACAGC 3000
A R A S Q D G T T V S S L S D T D L T G A A N T A T G K D V A M V F I T A D S 546

GGgtgctgtgtctgcccctgttcttattgagcgcagctgatttttacagTGAAGGATACCTTACTGTGCGAGGCAATGCAGGTGACCGTAAACGCTCCAGGCATGGCAGGAGTgt 3120
G E G Y L T V E G N A G D R N D L Q A W H G G 569

aagtgcgcctaacatcgtgactacatgcccgttatacacagacgctctagGATGCGCTGGTTCAGAGGTCCGCGAGCCACAACAAGAACAGATTTGCTGTATCAACAGTTCGGGACC 3240
D A L V Q V A S H N K N T I V V I N S V G P 592

TATCAATATGGAACGCTGGGTAACCCCGAACGTCACCGCTATTgtgagtgacctgtgctcattggagacctctgctgacgcccgtcggttaactagGTTGGAGTGGCCCTCCGTC 3360
I N M E A W V N H P N V T A I V W S G L P 613

GTCAGGAAGCAGgtatgcatatcgcgaagattcgtgtgcaaacactcaccctgtgcccagtagGCAACGCGTCACTGACGTTCTATTGGCGCGGTCAACCTGGTgtagctgtgctc 3480
G Q E A G N A V T D V L F G A V N P G 632

gacctgtgcttagggcgtaggacggtatgtgacaccgaccacagCGGCAAGCTGCCCTTACCATTCCGCAAGTGCAGTACTCCGCGCAGATCATACCACCGGCTCGGGCA 3600
G K L P F T I G K S I S D Y S A Q I I T T G S G 656

TCGTGCCATCCCGTACAACGAGGGCTCTTCATCGACTACCGGCACTTTGACCAAGgtgtgaccgggtgctgctgacgagcgcgagctgtgacggggcgctgtagCGGGCATCG 3720
I V P I P Y N E G L F I D Y R H F D Q A G I 678

CACCGCGTTCGAGTTCGGGTTCGGCTCTCGTACACGACGTTGACTACTCGAACCTCGTATCACGGGCTCGACGCGCGGCGCAGCGTCAGCCCCGGGGCCGGCTCGTCTCG 3840
A P R F E F G F G L S Y T T F D Y S N L V I T G S T A G G T R Q P P C G P G S S L 718

ACCCATGgtgcccatttcccttccgtaaaagtcaactcgtgacgcaggtgctgctgcccgcgagcagGCTGCACCACTCCGCTGCTCACCCTGACGACGACCAACCGGACCGGT 3960
D P W L H D S V V T V S F T L T N N G T V 739

CGAGGGACGGAGTCCGCGAGCTGTACTGAGCCCGCGGAGCGCGGCAAGGCGCGCGGCAAGGCTTCGACAGCGTATTCCTCCCGCGAGCGCTCGACCTGCTCTC 4080
D G T E V P L Y L S P P A S A K S A P Q N L K G F D S V F L P A G A S C T V S 779

TTTCGAGCTCTCGCGTACTCGTCTCCGCTGGGACGTCGCTTCCAGAGTGGCAGATCCCTGCTGGTGTACCAGGATCCTCCGTTGGCGCGGAGCAGAGATTTCGGCTCAAGGG 4200
F E L S R Y S F S V W D V V S Q S W Q I P A G V T G I S V G A S S R D L R L K G 819

CTCAATCAGGAACGAGG--CGAGAACGTCGCGGTAAAATGTATTCGTAGTGCATGTGCACTATCTTCTCGGCTTGACATATGTAACACTTCCCATTTCCGGACATAGCG 4318
S I T N T A CG 823

GAGGTACACTGACTGACTTTCACATGAGGACGCTTTGCCATATAAGCTAGTCTCCCTTCCGTCGCAACAGTAAACAGGCGGTAATAAATGTTTTCGACCCCAACCATGTACAAA 4438
ACTACAGATTAGGGGGATCAGCTCAGTGCAGAGAGTGTGCTAGAGTGAATATGGGGTAGTACAATGATATATCCAGACTCGGGGAGTTACGTCGCGAGTACGCTTTTGA 4555

FIG. 1. Nucleotide and deduced amino acid sequences of CBGL from *P. chrysosporium*. Genomic and amino acid sequences were derived from *cbgl-2*. The amino acid sequence deduced from the cDNA sequence of *cbgl-1* was the same as that of *cbgl-2* except at positions indicated in the line below the amino acid sequence in parentheses. The exon sequence of *cbgl-1* is the same as that of *cbgl-2* except at the positions indicated in the line above the gene sequence. Nucleotides and amino acids are numbered on the right. The potential signal peptide sequence is overlined. The potential CBD is boxed. Potential N-glycosylation sites are in boldface type.

Saf1 1 MLMIVQLLVFALGLAVAPIQNTQSPSQ-RDE-SSQWVSPHYPTPQGG--RLQDVWQBAVARAKATVGMQTTVEKVN
 Saf2 1 MLLILELLVLIIGLGVLPVQTHNLTNDNQFDEESSQWISPHYPTPQGG--RLQGVWQDAKAKALVSMQTTVEKVN
 Phc 1 -----VAN-----VSPWAAAVKAAQAAVAKLSVTDVNL
 Trr 1 -----MRYRTAAALALATGPFARADS-----HSTSGASAEAVVPP--AGTPWGTADKAKAALAKLNLDQKVGIL
 Asa 1 -----MKLSWLEAAALTAASVVSAD-----LAFSPFPYSPWAN--GQGEWAEAYQRAVAIVSQMTLDEKVN
 Sel 1 -----MVSSLFNIAALAGAVIALSHED-----QSKHPTIPTPTPDST--GEG-WKAAAEKAAADAVSRNLTKQVAL
 Pic 1 ----MKSTIILSVLAAATAKNIKSAEMENLEHWWSVGRSDPVYSPSEIS--GLGDWQFAQARAVLALMTNEKTNL
 Pia 1 ----MLLPYGLASFLVLSQAALVNTSAPQASNDPFPNHSFSPFYPTPQGGRI-NDGKWQAAYRARELVDMQSTAKKVN

Saf1 77 TTGTGWQLDPCVCGTGSVPRFG-IPNLCLODGPPLGVRFDVFTGYSPLATGATFNKDLFLQRGAALGHEFNKGVHIAL
 Saf2 79 TTGTGWQLGPCVCGTGSVPRFG-IPNLCLODGPPLGVRFLDFSTGYSPSMTATGATFNKDLFLQRGAALGHEFNKGVHIAL
 Phc 71 ATGVQVQGPCVCGTTPAISSIPGTFGLCLQDQSPVGVRYADGTSVFPPEINVAATWNRRTLMRQRGAALGAEFKGKGVHIAL
 Trr 63 VSGVGNWGGPCVCGTSPASKIS-YPSLCLQDGPPLGVRFDVFTGYSPLATGATFNKDLFLQRGAALGHEFNKGVHIAL
 Asa 63 TTGTGWLEKPCVCGTGGVPRLN-IGGMLCLODQSPGLGRSDYNSAIPAGVNVAAATWKNLALYLRGAALGHEFNKGVHIAL
 Sel 66 TTGTTAGLS-CNGGIAPPEIN-FSGLCLADGPIVSRADLATVPAGLTAATWDRQLIYERARALGSEFRKGSQVHL
 Pic 74 TFGSSGDTG-CSGEMISDVPDVF-PFGLCLQDAGNGVGRDMDVNAASGLHVQASWNRQLAYDRAVYMGAEFRHKGVNVL
 Pia 76 TTGVGSASGDFCSGNTGSVPRLN-ISSICVQDGPPLSVRAADLTVDVFCGMAASSSFNKQLIYDRAVYMGAEFRHKGVNVL

Saf1 156 GPAVGPPLGVKARGGRNEAFSGDPYLOGTAAATIKGQENNVMAVCVKHFIQNEQEKYRQDD----INPATNQTTKEA
 Saf2 158 GPAVGPPLGVKARGGRNEAFSGDPYLOGIAAAATIKGQENNVMAVCVKHFIQNEQDIYRQPSNS--KVDPEYDPATKES
 Phc 111 GPMMN-LMRVPAAGRNVEGGGDFPLSGEVAFETISGQSSGAQCAKHFINNEQEHFR-----DS
 Trr 142 GPVAGPCKTPOGGRNEGFGVDPYLTGIAMQTINGQSVGVQATAKHYILNEQELNR-----ET
 Asa 142 GPAAQGRSPDGGRNVEGFSDDPALTVLFAETIKGQDAGVVAATAKHYILNEQEHFRQVAE--AAGYGFNIDST
 Sel 144 GPASGALGRHLGGRNVEFSFDPYLSGVAMDPSIRGQEMGVQANRKHFIQNEQETQR--SN-----TFTDDGTETQA
 Pic 152 GPVVGPGRVATGGRNEGFTNDPPLAGALVYEATKQEN-VIACTKHFIQNEQETNR-----NPSG--TYNQS
 Pia 155 GPVYGPVGVKAAAGRGVEGCHGPDPLLEGVIAYLQITIGQSQGVVSTAKHLLIQNEQEHFRFAKKDKHAGRIDPGMFTSSS

Saf1 231 ISANIPDRAMHAYLWPFADSVRAGVCSMCSYNRVNNTYACENSYMNHLKKEELGFQGVVSDWGAQLSGVYSAISGL
 Saf2 235 ISANIPDRAMHAYLWPFADSVRAGVCSMCSYNRVNNTYSCENSYMIHLLKKEELGFQGVVSDWGAQMSGAYSATISGL
 Phc 171 SSSNVDDRTHEHEVYGHFFLRSVQANVASMCSYNQNGTFSCNEKTLGSLKGEYGFQGVVSDWVAATHSGAPAVNAGL
 Trr 203 ISSNPDDRTLHEHEVYTFPFADAVQANVASMCSYNKNTTWACEDQYTLQTLKDLQGFQGVVMTDWDVAQHTTVQSANSGL
 Asa 216 ISSNVDDRTLHEHEVYLPFFADAVRAGVCSMCSYNQNNISYGCNSYTLNKLKKEELGFQGVVSDWGAHHSVGSALAGL
 Sel 216 ISSNVDDRTLHEHEVYLPFFANAVRSGVCSMCSYNRNVNTYACENSYKLNGLKKEELGFQGVVSDWVAHHSVGSVNAAGL
 Pic 219 ISSANIDRTMHHEVYLPWFQDSVRAGVCSMCSYNRVNNTYACKNSKVLNGLKKEELGFQGVVSDWGAQHTGIASANAGL
 Pia 235 LSSELDDRAMHAYLWPFADSVRAGVCSMCSYNKNGSHACQNSYLLNYLKKEELGFQGVVMTDWDGALYSGIDAANAGL

Saf1 311 DMSMPGEVYGGWNTG---TSWCGNLTTRKALYNETVPIERDDMAIRILAALYATNSFPTD---HLPNFSWTTKEYGNK
 Saf2 315 DMSMPGELLGGWNTG---KSYWCGNLTTRKALYNETVPIERDDMAIRILAALYATNSFPTD---RLPNFSWTTKEYGNE
 Phc 251 DMTMPEGDELSSGTT---YCGNLTVAVNSGQVQARVDDMAIRILAALYLLGQDQNF---PAVNSWNSGQG---
 Trr 283 DMSMPG-TDFGNRR---LWCPALTNVANSQVTRVDDMVRILAALYLLTGQDQ---AGPFSNISR---
 Asa 296 DMSMPGDTTFDSATS---SWGNTLTIATVNLGTVPQWRVDDMAVRIIAAYKXVGRDRLY---QPPNFSWTRDEYGFK
 Sel 296 DMTMPEGLDPSSTALRPPPSYLGGLNTEATVNLGTVPEARVDDMAIRILAALYLLGQDQDPTVDPSTGCVFARTYNYPDE
 Pic 299 DMAMPS-----ST---YEEGLIEAVKNGTVDQSRDDMAIRILAALYKYARLDD---PGFMVPSYLAED---
 Pia 315 DMDMPECEA-----Q-----YCGNLTVAVNLGTVLQDRDDMAIRILAALYISGVHNP---DGPNYNAQTFLEGEH

Saf1 385 -YADNTTEIVKVNYNVDPNSDYFTEDTALKVAEESI VLLKNEENTLPIISPEKAKRLLLSGIAAG---PDPICYQ-----C
 Saf2 389 FFVDKTSFVVKVNHVDPNSD-FTEDTALKVAEESI VLLKNEENTLPIISPNKVRKLLLSGIAAG---PDPKGYE-----C
 Phc 319 -----QHVNVSNG--HASLIRITGAASQVLLKNANGALPLK--KPTTIGIIGAG---SNPNCNPN---AF
 Trr 345 -----NVQGN--HKTNVAIARDGI VLLKNDANILPLK--KPAIAVVGSAAI--IGNHARNSP--SC
 Asa 367 YFPYQEGPYEKVNHFVNQRN--HSEVIRLQADSTVLLKNNN-ALPDTG-KERKVAILEGDA---SNSYGAN---GC
 Sel 376 YLTLGGLDFYNPPARDVRGN--HSDIVRVAAGTVLKNVNNVPLK--EPKSVGIFGNGAA--DVEGLTFTGDSS
 Pic 358 -----HELVDARDP-AAASTFQGAVEGHVLLKNE-ALPLK--EPKYISLFGYDGVSTDVNTVGGFSFSS
 Pia 379 YFKQQEGDIVVLNKHVDVRS-D-INRAVALRVAVEGVLLKNEHELELREKVRISILGQAAAG---DDSKGTSS---C

Saf1 456 EDQSCNTCALFQGWGSGSVGSPKVVQVTFEEISYLARKNKMQFDYIRESYDLAQVTK--VASDAHLSVWVSAASGEGYI
 Saf2 460 SDQSCVDALFEGWGSVGYPKVVQVTFEEISANARKNKMQFDYIRESFDLTQVST--VASDAHMSVWVSAVSGEGYL
 Phc 375 SDRAGDVGLFALGWSGTANFP-YLVAPVDAITARASQDGTTVSSSLSDTLTGAAN--TATGDHVAVFTADSDGEGYL
 Trr 400 NDKGCDGALGMGWSGAVNYP-YFVAPYDANTRASSQGTQVTLNNTDNTSSGAS--AARGKDVAVFTADSDGEGYI
 Asa 436 SDRGCDNCTLAMAGWSGTAFFP-YLVTPQALQAEVLKHKGSVYAITDNWALSQVET--LAKQASVLFVNSDAGEGYI
 Sel 449 GPWAGDICALAVGSGSAGRHT-HLVSPLAARKRKTESVGGRVYQLLSNSRIVNDDFTSIYPTPEVCLVFKTWAREGTD
 Pic 422 DVKAIENKTLISGGGSG-TNTPSYVDAPFNFAVAKARED----NTFLSWDFTSAEP-VANPASDADDFINAAASGQYD
 Pia 450 SLRGCSSGALGTGYSAGTFS-YFVTPADGEGARAQKEKISYEFIGDSWNQAAMD--SALYADAATEVANSVAGBERIG

Saf1 534 TVDGNQGRKNLTLWNGDKLLETVAENCANTVVVVTSGQINFEGYADHPNVTAVVWAGPLGDRSGTANILFEGKANP
 Saf2 538 IIDGNRGRKNLTLWNSDNLKVAENCANTVVVVTSGQVDVSEADHPNVTAVVWAGPLGDRSGTANILFEGKANP
 Phc 452 TVEGNAGDRNDLQAWHGGLDALVQVVAASHNKNTVVVINSVGPINMEAVNHPNVTAVVWAGPLGQEAAGNATDVLFGAVNP
 Trr 476 TVEGNAGDRNDLDPWHNGNALVQAVAGANSVVVIVHVSVPAILLEQILALPQVRAVWAGPLSQESGNALVDVWLVGDSV
 Asa 513 SVDGNEGRNDLTLWNGDKLKAANNCNTVVVIVHVSVPAILVDEYDHPNVTAVVWAGPLGQESGNSLADVLYGRVNP
 Sel 528 RLS-YEND-----WN-STAVVNNVARRCNTVVVTHSGG-INTMPWADNANVTALAAHYQENGNSMDILYGDVNP
 Pic 495 RPN--LAD-----KY-SDKLVEAVASQCSNTEVVIHNAQIRLVDNWIENHNTVYLAHLPGQDTGTSILEVLYGNQSF
 Pia 527 DVVGNQGLNNTLWNAVPLKNTSINNTVVVVTSGQIDLEPVIDNENVTAVVYSSYLQDFGTVAKVLLEGDENP

Saf1 614 SGLLPPTAKT---DDYIPIETYSFSSG-EPED--NHLVENDLLVDYRYFEKNIERPVAFGGLSYNEYVSNAKVSA
 Saf2 618 SGLLPPTAKS---NDYIPIVTYNPPNG-EPED--NTLAEHDLLVDYRYFEKNIERPVAFGGLSYNEYKVSNAKVSA
 Phc 532 GGLLPPTAKS---ISDYSAQIITGSGI-VP-----IPYNEGLFDYRHFDQAGIAPREFGGLSYTTFDYSNLSVETG
 Trr 556 SGLLVYTIAS---PNYNTIRIVSGS-----DFFSEGLFDYKHFDANITPRYFEGGLSYTRKYNISRLSLS
 Asa 593 GAKSPFTWKTREAYGDYLVRELNNGGA-PQ-----DDFSEGLFDYRGFDKRNTEPIYFEGGLSYTTFNYSGLHLQV
 Sel 599 SGLLPPTAKL---ATDYDFPVVITNEA-QDYVWQADFTGELLDYRHFDARNITPLYFEGGLSYTTFEIEGVANLV
 Pic 566 SGLLVYTIARK---ASDYGLLWTEPEGLDLVFPQSNFTGCVYDYKFIQNTIPRYFEGGLSYTTFEIEGVANLV
 Pia 607 SGLLPPTAKD---VMDYIPVIEKVDVP--DP-----VDKFTESIYDYRYFEKNIKPVRYFEGGLSYSNISLSLDIEKQT

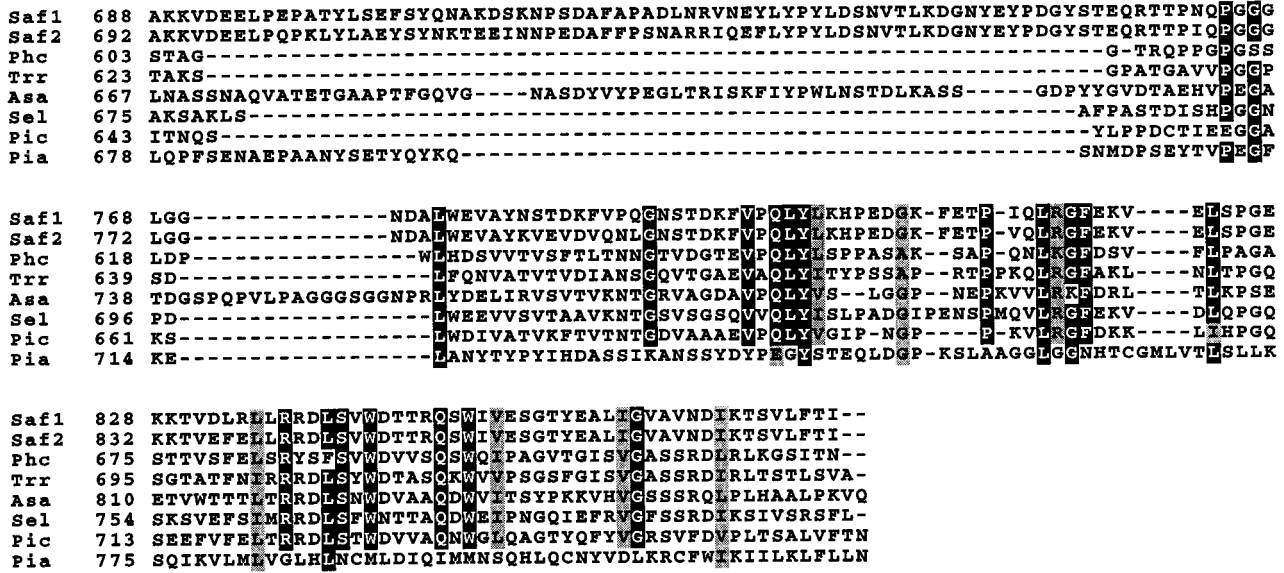


FIG. 2. Comparison of the catalytic domain sequence of CBGL with the β -glucosidases from *T. reesei* (Trr) (3), *A. aculeatus* (Asa) (15), *S fibuliger* (Saf1 and Saf2) (21), *Pichia anomala* (Pia) (17), *P. capsulata* (Pic) (14), and *Septoria lycopersici* (Sel) (24).

branes and hybridized to a ³²P-labeled 1.4-kb *SalI* cDNA fragment of *cbgl* (5).

Seventy-two positive clones of *cbgl* were isolated by immunoscreening of the *P. chrysosporium* cDNA library. A full-length clone was isolated by screening the cDNA library with a *MscI* + *NdeI* fragment from the genomic clone *cbgl-2*. Genomic clones were isolated by screening a λ EMBL3 genomic library of *P. chrysosporium* with a 500-bp *ApaI* fragment from the 3' region of the cDNA sequence. This screening yielded ~50 positive genomic clones. Restriction fragment analyses of five clones which hybridized strongly to the probe indicated that they were very similar, and one of them was subcloned and sequenced.

***cbgl* cDNA sequence.** Sequence analysis of the cDNA clone (2.4 kb) revealed an open reading frame consisting of 2,469 bp encoding 823 amino acids, including a 21-amino-acid N-terminal signal peptide sequence (Fig. 1). Prediction of the signal peptide cleavage site suggested that Gln22 was the N-terminal amino acid (22). The mature CBGL apparently consists of 802 amino acids and has an apparent molecular weight of 83,439. The CBGL molecular weight as determined by sodium dodecyl sulfate-polyacrylamide gel electrophoresis was 114,000. CBGL is a glycoprotein, and the difference in molecular weight could be attributable to the carbohydrate portion. The cDNA sequence revealed six potential N-glycosylation sites conforming to the general rule Asn-X-Thr/Ser, in which X is not a proline (4). In addition, numerous O-glycosylation sites are possible.

CBD. Analysis of the *cbgl* cDNA sequence suggested that the amino acid sequence from 22 to 57 has a high sequence similarity to the conserved cellulose-binding sequence of CBHII from *P. chrysosporium* and other fungal CBD sequences (11). This confirmed our prediction that CBGL possesses a CBD similar to that of cellulases (20). This domain is connected to the C-terminal catalytic domain via a 43-amino-acid linker region.

Catalytic domain. Approximately amino acids 101 to 823 at the C terminus form the catalytic domain. Sequence analysis suggests that CBGL should be categorized as a family 3 glycosylhydrolase along with the extracellular β -glucosidases from *Trichoderma reesei*, *Aspergillus aculeatus*, *Saccharomyopsis fi-*

buliger, and *Pichia capsulata* (3, 13, 14, 17, 21). Asp and Glu have been found in the active sites of numerous glycosidases, including β -glucosidase, cellulase, and amylase (7, 23, 28). Analysis of the catalytic domain sequences of eight family 3 fungal and yeast glycosidases indicated conservation of 13 acidic amino acid residues. Sequences surrounding seven conserved residues are preserved (Fig. 2). Potentially, any two of the conserved residues could be involved in catalysis.

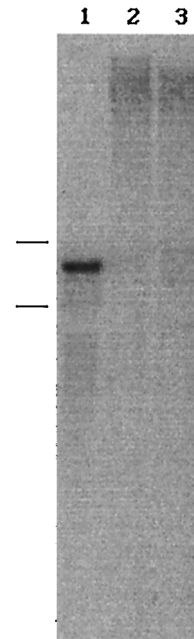


FIG. 3. Northern blot analysis of *P. chrysosporium* RNA. Total RNA was isolated from 11-day-old mycelia obtained from 1% cellulose (lane 1), glucose (lane 2), and cellobiose (lane 3) cultures. The blot was probed with ³²P-labeled CBGL cDNA. Bars to the left indicate the positions of 18S and 28S rRNA (from top to bottom).

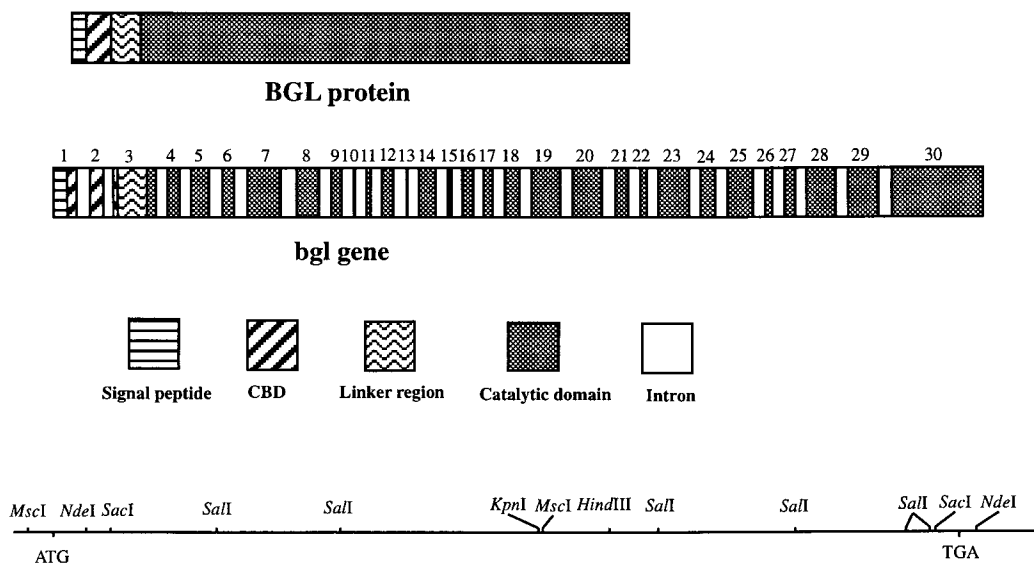


FIG. 4. Schematic representation of the protein and gene structures of CBGL and the restriction map of *cbgl-2*.

CBGL expression. *P. chrysosporium* produced CBGL abundantly only when cellulose was provided as the sole carbon source (2, 20). CBGL production was not observed in cultures supplemented with either glucose or cellobiose (2). To further understand CBGL expression in *P. chrysosporium*, total RNA was isolated from 11-day-old cellulose, cellobiose, or glucose cultures and analyzed by Northern blotting (Fig. 3). A band corresponding to 2.4 kb was observed only with the RNA isolated from cellulose-grown cells. Also, the size of this RNA was very similar to the size of the cDNA insert. These preliminary findings suggest that either cellulose or one of its degradation products might be controlling the expression of CBGL at the transcriptional level.

Gene sequence of *cbgl-2*. *cbgl-2* consisted of 4,555 bp, including 182 bp in the 5' flanking region and 339 bp in the 3' flanking region (Fig. 1). The 5' upstream region contained a potential TATAA box (TATAAGT) 64 bp upstream from the translation start codon. Comparison of the genomic and cDNA sequences of CBGL indicated the presence of 29 introns varying in size from 47 to 68 bp (Fig. 1). All of the intron splice junctions conformed to the GT-AG rule. Exon 1 codes for the signal peptide and a portion of the CBD (Fig. 4). Exon 2 codes only for the CBD. Exon 3 codes for the rest of the CBD, the linker peptide, and a small sequence of the catalytic domain. Exons 4 to 30 code for the catalytic domain. Interestingly, exons 10 and 13 code for one and two amino acids, respectively (Fig. 4). In contrast to *cbgl*, *T. reesei bglu1* has only two introns (3).

Exon sequences of *cbgl-2* exhibited 98% similarity to the *cbgl* cDNA sequence. A total of 50 bp in the sequences (in the exon regions) did not match the cDNA sequence; however, the amino acid sequences differed only at four locations (Fig. 1). Restriction analysis of *P. chrysosporium* DNA indicated that, except for *HindIII* restriction, only one fragment from all the other restrictions hybridized to a 1.4-kb *SalI* fragment of *cbgl-2* (Fig. 5). This result suggested that *cbgl* is probably encoded by two alleles (*cbgl-1* and *cbgl-2*) of a single gene. The cDNA sequence was presumably derived from *cbgl-1*.

***cbglu-1* and *cbglu-2* allelism.** *P. chrysosporium* is a heterokaryon with two or more genetically distinct nuclei (1). The genomic library from which the *cbgl* clones were isolated was derived from such a heterokaryon. However, the basidiospores

are homokaryons and contain two identical nuclei. If *cbgl-1* and *cbgl-2* are truly allelic, then they should segregate among homokaryons. Segregation of allelic variants of lignin peroxidase, glyoxal oxidase, and cellobiose dehydrogenase from

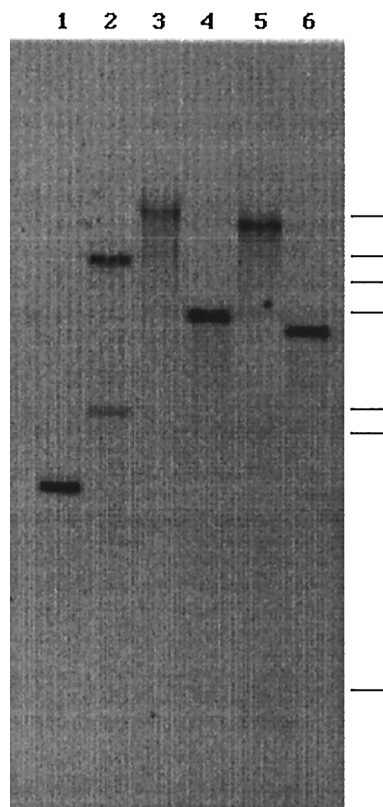


FIG. 5. Southern analysis of genomic DNA from *P. chrysosporium*. Genomic DNA, isolated by standard procedures, was digested with restriction enzymes *SalI* (lane 1), *HindIII* (lane 2), *BamHI* (lane 3), *NdeI* (lane 4), *EcoRI* (lane 5), and *SacI* (lane 6). The blot was probed with a ^{32}P -labeled 1.4-kb *SalI* fragment of *cbgl-2*. Bars indicate the positions of molecular size standards (from top to bottom: 23.1, 9.4, 6.6, 4.4, 2.3, 2.0, and 0.6 kb).

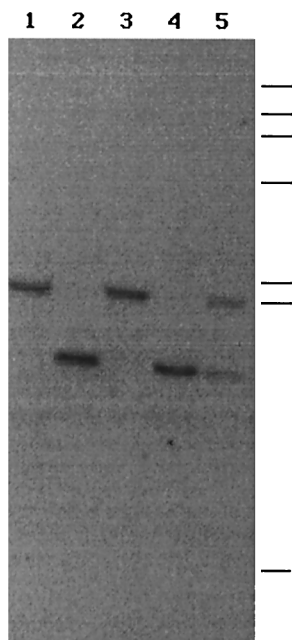


FIG. 6. Segregation of CBGL alleles into homokaryons. DNA from four separate single spore cultures (lanes 1 to 4) and one parental heterokaryon culture (lane 5) of *P. chrysosporium* OGC101 were restricted with *SalI*, size fractionated on an agarose gel, and probed with a 1.4-kb *SalI* of *cbgl-2*. Bars indicate the positions of molecular size standards (from top to bottom) of 23.1, 9.4, 6.6, 4.4, 2.3, 2.0, and 0.6 kb.

P. chrysosporium is known (10, 16, 19). A comparison of the cDNA sequences of *cbgl-1* and the exon sequences of *cbgl-2* suggested that *cbgl-2* has at least one extra *SalI* site at nucleotide 2866 (Fig. 4). This difference was utilized in identifying the two alleles. DNAs from homokaryotic and heterokaryotic cultures were restricted with *SalI* and probed with a 1.4-kb *SalI* fragment (bp 1446 to 2866). The probe was expected to hybridize to only a 2-kb fragment from *cbgl-1*, a 1.4-kb fragment from *cbgl-2*, and two fragments (1.4 and 2 kb) from the wild-type heterokaryon. Southern analysis suggested that only one fragment (1.4 or 2 kb) from homokaryon DNA and two fragments from wild-type DNA can hybridize to the probe (Fig. 6). These findings support the proposal that *cbgl-1* and *cbgl-2* are alleles.

β -Glucosidase multiplicity. Smith and Gold (26) partially purified an extracellular β -glucosidase (M_r , 90,000) from *P. chrysosporium* OGC101. CBGL is produced by the same strain in cultures optimized for low extracellular protease levels (2). Proteolytic hydrolysis of CBGL produces two non-cellulose-binding forms (M_r s, 96,000 and 98,000). Thus, the β -glucosidase isolated by Smith and Gold (26) was probably a degradation product of CBGL. At this time, there is no reason to believe that the low-molecular-weight glucosidase could arise from differential splicing of *cbgl-1* or *cbgl-2*. Deshpande et al. (9) have reported five extracellular β -glucosidases from *P. chrysosporium*, with molecular weights ranging from 165,000 to 182,000. A comparison with the current findings is not worthwhile because of strain differences and the variation in culturing conditions.

Nucleotide sequence accession numbers. The *P. chrysosporium* OGC101 CBGL cDNA and gene sequence data reported here have been deposited in GenBank under accession no. AF036873 and AF036872.

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