

# Gibberellins and Heterosis in Maize<sup>1</sup>

## II. RESPONSE TO GIBBERELIC ACID AND METABOLISM OF [<sup>3</sup>H]GIBBERELLIN A<sub>20</sub>

Received for publication July 14, 1982 and in revised form November 8, 1982

STEWART B. ROOD, TERENCE J. BLAKE, AND RICHARD P. PHARIS<sup>2</sup>

Faculty of Forestry, University of Toronto, Toronto, Ontario M5S 1A1 Canada (S. B. R., T. J. B.); and  
Department of Biology, University of Calgary, Calgary, Alberta T2N 1N4 Canada (R. P. P.)

### ABSTRACT

Two maize inbreds, CM7 and CM49, and CM7 × CM49, their F<sub>1</sub> hybrid (which displayed significant heterosis), were examined with regard to response to exogenous gibberellin A<sub>3</sub> (GA<sub>3</sub>), and in their ability to metabolize GA<sub>20</sub>, a native GA of maize. The leaf sheath elongation response to GA<sub>3</sub> was far greater for the inbreds than for their hybrid. The inbreds also displayed significant elongation of the leaf blades in response to GA<sub>3</sub>, whereas the hybrid was unaffected. Promotion of cell division in the leaf sheath of CM7 and the hybrid was effected by GA<sub>3</sub>, but no promotion of cell elongation was observed in CM49, even though significant leaf sheath elongation occurred. Shoot dry weight of both inbreds was significantly increased by GA<sub>3</sub>, but response by the hybrid in this parameter was slight and variable. Root dry weight of CM7 was significantly increased by GA<sub>3</sub>, but was unchanged in CM49 and the hybrid. Thus, inbred shoot dry weight increases effected by GA<sub>3</sub> were not at the expense of the root system. Rapid metabolism of [2,3-<sup>3</sup>H]GA<sub>20</sub> occurred in all genotypes, although genotypic differences were observed. The hybrid had the highest rates of metabolism to GA glucosyl conjugate-like substances. Oxidative metabolism was also fastest in the hybrid, followed by CM7, and slowest in CM49, the slowest-growing inbred. Thus, rate of GA<sub>20</sub> metabolism is under genetic control in normal (*i.e.* not dwarfed) maize genotypes. These results, taken together with previous reports that the hybrid has significantly enhanced levels of endogenous GA-like substances, suggest that GA play a role in the expression of heterosis in maize.

Although heterosis in yield and/or growth performance is agriculturally, horticulturally, and silviculturally important, its physiological basis has not been determined (21, 22). The involvement of plant hormones is a logical possibility inasmuch as they are known to act in regulatory roles in a wide range of processes, from seed germination through vegetative growth, reproductive development, and seed development. Heterosis has been reported for components of all of these aspects of plant growth and development (22).

In a previous paper in this series (21), we reported that increased level of endogenous GA<sup>3</sup>-like substances was correlated with heterosis for vigorous growth in a maize hybrid. Nickerson (13, 14) noted that responsiveness to exogenous GA<sub>3</sub> was correlated with degree of inbreeding in maize; inbreds were very responsive,

whereas hybrids were somewhat less affected. Nickerson's (13, 14) study thus suggests that a causal role for GA may exist in heterosis. Inasmuch as maize inbreds are very responsive to exogenous GA<sub>3</sub>, it is possible that their low endogenous GA content (21) may be one factor limiting their growth, and hence, underlying the growth depression shown by the inbreds.

Because heterosis for increased levels of endogenous GA-like substances occurs (21), it is of interest to investigate the metabolic basis for this heterosis. Is GA biosynthesis more rapid in hybrids, and/or does catabolism/conjugation proceed more slowly? Either possibility could lead to increased endogenous GA levels in the hybrids. Unfortunately, it is currently not possible to evaluate the biosynthesis of GA from GA precursors *in vivo*. However, we can examine biosynthesis of di- and tri-hydroxylated GA from monohydroxylated precursors, and we can also follow the catabolism and/or conjugation through the use of radioactive GA. The principal native GA of developing maize tassels have recently been characterized (8) and at least certain parts of the metabolic sequence have been tentatively determined (5, 9, 20). Consequently, we can examine rates of metabolism of native GA in inbreds and their hybrid.

Thus, in this second paper about GA and heterosis in maize, response to exogenous GA<sub>3</sub> and metabolism of [<sup>3</sup>H]GA<sub>20</sub> in two maize inbreds and their F<sub>1</sub> hybrid are described.

### MATERIALS AND METHODS

**Plant Materials.** Two early maturing maize inbreds, CM7 and CM49, were obtained from Dr. J. Giesbrecht, Agriculture Canada Research Station, Morden Manitoba. Self-pollinations and CM7 (male) × CM49 (female) crosses were made in the summer of 1980 at the Agriculture Canada Research Station, Lethbridge, Alberta. After black layer maturity had been reached, cobs were harvested and allowed to air-dry for 1 month prior to shelling. Seeds were stored at 5°C until planting.

On January 6, 1982, five seeds of one genotype were planted 2 cm deep in each 3-L plastic pot filled with 3:1 (v:v) mixture of vermiculite and peat moss. Growth took place in a greenhouse at Toronto, Ontario (latitude 43°N) at 25/15 (±3)°C (day/night), the natural daylength being extended with 24 h low-intensity incandescent light (50 μE s<sup>-1</sup> m<sup>-2</sup>) to prevent sex reversals (18) of the apical inflorescence. Following emergence, seedlings were thinned to the largest three plants. Pots were watered to saturation once daily except once every 10 d when 200 ml with 0.25 g/pot of 28-14-14 (N-P-K) fertilizer was added.

Numbers of seedlings emerged were assessed daily. Numbers of leaves (*e.g.* numbers of ligules emerged from the leaf whorl), and heights to the highest extended leaf tip were measured weekly. Five plants of each genotype were harvested weekly to determine total leaf areas (measured with a Wescor LI 3000 area meter) and dry weights of roots and shoots after oven drying for 2 d at 70°C. Collodion peels were used to measure lengths of long adaxial

<sup>1</sup> Supported in part by Natural Sciences and Engineering Research Council Grants A-7815 and A-2585 to T. J. B. and R. P. P., respectively.

<sup>2</sup> To whom requests for reprints should be sent.

<sup>3</sup> Abbreviations: GA, gibberellin(s); EtOH, ethanol; MeOH, methanol; EtOAc, ethyl acetate; C/D R GA<sub>20</sub>, C/D ring rearranged GA<sub>20</sub>; RC, radioactivity counting.

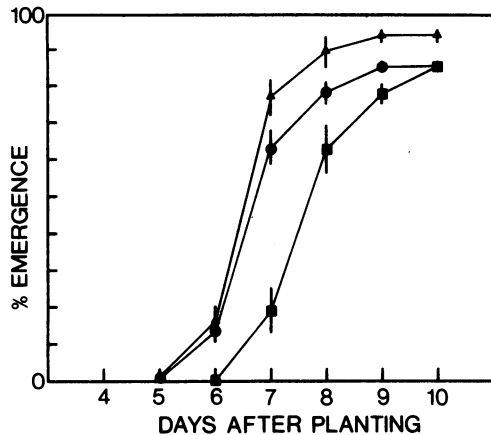


FIG. 1. Cumulative seedling emergence versus time for two maize inbreds (CM7 [●], CM49 [■]) and their F<sub>1</sub> hybrid (CM7 × CM49 [▲]) at about 25/15°C (day/night). Mean ± SE.

epidermal cells of the leaf sheath within the 10-cm region basal to the ligule of the fourth leaf. Cell number in a microscope field of view was counted as an inverse measure of cell length.

**Exogenous Application of GA<sub>3</sub>.** Crystalline GA<sub>3</sub> was dissolved in 95% EtOH and diluted to 10% aqueous EtOH containing 0, 0.1, 0.5, or 1.0 mg GA<sub>3</sub>/100 μl solution. On February 2 and 3, 1982, 50 μl of treatment solution was pipetted into the leaf whorl of each plant. Similar applications were carried out on February 9 and 10, 1982 on different plants. For each treatment, all three plants within a pot were treated similarly, and values from these three were averaged to produce a single experimental unit. Four replications of each treatment were carried out. Thus, 96 pots were included representing the three genotypes × two dates × four levels × four replicates, and 288 plants (96 pots × 3 plants) were studied.

After exogenous application of GA<sub>3</sub>, heights to the highest extended leaf tip, and to all emerged ligules, were measured every 2 d and destructive sampling for shoot and root dry weights and leaf areas occurred weekly. The significance of treatments was determined by one-way analyses of variance.

**Application of [<sup>3</sup>H]GA<sub>20</sub>.** Preparation and purification of [2,3-<sup>3</sup>H]GA<sub>20</sub> was described previously (20). In the present study, 0.5 μCi [<sup>3</sup>H]GA<sub>20</sub> in 2 μl of 20% aqueous EtOH was injected into the apical meristem of each of six plants of each genotype at 10 AM on

February 9, 1982. Three plants of each genotype were harvested 24 and at 48 h after application (e.g. six sample plants). The basal 2 cm of the shoot cylinder (which contained the terminal shoot apical meristem) was removed and immediately ground in 15 ml of 80% aqueous MeOH at -20°C. The six samples from each genotype were analyzed separately.

**Chromatography of Metabolites.** After 24 h of intermittent shaking at 2°C, the MeOH extracts were vacuum filtered and reduced to dryness *in vacuo* at 35°C. To promote removal of water, 20 ml of MeOH was added part way through solvent evaporation. Ten μl H<sub>2</sub>O (to dissolve highly water-soluble components), and then 1 ml of 50:50 MeOH:EtOAc was added, and this solution was loaded onto glass-fiber filter paper discs and dried. These discs were placed on silicic acid (SiO<sub>2</sub>) partition columns (6) (the column being made with 4.2 g Woelm SiO<sub>2</sub> for partition to which 20% H<sub>2</sub>O by weight was added, shaken, and allowed to equilibrate for 7 d before use), which were stepwise eluted with 50 ml 55:45 HCOOH-saturated EtOAc:hexane, 40 ml 95:05 EtOAc:hexane, and finally MeOH. Five-ml fractions were collected and elution solvents evaporated *in vacuo*. The fractions were solubilized in MeOH, and detection of <sup>3</sup>H was performed by liquid scintillation spectroscopy. The partition column SiO<sub>2</sub> system will elute acidic GA (excepting GA<sub>30</sub>) in the hexane:EtOAc while GA glucosyl conjugates are retained until the MeOH wash.

Radioactive fractions from the three SiO<sub>2</sub> partition columns of each replicate sample were bulked, and then chromatographed (1-min fractions, 1.8 ml) on reverse-phase C<sub>18</sub> HPLC (12, 20). Due to the abundance of pigments in certain SiO<sub>2</sub> fractions, additional aliquots were removed to verify levels of radioactivity ([<sup>3</sup>H]). Consequently, unequal proportions of fractions from the SiO<sub>2</sub> partition columns were loaded onto the C<sub>18</sub> HPLC columns, and reverse-phase elution profiles (Fig. 7) must be considered as qualitative in nature; they are not quantitatively representative. For both forms of chromatography, [<sup>3</sup>H]GA<sub>20</sub> and authentic standards of logical acidic metabolites and of conjugates of logical acidic metabolites were analyzed for comparative purposes (20).

## RESULTS AND DISCUSSION

**Growth.** Heterosis was observed very early in the life cycle of the hybrid, the order of seedling emergence rate was CM7 × CM49 > CM7 >> CM49 (Fig. 1). Final percentage seedling emergence was higher in the hybrid than in either parent (Fig. 1). The superior seedling emergence of the hybrid cannot be ex-

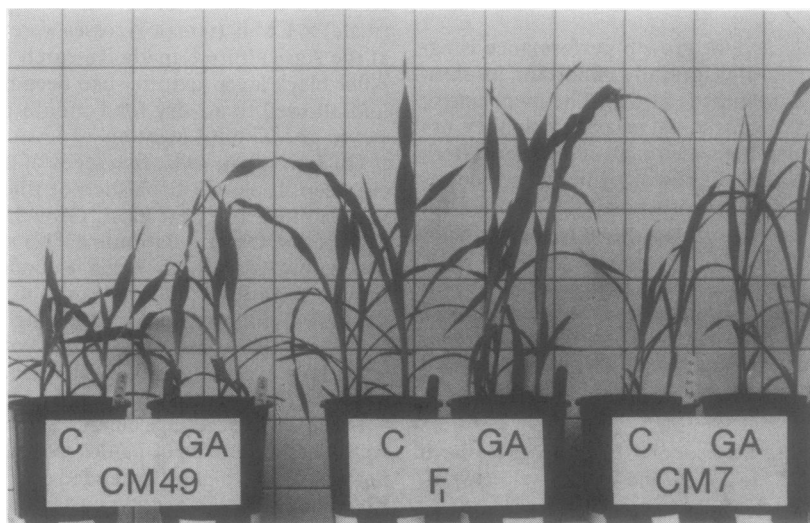


FIG. 2. Two maize inbreds and their F<sub>1</sub> hybrid 50 d after planting and 14 d after application of 100 μl 10% EtOH (C) or the same solution containing 0.5 mg GA<sub>3</sub> (+GA).

Table I. Average Lengths of Long Adaxial Epidermal Cells of the 10-cm Region Basal to the Ligule of the Fourth Leaf, and Shoot and Root Dry Weights of Two Maize Inbreds and Their F<sub>1</sub> Hybrid  
Measurements were taken 14 d after the addition of 0.5 mg GA<sub>3</sub> or a control (C) solution (mean ± SE).

| Treatment  | Cell Length     | Dry Wt        |               |
|------------|-----------------|---------------|---------------|
|            |                 | Shoot         | Root          |
|            | <i>mm</i>       |               | <i>g</i>      |
| Inbred     |                 |               |               |
| CM7        | C               | 0.155 ± 0.003 | 0.450 ± 0.053 |
|            | GA <sub>3</sub> | 0.161 ± 0.004 | 0.881 ± 0.141 |
| CM49       | C               | 0.084 ± 0.002 | 0.667 ± 0.039 |
|            | GA <sub>3</sub> | 0.119 ± 0.003 | 0.842 ± 0.043 |
| Hybrid     |                 |               |               |
| CM7 × CM49 | C               | 0.127 ± 0.003 | 2.007 ± 0.119 |
|            | GA <sub>3</sub> | 0.151 ± 0.005 | 1.967 ± 0.141 |

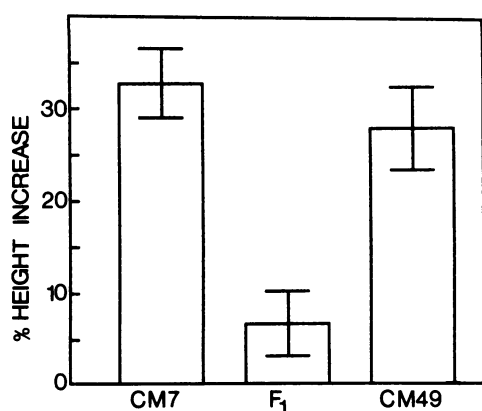


FIG. 3. Percentage change in total height of two inbreds and their F<sub>1</sub> hybrid 14 d after the addition of 0.5 mg GA<sub>3</sub>. Mean ± SE.

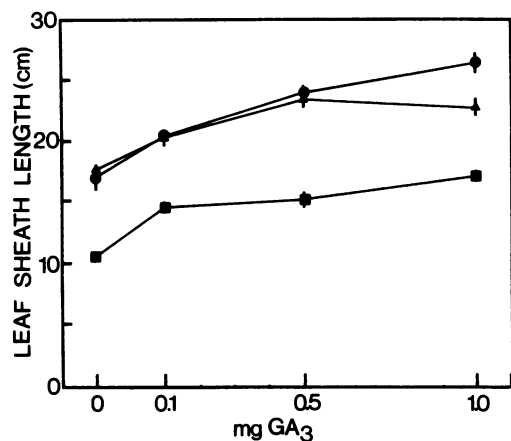


FIG. 4. Distance to uppermost ligule (e.g. leaf sheath length) of two maize inbred (CM7 [●], CM49 [■]) and their F<sub>1</sub> hybrid (CM7 × CM49 [▲]) 14 d after the addition of 0, 0.1, 0.5, or 1.0 mg GA<sub>3</sub>. Mean ± SE.

plained simply in terms of larger seeds since hybrid seed weights ( $0.177 \pm 0.009$  g seed<sup>-1</sup>) were similar to CM49 ( $0.192 \pm 0.007$  g), while CM7 seeds were smaller ( $0.114 \pm 0.004$  g). Thus, larger seeds do not necessarily support more vigorous seedlings. The similarity in seed weights for CM49 and the hybrid was expected as CM49 was the female parent. Consistent with our repeated previous observations (19, 21), growth of the hybrid continued to exceed growth of either inbred throughout the experiment (Figs. 2-4).

**Response to Exogenous GA.** Growth of the maize inbreds was

dramatically promoted following the exogenous application of GA<sub>3</sub> (Figs. 2-4), while growth promotion of the hybrid by GA<sub>3</sub> was nominal (Figs. 2-4). The growth promotion of both inbreds by GA<sub>3</sub> was statistically significant ( $P \geq 0.05$ ) at all levels of GA<sub>3</sub> on either date of application (data not presented). Leaf orientation of all three genotypes was changed such that leaf blades were more vertical (Fig. 2); the implications of this effect on photosynthetic efficiency of the standing crop warrant further study.

Elongation growth (expressed as a percentage) of the hybrid was only marginally altered, whereas growth of each parent was dramatic (Fig. 3). This trend is also present for absolute values; control and GA<sub>3</sub>-treated plants under the conditions in Figure 3 average  $52.3 \pm 1.7$  and  $69.6 \pm 2.0$  cm for CM7,  $43.6 \pm 0.6$  and  $55.8 \pm 2.0$  cm for CM49, and  $68.7 \pm 1.6$  and  $73.4 \pm 2.5$  cm for CM7 × CM49. Thus, while GA<sub>3</sub> elicited an increase of 4.7 cm in height growth for the hybrid, increases in the inbreds were 17.3 and 12.2 cm for CM7 and CM49, respectively.

Growth of the leaf sheaths of all genotypes was promoted ( $P \leq 0.05$ ) following exogenous application of GA<sub>3</sub> (Fig. 4). Again, growth promotion was most conspicuous in the inbreds. Elongation of the leaf sheath is one component of total plant height, the other being elongation of the leaf blades (internode elongation had not yet begun in these plants). Elongation of leaf blades was significantly promoted by GA<sub>3</sub> in both inbreds ( $P \leq 0.05$ ), but remained unaffected in the hybrid.

Epidermal cell length in CM7 leaf sheath number 4 was not significantly affected by GA<sub>3</sub>, but cell length of this region of CM49 was significantly increased (Table I). Since the leaf sheath of CM7 elongated without significant cell elongation, it is probable that increased cell division was responsible for the leaf sheath elongation. In CM49, however, the sheath elongation could be attributed largely to cell elongation (Table I). The modest sheath elongation noted in the hybrid after GA<sub>3</sub> application was also a result of cell elongation.

In response to GA<sub>3</sub>, shoot dry weight of both inbreds was increased by GA<sub>3</sub> ( $P \leq 0.05$ ) (Table I). Response of the hybrid was negligible and variable (Table I). The promotion of shoot growth in the inbreds was not at the expense of root growth as root dry weight was also increased by GA<sub>3</sub> in CM7, and remained unchanged in CM49 (Table I). Consequently, total plant dry weights of both inbreds (but not the hybrid) were promoted ( $P \leq 0.05$ ) by the application of GA<sub>3</sub>. Thus, almost all growth parameters were consistent in showing that exogenous GA<sub>3</sub> promoted growth (dry weight), particularly elongation, in these inbreds. Gibberellin A<sub>3</sub> had much less effect on the growth and elongation of the hybrid. Our observation that inbreds are more responsive to GA<sub>3</sub> than a hybrid is consistent with previous reports by Nickerson (13) and Nickerson and Embler (14).

**Metabolism of [<sup>3</sup>H]GA<sub>20</sub>.** Three principal regions of radioactiv-

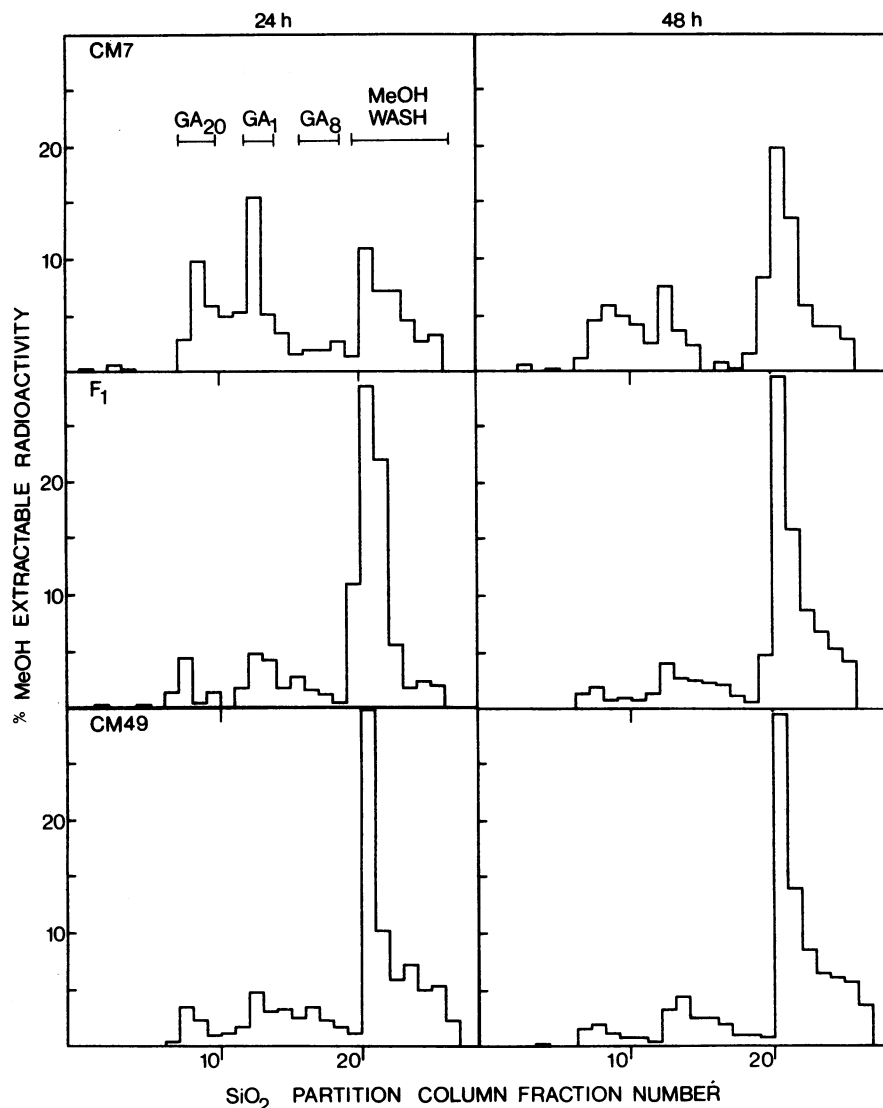


FIG. 5. Elution profiles of radioactivity  $^3\text{H}$  from typical replicate MeOH extracts chromatographed on a stepwise-eluted  $\text{SiO}_2$  partition column from two maize inbreds and their  $F_1$  hybrid (age, 24 d) harvested 24 or 48 h after feeding [ $^3\text{H}$ ]GA $_{20}$ . Retention times of authentic standards, and fraction grouping are shown to upper left figure. Any GA glucosyl conjugates present in the methanolic extract would be eluted in the MeOH wash fraction grouping.

ity were eluted from the  $\text{SiO}_2$  partition columns (Fig. 5). The first was coincidental with [ $^3\text{H}$ ]GA $_{20}$ , the second with [ $^3\text{H}$ ]GA $_1$  (and other GA, including GA $_{20}$ ), and the third occurred in the MeOH wash where GA glucosyl conjugates would elute. A minor region of radioactivity eluted coincidental with [ $^3\text{H}$ ]GA $_8$  (Fig. 5). In all genotypes, the amount of radioactivity in peak I ([ $^3\text{H}$ ]GA $_{20}$ ) decreased over the 48-h period (Figs. 5 and 6). However, there were significant ( $P \leq 0.05$ ) differences in the relative amount of radioactivity in peak I, samples from CM7 being consistently higher than CM49 and the hybrid (Fig. 6). The level of radioactivity in peak II was also considerably higher in CM7 than in CM49 and the hybrid (Figs. 5 and 6). The amount of radioactivity in the MeOH wash fraction (which would contain GA glucosyl conjugates) increased during the experimental period for all genotypes and was consistently highest in the hybrid (Fig. 6).

When fractions I and II (Fig. 6) were chromatographed on gradient-eluted  $C_{18}$  reverse-phase HPLC, at least four regions of radioactivity were eluted (Fig. 7). These were coincidental with [ $^3\text{H}$ ]GA $_8$ , [ $^3\text{H}$ ]GA $_1$ , [ $^3\text{H}$ ]GA $_{20}$ , and [ $^3\text{H}$ ]C/D R GA $_{20}$ , respectively. Amounts of radioactivity (Fig. 7) were inadequate for subsequent GLC-RC to be carried out on any of these four regions. However,

these identifications based on sequential  $\text{SiO}_2$  partition  $\rightarrow$  gradient-eluted reverse-phase  $C_{18}$  HPLC are consistent with isocratic  $C_{18}$  HPLC-RC and/or GLC-RC of similar [ $^3\text{H}$ ]GA $_{20}$  metabolites in maize (20). The C/D R GA $_{20}$ -like metabolite may be an experimental artifact (20), although, interestingly, only CM49 yielded detectable amounts of it (Fig. 7). The metabolic sequence of GA $_{20} \rightarrow$  GA $_1 \rightarrow$  GA $_8$  is both logical and consistent with previous work on several maize genotypes, including the hybrid CM7  $\times$  CM49 (5, 20). The [ $^3\text{H}$ ]GA $_8$ -like metabolite was observed only in the hybrid, while the [ $^3\text{H}$ ]GA $_1$ -like metabolite was observed in all genotypes (Figs. 5 and 7).

A number of GA glucosyl conjugate-like metabolites originating in the MeOH wash of the  $\text{SiO}_2$  partition column were eluted from the  $C_{18}$  HPLC (Fig. 8). Subsequent analyses by isocratic  $C_{18}$  HPLC or GLC have not been performed due to limited amounts. However, it is known that GA glucosyl ether or ester conjugates are eluted from  $C_{18}$  HPLC coincidental with, or a few min prior to, the acidic GA (12). Thus, HPLC peak I and II (Fig. 8) is probably a conjugate(s) of [ $^3\text{H}$ ]GA $_8$ . Peak IV from HPLC (Fig. 8) probably represents a conjugate(s) of [ $^3\text{H}$ ]GA $_1$ . Likewise, Peak VI probably represents a conjugate(s) [ $^3\text{H}$ ]GA $_{20}$ , and peak VII a conjugate(s)

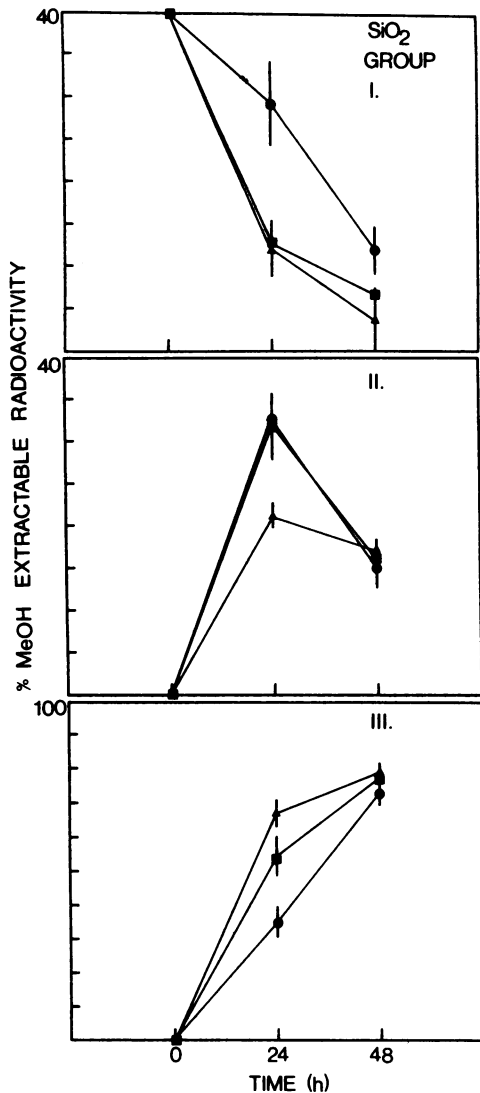


FIG. 6. Metabolism over 48 h of [ $^3\text{H}$ ]GA<sub>20</sub> into acidic and GA glycosyl conjugate-like substances by 24-d-old maize inbreds (CM7 (●) and CM49 (■)) and their F<sub>1</sub> hybrid (CM7 × CM49 (▲)). The SiO<sub>2</sub> partition column elution profile of typical replicate MeOH extracts is shown in Figure 5. Mean ± SE. SiO<sub>2</sub> group I would contain [ $^3\text{H}$ ]GA<sub>20</sub>, group II would contain [ $^3\text{H}$ ]GA<sub>1</sub> and GA<sub>8</sub>, whereas group III consisted of the MeOH wash.

of [ $^3\text{H}$ ]C/D R GA<sub>20</sub> (20).

The GA<sub>8</sub>-like metabolite (Fig. 7, peak I) occurred only in the hybrid, and the GA<sub>8</sub> glycosyl conjugate-like metabolite (Fig. 8, peak II) occurred only in the hybrid and CM7.

Results from SiO<sub>2</sub> partition (Fig. 5) and reverse-phase C<sub>18</sub> HPLC (Fig. 7) were consistent, and indicated that oxidative metabolism of [ $^3\text{H}$ ]GA<sub>20</sub> was more rapid in CM7 and the hybrid than in CM49 (Fig. 6). Also, rate of apparent conjugation (Fig. 6) was greater in the hybrid than in either inbred, and greater in CM49 than in CM7 (Figs. 5 and 6).

Thus, the rate of GA metabolism is under genetic control in normal (*i.e.* not dwarfed) maize genotypes. However, the results do not explain the apparent abundance of GA in the hybrid (21). They do suggest, however, that the rate of metabolism of [ $^3\text{H}$ ]GA<sub>20</sub>, and probably that of [ $^3\text{H}$ ]GA<sub>1</sub>, is more rapid in the hybrid than in either inbred parent. An intriguing question would be: does increased metabolism in the hybrid reflect increased endogenous GA levels.

**GA and Heterosis.** The data presented herein and in Rood *et al.* (21) contributes to an understanding of the possible role of GA

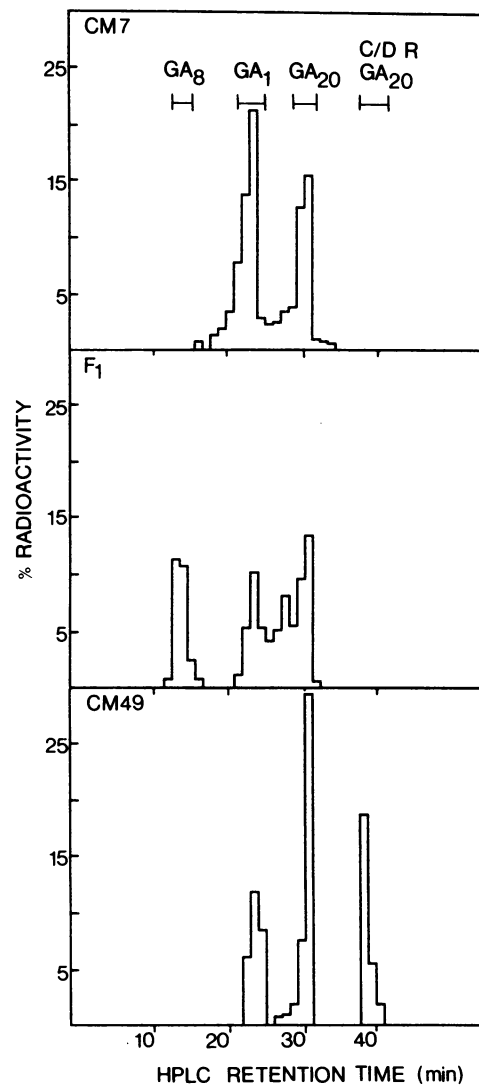


FIG. 7. Elution profile of radioactivity  $^3\text{H}$  from grouped fractions I + II (Fig. 6) chromatographed on gradient-eluted reverse-phase C<sub>18</sub> HPLC columns. Retention times of authentic standards are shown in top Figure. These profiles provide qualitative information only as transfers of SiO<sub>2</sub> fractions was not quantitative.

in heterosis in maize. Assessed in view of additional information (presented below), an integrated analysis of all information suggests that a causal association may exist between GA and hybrid vigor.

Five single gene mutants of maize have been identified which are deficient in, or contain reduced levels of GA-like substances (17). These mutants are phenotypically dwarfed and produce low grain yields. The dwarf phenotype can be totally overcome through exogenous application of GA<sub>3</sub> (17), and grain production of these mutants is also increased by GA<sub>3</sub>, suggesting that a GA deficiency not only limits growth but also reduces productivity.

The most conspicuous response to exogenous application of GA<sub>3</sub> by normal as well as dwarf maize genotypes is an increase in internode elongation, and consequently an increase in plant height. Application of GA<sub>3</sub> increases whole seedling dry weight (7), and increases in fodder and grain yields have also been reported following GA<sub>3</sub> application (4). Thus, exogenous application of GA<sub>3</sub> promotes growth and yield of normal maize genotypes.

In normal genotypes, content of endogenous GA-like substances is correlated with growth rate (18, 20) and a low growing temper-

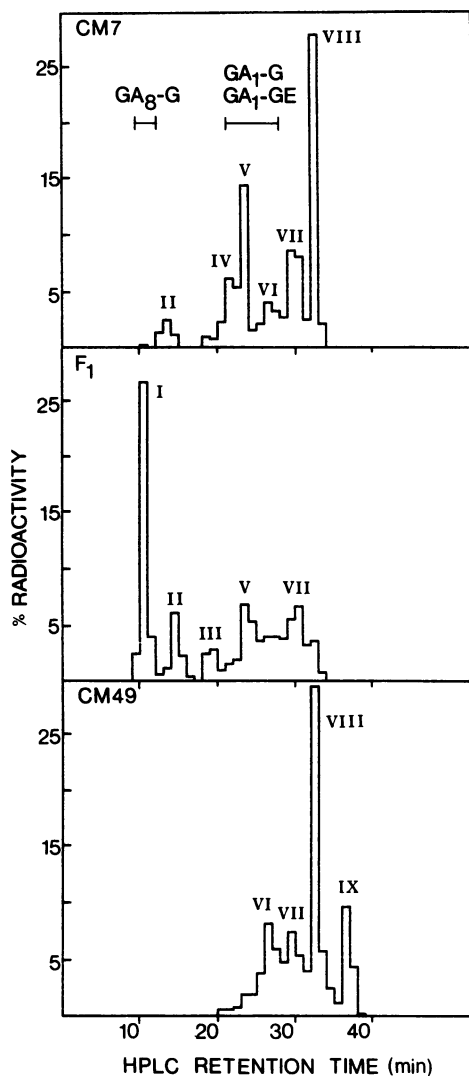


FIG. 8. Elution profiles of radioactivity  $^3\text{H}$  from grouped MeOH wash (Fig. 6) chromatographed on gradient-eluted reverse-phase  $\text{C}_{18}$  HPLC columns. Retention times of authentic GA glucosyl conjugates are shown in top figure, and glucosyl conjugates of  $\text{GA}_{20}$  and C/D R  $\text{GA}_{20}$  would be expected to elute coincidental with or just prior to the acidic GA moiety (Fig. 7). Peaks sharing the same roman numerals had similar retention times.

ature reduces growth rate as well as levels of endogenous GA-like substances (18). The three genotypes described in the present study have now been grown under three temperature regimes, and growth rates and levels of GA-like substances were increased as growing temperature was progressively raised (18, 21). Thus, rapid growth is correlated with high levels of endogenous GA-like substances. Under favorable (warm) temperature conditions, hybrid vigor for a number of growth parameters is observed in these three genotypes (21), and endogenous GA-like substances were higher in the hybrid than in either parental inbred (21). Thus, content of GA-like substances was correlated with heterosis for superior growth.

Nickerson (13, 14) previously reported that maize inbreds are more responsive to exogenous  $\text{GA}_3$  application than hybrids. Indeed, Nickerson further suggested that sensitivity to  $\text{GA}_3$  was correlated with the degree of inbreeding in maize (13, 14). The results of the present study (21) support Nickerson's conclusion, show that such a relationship holds for genetically related genotypes, and are consistent with the observation that inbreds contain

lower levels of endogenous GA-like substances. Hence, inbreds should be more responsive to GA application, and thus their low endogenous level of GA-like substances may be a major factor limiting growth. It is possible that the observed 'overdominance' for increased levels of GA-like substances (21) may underlie at least certain aspects of hybrid vigor in maize.

Other research also suggests that GA play a role in heterosis in maize. Parental pollen tube growth rates of germinating maize pollen are positively correlated with seedling dry weight, ear weight, and grain yield of the resultant hybrids (15). Pollen tube growth is promoted through exogenous application of GA (3, 11), and increases in endogenous GA of untreated pollen is positively correlated with increased pollen germination and rapid pollen tube growth (1, 10). Thus, the correlation between rapidity of pollen tube growth and hybrid vigor of the progeny (15) also appears interrelated with GA physiology.

In an early review, Paleg (16) concluded that GA play a role in endosperm mobilization and that this mobilization is probably correlated with the expression of heterosis. Although hybrid vigor cannot be attributed to an initial advantage of larger hybrid embryos (20), there is evidence (23) suggesting that early maize seedling performance is positively correlated with subsequent growth rate, and we noted (Fig. 1) that hybrid vigor is also displayed for early growth in maize.

Thus, maize inbreds, relative to a hybrid, are deficient in GA-like substances (21). These inbreds are also more responsive to  $\text{GA}_3$  application than their hybrid. These results, together with the other findings (1, 3, 4, 7, 10, 11, 13-18, 21, 23) noted above, lead us to conclude that (a) low endogenous GA level is one of the more important factors limiting growth and yield of maize inbreds, and (b) the high endogenous GA level in the hybrid may provide a phytohormonal basis for at least certain aspects of heterosis in maize.

It must be noted, however, that it is unlikely that a single mechanism is responsible for all aspects of hybrid vigor. It has been suggested that enzymic polymorphism may offer a general mechanism for heterosis (2, 22), and that enzymes from a number of metabolic pathways are probably involved. The enzymic polymorphism hypothesis suggests that the presence of different isozymes in the heterozygote may broaden an individual's biosynthetic potential, thus leading to superior performance. An increase in biosynthesis of specific regulatory compounds such as GA could further amplify the benefits of enzymic polymorphism, the increased GA promoting many aspects of maize growth and development which produce the heterotic phenotype.

#### LITERATURE CITED

- BARENDSE GWM, AS RODRIGUES PEREIRA, PA BERKERS, FM DREISSEN, A VAN EYDEN-EMONS, HF LINSKENS 1970 Growth hormones in pollen, styles and ovaries of *Petunia hybrida* and *Lolium* species. *Acta Bot Neerl* 19: 175-186
- BERGER E 1975 Heterosis and the maintenance of enzyme polymorphism. *Am Nat* 110: 823-829
- BOSE N 1959 Effect of gibberellin on the growth of pollen tubes. *Nature* 184: 1577
- CHERRY J, HA LUND, EB EARLEY 1960 Effect of gibberellic acid on growth and yield of corn. *Agron J* 52: 167-170
- DAVIES LJ, L RAPPAPORT 1975 Metabolism of tritiated gibberellins in *d-5* dwarf maize. II. [ $^3\text{H}$ ]Gibberellin A<sub>1</sub>, [ $^3\text{H}$ ]gibberellin A<sub>2</sub>, and related compounds. *Plant Physiol* 56: 60-66
- DURLEY RC, A CROZIER, RP PHARIS, GE McLAUGHLIN 1972 Chromatography of 33 gibberellins on a gradient eluted silica gel partition column. *Phytochemistry* 11: 3029-3033
- GUBBELLS GH 1976 Emergence, seedling vigor, and seed-yield of corn after pre-sowing treatments and the addition of phosphorous with the seed. *Can J Plant Sci* 56: 749-751
- HEDDEN P, BO PHINNEY, R HEUPEL, D FUJII, H COHEN, P GASKIN, J MACMILLAN, JE GRAEBE 1982 Hormones of young tassels of *Zea mays*. *Phytochemistry* 21: 390-393
- HEUPEL R, BO PHINNEY, P HEDDEN 1981 Gibberellin metabolism in *Zea mays* L. *Plant Physiol* 67: S-102
- KAMIENSKA A, RP PHARIS 1975 Endogenous gibberellins of pine pollen. II. Changes during germination of *Pinus attenuata*, *P. coulteri*, and *P. ponderosa*

- pollen. *Plant Physiol* 56: 655-659
11. KATO Y 1955 Responses of plant cells to gibberellin. *Bot Gaz* 117: 16-24
  12. KOSHIOKA M, J HARADA, K TAKENO, M NOMA, T SASSA, K OGIYAMA, JS TAYLOR, SB ROOD, RL LEGGE, RP PHARIS 1982 Reversed-phase  $C_{18}$  high-performance liquid chromatography of acidic and conjugated gibberellins. *J Chromatogr*. In press
  13. NICKERSON NH 1959 Sustained treatment with gibberellic acid of five different kinds of maize. *Ann Mo Bot Garden* 46: 19-37
  14. NICKERSON NH, TN EMBLER 1960 Studies involving sustained treatment of maize with gibberellic acid; further notes on responses of races. *Ann Mo Bot Garden* 47: 227-242
  15. OTTAVIANO E, M SARI-GORLA, DL MULCAHY 1980 Pollen tube growth rate in *Zea mays*: implications for genetic improvement of crops. *Science* 210: 437-438
  16. PALEG LG 1965 Physiological effects of gibberellins. *Annu Rev Plant Physiol* 16: 291-322
  17. PHINNEY BO 1961 Dwarfing genes in *Zea mays* and their relation to the gibberellins. In RM Klein, ed, *Plant Growth Regulation*. Iowa State University Press, Ames, pp 489-501
  18. ROOD SB 1981 Genetic, environmental and hormonal control of maize development. PhD thesis. University of Calgary, Calgary
  19. ROOD SB, DJ MAJOR 1981 Diallel analysis of leaf number, leaf development rate, and plant height of early maturing maize. *Crop Sci* 21: 867-873
  20. ROOD SB, KOSHIOKA M, TJ DOUGLAS, RP PHARIS 1982 Metabolism of tritiated gibberellin  $A_{20}$  in maize. *Plant Physiol* 70: 1614-1618
  21. ROOD SB, RP PHARIS, M KOSHIOKA, DJ MAJOR 1983 Gibberellins and heterosis in maize. I. Endogenous GA-like substances. *Plant Physiol* 71: 639-644
  22. SINHA SK, R KHANNA 1975 Physiological, biochemical and genetic basis of heterosis. In NC Brady, ed, *Adv Agron* 27: 123-174
  23. WOODSTOCK LW 1965 Initial respiration rates and subsequent growth in germinating corn seedlings. *BioScience* 15: 783-784