



Review

Novel Role of Molecular Hydrogen: The End of Ophthalmic Diseases?

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Abstract: Molecular hydrogen (H_2) is a colorless, odorless, and tasteless gas which displays non-toxic features at high concentrations. H_2 can alleviate oxidative damage, reduce inflammatory reactions and inhibit apoptosis cascades, thereby inducing protective and repairing effects on cells. H_2 can be transported into the body in the form of H_2 gas, hydrogen-rich water (HRW), hydrogen-rich saline (HRS) or H_2 produced by intestinal bacteria. Accumulating evidence suggest that H_2 is protective against multiple ophthalmic diseases, including cataracts, dry eye disease, diabetic retinopathy (DR) and other fields. In particular, H_2 has been tested in the treatment of dry eye disease and corneal endothelial injury in clinical practice. This medical gas has brought hope to patients suffering from blindness. Although H_2 has demonstrated promising therapeutic potentials and broad application prospects, further large-scale studies involving more patients are still needed to determine its optimal application mode and dosage. In this paper, we have reviewed the basic characteristics of H_2 , and its therapeutic effects in ophthalmic diseases. We also focus on the latest progress in the administration approaches and mechanisms underlying these benefits.

Keywords: molecular hydrogen; ophthalmic diseases; oxidation stress



Citation: Li, S.-Y.; Xue, R.-Y.; Wu, H.; Pu, N.; Wei, D.; Zhao, N.; Song, Z.-M.; Tao, Y. Novel Role of Molecular Hydrogen: The End of Ophthalmic Diseases? *Pharmaceuticals* **2023**, *16*, 1567. <https://doi.org/10.3390/ph16111567>

Academic Editors: Tyler W. LeBaron and Jan Slezak

Received: 31 August 2023

Revised: 3 October 2023

Accepted: 6 October 2023

Published: 7 November 2023



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1. Introduction

Molecular hydrogen (H_2) is a colorless, odorless and tasteless diatomic gas with small molecular size. H_2 can readily cross the biologic barriers, penetrate the tissue and efficiently remove surplus reactive oxygen species (ROS) [1,2]. In addition to its antioxidative capacity, H_2 also has the characteristics of anti-inflammation, anti-apoptosis and cytoprotection in mitochondria [3]. Emerging animal experiments and clinical studies have utilized the H_2 in the treatment of ocular diseases and have demonstrated promising therapeutic potential. For instance, H_2 can prevent a reduction in tear stability in patients and exerts therapeutic effects on the oxidative ocular diseases in animal models.

ROS are produced via the partial reduction of molecular oxygen under normal physiological conditions [4]. ROS encompass molecules derived from O_2 , including superoxide ($O_2^{\bullet-}$), hydrogen peroxide (H_2O_2), hydroxyl radical ($\bullet OH$), ozone and singlet oxygen [5]. Typically, oxidative stress is caused by excessive ROS production, mitochondrial dysfunction, an impaired antioxidant system or a combination of these factors [6–8]. Excessive ROS can disrupt the redox homeostasis and result in the production of a large amounts of peroxynitrite ($ONOO^-$) and $\bullet OH$ [9]. Peroxynitrite acid ($ONOOH$) is a form of protonation of $ONOO^-$, a strong oxidant that can exacerbate the risk of oxidative stress in the body [9]. At the same time, $ONOOH$ can also be decomposed to form intermediates ($\bullet OH$ and $\bullet NO_2$) [9]. Both $\bullet NO_2$ and $\bullet OH$ are indiscriminate in what they will oxidize, which creates the havoc called oxidative stress [5]. In this context, excessive ROS will cause oxidative damage to deoxyribonucleic acid (DNA), proteins and lipids. For instance, ROS can react with the nitrogen-containing bases of nucleic acids and the glycosylphosphate skeleton of DNA, resulting in chromosome and mitochondrial DNA (mtDNA) damage [10].

DNA damage can affect the protein-coding region and non-coding regulatory region of the gene, thus affecting protein expression and regulation [11,12]. DNA damage can also activate the I κ B kinase (IKK) complex, resulting in the phosphorylation, ubiquitination and degradation of I κ B α and the activation of nuclear factor kappaB (NF- κ B) [13]. Oxidative stress produces reactive oxygen intermediates (ROI), which activate caspase-1, causing the proteolytic cleavage of NF- κ B proteins, leading to apoptosis [14]. mtDNA damage causes further respiratory chain dysfunction and increase membrane permeability, resulting in the excessive production of ROS and a continuous oxidative stress cycle [15]. ROS also attack the structural and enzymatic proteins by oxidizing residual amino acids and pseudogroups, thereby forming cross-links, protein aggregates and proteolysis [16]. The inactivation of these proteins can lead to serious abnormalities in key metabolic pathways. Lipid peroxidation is the oxidation process of polyunsaturated fatty acids (PUFAs) [16,17]. Due to the existence of multiple double bonds in PUFA structures, lipid peroxidation is involved in the production of peroxides (compounds in which two oxygen atoms are connected via a single covalent bond), ROS and other active organic free radicals. Lipid peroxidation can also induce apoptosis, ferroptosis and autophagy during the cell death [18].

In the past few decades, oxidative stress has been recognized as a critical pathogenic factor of various ocular diseases, such as the endothelial corneal dystrophy, terygium, glaucoma, cataracts, uveitis, retinopathy and optic neuropathies [4,19]. Oxidative stress can impair tissue structure, increase vascular permeability and promote neovascularization in the eye [20–23]. Moreover, oxidative stress can initiate inflammatory response, ocular cell apoptosis and ferroptotic cell death [14,24,25]. In this context, oxidative stress may act as an effective target for the treatment of ocular disease.

2. Advantages and Potentials of Molecular Hydrogen in Treating Ocular Diseases

The molecular weight of H₂ is 2.01588, which is the smallest in nature. The density of H₂ is 0.089 g/L (101.325 kpa, 0 °C), which is 1/14 of the same volume of air [26]. The small molecular weight and low density endow H₂ with an extremely strong ability to penetrate tissue rapidly. H₂ behaves as an inert gas in the presence of catalysts at body temperature [1]. Only strong oxidants, like ONOO⁻, are able to oxidize H₂ [3]. On these bases, H₂ is very mild and safe in human tissue, neither disturbing normal metabolic redox reactions nor affecting the physiological function of ROS in cell signaling [3]. Once excessive ROS are produced to induce oxidative stress in cells, H₂ can efficiently attack •NO₂ and •OH, which are the main cytotoxic ROS [27]. In 2007, Ohsawa et al. reported for the first time that H₂ is a selective antioxidant when administered via gaseous rapid diffusion into tissues and cells [2]. Unlike other therapeutic gases, such as NO, CO or hydrogen sulfide, H₂ will not induce any toxicity even at high concentrations [1]. These advantages make H₂ an excellent candidate for treating ocular diseases (Figure 1). In ophthalmological practice, the dynamic and static biologic barriers are the most challenging, since they will affect the bioavailability of drugs in both the anterior and posterior segments. These biologic barriers will retract the passive absorption and reduce the delivery efficiency of therapeutic agents [28]. For instance, eye drops account for about 90% of ophthalmic medications for anterior segment eye diseases, but only 1–5% of the applied drug remains on the ocular surface for a sufficiently long time [29]. Furthermore, the static and dynamic barriers of the eye hinder the penetration of various therapeutic agents, including conventional small molecules and macromolecules, such as antioxidants (e.g., superoxide dismutase, catalase and vitamin C); anti-inflammatory agents (e.g., cefazolin, amikacin); anti-aggregation agents, like protein chaperones (e.g., clusterin and crystallins); small molecule chaperones (e.g., quinacrine, curcumin and chlorpromazine), etc. [30,31].

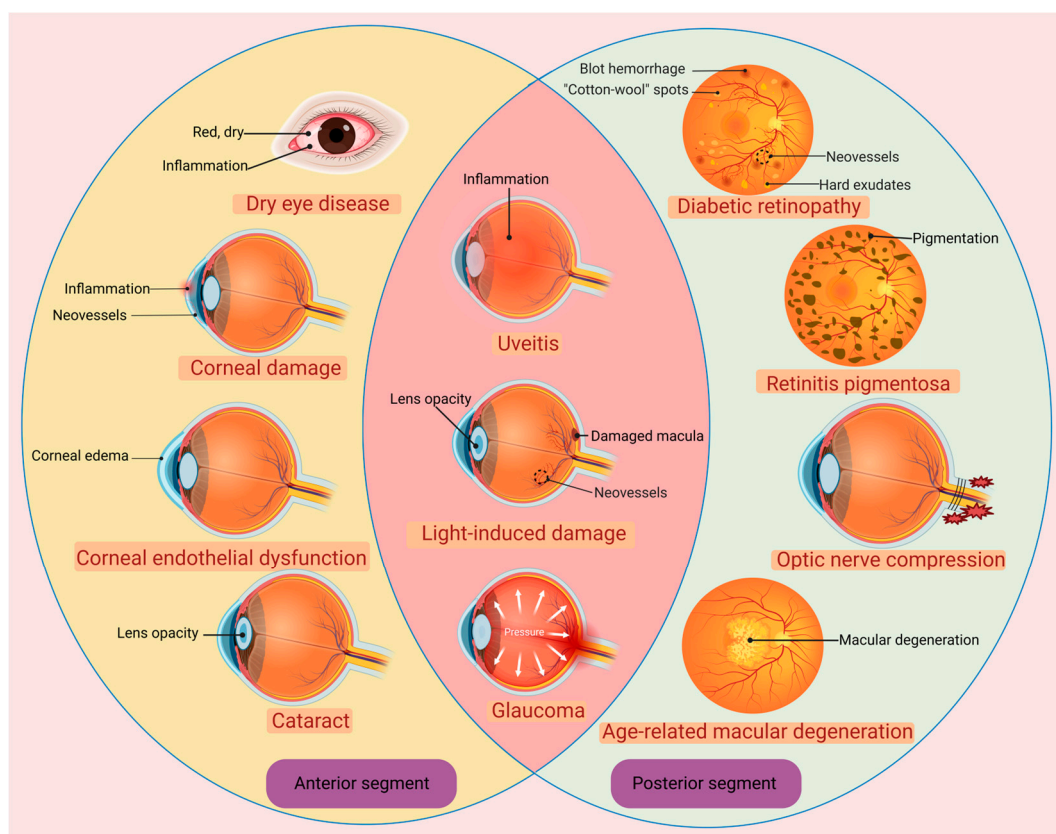


Figure 1. The emerging applications of hydrogen in the treatment of various ophthalmological diseases. Created with [BioRender.com](https://www.biorender.com).

The static barrier of the cornea is the outermost part of the ocular barrier. It is composed of the lipophilic corneal epithelium, hydrophilic corneal stroma, corneal endothelium and blood aqueous barrier [28]. Although the lipophilic corneal epithelium allows the penetration and absorption of hydrophobic drugs, the paracellular diffusion will still be restricted by the tight junctions between corneal epithelium cells [32,33]. Simultaneously, some efflux transporters in the plasma membrane of corneal epithelial cells also reduce the transportation rate of drugs [34]. Moreover, the dynamic barriers such as tear drainage, conjunctival lymph, blood and aqueous humor circulation would reduce sharply the penetrate efficiency of therapeutic molecules [29]. Even if a small proportion of drugs bypass the corneal barrier, the conjunctiva, sclera, choroid, Bruch's membrane and retinal barrier in the anatomic pathway are other conundrums [29]. In particular, the thickness of Bruch's membrane can increase with age, showing a gradually stronger blocking ability [28]. The blood–retinal barrier (BRB) is another challenge for efficient drug delivery [35]. BRB consists of the internal and external barriers. The internal barrier is composed of the adherens junction and zonula occludens between vascular endothelial cells. The external barrier is composed of the RPE cells and their connections, which can separate choroid interstitial fluid from retinal interstitial fluid [28]. Collectively, these layered barriers hinder the transportation of most therapeutic drugs into the eye. However, H_2 can quickly cross these biologic barriers and enter cellular membranes and even mitochondria and nuclei due to its powerful penetrating ability [1]. Additionally, H_2 can scavenge the ROS selectively on the basis of its unique reducibility in a selective manner without cytotoxicity. Hence, owing to its superior characteristics, H_2 gas holds immense potential for extensive investigation into diverse ophthalmic diseases (Table 1).

Table 1. Experimental studies on the ophthalmological application of H₂.

Ocular Diseases	Experimental Objects	Disease Models	Methods of H ₂ Uptake	Effect of H ₂	Refs.
Dry eye disease	Wistar rats	Scopolamine-induced DED	Intraperitoneal injection of HRS (0.6 mmol/L) at a dose of 5 mL/kg BW daily for 28 days; dropping HRS into the eye 1 time per hour, 9 times per day	Inhibiting the activity of NF-κB to reduce inflammation	[36]
Alkali burn of cornea	SOD-1 ^{-/-} mice or WT mice	Alkali burn	Irrigated with HRW (0.5–0.6 ppm) onto the cornea for 30 min	Reducing oxidative stress; inhibiting angiogenesis in cornea	[37]
	Wistar rats		Irrigated with HRW (1.2–1.6 ppm) onto the cornea for 5 min	Upregulating the expression of antioxidants	[38]
UVB	SOD-1 ^{-/-} mice or WT mice	Ultraviolet B ray-induced corneal damage model	Irrigated with HRW (0.5–0.6 ppm) onto the cornea	Reducing oxidative stress; inhibiting of angiogenesis in cornea	[37]
Corneal endothelial dysfunction	Rabbits	MNU-induced corneal endothelial cell injury	Irrigated with HRS (1.2 ppm) 3 times a day for 3 min and 3 drops per second for 7 days	Anti-apoptotic effect through the NF-κB/NLRP3 and FOXO3a/p53/p21 pathway	[39]
	Rabbits	Corneal endothelial dysfunction induced by phacoemulsification	Ultrasound oscillation with irrigation solution at almost 61% H ₂ dissolved concentration for 30 s	Reducing oxidative stress	[40]
Cataract	Rats	Selenite-induced cataract	Intraperitoneal injection of HRS (0.6 mmol/L) at a dose of 5 mL/kg BW daily from postnatal day 8 to postnatal day 17	Maintaining the activity of antioxidant enzymes, inhibiting lipid peroxidation	[41]
Uveitis	Rats	Endotoxin-induced uveitis	Inhaling mixed gas that consisted of 67% H ₂ and 33% O ₂ for once a day for 3 weeks	Suppressing the microglia activation	[42]
	Rats		Intraperitoneal injection of HRS (0.6 mM) at a dose of 10 mL/kg BW once a day for 1 week	Maintaining the integrity of the blood–aqueous barrier	[43]
RP	Rats	Rd6 rats	Drinking HRW (1.2–1.6 ppm) 3.42 ± 0.14 mL/day for 1 week	Neuroprotective effect	[44]
	Rats	MNU-induced RP	Intraperitoneal (10 mL/kg BW) and intravitreal (8 μL) injections of HRS (0.6 mmol/L)	Increasing the level of SOD, modulating the expressions of apoptosis-related genes	[45]
DR	Male rats	Rats with streptozotocin-induced diabetes mellitus	Intraperitoneal injections of HRS (0.86 mmol/L) at a dose of 5 mL/kg BW daily for 1 month	Reducing oxidative stress; preserving synaptophysin and BDNF levels	[46]
	C57BL/6J mice	Rats with diabetes mellitus	Intraperitoneal injections of HRS	Reducing the retinal neovascularization, and the expression of VEGF and MDA	[47]
Glaucoma	Rats	Retinal ischemia/reperfusion	Consecutive peritoneally injected of HRS (0.6 mM) at a dose of 5 mL/kg BW until the rats were sacrificed	Alleviating apoptosis of RGCs by overactivating PARP-1	[48]
			Inhaling mixed gas that consisted of 67% H ₂ and 33% O ₂ for 1 h daily for 7 days	Lessening RGCs loss; reducing the levels of IL-1-β, TNF-α and 4-HNE	[49]
AMD	Mice	NaIO ₃ -induced AMD	Intragastric administration of HRS (4.0 mg/L) at a dose of 10 mL/kg BW for 12 days.	Inhibiting cellular senescence; maintaining DNA homeostasis	[50]
			Intragastric administration of HRW (0.55–0.65 mM) at a dose of 1 mL/g three times daily for 12 days	Inhibiting oxidative stress and apoptosis	[51]
		Laser-induced CNV mouse	Inhaling mixed gas that consisted of 21% oxygen, 42% H ₂ and 37% nitrogen gas for 2/5 h daily for 15 days	Alleviating CNV leakage	[52]

Table 1. Cont.

Ocular Diseases	Experimental Objects	Disease Models	Methods of H ₂ Uptake	Effect of H ₂	Refs.
Light-induced retinal damage	Rats	Blue light-induced damage model	Intraperitoneal injection of saturated HRS (0.6 mmol/L) at a dose of 1 mL/100 g BW once a day before and during the exposure session	Suppressing photo-oxidative stress	[53]
			Intraperitoneal injection of saturated HRS (5 mL/kg) before and within 5 days after light exposure		[54]
Optic nerve injury	Rats	Establishing the optic nerve crush model via surgery	Intraperitoneal injection of saturated HRS (5 mL/kg) at 6:00 and 18:00 lasting for 2 weeks	Reducing the lipid peroxidation and apoptosis of RGCs	[55]
	Guinea pigs	Glutamate-induced retinal injury model	Intravitreal (almost 0.6 mmol/L) and/or peritoneal injection (5 mL/kg) of HRS	Clearing glutamate by increasing EAAT-1; reducing RGCs apoptosis of by upregulating GRP78	[56]
	RGCs cells	S-nitroso-N-acetylpenicillamine-induced oxidative stress model	Culturing cell in medium consisting of 5% O ₂ and 95% H ₂ (v/v) for 24–72 h	Suppressing ONOO ⁻ -mediated oxidative stress by clearing peroxynitrite	[57]

Abbreviations: H₂, molecular hydrogen; DED, dry eye disease; HRS, hydrogen-rich saline; NF-κB, nuclear factor kappa B; SOD-1, superoxide dismutase-1; HRW, hydrogen-rich water; MNU, N-Nitroso-N-methylurea; NLRP3, NOD-like receptor family pyrin domain containing 3; FOXO3a, forkhead box O3; BDNF, brain-derived neurotrophic factor; VEGF, vascular endothelial-derived growth factor; MDA, malondialdehyde; AMD, age-related macular degeneration; CNV, choroidal neovascularization; EAAT-1, excitatory amino acid transporter 1; RGCs, retinal ganglion cells; GRP78, glucose-regulated protein 78; WT, wild-type; BW, bodyweight; ONOO⁻, peroxynitrite.

3. Mechanisms Underlying the Therapeutic Effects of Molecular Hydrogen

H₂ can readily penetrate biofilms compared to classic anti-oxidants, such as superoxide dismutase (SOD), catalase (CAT) and alpha-tocopherol, and H₂ is able to specifically eliminate cytotoxic free radicals (Figure 2) [58]. In order to determine the free radical species reduced directly by H₂, researchers have tested the reactivity of H₂ with artificially prepared ROS or reactive nitrogen species (RNS) in cell-free experiments. The results show that H₂ can directly neutralize •OH and generate water, thereby inhibiting the primary free radical chain reactions. However, H₂ does not affect directly the levels of O₂•⁻, H₂O₂ and •NO, all of which play essential roles in signal transduction [2]. In addition to directly alleviating oxidative stress, H₂ can also promote the expression of endogenous antioxidant enzymes. Nuclear factor erythroid 2-related factor 2 (Nrf2) is a basic region leucine-zipper transcription factor that plays a pivotal role in the coordinated gene expression of antioxidant enzymes [59]. H₂ can promote Nrf2 transcriptional activation and induce its translocation into the nucleus, thereby enhancing the transcription of anti-oxidative enzymes such as CAT, glutathione 1 and glutathione reductase [26]. Furthermore, H₂ participates in the metabolic processes related to oxidation–reduction reactions. H₂ can decrease the levels of NADPH oxidase subunits, including p40 phox, p47 phox and p67 phox in the cell membrane, but increase their levels in the cytoplasm. By limiting the translocation of these molecules to the cell membrane, H₂ reduces the NADPH oxidase activity [26]. H₂ can also inhibit the activity of NADPH oxidase by blocking the apoptosis signal-regulating kinase 1 (ASK1) pathway and the downstream mediator p38 mitogen-activated protein kinase [60]. Thus, H₂ can indirectly decrease the production of ROS by inhibiting NADPH oxidase activity.

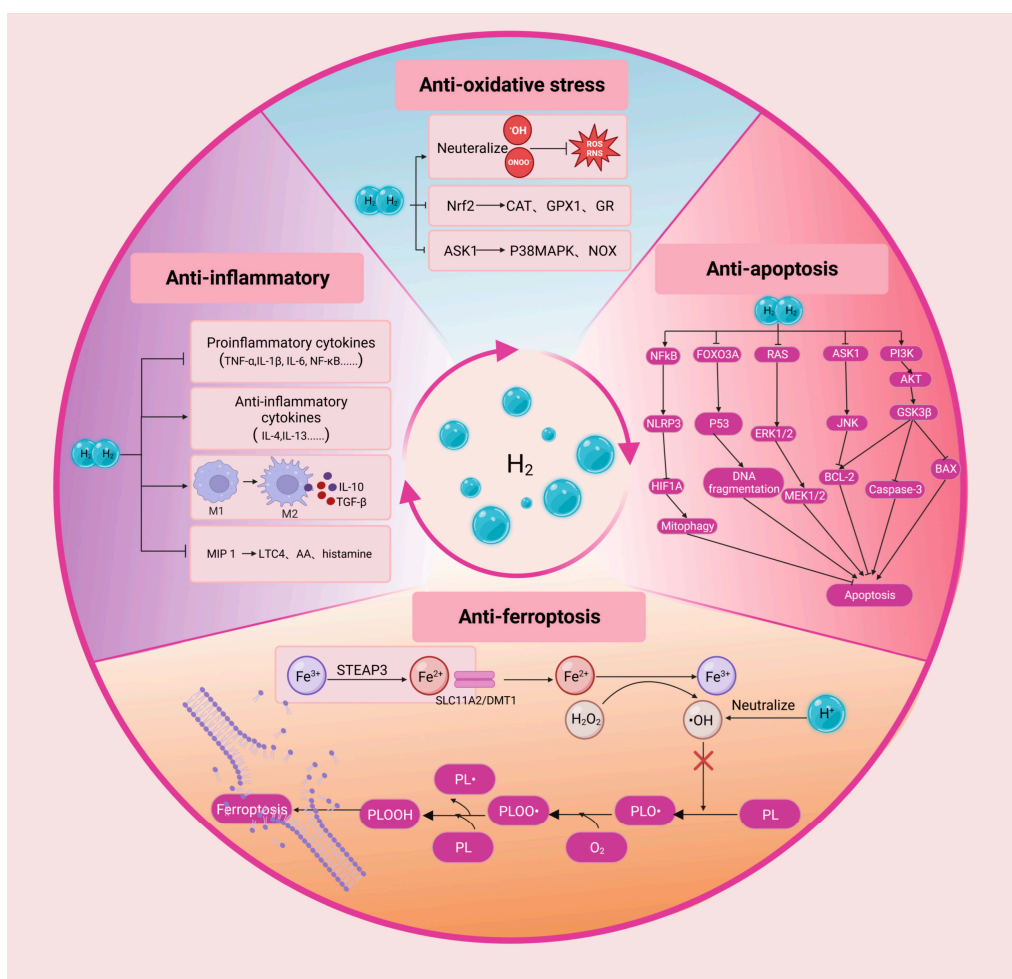


Figure 2. The fundamental principles underlying the therapeutic effects of H₂ in ophthalmological pathologies. H₂, molecular hydrogen; •OH, hydroxyl radical; ONOO⁻, peroxynitrite; ROS, reactive oxygen species; Nrf2, nuclear factor erythroid 2-related factor 2; CAT, catalase; GPX1, glutathione peroxidase 1; GR, glutathione reductase; ASK1, apoptosis signal-regulating kinase 1; P38MAPK, p38 mitogen activated protein kinase; NOX, NADPH Oxidases; TNF-α, tumor necrosis factor alpha; IL, interleukin; NF-κB, nuclear factor kappaB; TGF-β, transforming growth factor-beta; MIP 1, macrophage inflammatory protein 1. LTC4, leukotriene C4; AA, arachidonic acid; STEAP3, six-transmembrane epithelial antigen of the prostate 3; SLC11A2, solute carrier family 11 member 2; DMT1, divalent metal transporter 1; H₂O₂, hydrogen peroxide; PL, phospholipid; PLOOH, phospholipid hydroperoxides; NLRP3, NOD-like receptor family pyrin domain containing 3; RAS, rat sarcoma; ERK1/2, extracellular signal-related kinases 1 and 2; JNK, Jun N-terminal kinase; BCL-2, B cell lymphoma protein-2; PI3K, phosphatidylinositol 3-kinases; GSK3β, glycogen synthase kinase-3beta. Created with [BioRender.com](https://www.biorender.com).

H₂ also exerts protective effects through anti-inflammatory properties. Emerging evidence suggests that H₂ can inhibit the release of proinflammatory cytokines, including interleukin (IL)-1β, IL-6, prostaglandin E₂, tumor necrosis factor-α (TNF-α), NF-κB, interferon-γ and high-mobility group box 1 [3,26,61]. It can also enhance the expression level of anti-inflammatory cytokines, such as IL-4 and IL-13 [26,61]. Moreover, H₂ facilitates macrophage polarization from proinflammatory M1 type to anti-inflammatory M2 type, which in turn generates additional anti-inflammatory cytokines, including IL-10 and transforming growth factor-β [62]. A recent study show that H₂ is able to enhance macrophage-mediated phagocytosis and inhibit the neutrophil recruitment at lesion locations [60]. Moreover, H₂ has been documented to alleviate the inflammation by mitigating

the expression level of macrophage inflammatory protein 1 (MIP 1) family members [61]. MIP 1 exacerbates the acute and chronic inflammatory responses when the tissue is exposed to trauma or infection. This cytokine can induce the release of proinflammatory mediators such as leukotriene C4, arachidonic acid or histamine [63].

H₂ can inhibit the activation of apoptotic cascades in multiple types of tissue. Apoptosis is caused by the successive activation of a series of cysteine proteases known as caspases [60]. Apoptosis is involved in the pathology of various ophthalmological disease, such as glaucoma, retinitis pigmentosa (RP), cataracts, retinoblastoma, retinal ischemia and diabetic retinopathy (DR) [64]. H₂ can protect ocular tissue by modulating the expression of apoptosis-related factors, such as inhibiting the expressions of proapoptotic factors BAX, caspase-3, caspase-8 and caspase-12, while upregulating the expressions of antiapoptotic factors, such as B cell lymphoma protein-2 (BCL-2) and B-Cell lymphoma-extra-large (BCL-XL) [26]. H₂ also inhibits apoptosis by the regulating signal transduction within and between specific pathways. For example, the H₂-mediated neuroprotective effect is at least partially associated with anti-apoptotic protein kinase B (AKT) pathway activation in neurons [3]. In addition, there are other signaling pathways that play a critical role in H₂ inhibiting cell apoptosis, such as PI3K/Akt/GSK3 β , ASK1/JNK and Ras-ERK1/2-MEK1/2 pathways. Notably, H₂ can promote apoptosis in some cases. Apoptosis evasion is a prominent hallmark of cancer, where H₂ inhibits tumor cell proliferation and migration as well as invasion and promotes cell apoptosis in lung cancer [65].

Ferroptosis is a cell death modality that is characterized by the iron-dependent accumulation of lipid peroxidation. Ferroptosis is closely related to the pathogenesis of various ocular diseases. For example, the progressive death of RPE cells and overlying photoreceptors is the hallmark feature of age-related macular degeneration (AMD) pathology [66]. Recent studies suggest that ferroptosis is the momentous mechanism underlying the RPE/photoreceptor cells death in AMD [67]. Malondialdehyde (MDA) is one of the final products of polyunsaturated fatty acids peroxidation. A study shows that drinking hydrogen-rich saline (HRS) can significantly reduce the serum MDA level of older adults [68]. 4-hydroxynonenal (4-HNE) is the most well-studied and seemingly the most relevant product of lipid peroxidation in ferroptosis [69]. Encouragingly, researchers find that H₂ dissolved in medium can decrease the levels of 4-HNE and alleviating the pheochromocytoma cells death in a dose-dependent manner [2]. In addition to protecting cell integrity by reducing the production of lipid peroxidation elements, H₂ can also inhibit ferroptosis by ameliorating oxidative stress. This should be ascribed to some mutual commonalities and overlap between the H₂ signaling and ferroptosis pathways [70]. Oxidative stress can induce the lipid peroxidation of phospholipid (PL) in the cell membrane, leading to the ferroptosis of cells. H₂ may significantly block the occurrence of this process by inhibiting lipid peroxidation [70]. In this context, the anti-ferroptosis mechanism should be partially responsible for the H₂-mediated beneficial effects.

4. Current Administration Approaches of Molecular Hydrogen

4.1. Hydrogen Gas

In the pioneering studies, H₂ gas can be produced through water electrolysis. Currently, the common approaches of H₂ gas therapy include delivery through a facemask, nasal cannula or ventilator circuit [71–73]. H₂ gas therapy has demonstrated many advantages in the treatment of certain diseases. Firstly, it acts rapidly and is particularly suitable for addressing acute oxidative stress. ROS can cause damage to cellular components (Figure 3). The main advantage of H₂ gas relies on its ability to penetrate rapidly cell membranes and enter the organelles, where it specifically targeting and neutralizing cytotoxic ROS [2,74]. Secondly, H₂ gas therapy does not have adverse effects on blood pressure, which is recognized as a crucial risk factor under certain conditions. H₂ gas therapy does not produce side effects on the cardiovascular system, making it a relatively safe treatment option [2]. However, H₂ gas therapy also has some drawbacks. Firstly, it is not as convenient as other portable medications or treatment devices. H₂ needs to be

prepared and stored using gas generators or pressure tanks, which makes H₂ gas therapy challenging to implement in certain situations. Secondly, there is potential risk of explosion when using H₂ gas. Although H₂ gas leakage is rare in clinical practice, the careful management of the delivery process is necessary to ensure the safety of patients and healthcare professionals. Thus, the definitive quantitative data and uniform standards for the application of H₂ gas in ophthalmic practice remain unset. Typically, scientific studies involving the inhalation of H₂ gas require diluting the H₂ gas to an appropriate ratio, with 66% H₂ gas being used in some ophthalmology animal experiments [42,49]. However, the precision in defining both the volume and intensity of H₂ inhalation, under secure conditions, and its further applications in the treatment of ophthalmic diseases, as opposed to other ailments, remain a domain that requires exploration and experimentation. In this context, H₂ gas administration has some limitations which might impede its application and therapeutic efficacy in clinical practice.

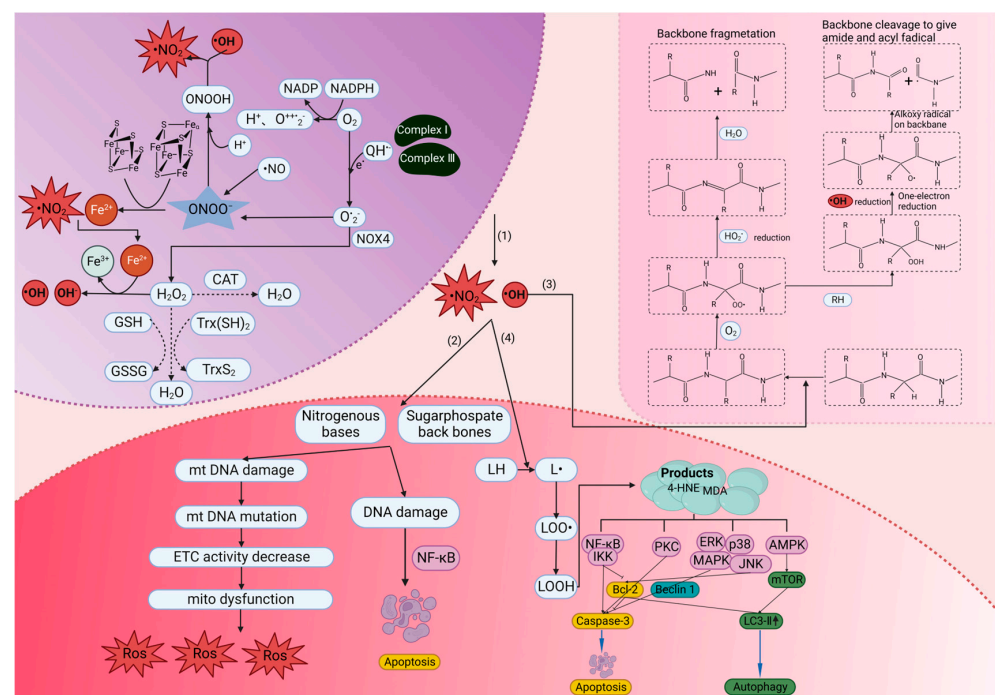


Figure 3. The mechanism of oxidative stress and its damage to DNA, lipids and proteins. (1) $\bullet\text{OH}$ and ONOO^- production mechanism. (2) Damage of chromosome and mitochondrial DNA by $\bullet\text{OH}$ and $\bullet\text{NO}_2$ and its possible consequences. (3) The α -carbon site of amino acid residues can form stable carbon-centered radicals with hydroxyl radicals and react with O_2 to form alkyl peroxy groups. Peroxyl radicals can be removed via an elimination reaction that releases $\text{HO}_2\bullet$ and generates an imine, which subsequently undergoes hydrolysis and thus gives rise to backbone fragmentation. In addition, peroxy radicals can extract hydrogen atoms from another species to produce hydroperoxide. The subsequent decomposition of these hydroperoxides to radicals can also result in backbone fragmentation via an alkoxy-radical-mediated process. (4) LH is a lipid with allylic hydrogens, which are present in polyunsaturated fatty acids, including arachidonic acid. The reaction of $\bullet\text{OH}$ and LH leads to lipid peroxidation and eventually causes autophagy and apoptosis. LH, luteinizing hormone; CAT, catalase; $\text{L}\bullet$, an alkoxy radical; $\text{LOO}\bullet$, peroxy radical; LOOH , a lipid hydroperoxide; $\text{QH}\bullet^-$, ubiquinone; $\bullet\text{OH}$, hydroxyl radical; H_2O_2 , hydrogen peroxide; $\text{O}_2^{\bullet-}$, superoxide; ONOO^- , peroxynitrite; $\bullet\text{NO}$, nitric oxide; $\text{HO}_2\bullet$, hydroperoxyl radical; ETC, electron transport chain; NOX4, NADPH oxidase 4; ONOOH , peroxynitrous acid; MAPKs, mitogen-activated protein kinases; JNK, Jun N-terminal kinase; ERK, extracellular signal-regulated kinase; PKC, protein kinase C; AMPK, adenosine monophosphate-activated protein kinase; mTOR, mammalian target of rapamycin; IKK, I κ B kinase; NF- κ B, nuclear factor κ B. Created with [BioRender.com](https://www.biorender.com).

4.2. Hydrogen-Rich Water/Hydrogen-Rich Saline

Hydrogen-rich water (HRW)/HRS refers to water or saline with a higher concentration of H₂. Currently, several common methods have been developed to produce HRW/HRS. One method is to inject H₂ into water or saline (usually 0.9% NaCl solution) under high pressure, allowing the water or saline to dissolve the H₂. Another method is to produce H₂ through electrolysis and then dissolve it in water or saline. The third method mainly involves the reaction of metal elements with acids (primarily sulfuric acid and hydrochloric acid). By utilizing metals with higher reactivity than H₂, atoms are displaced from H₂O to generate H₂ gas. Finally, these preparation procedures enrich the water or saline with H₂ and endow it with therapeutic potentials. The quantity of H₂ dissolved in HRW or HRS can vary according to the preparing method. Typically, HRW/HRS used in experiments contains a dissolved H₂ concentration of 0.6 mM (1.2 ppm) or above [44,47].

HRW/HRS can be administered in various ways, such as through oral intake, drops, or direct injection [38,75,76] (Figure 4). In ophthalmologic practice, HRW/HRS is always applied through local eye drops. Through this delivery approach, HRW/HRS can directly contact the ocular tissues, rapidly penetrate corneal epithelium and enter the interior eyeball [36]. Experimental results have shown that when HRS eye drops are typically administered on the ocular surface of rat pups, a certain amount of H₂ can be detected in the vitreous humor. Only two minutes after administration, the concentration of H₂ in the vitreous humor begins to increase and peaks at 15 min. At this point, the H₂ concentration in the vitreous humor is approximately 20% of that in the eye drops [77]. Additionally, HRW/HRS can also be administered intravitreal (IV) injections [56]. Unlike oral intake, in which H₂ can both escape over time and be lost in the stomach or intestines, IV injections achieve a more precise control of the H₂ concentration [78]. The precise locale for the injection on the ocular surface is determined, typically about 4 mm from the limbus. Once the needle has secured its position properly, the HRS is judiciously released into the vitreous humor. In this approach, HRW/HRS is directly injected into the vitreous humor of the eye, placing it in direct contact with intraocular cells. This administrative method is typically used for posterior segment pathologies, which is challenging for conventional delivery approach.

HRW/HRS is advantageous in the treatment of ocular diseases. Firstly, it is portable, convenient and safe. Patients can carry HRW/HRS with them without the need for complex equipment or special administrative procedures [79]. Secondly, the therapeutic effect of HRW/HRS is comparable to that of inhaling H₂ gas. H₂ affords antioxidant effects by selectively neutralizing the toxic free radicals in the body. Through oral intake or direct contact with HRW/HRS, the body can absorb water enriched with H₂, thereby achieving the comparable effects of inhaling H₂ gas [80]. Notably, HRW/HRS administration also has some drawbacks: firstly, H₂ gradually escapes from the water over time, making it difficult to store HRW/HRS for an extended period of time. Therefore, patients need to consume or use HRW/HRS within a certain time frame to ensure its efficacy. Secondly, when taken orally, some H₂ may be lost, making it challenging to precisely control the H₂ concentration. Compared to inhaling H₂ gas directly, the oral intake of HRW/HRS may result in reduced absorption efficiency. Furthermore, although HRW/HRS can be administered more easily, the duration of H₂ retention in the body is only approximately 20–30 min after administration, which cannot provide a reliable supply of H₂ like the direct inhalation of H₂ gas [81]. Therefore, careful dosage and frequency control are necessary during HRW/HRS treatment. In clinical practice, a comprehensive consideration of its strengths and limitations is required to ensure that patients obtain an effective concentration of H₂.

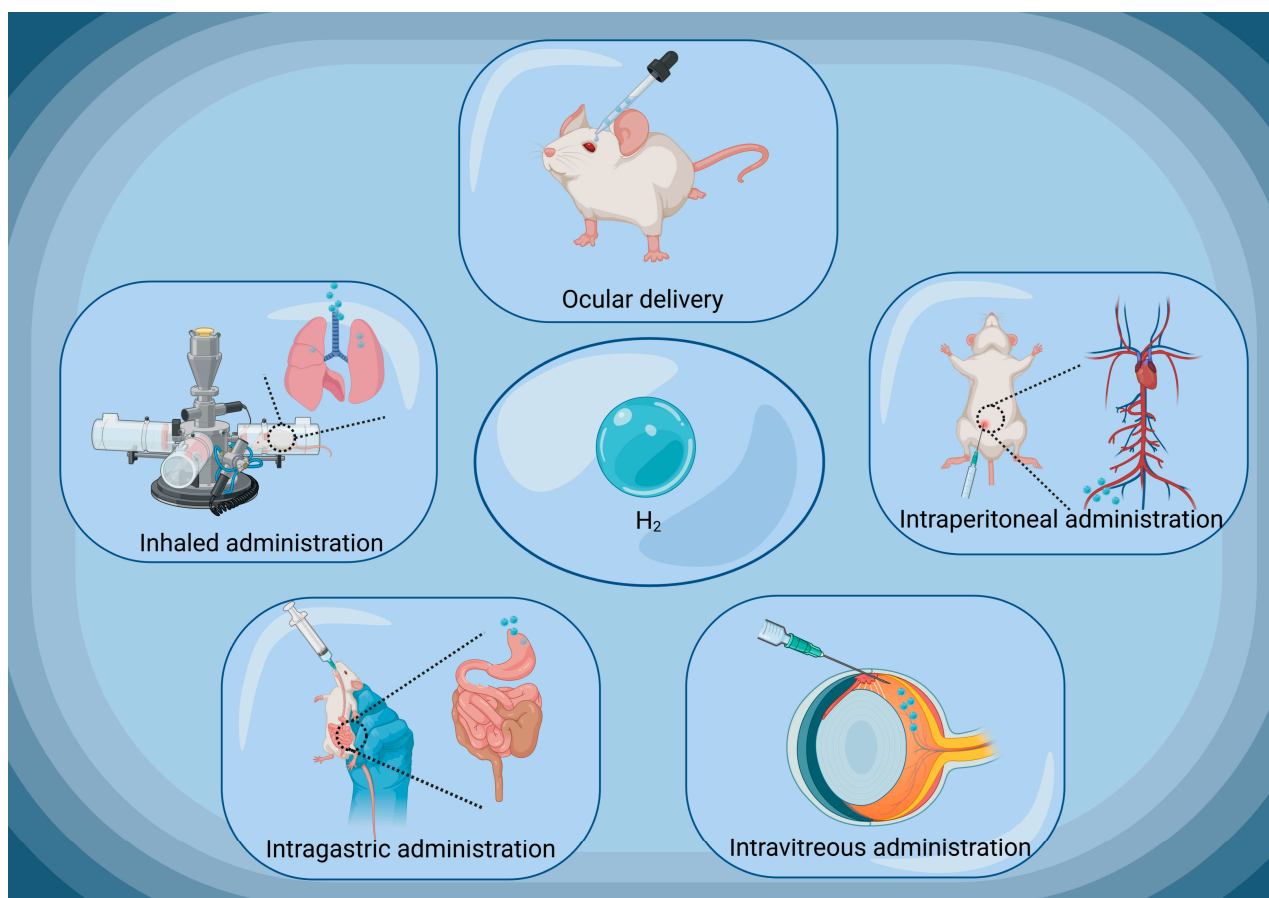


Figure 4. Common routes of H₂ administration in ophthalmology. H₂, molecular hydrogen. Created with BioRender.com.

4.3. Molecular Hydrogen Produced by Intestinal Bacteria

The direct use of H₂ gas poses some safety risks and inconveniences. On the other hand, the short duration of HRW in the body limits its application in clinical practice (Table 2) [81]. In recent years, a novel delivery method has gained abundant interest, in which the researcher utilizes gut bacteria to produce a large amount of H₂ in the body. When indigestible components reach the colon, gut bacteria utilize them to produce H₂, carbon dioxide, short-chain fatty acids, and other endogenous substances [82]. Previous studies have shown that H₂ produced by gut microbiota can influence the central nervous system [83]. Experimental evidence has suggested that certain bifidobacteria in mouse intestines may affect H₂ production through the ingestion of dietary substances [84].

Table 2. Current H₂ therapy methods.

Method	Advantages	Disadvantages	Preparation Method	Administration Method	Ref.
H ₂ gas	Rapid and reliable	Risk of explosion, requires strict management and monitoring	Prepared using H ₂ generators	Direct inhalation of H ₂ gas under professional guidance	[2,74]
	Insignificant effect on blood pressure	Inconveniences during usage			

Table 2. Cont.

Method	Advantages	Disadvantages	Preparation Method	Administration Method	Ref.
HRW/HRS	Simple administration through drinking or injection, easily accepted by patients	Limited storage and duration in the body, limited treatment effect	H ₂ gas injection under high pressure	Oral ingestion or drinking, controlled dosage and frequency as per medical advice	[36,38,56,75–79,81]
	Partly mitigates the risk of direct H ₂ gas usage		Reaction between metals and water	Injection under professional healthcare personnel	
			Electrolysis to produce HRW/HRS	Through drip infusion, controlled drip rate and dosage Local eye drops	
H ₂ produced by intestinal bacteria	Production of H ₂ by intestinal bacteria, providing longer duration of effect	Variability due to individual differences in gut microbiota affecting H ₂ production	Induced by ingesting non-digestible substances	Through normal dietary intake of non-digestible components	[82,84]

Abbreviations: H₂, molecular hydrogen; HRS, hydrogen-rich saline; HRW, hydrogen-rich water.

This method provides new possibilities for the convenient application of H₂ in the treatment of ophthalmic diseases. Researchers have used the H₂-producing milk to prevent the short-fluorescein tear film breakup time (fTBUT) type of dry eye symptom, providing new insights into the clinical application of H₂ [85]. Previous studies have shown that pectin and high amylose corn starch can increase H₂ production in the cecum of rats [86]. Moreover, the results of a clinical trial also show that eating turmeric can also increase the production of respiratory H₂ [87]. It is reasonable to speculate that these substances may also be used like H₂-rich milk in the future.

There are more than 100 trillion kinds of intestinal bacteria from 1000 different species in our large intestine. Among these bacteria, 70% are H₂-producing bacteria [88]. They can ferment substances that cannot be absorbed in the intestines into short-chain fatty acids, releasing carbon dioxide and H₂. In the colon, three different types of microorganisms, including methane-producing bacteria, sulfate-reducing bacteria and acetate-forming bacteria can consume the H₂ in the gut [89–91]. Under physiologic conditions, they do not receive enough H₂, which hinders their significant proliferation. However, once the production of H₂ in the gut is enhanced, the H₂-consuming bacteria will multiply accordingly, resulting in a decrease in the H₂ concentration expelled from the colon [92]. Although certain bacteria in the gut consume H₂, the surplus produced can still make its way to the bloodstream. An increase in H₂ production may surpass the rate of augmentation in the H₂-consuming bacterial population, enabling sufficient H₂ formation within the body. Experimental evidence now substantiates that the ingestion of H₂-producing foods can enhance internal H₂ levels [85,87,93]. Therefore, despite the presence of H₂-consuming microbial communities in the gut, a robust supply of H₂ may still retain potential therapeutic efficacy against eye diseases, assuming sufficient systemic H₂ availability is maintained. If further research can delve into the exact relationship between gut microbiota and H₂ production, it may offer new insights for optimizing the H₂ therapeutic strategies.

5. Molecular Hydrogen-Mediated Therapeutic Effects on Ocular Diseases

Light permeates through every layer of the eye, leading to the generation of a significant volume of ROS. External factors that harm the eye, such as prolonged hyperglycemia and increased intraocular pressures, expedite the process of oxidative stress. Consequently, research focused on ROS has expanded in various ocular pathologies, encompassing conditions like glaucoma, cataract, AMD and retinopathies. It is hypothesized that the development of these diseases occurs when there is an imbalance between ROS generation

and the body's capacity to neutralize them with natural defense mechanisms, including the production of endogenous antioxidants. H₂ is very suitable for preventing or improving ocular diseases due to its unique properties. Compared to most known antioxidants, H₂ possesses advantageous distribution characteristics. It has the ability to permeate biomembranes and diffuse throughout key cellular compartments such as the cytosol, mitochondria, and nucleus. Then, H₂ is able to specifically eliminate the cytotoxic free radical ($\bullet\text{OH}$ and ONOO^-), as a typical redox reaction. H₂ can also increase the expression of Nrf2 mechanisms and endogenous antioxidant enzymes, including NRF2, SOD, PGC-1 α , glutathione peroxidase, glutathione reductase, glutathione S-transferase and so on. Additionally, H₂ therapy can also reduce the expressions of stress transcription factors such as NF- κ B, thereby exerting leads anti-inflammatory and anti-apoptotic effects on ocular cells.

5.1. Molecular Hydrogen-Mediated Therapeutic Effects on Dry Eye Disease (DED)

DED is a common ocular surface disease that can result in inflammation, foreign body sensation and blurred vision in patients [94]. DED can be caused by the reduced production of tears, the excessive evaporation of tears, or a combination of these two processes [95]. Accumulating evidence suggests that ultraviolet radiation, pollutants (particular PM 2.5, gaseous, ozone), aging and microbial antigens all lead to increased ROS in the tear fluid film. These unopposed ROS can directly damage the structures such as the tear lipid layer and the myelin sheath of the ocular surface nerve. ROS also cause cellular stress, leading to ocular surface epithelial and goblet cell dysfunction, inflammation and vascular endotheliopathy, ultimately resulting in tear instability [96]. In an animal experiment, researchers established a scopolamine-induced DED model using Wistar rats. The HRS is administered through intraperitoneal (IP) injection in the DED model. The results show that HRS is effective in the treatment of DED [36]. Typically, when indigestible components reach the large intestine, intestinal bacteria utilize them to produce H₂, together with short-chain fatty acids, carbon dioxide and other substances. H₂ is absorbed through the small intestinal mucosa and enters the bloodstream through the capillaries in the small intestinal villi. Once absorbed into the bloodstream, H₂ is transported via the circulatory system to various cells and tissues throughout the body, including the eyeball [97,98]. Via this mechanism, Kawashima, M., et al. developed a novel method to supply large amounts of H₂ to the human body through intestinal digestion. They added galacto-oligosaccharides (2 g), maltol (2 g) and glucomannan (0.2 g) to a milk solution to generate H₂-producing milk. H₂ can be produced in the colon of patients after they have drunk H₂-producing milk [81]. The results show that the mean value of tTBUT in these DED patients is slowed after drinking. The area under the curve (AUC) of breath H₂ is approximately 5-fold higher for H₂-producing milk than for HRW [85].

5.2. Molecular Hydrogen Protects the Cornea from Alkali Burn

Alkali burns can cause acute corneal inflammation and corneal neovascularization (NV), both of which are the critical cause of blindness. After alkali burns, the level of ROS in the cornea tissue increases rapidly and subsequently activates the nuclear factor kappa B (NF- κ B) [99]. NF- κ B plays a significant role in regulating the expression of genes associated with cell growth, inflammation and apoptosis [100]. It translocates into the cell nucleus and promotes the expression of inflammatory cytokines, including vascular endothelial-derived growth factor (VEGF), monocyte chemoattractant protein-1 (MCP-1), IL and tumor necrosis factor alpha (TNF- α) [101]. These cytokines not only promote growth of NV but also recruit more inflammatory cells, which would exacerbate inflammatory response [37]. An experiment studies the H₂-induced effects on a mouse model with corneal alkali burns. They find that the irrigation of the cornea with H₂ solution exerts anti-angiogenic effects on the alkali-burned region by inhibiting the mRNA level of NF- κ B [37,38].

H₂ exerts these beneficial effects not only through direct reactions with free radicals but also through indirect pathways involving various anti-oxidative factors. Proliferator-activated receptor gamma coactivator 1 α (PGC-1 α) is a transcription coactivator that

has been shown to mitigate the production of ROS [77]. By binding with nuclear receptor transcription factors and similar elements, it modulates the expression levels of target genes, thereby regulating cellular energy metabolism [102,103]. Functioning as an antioxidant enzyme, the SOD1 protein transforms a pair of superoxide anions byproducts stemming from cellular respiration into oxygen and H₂O₂. SOD1 protein plays a crucial role in activating a significant portion of SOD activity, making it an essential element in the body's defense against oxidative stress [104]. Experimental results show that instilling HRS eyedrops onto the ocular surface of normal rats can increase the activity of SOD1 and PGC-1 α in the corneal epithelial tissue. Furthermore, the expression level of VEGF mRNA in the corneal tissue decreases after H₂ treatment, confirming the H₂-mediated anti-angiogenic effects on the cornea. Even after ceasing H₂ administration for over 6 h, these beneficial activities could still be detected in the cornea tissue [38].

5.3. Molecular Hydrogen-Mediated Therapeutic Effects on Ultraviolet B Ray-Induced Corneal Damage

Ultraviolet (UV) radiation does not affect our visual perception. However, it primarily causes damage to corneal structures. Compared to visible light, UV carries higher energy, and can induce a range of effects and interactions. For instance, it can trigger chemical reactions and more readily ionize atoms and molecules. Additionally, it has the capability of damaging DNA molecules, posing harm to living organisms. Although the cornea efficiently absorbs more than 90% of ultraviolet B (UVB) radiation, excessive exposure to UVB can lead to serious eye problems, including photokeratitis, corneal epithelial apoptosis and corneal edema [105,106].

A critical mechanism underlying UVB-induced corneal damage can be ascribed to the over-production of ROS. Under physiological condition, xanthine oxidase activity is distributed in the corneal epithelium, corneal endothelium and lens epithelium. After excessive exposure to UVB, xanthine oxidase activity increases significantly and a notable decrease in xanthine dehydrogenase activity is seen. The primary function of xanthine dehydrogenase enzyme is not the production of O₂^{•-} and H₂O₂, but rather the conversion of xanthine into uric acid, which serves as an effective antioxidant. Xanthine oxidase functions as a xanthine oxidoreductase, generating ROS and exacerbating oxidative stress on the cornea. The reduction in antioxidants, coupled with an increase in ROS, further intensifies corneal damage [107]. Moreover, excessive UVB exposure can result in the reduction in antioxidant level in the cornea. During UVB irradiation, antioxidant enzymes in the corneal epithelium, including SOD, glutathione peroxidase and CAT, are down-regulated. Consequently, a larger burst of ROS will not be counteracted in a timely manner by the antioxidant enzymes and thus accumulate in the cornea [108]. To this end, the exogenous supplement of antioxidants is crucial for protecting the cornea from oxidative damage. A recent study shows that the irrigation of the cornea with H₂ solution can effectively suppress corneal oxidative stress caused by UVB irradiation in SOD-1-deficient mice or wild-type mice, thereby reducing corneal inflammation and promoting beneficial corneal healing [37].

5.4. The Potential Therapeutic Effects on Corneal Endothelial Dysfunction

Endothelial cells play a crucial role in maintaining the transparency of cornea by supporting the pump function [109]. The human corneal endothelium lacks regeneration ability and leaves no opportunity for endogenous supplementation. Corneal endothelial dysfunction can result in stromal edema and the loss of transparency, thereby inducing severe visual impairments in patients [110]. Several etiologies, including Fuchs' endothelial corneal dystrophy, posterior polymorphous corneal dystrophy and phacoemulsification surgery, can cause endothelial cell dysfunction [111]. For instance, clinical studies have shown that a decreased corneal endothelial density and an increase in cell variability may occur after phacoemulsification surgery [112]. During the phacoemulsification procedure, the probe oscillates at ultrasonic frequencies, creating intense ultrasonic waves that travel

through the liquid medium. In this context, small cavitation bubbles are formed, which subsequently enlarge and implode. The heat generated by the implosion of these bubbles leads to the decomposition of water molecules and the production of highly active free radicals ($\text{H}_2\text{O} \rightarrow \bullet\text{OH} + \bullet\text{H}$). These toxic ROS can lead to corneal oxidative damage in patients [113,114]. Although corneal transplant is considered the definitive treatment for corneal endothelial dysfunction, there is a global shortage of donated corneal grafts. This dilemma has prompted researchers to explore alternative therapeutic options [115]. Excitingly, a clinical study has shown that the H_2 dissolved in the flushing solution can reduce corneal endothelial damage during phacoemulsification [116]. Another animal experiment also shows that H_2 dissolved in the ocular irrigating solution is protective against the endothelial cell damage caused by phacoemulsification in rabbit eyes [40]. Researchers have successfully established a corneal endothelium decompensation rabbit model [39]. They find that irrigating rabbit eyes with HRS can improve the survival and physiological function of corneal endothelial cells by inhibiting apoptotic cascades. Furthermore, the H_2 can mitigate the oxidative stress in corneal tissue via the NF- κ B/NLRP3 pathway and the FOXO3A/P53/P21 pathway.

5.5. Molecular Hydrogen-Mediated Therapeutic Effects on Cataract

Cataracts are caused by the gradual aggregation or misfolding of crystallin proteins within the lens [117]. Currently, 94 million people worldwide are blind or visually impaired, with cataracts (the clouding of the lens) being the most common cause [118]. The conformational changes of lens proteins are mainly caused by oxidative stress, changes in osmotic pressure and phase separation of proteins and water [119]. Notably, the initial mechanism of cataracts should be ascribed to the lipid peroxidation caused by free radicals (especially $\bullet\text{OH}$) [120]. Lipid peroxidation will affect the permeability of cell membrane, change the internal composition and configuration of lens epithelial cells, lead to malfunction of lens protein and finally cause the occurrence of cataract [19]. H_2O_2 is a type of ROS that exists in the aqueous humor of mammals. It can activate a series of signaling transduction events, such as the BCL-2 family, the caspases, NF- κ B pathways and the mitogen-activated protein kinases. These signaling will induce the apoptosis of the lens epithelial cell and cause lens opacity [121]. Yang, C. X., et al. injected sodium selenite (25 $\mu\text{mol}/\text{kg}$ bodyweight) subcutaneously into neonatal Sprague Dawley rats to induce cataracts. Subsequently, they injected HRS (5 mL/kg bodyweight) intraperitoneally every day from 8 to 17 days after the birth of the rats. The severity and development rate of cataracts in the HRS-treated rats was alleviated compared with untreated controls. After an IP injection of HRS, the activity of SOD, CAT, glutathione peroxidase, glutathione reductase and glutathione S-transferase was significantly higher than that of untreated mice [41]. Taken together, these findings suggest that HRS has a salvage effect on the development of cataracts.

5.6. The Potential Therapeutic Effects against Uveitis

Uveitis is a medical nomenclature used to describe a collection of diseases characterized by the inflammation of the uvea, which encompasses the iris, ciliary body, and choroid. As a type of intraocular inflammation, the etiology of uveitis is multifaceted, and the pathogenic mechanism is yet unclear [122]. Emerging evidence suggest that ROS plays a critical role in the inflammatory reaction of uveitis. The increased ROS production exacerbates inflammation by promoting the release of pro-inflammatory cytokines and chemokines. Additionally, these cytokines and chemokines contribute to the infiltration of polymorphonuclear leukocytes and macrophages, which further activate the inflammatory cascade [123]. Researchers find that H_2 , may act as a promising medication to treat uveitis. For example, continuous H_2 inhalation can suppress the release of inflammatory cytokine (MCP-1 and IL-6), thereby alleviating the microglia activation in a rat model of endotoxins-induced uveitis. Moreover, continuous H_2 inhalation can help to maintain the integrity of the blood–aqueous barrier by reducing the level of prostaglandin E_2 , which mediates the exudation of protein into the aqueous humor [42]. Another study using the

same uveitis model also shows that IP injection of HRS is beneficial for the normal function of the blood–aqueous barrier [43]. Therefore, H₂ can help to mitigate the ROS production in uvea tissue and alleviate the symptoms in uveitis patients.

5.7. Molecular Hydrogen-Mediated Therapeutic Effects on Retinitis Pigmentosa

RP is a degenerative retinopathy with genetically heterogeneous features. Photoreceptor degeneration in the RP patients moves from the mid-periphery of the fundus and then progresses toward the macular and foveal regions, with typical symptoms of night blindness emerge first, followed by reduced visual field and, eventually, legal blindness occurs in these RP patients [124]. In RP pathogenesis, the mechanism of rod cell death varies depending on the mutated gene. The initial Rod cell loss would mitigate the oxygen consumption, resulting in elevated oxygen levels in the retinal tissue. Excess oxygen stimulates the production of O₂^{•−} through mispairing of the electron transport chain in mitochondria and stimulating the NADPH oxidase activity in the cytoplasm. High levels of free O₂^{•−} can overwhelm antioxidant defenses and produce more reactive species, like ONOO[−], which is extremely difficult to detoxify. These metabolism alterations will impose constant oxidative stress on the cones. In this context, H₂ is used to promote cone cell survival and maintain the visual function in RP model [125]. A study has shown that drinking HRW improves the photoreceptor survival and function in rd6 mice [44]. As evidenced by the optical coherence tomography examination, the retinal thickness of the HRW-treated group is significantly higher than that of untreated controls. Histopathological and morphological analysis show that the thickness of outer nuclear layer and the number of opsin red/green (cone opsin) positive cells in HRW-treated mice are higher than those in the control group. RNA sequencing (RNA-seq) analysis shows that there are significant differences in the expression levels of 1996 genes between the two groups. In greater detail, gene and pathway ontology analysis show that the genes responsible for phototransduction are significantly up-regulated in HRW-treated mice. Collectively, these results show that drinking HRW (1.2–1.6 ppm) exerts neuroprotective effect on the cone cell death in RP mice [44]. In another study, researchers deliver HRS into a chemical-induced RP rat model through IP and IV injections. The results show that the average cone density of HRS treatment rats is significantly larger than that of untreated controls, IV injection group has a better therapeutic effect than the IP injection group. Mechanism study shows that HRS treatment increases the level of retinal SOD and reduces the level of retinal MDA in the RP rat model. HRS treatment can modulate the expression level of severe apoptosis-associated genes, such as the BAX, BCL-2, Calpain-2, and caspase-3s [45].

5.8. Molecular Hydrogen-Mediated Therapeutic Effects on Diabetic Retinopathy

DR is a progressive complication of diabetes with complex metabolic and pathological features. It is characterized by the microvascular damage and visual impairments. It stems from the chronic and progressive effects of diabetes, causing a sequence of retinal microvascular leaks and obstructions. This cascade results in an array of retinal abnormalities, including microaneurysms, hard exudates, cotton-wool spots, neovascularization, vitreous proliferation, macular edema and, potentially, retinal detachment. The excessive production of ROS can induce structural and functional abnormalities in the retinal microvasculature, leading to the breakdown of the BRB and promoting the progression of DR [126,127]. Additionally, ROS can upregulate the expression level of VEGF and enhance the sessility of VEGF receptors. The overactivation of VEGF leads to the growth of the choroid neovascular, which are also known as choroidal neovascularization (CNV) [128,129].

Compared to other tissues, the retina has the highest oxygen consumption and glucose oxidation levels, making it extremely susceptible to the impacts of oxidative stress [130]. In diabetic patients, there is a decrease in the capacity of antioxidant enzymes to clear free radicals and maintain the redox homeostasis in the retina [131]. For instance, the levels of intracellular antioxidant glutathione are significantly reduced in the diabetic retina [132]. On the other hand, studies have found elevated levels of O₂^{•−} and H₂O₂ in the retinas

of a DR rat model and retinal cell lines cultured in high-glucose media [133,134]. Studies indicate that oxidative stress not only propels the progression of DR but also poses a challenge for its reversal due to the buildup of damaged molecules and ROS, even under well-maintained blood glucose levels [135]. In pioneering studies, HRS has been used to treat the streptozotocin (STZ)-induced DR rats. HRS treatment can inhibit the apoptosis caspase in retinal cells, reduce vascular permeability and alleviate the retinal thinning in the DR rat model [46]. This study evaluates HRS-induced effects on the neurovascular and oxidative stress in DR rats. HRS treatment mitigated diminished b-wave amplitudes and oscillatory potentials, BRB breakdown and histological changes in the inner retina in STZ-diabetic rats. HRS ameliorates the oxidative stress and enhance the antioxidant enzyme activity in the DR rats [46]. Additionally, another study uses HRS to treat the oxygen-induced retinopathy in mice. Researchers find that HRS treatment alleviates the retinal neovascularization by inhibiting the mRNA and protein levels of VEGF expressions [47].

5.9. Therapeutic Potential of Molecular Hydrogen in Glaucoma

Glaucoma is a group of neurodegenerative diseases that is characterized by optic nerve head damage, the progressive death of RGCs (retinal ganglion cells) and visual field defects [136,137]. Despite the heterogeneous backgrounds of glaucoma, these phenotypes eventually cause the pathological outcome of RGC demise [138,139]. Accumulated evidence obtained from both clinical and experimental research convincingly suggests that oxidative stress is implicated in the RGC loss of glaucoma [140,141]. Oxidative stress can impair the structure of trabecular meshwork, leading to the obstruction of aqueous humor. The intraocular pressure increases significantly, which subsequent results in the retinal ischemia/reperfusion (I/R) injury and RGCs [142,143]. Oxidative stimulus can also attack directly the RGCs degeneration. For example, ROS has been shown to trigger RGC-5 cells (an immortalized RGC line) apoptosis *in vitro* through the caspase-independent pathways [144]. H₂ can slow down the development of glaucoma and rescue RGCs. In a rat retinal I/R model, researchers find that the IP injection of HRS can alleviate RGCs apoptosis through the mitigation of DNA oxidation and the overactivation of poly (ADP-ribose) polymerase-1, a nuclear enzyme that promotes caspase-mediated cell apoptosis [48]. In another therapeutic experiment, it has been found that inhaling H₂ for 1 h daily for 7 consecutive days can significantly lessen RGC loss in a rat retinal I/R model [48]. A mechanism study shows that H₂ significantly reduces the levels of IL1- β , TNF- α and 4-HNE in the retinal tissue [49]. These findings highlight that H₂ may be developed into an efficient clinical treatment for glaucoma.

5.10. Therapeutic Potential of Molecular Hydrogen in Age-Related Macular Degeneration

AMD is a degenerative retinopathy that causes severe visual impairments in the elder population [145]. Oxidative stress-induced RPE cell dysfunction plays a crucial role in the pathogenesis of AMD [146]. Oxidative stress can lead to mtDNA damage and mitochondrion malfunction in RPE cells. These impairments accumulate with age in the macular region [147]. RPE cells play an important role in regulating the retinal function, including nutrient supply, maintaining orientation and clearing metabolic products [148]. Therefore, RPE degradation can cause photoreceptor death and the loss of central vision [149]. Clinical studies have shown that dietary supplements of antioxidants can effectively improve the vision quality of AMD patients [150]. In this context, it is conceivable that exogenous H₂ supplementation can serve as a measure to safeguard the retina functioning and alleviate oxidative stress in AMD patients. Researchers have shown that intragastric administration of HRS can alleviate the oxidative stress in an NaIO₃-induced AMD mice model by enhancing the aging-related protein (SIRTUIN 3) expression and inhibiting cell senescence [50]. Moreover, they found that HRS can protect the retina tissue by reducing DNA alkylation and promoting DNA repair. Another experiment also provides a more comprehensive description of the therapeutic effect of HRW on an NaIO₃-induced AMD mice model [51]. It demonstrated that the intragastric administration of HRW can reduce the deposition

of yellow-white drusen-like structures and improve the rate of photoreceptor survival and the integrity of retinal vessels. These beneficial effects should be attributed to the reduced MDA content and upregulated SOD level. H₂ can also alleviate the photoreceptor apoptosis by inhibiting the expressions of caspase 8 and caspase 9. Some researchers use a laser-induced CNV mouse model to simulate the neovascular AMD [52]. The results show that H₂ inhalation can inhibit the inflammation of an injured retina after Bruch's membrane disruption, suppress CNV development and decrease the leakage of the CNV. The potential mechanism of action may involve the suppression of the VEGF-dependent pathway of CNV formation through the downregulation of VEGF and HIF-1 α . Additionally, it may inhibit the expression of pro-inflammatory mediators, such as TNF- α and IL-6. This mechanism could help reduce the pathological angiogenesis associated with CNV and mitigate the inflammatory response in the retina tissues.

Excessive exposure to blue light is a risk factor of AMD. Blue light between 450 and 500 nanometers can adjust biological rhythms, while the short-wave blue light within 400 to 450 nm is truly harmful. Short wave blue light has high energy, which enables them to penetrate the anterior segment and directly contact the retina. Typically, excessive exposure of blue light can lead to photochemical damage in the retina. Blue light itself can cause a significant increase in ROS production, which will lead to lipid peroxidation and photoreceptor apoptosis [151]. According to previous reports, blue light induces high levels of H₂O₂ and O₂^{•-} and disrupts the mitochondrial respiratory chain, leading to intracellular oxidative stress [152,153]. Notably, blue light at 415–455 nm is the spectral band that most conducive to produce larger bursts of ROS [151]. These ROS can activate the endoplasmic reticulum stress-C/EBP homologous protein (ER stress-CHOP) apoptosis signaling in RPE [154]. The RPE loss can directly lead to the dysfunction and degradation of photoreceptors, thereby accelerating the development of AMD. Excitingly, H₂ can alleviate the blue light-induced retinal damage. Researchers use blue light irradiation for 6 h to induce retinal damage in rats [53]. The experimental results show that the retinas in HRS-treated rats are less damaged compared with those in the untreated controls. Another independent study shows that the amplitude of electroretinogram waveform and the average thickness of outer nuclear layer in rats injected intraperitoneally with HRS are significantly higher than those in the untreated controls, indicating that HRS protects photoreceptors from light-induced damage [53].

5.11. Therapeutic Potential of Molecular Hydrogen in Optic Nerve Crush

As a neuroprotective agent, H₂ can prevent oxidative stress and inflammation in many neurological diseases. In ophthalmic practice, H₂ can afford some beneficial effects in treating optic neuropathy. Traumatic optic nerve injury is a serious complication of traumatic brain injury, where the mechanical pressure crushes the optic nerve and causes axonal degeneration. This damage eventually causes neural degeneration, leading to RGC loss and complete blindness in patients [155,156]. Nevertheless, oxidative stress is now considered as one of the most important factors that mediate the above process. In addition, glutamate-induced retinal excitotoxicity acts as another pathogenic mechanism of the neuronal damage in optic nerve crush (ONC). A study shows that the IP injection of HRS for consecutive 14 days can promote the survival of recurrent ganglia cells in a ONC rat model [55]. Researcher have also shown that IP or IV of HRS in rats can promote glutamate clearance and enhance the survival of RGCs [56]. During the early stages of neuronal damage, microglia are activated by glutamate to release inflammatory mediators and toxic substances in the retina [157]. Studies report that HRS can inhibit the microglia the activation in the glutamate-induced retinal toxic injury model in white guinea pigs [56]. NO is an essential signaling factor in oxidative-mediated neuron damage. Another study shows that introducing H₂ gas into a retina incubator can alleviate the cell loss in the GCL and the INL through neutralizing the ONOO⁻ [57]. Accordingly, the H₂-mediated effects on oxidative stress or glutamate toxicity may support the hypothesis that H₂ can protect the optic nerve in ONC through the aforementioned effects.

6. Conclusions and Prospects

As a powerful antioxidant, the H₂-mediated benefits are mainly based on its antioxidant, anti-inflammatory and anti-apoptotic properties. It exhibited remarkable potential for the treatment of diseases of the nervous system (such as stroke, Parkinson's disease), cardiovascular diseases (such as I/R injury, hypertension) and liver diseases (such as fatty liver, liver fibrosis) [158–163]. Between 2016 and 2023, H₂ therapy gained substantial recognition as a treatment approach for ophthalmic diseases. Notably, researchers have forayed into novel territory by employing H₂ in animal models to address a broad spectrum of ophthalmic conditions, including the corneal endothelial injury, uveitis and AMD, among others. Furthermore, H₂ has formally entered the realm of clinical research for ophthalmic diseases, encompassing conditions like cataracts and dry eye disease. This progression marks a significant stride in harnessing the potential of H₂ in ophthalmologic practice. In clinical practice, the bag containing the ocular irrigation solution is placed in an acrylic vacuum chamber (SNS-type, Sanplatec Corporation, Osaka, Japan) for a period of 24 h. Throughout this treatment, the chamber is purged of air and replaced with 100% H₂ gas. Subsequent analysis suggests that the concentration of dissolved H₂ within the solution has successfully achieved a level of 61.9%. During phacoemulsification surgery for cataract patients, this solution containing dissolved H₂ has been used for irrigation. The endothelial cell density reduction rate in the H₂ group is significantly smaller compared with that observed in the control group at 1 day, 1 week and 3 weeks postoperatively. Additionally, the results of slit-lamp photography and specular microscopy show that the corneal edema is apparently milder in eyes treated with H₂ solution post operation [116]. Currently, H₂ drinks for clinical use are also available. They are packaged as HRW in 420 mL plastic aluminum packs equipped with a gas-tight cap (Aquistamina-R, Nutristamina, Ostrava, Czech Republic). As per the manufacturer's specifications, HRW is generated from drinking water that has undergone chlorine removal and the subsequent infusion of H₂ directly into the water under high pressure. This manufacturing process ensures the creation of a H₂-rich beverage with specific packaging to maintain its integrity and therapeutic potential for clinical applications. It has been clinically used for the treatment of glaucoma. However, the rapid intake of 1260 mL of both HRW and H₂-free water causes a statistically significant increase in IOP compared to the baseline in healthy individuals. It can be explained that just drinking water could cause parasympathetic nervous system stimulation, which may disrupt the functionality of Schlemm's canal, ultimately resulting in an elevation of intraocular pressure [164]. Current limitations in usage method, including special requirements for H₂ preparation and storage, as well as inconveniences and potential explosion risks, may hinder the widespread application of H₂. It is difficult to realize the stable and sustained release of H₂ through ingesting HRW/HRS. The optimal dosage, injection approach and treatment duration for HRW/HRS have not been clearly defined, demanding the further optimization of therapeutic plans. However, H₂ production by intestinal bacteria has emerged as a novel method for H₂ treatment. Thus far, the H₂-producing milk has been used to treat DED in clinical experiment. Some large-scale clinical studies are necessary to validate its effectiveness and safety during application. At present, research has demonstrated the ability of H₂ to induce tumor cell damage and apoptosis [165]. The therapeutic effect of H₂ gas in ocular tumors may be a fruitful area for further work. As a therapeutic drug that is almost flawless, the therapeutic effect of H₂ on ophthalmic diseases is undisputed. Considerably more prospective clinical studies with larger sample sizes are needed to ensure that standardized H₂ medication goes to the market.

Author Contributions: Conceptualization, S.-Y.L., Z.-M.S. and Y.T.; methodology, S.-Y.L., Z.-M.S. and Y.T.; formal analysis, S.-Y.L., R.-Y.X., H.W., N.P., D.W. and N.Z.; writing—original draft preparation, S.-Y.L., R.-Y.X., H.W., N.P., D.W. and N.Z.; writing—review and editing, S.-Y.L., R.-Y.X., H.W., N.P., D.W., Z.-M.S. and Y.T.; supervision, Z.-M.S. and Y.T. All authors have read and agreed to the published version of the manuscript.

Funding: This work was funded by the National Natural Science Foundation of China, grant number No. 82101162, Health science and Technology Innovation Outstanding Young talents training program of Henan Province, grant number No. YXKC2021027, and Basic Research Project of Henan Eye Institute, grant number No. 21JCQN002 and 21JCZD002.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: Data sharing is not applicable.

Conflicts of Interest: The authors declare no conflict of interest.

References

- Ohta, S. Recent progress toward hydrogen medicine: Potential of molecular hydrogen for preventive and therapeutic applications. *Curr. Pharm. Des.* **2011**, *17*, 2241–2252. [[CrossRef](#)]
- Ohsawa, I.; Ishikawa, M.; Takahashi, K.; Watanabe, M.; Nishimaki, K.; Yamagata, K.; Katsura, K.; Katayama, Y.; Asoh, S.; Ohta, S. Hydrogen acts as a therapeutic antioxidant by selectively reducing cytotoxic oxygen radicals. *Nat. Med.* **2007**, *13*, 688–694.
- Cejka, C.; Kubinova, S.; Cejkova, J. The preventive and therapeutic effects of molecular hydrogen in ocular diseases and injuries where oxidative stress is involved. *Free Radic. Res.* **2019**, *53*, 237–247.
- Nita, M.; Grzybowski, A. The Role of the Reactive Oxygen Species and Oxidative Stress in the Pathomechanism of the Age-Related Ocular Diseases and Other Pathologies of the Anterior and Posterior Eye Segments in Adults. *Oxid. Med. Cell. Longev.* **2016**, *2016*, 1–23.
- Karkucinska-Wieckowska, A.; Simoes, I.C.M.; Kalinowski, P.; Lebiezinska-Arciszewska, M.; Zieniewicz, K.; Milkiewicz, P.; Górska-Ponikowska, M.; Pinton, P.; Malik, A.N.; Krawczyk, M.; et al. Mitochondria, oxidative stress and Non-Alcoholic Fatty Liver Disease: A complex relationship. *Eur. J. Clin. Investig.* **2022**, *52*, e13622.
- Cui, H.; Kong, Y.; Zhang, H. Oxidative stress, mitochondrial dysfunction, and aging. *J. Signal. Transduct.* **2012**, *2012*, 646354.
- Franceschelli, S.; Gatta, D.M.P.; Pesce, M.; Ferrone, A.; Di Martino, G.; Di Nicola, M.; De Lutiis, M.A.; Vitacolonna, E.; Patruno, A.; Grilli, A.; et al. Modulation of the oxidative plasmatic state in gastroesophageal reflux disease with the addition of rich water molecular hydrogen: A new biological vision. *J. Cell. Mol. Med.* **2018**, *22*, 2750–2759.
- Abad-Jiménez, Z.; López-Domènech, S.; Gómez-Abril, S.Á.; Periañez-Gómez, D.; de Marañón, A.M.; Bañuls, C.; Morillas, C.; Víctor, V.M.; Rocha, M. Effect of Roux-en-Y Bariatric Bypass Surgery on Subclinical Atherosclerosis and Oxidative Stress Markers in Leukocytes of Obese Patients: A One-Year Follow-Up Study. *Antioxidants* **2020**, *9*, 734.
- Forman, H.J.; Zhang, H. Targeting oxidative stress in disease: Promise and limitations of antioxidant therapy. *Nat. Rev. Drug Discov.* **2021**, *20*, 689–709.
- Shokolenko, I.; Venediktova, N.; Bochkareva, A.; Wilson, G.L.; Alexeyev, M.F. Oxidative stress induces degradation of mitochondrial DNA. *Nucleic Acids Res.* **2009**, *37*, 2539–2548. [[CrossRef](#)]
- Finkel, T. Signal transduction by reactive oxygen species. *J. Cell. Biol. Vol.* **2011**, *194*, 7–15.
- Evans, M.D.; Dizdaroglu, M.; Cooke, M.S. Oxidative DNA damage and disease: Induction, repair and significance. *Mutat. Res./Rev. Mutat. Res.* **2004**, *567*, 1–61.
- Janssens, S.; Tschopp, J. Signals from within: The DNA-damage-induced NF-kappaB response. *Cell Death Differ.* **2006**, *13*, 773–784. [[PubMed](#)]
- Krishnamoorthy, R.R.; Crawford, M.J.; Chaturvedi, M.M.; Jain, S.K.; Aggarwal, B.B.; Al-Ubaidi, M.R.; Agarwal, N. Photo-oxidative stress down-modulates the activity of nuclear factor-kappaB via involvement of caspase-1, leading to apoptosis of photoreceptor cells. *J. Biol. Chem.* **1999**, *274*, 3734–3743. [[PubMed](#)]
- Bahr, T.; Welburn, K.; Donnelly, J.; Bai, Y. Emerging model systems and treatment approaches for Leber’s hereditary optic neuropathy: Challenges and opportunities. *Biochim. Biophys. Acta (BBA)-Mol. Basis Dis.* **2020**, *1866*, 165743.
- Blasiak, J.; Petrovski, G.; Veréb, Z.; Facsó, A.; Kaarniranta, K. Oxidative Stress, Hypoxia, and Autophagy in the Neovascular Processes of Age-Related Macular Degeneration. *Biomed. Res. Int.* **2014**, *2014*, 1–7.
- Nam, T. Lipid Peroxidation and Its Toxicological Implications. *Toxicol. Res.* **2011**, *27*, 1–6. [[PubMed](#)]
- Su, L.; Zhang, J.; Gomez, H.; Murugan, R.; Hong, X.; Xu, D.; Jiang, F.; Peng, Z. Reactive Oxygen Species-Induced Lipid Peroxidation in Apoptosis, Autophagy, and Ferroptosis. *Oxid. Med. Cell. Longev.* **2019**, *2019*, 1–13.
- Hsueh, Y.J.; Chen, Y.N.; Tsao, Y.T.; Cheng, C.M.; Wu, W.C.; Chen, H.C. The Pathomechanism, Antioxidant Biomarkers, and Treatment of Oxidative Stress-Related Eye Diseases. *Int. J. Mol. Sci.* **2022**, *23*, 1255.
- Sadowska-Bartosz, I.; Bartosz, G.; Grune, T.; Sereikaite, J. Role of Oxidative, Nitrative, and Chlorinative Protein Modifications in Aging and Age-Related Diseases. *Oxid. Med. Cell. Longev.* **2018**, *2018*, 1–2. [[CrossRef](#)]
- Hull, D.S.; Green, K. Oxygen free radicals and corneal endothelium. *Lens Eye Toxic. Res.* **1989**, *6*, 87–91. [[PubMed](#)]
- Kowluru, R.A.; Chan, P. Oxidative Stress and Diabetic Retinopathy. *Exp. Diabetes Res.* **2007**, *2007*, 1–12.
- Beatty, S.; Koh, H.H.; Phil, M.; Henson, D.; Boulton, M. The role of oxidative stress in the pathogenesis of age-related macular degeneration. *Surv. Ophthalmol.* **2000**, *45*, 115–134. [[PubMed](#)]

24. Tian, L.; Wen, Y.; Li, S.; Zhang, P.; Wang, Y.; Wang, J.; Cao, K.; Du, L.; Wang, N.; Jie, Y. Benefits and Safety of Astaxanthin in the Treatment of Mild-To-Moderate Dry Eye Disease. *Front. Nutr.* **2022**, *8*, 796951. [[PubMed](#)]
25. Totsuka, K.; Ueta, T.; Uchida, T.; Roggia, M.F.; Nakagawa, S.; Vavvas, D.G.; Honjo, M.; Aihara, M. Oxidative stress induces ferroptotic cell death in retinal pigment epithelial cells. *Exp. Eye Res.* **2019**, *181*, 316–324.
26. Fu, Z.; Zhang, J.; Zhang, Y. Role of Molecular Hydrogen in Ageing and Ageing-Related Diseases. *Oxid. Med. Cell. Longev.* **2022**, *2022*, 2249749.
27. Yamamoto, H.; Ichikawa, Y.; Hirano, S.; Sato, B.; Takefuji, Y.; Satoh, F. Molecular Hydrogen as a Novel Protective Agent against Pre-Symptomatic Diseases. *Int. J. Mol. Sci.* **2021**, *22*, 7211. [[CrossRef](#)]
28. Gote, V.; Sikder, S.; Sicotte, J.; Pal, D. Ocular Drug Delivery: Present Innovations and Future Challenges. *J. Pharmacol. Exp. Ther.* **2019**, *370*, 602–624.
29. Lanier, O.L.; Manfre, M.G.; Bailey, C.; Liu, Z.; Sparks, Z.; Kulkarni, S.; Chauhan, A. Review of Approaches for Increasing Ophthalmic Bioavailability for Eye Drop Formulations. *AAPS PharmSciTech.* **2021**, *22*, 107.
30. Kulkarni, S.S.; Kompella, U.B. Potential Drug Delivery Approaches for XFS-associated and XFS-associated Glaucoma. *J. Glaucoma* **2014**, *23*, S77–S79. [[CrossRef](#)]
31. Yannuzzi, N.A.; Si, N.; Relhan, N.; Kuriyan, A.E.; Albin, T.A.; Berrocal, A.M.; Davis, J.L.; Smiddy, W.E.; Townsend, J.; Miller, D.; et al. Endophthalmitis After Clear Corneal Cataract Surgery: Outcomes Over Two Decades. *Am. J. Ophthalmol.* **2017**, *174*, 155–159. [[CrossRef](#)]
32. Uang, H.; Schoenwald, R.D.; Lach, J.L. Corneal Penetration Behavior of P-Blocking Agents 111: In Vitro-In Vivo Correlations. *J. Pharm. Sci.* **1983**, *72*, 1279–1281.
33. Hornof, M.; Toropainen, E.; Urtili, A. Cell culture models of the ocular barriers. *Eur. J. Pharm. Biopharm.* **2005**, *60*, 207–225.
34. Gote, V.; Ansong, M.; Pal, D. Prodrugs and nanomicelles to overcome ocular barriers for drug penetration. *Expert Opin. Drug Met.* **2020**, *16*, 885–906.
35. Nguyen, H.; Eng, S.; Ngo, T.; Dass, C.R. Delivery of therapeutics for deep-seated ocular conditions—Status quo. *J. Pharm. Pharmacol.* **2018**, *70*, 994–1001.
36. Chu, Y.Y.; Hua, N.; Ru, Y.S.; Zhao, S.Z. The protection of hydrogen-rich saline on a rat dry eye model induced by scopolamine hydrobromide. *Zhonghua Yan Ke Za Zhi* **2017**, *11*, 363–371.
37. Kubota, M.; Shimmura, S.; Kubota, S.; Miyashita, H.; Kato, N.; Noda, K.; Ozawa, Y.; Usui, T.; Ishida, S.; Umezawa, K.; et al. Hydrogen and N-acetyl-L-cysteine rescue oxidative stress-induced angiogenesis in a mouse corneal alkali-burn model. *Investig. Ophthalm. Vis. Sci.* **2011**, *52*, 427–433. [[CrossRef](#)]
38. Kasamatsu, M.; Arima, T.; Ikebukuro, T.; Nakano, Y.; Tobita, Y.; Uchiyama, M.; Shimizu, A.; Takahashi, H. Prophylactic Instillation of Hydrogen-Rich Water Decreases Corneal Inflammation and Promotes Wound Healing by Activating Antioxidant Activity in a Rat Alkali Burn Model. *Int. J. Mol. Sci.* **2022**, *23*, 9774. [[PubMed](#)]
39. Li, R.; Qu, Y.; Li, X.; Tao, Y.; Yang, Q.; Wang, J.; Diao, Y.; Li, Q.; Fang, Y.; Huang, Y.; et al. Molecular Hydrogen Attenuated N-methyl-N-Nitrosourea Induced Corneal Endothelial Injury by Upregulating Anti-Apoptotic Pathway. *Investig. Ophthalm. Vis. Sci.* **2021**, *62*, 2.
40. Igarashi, T.; Ohsawa, I.; Kobayashi, M.; Igarashi, T.; Suzuki, H.; Iketani, M.; Takahashi, H. Hydrogen prevents corneal endothelial damage in phacoemulsification cataract surgery. *Sci. Rep.* **2016**, *6*, 31190.
41. Yang, C.; Yan, H.; Ding, T. Hydrogen saline prevents selenite-induced cataract in rats. *Mol. Vis.* **2013**, *19*, 1684–1693.
42. Yan, W.; Chen, T.; Long, P.; Zhang, Z.; Liu, Q.; Wang, X.; An, J.; Zhang, Z. Effects of Post-Treatment Hydrogen Gas Inhalation on Uveitis Induced by Endotoxin in Rats. *Med. Sci. Monit.* **2018**, *24*, 3840–3847. [[CrossRef](#)]
43. Yan, W.M.; Zhang, L.; Chen, T.; Zhao, G.H.; Long, P.; An, J.; Zhang, Z.M. Effects of hydrogen-rich saline on endotoxin-induced uveitis. *Med. Gas Res.* **2017**, *7*, 9–18.
44. Igarashi, T.; Ohsawa, I.; Kobayashi, M.; Miyazaki, K.; Igarashi, T.; Kameya, S.; Shiozawa, A.L.; Ikeda, Y.; Miyagawa, Y.; Sakai, M.; et al. Drinking hydrogen water improves photoreceptor structure and function in retinal degeneration 6 mice. *Sci. Rep.* **2022**, *12*, 13610.
45. Tao, Y.; Chen, T.; Fang, W.; Yan, Z.; Yang, Q.; Huang, Y.; Yu, L.; Fan, L. The Comparative Efficiency of Intraperitoneal and Intravitreal Injection of Hydrogen Rich Saline against N-Methyl-N-Nitrosourea Induced Retinal Degeneration: A Topographic Study. *Front. Pharmacol.* **2017**, *8*, 587. [[CrossRef](#)] [[PubMed](#)]
46. Feng, Y.; Wang, R.; Xu, J.; Sun, J.; Xu, T.; Gu, Q.; Wu, X. Hydrogen-Rich Saline Prevents Early Neurovascular Dysfunction Resulting from Inhibition of Oxidative Stress in STZ-Diabetic Rats. *Curr. Eye Res.* **2013**, *38*, 396–404. [[CrossRef](#)] [[PubMed](#)]
47. Huang, L.; Zhao, S.; Zhang, J.H.; Sun, X. Hydrogen Saline Treatment Attenuates Hyperoxia-Induced Retinopathy by Inhibition of Oxidative Stress and Reduction of VEGF Expression. *Ophthalmic Res.* **2012**, *47*, 122–127. [[CrossRef](#)] [[PubMed](#)]
48. Liu, H.; Hua, N.; Xie, K.; Zhao, T.; Yu, Y. Hydrogen-rich saline reduces cell death through inhibition of DNA oxidative stress and overactivation of poly (ADP-ribose) polymerase-1 in retinal ischemia-reperfusion injury. *Mol. Med. Rep.* **2015**, *12*, 2495–2502.
49. Wang, R.; Wu, J.; Chen, Z.; Xia, F.; Sun, Q.; Liu, L. Postconditioning with inhaled hydrogen promotes survival of retinal ganglion cells in a rat model of retinal ischemia/reperfusion injury. *Brain Res.* **2016**, *1632*, 82–90.
50. Li, R.; Liu, Y.; Xie, J.; Huang, X.; Zhang, L.; Liu, H.; Li, L. Sirt3 mediates the protective effect of hydrogen in inhibiting ROS-induced retinal senescence. *Free. Radic. Biol. Med.* **2019**, *135*, 116–124.

51. Liu, Y.; Li, R.; Xie, J.; Hu, J.; Huang, X.; Ren, F.; Li, L. Protective Effect of Hydrogen on Sodium Iodate-Induced Age-Related Macular Degeneration in Mice. *Front. Aging Neurosci.* **2018**, *10*, 389. [[PubMed](#)]
52. Liang, I.; Ko, W.; Hsu, Y.; Lin, Y.; Chang, Y.; Zong, X.; Lai, P.; Chang, D.; Hung, C. The Anti-Inflammatory Effect of Hydrogen Gas Inhalation and Its Influence on Laser-Induced Choroidal Neovascularization in a Mouse Model of Neovascular Age-Related Macular Degeneration. *Int. J. Mol. Sci.* **2021**, *22*, 12049. [[CrossRef](#)] [[PubMed](#)]
53. Feng, M.; Wang, X.; Yang, X.; Xiao, Q.; Jiang, F. Protective effect of saturated hydrogen saline against blue light-induced retinal damage in rats. *Int. J. Ophthalmol.-Chi* **2012**, *5*, 151–157.
54. Tian, L.; Zhang, L.; Xia, F.; An, J.; Sugita, Y.; Zhang, Z. Hydrogen-rich saline ameliorates the retina against light-induced damage in rats. *Med. Gas Res.* **2013**, *3*, 19. [[CrossRef](#)]
55. Sun, J.; Xu, T.; Zuo, Q.; Wang, R.; Qi, A.; Cao, W.; Sun, A.; Sun, X.; Xu, J.; Agudo-Barriuso, M. Hydrogen-rich saline promotes survival of retinal ganglion cells in a rat model of optic nerve crush. *PLoS ONE* **2014**, *9*, e99299.
56. Wei, L.; Ge, L.; Qin, S.; Shi, Y.; Du, C.; Du, H.; Liu, L.; Yu, Y.; Sun, X. Hydrogen-rich saline protects retina against glutamate-induced excitotoxic injury in guinea pig. *Exp. Eye Res.* **2012**, *94*, 117–127. [[CrossRef](#)] [[PubMed](#)]
57. Yokota, T.; Kamimura, N.; Igarashi, T.; Takahashi, H.; Ohta, S.; Oharazawa, H. Protective effect of molecular hydrogen against oxidative stress caused by peroxynitrite derived from nitric oxide in rat retina. *Clin. Exp. Ophthalmol.* **2015**, *43*, 568–577. [[CrossRef](#)]
58. Penders, J.; Kissner, R.; Koppenol, W.H. ONOOH does not react with H₂: Potential beneficial effects of H₂ as an antioxidant by selective reaction with hydroxyl radicals and peroxynitrite. *Free Radic. Biol. Med.* **2014**, *75*, 191–194. [[CrossRef](#)]
59. Zgorzynska, E.; Dziedzic, B.; Walczewska, A. An Overview of the Nrf2/ARE Pathway and Its Role in Neurodegenerative Diseases. *Int. J. Mol. Sci.* **2021**, *22*, 9592.
60. Fu, Z.; Zhang, J. Molecular hydrogen is a promising therapeutic agent for pulmonary disease. *J. Zhejiang Univ.-Sci. B* **2022**, *23*, 102–122.
61. Nogueira, J.E.; Branco, L.G.S. Recent Advances in Molecular Hydrogen Research Reducing Exercise-Induced Oxidative Stress and Inflammation. *Curr. Pharm. Des.* **2021**, *27*, 731. [[PubMed](#)]
62. Yao, W.; Guo, A.; Han, X.; Wu, S.; Chen, C.; Luo, C.; Li, H.; Li, S.; Hei, Z. Aerosol inhalation of a hydrogen-rich solution restored septic renal function. *Aging* **2019**, *11*, 12097–12113.
63. Maurer, M.; von Stebut, E. Macrophage inflammatory protein-1. *Int. J. Biochem. Cell Biol.* **2004**, *36*, 1882–1886. [[CrossRef](#)] [[PubMed](#)]
64. Nickells, R.W.; Zack, D.J. Apoptosis in ocular disease: A molecular overview. *Ophthalmic Genet.* **1996**, *17*, 145.
65. Wang, D.; Wang, L.; Zhang, Y.; Zhao, Y.; Chen, G. Hydrogen gas inhibits lung cancer progression through targeting SMC3. *Biomed. Pharmacother.* **2018**, *104*, 788–797. [[PubMed](#)]
66. Yang, M.; So, K.; Lam, W.C.; Lo, A.C.Y. Novel Programmed Cell Death as Therapeutic Targets in Age-Related Macular Degeneration? *Int. J. Mol. Sci.* **2020**, *21*, 7279.
67. Zhao, T.; Guo, X.; Sun, Y. Iron Accumulation and Lipid Peroxidation in the Aging Retina: Implication of Ferroptosis in Age-Related Macular Degeneration. *Aging Dis.* **2021**, *12*, 529.
68. Zanini, D.; Todorovic, N.; Korovljev, D.; Stajer, V.; Ostojic, J.; Purac, J.; Kojic, D.; Vukasinovic, E.; Djordjievski, S.; Sopic, M.; et al. The effects of 6-month hydrogen-rich water intake on molecular and phenotypic biomarkers of aging in older adults aged 70 years and over: A randomized controlled pilot trial. *Exp. Gerontol.* **2021**, *155*, 111574.
69. Milkovic, L.; Zarkovic, N.; Marusic, Z.; Zarkovic, K.; Jaganjac, M. The 4-Hydroxynonenal-Protein Adducts and Their Biological Relevance: Are Some Proteins Preferred Targets? *Antioxidants* **2023**, *12*, 856.
70. Yang, M.; Dong, Y.; He, Q.; Zhu, P.; Zhuang, Q.; Shen, J.; Zhang, X.; Zhao, M. Hydrogen: A Novel Option in Human Disease Treatment. *Oxid. Med. Cell. Longev.* **2020**, *2020*, 1–17.
71. Abraini, J.H.; Gardette-Chauffour, M.C.; Martinez, E.; Rostain, J.C.; Lemaire, C. Psychophysiological reactions in humans during an open sea dive to 500 m with a hydrogen-helium-oxygen mixture. *J. Appl. Physiol.* **1994**, *76*, 1113–1118. [[PubMed](#)]
72. Buchholz, B.M.; Kaczorowski, D.J.; Sugimoto, R.; Yang, R.; Wang, Y.; Billiar, T.R.; McCurry, K.R.; Bauer, A.J.; Nakao, A. Hydrogen Inhalation Ameliorates Oxidative Stress in Transplantation Induced Intestinal Graft Injury. *Am. J. Transpl.* **2008**, *8*, 2015–2024.
73. Roberts, B.J.; Fife, W.P.; Corbett, T.H.; Schabel, F.M., Jr. Response of five established solid transplantable mouse tumors and one mouse leukemia to hyperbaric hydrogen. *Cancer Treat. Rep.* **1978**, *62*, 1077–1079.
74. Hong, Y.; Chen, S.; Zhang, J.M. Hydrogen as a selective antioxidant: A review of clinical and experimental studies. *J. Int. Med. Res.* **2010**, *38*, 1893–1903. [[CrossRef](#)] [[PubMed](#)]
75. Zhang, X.; Yu, P.; Hong, N.; Liu, F.; Shan, Y.; Wu, Y.; An, B.; Sang, H.; Kong, Q. Effect and mechanism of hydrogen-rich bath on mice with imiquimod-induced psoriasis. *Exp. Derm.* **2023**, 1–8. [[CrossRef](#)]
76. Chen, T.; Tao, Y.; Yan, W.; Yang, G.; Chen, X.; Cao, R.; Zhang, L.; Xue, J.; Zhang, Z. Protective effects of hydrogen-rich saline against N-methyl-N-nitrosourea-induced photoreceptor degeneration. *Exp. Eye Res.* **2016**, *148*, 65–73.
77. Oharazawa, H.; Igarashi, T.; Yokota, T.; Fujii, H.; Suzuki, H.; Machide, M.; Takahashi, H.; Ohta, S.; Ohsawa, I. Protection of the retina by rapid diffusion of hydrogen: Administration of hydrogen-loaded eye drops in retinal ischemia-reperfusion injury. *Investig. Ophthalm. Vis. Sci.* **2010**, *51*, 487–492. [[CrossRef](#)]
78. Cai, J.; Kang, Z.; Liu, K.; Liu, W.; Li, R.; Zhang, J.H.; Luo, X.; Sun, X. Neuroprotective effects of hydrogen saline in neonatal hypoxia-ischemia rat model. *Brain Res.* **2009**, *1256*, 129–137.

79. Nagata, K.; Nakashima-Kamimura, N.; Mikami, T.; Ohsawa, I.; Ohta, S. Consumption of molecular hydrogen prevents the stress-induced impairments in hippocampus-dependent learning tasks during chronic physical restraint in mice. *Neuropsychopharmacology* **2009**, *34*, 501–508.
80. Nakashima-Kamimura, N.; Mori, T.; Ohsawa, I.; Asoh, S.; Ohta, S. Molecular hydrogen alleviates nephrotoxicity induced by an anti-cancer drug cisplatin without compromising anti-tumor activity in mice. *Cancer Chemother. Pharm.* **2009**, *64*, 753–761.
81. Shimouchi, A.; Nose, K.; Yamaguchi, M.; Ishiguro, H.; Kondo, T. Breath hydrogen produced by ingestion of commercial hydrogen water and milk. *Biomark. Insights* **2009**, *4*, 27–32.
82. Flint, H.J.; Bayer, E.A.; Rincon, M.T.; Lamed, R.; White, B.A. Polysaccharide utilization by gut bacteria: Potential for new insights from genomic analysis. *Nat. Rev. Microbiol.* **2008**, *6*, 121–131. [[PubMed](#)]
83. Zhai, X.; Chen, X.; Shi, J.; Shi, D.; Ye, Z.; Liu, W.; Li, M.; Wang, Q.; Kang, Z.; Bi, H.; et al. Lactulose ameliorates cerebral ischemia–reperfusion injury in rats by inducing hydrogen by activating Nrf2 expression. *Free Radic. Biol. Med.* **2013**, *665*, 731–741.
84. Nishimura, N.; Tanabe, H.; Komori, E.; Sasaki, Y.; Inoue, R.; Yamamoto, T. Transplantation of High Hydrogen-Producing Microbiota Leads to Generation of Large Amounts of Colonic Hydrogen in Recipient Rats Fed High Amylose Maize Starch. *Nutrients* **2018**, *10*, 144. [[CrossRef](#)] [[PubMed](#)]
85. Kawashima, M.; Tsuno, S.; Matsumoto, M.; Tsubota, K. Hydrogen-producing milk to prevent reduction in tear stability in persons using visual display terminals. *Ocul. Surf.* **2019**, *17*, 714–721. [[PubMed](#)]
86. Nishimura, N.; Tanabe, H.; Sasaki, Y.; Makita, Y.; Ohata, M.; Yokoyama, S.; Asano, M.; Yamamoto, T.; Kiriyama, S. Pectin and high-amylose maize starch increase caecal hydrogen production and relieve hepatic ischaemia–reperfusion injury in rats. *Br. J. Nutr.* **2012**, *107*, 485–492.
87. Shimouchi, A.; Nose, K.; Takaoka, M.; Hayashi, H.; Kondo, T. Effect of Dietary Turmeric on Breath Hydrogen. *Dig. Dis. Sci.* **2009**, *54*, 1725–1729. [[CrossRef](#)]
88. Wolf, P.G.; Biswas, A.; Morales, S.E.; Greening, C.; Gaskins, H.R. H₂ metabolism is widespread and diverse among human colonic microbes. *Gut Microbes* **2016**, *7*, 235–245. [[CrossRef](#)]
89. GR, G. Physiology and ecology of the sulphate-reducing bacteria. *J. Appl. Bacteriol.* **1990**, *69*, 769–797.
90. Lajoie, S.F.; Bank, S.; Miller, T.L.; Wolin, M.J. Acetate production from hydrogen and [13C]carbon dioxide by the microflora of human feces. *Appl. Environ. Microb* **1988**, *54*, 2723–2727. [[CrossRef](#)]
91. Gibson, G.R.; Cummings, J.H.; Macfarlane, G.T. Competition for hydrogen between sulphate-reducing bacteria and methanogenic bacteria from the human large intestine. *J. Appl. Bacteriol.* **1988**, *65*, 241–247.
92. Stocchi, A.; Levitt, M.D. Maintaining intestinal H₂ balance: Credit the colonic bacteria. *Gastroenterology* **1992**, *102*, 1424–1426.
93. Fan, D.; Hu, H.; Sun, X.; Meng, X.; Zhang, Y.; Pan, S. Oral administration of lactulose: A novel therapy for acute carbon monoxide poisoning via increasing intestinal hydrogen production. *Undersea Hyperb. Med.* **2016**, *43*, 45–48.
94. Schiffman, R.M. Reliability and Validity of the Ocular Surface Disease Index. *Arch. Ophthalmol.* **2000**, *118*, 615. [[CrossRef](#)] [[PubMed](#)]
95. Kristin, W. What Is Dry Eye Disease? *JAMA-J. Am. Med. Assoc.* **2022**, *328*, 84.
96. Sophia, S.; Tong, L. Dry eye disease and oxidative stress. *Acta Ophthalmol.* **2018**, *96*, e412–e420.
97. Levitt, M.D. Production and excretion of hydrogen gas in man. *N. Engl. J. Med.* **1969**, *281*, 122. [[CrossRef](#)]
98. Hammer, H.F. Colonic hydrogen absorption: Quantification of its effect on hydrogen accumulation caused by bacterial fermentation of carbohydrates. *Gut* **1993**, *34*, 818–822. [[CrossRef](#)]
99. Finkel, T.; Holbrook, N.J. Oxidants, oxidative stress and the biology of ageing. *Nature* **2000**, *408*, 239–247.
100. Viatour, P.; Merville, M.; Bours, V.; Chariot, A. Phosphorylation of NF- κ B and I κ B proteins: Implications in cancer and inflammation. *Trends Biochem. Sci.* **2005**, *30*, 43–52.
101. Baldwin, A.J. The NF-kappa B and I kappa B proteins: New discoveries and insights. *Annu. Rev. Immunol.* **1996**, *14*, 649–683. [[CrossRef](#)] [[PubMed](#)]
102. Rowe, G.C.; Jiang, A.; Arany, Z. PGC-1 Coactivators in Cardiac Development and Disease. *Circ. Res.* **2010**, *107*, 825–838. [[PubMed](#)]
103. Han, B.; Jiang, W.; Liu, H.; Wang, J.; Zheng, K.; Cui, P.; Feng, Y.; Dang, C.; Bu, Y.; Wang, Q.M.; et al. Upregulation of neuronal PGC-1 α ameliorates cognitive impairment induced by chronic cerebral hypoperfusion. *Theranostics* **2020**, *10*, 2832–2848. [[CrossRef](#)] [[PubMed](#)]
104. Crapo, J.D.; Oury, T.; Rabouille, C.; Slot, J.W.; Chang, L.Y. Copper, zinc superoxide dismutase is primarily a cytosolic protein in human cells. *Proc. Natl. Acad. Sci. USA* **1992**, *89*, 10405–10409. [[CrossRef](#)]
105. Izadi, M.; Jonaidi-Jafari, N.; Pourazizi, M.; Alemzadeh-Ansari, M.H.; Hoseinpourfard, M.J. Photokeratitis induced by ultraviolet radiation in travelers: A major health problem. *J. Postgrad. Med.* **2018**, *64*, 40.
106. Zhao, C.; Li, W.; Duan, H.; Li, Z.; Jia, Y.; Zhang, S.; Wang, X.; Zhou, Q.; Shi, W. NAD(+) precursors protect corneal endothelial cells from UVB-induced apoptosis. *Am. J. Physiol.-Cell Physiol.* **2020**, *318*, C796–C805. [[CrossRef](#)]
107. Čejková, J.; Lojda, Z. Histochemical study on xanthine oxidase activity in the normal rabbit cornea and lens and after repeated irradiation of the eye with UVB rays. *Acta Histochem.* **1996**, *98*, 47–52. [[CrossRef](#)]
108. Cejkova, J.; Stipek, S.; Crkovska, J.; Ardan, T. Changes of superoxide dismutase, catalase and glutathione peroxidase in the corneal epithelium after UVB rays. Histochemical and biochemical study. *Histol. Histopathol.* **2000**, *15*, 1043–1050.
109. Gupta, P.K.; Berdahl, J.P.; Chan, C.C.; Rocha, K.M.; Yeu, E.; Ayres, B.; Farid, M.; Lee, W.B.; Beckman, K.A.; Kim, T.; et al. The corneal endothelium: Clinical review of endothelial cell health and function. *J. Cataract. Refract. Surg.* **2021**, *47*, 1218–1226.

110. Parekh, M.; Ferrari, S.; Sheridan, C.; Kaye, S.; Ahmad, S. Concise Review: An Update on the Culture of Human Corneal Endothelial Cells for Transplantation. *Stem Cell Transl. Med.* **2016**, *5*, 258–264.
111. Feizi, S. Corneal endothelial cell dysfunction: Etiologies and management. *Adv. Ophthalmol.* **2018**, *10*, 2137551084.
112. Perone, J.M.; Boiche, M.; Lhuillier, L.; Ameloot, F.; Premy, S.; Jeancolas, A.; Goetz, C.; Neiter, E. Correlation Between Postoperative Central Corneal Thickness and Endothelial Damage After Cataract Surgery by Phacoemulsification. *Cornea* **2018**, *37*, 587–590.
113. Holst, A.; Rolfsen, W.; Svensson, B.; Ollinger, K.; Lundgren, B. Formation of free radicals during phacoemulsification. *Curr. Eye Res.* **1993**, *12*, 359. [[CrossRef](#)]
114. Takahashi, H. Corneal Endothelium and Phacoemulsification. *Cornea* **2016**, *35*, 3–7.
115. Price, M.O.; Mehta, J.S.; Jurkunas, U.V.; Price, F.W. Corneal endothelial dysfunction: Evolving understanding and treatment options. *Prog. Retin. Eye Res.* **2021**, *82*, 100904. [[PubMed](#)]
116. Igarashi, T.; Ohsawa, I.; Kobayashi, M.; Umemoto, Y.; Arima, T.; Suzuki, H.; Igarashi, T.; Otsuka, T.; Takahashi, H. Effects of Hydrogen in Prevention of Corneal Endothelial Damage During Phacoemulsification: A Prospective Randomized Clinical Trial. *Am. J. Ophthalmol.* **2019**, *207*, 10–17. [[PubMed](#)]
117. Moreau, K.L.; King, J.A. Protein misfolding and aggregation in cataract disease and prospects for prevention. *Trends Mol. Med.* **2012**, *18*, 273–282.
118. Cicinelli, M.V.; Buchan, J.C.; Nicholson, M.; Varadaraj, V.; Khanna, R.C. Cataracts. *Lancet* **2023**, *401*, 377–389.
119. Sunkireddy, P.; Jha, S.N.; Kanwar, J.R.; Yadav, S.C. Natural antioxidant biomolecules promises future nanomedicine based therapy for cataract. *Colloids Surf. B Biointerfaces* **2013**, *112*, 554–562.
120. Varma, S.D.; Chand, D.; Sharma, Y.R.; Kuck, J.J.F.; Richards, R.D. Oxidative stress on lens and cataract formation: Role of light and oxygen. *Curr. Eye Res.* **1984**, *3*, 35.
121. Peng, J.; Zheng, T.; Liang, Y.; Duan, L.; Zhang, Y.; Wang, L.; He, G.; Xiao, H.P. Coumaric Acid Protects Human Lens Epithelial Cells against Oxidative Stress-Induced Apoptosis by MAPK Signaling. *Oxid. Med. Cell. Longev.* **2018**, *2018*, 1–7.
122. Krishna, U.; Ajanaku, D.; Denniston, A.K.; Gkika, T. Uveitis: A sight-threatening disease which can impact all systems. *Postgrad. Med. J.* **2017**, *93*, 766–773.
123. Ung, L.; Pattamatta, U.; Carnt, N.; Wilkinson-Berka, J.L.; Liew, G.; White, A.J.R. Oxidative stress and reactive oxygen species: A review of their role in ocular disease. *Clin. Sci.* **2017**, *131*, 2865–2883.
124. Hartong, D.T.; Berson, E.L.; Dryja, T.P. Retinitis pigmentosa. *Lancet* **2006**, *368*, 1795–1809. [[PubMed](#)]
125. Campochiaro, P.A.T.A. The mechanism of cone cell death in Retinitis Pigmentosa. *Prog. Retin. Eye Res.* **2018**, *62*, 24–37.
126. Zheng, Z.; Chen, H.; Ke, G.; Fan, Y.; Zou, H.; Sun, X.; Gu, Q.; Xu, X.; Ho, P.C.P. Protective Effect of Perindopril on Diabetic Retinopathy Is Associated with Decreased Vascular Endothelial Growth Factor-to-Pigment Epithelium-Derived Factor Ratio. *Diabetes* **2009**, *58*, 954–964.
127. Forbes, J.M.; Coughlan, M.T.; Cooper, M.E. Oxidative Stress as a Major Culprit in Kidney Disease in Diabetes. *Diabetes* **2008**, *57*, 1446–1454.
128. Guma, M.; Rius, J.; Duong-Polk, K.X.; Haddad, G.G.; Lindsey, J.D.; Karin, M. Genetic and pharmacological inhibition of JNK ameliorates hypoxia-induced retinopathy through interference with VEGF expression. *Proc. Natl. Acad. Sci. USA* **2009**, *106*, 8760–8765. [[CrossRef](#)]
129. Dong, A.; Xie, B.; Shen, J.; Yoshida, T.; Yokoi, K.; Hackett, S.F.; Campochiaro, P.A. Oxidative stress promotes ocular neovascularization. *J. Cell Physiol.* **2009**, *219*, 544–552.
130. Anderson, R.E.; Rapp, L.M.; Wiegand, R.D. Lipid peroxidation and retinal degeneration. *Curr. Eye Res.* **1984**, *3*, 223–227.
131. Kowluru, R.A.; Tang, J.; Kern, T.S. Abnormalities of retinal metabolism in diabetes and experimental galactosemia. VII. Effect of long-term administration of antioxidants on the development of retinopathy. *Diabetes* **2001**, *50*, 1938–1942. [[CrossRef](#)] [[PubMed](#)]
132. Kern, T.S.; Kowluru, R.A.; Engerman, R.L. Abnormalities of retinal metabolism in diabetes or galactosemia: ATPases and glutathione. *Investig. Ophthalmol. Vis. Sci.* **1994**, *35*, 2962–2967.
133. Kowluru, R.A.; Abbas, S.N. Diabetes-induced mitochondrial dysfunction in the retina. *Investig. Ophthalm. Vis. Sci.* **2003**, *44*, 5327. [[CrossRef](#)] [[PubMed](#)]
134. Ellis, E.A.; Guberski, D.L.; Somogyi-Mann, M.; Grant, M.B. Increased H₂O₂, vascular endothelial growth factor and receptors in the retina of the BBZ/WOR diabetic rat. *Free Radic. Biol. Med.* **2000**, *28*, 91–101. [[CrossRef](#)]
135. Wohaieb, S.A.; Godin, D.V. Alterations in Free Radical Tissue-Defense Mechanisms in Streptozocin-Induced Diabetes in Rat: Effects of Insulin Treatment. *Diabetes* **1987**, *36*, 1014–1018. [[CrossRef](#)] [[PubMed](#)]
136. Weinreb, R.N.; Aung, T.; Medeiros, F.A. The Pathophysiology and Treatment of Glaucoma: A Review. *JAMA J. Am. Med. Assoc.* **2014**, *311*, 1901–1911. [[CrossRef](#)] [[PubMed](#)]
137. Weinreb, R.N. Glaucoma neuroprotection: What is it? Why is it needed? *Can. J. Ophthalmol.* **2007**, *42*, 396–398. [[CrossRef](#)]
138. Dong, Z.; Shinmei, Y.; Dong, Y.; Inafuku, S.; Fukuhara, J.; Ando, R.; Kitaichi, N.; Kanda, A.; Tanaka, K.; Noda, K.; et al. Effect of geranylgeranylacetone on the protection of retinal ganglion cells in a mouse model of normal tension glaucoma. *Heliyon* **2016**, *2*, e00191. [[PubMed](#)]
139. Manassakorn, A.; Aupapong, S. Retinal nerve fiber layer defect patterns in primary angle-closure and open-angle glaucoma: A comparison using optical coherence tomography. *Jpn. J. Ophthalmol.* **2011**, *55*, 28–34. [[CrossRef](#)]
140. Malishevskaya, T.N.; Yusupov, A.R.; Shatskikh, S.V.; Antipina, N.A.; Klindyuk, T.S.; Bogdanova, D.S. Efficacy and safety of neuroprotection in patients with primary open-angle glaucoma. *Vestn. Oftal'mol.* **2019**, *135*, 83. [[CrossRef](#)]

141. Liu, M.; Li, H.; Yang, R.; Ji, D.; Xia, X. GSK872 and necrostatin-1 protect retinal ganglion cells against necroptosis through inhibition of RIP1/RIP3/MLKL pathway in glutamate-induced retinal excitotoxic model of glaucoma. *J. Neuroinflamm.* **2022**, *19*, 262.
142. Izzotti, A.; Bagnis, A.; Sacca, S. The role of oxidative stress in glaucoma. *Mutat. Res./Rev. Mutat. Res.* **2006**, *612*, 105–114. [[CrossRef](#)] [[PubMed](#)]
143. Tao, Y.; Geng, L.; Xu, W.; Qin, L.; Peng, G.; Huang, Y. The potential utilizations of hydrogen as a promising therapeutic strategy against ocular diseases. *Clin. Risk Manag.* **2016**, *12*, 799–806.
144. Li, G.; Osborne, N.N. Oxidative-induced apoptosis to an immortalized ganglion cell line is caspase independent but involves the activation of poly(ADP-ribose)polymerase and apoptosis-inducing factor. *Brain Res.* **2008**, *1188*, 35–43. [[CrossRef](#)]
145. Tan, W.; Zou, J.; Yoshida, S.; Jiang, B.; Zhou, Y. The Role of Inflammation in Age-Related Macular Degeneration. *Int. J. Biol. Sci.* **2020**, *16*, 2989–3001. [[CrossRef](#)]
146. Feher, J.; Kovacs, I.; Artico, M.; Cavallotti, C.; Papale, A.; Balacco Gabrieli, C. Mitochondrial alterations of retinal pigment epithelium in age-related macular degeneration. *Neurobiol. Aging* **2006**, *27*, 983–993.
147. Plafker, S.M.; O’Mealey, G.B.; Szveda, L.I. Mechanisms for countering oxidative stress and damage in retinal pigment epithelium. *Int. Rev. Cell Mol. Biol.* **2012**, *298*, 135–177. [[PubMed](#)]
148. Tonade, D.; Kern, T.S. Photoreceptor cells and RPE contribute to the development of diabetic retinopathy. *Prog. Retin. Eye Res.* **2021**, *83*, 100919.
149. Kaarniranta, K.; Uusitalo, H.; Blasiak, J.; Felszeghy, S.; Kannan, R.; Kauppinen, A.; Salminen, A.; Sinha, D.; Ferrington, D. Mechanisms of mitochondrial dysfunction and their impact on age-related macular degeneration. *Prog. Retin. Eye Res.* **2020**, *79*, 100858.
150. Tao, Y.; Jiang, P.; Wei, Y.; Wang, P.; Sun, X.; Wang, H. α -Lipoic Acid Treatment Improves Vision-Related Quality of Life in Patients with Dry Age-Related Macular Degeneration. *Tohoku J. Exp. Med.* **2016**, *240*, 209–214. [[CrossRef](#)]
151. Marie, M.; Bigot, K.; Angebault, C.; Barrau, C.; Gondouin, P.; Pagan, D.; Fouquet, S.; Villette, T.; Sahel, J.; Lenaers, G.; et al. Light action spectrum on oxidative stress and mitochondrial damage in A2E-loaded retinal pigment epithelium cells. *Cell Death Dis.* **2018**, *9*, 287.
152. Alaimo, A.; Liñares, G.G.; Bujjamer, J.M.; Gorojod, R.M.; Alcon, S.P.; Martínez, J.H.; Baldessari, A.; Grecco, H.E.; Kotler, M.L. Toxicity of blue led light and A2E is associated to mitochondrial dynamics impairment in ARPE-19 cells: Implications for age-related macular degeneration. *Arch. Toxicol.* **2019**, *93*, 1401–1415. [[PubMed](#)]
153. Park, S.; Jang, Y.P. The Protective Effect of Brown-, Gray-, and Blue-Tinted Lenses against Blue LED Light-Induced Cell Death in A2E-Laden Human Retinal Pigment Epithelial Cells. *Ophthalmic Res.* **2017**, *57*, 118–124. [[CrossRef](#)] [[PubMed](#)]
154. Zha, X.; Wu, G.; Zhang, H.; Yang, Y.; Zhang, Y.; Ma, L. PRDX6 regulates the H₂O₂ and blue light-induced APRE-19 cell apoptosis via down-regulating and interacting with RARA. *Anim. Cells Syst.* **2019**, *23*, 241–245. [[CrossRef](#)]
155. Bien, A.; Seidenbecher, C.I.; Bockers, T.M.; Sabel, B.A.; Kreutz, M.R. Apoptotic versus necrotic characteristics of retinal ganglion cell death after partial optic nerve injury. *J. Neurotraum* **1999**, *16*, 153–163. [[CrossRef](#)]
156. Kreutz, M.R.; Seidenbecher, C.I.; Sabel, A.B.A. Molecular plasticity of retinal ganglion cells after partial optic nerve injury. *Restor. Neurol. Neurosci.* **1999**, *14*, 127–134. [[PubMed](#)]
157. Kreutzberg, G.W. Microglia: A sensor for pathological events in the CNS. *Trends Neurosci.* **1996**, *19*, 312–318. [[CrossRef](#)]
158. Li, H.; Luo, Y.; Yang, P.; Liu, J. Hydrogen as a complementary therapy against ischemic stroke: A review of the evidence. *J. Neurol. Sci.* **2019**, *396*, 240–246. [[CrossRef](#)]
159. Yoritaka, A.; Kobayashi, Y.; Hayashi, T.; Saiki, S.; Hattori, N. Randomized double-blind placebo-controlled trial of hydrogen inhalation for Parkinson’s disease: A pilot study. *Neurol. Sci.* **2021**, *42*, 4767–4770. [[CrossRef](#)]
160. Nie, C.; Ding, X.; Rong, A.; Zheng, M.; Li, Z.; Pan, S.; Yang, W. Hydrogen gas inhalation alleviates myocardial ischemia-reperfusion injury by the inhibition of oxidative stress and NLRP3-mediated pyroptosis in rats. *Life Sci.* **2021**, *272*, 119248. [[CrossRef](#)]
161. Lazar, H.L. Molecular hydrogen: A novel therapy for the treatment of pulmonary hypertension. *J. Thorac. Cardiovasc. Surg.* **2015**, *150*, 654–655. [[CrossRef](#)] [[PubMed](#)]
162. Jackson, K.; Dressler, N.; Ben-Shushan, R.S.; Meerson, A.; LeBaron, T.W.; Tamir, S. Effects of alkaline-electrolyzed and hydrogen-rich water, in a high-fat-diet nonalcoholic fatty liver disease mouse model. *World J. Gastroenterol. WJG* **2018**, *24*, 5095–5108. [[PubMed](#)]
163. Koyama, Y.; Taura, K.; Hatano, E.; Tanabe, K.; Yamamoto, G.; Nakamura, K.; Yamanaka, K.; Kitamura, K.; Narita, M.; Nagata, H.; et al. Effects of oral intake of hydrogen water on liver fibrogenesis in mice. *Hepatol. Res.* **2014**, *44*, 663–677. [[PubMed](#)]
164. Najmanová, E.; Manethová, H.; Botek, M.; Pluháček, F. Vliv Rychlého Příjmu Vodíkem Sycené Vody na Nitrooční Tlak u Zdravých Osob. *Cesk. Slov. Oftalmol.* **2023**, *79*, 180–184.
165. Trachootham, D.; Alexandre, J.; Huang, P. Targeting cancer cells by ROS-mediated mechanisms: A radical therapeutic approach? *Nat. Rev. Drug Discov.* **2009**, *8*, 579–591.

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