RHEUMATOLOGY

Concise report

Identification of a shared genetic risk locus for Kawasaki disease and immunoglobulin A vasculitis by a cross-phenotype meta-analysis

Elio G. Carmona^{1,2}, Jose A. García-Giménez², Raquel López-Mejías³, Chiea Chuen Khor⁴, Jong-Keuk Lee⁵, Ekim Taskiran⁶, Seza Ozen⁷, Alojzija Hocevar⁸, Lili Liu⁹, Mario Gorenjak¹⁰, Uroš Potočnik¹⁰, Krzysztof Kiryluk⁹, Norberto Ortego-Centeno^{11,12}, María C. Cid¹³, José Hernández-Rodríguez¹³, Santos Castañeda¹⁴, Miguel A. González-Gay³, David Burgner^{15,16,17,18}, Javier Martín² and Ana Márquez^{1,2}; Spanish IgA Vasculitis Consortium;* International Kawasaki Disease Genetics Consortium*

Abstract

Objectives. Combining of genomic data of different pathologies as a single phenotype has emerged as a useful strategy to identify genetic risk loci shared among immune-mediated diseases. Our study aimed to increase our knowledge of the genetic contribution to Kawasaki disease (KD) and IgA vasculitis (IgAV) by performing the first comprehensive large-scale analysis on the genetic overlap between them.

Methods. A total of 1190 vasculitis patients and 11 302 healthy controls were analysed. First, in the discovery phase, genome-wide data of 405 KD patients and 6252 controls and 215 IgAV patients and 1324 controls, all of European origin, were combined using an inverse variance meta-analysis. Second, the top associated polymorphisms were selected for replication in additional independent cohorts (570 cases and 3726 controls). Polymorphisms with *P*-values $\leq 5 \times 10^{-8}$ in the global IgAV-KD meta-analysis were considered as shared genetic risk loci.

Results. A genetic variant, rs3743841, located in an intron of the *NAGPA* gene, reached genome-wide significance in the cross-disease meta-analysis ($P = 8.06 \times 10^{-10}$). Additionally, when IgAV was individually analysed, a strong association between rs3743841 and this vasculitis was also evident [$P = 1.25 \times 10^{-7}$; odds ratio = 1.47 (95% CI 1.27, 1.69)]. *In silico* functional annotation showed that this polymorphism acts as a regulatory variant modulating the expression levels of the *NAGPA* and *SEC14L5* genes.

Conclusion. We identified a new risk locus with pleiotropic effects on the two childhood vasculitides analysed. This locus represents the strongest non-HLA signal described for IgAV to date.

Key words: IgA vasculitis, Kawasaki disease, genome-wide association study, single-nucleotide polymorphism

¹Unidad de Enfermedades Autoinmunes Sistémicas, Hospital Clínico San Cecilio, Instituto de Investigación Biosanitaria de Granada ibs.GRANADA, ²Instituto de Parasitología y Biomedicina ¹López-Neyra', CSIC, PTS Granada, Granada, ³Epidemiology, Genetics and Atherosclerosis Research Group on Systemic Inflammatory Diseases, Hospital Universitario Marqués de Valdecilla, IDIVAL, Santander, Spain, ⁴Genome Institute of Singapore, Singapore, ⁵Asan Institute for Life Sciences, University of Ulsan College of Medicine, Seoul, Korea, ⁶Department of Medical Genetics, Faculty of Medicine, Hacettepe University, ⁷Department of Paediatrics, Division of Rheumatology, Hacettepe University Faculty of Medicine, Ankara, Turkey, ⁸Department of Rheumatology, University Medical Centre Ljubljana, Ljubljana, Slovenia, ⁹Division of Nephrology, Department of Medicine, Vagelos College of Physicians and Surgeons, Columbia University, New York, NY, USA, ¹⁰Centre for Human Molecular Genetics and Pharmacogenomics, Faculty of Medicine, Linversity of Marithor, Slovenia, ⁹Divenia

Faculty of Medicine, University of Maribor, Maribor, Slovenia, ¹¹Systemic Autoimmune Diseases Unit, Hospital Universitario San Cecilio¹²School of Medicine, University of Granada, Instituto de Investigación Biosanitaria de Granada ibs.GRANADA, Granada, ¹³Department of Autoimmune Diseases, Hospital Clinic, IDIBAPS, University of Barcelona, Barcelona, ¹⁴Rheumatology Division, Hospital de La Princesa, IIS-Princesa, Universidad Autónoma de Madrid, Madrid, Spain, ¹⁵Murdoch Children's Research Institute, Royal Children's Hospital, ¹⁶Department of Paediatrics, University of Melbourne, ¹⁷Department of General Medicine, Royal Children's Hospital, Parkville and ¹⁸Department of Paediatrics, Monash University, Clayton, Victoria, Australia

Submitted 5 March 2021; accepted 12 May 2021

Correspondence to: Ana Márquez, Instituto de Parasitología y Biomedicina 'López-Neyra', Parque Tecnológico Ciencias de la Salud, Avda del Conocimiento 17, 18016 Armilla, Granada, Spain. E-mail: anamaort@ipb.csic.es

*See acknowledgements section for a list of the members of the International Kawasaki Disease Genetics Consortium and the Spanish IgA Vasculitis Consortium who contributed to this study.

Rheumatology key messages

- The rs3743841 genetic variant has a pleiotropic effect on IgA vasculitis and Kawasaki disease.
- This locus represents the strongest non-HLA signal described for IgA vasculitis to date.
- The rs3743841 polymorphism affects expression levels of the NAGPA and SEC14L5 genes.

Introduction

Systemic vasculitis comprises a heterogeneous group of chronic diseases characterized by arterial wall inflammation. Vasculitis can be classified pathologically into three main categories by vessel size: large (LVV), medium (MVV) and small vessel vasculitis (SVV) [1]. However, clinical phenotypes of the different forms of vasculitis seem to be independent of the diameter of the vessels and, indeed, some of these diseases can affect vessels of variable types and sizes.

Kawasaki disease (KD), an MVV, and IgA vasculitis (IgAV), an SVV, are the first and second most common paediatric vasculitides in European countries, respectively [2, 3]. KD predominantly affects children <5 years of age, with an increased incidence in Asian countries. It is characterized by fever, rash, redness and edema of the hands and feet, mucosal changes, conjunctival injection, lymphadenopathy and, in some cases, arthritis [2]. IgAV largely affects children, but it also occurs in adults. The main features include palpable purpura, gastrointestinal pain and bleeding, arthritis and glomerulonephritis [3].

In both diseases, the aetiology is unknown and the heightened inflammatory pathogenesis is driven by environmental and genetic factors. In the last decade, genome-wide association studies (GWASs) have revolutionized the study of the genetics of systemic vasculitis, including IgAV and KD [4]. In this regard, several genetic risk loci related to immune responses and regulation have shown robust associations with KD, such as *ITPKC*, *FCGR2A*, *CASP3*, *BLK* and *CD40* [5], whereas only the HLA region has been consistently associated with IgAV to date [6].

However, the low prevalence of systemic vasculitis and the moderate effect conferred by diseaseassociated variants mean GWASs require large sample sizes for adequate power to meet the accepted standards for genome-wide significance [7]. In this regard, the combined analysis of genomic data from rare phenotypes of similar pathophysiology allows a significant increase in sample size and may identify common genetic determinants indicative of shared pathogenesis. This approach, called cross-phenotype meta-analysis, has been successfully applied to the study of the genetic component shared across different vasculitides, including giant cell arteritis, Takayasu's arteritis, ANCAassociated vasculitis and IgAV [8, 9], suggesting that common molecular mechanisms can be implicated in these diseases.

Interestingly, coexistence of both IgAV and KD has been described in several patients [10–12]. Moreover, additional evidence points to the existence of shared pathogenic mechanisms between both disorders. In this regard, following the identification of a murine model of KD showing deposition of IgA–C3 immune complexes in cardiovascular lesions and kidneys, it has been proposed that KD may be a form of IgAV [13].

Considering this, the aim of this study was to investigate, for the first time, the common genetic components of two forms of paediatric vasculitis by performing a cross-disease meta-analysis of KD and IgAV GWAS data.

Materials and methods

Study design

This study was conducted in two stages: a discovery stage, in which IgAV and KD GWAS data from previously published studies [14, 15] were combined as a single phenotype, with the aim of identifying potential new susceptibility variants for both vasculitides, and a validation stage, in which the most associated single nucleotide polymorphisms (SNPs) in the cross-phenotype metaanalysis were tested in additional independent cohorts. A summarized workflow of the study can be found in **Supplementary Fig. S1**, available at *Rheumatology* online.

Approval from the local ethical committees of the different participating centres and informed written consent from all participants were obtained in accordance with the tenets of the Declaration of Helsinki.

Discovery stage

Study population

Genomic data from a total of 620 paediatric cases and 7576 healthy controls of European descent, including 215 IgAV patients and 1324 controls from Spain and 405 KD patients and 6252 controls from Australia, The Netherlands, the UK and the USA, were included in the cross-disease meta-analysis. A table summarizing the analysed cohorts is available in Supplementary Table S1, available at *Rheumatology* online.

Quality control and imputation

GWAS data were filtered prior to imputation using PLINK version 1.9 software [16]. SNPs were removed based on a low genotyping rate (<95%), low allele

frequency (minor allele frequency <1%) and deviation from Hardy–Weinberg equilibrium (P < 0.001). In addition, individuals with successful call rates <95% were discarded, as well as duplicates and first-degree relatives.

SNP genotype imputation was performed using the Michigan Imputation Server (https://imputationserver. sph.umich.edu), with phase 3 of the 1000 Genomes Project as the reference panel and a probability threshold for merging genotypes of 0.9. Imputed data were subsequently filtered as described above. Additionally, principal component analysis was performed with PLINK version 1.9 and GCTA (Genome-wide Complex Trait Analysis) software [17], using the first 10 principal components to detect possible biased associations by ancestry. Individuals who surpassed 4 standard deviations from the centroid were excluded by considering them as outliers. Principal component plots are shown in Supplementary Fig. S2, available at *Rheumatology* online.

Statistical analysis

PLINK version 1.9 was used to perform logistic regression analysis on each individual disease, including the first 10 principal components and sex as covariates. Quantile–quantile plots are shown in Supplementary Fig. S3, available at *Rheumatology* online. Summary statistics of each disease were then meta-analysed by applying the inverse variance method, to identify shared genetic variants with the same effect in both diseases. In addition, to identify genetic variants with opposite allelic effects, the direction of association was flipped, that is, 1/OR instead of OR, in the IgAV dataset before the KD–IgAV meta-analysis.

SNPs that met our criteria (P < 0.01 in each individual disease and $P < 5 \times 10^{-5}$ in the cross-disease metaanalysis) were selected for the validation stage.

Validation stage

Study population and data sources

A total of 570 cases and 3726 healthy controls, including 186 KD patients and 600 controls from Korea and 384 IgAV patients and 3126 controls from four independent cohorts (Spain, Turkey, Slovenia and the USA) were analysed (Supplementary Table S1, available at *Rheumatology* online).

For the KD validation cohort, summary statistics of the selected SNPs were obtained from a previously published GWAS [18]. In the case of the IgAV validation cohorts, genotyping data from part of the samples (cases and controls from the USA and controls from Slovenia) were obtained using SNP genotyping arrays, while data from the remaining samples (cases from Slovenia and cases and controls from Spain and Turkey) were genotyped for the selected SNPs by allelic discrimination assay using TaqMan probes (Applied Biosystems, Waltham, MA, USA) (Supplementary Table S1, available at *Rheumatology* online).

Statistical analysis

Each independent cohort was individually analysed by logistic regression analysis. Subsequently, KD and IgAV cohorts were combined using the inverse variance method. A fixed effects model was used in the absence of heterogeneity across studies. Instead, when heterogeneity was detected, a random effects approach (RE2), implemented in METASOFT (http://genetics.cs.ucla.edu/ meta/), was applied. After meta-analysis, those SNPs reaching a genome-wide significance level ($P \le 5 \times 10^{-8}$) in the cross-disease meta-analysis, including both discovery and validation cohorts, and showing disease-specific *P*-values ≤ 0.05 in the validation stage were considered as common susceptibility loci for IgAV and KD.

In silico functional characterization

To evaluate the potential functional role of the most strongly associated polymorphism of each shared locus, and their proxies ($r^2 > 0.8$), publicly available functional annotation data were assessed. Specifically, data from Haploreg version 4.1 (https://pubs.broadinstitute.org/mammals/haploreg/haploreg.php) and the GTEx (https://gtexportal.org) and ENCODE (https://www.encodepro ject.org/) projects were explored in order to determine whether associated SNPs overlapped with published expression quantitative trait loci (eQTLs), regulatory chromatin marks, DNase I hypersensitive sites (DHSs) and/or transcription factor binding sites (TFBSs) in immune cell lines and/or whole blood.

Results

Meta-analysis

Discovery stage

After quality control, 617 cases (215 IgAV cases and 402 KD cases) and 7538 controls were available for analysis. A total of 5 690 841 SNPs that overlapped between the discovery datasets were included in the cross-disease meta-analysis (Fig. 1).

After combined analysis of IgAV and KD GWAS data, assuming same allelic effects in both diseases, genetic variants at seven loci satisfied the established criteria for the discovery phase ($P < 5 \times 10^{-5}$ in the combined analysis; P < 0.01 in each independent disease analysis) (Table 1 and Fig. 1). Additionally, when taking into consideration potential opposite allele effects in both diseases, an additional SNP (rs2659608) fulfilled the established criteria (Table 1 and Fig. 1).

Validation stage

Subsequently the top signals within each locus were analysed in independent additional cohorts (Supplementary Table 1, available at *Rheumatology* online). As shown in Table 1, rs3743841, located in an intron of the *NAGPA* (N-acetylglucosamine-1-phosphodiester alpha-N-acetylglucosaminidase) gene, reached genome-wide significance in the IgAV-KD meta-analysis, including both



Fig. 1 Manhattan plot representing the results of the cross-disease meta-analysis for the discovery phase

The $-\log_{10}$ of the *P*-values are plotted against their physical chromosomal position. The red line represents the genome-wide level of significance ($P < 5 \times 10^{-8}$). The established significance threshold for the discovery stage ($P < 5 \times 10^{-5}$) is highlighted in blue. Loci have been annotated only for those polymorphisms reaching the established criteria for the discovery phase ($P < 5 \times 10^{-5}$ in the combined analysis; P < 0.01 in each independent disease analysis). Ochre dots represent those signals showing opposite allelic effect in both vasculitides.

discovery and replication datasets ($P = 8.06 \times 10^{-10}$) (Supplementary Fig. S4, available at *Rheumatology* online). Furthermore, this polymorphism showed statistical significance in the IgAV replication analysis [$P = 5.48 \times 10^{-4}$; OR = 1.39 (95% CI 1.15, 1.68)], although no association was observed in the KD replication cohort [P = 0.76; OR = 1.04 (95% CI 0.82, 1.31)] (Table 1).

It should be noted that the rs3743841 variant was near genome-wide significant in the IgAV-only metaanalysis [$P = 1.25 \times 10^{-7}$; OR = 1.47 (95% CI 1.27, 1.69)], including both discovery and replication cohorts. The minor allele frequency of rs3743841 in each of the study cohorts is shown in Supplementary Table S2, available at *Rheumatology* online.

Functional annotation

In order to provide biological context to our findings, we evaluated putative regulatory effects of the rs3743841 polymorphism, using different functional annotation databases. The rs3743841 genetic variant, and its only proxy ($r^2 = 0.99$), appeared to act as eQTLs, affecting expression levels of the *NAGPA* (3.89 × 10⁻²³) and *SEC14L5* (5.21 × 10⁻⁸) genes in whole blood [19] and in whole blood and monocytes [20], respectively.

Discussion

In the present study, we report the first large-scale cross-disease meta-analysis of IgAV and KD, identifying a potential novel risk locus shared between both paediatric diseases. Specifically, the rs3743841 variant, located in an intron of the *NAGPA* gene, reached genome-wide significance in the overall cross-disease meta-analysis. This suggests a possible shared genetic component and, presumably, common molecular mechanisms involved in the development of both forms of vasculitis. In addition, the *NAGPA* variant showed a strong association with IgAV ($P = 1.25 \times 10^{-7}$; OR = 1.47), thus representing the strongest non-HLA signal described for this disease to date.

Interestingly, in silico functional analysis showed that this polymorphism, and its only proxy, acts as an eQTL, altering NAGPA and SEC14L5 gene expression levels in whole blood and monocytes. SEC14L5 encodes a member of the SEC14 family of proteins, which drives interactions between proteins and phospholipids that are essential for protein targeting, signal transduction, lipid metabolism and transport and compartment maintenance [21]. The NAGPA gene is involved in mannose 6phosphate formation and encodes the uncovering enzyme that catalyzes the second step in the formation of mannose 6-phosphate, a recognition marker on lysosomal hydrolases. It has been reported that inactivation of the NAGPA gene in mice results in elevated levels of several acid hydrolases in plasma, which showed a decreased affinity for the cation-independent mannose 6-phosphate receptor and failed to bind to the cationdependent mannose 6-phosphate receptor [22]. Interestingly, a central role of the lysosomal compartment in regulating different processes of the immune response, such as secretion of inflammatory cytokines or antigen presentation, has been described [23]. Furthermore, lysosomal enzyme activity may be involved in autoantigen generation and, indeed, changes in lysosomal hydrolases have been implicated in different autoimmune diseases [23]. Finally, lysosomes have also been implicated in other vasculitides, specifically in

-
S
Ω.
2
g
a
4
ž
æ
¥
4
∢
ත
<u>م</u>
Ĕ
Ξ
⊒.
σ
ŝ
0
ő
ğ
ő
.≅
2
g
Ę
÷
÷≓
2
-=
눈
ы
õ
۲
4
ŝ
ě
ъ
ъ
Ē
ō
Ë
8
÷
ŝ
ĥ
Ĕ
g
\geq
ē
2
8
<u>.</u>
σ
Φ
다
Ē
-=
ő
Ĕ
aji.
đ
6
õ
Ë
ß
ĕ
2
2
ĭ
<u>ש</u>
g
S
ŝ
∢
-
ш
Balled
닕

3p13.3

Region	Gene	SNP	Test allele			Discovery				Valid	ation		Discovery + validation
					IgAV		KD	IgAV-KD		IgAV		KD	lgAV-KD
				P-value	OR (95% CI)	P-value	OR (95% CI)	<i>P</i> -value	P-value	OR (95% CI)	P-value	OR (95% CI)	P-value
16p13.3	NAGPA	rs3743841	 -	4.32E-05	1.58 (1.27, 1.97)	6.08E-05	1.39 (1.18, 1.63)	1.61E-08	5.48E-04	1.39 (1.15, 1.68)	0.76	1.04 (0.82, 1.31)	8.06E-10 [*]
5q35.3	GRM6	rs67428898	F	2.70E-05	1.61 (1.29, 2.01)	2.30E-03	1.28 (1.09, 1.50)	8.64E-07	7.63E-01	0.96 (0.71, 1.28)	0.37	1.12 (0.88, 1.42)	1.61E-05 [*]
18p11.31	EPB41L3	rs73379591	с U	4.52E-03	0.66 (0.49, 0.88)	8.27E-05	0.63 (0.50, 0.79)	1.25E-06	3.95E-03	0.69 (0.53, 0.89)	0.86	0.96 (0.61, 1.50)	6.90E-08
3p25.1	TPRXL	rs2733560	വ	1.00E-04	1.52 (1.23, 1.87)	2.00E-03	1.27 (1.09, 1.48)	1.70E-06	2.77E-02	0.82 (0.69, 0.98)	0.71	1.04 (0.82, 1.33)	1.48E-04 [*]
11q25	OPCML	rs2659608	с О	1.91E-05	1.6 (1.29, 1.99)	7.22E-03	0.8 (0.69, 0.94)	3.30E-06	6.18E-01	0.95 (0.79, 1.15)	0.69	1.62 (0.15, 17.86)	1.83E-05 [*]
10p14	LINC00707	rs10508313	с U	3.64E-03	1.45 (1.13, 1.87)	2.98E-04	1.43 (1.18, 1.73)	3.53E-06	2.02E-02	1.35 (1.05, 1.75)	0.70	0.95 (0.74, 1.22)	1.17E-07 [*]
5q23.2	I	rs11957435	с U	1.30E-03	1.5 (1.17, 1.91)	1.52E-03	1.34 (1.12, 1.60)	8.16E-06	6.42E-01	0.95 (0.76, 1.19)	0.82	0.97 (0.75, 1.25)	1.18E-04 [*]
17q11.2	ASIC2	rs4426392	G	1.76E-03	0.7 (0.56, 0.87)	1.31E-03	0.77 (0.66, 0.90)	9.57E-06	6.71E-03	0.77 (0.63, 0.93)	0.32	1.13 (0.89, 1.43)	1.65E-05
The polymo	rphism asso.	ciated at the	genon	ne-wide sig	nificance level ir	the IgAV-	-KD meta-analys	is is marke	ed in bold.	* P-values obtair	ted using	a random effects	model (RE2).

ANCA-associated vasculitis, which are characterized by the presence of autoantibodies targeted against lysosomal enzymes of monocytes and neutrophils [24].

Our study has some strengths. First, this work represents a great international collaborative effort that has made possible the analysis of a large cohort of paediatric vasculitis patients using common case definitions. Second, this is the first study comprehensively examining the genetic overlap between IgAV and KD. However, we acknowledge some limitations. Although the rs3743841 variant reached the genome-wide significance level in the cross-disease meta-analysis, it was not statistically significant in the KD validation cohort. One possible explanation could be that this polymorphism is a genetic marker in linkage disequilibrium (LD) with the real causal variant. In this situation, differences in the LD pattern between Europeans and Asians might be responsible for the observed results. Considering this possibility, the associated polymorphism could be targeting the causal variant in European but not in the Asian population. Alternatively, the absence of association observed in the Korean cohort could be due to a lack of statistical power. Nevertheless, the statistical power for this analysis was almost 80%, considering the minor allele frequency reported in East Asian populations in phase 3 of the 1000 Genomes Project (49%) and the effect of the allele risk observed in the discovery phase (OR = 1.39). Therefore it is unlikely that the absence of association observed in the KD replication cohort was due to limited statistical power, although we cannot discard this hypothesis (Supplementary Table S3, available at Rheumatology online). Finally, as shown in Supplementary Table S3, available at Rheumatology online, for all the selected SNPs, the statistical power of the IqAV replication analysis was >80% to detect the effect size observed in the discovery IgAV cohort; however, the KD validation analysis did not reach enough statistical power, except for rs3743841 and rs10508313, and therefore, for the remaining six polymorphisms, the lack of replication in KD could be due to the low statistical power of the analysis.

In summary, in the first combined analysis of IgAV and KD GWAS data, we identified a putative susceptibility risk factor shared between both diseases as well as the strongest non-HLA signal described for IgAV to date. Our results point to a potential role of the lysosomal pathway in the IgAV pathogenesis and suggest that this process could represent a common pathogenic mechanism for these two most common types of paediatric vasculitides.

Acknowledgements

The authors thank Sofía Vargas and Gema Robledo for their excellent technical assistance and all the patients and healthy controls for their essential collaboration. A.M. and J.M. were involved in the conception and design of the study. A.M., E.G.C., E.T. and J.A.G.G. performed the analyses. R.L.M., C.C.K., J.K.L., E.T., S.O., A.H., L.L., M.G., U.P., K.K., N.O.C., M.C.C., J.H.R.,

S.C., M.A.G.G. and D.B. were involved in study subject and data recruitment and participated in interpretation of data. A.M. and E.G.C. drafted the manuscript. All authors critically revised the manuscript.

Other members of the Spanish IqA Vasculitis Consortium who contributed to this work: Fernanda Genre, Sara Remuzgo-Martínez, Begoña Ubilla, Verónica Mijares, Trinitario Pina and Ricardo Blanco (Epidemiology, Genetics and Atherosclerosis Research Group on Svstemic Inflammatory Diseases. Rheumatology Division, Hospital Universitario Marqués de Valdecilla, IDIVAL, Santander, Spain); Belén Sevilla-Perez (Medicine Department, Hospital Universitario San Cecilio, Granada, Spain); Javier Llorca [Epidemiology and Computational Biology Department, School of Medicine. University of Cantabria, and CIBER Epidemiología y Salud Pública (CIBERESP), IDIVAL, Santander, Spain]; José A. Miranda-Filloy (Division of Rheumatology, Hospital Universitario Lucus Augusti, Lugo, Spain); Antonio Navas Parejo (Nephrology Department, Hospital Universitario San Cecilio, Granada, Spain); Diego de Argila and Maximiliano Aragües (Dermatology Department, Hospital Universitario La Princesa, IIS-Princesa, Madrid, Spain); Esteban Rubio and Manuel León Lugue (Rheumatology Department, Hospital Universitario Virgen del Rocío, Sevilla, Spain): Juan María Blanco-Madrigal and Eva Galíndez-Aguirregoikoa (Rheumatology Department, Hospital Universitario de Basurto, Bilbao, Spain); Marc Corbera-Bellalta and Sergio Prieto-González (Department of Autoimmune Diseases. Hospital Clinic. IDIBAPS. University of Barcelona).

Other members of the International Kawasaki Disease Genetics Consortium who contributed to this work: Michael Levin (Department of Pediatrics, Imperial College London, London, UK); Jane C Burns (Department of Pediatrics, University of California San Diego School of Medicine, La Jolla, CA, USA; Department of Pediatrics, Rady Children's Hospital, San Diego, CA, USA); Taco W. Kuijpers (Department of Pediatric Hematology, Immunology and Infectious Diseases, Emma Children's Hospital Academic Medical Center, Amsterdam, The Netherlands; Department of Blood Cell Research, Sanguin Research and Landsteiner Laboratory, University of Amsterdam, Amsterdam, The Netherlands); Martin L. Hibberd (Infectious Diseases, Genome Institute of Singapore, Singapore; Department of Paediatrics, Faculty of Medicine, National University of Singapore, Singapore).

Funding: This work was supported by the Cooperative Research Thematic Network programme (RD16/0012/ 0013 and RD16/0012/0009) from the Instituto de Salud Carlos III (ISCIII, Spanish Ministry of Economy, Industry and Competitiveness). A.M. is a recipient of a Miguel Servet fellowship (CP17/00008) from the ISCIII (Spanish Ministry of Economy, Industry and Competitiveness). R.L.M. is a recipient of a Miguel Servet type I programme fellowship from ISCIII (Spanish Ministry of Economy, Industry and Competitiveness), co-funded by European Social Fund (ESF) ('Investing in your future') (grant CP16/00033).

Disclosure statement: The authors declare no conflicts of interest.

Data availability statement

Results of the KD–IgAV cross-phenotype meta-analysis are available from the corresponding author on reasonable request.

Supplementary data

Supplementary data are available at *Rheumatology* online.

References

- Jennette JC, Falk RJ, Bacon PA *et al.* 2012 revised International Chapel Hill consensus Conference Nomenclature of Vasculitides. Arthritis Rheum 2013;65: 1–11.
- 2 Newburger JW, Takahashi M, Burns JC. Kawasaki disease. J Am Coll Cardiol 2016;67:1738–49.
- 3 González-Gay MA, López-Mejías R, Pina T, Blanco R, Castañeda S. IgA vasculitis: genetics and clinical and therapeutic management. Curr Rheumatol Rep 2018;20: 1–7.
- 4 Ozen S, Batu ED. Vasculitis pathogenesis: can we talk about precision medicine? Front Immunol 2018;9:1–10.
- 5 Kumrah R, Vignesh P, Rawat A, Singh S. Immunogenetics of Kawasaki disease. Clin Rev Allergy Immunol 2020;59:122–39.
- 6 López-Mejías R, Castañeda S, Genre F et al. Genetics of immunoglobulin-A vasculitis (Henoch-Schönlein purpura): an updated review. Autoimmun Rev 2018;17:301–15.
- 7 Tam V, Patel N, Turcotte M *et al.* Benefits and limitations of genome-wide association studies. Nat Rev Genet 2019;20:467–84.
- 8 Ortiz-Fernández L, Carmona FD, López-Mejías R et al. Cross-phenotype analysis of Immunochip data identifies KDM4C as a relevant *locus* for the development of systemic vasculitis. Ann Rheum Dis 2018;77:589–95.
- 9 Carmona FD, Coit P, Saruhan-Direskeneli G et al. Analysis of the common genetic component of largevessel vasculitides through a meta-Immunochip strategy. Sci Rep 2017;7:43953.
- 10 Miura M, Mochizuki T, Fukushima H, Ishihara J. A case of Kawasaki disease accompanied by Henoch-Schönlein purpura. Clin Exp Rheumatol 2004;22:377–8.
- 11 Vedagiriswaran VV, Amperayani S, Ramamoorthy RK, Ranjith MS. A case of Henoch-Schönlein purpura with Kawasaki disease. Indian J Pediatr 2014;81:408–9.
- 12 Heldrich FJ, Jodorkovsky RA, Lake AM, Parnes CA. Kawasaki syndrome: HUS and HSP complicating its course and management. Md Med J 1987;36:764–6.

- 13 Noval Rivas M, Wakita D, Franklin MK *et al.* Intestinal permeability and IgA provoke immune vasculitis linked to cardiovascular inflammation. Immunity 2019;51: 508–521.e6.
- 14 Khor CC, Davila S, Breunis WB *et al.* Genome-wide association study identifies *FCGR2A* as a susceptibility locus for Kawasaki disease. Nat Genet 2011;43: 1241–6.
- 15 López-Mejías R, Carmona FD, Castañeda S et al. A genome-wide association study suggests the HLA Class II region as the major susceptibility locus for IgA vasculitis. Sci Rep 2017;7:5088.
- 16 Purcell S, Neale B, Todd-Brown K et al. PLINK: a tool set for whole-genome association and population-based linkage analyses. Am J Hum Genet 2007;81:559–75.
- 17 Yang J, Lee SH, Goddard ME, Visscher PM. GCTA: a tool for genome-wide complex trait analysis. Am J Hum Genet 2011;88:76–82.
- 18 Kim JJ, Hong YM, Sohn S *et al.* A genome-wide association analysis reveals 1p31 and 2p13.3 as susceptibility loci for Kawasaki disease. Hum Genet 2011;129: 487–95.

- 19 Westra HJ, Peters MJ, Esko T *et al.* Systematic identification of trans eQTLs as putative drivers of known disease associations. Nat Genet 2013;45:1238–43.
- 20 Zeller T, Wild P, Szymczak S *et al.* Genetics and beyond—the transcriptome of human monocytes and disease susceptibility. PLoS One 2010;5: e10693.
- 21 Saito K, Tautz L, Mustelin T. The lipid-binding SEC14 domain. Biochim Biophys Acta Mol Cell Biol Lipids 2007; 1771:719–26.
- 22 Boonen M, Vogel P, Platt KA, Dahms N, Kornfeld S. Mice lacking mannose 6-phosphate uncovering enzyme activity have a milder phenotype than mice deficient for N-acetylglucosamine-1-phosphotransferase activity. Mol Biol Cell 2009;20:4381–9.
- 23 Ge W, Li D, Gao Y, Cao X. The roles of lysosomes in inflammation and autoimmune diseases. Int Rev Immunol 2015;34:415–31.
- 24 Ramponi G, Folci M, De Santis M *et al.* The biology, pathogenetic role, clinical implications, and open issues of serum anti-neutrophil cytoplasmic antibodies. Autoimmun Rev 2021;20:102759.