

Disuse-driven plasticity in the human thalamus and putamen

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Abstract:

Motor adaptation in cortico-striato-thalamo-cortical loops has been studied mainly in animals using invasive electrophysiology. Here, we leverage functional neuroimaging in humans to study motor circuit plasticity in the human subcortex. We employed an experimental paradigm that combined two weeks of upper-extremity immobilization with daily resting-state and motor task fMRI before, during, and after the casting period. We previously showed that limb disuse leads to decreased functional connectivity (FC) of the contralateral somatomotor cortex (SM1) with the ipsilateral somatomotor cortex, increased FC with the cingulo-opercular network (CON) as well as the emergence of high amplitude, fMRI signal pulses localized in the contralateral SM1, supplementary motor area and the cerebellum. From our prior observations, it remains unclear whether the disuse plasticity affects the thalamus and striatum. We extended our analysis to include these subcortical regions and found that both exhibit strengthened cortical FC and spontaneous fMRI signal pulses induced by limb disuse. The dorsal posterior putamen and the central thalamus, mainly CM, VLP and VIM nuclei, showed disuse pulses and FC changes that lined up with fmri task activations from the Human connectome project motor system localizer, acquired before casting for each participant. Our findings provide a novel understanding of the role of the cortico-striato-thalamo-cortical loops in human motor plasticity and a potential link with the physiology of sleep regulation. Additionally, similarities with FC observation from Parkinson Disease (PD) questions a pathophysiological link with limb disuse.

Introduction

Brain networks must simultaneously exhibit stability, to preserve acquired skills, but also flexibility, to learn and adapt to environmental changes¹. Motor behavior is generated by complex cortico-subcortical circuits including the thalamus, basal ganglia and cerebellum²⁻⁴. Cortico-striato-thalamo-cortical loops have been intensively studied to understand pathways involved in motor learning. Prior studies of plasticity in this circuit have been pursued largely by invasive electrophysiology in patients undergoing deep brain stimulation [DBS]⁵⁻⁷, and animal models^{8,9}. Human neuroimaging studies of motor plasticity have focused on the cerebral cortex because of the low signal-to-noise ratio of blood oxygen dependent (BOLD) signals in subcortical structures^{10,11} and the limited effect sizes typically seen in plasticity paradigms¹²⁻¹⁴.

To study plasticity mechanisms in humans, we developed an experimental paradigm that induces disuse by restraining the upper-extremity using a full length cast¹⁵⁻¹⁷. This approach is similar to classical animal plasticity studies, which impose motor or sensory restrictions (e.g., limb constraint, deafferentation, monocular deprivation) in a small number of intensively studied individuals^{18,19}. We casted the dominant (right) upper extremity of three participants (Nico, Ashley and Omar) for two weeks and collected around-the-clock actigraphy and daily task and resting state functional MRI (fMRI) over a 6-week experimental protocol (2 weeks pre-cast, 2 weeks casting, 2 weeks post-cast). Limb disuse was documented by the actigraphy data and showed 15 to 24% increased use of non-dominant hand use. The participants only exhibited a reduced grip strength of the casted extremity measured at cast removal and recovered within days with no persistent deficits.

Dense longitudinal fMRI sampling enabled us to perform within participant analyses using our individual-specific Precision Functional Mapping (PFM) methodology²⁰. Resting state functional connectivity (FC) noninvasively maps functional networks within the brain and how they change in response to disuse. Previous analyses of the fMRI data acquired in the casting experiment focused on the cerebral cortex^{16,17}. fMRI signals in the left and right upper extremity primary somatomotor cortex (SM1_{ue}) are strongly correlated at baseline. Casting induced a marked decrease in FC (-0.23 to -0.86 change in correlation from around 0.8 before casting) restricted to the upper extremity specific parts of SM1_{ue}. We also detected increased FC between left SM1_{ue} and the cingulo-opercular network (CON)¹⁶, an executive control system responsible for initiating and maintaining actions^{21,22}. Unexpectedly, high amplitude fMRI signal pulses arising in left SM1_{ue} emerged during the casting period and were detected in supplementary motor area and cerebellum¹⁷.

Recently, we also described the previously unrecognized Somato-Cognitive Action network (SCAN), which is interleaved with effector-specific motor regions along the central sulcus²³. The SCAN is strongly functionally connected to the CON, and seems to serve as its downstream actuator, turning abstract plans into integrated whole-body actions. Becoming aware of the sharp divisions between SCAN and effector-specific

motor regions also provided motivation to re-evaluate our prior interpretations of some of the upper-extremity disuse-driven plasticity effects.

Motor and action control cannot be understood without subcortical nodes. Basal ganglia (putamen, caudate, globus pallidus, etc.) and thalamic nodes are intricately networked to facilitate the selection of intended actions while simultaneously suppressing or inhibiting potentially conflicting or undesirable ones²⁴. The posterior putamen specifically has been shown to be important for slow but long-term establishment of habits^{25,26}.

The central thalamus also plays a significant role in motor adaptation and control by serving as a relay and integration center between various brain regions involved in motor functions²⁷. The thalamus comprises a multitude of distinct nuclei, including the ventralis intermedius (VIM), the centromedian (CM), and the ventroposterior lateral (VPL)^{28,29}. The VPL receives sensory information and relay information for fine tuning of movement²⁹. The VIM, in comparison, is primarily involved in motor control functions such as planning, initiation, and execution of voluntary movements³⁰. The VIM is the deep brain stimulation (DBS) target in Essential Tremor and tremor-predominant Parkinson's Disease (PD)³¹⁻³⁵. A case report from a patient with longstanding bilateral upper extremity loss undergoing DBS suggested plasticity of VIM neurons, leading to an over-representation of shoulder movements³⁶.

The centromedian nucleus (CM) of the thalamus has been classified as part of the 'non-specific' nuclei of the thalamus and shows specific projections to the sensorimotor regions and anterior cingulate⁸. The CM plays an important role in the regulation of the cortical parvalbumin neurons that promote Hebbian plasticity³⁷⁻⁴⁰, and the overall level of cortical activity and arousal⁴¹. CM has mainly been targeted with DBS in refractory generalized epilepsy⁴²⁻⁴⁴ due to its role in sleep-wake regulation and arousal. The central thalamus plays an important role in regulating sleep stages and generates sleep spindles important for memory consolidation⁴⁵⁻⁴⁷, including procedural memory.

Given the importance of subcortical structures for motor control and skill learning, we expanded our prior analyses to investigate disuse-driven plasticity in human subcortical circuits for the first time.

Results

Disuse strengthens motor cortex functional connectivity with central thalamus and posterior putamen

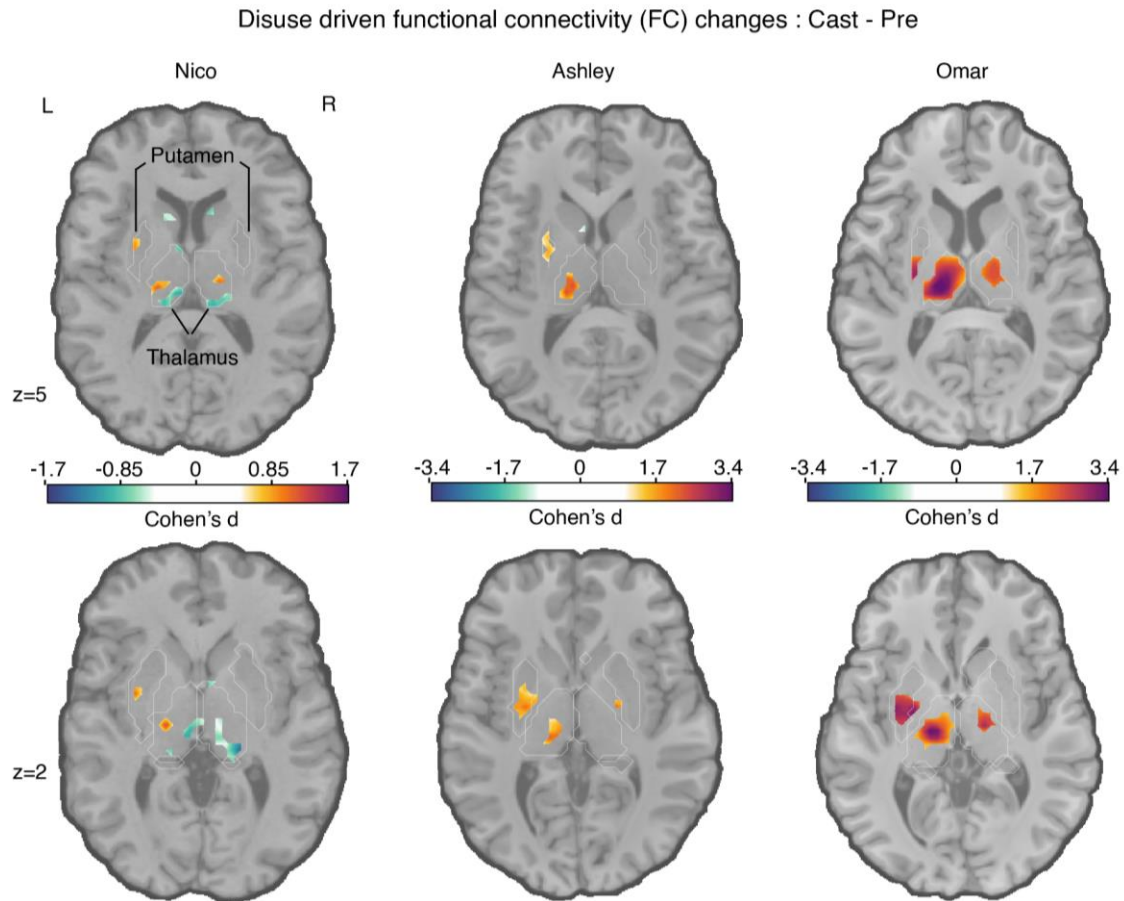


Figure 1: Disuse-driven changes in functional connectivity (FC) of the left effector-specific primary somatomotor cortex (L-SM1_{ue}) in subcortex. Individual-specific plasticity effect size (Cohen's d) maps showing changes in FC during right arm casting (Cast – Pre) for the L-SM1_{ue} for each participant (left to right columns: Nico, Ashley, Omar). For reference, a Cohen's d of 0.8 is generally considered a large effect size. Only significant effects after cluster correction at $p < 0.05$ (see Methods) are displayed. Please note, Nico's data were collected using an earlier pulse sequence with a TR that was twice as long (2.2 s) compared to Ashley and Omar (1.1s). Nico's effect sizes are about half the size of the other participants. The FreeSurfer based anatomical borders of the putamen and thalamus are shown as white outlines (bottom row).

Measures of FC in the subcortex are smaller than in the cerebral cortex. This observation is, at least partly because fMRI signal-to-noise ratio drops off towards the center of the brain as distance from the MRI coil elements increases. Consequently, the

magnitude of FC changes between different conditions (e.g., Cast - Pre) are greater in the cortex than in the subcortex. Therefore, to measure FC changes in the subcortex, we used standardized effect size (Cohen's d) to account for the signal-to-noise differences. The present effects, expressed in terms of Cohen's d, match our previously reported parcel-based FC findings¹⁶. Use-driven FC changes (Cohen's d) of the left SM1_{ue} increases with the CON regions (purple border, figure S1) for all three participants in cortex and with hand motor regions in the cerebellum (Figure S1, S3). We note that disuse-driven FC changes spared the Somato-Cognitive Action Network (SCAN; Figure S1, maroon outlines), a set of effector-general action control regions in motor cortex²³ strongly functionally connected with the CON (see also Figure S2). Thus, the two SCAN regions around the upper-extremity specific primary motor region (black seed region, left SM1_{ue}) did not show significant FC changes due to casting.

In the subcortex, all three participants showed statistically significant increases in FC between disused left SM1_{ue} and the central thalamus, as well as the posterior putamen (Figure 1, bottom row; cluster-based thresholding; see Methods). The subcortical effects (Cohen's d) were comparable in magnitude to effects observed in SM1_{ue} (Table 1).

Other FC changes were significant in some but not all participants. In particular, in Ashley and Omar, a small area of significant FC increase was observed in the left posterior globus pallidus bilaterally. Ashley and Omar's effect sizes were roughly twice those observed in Nico. These differences could be attributable to inter-individual differences, or due to differences in the fMRI pulse sequence (TR 1.1s Ashley, Omar vs. 2.2s Nico), or fMRI signal signal-to-noise ratios⁴⁸.

	Nico	Ashley	Omar
Left posterior Putamen	-31 -6 0 N _{vox} = 43 Max = 1.17	-25 -12 0 N _{vox} = 76 Max = 2.56	-31 -9 3 N _{vox} = 71 Max = 3.09
Left Thalamus	-16 -21 6 N _{vox} = 68 Max = 1.24	-7 -21 0 N _{vox} = 72 Max = 2.83	-13 -24 6 N _{vox} = 219 Max = 3.86
Cerebellum	Max = 2.22	Max = 2.68	Max = 4.79
Cortex	Max = 2.43	Max = 4.01	Max = 4.49

Table 1: Disuse-driven functional connectivity changes in subcortex. Peak coordinate (x,y,z) in MNI space and cluster size (N_{vox}) for subcortical regions with disuse-driven increases in functional connectivity with L-SM1_{ue}, as well as maximum Cohen's d values (Max) of the left putamen, left thalamus, cortex and cerebellum

Disuse pulses in the central thalamus and motor cerebellum

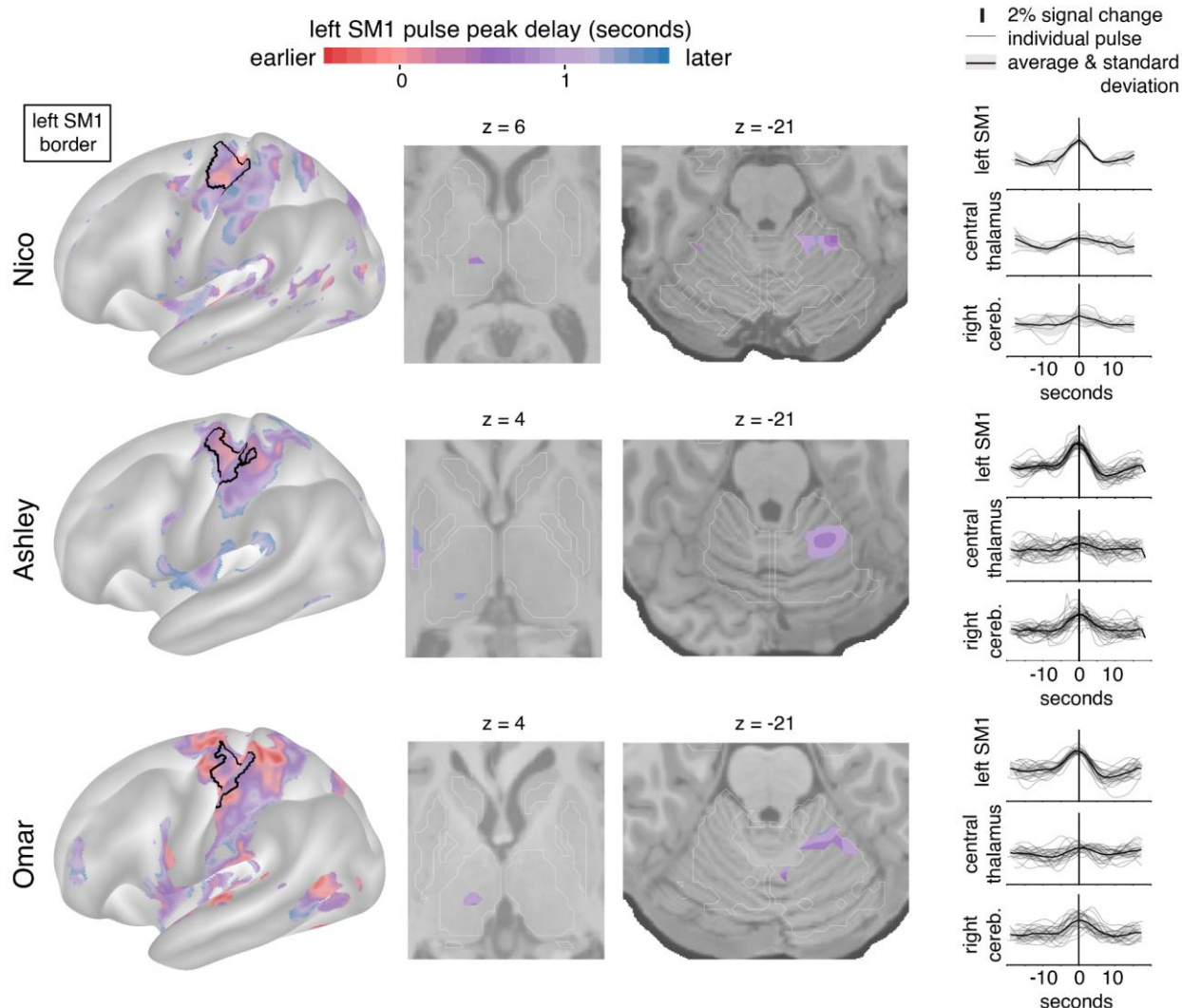


Figure 2: Disuse pulse distribution in cortex and subcortex. The timecourse of each disuse pulse observed during casting was modeled using a Hemodynamic Response Function (HRF) (see Methods). The left hemisphere cortical surface (left), and subcortical axial slices (center: thalamic and cerebellar view) show where pulses were detected in each of the participants (Nico, top; Ashley, middle; Omar, bottom). The color scale spans 2 seconds bracketing the average left SM1_{ue} pulse peak. The maps display the top 20th percentile of highest pulse detectability. The participant-specific upper extremity somatomotor region is outlined in black (left). On the right the individual (thin lines) and average (thick line) pulse timecourses are shown (y-axes: percent signal change) for the left SM1_{ue}, the left thalamus and right cerebellum.

We previously reported the emergence of large fMRI signal pulses in the disused motor cortex after 12-48 hours of arm casting. To identify disuse pulses in the subcortex, we developed an HRF-based pulse detection method that accounts for spatial variability in HRF shape (see Methods). Using the HRF-based detection method, we were able to

detect the presence of disuse pulses in the subcortex on top of confirming their spatial distribution in the cortex (Figure 2; left) and cerebellum (Figure 2; third column) ¹⁷. Disuse pulses in the central thalamus were observed in all participants (Figure 2; second column). The peak pulse percent signal change in the thalamus was lower compared to cortex (Figure 2, right column). In the participant with the most disuse pulses (Ashley), we also detected them in the posterior putamen. The least number of pulses were detected in Nico, who also exhibited the smallest FC changes.

Disuse pulses propagate through the brain in a specific temporal sequence. We previously reported that SMA regions peak earlier than left SM1_{ue}, followed by the cerebellum. On average, the central thalamus pulses peaked later than in left SM1_{ue} (Figure 2; first and second columns, Nico +0.75 seconds (sd 0.16); Ashley +1.07s (sd 0.02); Omar +0.95s (sd 0.09)).

FC changes and disuse pulses overlap in the central thalamus

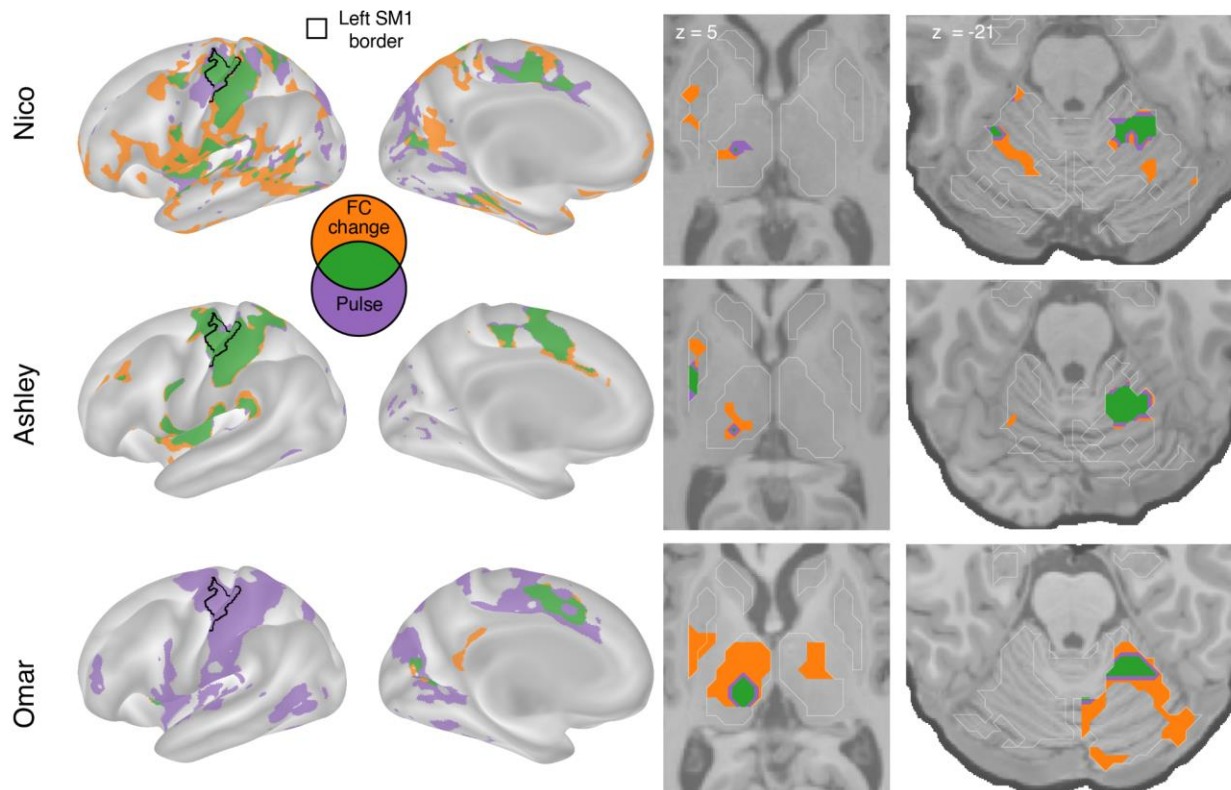


Figure 3: Spatial overlap of functional connectivity (FC) increases and disuse pulses. The strongest disuse-driven FC increases (orange, *Cast > Pre*, cluster corrected) and disuse pulses (purple, top 20% threshold), as well as their overlap (green) are shown on the cortical surface (left), in the thalamus and putamen (middle) and the cerebellum (right). Results are displayed on the lateral left hemisphere surface, medial left hemisphere surface, and two axial slices (MNI $z = 5$ and -21). White borders on axial slices defined individual specific FreeSurfer based anatomical structures ($z = 5$: putamen, globus pallidus, caudate, thalamus; $z = -21$: cerebellum, hippocampus).

Figure 3 illustrates the topography of pulses in relation to FC changes. All participants showed overlap (green) between disuse-driven FC changes and pulses in the dorsal medial cortex (SMA, pre-SMA, dACC), the central thalamus and the effector-specific motor regions of the cerebellum (Figure 3 and Figure S3). Ashley's pulses in the posterior putamen also overlapped with FC increases.

Subcortical disuse-driven plasticity overlaps with fMRI task activations

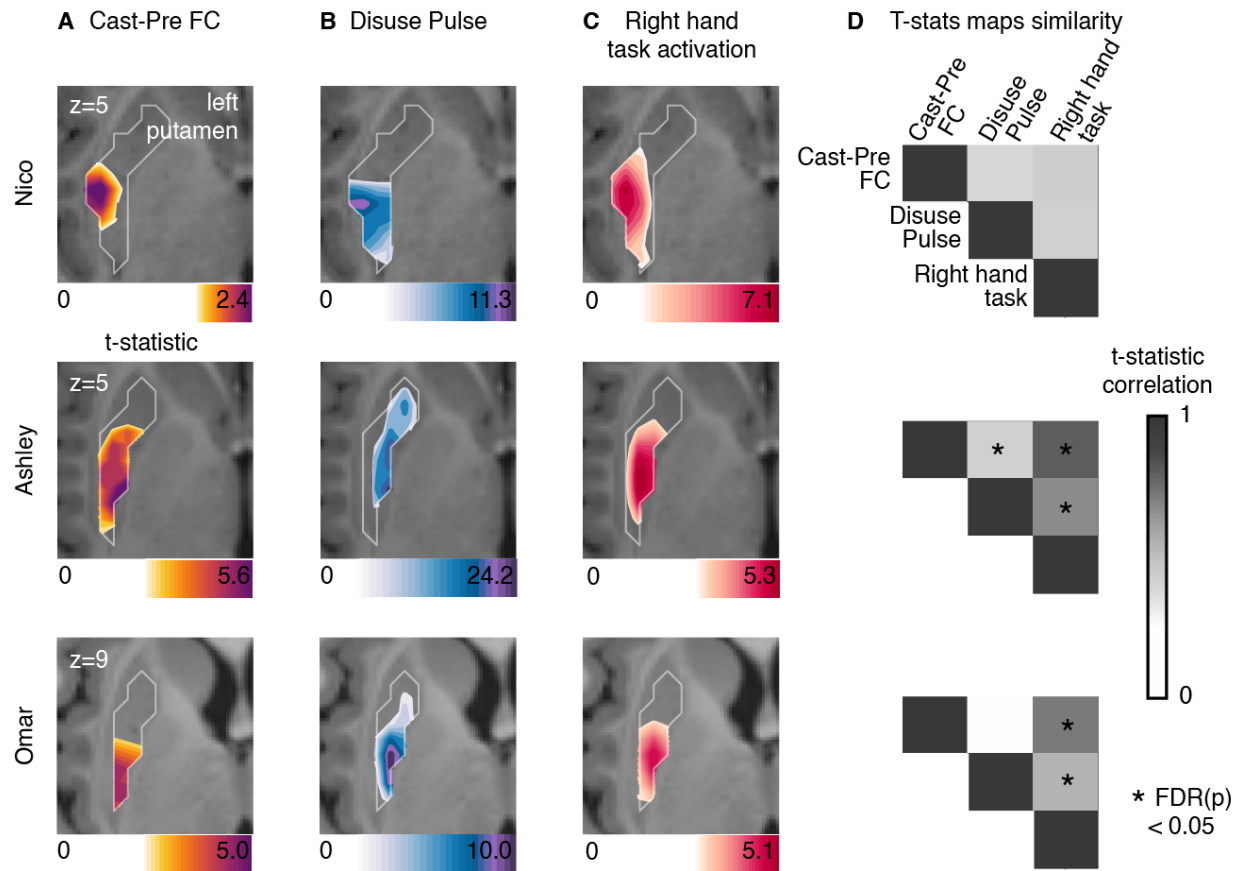


Figure 4: Putamen representations of disuse-driven FC changes, pulses and hand movement task fMRI activations. (A) Map of disuse-driven increases in FC with the left SM1_{UE} region of interest (top 30th percentile *t* statistics). (B) Map of disuse pulses (top 30th percentile *t* statistics). (C) Map of pre-casting task fMRI contrast: right hand movement vs baseline (top 30th percentile *t* statistics). For all three maps, color scales are represented at the bottom of the map with maximum value at 99.5th percentile. (D) Correlation between left putamen *t* statistic maps for disuse-driven FC increases, pulse and activation during hand movement (right hand vs baseline). Correlations between unthresholded *t* statistics maps were tested against individual-specific null distribution effects for each participant (top to bottom : Nico, Ashley, Omar). Reported significant *p* < 0.05 corrected for FDR (black *).

To determine whether FC changes and disuse pulses spatially coincided with regions active during upper extremity movement, we analyzed motor task fMRI data collected at baseline (Figures 4,5). The motor task includes simple hand, tongue, and foot movements in a block design (same as used in the Human Connectome Project). This paradigm elicited somatotopic specific responses in the primary motor cortex (see Figures S6-9).

All three participants showed motor task fMRI activations in the posterior portion of the putamen during right hand movement (Figure 4). In each participant, task activations overlapped with the FC change and disuse pulses. This similarity was confirmed by the strength of correlation between t-statistic maps. All participants showed strong (significant for Ashley and Omar) topographic similarities between motor task responses, FC change, and pulse density maps. Statistical significance of map similarity (Figure 4D) was assessed using individual-specific null distributions (see Methods).

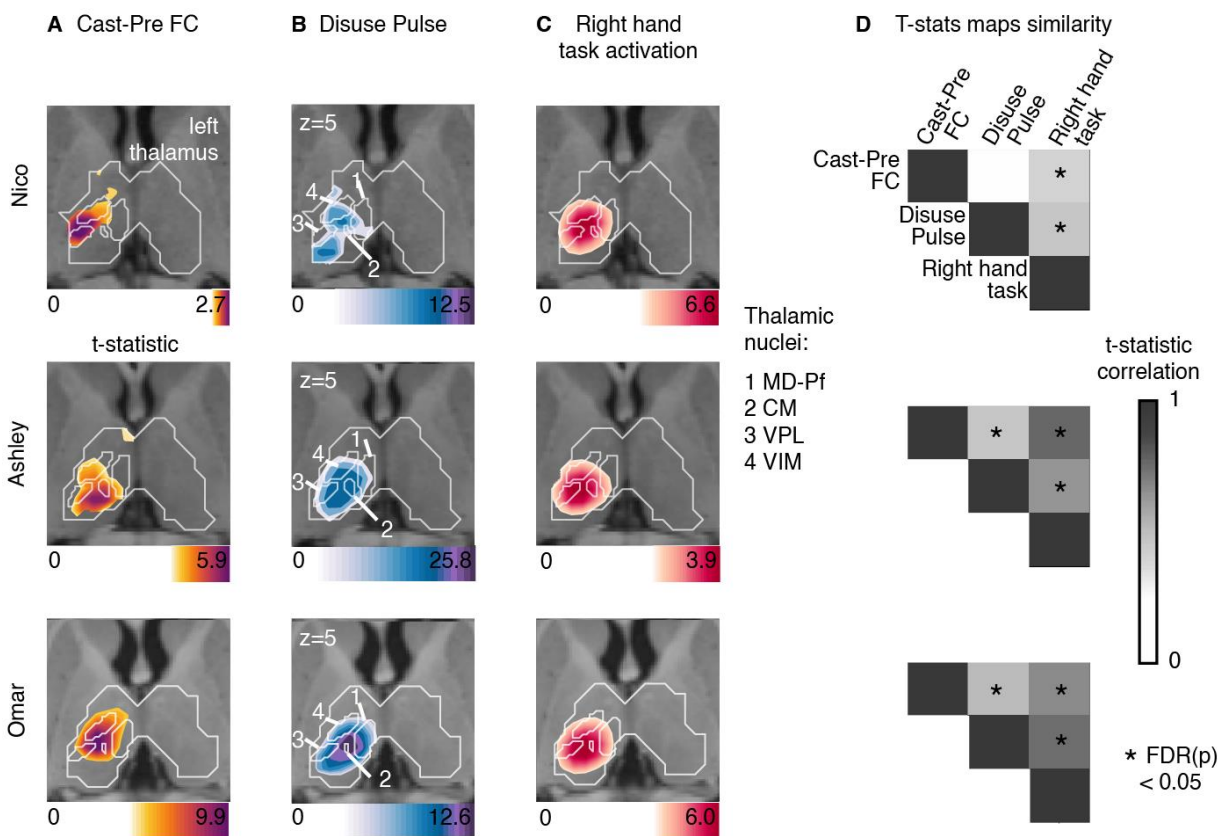


Figure 5: Thalamus representations of disuse-driven FC changes, pulses and hand movement task fMRI activations. (A) Map of disuse-driven increases in FC with left $SM1_{ue}$ region of interest (top 30th percentile t statistics). (B) Map of disuse pulses (top 30th percentile t statistics). (C) Map of precasting task fMRI contrast : right hand movement vs baseline (top 30th percentile t statistics). For all three maps, color scales are represented at the bottom of the map with maximum value at 99.5th percentile. For all three maps, color scales are represented at the bottom of the map with maximum

*value at 99.5th percentile. Nucleus borders from the THOMAS individual-based segmentation overlapping with the effects are visible in white. Four nuclei are displayed : CM: centro-median , VPL: ventro-posterior lateral , VIM(VLPv): ventral intermediate (ventro-lateral posterior ventral), MD-Pf: medio-dorsal. Average t-statistics values for all thalamic nuclei are represented in bar plots (see figure S4) (D) Correlation between left thalamic t statistic maps for disuse-driven FC increases, pulse and activation during hand movement (right hand vs baseline). Correlations between unthresholded t statistics maps were tested against individual-specific null distribution effects for each participant (top to bottom : Nico, Ashley, Omar). Reported significant $p < 0.05$ corrected for FDR (black *).*

All three participants showed task fMRI activations in central thalamus (Figure 5). Task activations overlapped with FC changes and disuse pulses in each participant. This similarity was confirmed by the strength of correlation between t-statistic maps, showing significant similarity in comparison to spatial null distributions (see Methods, Figure 5D). Individual differences in effect sizes were similar in all three measures (as in Table 1). Overall, thalamic FC changes and disuse pulses were similar to task fMRI activations when moving the right hand.

To delineate the specific thalamic distribution of cast-induced plasticity effects in comparison to hand motor circuitry, we quantified the average t-statistic values within each thalamic nucleus from the THOMAS atlas (Figure 5, S4, individual THOMAS atlas segmentation, see Method). Thalamic nuclei showing the greatest average t-statistic for both plasticity effects were the centro-median (CM), the ventroposterior lateral (VPL) and ventral intermediate (VIM) (see Figure S6 for all participants). Right hand movement showed consistent activations of CM, VPL, VIM across all participants. This set of nuclei remained specific when quantifying t-statistic average values from right > left hand movement task contrast, thus confirming the specificity of the right-hand movement circuitry (Figures S7-9). CM, VPL and VIM showed disuse pulse and increased FC changes with left SM1_{ue}.

Discussion

Disuse driven plasticity effects are not limited to cortex

Using analyses optimized for characterizing fMRI signal differences across the cortex and subcortex, we observed significant subcortical FC changes and disuse pulses in the central thalamus (VIM, CM) and posterior putamen as a result of dominant upper extremity disuse, due to casting. These findings suggest that disuse can drive network changes at all levels of motor circuitry: cortex, cerebellum, thalamus and striatum. In the cortex, the SCAN regions do not show an increase in FC or pulse presence across participants, which highlights the functional differences between SCAN and effector

specific networks. Throughout motor and action control circuits, as verified by task fMRI activations, FC increases partially overlapped with the presence of disuse pulses, suggesting that they might represent a brain-wide disuse plasticity mechanism. We previously demonstrated that the presence of disuse pulse does not fully explain the changes in FC¹⁶. While the presence of disuse pulses in the thalamus suggests a more general phenomenon, other plasticity mechanisms for neuronal and synaptic adaptation likely act in parallel.

Disuse strengthens FC in motor and action sub-circuitry

Increased FC between subcortical and cortical motor regions, in the presence of pulses, is consistent with our previously reported finding of strengthened FC between motor cortex and the CON¹⁶. However, these results cannot be fully accounted for by a simple Hebbian-like process^{39,49,50}, which one would predict should weaken FC in the setting of disuse (not firing together). Instead, we speculate that pulses, by creating co-activation in the disused motor control circuit, could help within network synchronization and information transfer, therefore playing a role in plasticity in order to help maintain the integrity of disused subcircuits¹⁷. Pulses in the disused subcircuits may enable the rapid recovery of both behavior (actigraphy) and functional connectivity (within days) after cast removal¹⁷.

Alternatively, the circuit can be going through integration of new motor programs related to disuse. Prior work in animal models suggests that neural plasticity in response to changes in auditory or visual stimuli starts with a reduction of inhibitory interneuron activity at the population level, increasing excitatory activity which facilitates Hebbian processes^{37,51-53}. Therefore, the observed FC increase within the motor execution circuit could indicate an increased simultaneous firing, eliciting Hebbian plasticity and information retention. This rise in local activity could increase the likelihood of spontaneous activity pulses, whether they represent runaway activity or necessary homeostatic regulation processes or windows for cortico-thalamic information exchange.

Disuse-driven plasticity affects posterior putamen involved in habit formation

Rapid, goal-directed learning is thought to primarily involve the dorsomedial striatum, including the caudate, whereas the slower acquisition of habits, which are insensitive to changes in the reward value of the outcome, is thought to depend more on the dorsolateral striatum, including the posterior putamen⁵³⁻⁵⁶. The increased FC with the posterior putamen suggests that it might specifically be related to protecting existing motor skills or acquiring new ones. Indeed, the first day removing the cast, our participants kept using their non casted arm more than their recently freed dominant arm, suggesting that participants learned to suppress movements of the casted arm. This trend disappeared on day 2 after cast removal and no fine motor or coordination motor impairment the day of cast removal.¹⁷

Plasticity in thalamic nuclei for motor execution

The central thalamus is important for motor and action execution and studies have tied movement performance to activity in VIM, VPL and CM thalamic nuclei^{6,57}, which is

consistent with our motor task fMRI activations (Figure S4, S6-8). The VIM is part of the thalamic ventro-lateral region (VL), primarily involved in motor control and relays information between the basal ganglia, cerebellum and motor cortex, whereas the VPL is part of the somatosensory system and relays sensory information related to touch, temperature, and pain from the body to the primary somatosensory cortex^{58–60}. VIM is a common and effective DBS target for tremor. In cases of neurological injury or disease, such as stroke or neurodegenerative disorders, the VIM can undergo plasticity as part of the brain's adaptation and recovery mechanisms^{36,61}. In rats, for example, VIM stroke results in specific skilled locomotor impairment (e.g., impaired ladder mobility, but not flat walk)⁶². Reorganization of activity in VIM after the stroke correlates with restored mobility^{63,64}. In fact, successful movement performance, such as walking on a complex surface, after a VIM partial lesion are compensated within a few days, showing the highly effective plasticity of the thalamus⁶². While the thalamus is essential to cortical organization and specialization during development⁶⁵, plastic reorganization of specific motor programs can also occur in response to motor demands driving functional adaptation in adults. For example, in humans, in the context of amputation, one study observed changes in the representation of the affected body parts within the VIM. Electrophysiological recording in a patient with arm amputation and long-term use of a prosthetic has shown an enlargement of the shoulder representation in the VIM, which now is used for prosthetic grip control by the patient³⁶.

Thalamic plasticity pulses could carry local sleep spindles

In addition to the VIM and VPL, the CM also showed FC increases, pulses and task fMRI activation. The CM plays a special role in regulating arousal⁶⁶. While the VIM is the DBS target of choice for treatment of tremor, the CM is targeted in the treatment of Tourette syndrome and, with increasing frequency, also epilepsy and disorders of consciousness⁶⁷.

Indeed, the thalamus has a specific role in sleep pressure and wakefulness regulation⁶⁸. The thalamus regulates sleep stages and slow oscillations in deep sleep⁴⁵. Sleep events like thalamo-cortical spindles are thought to help memory and skill consolidation^{69–71}. During deep sleep, slow waves of activity are observed across the cortex and relayed in the thalamus⁷². These slow waves are thought to help with homeostasis of neural activity after a day of novel experiences^{49,73,74}. The slow waves help networks to synchronize and increase phase locked communication between regions^{75,76}.

The occurrence of disuse pulses in the central thalamus, most reliably in the CM, raises the question whether they might be related to thalamocortical sleep spindles. Could the disuse pulses represent a circuit-specific, sleep-like phenomenon happening during the awake resting-state? Indeed, local sleep can be observed during wakefulness⁷⁷. After a long period in an awake state, EEG recordings in awake rats can capture local 'offline' stages similar to sleep, together with slow waves^{78,79}. Consistent with this idea, the one participant who was always scanned in the morning (Nico) also had the lowest number of pulses. In contrast, Ashley and Omar were always scanned in the evening. Thus, hours spent active since awakening could have driven the frequency of circuit-specific spontaneous activity pulses at the time of scanning.

Cortico-striato-thalamic FC increase as a marker of disuse in Parkinson's Disease

In our arm immobilization paradigm, we observed increases in FC in the disused motor circuit, but after removing the cast, these FC changes reversed rapidly and motor behavior was unimpaired¹⁶. In Parkinson's disease, action and motor impairments include tremor, rigidity and bradykinesia on top of a general paucity and slowness, and follow subcortical progressive loss of dopamine neurons projecting then affecting the caudate and putamen⁵. Similarly to our cast experiment, patients with Parkinson's disease show an increase in FC between putamen, central thalamus and cortical motor area, especially during akinesia^{80–89}. This suggests a potential pathophysiological link between limb disuse and Parkinson's disease. If increased FC in the motor and action circuits is a marker of increased neuronal excitability when a brain circuit is idling^{90–93}.

VIM is a target for DBS treatment of tremor. Electrophysiological studies in patients with Parkinson disease and parkinsonian mouse models have revealed increased beta power and prolongation of beta burst discharges propagating throughout cortico-basal ganglia circuits^{94–97}. These beta bursts disappear when moving or with L-dopa treatment^{98–100}. Further research is needed to determine whether beta bursts detected with electrophysiology are Parkinson's specific or perhaps disuse-related. Through this parallel, a potential role of disuse pulses would be to behave like sleep thalamic spindle, that are characterized by high frequency such as beta¹⁰¹, and to open synchronization windows for information updating in the motor network.

Plasticity and stability in subcortex

Disuse-driven FC changes and spontaneous activity pulses are not confined to the cortex but extend into the putamen and central thalamus (VIM, CM, VPL). The anatomical pattern, especially the prominence of the CM evokes parallels to sleep-related mechanisms of consolidation and plasticity, as well as to the beta bursts seen in Parkinson's patients. These findings open up intriguing new avenues for studying disorders such as Parkinson's disease and sleep physiology. They also raise the interesting possibility that the mechanisms for changing the brain and for maintaining it are one and the same.

Methods

Human participants: We analyzed a previously published dataset that comprised three healthy adult volunteers. The first participant (Nico) was 35 years old at the time of scanning and is male. The second participant (Ashley) was 25 years old and female. The third participant (Omar) was 27 years old and male. All participants were right-handed, as assessed by the Edinburgh Handedness Inventory¹⁰² (Nico: +100, right handed; Ashley: +91, right-handed; Omar: +60, right-handed). The Washington University School of Medicine Institutional Review Board approved the study protocol and provided experimental oversight. Participants provided informed consent for all aspects of the study and were paid for their participation.

Experimental setup: Arm immobilization was conducted by constraining the participant's dominant (right) arm for two weeks (cast period). The immobilization followed a two-week experimental baseline (pre-cast period) and was followed by a recovery period of two weeks (post-cast period). For one participant (Nico), the pre-cast period was acquired one month before the cast period and data was consistently acquired at 5 AM, while for the two other participants, fMRI was acquired at 9 PM. For one participant (Omar), the cast was removed and reapplied after one day of the cast period to adjust for finger comfort. Details of cast construction are described in Newbold et al. ¹⁷

Imaging data: On each day of the experiment, a scan session was conducted to acquire structural and functional data. Structural MRI consisted of four T1-weighted images (sagittal acquisition, 0.8 mm isotropic resolution, 3D MP-RAGE, Gradient echo) and four T2-weighted images (sagittal acquisition, 0.8 mm isotropic resolution, 3D T2-SPC, Spin echo). A 30 minutes resting state fMRI (rs-fMRI) run was acquired during each session, and two runs of the HCP Motor strip mapping task ^{103,104} were acquired for each pre and post cast session. Ashley and Omar's fMRI was acquired with an improved fMRI sequence (all 2D Gradient echo, echo planar, TR: 1.1 vs. 2.2 seconds in Nico, 2.6 vs. 4 mm isotropic resolution in Nico). All data were resampled at 3mm in atlas space. Acquisition parameters and procedures are detailed in Newbold et al. ¹⁷.

Precision functional analysis: All following data processing and analysis is conducted at the participant level using individually defined functional and anatomical boundaries. All statistical testing was done against null distributions built for each participant. All major results were replicated across all participants.

MR Image Processing. Preprocessing of structural and functional images, denoising of rs-fMRI data, and cortical surface projection were performed as previously described¹⁷. Functional image processing followed a previously published pipeline ¹⁰⁵ and involved temporal interpolation to correct for differences in slice acquisition timing, rigid-body correction of head movements, correction for susceptibility inhomogeneity-related distortions, and alignment to atlas space. The present results are reported in MNI152 space. For cortical surface projection and creation of cifti images, individual-specific surfaces were created defining the cortical gray-matter boundaries derived from T1-weighted images using FreeSurfer ¹⁰⁶. Subcortical boundaries from the FreeSurfer segmentation were used to select voxels of interest for building the volume part of the cifti image. There were no systematic differences in head movement (mean FD or number of frames removed) between phases of the casting protocol.

rs-fMRI data denoising involved replacement of high-motion frames (framewise displacement [FD] > 0.1 mm) by temporal linear interpolation, band-pass filtering (0.005 to 0.1 Hz), and regression of nuisance time series, including head movement parameters, the global signal averaged across all gray-matter voxels, and orthogonalized waveforms extracted from ventricles, white matter, and extracranial tissues. The rs-fMRI for functional connectivity is then projected to cortical surface as the last step and smoothed using a two-dimensional 6-mm full-width half-maximum (FWHM) smoothing kernel and volume data were smoothed using a three-dimensional 4.7-mm FWHM kernel.

Hemodynamic response function modeling was used for the pulse spatio-temporal representation analyses using rs-fMRI and for the HCP task data. For these analyses, denoising of data involved high-pass filtering (0.1Hz) before surface projection and regression of

nuisances time series (head movements parameters, individual high-motion frames). Nuisance regression was done at the surface and subcortical volume level within a GLM design. Cortical surface data were smoothed at the last stage of each analysis using a two-dimensional 6-mm full-width half-maximum (FWHM) smoothing kernel and volume data were smoothed using a three-dimensional 4.7-mm FWHM kernel.

Fully processed data are available in the Derivatives folder of the Cast-Induced Plasticity dataset in OpenNeuro (<https://openneuro.org/datasets/ds002766>).

Primary somatomotor upper extremity region of interest: The upper extremity SM1 region was defined in individual using a task-based approach combined with automatic labeling by FreeSurfer. Details were previously described in Newbold et al. ¹⁷.

Individual network representation: A set of 18 canonical functional networks was defined for each participant using a graph theory-based community detection algorithm with anatomical priors ²⁰(see figure S1), Infomap algorithm ¹⁰⁷(<https://www.mapequation.org/>). This algorithm assigns grayordinates to communities. Subsequently, these communities are categorized based on their similarity to established group-average networks recently updated to include the SCAN ^{20,23}. Final cortical resting state networks were derived from the consensus network assignments obtained through aggregation across thresholds.

Thalamic nuclei segmentation using THOMAS: The Thalamus-Optimized Multi-Atlas Segmentation (THOMAS v 2.1) method, a promising approach for identifying nuclei, was employed ²⁸. The choice of THOMAS followed the latest consensus of nuclei naming and automatic segmentation algorithm improvement. We also aimed to interpret our results in relation to clinical application and needed a robust localization of VIM, which has been shown to co-localized with the segment labeled VLPv (Ventral-Lateral-Posterior ventral) using the THOMAS segmentation ³¹. For the thalamic nuclei segmentation in our precision mapping participant, the `hips_thomas.csh` function from version 2.1 was utilized. This version has been validated exclusively for T1 acquisition ^{108–110} and is accessible through Docker (https://github.com/thalamicseg/thomas_new). The average T1 acquisition, generated for the registration of all functional data, was employed for this purpose. Nuclei were mapped into cifti format and the resolution of functional image (3mm) to quantify overlap between brain maps and thalamic nuclei.

rs-fMRI Functional connectivity (FC): Average BOLD time series were calculated for each individual specific vertices/voxels (i.e., cifti grayordinates) to construct a full brain FC for each rs-fMRI session. Seed FC was estimated as the average of FC maps for each voxel in the seed. Pre-cast FC maps were averaged over sessions to define the baseline FC.

rs-fMRI FC change: For casting sessions comparison to baseline (pre-cast) sessions, Cohen's *d* was calculated at each grayordinate. Cohen's *d* indexes change in FC effect size as it accounts for standard deviation. This allows higher sensitivity to subcortical casting effects as the subcortical correlation values are lower in comparison to cerebral cortex but consistent across sessions. Cohen's *d* is also more stable than the *t*-statistic to variation in the number of measurements between participants or between casting protocol phases. Due to repeated measure design, the sample size is small (only 14 sessions per phase). Thus, loss of a single session impacts the interpretation of *t*-statistic more than Cohen's *d*. Statistical significance was assessed within participants using individually generated nulls. To define significant effect, we computed null distribution FC maps by randomizing session labels 1000 times and computing a

multi threshold cluster based correction. 10 thresholds were defined within the range of values of the original data, using values of every 10th percentile of the values distribution. Cluster of data passing cluster size correction at $p < 0.05$ for at least two thresholds are displayed in figure 1. Cluster size correction was estimated for each anatomical structure (cortical and subcortical) independently.

Pulse detection and modeling: To detect large amplitude fluctuation in Left SM1_{ue} regions characteristic to a “pulse”, we look for variation of fMRI activity in Left SM1_{ue} above a standard activity and unilateral. To do so, the average BOLD time series for the Left and Right SM1_{ue} were computed for each rs-fMRI run and normalized across all runs. The difference between Left and Right time series were computed and the 2.3 times the standard deviation of this difference was used as threshold for pulse detection. A Left SM1_{ue} pulse was defined by two criteria. The first criterion was an increase of Left to Right time series difference (above 2.3 times the Left to Right time series difference standard deviation). The second criterion was an increase in signal of Left SM1_{ue} time series above 2.3 times its average time series standard deviation. To avoid movement related to false positives, Pulses with a high correlation (> 0.8) with head movement (on a 18s-window centered on the pulse peak) were removed.

In order to study the subcortical pattern of the disuse pulse described in Newbold et al. ¹⁷, we used a pulse detection analysis sensitive to potential differences in hemodynamic response function (HRF) shape ¹¹¹ in subcortical regions. We modeled, at each grayordinate, a pulse waveform using the HRF shape at each pulse (double gamma HRF function available in nipy (SPM based hrf double gamma, `nipy.modalities.fmri.hrf`) and optimized using scipy (`scipy.optimize.curve_fit`). For grayordinates that did not show a pulse activity, the model did not converge within the parameter range. Regions with the most frequent pulse activity (20% highest pulse detection) are displayed in figure 2. Pulse latency at each pulse locus was computed as the temporal difference between peaks relative to Left SM1_{ue}. The displayed map was thresholded at 1.1 seconds (TR) after the Left SM1_{ue} pulse peak.

Motor Task analysis: Task block designs for each movement condition (Tongue, Left Hand, Right Hand, Left Foot, Right Foot) were modeled using a double gamma HRF in a GLM analysis from FSL `feat`¹⁰³. Block onset and offset were modeled as independent events ¹¹². Analysis was conducted independently on surface and subcortical grayordinates, following the HCP pipeline steps (release v4.3). Second level analysis across runs within participants was performed with FSI and the resulting t-scores were used to study thalamic responses.

VIM localization: Voxels representing VIM were determined using the anterior commissure - posterior commissure (ACPC) formula ¹¹³ as follows. The T1 was aligned on the ACPC line using ACPC detect ¹¹⁴. We used the FreeSurfer segmentation of the third ventricle to estimate VIM coordinates. The coronal range was estimated as 1/4 of the third ventricle length from the posterior limit of the ventricle with 2 mm anterior range. The sagittal range was estimated between 14 mm from the center of the third ventricle and 11 mm from the border of the third ventricle. The voxels between the three axis range were brought to functional orientation and to the functional resolution of 3mm.

Quantification and testing of effects against individual null distribution: To test the significance of results per anatomical region (e.g. subcortical structure or thalamic nuclei), given spatial autocorrelation in BOLD signal, we used null distribution testing. We generated 1000

random representations at the grayordinates level, using Moran spectral randomization, an algorithm that gets informed by spatial distances between vertices^{115,116}. We compute the values of interest, i.e. average values per nuclei or correlation across voxels of the thalamus and compare the true to the random null distribution data. Tests were corrected for multiple comparisons across participants using false discovery rate.

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Competing Interests

A.N.V. and N.U.F.D. have a financial interest in Turing Medical Inc. and may benefit financially if the company is successful in marketing FIRMM motion monitoring software products. A.N.V. and N.U.F.D. may receive royalty income based on FIRMM technology developed at Washington University School of Medicine and Oregon Health and Sciences University and licensed to Turing Medical Inc. N.U.F.D. are co-founders of Turing Medical Inc. These potential conflicts of interest have been reviewed and are managed by Washington University School of Medicine, Oregon Health and Sciences University and the University of Minnesota. A.N.V. is now an employee of Turing Medical Inc. The other authors declare no competing interests.

Reference

1. von Bernhardi R, Bernhardi LE von, Eugenín J. What Is Neural Plasticity? In: von Bernhardi R, Eugenín J, Muller KJ, eds. *The Plastic Brain*. Advances in Experimental Medicine and Biology. Springer International Publishing; 2017:1-15. doi:10.1007/978-3-319-62817-2_1
2. Vicente AM, Martins GJ, Costa RM. Cortico-basal ganglia circuits underlying dysfunctional control of motor behaviors in neuropsychiatric disorders. *Curr Opin Genet Dev*. 2020;65:151. doi:10.1016/j.gde.2020.05.042
3. Jahanshahi M, Obeso I, Rothwell JC, Obeso JA. A fronto-striato-subthalamic-pallidal network for goal-directed and habitual inhibition. *Nat*

- Rev Neurosci*. 2015;16(12):719-732. doi:10.1038/nrn4038
4. La Terra D, Bjerre AS, Rosier M, Masuda R, Ryan TJ, Palmer LM. The role of higher-order thalamus during learning and correct performance in goal-directed behavior. Abdus-Saboour I, Gold JI, eds. *eLife*. 2022;11:e77177. doi:10.7554/eLife.77177
 5. Beitz JM. Parkinson's disease: a review. *Front Biosci Sch Ed*. 2014;6:65-74.
 6. Opri E, Cernera S, Okun MS, Foote KD, Gunduz A. The Functional Role of Thalamocortical Coupling in the Human Motor Network. *J Neurosci*. 2019;39(41):8124-8134. doi:10.1523/JNEUROSCI.1153-19.2019
 7. Ilyas A, Pizarro D, Romeo AK, Riley KO, Pati S. The centromedian nucleus: Anatomy, physiology, and clinical implications. *J Clin Neurosci*. 2019;63:1-7. doi:10.1016/j.jocn.2019.01.050
 8. Van der Werf YD, Witter MP, Groenewegen HJ. The intralaminar and midline nuclei of the thalamus. Anatomical and functional evidence for participation in processes of arousal and awareness. *Brain Res Rev*. 2002;39(2):107-140. doi:10.1016/S0165-0173(02)00181-9
 9. Bonelli RM, Cummings JL. Frontal-subcortical circuitry and behavior. *Dialogues Clin Neurosci*. 2007;9(2):141-151.
 10. Bennett CM, Miller MB. How reliable are the results from functional magnetic resonance imaging? *Ann N Y Acad Sci*. 2010;1191(1):133-155. doi:10.1111/j.1749-6632.2010.05446.x
 11. Maugeri L, Moraschi M, Summers P, et al. Assessing denoising strategies to increase signal to noise ratio in spinal cord and in brain cortical and subcortical regions. *J Instrum*. 2018;13(02):C02028. doi:10.1088/1748-0221/13/02/C02028
 12. Appelbaum LG, Shenasa MA, Stolz L, Daskalakis Z. Synaptic plasticity and mental health: methods, challenges and opportunities. *Neuropsychopharmacology*. 2023;48(1):113-120. doi:10.1038/s41386-022-01370-w
 13. Kolb B, Gibb R. Brain Plasticity and Behaviour in the Developing Brain. *J Can Acad Child Adolesc Psychiatry*. 2011;20(4):265-276.
 14. Mateos-Aparicio P, Rodríguez-Moreno A. The Impact of Studying Brain Plasticity. *Front Cell Neurosci*. 2019;13. Accessed February 21, 2023. <https://www.frontiersin.org/articles/10.3389/fncel.2019.00066>
 15. Newbold DJ, Dosenbach NU. Tracking plasticity of individual human brains. *Curr Opin Behav Sci*. 2021;40:161-168. doi:10.1016/j.cobeha.2021.04.018
 16. Newbold DJ, Gordon EM, Laumann TO, et al. Cingulo-opercular control network and disused motor circuits joined in standby mode. *Proc Natl Acad Sci*. 2021;118(13):e2019128118. doi:10.1073/pnas.2019128118
 17. Newbold DJ, Laumann TO, Hoyt CR, et al. Plasticity and Spontaneous Activity Pulses in Disused Human Brain Circuits. *Neuron*. 2020;107(3):580-589.e6. doi:10.1016/j.neuron.2020.05.007

18. Hubel DH, Wiesel TN. Receptive fields and functional architecture in two nonstriate visual areas (18 and 19) of the cat. *J Neurophysiol.* 1965;28(2):229-289.
19. Merzenich MM, Kaas JH, Wall J, Nelson RJ, Sur M, Felleman D. Topographic reorganization of somatosensory cortical areas 3b and 1 in adult monkeys following restricted deafferentation. *Neuroscience.* 1983;8(1):33-55. doi:10.1016/0306-4522(83)90024-6
20. Gordon EM, Laumann TO, Gilmore AW, et al. Precision Functional Mapping of Individual Human Brains. *Neuron.* 2017;95(4):791-807.e7. doi:10.1016/j.neuron.2017.07.011
21. Dosenbach NUF, Fair DA, Cohen AL, Schlaggar BL, Petersen SE. A dual-networks architecture of top-down control. *Trends Cogn Sci.* 2008;12(3):99-105. doi:10.1016/j.tics.2008.01.001
22. D'Andrea CB, Laumann TO, Newbold DJ, et al. Substructure of the brain's Cingulo-Opercular network. Published online October 10, 2023:2023.10.10.561772. doi:10.1101/2023.10.10.561772
23. Gordon EM, Chauvin RJ, Van AN, et al. A somato-cognitive action network alternates with effector regions in motor cortex. *Nature.* 2023;617(7960):351-359. doi:10.1038/s41586-023-05964-2
24. Lanciego JL, Luquin N, Obeso JA. Functional Neuroanatomy of the Basal Ganglia. *Cold Spring Harb Perspect Med.* 2012;2(12):a009621. doi:10.1101/cshperspect.a009621
25. Jahanshahi M, Obeso I, Rothwell JC, Obeso JA. A fronto-striato-subthalamic-pallidal network for goal-directed and habitual inhibition. *Nat Rev Neurosci.* 2015;16(12):719-732. doi:10.1038/nrn4038
26. Pekny SE, Izawa J, Shadmehr R. Reward-Dependent Modulation of Movement Variability. *J Neurosci.* 2015;35(9):4015-4024. doi:10.1523/JNEUROSCI.3244-14.2015
27. Rae CL, Hughes LE, Anderson MC, Rowe JB. The Prefrontal Cortex Achieves Inhibitory Control by Facilitating Subcortical Motor Pathway Connectivity. *J Neurosci.* 2015;35(2):786-794. doi:10.1523/JNEUROSCI.3093-13.2015
28. Su JH, Thomas FT, Kasoff WS, et al. Thalamus Optimized Multi Atlas Segmentation (THOMAS): fast, fully automated segmentation of thalamic nuclei from structural MRI. *NeuroImage.* 2019;194:272-282. doi:10.1016/j.neuroimage.2019.03.021
29. Mai JK, Majtanik M. Toward a Common Terminology for the Thalamus. *Front Neuroanat.* 2019;12. Accessed November 2, 2023. <https://www.frontiersin.org/articles/10.3389/fnana.2018.00114>
30. Strick PL. Activity of ventrolateral thalamic neurons during arm movement. *J Neurophysiol.* 1976;39(5):1032-1044. doi:10.1152/jn.1976.39.5.1032
31. Su JH, Choi EY, Tourdias T, et al. Improved Vim targeting for focused ultrasound ablation treatment of essential tremor: A probabilistic and

- patient-specific approach. *Hum Brain Mapp.* 2020;41(17):4769-4788. doi:10.1002/hbm.25157
32. Bertino S, Basile GA, Bramanti A, et al. Ventral intermediate nucleus structural connectivity-derived segmentation: anatomical reliability and variability. *NeuroImage.* 2021;243:118519. doi:10.1016/j.neuroimage.2021.118519
 33. Fama R, Sullivan EV. Thalamic structures and associated cognitive functions: Relations with age and aging. *Neurosci Biobehav Rev.* 2015;54:29-37. doi:10.1016/j.neubiorev.2015.03.008
 34. Shine JM, Lewis LD, Garrett DD, Hwang K. The impact of the human thalamus on brain-wide information processing. *Nat Rev Neurosci.* 2023;24(7):416-430. doi:10.1038/s41583-023-00701-0
 35. Mai JK, Majtanik M. Toward a Common Terminology for the Thalamus. *Front Neuroanat.* 2019;12:114. doi:10.3389/fnana.2018.00114
 36. Whatley BP, Chopek JW, Hill R, Brownstone RM. Case Studies in Neuroscience: Evidence of motor thalamus reorganization following bilateral forearm amputations. *J Neurophysiol.* 2018;120(4):1776-1780. doi:10.1152/jn.00120.2018
 37. Froemke RC. Plasticity of Cortical Excitatory-Inhibitory Balance. *Annu Rev Neurosci.* 2015;38:195-219. doi:10.1146/annurev-neuro-071714-034002
 38. Avoli M, de Curtis M, Gnatkovsky V, et al. Specific imbalance of excitatory/inhibitory signaling establishes seizure onset pattern in temporal lobe epilepsy. *J Neurophysiol.* 2016;115(6):3229-3237. doi:10.1152/jn.01128.2015
 39. Dehghani N, Peyrache A, Telenczuk B, et al. Dynamic Balance of Excitation and Inhibition in Human and Monkey Neocortex. *Sci Rep.* 2016;6(1):23176. doi:10.1038/srep23176
 40. Toader O, Forte N, Orlando M, et al. Dentate gyrus network dysfunctions precede the symptomatic phase in a genetic mouse model of seizures. *Front Cell Neurosci.* 2013;7:138. doi:10.3389/fncel.2013.00138
 41. Rupert DD, Shea SD. Parvalbumin-Positive Interneurons Regulate Cortical Sensory Plasticity in Adulthood and Development Through Shared Mechanisms. *Front Neural Circuits.* 2022;16:886629. doi:10.3389/fncir.2022.886629
 42. Yang JC, Bullinger KL, Isbaine F, et al. Centromedian thalamic deep brain stimulation for drug-resistant epilepsy: single-center experience. *J Neurosurg.* 2022;137(6):1591-1600. doi:10.3171/2022.2.JNS212237
 43. Cukiert A, Cukiert CM, Burattini JA, Mariani PP. Seizure outcome during bilateral, continuous, thalamic centromedian nuclei deep brain stimulation in patients with generalized epilepsy: a prospective, open-label study. *Seizure.* 2020;81:304-309. doi:10.1016/j.seizure.2020.08.028
 44. Agashe S, Burkholder D, Starnes K, et al. Centromedian Nucleus of the

- Thalamus Deep Brain Stimulation for Genetic Generalized Epilepsy: A Case Report and Review of Literature. *Front Hum Neurosci*. 2022;16. Accessed August 24, 2023. <https://www.frontiersin.org/articles/10.3389/fnhum.2022.858413>
45. Schreiner T, Kaufmann E, Noachtar S, Mehrkens JH, Staudigl T. The human thalamus orchestrates neocortical oscillations during NREM sleep. *Nat Commun*. 2022;13(1):5231. doi:10.1038/s41467-022-32840-w
 46. Timofeev I, Chauvette S. The Spindles: Are They Still Thalamic? *Sleep*. 36(6):825. doi:10.5665/sleep.2702
 47. Bastuji H, Lamouroux P, Villalba M, Magnin M, Garcia-Larrea L. Local sleep spindles in the human thalamus. *J Physiol*. 2020;598(11):2109-2124. doi:10.1113/JP279045
 48. Snyder AZ, Nishino T, Shimony JS, et al. Covariance and Correlation Analysis of Resting State Functional Magnetic Resonance Imaging Data Acquired in a Clinical Trial of Mindfulness-Based Stress Reduction and Exercise in Older Individuals. *Front Neurosci*. 2022;16:825547. doi:10.3389/fnins.2022.825547
 49. Turrigiano GG. The Self-Tuning Neuron: Synaptic Scaling of Excitatory Synapses. *Cell*. 2008;135(3):422-435. doi:10.1016/j.cell.2008.10.008
 50. Fox K, Stryker M. Integrating Hebbian and homeostatic plasticity: introduction. *Philos Trans R Soc B Biol Sci*. 2017;372(1715):20160413. doi:10.1098/rstb.2016.0413
 51. Mackwood O, Naumann LB, Sprekeler H. Learning excitatory-inhibitory neuronal assemblies in recurrent networks. Palmer SE, Ivry RB, eds. *eLife*. 2021;10:e59715. doi:10.7554/eLife.59715
 52. Bannon NM, Chistiakova M, Volgushev M. Synaptic Plasticity in Cortical Inhibitory Neurons: What Mechanisms May Help to Balance Synaptic Weight Changes? *Front Cell Neurosci*. 2020;14. Accessed August 31, 2022. <https://www.frontiersin.org/articles/10.3389/fncel.2020.00204>
 53. Yin HH, Knowlton BJ. The role of the basal ganglia in habit formation. *Nat Rev Neurosci*. 2006;7(6):464-476. doi:10.1038/nrn1919
 54. Balleine BW, Delgado MR, Hikosaka O. The role of the dorsal striatum in reward and decision-making. *J Neurosci Off J Soc Neurosci*. 2007;27(31):8161-8165. doi:10.1523/JNEUROSCI.1554-07.2007
 55. Turner KM, Svegborn A, Langguth M, McKenzie C, Robbins TW. Opposing Roles of the Dorsolateral and Dorsomedial Striatum in the Acquisition of Skilled Action Sequencing in Rats. *J Neurosci*. 2022;42(10):2039-2051. doi:10.1523/JNEUROSCI.1907-21.2022
 56. Vandaele Y, Mahajan NR, Ottenheimer DJ, Richard JM, Mysore SP, Janak PH. Distinct recruitment of dorsomedial and dorsolateral striatum erodes with extended training. *eLife*. 8:e49536. doi:10.7554/eLife.49536
 57. Charyasz E, Heule R, Molla F, et al. Functional mapping of sensorimotor activation in the human thalamus at 9.4 Tesla. *Front Neurosci*. 2023;17:1116002. doi:10.3389/fnins.2023.1116002

58. Vázquez-García M, Wallman MJ, Timofeev I. Somatotopic organization of ferret thalamus. *Front Integr Neurosci*. 2014;8:90. doi:10.3389/fnint.2014.00090
59. Kaas JH, Nelson RJ, Sur M, Dykes RW, Merzenich MM. The somatotopic organization of the ventroposterior thalamus of the squirrel monkey, *Saimiri sciureus*. *J Comp Neurol*. 1984;226(1):111-140. doi:10.1002/cne.902260109
60. Hong JH, Kwon HG, Jang SH. Probabilistic Somatotopy of the Spinothalamic Pathway at the Ventroposterolateral Nucleus of the Thalamus in the Human Brain. *Am J Neuroradiol*. 2011;32(7):1358-1362. doi:10.3174/ajnr.A2497
61. Beloozerova IN. Neuronal activity reorganization in motor cortex for successful locomotion after a lesion in the ventrolateral thalamus. *J Neurophysiol*. 2022;127(1):56-85. doi:10.1152/jn.00191.2021
62. Beloozerova IN. Neuronal activity reorganization in motor cortex for successful locomotion after a lesion in the ventrolateral thalamus. *J Neurophysiol*. 2022;127(1):56-85. doi:10.1152/jn.00191.2021
63. Favre I, Zeffiro TA, Detante O, Krainik A, Hommel M, Jaillard A. Upper limb recovery after stroke is associated with ipsilesional primary motor cortical activity: a meta-analysis. *Stroke*. 2014;45(4):1077-1083. doi:10.1161/STROKEAHA.113.003168
64. Ward NS, Brown MM, Thompson AJ, Frackowiak RSJ. Neural correlates of outcome after stroke: a cross-sectional fMRI study. *Brain J Neurol*. 2003;126(Pt 6):1430-1448. doi:10.1093/brain/awg145
65. Rockland KS. A Closer Look at Corticothalamic "Loops." *Front Neural Circuits*. 2021;15. Accessed December 6, 2023. <https://www.frontiersin.org/articles/10.3389/fncir.2021.632668>
66. Gottshall JL, Adams ZM, Forgacs PB, Schiff ND. Daytime Central Thalamic Deep Brain Stimulation Modulates Sleep Dynamics in the Severely Injured Brain: Mechanistic Insights and a Novel Framework for Alpha-Delta Sleep Generation. *Front Neurol*. 2019;10:20. doi:10.3389/fneur.2019.00020
67. Arnts H, Coolen SE, Fernandes FW, et al. The intralaminar thalamus: a review of its role as a target in functional neurosurgery. *Brain Commun*. 2023;5(3):fcad003. doi:10.1093/braincomms/fcad003
68. Gent TC, Bandarabadi M, Herrera CG, Adamantidis AR. Thalamic dual control of sleep and wakefulness. *Nat Neurosci*. 2018;21(7):974-984. doi:10.1038/s41593-018-0164-7
69. Latchoumane CFV, Ngo HVV, Born J, Shin HS. Thalamic Spindles Promote Memory Formation during Sleep through Triple Phase-Locking of Cortical, Thalamic, and Hippocampal Rhythms. *Neuron*. 2017;95(2):424-435.e6. doi:10.1016/j.neuron.2017.06.025
70. Hahn MA, Heib D, Schabus M, Hoedlmoser K, Helfrich RF. Slow oscillation-spindle coupling predicts enhanced memory formation from

- childhood to adolescence. Haegens S, Colgin LL, eds. *eLife*. 2020;9:e53730. doi:10.7554/eLife.53730
71. Klinzing JG, Mölle M, Weber F, et al. Spindle activity phase-locked to sleep slow oscillations. *NeuroImage*. 2016;134:607-616. doi:10.1016/j.neuroimage.2016.04.031
 72. Gent TC, Bassetti CL, Adamantidis AR. Sleep-wake control and the thalamus. *Curr Opin Neurobiol*. 2018;52:188-197. doi:10.1016/j.conb.2018.08.002
 73. Frank MG. Erasing synapses in sleep: is it time to be SHY? *Neural Plast*. 2012;2012:264378. doi:10.1155/2012/264378
 74. Tononi G, Cirelli C. Sleep and the Price of Plasticity: From Synaptic and Cellular Homeostasis to Memory Consolidation and Integration. *Neuron*. 2014;81(1):12-34. doi:10.1016/j.neuron.2013.12.025
 75. Esser SK, Hill SL, Tononi G. Sleep Homeostasis and Cortical Synchronization: I. Modeling the Effects of Synaptic Strength on Sleep Slow Waves. *Sleep*. 2007;30(12):1617-1630.
 76. Guo D, Thomas RJ, Liu Y, Shea SA, Lu J, Peng CK. Slow wave synchronization and sleep state transitions. *Sci Rep*. 2022;12(1):7467. doi:10.1038/s41598-022-11513-0
 77. Lewis LD. The interconnected causes and consequences of sleep in the brain. *Science*. 2021;374(6567):564-568. doi:10.1126/science.abi8375
 78. Vyazovskiy VV, Olcese U, Hanlon EC, Nir Y, Cirelli C, Tononi G. Local sleep in awake rats. *Nature*. 2011;472(7344):443-447. doi:10.1038/nature10009
 79. Laumann TO, Snyder AZ. Brain activity is not only for thinking. *Curr Opin Behav Sci*. 2021;40:130-136. doi:10.1016/j.cobeha.2021.04.002
 80. Albano L, Agosta F, Basaia S, et al. Functional connectivity in Parkinson's disease candidates for deep brain stimulation. *Npj Park Dis*. 2022;8(1):1-12. doi:10.1038/s41531-021-00268-6
 81. Hensel L, Hoffstaedter F, Caspers J, et al. Functional Connectivity Changes of Key Regions for Motor Initiation in Parkinson's Disease. *Cereb Cortex N Y NY*. 2019;29(1):383-396. doi:10.1093/cercor/bhy259
 82. Gao L lin, Wu T. The study of brain functional connectivity in Parkinson's disease. *Transl Neurodegener*. 2016;5(1):18. doi:10.1186/s40035-016-0066-0
 83. Owens-Walton C, Jakabek D, Power BD, et al. Increased functional connectivity of thalamic subdivisions in patients with Parkinson's disease. *PLOS ONE*. 2019;14(9):e0222002. doi:10.1371/journal.pone.0222002
 84. Yu R, Liu B, Wang L, Chen J, Liu X. Enhanced Functional Connectivity between Putamen and Supplementary Motor Area in Parkinson's Disease Patients. *PLOS ONE*. 2013;8(3):e59717. doi:10.1371/journal.pone.0059717
 85. Wu T, Wang L, Hallett M, Chen Y, Li K, Chan P. Effective connectivity of brain networks during self-initiated movement in Parkinson's disease.

- NeuroImage*. 2011;55(1):204-215.
doi:10.1016/j.neuroimage.2010.11.074
86. Simioni AC, Dagher A, Fellows LK. Compensatory striatal–cerebellar connectivity in mild–moderate Parkinson’s disease. *NeuroImage Clin*. 2016;10:54-62. doi:10.1016/j.nicl.2015.11.005
 87. Binder T, Hobert MA, Pfrommer T, et al. Increased functional connectivity in a population at risk of developing Parkinson’s disease. *Parkinsonism Relat Disord*. 2021;92:1-6. doi:10.1016/j.parkreldis.2021.09.026
 88. Gratton C, Koller JM, Shannon W, et al. Emergent Functional Network Effects in Parkinson Disease. *Cereb Cortex N Y N 1991*. 2019;29(6):2509-2523. doi:10.1093/cercor/bhy121
 89. Hacker CD, Perlmutter JS, Criswell SR, Ances BM, Snyder AZ. Resting state functional connectivity of the striatum in Parkinson’s disease. *Brain J Neurol*. 2012;135(Pt 12):3699-3711. doi:10.1093/brain/aws281
 90. Fukai T. Computational models of Idling brain activity for memory processing. *Neurosci Res*. 2023;189:75-82. doi:10.1016/j.neures.2022.12.024
 91. Wichmann T. Changing views of the pathophysiology of Parkinsonism. *Mov Disord Off J Mov Disord Soc*. 2019;34(8):1130-1143. doi:10.1002/mds.27741
 92. Cheung THC, Ding Y, Zhuang X, Kang UJ. Learning critically drives parkinsonian motor deficits through imbalanced striatal pathway recruitment. *Proc Natl Acad Sci*. 2023;120(12):e2213093120. doi:10.1073/pnas.2213093120
 93. Ren J, Zhang W, Dahmani L, et al. The somato-cognitive action network links diverse neuromodulatory targets for Parkinson’s disease. Published online December 14, 2023:2023.12.12.571023. doi:10.1101/2023.12.12.571023
 94. Tai CH. Subthalamic burst firing: A pathophysiological target in Parkinson’s disease. *Neurosci Biobehav Rev*. 2022;132:410-419. doi:10.1016/j.neubiorev.2021.11.044
 95. Cagnan H, Mallet N, Moll CKE, et al. Temporal evolution of beta bursts in the parkinsonian cortical and basal ganglia network. *Proc Natl Acad Sci*. 2019;116(32):16095-16104. doi:10.1073/pnas.1819975116
 96. West TO, Berthouze L, Halliday DM, et al. Propagation of beta/gamma rhythms in the cortico-basal ganglia circuits of the parkinsonian rat. *J Neurophysiol*. 2018;119(5):1608-1628. doi:10.1152/jn.00629.2017
 97. Sharott A, Vinciati F, Nakamura KC, Magill PJ. A Population of Indirect Pathway Striatal Projection Neurons Is Selectively Entrained to Parkinsonian Beta Oscillations. *J Neurosci Off J Soc Neurosci*. 2017;37(41):9977-9998. doi:10.1523/JNEUROSCI.0658-17.2017
 98. Torrecillos F, Tinkhauser G, Fischer P, et al. Modulation of Beta Bursts in the Subthalamic Nucleus Predicts Motor Performance. *J Neurosci*.

- 2018;38(41):8905-8917. doi:10.1523/JNEUROSCI.1314-18.2018
99. Vinding MC, Tsitsi P, Waldthaler J, et al. Reduction of spontaneous cortical beta bursts in Parkinson's disease is linked to symptom severity. *Brain Commun.* 2020;2(1):fcaa052. doi:10.1093/braincomms/fcaa052
100. Ray NJ, Jenkinson N, Wang S, et al. Local field potential beta activity in the subthalamic nucleus of patients with Parkinson's disease is associated with improvements in bradykinesia after dopamine and deep brain stimulation. *Exp Neurol.* 2008;213(1):108-113. doi:10.1016/j.expneurol.2008.05.008
101. Hashemi NS, Dehnavi F, Moghimi S, Ghorbani M. Slow spindles are associated with cortical high frequency activity. *NeuroImage.* 2019;189:71-84. doi:10.1016/j.neuroimage.2019.01.012
102. Oldfield RC. The assessment and analysis of handedness: The Edinburgh inventory. *Neuropsychologia.* 1971;9(1):97-113. doi:10.1016/0028-3932(71)90067-4
103. Barch DM, Burgess GC, Harms MP, et al. Function in the human connectome: Task-fMRI and individual differences in behavior. *NeuroImage.* 2013;80:169-189. doi:10.1016/j.neuroimage.2013.05.033
104. Bizzi A, Blasi V, Falini A, et al. Presurgical functional MR imaging of language and motor functions: Validation with intraoperative electrocortical mapping. *Radiology.* 2008;248(2):579-589. doi:10.1148/radiol.2482071214
105. Raut RV, Mitra A, Snyder AZ, Raichle ME. On time delay estimation and sampling error in resting-state fMRI. *NeuroImage.* 2019;194:211-227. doi:10.1016/j.neuroimage.2019.03.020
106. Fischl B. FreeSurfer. *NeuroImage.* 2012;62(2):774-781. doi:10.1016/j.neuroimage.2012.01.021
107. Rosvall M, Bergstrom CT. Maps of random walks on complex networks reveal community structure. *Proc Natl Acad Sci U S A.* 2008;105(4):1118-1123. doi:10.1073/pnas.0706851105
108. Pfefferbaum A, Sullivan EV, Zahr NM, Pohl KM, Saranathan M. Multi-atlas thalamic nuclei segmentation on standard T1-weighted MRI with application to normal aging. *Hum Brain Mapp.* 2023;44(2):612-628. doi:10.1002/hbm.26088
109. Vidal JP, Danet L, Péran P, et al. Robust thalamic nuclei segmentation from T1-weighted MRI. arXiv.org. Published April 14, 2023. Accessed January 10, 2024. <https://arxiv.org/abs/2304.07167v1>
110. Williams B, Nguyen D, Vidal J, Initiative ADN, Saranathan M. Thalamic nuclei segmentation from T₂-weighted MRI: unifying and benchmarking state-of-the-art methods with young and old cohorts. arXiv.org. Published September 26, 2023. Accessed January 10, 2024. <https://arxiv.org/abs/2309.15053v1>
111. Bielsczyk NZ, Llera A, Buitelaar JK, Glennon JC, Beckmann CF. The impact of hemodynamic variability and signal mixing on the identifiability

- of effective connectivity structures in BOLD fMRI. *Brain Behav.* 2017;7(8):n/a-n/a. doi:10.1002/brb3.777
112. Dosenbach NUF, Visscher KM, Palmer ED, et al. A core system for the implementation of task sets. *Neuron.* 2006;50(5):799-812. doi:10.1016/j.neuron.2006.04.031
113. Acar F, Miller JP, Berk MC, Anderson G, Burchiel KJ. Safety of anterior commissure-posterior commissure-based target calculation of the subthalamic nucleus in functional stereotactic procedures. *Stereotact Funct Neurosurg.* 2007;85(6):287-291. doi:10.1159/000107361
114. Ardekani BA, Bachman AH. Model-based automatic detection of the anterior and posterior commissures on MRI scans. *NeuroImage.* 2009;46(3):677-682. doi:10.1016/j.neuroimage.2009.02.030
115. Markello RD, Misic B. Comparing spatial null models for brain maps. *NeuroImage.* 2021;236:118052. doi:10.1016/j.neuroimage.2021.118052
116. Markello RD, Hansen JY, Liu ZQ, et al. neuromaps: structural and functional interpretation of brain maps. *Nat Methods.* Published online October 6, 2022:1-8. doi:10.1038/s41592-022-01625-w