

The role of miRNA-144-3p/Oprk1/KOR in nicotine dependence and nicotine withdrawal in male rats

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Abstract

Introduction: The kappa-opioid receptor (KOR) has been implicated in mediating the behavioral and biochemical effects associated with nicotine reward and withdrawal; however, its underlying mechanisms remain to be further explored.

Methods: Adult male Sprague-Dawley rats were used to establish a nicotine dependence and withdrawal model by injecting nicotine (3 mg/kg/ day, s.c.) or vehicle for 14 days, followed by the termination of nicotine for 7 days. Body weight gain, pain behaviors, and withdrawal scores were assessed in succession. MicroRNA (miRNA) sequencing was performed, and quantitative real-time PCR was used to detect the expression of candidate miRNAs and Oprk1. Western blotting was performed to examine KOR protein expression of KOR. Luciferase assay was conducted to validate the relationship of certain miRNAs/Oprk1.

Results: The behavioral results showed that nicotine dependence and withdrawal induced behavioral changes. Biochemical analyses demonstrated that miR-144-3p expression decreased and Oprk1/KOR expression increased in the prefrontal cortex, nucleus accumben, and hippocampus. Further investigation suggested that miR-144-3p exerted an inhibitory effect on Oprk1 expression in PC12 cells.

Conclusions: This study revealed that miR-144-3p/Oprk1/KOR might be a potential pathway underlying the adverse effects induced by nicotine dependence and withdrawal, and might provide a novel therapeutic target for smoking cessation.

Implications: This study demonstrates an impact of nicotine dependence and nicotine withdrawal on behavioral outcomes and the expressions of miR-144-3p/Oprk1/KOR in male rats. These findings have important translational implications given the continued use of nicotine and the difficulty in smoking cessation worldwide, which can be applied to alleviated the adverse effects induced by nicotine dependence and withdrawal, thus assist smokers to quit smoking.

Introduction

Cigarette smoking is a primary preventable cause of morbidity and mortality worldwide.¹⁻³ Smoking cessation causes adverse withdrawal symptoms, which in turn make it challenging to maintain abstinence and contributes to relapse.⁴⁻⁶

Nicotine withdrawal symptoms are characterized by somatic symptoms (eg, gastrointestinal discomfort, increased appetite, and tremors), affective symptoms (such as depression, anxiety, hyperalgesia, and irritability), attention deficits, and cognitive deficits.^{7,8} Generally, nicotine withdrawal symptoms peak during the first week of smoking abstinence and may last for up to 4 weeks. Current smoking cessation methods are inadequate, and novel therapeutic targets need to be explored.^{9,10}

The dynorphin/kappa-opioid receptor (KOR) is one of the endogenous opioid receptor systems, which has been implicated in the biological processes of nicotine.¹¹⁻¹³ Ugur et al. observed that chronic nicotine treatment modulated DOR and KOR gene expression in the mesocorticolimbic system.¹⁴ Ward et al. reported that KOR agonist \pm U50,488 H increased the aversive effects of nicotine, whereas the KOR antagonist nor-BNI attenuated them.¹⁵ Other studies have also suggested that the KOR antagonist is a potential therapeutic agent for managing withdrawal syndromes in smoking cessation.¹⁶⁻¹⁸ In addition, KOR was found to affect the energy balance. Seoane-Collazo et al. reported that nicotine treatment decreased body weight in wild-type mice but not in KOR-null mice. Further investigation revealed that the effect of nicotine on weight was dependent on KOR, specifically in the lateral hypothalamic area.¹⁹ Hence, KOR likely plays an important role in nicotine reward and withdrawal in several ways and could be a promising target for smoking cessation.

MicroRNAs (miRNAs) are endogenous short noncoding RNAs (~22 nucleotides long) that usually direct posttranscriptional suppression through translational inhibition or mRNA destabilization by pairing to the 3' untranslated (UTR) region of mRNAs.²⁰ miRNAs are involved in the biological functions of nicotine and are potential biomarkers or therapeutic targets.²¹⁻²⁴

We hypothesized that the miRNA-mediated regulations of Oprk1/KOR in different brain regions are one of the mechanisms underlying nicotine administration and withdrawal symptoms. Therefore, we identified and compared miRNAs from different brain regions in nicotine-administered

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and nicotine-withdrawn rats with those in non-nicotineadministered rats. The expression of the candidate differentially expressed miRNAs (DEmiRNAs) and Oprk1/KOR was examined both in vivo and in vitro. Finally, the relationship between specific DEmiRNAs and Oprk1/KOR was validated using the luciferase reporter assay.

Materials and Methods

Animals and Grouping

A total of 36 male Sprague-Dawley rats (8 weeks old) were acquired from Hunan SJA Laboratory Animal Company (Changsha, Hunan, China). All rats had free access to food and water and were housed under a 12/12 dark/light cycle at $22 \pm 1^{\circ}$ C. After 7 days of adaptation, the rats were randomly divided into two groups (n = 18): Group NS and Group NI. The animal study protocol was approved by the Animal Care and Use Committee of Hunan Cancer Hospital (Permit Number: 2020-11).

Establishment of Nicotine Dependence Model

(-)-Nicotine hydrogen tartrate salt [(-)-1-methyl-2-(3-pyridyl) pyrrolidine (+)-bitartrate salt] was purchased from Sigma-Aldrich Inc. (St. Louis, MO, USA). Nicotine was dissolved in 0.9% sodium chloride. In Group NI, the rats were subcutaneously injected with nicotine (3 mg/kg/day) twice daily at 08:00–09:00 AM and 16:00–17:00 PM for 14 days. Rats in Group NS were injected with an equal volume of 0.9% saline following the same schedule.

Spontaneous Nicotine Withdrawal

After 14 days of consecutive injections, nicotine or saline injections were ceased. On day 1 after nicotine withdrawal, a 15-min observation of somatic signs was observed, including paw and body tremors, head shakes, backing, jumps, curls, and ptosis.^{7,25} The total number of somatic signs was tallied for each rat, and group averages were plotted. Hyperalgesia was evaluated using the von-Frey fibers and hot plate test immediately following the somatic sign observation period as previously described.^{26,27} All experiments were performed by an observer blinded to the experimental treatments.

Pain Behavioral Assessments

To assess the effects of nicotine dependence and withdrawal on pain perception, we tested mechanical withdrawal threshold (MWT) and thermal withdrawal latency (TWL) to monitor the changes in pain. Based on previous reports,²⁸ von-Frey filaments (Touch-Test Sensory Evaluator, Wood Dale, IL, USA) were used for the MWT test. Before the experiment, the rats were allowed to adapt to the test environment for 15–30 min. Each rat was tested at least five times at an interval of 15 s. The 50% threshold for further analysis was calculated using the up- and down-method.

The TWL was measured using a Hargreaves apparatus (Ugo Basile, Italy). During the test, each rat was tested at least three times at an interval of 5 min. The time from the onset of radiant heat stimulation to the withdrawal of the hind paw was recorded.

RNA Isolation, Sequencing, and Target Gene Prediction

On day 14, 24 h after the last injection, the rats were decapitated under deep anesthesia with 4% (v/v) isoflurane

inhalation for 3 min. Hippocampi were quickly dissected, and collected on ice. Total RNA was extracted from the hippocampi using TRIzol Reagent (Invitrogen, Carlsbad, CA, USA), and the concentration and purity of the RNA samples were validated using a NanoDrop spectrophotometer. Next, small RNA sequencing was conducted to detect the DEmiRNAs. A cut-off value of 1.5-fold was used to determine the differences in miRNA levels. Finally, a list of target transcripts for miR-144-3p (256 transcripts) was obtained from miRDB.

Western Blot Analyses

Total protein was extracted from the prefrontal cortex (PFC), nucleus accumben (NAc), and hippocampus (Hippo) using radioimmunoprecipitation assay (RIPA) lysis buffer, and protein concentration was determined by BCA Assay kit. Then the samples were loaded onto a 10% SDS-PAGE gel. The primary antibodies used were against KOR (Abcam, Cambridge, MA, USA) and GAPDH (Proteintech, China). Protein bands were visualized using enzyme-catalyzed chemiluminescence (ECL). Images were analyzed by Image J v1.8.0 software (Bio-Rad, Hercules, CA, USA).

Quantitative Real-Time PCR

Quantitative real-time PCR (RT-qPCR) analyses were performed to confirm the expressions of miR-144-3p and Oprk1 in the PFC, NAc, Hippo, and PC12 cells. U6 and actin serve as endogenous reference genes (controls). The primers used were as follows: U6 (F: 5'-GCTTCGGCAGCACATATAC TAAAAT-3', R: 5'-CGCTTCACGAATTTGCGTGTCAT-3'), 5'-ACATCCGTAAAGACCTCTATGCC-3', actin (F: R: 5'-TACTCCTGCTTGCT GATCCAC-3'), miR-144-3p 5'-GGGGGGGTACAGTATAGATGA-3', (GSP: 5'-CAGTGCGTGTCGTGGA-3'), and R: Oprk1(F: 5'-ACTTCTGCATTGCCTTGGGTT-3', R٠ 5'-AAGCAGAAGTC CCTAA AACACCG-3').

Vector Construction, Mutagenesis, and Luciferase Assay

To elucidate the regulatory relationship between Oprk1 and rno-miR-144-3p, the 3' UTR sequences of Oprk1 containing binding sites for rno-miR-144-3p were cloned into pcDNA luciferase vectors (Promega, Madison, WI, USA) to produce vectors for wide-type Oprk1 3' UTRs. Then, site-directed mutagenesis was carried out using a Quick Change II site-directed mutagenesis assay kit (Stratagene, San Diego, CA) to generate mutant 3' UTR sequences of Oprk1 harboring the mutated binding sites for rno-miR-144-3p. Furthermore, the mutant 3' UTR sequences were cloned into pcDNA luciferase vectors to produce vectors for mutant Oprk1 3' UTRs. In the next step, PC12 cells (Zhong Qiao Xin Zhou Biotechnology Company, Shanghai, China) were cotransfected with rno-miR-144-3p in conjunction with wild-type or mutant 3' UTR of Oprk1 using Lipofectamine 3000. At 48 h post-transfection, the luciferase activity of the transfected cells was assayed using a Bright Glo Luciferase kit (Promega, Madison, WI, USA) following the manufacturer's instructions.

Statistical Analysis

All data were expressed as mean \pm SD and analyzed using GraphPad Prism 3.0. Image quantification of western blotting bands was presented as values relative to the control. Differences between the groups were assessed using two-way

or one-way analysis of variance (ANOVA), followed by Bonferroni or Dunnett's post hoc tests. Statistical significance was set at p < .05.

Results

Nicotine Dependence and Withdrawal Caused Changes in MWT, TWL, Body Weight Gain, and Withdrawal Scores

To explore the effects of nicotine dependence and withdrawal on the behaviors of rats, we established a rat model by injecting 3 mg/kg/day nicotine s.c. for 14 days followed by the spontaneous withdrawal for 7 days. As shown in Figure 1A and B, the 50% MWT and TWL of rats in Group NI (nicotine administration) were significantly higher than those in Group NS (saline administration) at 1 and 2 weeks postinjection. The 50% MWT and TWL in Group NI declined and were lower than those of Group NS on day 1 of withdrawal. The hyperalgesic state lasted for several days before returning to normal levels. Regarding body weight gain, the rats in Group NI gained less weight than those in Group NS after nicotine administration, whereas after nicotine withdrawal, the weight gain of rats in Group NI (nicotine administration) was higher than those of Group NS (saline administration) (Figure 1C). As shown in Figure 1D, compared with Group NS, the withdrawal scores were significantly higher in Group NI. These tests indicated that 3 mg/kg/day nicotine successfully established a rat nicotine dependence and withdrawal model. Nicotine administration resulted in hypoalgesia and decreased body weight gain, while nicotine withdrawal caused hyperalgesia, increased body weight gain, and higher withdrawal scores.

Expression Profile, Validation, and Target Genes Prediction of miRNAs in the Hippocampi of Nicotine Dependence Rats

To determine the effect of nicotine administration on the expression of miRNAs, we tested the expression profile of miRNAs in the hippocampi of rats using microarray. According to the criteria of a fold change > 1.5 and a *p*-value < .05, a total of 14 DEmiRNAs were identified, including 9 upregulated and 5 downregulated miRNAs (Figure 2A), and the hierarchical clustering heat map for the 14 DEmiRNAs was displayed in Figure 2B. The prediction of target genes suggested that Oprk1 was likely one of the downstream genes of miR-144-3p (Figure 2D). Previous reports have indicated that miR-144-3p as the candidate for further study.

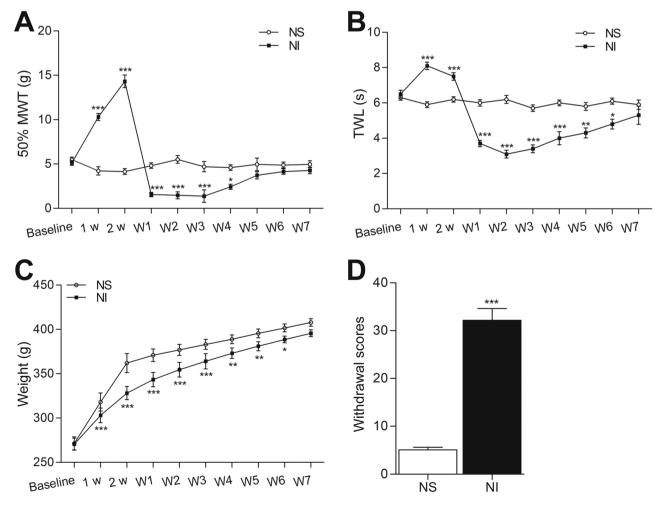


Figure 1. Nicotine dependence and withdrawal cause changes in mechanical withdrawal threshold (MWT), thermal withdrawal latency (TWL), weight gain, and the withdrawal scores in rats. (A) Changes in MWT, (B) changes in TWL, (C) changes in weight gains, and (D) changes in withdrawal scores. Each point represents the mean \pm SD (n = 6-12/group). *p < .05, **p < .005, **p < .001 versus Group NS.

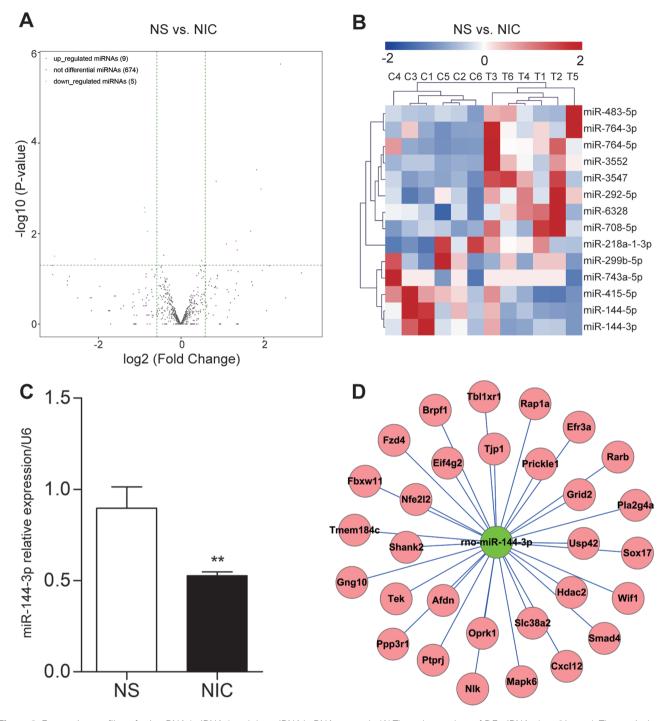


Figure 2. Expression profiles of microRNA (miRNAs) and the miRNA/mRNA network. (A) The volcano plots of DEmiRNAs (n = 6/group). The vertical dotted lines manifested 1.5-fold changes (log2 scaled) up and down, respectively, and the horizontal dotted line showed a p-value of .05 (–log10 scaled). The red and green points denoted the significantly up- and downregulated miRNAs. Gray dots indicated nondifferentially expressed miRNAs. (B) Hierarchical clustering heat map for the 14 DEmiRNAs. The up- and downregulated miRNAs were colored in red and blue, respectively. (C) RT-qPCR verification of miR-144-3p. Compared with Group NS (day 14 after saline administration), miR-144-3p was significantly downregulated in Group NIC (day 14 after nicotine administration). The data were presented as the mean \pm SD. **p < .005. (D) The network of miR-144-3p and its putative mRNAs. Nodes in red color were the putative mRNAs, and nodes in green color were the downregulated miR-144-3p.

The RT-qPCR results revealed that miR-144-3p was downregulated in Group NIC (day 14 after nicotine administration), compared with Group NS (day 14 after saline administration) (n = 6 in each group, p < .005) (Figure 2C), which was in line with the sequencing data. In summary, the expression of miRNAs was altered in the hippocampi of nicotine-dependent rats, specifically, miR-144-3p was downregulated.

Nicotine Dependence and Withdrawal Downregulated miR-144-3p and Upregulated Oprk1 in Rats

Previous studies have implied the role of KOR in nicotine dependence and withdrawal.²⁹ Based on these results, we hypothesized that miR-144-3p participated in the biological processes of nicotine dependence and withdrawal by regulating the expression of Oprk1 (the gene encoding KOR). Moreover, recent studies have correlated brain regions (such as PFC, NAc, Hippo) with nicotine use.^{3,30} Hence, we detected the expressions of miR-144-3p, Oprk1, and KOR in PFC, NAc, and Hippo on day 14 after nicotine administration (NIC) and day 1 after withdrawal (NW). The results suggested that miR-144-3p and Oprk1 exhibited opposite trends after nicotine administration and withdrawal, such that miR-144-3p decreased and Oprk1 increased at the RNA level (Figure 3A–C). Western blotting indicated that KOR was highly expressed in Group NIC and NW (Figure 3D–F). These data demonstrate that nicotine administration and withdrawal influence the expression of miR-144-3p and Oprk1/KOR in multiple brain regions.

Nicotine Exposure Decreased miR-144-3p and Increased Oprk1 in PC12 Cells

To explore the effects of nicotine on the expression of miR-144-3p and Oprk1, we exposed PC12 cells to $10 \ \mu$ M nicotine for 6 h and observed that nicotine exposure could reduce the expression of miR-144-3p and increase the expression of Oprk1 (Figure 4A and C). Transfection of miR-144-3p mimic into PC12 cells could reduce the expression of Oprk1/KOR (Figure 4B and D); however, when we treated the transfected cells with 10 μ M nicotine for 6 h, we observed that nicotine exposure could neutralize the effect of miR-144-3p mimic on the expression of Oprk1/KOR (Figure 4E). Similar to the in vivo studies, in vitro, nicotine exposure could modulate the expression of miR-144-3p and Oprk1/KOR.

miR-144-3p Inhibited the Luciferase Activity of Wild-Type Oprk1 Through Binding to its 3' UTR

To further elucidate the relationship between miR-144-3p and Oprk1, luciferase vectors containing wild-type and mutant Oprk1 3' UTRs were established and cotransfected with miR-144-3p into PC12 cells. miR-144-3p suppressed the luciferase activity of wild-type Oprk1 but had no inhibitory effect on

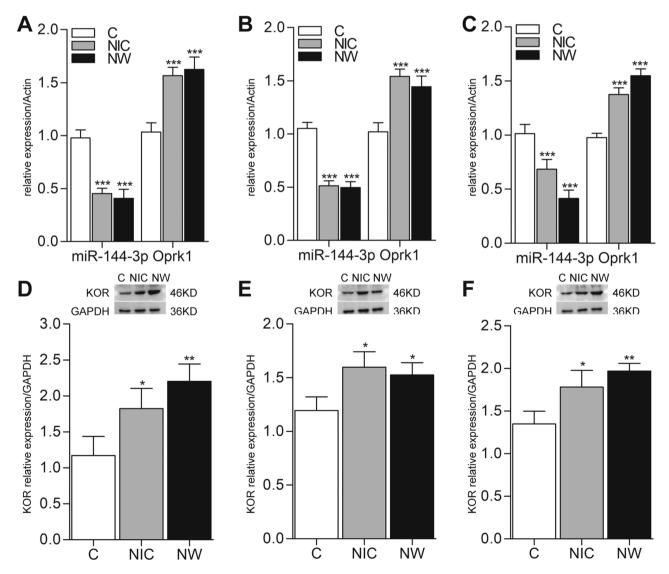


Figure 3. Nicotine dependence and withdrawal change the expressions of miR-144-3p and kappa-opioid receptor (KOR) in rats. (A–C) shown the expressions of miR-144-3p and Oprk1 (the gene of KOR) at RNA level in PFC, hippocampus, and NAc. (D–F) The expressions of KOR at protein level in PFC, hippocampus, and NAc. For RT-qPCR, actin was used as the housekeeping gene, and the relative expression of rno-miR-144-3p, Oprk1 were quantified using the $2^{-\Delta\Delta CT}$ method. For western blot, GAPDH was used as a loading control. Data were expressed as normalized ratio of protein band density of KOR against GAPDH and presented as the mean \pm SD (n = 6). Group NIC represents day 14 after nicotine administration; Group NW represents day 1 after withdrawal. *p < .05, **p < .005, **p < .001 versus Group C.

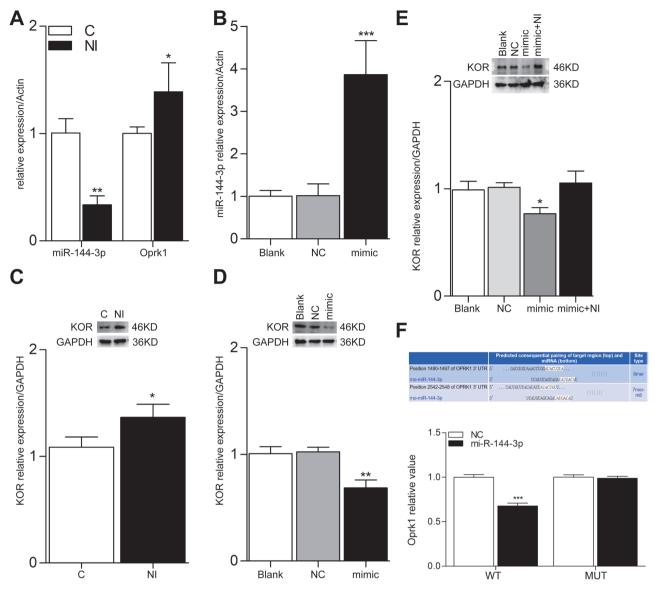


Figure 4. Nicotine exposure regulates the expressions of miR-144-3p and KOR in PC12 cells. (A) and (B) The RT-qPCR results of nicotine exposure and the transfection of miR-144-3p mimic, respectively. Actin was used as the housekeeping gene. (C) and (D) The protein bands and its quantification analysis of western blot of nicotine exposure and the transfection of miR-144-3p mimic, respectively. GAPDH was used as a loading control. (E) The protein bands and its quantification analysis of western blot of the transfection of miR-144-3p mimic combined with exposure to nicotine. (F) The luciferase reporter. Rno-miR-144-3p inhibited the luciferase activity of wild-type Oprk1 through binding to its 3' UTR. Data were presented as the mean \pm SD. *p < .05, **p < .005, **p < .001.

mutant Oprk1(Figure 4F). These results indicate that miR-144-3p directly targets Oprk1.

Discussion

In this study, we explored the role of miR-144-3p/Oprk1/ KOR in the adverse behaviors induced by nicotine administration and withdrawal for the first time. The results revealed that nicotine administration and withdrawal altered the expressions of miRNA-144-3p, Oprk1, and KOR in multiple brain regions, indicating that miRNA-144-3p negatively modulates the expression of Oprk1, which might be the potential pathway underlying nicotine-induced adverse effects.

Nicotine use and withdrawal can cause neuroadaptive changes in the brain and, in turn, alter the behaviors. In rats, repeated subcutaneous nicotine injection can induce dependence, and termination of nicotine injections can lead to spontaneous withdrawal symptoms such as hyperalgesia and somatic withdrawal signs.³¹ Body weight gain in rats can also be affected. Specifically, nicotine exposure reduced body weight gain, whereas increased the body weight gain for a short time after cessation of nicotine injections and finally returned to baseline levels. Based on previous reports,7,32,33 we successfully established a nicotine dependence and spontaneous withdrawal model in rats via subcutaneous injection of 3 mg/kg/day nicotine for 14 days, followed by spontaneous termination of injections for 7 days. Behavioral tests revealed that the MTW and TWL increased during nicotine administration, but decreased during nicotine withdrawal in rats. The rats in the Group NI gained less weight than in Group NS, whereas those in the Group NI gained more weight than in the NS group after spontaneous withdrawal. In addition, compared to the rats in Group NS, rats in Group NI exhibited higher withdrawal scores. All of the above results are consistent with those of previous studies.

Alterations in gene expressions are one of the underlying mechanisms of nicotine-associated behavioral changes. Recent research has demonstrated that smoking or nicotine consumption can alter the expression of miRNAs in various cell lines, organisms, and tissues and that miRNAs regulate addiction-related neuroplasticity in the brain.^{34–37} Kameshwar et al. reported that exosomal miRNAs were differentially expressed in cigarette smokers and might be the novel circulating biomarkers for smokers.²² Rauthan et al. reported that miR-238 mediated nicotine withdrawal response in Caenorhabditis elegans following chronic nicotine exposure by suppressing nAChR ACR-19 expression.³⁸ This alteration in miRNA has also been reported in other studies.^{39,40} Besides, the hippocampus, PFC, and NAc are well-studied areas in nicotine research.³ Therefore, in this study, we detected 14 DEmiRNAs, including 9 upregulated and 5 downregulated miRNAs, in the hippocampi of nicotine-dependent rats. The results of previous studies suggested that the downregulated DEmiRNA miRNA-144-3p was involved in many biological processes, such as cell proliferation, carbohydrate metabolism, and the pathophysiology of depression.⁴¹⁻⁴³ Predictions of target genes of miRNA-144-3p indicated Oprk1 as one of its target genes. Hence, we considered miRNA-144-3p as the most promising candidate. We validated the expression of miRNA-144-3p in several crucial brain regions (Hippo, PFC, and NAc) and observed that the expression of miRNA-144-3p was downregulated in these regions.

Studies have indicated that opioid receptors are involved in the development of nicotine dependence and abstinenceinduced withdrawal syndromes.12,13,44 However, recent studies have mainly focused on µ-opioid receptor (MOR) and δ opioid receptor (DOR). Studies on the involvement of KOR are limited. Banks et al. outlined that KOR was altered during drug abuse, and functioned as a promising therapeutic target.²⁹ In the present study, we found that both nicotine administration and withdrawal upregulated the expression of Oprk1/KOR in the Hippo, PFC, and NAc. This trend was confirmed in an in vitro study. The expression of miRNA-144-3p was downregulated in the same brain tissues and the cell model. Further bioinformatic analysis indicated that Oprk1 was likely a putative downstream gene of miRNA-144-3p. Hence, we hypothesized that miRNA-144-3p negatively regulated KOR expression. To date, several KOR-selective antagonists, including nor-BNI, JDTic, and LY2456302, as well as their agonists (such as U50,488 and ICI 204,448) have been shown to modulate nicotine administration and nicotine withdrawal-induced adverse effects.^{17,18,45} The efficacy of KOR antagonists and agonists in human trials has been unsatisfactory. Therefore, it is necessary to explore novel molecules that target KOR. Our investigation identified miRNA-144-3p as the most suitable potential target.

To further clarify the relationship between miR-144-3p and Oprk1, we performed in vitro experiments in PC12 cells. The results revealed that nicotine exposure reduced the expression of miR-144-3p and increased that of Oprk1/KOR, similar to in vivo results. Overexpression of miR-144-3p by the transfection with a miR-144-3p mimic markedly decreased Oprk1/KOR expression in PC12 cells. Further investigation revealed that nicotine exposure neutralized the effect of the miR-144-3p mimic on the expression of Oprk1/KOR.

Through in silico analyses, we observed that Oprk1 3' UTR region contains a matching site for miR-144-3p. Finally, dualluciferase activity assay validated that miR-144-3p directly targeted Oprk1, which indicated that miR-144-3p probably have a negative regulatory effect on the expression of Oprk1/ KOR. Further investigation is need in vivo.

This study has several limitations. First, although we tested the relationship between miR-144-3p/Oprk1/KOR in PC12 cells, we did not validate it in a rat model. Second, the behavioral assessments of nicotine-induced withdrawal symptoms were inadequate, we did not observe affective symptoms (such as depression and anxiety), attention deficits, and cognitive deficits. Third, only male rats, one drug, and one dose were included in this study. In addition, the expressions of KOR and miR-144-3p should be detected dynamically over time. In future studies, we will try to construct the lentivirus vector, or the agomir and antagomir for miR-144-3p to perform in vivo study and further verify this specific modulatory mechanism. Meanwhile, we will explore the effects of different concentrations of nicotine on behavioral and molecular changes, repeat this experimental protocol in female rats, and detect these molecules at multiple time points, to better elucidate the specific modulatory mechanisms.

Conclusions

KOR has been shown to be involved in nicotine-induced adverse effects, and further investigation indicated that KOR agonists aggravated the adverse effects caused by nicotine administration and withdrawal, and KOR antagonists alleviated these adverse effects in animal models. However, these effects were not observed in smokers, and KOR antagonists failed to relieve withdrawal symptoms. We found that in several brain regions (PFC, NAc, and Hippo) of rats, the adverse effects induced by nicotine dependence and withdrawal were associated with the downregulated expression of miR-144-3p and the upregulated expression of Oprk1/KOR. It is crucial to clarify the specific regulatory effects on adverse reactions and verify their roles in future human trials. In summary, our findings indicated that miR-144-3p/Oprk1/KOR is a potential pathway underlying nicotine-induced adverse effects, thereby providing a novel target for smoking cessation.

Supplementary Material

A Contributorship Form detailing each author's specific involvement with this content, as well as any supplementary data, are available online at https://academic.oup.com/ntr.

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Declaration of Interests

All authors declare that they have no competing interests.

Author Contributions

Jiali Shao (Formal analysis [Equal], Investigation [Equal], Methodology [Equal], Software [Equal], Visualization [Equal], Writing – original draft [Equal]), Yanxia Fei (Data curation [Equal], Formal analysis [Equal], Investigation [Equal], Validation [Equal]), Ji Xiao (Data curation [Equal], Methodology [Equal], Validation [Equal], Writing – review & editing [Equal]), Lijuan Wang (Investigation [Equal], Software [Equal], Validation [Equal], Visualization [Equal]), Shuangfa Zou (Conceptualization [Equal], Data curation [Equal], Methodology [Equal], Project administration [Equal], Writing – review & editing [Equal]), and Jinfeng Yang (Conceptualization [Equal], Funding acquisition [Equal], Methodology [Equal], Funding acquisition [Equal], Methodology [Equal], Project administration [Equal], Methodology [Equal], Project administration [Equal], Resources [Equal], Supervision [Equal], Writing – review & editing [Equal], Writing – review & editing [Equal], Writing – review & editing [Equal], Nethodology [Equal], Project administration [Equal], Resources [Equal], Supervision [Equal], Writing – review & editing [Equal])

Data Availability Statement

The data that support the findings of this study will be openly available at OSF (https://osf.io/ ek6wx), upon manuscript acceptance.

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