



In silico prospection of *Lactobacillus acidophilus* strains with potential probiotic activity

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Abstract

Lactic acid bacteria (LAB) are fermentative microorganisms and perform different roles in biotechnological processes, mainly in the food and pharmaceutical industries. Among the LAB, *Lactobacillus acidophilus* is a species that deserves to be highlighted for being used both in prophylaxis and in the treatment of pathologies. Most of the metabolites produced by this species are linked to the inhibition of pathogens. In this study, we utilized a pangenomic and metabolic annotation analysis using Roary and BlastKOALA, ML-based probiotic activity prediction with iProbiotic and whole-genome similarity using ANI to identify strains of *L. acidophilus* with potential probiotic activity. According to the results in BlastKOALA and iProbiotics, *L. acidophilus* NCTC 13721 had the greatest potential among the 64 strains tested, both in terms of its ability to be a *Lactobacillus* spp. probiotic, when in the amount of genes involved in the metabolism of organic acids and quorum sensing. In addition, DSM 20079 proved to be promising for prospecting new probiotic *Lactobacillus* from BlastKOALA analyses, as they presented similar results in the number of genes involved in the production of lactic acid, acetic acid, hydrogen peroxide, except for quorum sensing where the NCTC 13721 strain had 14 more genes. *L. acidophilus* NCTC 13721 and *L. acidophilus* La-5 strains showed greater ability to be *Lactobacillus* spp. probiotic capacity, showing 84.8% and 51.9% capacity in the iProbiotics tool, respectively. When analyzed in ANI, none of the evaluated strains showed genomic similarity with NCTC 13721. In contrast, the DSM 20079 strain showed genomic similarity with all evaluated strains except NCTC 13721. Furthermore, eight strains with characteristics with approximately 100% genomic similarity to La-5 were listed: S20_1, LA-5, FSI4, APC2845, LA-G80-111, DS1_1A, LA1, and BCRC 14065. Therefore, according to the findings in iProbiotics and BlastKoala, among the 64 strains evaluated, NCTC 13721 is the most promising strain to be used for future in vitro studies.

Keywords Bioinformatics · Probiotics · Lactic acid bacteria · *Lactobacillus* · Lactic acid · Pangenome

Introduction

Probiotics are defined as live microorganisms that confer a health benefit on the host if administered in adequate amounts [1]. Among the various probiotics, species belonging to the previously known *Lactobacillus* genus represent

a group of bacteria with heterogeneous characteristics, having the following classification: phylum *Firmicutes*, class *Bacilli*, order *Lactobacillales*, and family *Lactobacillaceae* [2]. They are non-spore-forming, catalase-negative, gram-positive, and facultatively anaerobic rods [3]. It is also important to point out that *Lactobacillus* can ferment carbohydrates into lactic acid in their primary metabolism, which characterizes them as lactic acid bacteria (LAB) [4]. These probiotic microorganisms inhabit the human gastrointestinal and vaginal microbiota and have been gaining prominence due to their beneficial effects on the health of the host, which can occur directly between cells or indirectly through their metabolites [5, 6]. Among the many protective effects of *Lactobacillus* spp. is the modulation of the intestinal microbiota and immune system, the intestinal barrier's reinforcement, and the regulation of crucial pathways in epithelial cells [7].

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Various *L. acidophilus* strains have already been described with probiotic properties [8]. This species contains important probiotics, as it acts in several functions related to the health of the host, mainly in inhibiting pathogenic microorganisms, regulating the intestinal epithelial barrier, and anti-inflammatory effect, increasing its use in the food and pharmaceutical area [9]. In addition, *L. acidophilus* stands out for having characteristics such as resistance to bile salts, low pH, good ability to adhere to human colon cells in cell culture, regulation of host immune responses, and promising in the prophylaxis and treatment of infections. Among its functional mechanisms, the current study has shown that *L. acidophilus* regulates the intestinal microbiota by decreasing pH and producing metabolites [8].

The fermentation process with LAB generates an accumulation of organic acids, having lactic acid as the principal end product of carbohydrate metabolism. The accumulation of this acid, with the consequent reduction in pH, is responsible for a broad-spectrum inhibitory activity for both gram-positive and gram-negative bacteria. When LAB is in the presence of oxygen, hydrogen peroxide (H₂O₂) is produced. On the other hand, superoxide anions in the presence of H₂O₂ form hydroxy radicals, which can lead to peroxidation of membrane lipids and an increase in the membrane. The bactericidal effect obtained from these metabolites is believed to result from the oxidizing effect on the bacterial cell, the destruction of nucleic acids, and cellular proteins [10].

Quorum sensing (QS) is the mechanism through which bacterial cells communicate between and within species after reaching a certain level of cell density [11]. Communication occurs through signaling by autoinducer molecules (AI) produced by cells. QS inhibition appears to be how probiotics modulate the intestinal system and lessen the harmful effects of pathogenic bacteria [12].

Given the need for a better understanding of probiotic strains, using bioinformatics tools is a viable alternative. Bioinformatics is an interdisciplinary area of knowledge that uses tools such as computer science, mathematics, and biology, to promote a greater understanding of biological data, benefiting biomedical research in several aspects to understand the relationship between genes and the system stimulus [13]. Studies on pangenomics have become a powerful tool, as genetic analysis and comparison can be helpful to explore and characterize a shared pattern among microorganisms, providing a better understanding of the function and evolution of genomes. Furthermore, researchers can evaluate specific genomic characteristics to determine and characterize a species with better performance in the production of a particular protein or metabolite, for example [14].

This work aimed to prospect, through bioinformatics, strains of *L. acidophilus* that have potential probiotic activity through pangenomic analysis and predictive models. In addition, we

identified through bioinformatics the number of genes involved in the production of lactic acid, acetic acid, hydrogen peroxide, and quorum sensing related to probiotic activity.

Materials and methods

Data acquisition

Complete genome sequences of *L. acidophilus* strains were obtained from the National Center for Biotechnology Information (NCBI) using the Datasets portal (<https://www.ncbi.nlm.nih.gov/datasets/>). A total of 64 *L. acidophilus* records were retrieved, 13 complete genomes and 51 draft genomes. *L. acidophilus* phage records were excluded, along with records without a FASTA and GBFF (GenBank File Format) file available. Names, accession codes, completion levels, and references of the strains used in the present work are presented in Table 1.

Pangenome analysis

After selecting the *Lactobacillus* strains, the Roary v3.13 software—native to Linux— [26] was used, which receives GFF3 files (General Feature Formats version 3) as input. The present study converted GenBank data to the GFF3 format using BioPerl (<https://bioperl.org/>). It was established as part of the core gene, the genes present in at least 95% of the genomes, and the minimum similarity between two genes must be 70% for them to belong to the same orthologous group. The mentioned software provides several files with statistics of genes shared by a large part of the lineage or throughout (soft core and core genes) and some genomes (accessories, subdivided into cloud and shell genes) [27].

Identification of genes associated with the production of metabolites and probiotic activity

The genes involved in the production of lactic acid were obtained from the KEGG database, which links biological functions. Subsequently, the BlastKOALA [28] program was applied, a tool used for annotation and can identify proteins involved in signal transduction, catabolism, transport, biosynthesis, and glycan metabolism, among other metabolic pathways available from the KEGG database [29]. After the metabolic annotation of the pangenome (protein-coding genes in FASTA format, generated by Roary), the identified results were compared with the EC codes (Enzyme Commission Number) described in the KEGG database as associated with the production of lactic acid, acetic acid, hydrogen peroxide, and quorum sensing. This step used an in-house Python script and the libraries BioPython (<https://biopython.org/>).

Table 1 Identification and access codes of the *L. acidophilus* strains used in the present study

Strain	BioSample	BioProject	Code NCBI	Completion level	Reference
NCFM	SAMN02603047	PRJNA82	GCA_000011985.1	Complete	[15]
FSI4	SAMN03274004	PRJNA271341	GCA_000934625.1	Complete	[16]
LA1	SAMN05631052	PRJNA340059	GCA_002286215.1	Complete	[17]
YT1	SAMN08142761	PRJNA421407	GCA_003952845.1	Complete	-
LA-G80-111	SAMN15165794	PRJNA638040	GCA_013342945.1	Complete	-
NC55	SAMN23011956	PRJNA779097	GCA_020883435.1	Complete	-
5460	SAMN24563600	PRJNA793589	GCA_021432145.1	Complete	-
La-14	SAMN02603216	PRJNA196176	GCF_000389675.2	Complete	[18]
ATCC 53544	SAMN07357495	PRJNA394684	GCF_002224305.1	Complete	-
DSM 20079	SAMN06606133	PRJNA379350	GCF_003047065.1	Complete	-
HN017	SAMN29766956	PRJNA859117	GCF_024397395.1	Complete	-
LA-2	SAMN29862208	PRJNA860779	GCF_024665075.1	Complete	-
LA-5	SAMN29862214	PRJNA860779	GCF_024665555.1	Complete	-
NCTC13721	SAMEA3881062	PRJEB6403	GCA_900452495.1	Draft	-
KLDS 1.0901	SAMN05949236	PRJNA218564	GCF_001868765.1	Draft	[19, 20]
LA1063	SAMN14422845	PRJNA613973	GCF_017009725.1	Draft	[21]
LMG P-21904	SAMN07187785	PRJNA388854	GCF_002914945.1	Draft	-
BCRC 14065	SAMN14363925	PRJNA612162	GCF_017009515.1	Draft	[21]
BCRC 17008	SAMN14371317	PRJNA612399	GCF_017009595.1	Draft	[21]
BCRC 17481	SAMN14371318	PRJNA612401	GCA_017009655.1	Draft	[21]
NBRC 13951	SAMD00046914	PRJDB1353	GCF_001591845.1	Draft	-
La-5	SAMN14401351	PRJNA613347	GCA_017009715.1	Draft	[21]
CIRM-BIA 442	SAMEA2272381	PRJEB1530	GCF_000442865.1	Draft	-
ATCC 4356	SAMN03105773	PRJNA263693	GCA_000786395.1	Draft	[22]
BCRC 16092	SAMN14363928	PRJNA612164	GCA_017009575.1	Draft	[21]
BCRC 16099	SAMN14371312	PRJNA612394	GCA_017009605.1	Draft	[21]
NBIMCC 8242 (180)	SAMN23827470	PRJNA787572	GCF_021229035.1	Draft	-
DSM 20242	SAMEA2272474	PRJEB1533	GCA_000442825.1	Draft	-
QAULAN51	SAMN20114166	PRJNA744373	GCA_022509485.1	Draft	-
s-4	SAMN15579838	PRJNA647640	GCF_013867555.1	Draft	-
BCRC 14079	SAMN14363926	PRJNA612163	GCF_017009475.1	Draft	[21]
CIRM-BIA 445	SAMEA2272655	PRJEB1531	GCA_000469765.1	Draft	-
s-13	SAMN15579847	PRJNA647640	GCF_013867605.1	Draft	-
BCRC 80064	SAMN14371424	PRJNA612405	GCF_017009695.1	Draft	[21]
L3_101_000G1_ dasL3_101_000G1_metabat. metabat.48	SAMN17800807	PRJNA698986	GCA_018367455.1	Draft	[23]
BCRC 17486	SAMN14371319	PRJNA612404	GCF_017009585.1	Draft	[21]
PNW3	SAMN10979321	PRJNA504734	GCA_004348805.1	Draft	-
MGYG-HGUT-02379	SAMEA5851883	PRJEB33885	GCF_902386525.1	Draft	-
DSM 9126	SAMEA2272239	PRJEB1839	GCA_000469745.1	Draft	-
LA_AVK2	SAMN13198280	PRJNA587688	GCF_009741835.1	Draft	-
LA_AVK1	SAMN13198235	PRJNA587652	GCF_009742735.1	Draft	-
BCRC 12255	SAMN14363914	PRJNA612160	GCF_017009485.1	Draft	[21]
DS9_1A	SAMN05583792	PRJNA336518	GCF_003061925.1	Draft	[24]
DS5_1A	SAMN05583788	PRJNA336518	GCF_003061985.1	Draft	[24]
BIO6307	SAMN12856535	PRJNA574342	GCF_008868625.1	Draft	-
DSM 20079	SAMN02369388	PRJNA222257	GCA_001433895.1	Draft	[25]
DS24_1	SAMN06464090	PRJNA336518	GCA_003053135.1	Draft	[24]
DS8_1A	SAMN05583791	PRJNA336518	GCA_003061945.1	Draft	[24]

Table 1 (continued)

Strain	BioSample	BioProject	Code NCBI	Completion level	Reference
DS10_1A	SAMN05583778	PRJNA336518	GCA_003053245.1	Draft	[24]
CIP 76.13	SAMEA2272342	PRJEB1532	GCF_000469705.1	Draft	-
DS11_1A	SAMN05583779	PRJNA336518	GCF_003062025.1	Draft	[24]
UBLA-34	SAMN10136005	PRJNA493554	GCF_003641085.1	Draft	-
DS20_1	SAMN06464087	PRJNA336518	GCA_003061885.1	Draft	[24]
DS1_1A	SAMN05583775	PRJNA336518	GCA_003062045.1	Draft	[24]
DS13_1B	SAMN05583783	PRJNA336518	GCF_003061905.1	Draft	[24]
ATCC 4796	SAMN00001471	PRJNA31477	GCA_000159715.1	Draft	-
DS13_1A	SAMN05583782	PRJNA336518	GCF_003061965.1	Draft	[24]
APC2845	SAMN13342918	PRJNA590940	GCA_017695935.1	Draft	-
LA-G80	SAMN18679498	PRJNA720781	GCF_018252545.1	Draft	-
PB2021-BA04	SAMN18297454	PRJNA714263	GCA_023093425.1	Draft	-
P2	SAMN07665576	PRJNA407882	GCF_002406675.1	Draft	-
WG-LB-IV	SAMN04628015	PRJNA317797	GCF_001639165.1	Draft	-
DS2_1A	SAMN05583785	PRJNA336518	GCA_003062005.1	Draft	[24]
CFH	SAMN02401339	PRJNA227335	GCF_000497795.1	Draft	-

org/) and BioServices (<https://pypi.org/project/bioservices/>). The number of genes identified for each biological process in each strain was then scaled based on the greatest number of gene occurrences for each biological process across all strains. Based on this value, an average was calculated to reflect an empirical score.

Probiotic capacity analysis

For the analysis of the potential probiotic capacity of the different strains of *L. acidophilus*, the tool called iProbiotics was used, which facilitates the rapid screening of probiotics, based on the prediction of probiotic activity in silico, from the genome, which was obtained in FASTA format [30]. iProbiotics has three different parameters: a predictor of probiotic and non-probiotic strains (model one); a predictor of *Lactobacillus* probiotics, *Bifidobacterium* probiotics, and other probiotics (model two); and a predictor of probiotic *Lactobacillus* and non-probiotic *Lactobacillus* (model three), models three being used in this work, as it is more specific for *Lactobacillus* spp. This tool uses characteristics to define probiotic capacity, such as the composition of oligonucleotides, since it plays the role of a molecular marker and genes related to probiotic function, such as adsorption gene, competitiveness gene, a gene linked to growth rate, hydrolase gene of bile salts and gene related to retention.

Similarity analysis between genomes

Similarity analysis between genomes was performed using the FastANI tool, which is a method that estimates the Average Nucleotide Identity (ANI) through sequence comparison

without alignment [31]. In all, 4096 ANI comparisons were produced since the analysis is carried out in an “all against all” (all-vs-all) way. Briefly, the ANI technique allows for estimating a global similarity between two genomes, also serving as an indicator for the taxonomic classification of genera and species [32].

Results

Table 2 shows the numbers of genes identified by the Roary tool from the pangenome, showing the core genome, pangenome, and accessory genome, with 1506 genes, 2643 genes, and 4149 genes, respectively. A representation of the size distribution of the pangenome and core genome is shown in Figure 1. Based on this analysis, it is possible to observe a trend in the growth of the pangenome as more strains are added (considering the average in different permutations), while there is stability in the core genome, which indicates

Table 2 Number of genes present in the pangenome (total genes), core genome (core genes and soft core genes), and accessory genome (shell genes and cloud genes), as calculated by the Roary tool. The core genome is composed of conserved genes in at least 95% of the analyzed strains

Gene pool	Abundance	Amount
Core genes	(95% <= strains <= 100%)	1506
Soft core genes	(94% <= strains < 95%)	0
Shell genes	(15% <= strains < 94%)	485
Cloud genes	(0% <= strains < 15%)	2158
Total genes	(0% <= strains <= 100%)	4149

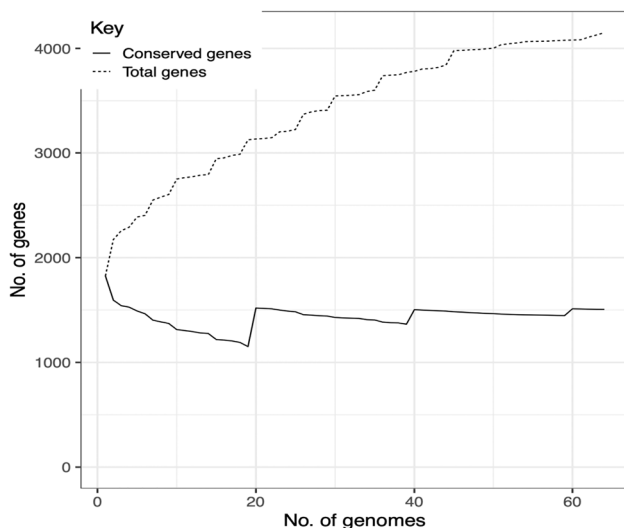


Fig. 1 Graph showing the size of the pangenome and conserved genes (core genome) for different iterations from Roary analysis

that the pangenome is “open” (the gene repertoire of the species is more suitable to grow). Furthermore, it is visible that at some points, there is discontinuity, indicating diversity in the genomes of the strains used.

As presented in Table 3, we noticed a variable distribution of genes involved in producing metabolites such as acetic acid, hydrogen peroxide, lactic acid, and quorum sensing among the strains, in addition to the score generated by the presence of these genes. Among the 64 strains of *L. acidophilus* tested, NCTC 13721 stood out, as it presented a more significant number of genes involved in the quorum sensing process when compared to the other strains and a considerable number of genes involved in hydrogen peroxide, lactic acid, and acetic acid metabolism, with a score of 91%. Followed by NCTC 13721, DSM 20079 showed a relevant amount of hydrogen peroxide, lactic acid, and acetic acid metabolism genes. However, the number of genes involved in QS was a little lower compared to DSM 20079, which obtained a score of 77%.

Table 4 shows the 64 strains of *L. acidophilus* and their probability to be a probiotic. Results were presented in descending order and generated through iProbiotics using models one (probiotics probability) and three (probiotic *Lactobacillus* probability). Strains are sorted based on the model’s three results. As shown, the iProbiotics model could not discriminate the strains, and all of them were predicted as potential probiotics, while model three predicted as probiotics only the strains NCTC 13721 and La-5 with probabilities of 84.8% and 51.9%, respectively.

The phylogenetic tree produced by the multiple sequence alignment of the proteins encoded by the core genome is displayed in Figure 2. The NCBI genome assembly accessions of the three strains were identified as more promising

for probiotics activity, as predicted by iProbiotics and BlastKOALA, as indicated in bold. Finally, ANI similarity values are present for each pair of strains in Supplementary Data 1. In this analysis, the NCTC 13721 strain did not present genomic similarity with any of the strains tested, considering the minimum threshold of 95% for species. DSM 20079, on the other hand, showed genomic similarity with all strains except NCTC 13721. La-5 obtained about 100% of genomic similarity with eight different strains: *L. acidophilus* 20_1, *L. acidophilus* LA-5, *L. acidophilus* FSI4, *L. acidophilus* APC2845, *L. acidophilus* LA-G80-111, *L. acidophilus* DS1_1A, *L. acidophilus* LA1, and *L. acidophilus* BCRC 14065.

Software installation and usage

BlastKoala and iProbiotics were accessed from their respective web servers (<https://www.kegg.jp/blastkoala/> and <http://bioinfor.imu.edu.cn/iprobiotics/public/>). Roary, Python, BioPython, BioServices, and FastANI were installed locally using conda (<https://docs.conda.io/en/latest/>) and Python “pip.”

Discussion

According to the results, of the 64 strains tested in silico, *L. acidophilus* NCTC 13721, *L. acidophilus* DSM 20079, and *L. acidophilus* La-5 showed more significant potential for future in vitro studies. NCTC 13721 presented a higher amount of QS genes compared to the other strains. In addition, NCTC 13721 and DSM 20079 showed similar results in the number of acetic acid, lactic acid, and hydrogen peroxide metabolism genes.

L. acidophilus NCTC 13721, obtained from the vaginal microbiota of a volunteer patient in the United Kingdom and available from the National Collection of Type Cultures bank (NCTC: 13721), showed promising results based on the BlastKOALA and iProbiotics analysis with model one and three; however, this strain is still poorly characterized and have not been evaluated as probiotic to our knowledge. It is relevant to mention that NCTC 13721 presented a total of 90 genes related to processes involved in the probiotic activity (production of acid acetic, hydrogen peroxide, and acid lactic), followed by the DSM 20079 strain with 77 genes. In addition, NCTC 13721 exhibited a significant number of quorum sensing (QS) genes compared to the other strains. These results are important, as lactic acid bacteria with probiotic capacities are related to producing organic acids, bacteriocins, hydrogen peroxide, and biosurfactants [8]. Additionally, they can act by inhibiting bacterial QS and other microorganisms present in the same environment [33]. The QS is fundamental in forming the biofilm, which can

Table 3 Number of genes involved in the metabolic production of different strains of *L. acidophilus* identified from the analysis with BLASTKOALA

Strain	Genome	Production metabolic				Score
		Acid acetic	Hydrogen peroxide	Acid lactic	Quorum sensing	
NCTC 13721	GCA_900452495.1	21	21	10	38	0.91
DSM 20079	GCF_003047065.1	22	22	9	24	0.77
APC2845	GCA_017695935.1	21	21	7	26	0.71
DSM 20079	GCA_001433895.1	20	20	7	29	0.71
ATCC 4796	GCA_000159715.1	20	20	7	28	0.70
NCFM	GCA_000011985.1	20	20	7	27	0.70
DSM 20242	GCA_000442825.1	20	20	7	27	0.70
DSM 9126	GCA_000469745.1	20	20	7	27	0.70
DS24_1	GCA_003053135.1	20	20	7	26	0.70
DS10_1A	GCA_003053245.1	20	20	7	26	0.70
DS20_1	GCA_003061885.1	20	20	7	26	0.70
DS8_1A	GCA_003061945.1	20	20	7	26	0.70
DS2_1A	GCA_003062005.1	20	20	7	26	0.70
DS1_1A	GCA_003062045.1	20	20	7	26	0.70
CIRM-BIA 445	GCA_000469765.1	21	21	7	28	0.69
YT1	GCA_003952845.1	20	20	7	25	0.69
La-5	GCA_017009715.1	19	19	7	27	0.69
LA1	GCA_002286215.1	19	19	7	26	0.68
LA-G80-111	GCA_013342945.1	19	19	7	26	0.68
BCRC 16092	GCA_017009575.1	19	19	7	26	0.68
BCRC 16099	GCA_017009605.1	19	19	7	26	0.68
BCRC 17481	GCA_017009655.1	19	19	7	26	0.68
L3_101_000G1_ dasL3_101_000G1_meta- bat.metabat.48	GCA_018367455.1	19	19	7	26	0.68
BIO6307	GCF_008868625.1	19	19	7	26	0.68
LA_AVK2	GCF_009741835.1	19	19	7	26	0.68
LA1063	GCF_017009725.1	19	19	7	26	0.68
ATCC 4356	GCA_000786395.1	19	19	7	25	0.68
FSI4	GCA_000934625.1	19	19	7	25	0.68
QAULAN51	GCA_022509485.1	19	19	7	25	0.68
PB2021-BA04	GCA_023093425.1	19	19	7	25	0.68
La-14	GCF_000389675.2	19	19	7	25	0.68
CIRM-BIA 442	GCF_000442865.1	19	19	7	25	0.68
CIP 76.13	GCF_000469705.1	19	19	7	25	0.68
CFH	GCF_000497795.1	19	19	7	25	0.68
WG-LB-IV	GCF_001639165.1	19	19	7	25	0.68
KLDS 1.0901	GCF_001868765.1	19	19	7	25	0.68
ATCC 53544	GCF_002224305.1	19	19	7	25	0.68
P2	GCF_002406675.1	19	19	7	25	0.68
BA05	GCF_002914945.1	19	19	7	25	0.68
DS9_1A	GCF_003061925.1	19	19	7	25	0.68
DS13_1A	GCF_003061965.1	19	19	7	25	0.68
DS5_1A	GCF_003061985.1	19	19	7	25	0.68
DS11_1A	GCF_003062025.1	19	19	7	25	0.68
UBLA-34	GCF_003641085.1	19	19	7	25	0.68
s-13	GCF_013867605.1	19	19	7	25	0.68
BCRC 12255	GCF_017009485.1	19	19	7	25	0.68
BCRC 14065	GCF_017009515.1	19	19	7	25	0.68

Table 3 (continued)

Strain	Genome	Production metabolic				Score
		Acid acetic	Hydrogen peroxide	Acid lactic	Quorum sensing	
BCRC 80064	GCF_017009695.1	19	19	7	25	0.68
LA-G80	GCF_018252545.1	19	19	7	25	0.68
LA-5	GCF_024665555.1	19	19	7	25	0.68
NBRC 13951	GCF_001591845.1	19	19	7	24	0.68
DS13_1B	GCF_003061905.1	19	19	7	24	0.68
LA_AVK1	GCF_009742735.1	19	19	7	24	0.68
s-4	GCF_013867555.1	19	19	7	24	0.68
NBIMCC 8242 (180)	GCF_021229035.1	19	19	7	24	0.68
MG-HGUT-02379	GCF_902386525.1	18	18	7	27	0.67
5460	GCA_021432145.1	18	18	7	26	0.67
HN017	GCF_024397395.1	18	18	7	26	0.67
LA-2	GCF_024665075.1	18	18	7	26	0.67
NC55	GCA_020883435.1	18	18	7	25	0.67
BCRC 17008	GCF_017009595.1	18	18	7	25	0.67
BCRC 17486	GCF_017009585.1	19	19	7	25	0.66
BCRC 14079	GCF_017009475.1	18	18	7	25	0.64
PNW3	GCA_004348805.1	19	19	7	23	0.63

increase the time the bacteria will survive in the intestine, increasing its colonization capacity and, thus, making the exchange of nutrients between the host and the microbiota. Furthermore, QS is capable of causing a cooperative change in the expression of bacterial genes, such as the expression of virulence factors [34].

L. acidophilus La-5 strain, correctly predicted as probiotics in our analysis, has already been widely studied in different applications in human health, ranging from in vitro [35–37] to clinical trials [32, 33] studies. Formulations containing this strain are usually prepared in combination with *Bifidobacterium animalis* subsp. lacti and have already been evaluated for a variety of conditions, including diarrhea [38], dermatitis [39], diabetes mellitus [40], and ventilator-associated pneumonia [41].

Regarding the genes involved in producing lactic acid, strains NCTC13721 and DSM 20079 showed higher abundance when compared to other strains. The first showed ten genes, the second 9 genes, and the other 62 strains showed seven genes involved in producing this metabolite. Among the most promising activities of probiotics, the antimicrobial stand out, helping to compete with opportunistic pathogens and inhibiting their adhesion to the mucosa. Lactic acid is an elementary antimicrobial factor [42], responsible for reducing the pH, which leads to inhibition of the growth of pathogenic bacteria [43], since the pH of several pathogenic bacteria is slightly alkaline [8]. The inhibitory and biocidal effects of pure lactic acid in vitro, it is able to act against Gram-negative bacteria: *Salmonella enteritidis*, *Escherichia coli*, and *Pseudomonas aeruginosa*, and also against Gram-positive:

Enterococcus faecalis, *Staphylococcus aureus*, *Bacillus cereus*, and *Listeria monocytogenes* [44]. This inhibitory effect occurs since lactic acid can reduce the pH, preventing the activity of the pathogen's urease, making this microorganism unable to grow at the adhesion site, thus acting as a bactericidal agent. Furthermore, this acid suppresses pro-inflammatory responses mediated by immune cells causing intestinal and immunological homeostasis through enterocyte renewal and macrophage mobilization [45, 46]. Thus, strains with many genes involved in this metabolic process can be prominent allies against different pathogens.

Based on the analysis using iProbiotics, the NCTC 13721 strain showed close results when tested in model one and model three. It is important to mention that the third is more specific for *Lactobacillus* spp. with probiotic capacity. In comparison, La-5 showed different results when compared in the two models; in model one, it presented 99%, and in model three, 51.9%, respectively. It is relevant to say that we noticed that iProbiotics was little used until writing this article. This tool uses machine learning, not mapping the mechanisms of action of the probiotic, which are essential to analyze whether or not an organism has the capacity to perform such a function [30]. It is already known that each probiotic will have specific characteristics, and its activities beneficial to the host may not be the same since each strain has its individual particularities [47]. Therefore, in this study, we made a prediction of probiotics based on the criteria used as probiotic mechanisms dictated by the iProbiotics program. We were able to observe that; when we compare model one with model three, the first presents approximately

Table 4 Different strains of *L. acidophilus* and their respective ability to be a probiotic according to model one of the iProbiotics web server

Strain	Genome	iProbiotics models	
		One	Three
NCTC 13721	GCA_900452495.1	84.26%	84.8%
La-5	GCA_017009715.1	99.0%	51.9%
DS20_1	GCA_003061885.1	98.51%	34.6%
L3_101_000G1_ dasL3_101_000G1_meta- bat.metabat.48	GCA_018367455.1	99.25%	32.7%
BA05	GCF_002914945.1	99.56%	32.6%
DS5_1A	GCF_003061985.1	99.65%	31.2%
DS10_1A	GCA_003053245.1	99.02%	30.3%
KLDS 1.0901	GCF_001868765.1	99.53%	30.0%
CFH	GCF_000497795.1	99.07%	28.6%
UBLA-34	GCF_003641085.1	98.83%	27.4%
BCRC 17486	GCF_017009585.1	99.63%	26.3%
LA-G80	GCF_018252545.1	99.24%	24.9%
PB2021-BA04	GCA_023093425.1	99.47%	24.6%
P2	GCF_002406675.1	99.61%	23.2%
LA1063	GCF_017009725.1	99.6%	22.5%
APC2845	GCA_017695935.1	99.75%	22.1%
DS13_1A	GCF_003061965.1	97.49%	21.8%
BCRC 17008	GCF_017009595.1	99.44%	21.6%
DS9_1A	GCF_003061925.1	99.53%	21.2%
s-13	GCF_013867605.1	99.74%	21.0%
PNW3	GCA_004348805.1	99.64%	20.3%
MG-HGUT-02379	GCF_902386525.1	99.42%	20.0%
LA_AVK1	GCF_009742735.1	99.6%	19.6%
BCRC 12255	GCF_017009485.1	99.82%	19.6%
BCRC 14079	GCF_017009475.1	99.79%	18.8%
DSM 20079	GCA_001433895.1	99.67%	18.3%
DS11_1A	GCF_003062025.1	98.89%	17.9%
BCRC 80064	GCF_017009695.1	99.79%	17.7%
QAULAN51	GCA_022509485.1	98.86%	17.2%
s-4	GCF_013867555.1	98.86%	17.2%
WG-LB-IV	GCF_001639165.1	99.46%	17.1%
BCRC 16092	GCA_017009575.1	99.73%	17.0%
DS8_1A	GCA_003061945.1	99.49%	16.2%
LA_AVK2	GCF_009741835.1	99.01%	14.3%
BCRC 17481	GCA_017009655.1	99.74%	14.3%
DS1_1A	GCA_003062045.1	99.57%	14.0%
BCRC 16099	GCA_017009605.1	99.3%	12.7%
BIO6307	GCF_008868625.1	99.54%	12.1%
BCRC 14065	GCF_017009515.1	99.73%	10.7%
NBIMCC 8242 (180)	GCF_021229035.1	99.76%	10.7%
DS24_1	GCA_003053135.1	98.94%	9.4%
NBRC 13951	GCF_001591845.1	99.47%	9.3%
DS13_1B	GCF_003061905.1	99.67%	8.7%
HN017	GCF_024397395.1	99.45%	8.6%
LA-2	GCF_024665075.1	99.5%	8.3%

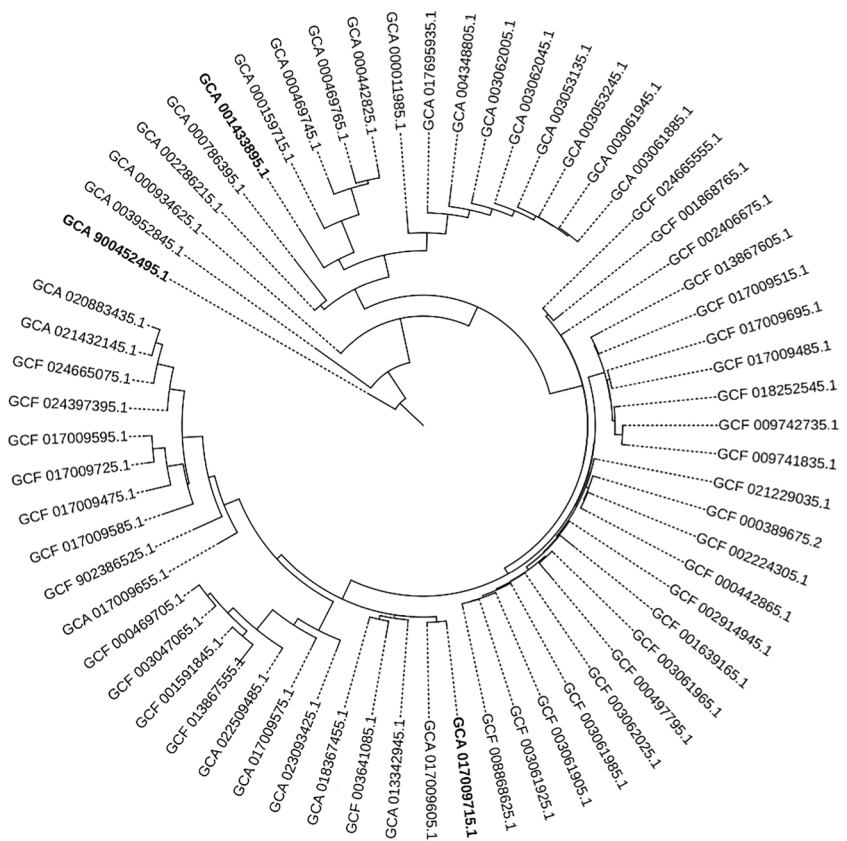
Table 4 (continued)

Strain	Genome	iProbiotics models	
		One	Three
5460	GCA_021432145.1	99.5%	8.3%
YT1	GCA_003952845.1	99.47%	7.9%
NC55	GCA_020883435.1	99.49%	7.2%
DS2_1A	GCA_003062005.1	99.57%	7.1%
ATCC 4356	GCA_000786395.1	99.67%	5.4%
DSM 9126	GCA_000469745.1	99.69%	5.4%
CIP 76.13	GCF_000469705.1	99.69%	5.3%
CIRM-BIA 445	GCA_000469765.1	99.73%	5.0%
DSM 20242	GCA_000442825.1	99.59%	4.7%
CIRM-BIA 442	GCF_000442865.1	99.64%	4.2%
ATCC 53544	GCF_002224305.1	99.69%	4.0%
NCFM	GCA_000011985.1	99.71%	3.7%
La-14	GCF_000389675.2	99.7%	3.7%
LA1	GCA_002286215.1	99.69%	3.7%
LA-G80-111	GCA_013342945.1	99.7%	3.7%
FSI4	GCA_000934625.1	99.7%	3.7%
LA-5	GCF_024665555.1	99.7%	3.7%
DSM 20079	GCF_003047065.1	99.72%	3.3%
ATCC 4796	GCA_000159715.1	99.8%	0.16%

99% of the strains with probiotic capacity above 90%. Therefore, it is understood that model three is more specific for *Lactobacillus* spp. with probiotic capacity, informing different percentages for each strain.

According to ANI-based molecular classification standards (Supplementary Data 1), values below 96% similarity and 90% global alignment indicate that the isolates may belong to different species [32]. The NCTC13721 strain showed approximately 81% similarity in genomics compared to the others, including La-5. Therefore, further characterization studies of the NCTC13721 strain are needed to confirm its classification. The genus *Lactobacillus* spp. has heterogeneous characteristics and includes species with diverse physiological and biochemical features. At present, the definition of *L. acidophilus* is shown in DNA-DNA hybridization, with its CG content of species ranging from 32% to 50%, exceedingly higher than is reported for well-defined bacterial genera [48, 49]. We highlight that, from the phylogenetic tree, the most promising strains: NCTC 13721, DSM 20079, and La-5, are not similar in their evolutionary characteristics, reinforcing the idea of heterogeneity. *L. acidophilus* La-5 showed about 100% genomic similarity with eight strains, namely: *L. acidophilus* DS20_1, *L. acidophilus* LA-5, *L. acidophilus* FSI4, *L. acidophilus* APC2845, *L. acidophilus* LA-G80-111, *L. acidophilus* DS1_1A, *L. acidophilus* LA1, and *L. acidophilus* BCRC 14065. However, these strains have few reports in the literature, consequently, few in vitro evaluations.

Fig. 2 Phylogenetic tree made from the 64 strains of *L. acidophilus*, with GCA_001433895.1 (DSM 20079), GCA_900452495.1 (NCTC 13721), and GCA_017009715.1 (La-5). In this analysis, these strains were evolutionarily distant



Conclusion

As demonstrated by the in silico analysis, NCTC 13721 strain presents genomic features that are desirable for probiotic bacteria, including a higher number of genes involved in QS metabolism. In addition, NCTC 13721 and DSM 20079 showed more genes involved in the production of metabolites involved in the probiotic activity (lactic acid, acetic acid, and hydrogen peroxide) in relation to the microbial inhibitory effect. However, more studies are needed to better characterize the NCTC 13721 strain since the ANI analysis showed a lower similarity with the other strains from the same species.

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Declarations

Competing interests The authors declare no competing interests.

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