Lipoprotein(a) and calcific aortic valve disease initiation and progression: a systematic review and meta-analysis

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Abstract

Although evidence indicates the association of lipoprotein(a) [Lp(a)] with atherosclerosis, the link with calcific aortic valve disease (CAVD) is unclear. This systematic review and meta-analysis explores the connection between Lp(a) and aortic valve calcification and stenosis (AVS). We included all relevant studies, indexed in eight databases, up to February 2023. A total of 44 studies (163 139 subjects) were included, with 16 of them being further meta-analysed. Despite considerable heterogeneity, most studies support the relationship between Lp(a) and CAVD, especially in younger populations, with evidence of early aortic valve micro-calcification in elevated-Lp(a) populations. The quantitative synthesis showed higher Lp(a) levels, by 22.63 nmol/L (95% CI: 9.98–35.27), for patients with AVS, while meta-regressing the data revealed smaller Lp(a) differences for older populations with a higher proportion of females. The meta-analysis of eight studies providing genetic data, revealed that the minor alleles of both rs10455872 and rs3798220 LPA gene loci were associated with higher risk for AVS (pooled odds ratio 1.42; 95% CI: 1.34–1.50 and 1.27; 95% CI: 1.09–1.48, respectively). Importantly, high-Lp(a) individuals displayed not only faster AVS progression, by a mean difference of 0.09 m/s/year (95% CI: 0.09–0.09), but also a higher risk of serious adverse outcomes, including death (pooled hazard ratio 1.39; 95% CI: 1.01–1.90). These summary findings highlight the effect of Lp(a) on CAVD initiation, progression and outcomes, and support the early onset of Lp(a)-related subclinical lesions before clinical evidence.

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Graphical Abstract



Keywords

Lipoprotein(a) • Lp(a) • Calcific aortic valve disease • Aortic valve stenosis • Meta-analysis

1. Introduction

Calcific aortic valve stenosis (AVS) is present in approximately 0.4% of the general population and 2% of individuals over 65 years, constituting one of the most common age-related valvulopathies.^{1,2} Beyond traditional risk factors, lipoprotein(a) [Lp(a)] has also emerged as a new risk factor for calcific aortic valve disease (CAVD), mediating aortic valve calcification (AVC) and AVS.^{3,4} The distinctive 'footprint' of the lipo-proteinic Lp(a) molecule, apolipoprotein(a) [apo(a)], involves two kringles (10 subtypes of KIV, with subtype KIV₂ appearing with a variable number of copies, and a single copy of KV) and an inactive protease domain, which grants its multiple atherogenic and proinflammatory actions.⁵ The levels of Lp(a) are mainly genetically determined, mostly by the LPA gene, which dictates the size of apo(a) and concentration of Lp(a).⁶ Although evidence exists to link CAVD with Lp(a) levels, the level of awareness among physicians is still low, with Lp(a) being measured at rates lower than 5% for populations at risk, ' even in the light of recent guidelines that proclaim its value.⁸ Moreover, the detailed picture of this connection, particularly with regard to demographic, genetic and other interfering factors, is still obscure, a fact reflected by the absence of concrete summary risk estimates regarding the Lp(a)-related outcomes.

This systematic review and meta-analysis summarises the existing evidence regarding the role of elevated Lp(a) in AVC and AVS onset and progression, and aims to highlight the heterogeneity of related data, providing an up-to-date, comprehensive view of this relationship.

2. Methods

2.1 Search strategy

This systematic review was conducted following the Preferred Reporting Items for Systematic Review and Meta-Analysis Protocols statement. The protocol has been prospectively registered in PROSPERO (ID: CRD42022311283). We systematically searched PubMed, Embase, Scopus, Web of Science, ScienceDirect, Cochrane Library, OpenGrey, and LILACS, for articles examining the effect of lipoprotein(a) and relevant genetic factors on AVC and AVS, published from inception until February 2023. The full list of queries per database can be found in Supplementary material online, *Appendix S1*. Two researchers independently assessed the articles for eligibility, based on predefined selection criteria. Any discrepancies were resolved through repeated reviewing and consensus among the authors.

2.2 Selection criteria and data extraction

All studies meeting the following criteria were included in the qualitative synthesis: All types of observational studies (cohort, registry-based cohort, case-control and cross-sectional) in English language, reporting original data published in peer-reviewed journals, aligning with the following PECO framework: (i) Participants—General population or specific population groups; (ii) Exposure—High soluble plasma levels of Lp(a); (iii) Comparator-Normal/low Lp(a) levels; and (iv) Outcomes-AVC or AVS. We also retrieved data from studies exploring the association between relevant genetic risk factors [LPA single-nucleotide polymorphisms (SNPs) and KIV₂ repeats] and AVC or AVS. First author's name, study type and setup, sample size, demographics and other characteristics, Lp(a) levels and method/unit of measurement, relevant LPA SNPs and KIV₂ repeats with their distribution among groups, the ascertainment method of reported outcomes, risk estimates [risk (RR), odds (OR), or hazard (HR) ratio] for AVC/AVS and related outcomes or Lp(a) level differences between compared groups (depending on study design) and, finally, information regarding the risk of bias, were extracted (see Supplementary material online, Appendix S2). The Newcastle-Ottawa Scale (NOS) tool was used to evaluate the risk of bias (see Supplementary material online, Appendix S3).

2.3 Statistical analysis

Meta-analysis was performed to pool the standardised mean difference in Lp(a) exposure levels (measured in nmol/L or mg/dL, on a continuous scale), between AVS and non-AVS patient groups. A sensitivity analysis was performed by excluding studies reporting Lp(a) in mg/dL. We also pooled the Lp(a) level differences between patients with severe/requiring intervention AVS and those with milder disease. The pooled mean difference in annualised peak aortic velocity change (measured in m/s/year), between high- and low-Lp(a) individuals, was calculated, along with the pooled risk of serious adverse outcomes, including death, aortic valve replacement and AVS-related hospitalisation. We also calculated the pooled OR for certain LPA SNPs (rs10455872 and rs3798220) between AVS and non-AVS subjects. Finally, we performed meta-regression to investigate the effect of cofactors on pooled effects, using maximumlikelihood as τ^2 estimator. The significance threshold was set to 0.05. All statistical analyses were performed in R (version 4.2.0). A detailed description of the methodology can be found in Supplementary material online, Appendix S2.

3. Results

3.1 Search results and study characteristics

From the initial 1460 titles, 44 of them were finally included in the systematic review, ^{7,9–51} with 16 being eligible for meta-analysis (*Figure 1*).^{7,12,15,} ^{18–21,24,35–37,41,45,49–51} A total sample size of n = 163 139 subjects, with data on CAVD and Lp(a) levels, was considered (accounting for duplicate cohorts leveraged in more than one study), with a mean/median age from 45 to 80.5 years and a sample-weighted average female-to-male ratio of 1.13/1 (range: 0.16–2.03/1). Most studies (43%) were case-control, while 25% adopted a cohort and 32% a cross-sectional design. In total, 31 (70.5%) studies contained data related to stenosis and 19 (43.2%) to calcification of the aortic valve. While most projects were based in the USA the majority of included subjects originated from Denmark (approximately 47.6% of the total sample size), mostly belonging to the Copenhagen General Population Study. Samples from this cohort, along with subjects from the European Prospective Investigation into Cancer-Norfolk study, the Cardiovascular Health Study, the Aortic Stenosis Progression Observation: Measuring Effects of Rosuvastatin trial, the Multi-Ethnic Study of Atherosclerosis (MESA), the Copenhagen City Heart Study and the Atherosclerosis Risk in Communities (ARIC) study, were included in more than one studies. Only 10 (22.7%) studies used Lp(a) measurement kits with molar quantification (reporting in nmol/L), while the remaining 34 measured Lp(a) solely in mg/dL or quantified it in different ways (e.g. the cholesterol content mass). An additional total of 17 studies provided data on Lp(a)-related genetic risk and CAVD,^{9,20,23,29,31,32,41,44,52–60} with eight of them being further meta-analysed.^{9,41,52–57} A detailed description of all study characteristics and findings can be found in *Table 1*, with an extended version in Supplementary material online, *Appendix S4*.

3.2 Quality assessment

The overall quality was found high for 33 studies (75%) and moderate for the remaining 11 (25%). Cohort studies displayed an average score of 7.2 out of 9, with nine (82%) being of high quality. The same figure was 7.4/9 for case-control and 7.2/10 for cross-sectional studies (n = 15, 79% and n = 9, 64% of high quality, respectively). Supplementary material online, *Appendix S3* includes a detailed report of the NOS quality assessment results.

3.3 Qualitative synthesis 3.3.1 Lp(a) and aortic valve stenosis

Despite the considerable heterogeneity in the design and findings of included studies, an overall trend toward increased risk of calcific AVS was observed for higher-Lp(a) groups. Several studies reported significantly higher Lp(a) levels for AVS patient groups, as compared to subjects without stenosis, with differences reaching 41.62 mg/dL. 9,24,32,41,43,45,49,50 Elevated Lp(a) was found to raise the risk for AVS, ranging from 1.70 (95% CI: 1.33-2.19; 17 745 patients),⁵⁰ to 3.4 (95% CI: 1.1-11.2; 202 patients),²¹ with two analyses suggesting a dose-dependent relationship.^{30,31} In most studies, AVS was associated mainly with two SNPs, rs10455872 and rs3798220.^{9,54,55,57,58} also in a dose-dependent fashion since homozygotes for minor alleles (G and C, respectively) presented with a 2- to nearly 3-fold higher risk, as compared with heterozygotes.^{9,57} Furthermore, an inverse relationship was widely found between KIV_2 repeats and $AVS_2^{23,31}$ with KIV2 number inversely affecting Lp(a) levels.⁴⁴ Seven studies did not find Lp(a) to associate with AVS, including one concerning patients with familial hypercholesterolemia (FH) of younger age (~49 years),²⁵ another investigating an older population with a mean age of 80 years³⁷ and two more studies suggesting only an indirect relationship through autotaxin (ATX).^{12,39}

3.3.2 Lp(a) and aortic valve calcification

The majority of studies showed a clear association between Lp(a) and AVC, with the adjusted risk of AVC ranging from 1.05 (95% CI: 1.02-1.08) per 1-SD of Lp(a) increase¹¹ to 1.79 (95% Cl: 1.32–2.43) for Lp(a) levels above 50 mg/dL⁴⁰ AVC groups displayed higher Lp(a) levels (by 5.7–7.8 mg/dL^{11,13, $\frac{7}{48}$}), as well as an association with the minor alleles of certain LPA SNPs (rs10455872^{29,58,59} and rs3798220²⁹) and fewer KIV2 repeats.^{29,44} Three studies also investigated microcalcification differences between high- and low-Lp(a) groups, by quantifying 18F-sodium fluoride (¹⁸FNaF) uptake with positron emission tomography (PET) scanning. Zheng et al. found that patients with elevated Lp(a) (>35 mg/dL) displayed significantly increased valve micro-calcification, despite their similar peak aortic velocity and calcium score.⁵¹ After a 2- to 3-year follow-up, high-Lp(a) patients of the same study were found with increased calcium score progression and higher annualised peak aortic velocity change. Another study showed that patients with no visible AVC, yet elevated Lp(a), presented with significantly increased valve microcalcification.¹⁹ On the contrary, Kaiser et al. found no difference between high- and low-Lp(a) in patients with non-severe AVS, both in terms of calcium score and ¹⁸FNaF uptake,²⁶ while in another study, increased Lp(a) was associated with the new-onset of AVC, but not with its progression.²⁸ Noteworthy is evidence indicating a dose-dependent relationship also



for calcification, with higher Lp(a) values multiplying the risk of AVC, 14,27,47 yet in a nonlinear fashion. 48

3.3.3 Race/ethnicity heterogeneity and special populations

While most studies leverage samples of single origin or race/ethnicity, evidence from three large, multi-race/ethnic cohorts with more than 7500 subjects, provides head-to-head comparisons among such subpopulations.^{10,14,38,40} Leveraging the MESA and 'Mediators of Atherosclerosis in

South Asians Living in America' cohorts, Makshood *et al.* suggest the contribution of race/ethnicity to Lp(a) levels, with Blacks and South Asians presenting with significantly higher median Lp(a) values than Whites, Hispanics and Chinese Americans.³⁸ Moreover, race/ethnicity was found to mediate the Lp(a) effect on AVC. AVC prevalence was higher in Whites (14.6%) and Hispanics (13.2%), as compared to South Asians (10.7%), Blacks (11.7%) and Chinese (6.6%), with only Whites and Blacks demonstrating a significant association between AVC and Lp(a) levels. Two additional analyses of the ARIC cohort revealed the same trend, since AVC was more

Study	Sample size	Age (yrs)	Sex (female)	Compared groups	Key Findings
Arsenault et <i>al.</i> 9	17 553	59.1 (n/a)	56%	AVS vs. non-AVS	• Higher Lp(a) levels in AVS pts [16.2 (6.3–44.5) vs. 11.6 (6.2–27.6) mg/dL; $P = 0.025$]. • 2-fold risk for AVS in pts with Lp(a) > 50 mg/dL (adjusted for sex, are, smoking and LDLd).
Boakye et al. ¹⁰	2283	80.5 ± 4.3	61.4%	AVC vs. non-AVC	• AVC present in 44.8% of participants, with higher rates among older, White males (58.2%) • Lp(a) independently associated with AVC [adjusted prevalence ratio: 1.09; 95% CI: 1.04– 1.15: $P = 0.0011$.
Bortnick et al. ¹¹ Bourgeois et al. ¹²	3426 232	72 ± 5 64.8 ± 9	63% 42.7%	AVC vs. non-AVC CAVS vs. non-CAVS	 Higher risk of AVC for elevated Lp(a) (aRR: 1.05 per 1-SD increase; 95% CI: 1.02–1.08) Borderline difference in Lp(a) between CAVS and non-CAVS (89.6 ± 119.0 vs. 63.3 ± 87.9 nmol/L; P = 0.055).
Bozbas et al. ¹³	285	70.2 ± 7.3	67.3%	AVC vs. non-AVC	 Lp(a)-bound ATX higher in CAVS pts (aOR: 2.80; 95% CI: 1.39–5.66; P = 0.003). Higher Lp(a) in AVC pts [27.4 (range: 13.0–47.5) vs. 19.9 (range: 10.7–36.1) mg/dL; P = 0.0331
Cao et <i>a</i> l. ¹⁴	4593	Range for age medians: 61–62	Range: 51.4– 61.2%	Multitethnic sample grouped by Lp(a) mass, molarity and cholesterol	 CAVD positively associated with Lp(a) mass, molarity and cholesterol content, either with their upper 25th or 15th percentile as cut-off (P < 0.001, for all associations). Blacks had higher Lp(a) levels while Whites and Hispanics showed higher AVC prevalence
Capoulade et al. ¹⁵	220	58 <u>±</u> 13	40%	Mild/moderate AVSAssociation of Lp(a) and OxPL with AVS progression	 AVS progression faster for top Lp(a) tertiles (V_{peak} change: +0.26 ± 0.26 vs. + 0.17 ± 0.21 m/s/year; P = 0.005) and OxPL-apoB tertiles (+0.26 ± 0.26 m/s/year vs. +0.17 ± 0.21 m/s/year; P = 0.01), during 3.5 ± 1.2 yrs of follow-up. Pts in the top tertiles of Lp(a) or OxPL-apoB were at increased risk of AV replacement and cardiac death.
Capoulade et <i>a</i> l. ¹⁶	220	58±13	40%	Mild/moderate AVS pts— Association of Lp(a) with the outcomes	• CAVS progression associated with Lp(a) levels (OR: 1.10 per 10-mg/dL increase; 95% CI: 1.03–1.19; $P = 0.006$), and OxPL (~3.5 yrs of follow-up). • Stronger association for younger ages (OR: 1.19 per 10-mg/dL increase; 95% CI: 1.07–1.33; $P = 0.002$).
Capoulade et al. ¹⁷ Chen et al. ¹⁸	218 3067	58 ± 13 61.1 ± 7.5	40% 53.4%	Mild/moderate AVS pts, grouped by Lp(a) levels CAVS vs. non-CAVS	• Elevated Lp(a) associated with faster progression rate of AVS ($P < 0.001$ for V _{peak} change, and $P = 0.03$ for risk of AV replacement/cardiac death), over a median of 3.5 yrs. • Lp(a) was higher in CAVS pts [9.1 (3.7–24.8) vs. 6.1 (3.2–14.3) mg/dL; $P < 0.001$], with a multi-orient-blu adjurted 2.001 at 0.07. 0.2. 0.013
Despres et al. ¹⁹	496	68.2 (n/a)	38.36%	CAVS vs. non-CAVS	• Higher Lp(a) in CAVS pts [28.7 (8.2–116.6) vs. 10.9 (3.6–28.8) nmol/L; $P < 0.0001$). • Higher Lp(a) in CAVS pts l28.7 (8.2–116.6) vs. 10.9 (3.6–28.8) nmol/L; $P < 0.0001$). Similarly, CAVS pts had higher OXPL ($P < 0.001$). • Higher TBR
Dong et al. ²⁰	219	63.72 (n/a)	53.5%	CAVD vs. CAD vs. non-CAVD/CAD	• Higher Lp(a) levels in CAVD [37.2 (16.5–79.6) nmol/L] and CAD [46.7 (21.5–104.6) nmol/L] pts, as compared to controls [23.6 (9.4–48.6) nmol/L; $P < 0.001$].
Glader et <i>al.</i> ²¹	202	71.3 ± 8.5	40.6%	Significant AVS vs. non-AVS matched controls	• High Lp(a) (≥48 mg/dL) pts in greater risk for AVS requiring intervention (aOR: 3.4; 95% CI: 1.1–11.2).
Gotoh et <i>a</i> l. ²²	784	62±11	55.7%	High (≥30 mg/dL) vs. low (<30 mg/dL) Lp(a)	• Higher rates of AVC for Lp(a) \ge 30 mg/dL (36.1% vs. 12.7%; P < 0.001). • Lp(a) levels higher in women (P < 0.01) and not related with age. Difference in Lp(a), between AVC and non-AVC groups, higher in female subpopulation.

Continued

Table 1 Con	tinued				
Study	Sample size	Age (yrs)	Sex (female)	Compared groups	Key Findings
Gudbjartsson et <i>al.</i> ²³	12137 with Lp(a) levels	n/a	n/a	Association of Lp(a) levels and KIV-2 repeats with AVS	 AVS associated with higher Lp(a), with aOR: 1.17 per 50 nmol/L increase (95% CI: 1.12–1.22; P < 0.0001). Fewer KIV-2 repeats related to higher Lp(a) molarity and larger apo(a) isoform size. Carriers of G4925A (KIV-2 mutation) had small apo(a) and lower Lp(a). KIV-2 repeats associated with CVD. only before adjusting for Lp(a) molarity.
Hojo et <i>al.</i> ²⁴	861	73 (66–78)	20.9%	Pts with PAD, grouped by AVS presence and other valvulopathies	• Higher Lp(a) in AVS pts [34.0 (16.7–50.0) vs. 20.0 (11.0–35.0) mg/dL; $P = 0.002$].
Hovland et al. ²⁵	78	49 \pm 15 for FH pts; 44 \pm 12 for controls	55% for FH pts; 57% for controls	Lp(a) levels and controls	• Lp(a) levels similar for FH and controls, measured either in mass or molarity. • AVA and V_{peak} did not differ between high (>75 nmol/L) and low Lp(a) FH pts ($P = 0.3$ and $P = 0.65$, respectively). • Higher V_{peak} and smaller AVA for FH pts [1.2 (1.1–1.5) vs. 1.0 (1.0–1.1) m/s; $P = 0.02$, and $2.5 \pm 0.6 \text{ xs}^{-2}$, $P = 0.04$, respectively).
Kaiser et al. ²⁶	58	66.4 ± 5.6	15.4%	Mild/moderate AVS pts, divided by Lp(a) levels (cut-off. 50 mg/dL)	• No difference between Lp(a) groups, in AV calcium score [1388 (450–2424) vs. 1173 (927–1628) AU, respectively; $P = 0.839$] or ¹⁸ FNaF PET uptake (TBR _{mean} : 3.02 ± 1.26 vs. 3.05 + 0.96, respectively; $P = 0.902$).
Kaiser et al. ²⁷	3271	63.3 ± 13.3	53%	Association of AVC with Lp(a)	• Dose-dependent relationship between Lp(a) and AVC, with aOR: 1.89 (95% CI: 1.48–2.42; $P < 0.001$) for 80th-94th and aOR: 2.84 (95% CI: 1.96–4.10; $P < 0.001$) for >95th percentiles.
Kaiser et al. ²⁸	922	66 ± 4.2	52.3%	Three groups according to AVC status at baseline and follow-up	 Lp(a) levels associated with baseline and new-onset AVC [aOR: 1.43; 95% CI: 1.15–1.79 and aOR: 1.30; 95% CI: 1.02–1.65, respectively, for each 50 mg/dL increase in Lp(a), but not with AVC progression.
Kaltoft et <i>a</i> l. ²⁹	12 006 with CT/ 85 884 in total	59.2 (51.1–67) for the sub-cohort with CT	57%	Association of Lp(a) and LPA genotypes with AVC and AVS	 Elevated Lp(a) increased AVC and AVS risk (aOR: 1.62; 95% CI: 1.48–1.77, and aHR: 1.54, 95% CI: 1.38–1.71, respectively, for 10-fold increase in Lp(a)). KIV-2 repeats and rs10455872 associated with increased AVC (aOR: 2.23; 95% CI: 1.81–2.76, and aOR: 1.86; 95% CI: 1.64–2.13, respectively.
Kaltoft et <i>al.</i> ³⁰	69 988	60 (range: 20-100)	54%	Grouped by Lp(a) levels: Low, ≤ 9 vs. moderate, 10–68 vs. high, ≥ 69 mg/dL	 Moderate and high Lp(a) associated with increased CAVD risk, over ~7.4 yrs (aHR: 1.28, 95% CI: 1.13–1.44, and 1.86; 95% CI: 1.57–2.21, respectively). Men and pts with increased BMI had a higher 10-year CAVD risk.
Kamstrup et al. ³¹	77 680	58 (47–67)	56%	Risk of AVS for different Lp(a) percentiles and LPA genotypes	• Dose-dependent relationship between Lp(a) and AVS risk, with aHR: 1.2 (95% CI: 0.8–1.7) for 22nd-66th, 1.6 (95% CI: 1.1–2.4) for 67th-89th, 2.0 (95% CI: 1.2–3.4) for 90th- 95th and 2.9 (95% CI: 1.8–4.9) for >95th Lp(a) percentiles. • Higher Lp(a) levels and AVS risk associated with rs10455872 and rs3798220 minor alleles, and low KIV-2 repeats ($P < 0.05$).
Kamstrup et al. ³²	2138	74 (67–79)	37%	CAVD vs. non-CAVD	 Lp(a) higher in CAVD group [12 (4–48) vs. 8 (4–24) mg/dL; P < 0.001]. For every 10 mg/dL-increase in Lp(a) aOR: 1.10 (95% CI: 1.06–1.13) for CAVD. OxPL-apoB and OxPL-apo(a) correlated with Lp(a) and associated with CAVD.
Langsted et al. ³³	52 652 with Lp(a) levels	58 (48–68)	54.3%	Association of Lp(a) and <i>LPA</i> genotypes with AVS	 Lp(a) associated with higher risk for AVS (aHR: 1.23 for every 1-SD increase; 95% CI: 1.06–1.41). LPA SNPs (rs10455872, rs3798220) and KIV-2 repeats also associated with higher AVS risk.

Study	Sample size	Age (yrs)	Sex (female)	Compared groups	Key Findings
Littmann et al. ³⁴	1860	48土16	44%	Pts with type 1 diabetes mellitus grouped by LD(a) levels	 Elevated Lp(a) associated with higher risk for CAVD (aRR = 2.03; P < 0.05, for Lp(a) > 120 nmol/L).
Liu et <i>al.</i> ³⁵	652 at baseline/ 359 with follow-up	62 ±17	41.9%	Mild/moderate CAVS pts, grouped by Lp(a) levels (cut-off: 38.15 mg/dL)	 Higher baseline V_{peak} (3.70 ± 1.12 vs. 3.43 ± 1.14 m/s; P = 0.012) and higher rates of severe AVS (aOR: 1.78; 95% CI: 1.18–2.66; P = 0.006) for pts with elevated Lp(a). After 3.16 ± 2.74 yrs of follow-up (excluding severe AVS cases), only age was a significant predictor of AV-related surgery or death.
Ljungberg et <i>al.</i> ³⁶	955 with Lp(a) levels	56.7	48%	AVS pts requiring surgery vs. non-AVS controls*	 Higher Lp(a) in AVS pts [55.6 (47.2–64.0) vs. 40.1 (35.6–44.7) nmol/L; P = 0.005] Severe AVS [52.9 (44.4–61.4) nmol/L] corresponded to lower Lp(a) than mild/moderate [77.8 (47.7–107.0)].
Mahabadi et <i>al.</i> ³⁷	968	80±5	48%	Severe AVS vs. non-AVS	• Median Lp(a) did not differ between severe-AVS and non-AVS pts [17 (8–56) vs. 18.5 (8.5–57) mg/dL; $P = 0.56$). Outcome retained in risk factor-adjusted model (aOR: 0.98; 95% CI: 0.90–1.06; $P = 0.57$).
Makshood et al. ³⁸	5366	Range for age mean: 59.3–62.4	Range: 51.4– 57.0%	Grouped by race/ethnicity	 Higher Lp(a) levels for South Asians (17.0 mg/dL) than all other groups (12.9–13.1 mg/dL), except Blacks (35.1 mg/dL). Lower AVC rates in Chinese (6.6%), with higher in Whites (14.6%) and Hispanics (13.2%). Lp(a) positively associated with AVC only in Blacks and Whites.
Nsaibia et <i>a</i> l. ³⁹	008	71 ± 9	35%	CAVS & CAD vs. only CAD	• Higher levels of Lp(a) when CAVS was present (32.5 \pm 36.2 vs. 23.7 \pm 29.5 mg/dL; P = 0.003).
Obisesan et al. ⁴⁰	2083	59.2 ± 4.3	62.2%	Low (≤50) vs. high (>50 mg/dL) Lp(a) levels	 Lp(a) levels >50 mg/dL independently increased the risk for AVC [aOR: 1.79; 95% CI: 1.32–2.43]. Race or sex did not significantly affect this relationship.
Ozkan et <i>a</i> l. ⁴¹	152	72.23 (n/a)	50.7%	CAVD vs. non-CAVD	 Higher Lp(a) in CAVD bts [68.67 (67.17–70.16) vs. 27.05 (25.86–28.23) mg/dL; P < 0.001]. Milder AVS corresponded to higher Lp(a) (non-significant outcome) [mild: 70.28 (68.34–72.23) vs. moderate: 69.10 (67.30–70.90) vs. severe: 67.44 (64.26–70.62); P = 0.388]. Rs1055872 and rs3798220 associated with CAVD.
Simony et al. ⁴²	70 042	60 (50–69)	53.6%	Risk of AVS/other CVD for high Lp(a) levels	 In women, Lp(a) levels increased by 27% after menopause and decreased by 12% with hormone replacement therapy. Lp(a) > 40 mg/dL independently associated with AVS. MI. CAD for both sexes.
Stewart et al. ⁴³	5114	72.7 (n/a)	57.9%	CAVD (sclerosis or stenosis) vs. non-CAVD	• Higher Lp(a) levels for CAVD pts (62.3 \pm 71.4 vs. 50.7 \pm 48.5 mg/dL; P < 0.001). • Lp(a) strongly associated with CAVD (aOR: 1.23; 95% CI: 1.14–1.32; P < 0.001). • Age and male sex double the risk for CAVD.
Sticchi et al. ⁴⁴	69	45 (30–53)	20.3%	Bicuspid aortic valve pts, grouped by AVC and AVS status	• Higher Lp(a) levels associated AVC ($P = 0.008$) and AVS status ($P = 0.043$).
Vassiliou et al. ⁴⁵	165	75.3 (n/a)	29.7%	AVS (severe or mild/moderate) vs. non-AVS	 Higher Lp(a) in AVS pts [30.9 (7.5–68.8) vs. 10.0 (4.1–26.6) mg/dL; P < 0.001]. Severe AVS related to lower median Lp(a) levels than mild/moderate (non-significant outcome) [24.2 (7.2–70.0) vs. 38.4 (9.1–65.6) mg/dL; P = 0.64].
					Continued

Table 1 Cont	tinued				
Study	Sample size	Age (yrs)	Sex (female)	Compared groups	Key Findings
Vongpromek et al. ⁴⁶	129	51±8	37.2%	Pts with heterozygous FH— Association of AVC with Lp(a)	 AVC present in 38.8% of pts (Ca-Score > 0). Elevated Lp(a) increased AVC risk (aOR: 1.11 per 10-mg/dL increase; 95% CI 1.01–1.20; P = 0.03).
Wang et al. ⁴⁷	152	70	43.4%	Symptomatic pts grouped by AVC status	 Lp(a) associated with calcification grade (1.21 ± 0.30 in heavy, 1.41 ± 0.32 in moderate, 1.61 ± 0.34 in mild and 1.63 ± 0.38 in absent AVC; P < 0.01, log-transformed data in nmol/L). Lp(a) associated with AVC in multifactorial model (aOR: 1.04; 95% CI: 1.01–1.06; P = 0.005), along with PCSK9 and age.
Wang et <i>a</i> l. ⁴⁸	410	58.6 ± 10.8	14.1%	AVC vs. non-AVC	 Lp(a) was higher in AVC pts with new-onset MI [23.2 (11.1–42.5) vs. 15.4 (6.8–30.8) mg/ dL; P < 0.001]. Lp(a) levels were independently associated with CAVS in a non-linear fashion.
Wilkinson et al. ⁷	4079	75 (n/a)	47%	CAVS vs. non-CAVS	 Low rates (<5%) of Lp(a) measurement. Non-significant difference in Lp(a) levels for CAVS and non-CAVS [14 (6–48) vs. 15.5 (6.5–63) mg/dL; P = 0.734].
Wodaje et al. ⁴⁹	23 298	55.5 ± 17.1	52%	CAVS vs. non-CAVS	 Higher Lp(a) levels for CAVS pts [20.2 (7.6–63.7) vs. 17 (6.6–43.6) mg/dL; P = 0.009; n = 19 151], for both sexes. Lp(a) levels >90th percentile increased the risk for AVS (aHR: 1.53; 95% CI: 1.08–2.15; P = 0.016; age, sex adjusted).
Zheng et al. ⁵⁰	17 745	59.2 ± 9.1	55.1%	AVS vs. non-AVS	 Higher levels of Lp(a) for AVS pts [15.3 (7.0–41.7) vs. 11.7 (6.3–27.7) mg/dL; P < 0.001]. Lp(a) > 50 mg/dL independent risk factor for AVS (aHR: 1.70; 95% CI: 1.33–2.19; P < 0.001; age, sex, LDLc, CAD adjusted).
Zheng et al. ⁵¹	145	70.3 ± 9.9	31.7%	Pts with AVS, grouped by Lp(a) levels (cut-off: 35 mg/dL)	• At baseline, top Lp(a) tertile pts (>35 mg/dL) showed increased ¹⁸ FNaF PET uptake (TBR _{mean} : 2.16 vs. 1.97; $P = 0.043$), but did not differ in V _{peak} ($P = 0.150$) or Ca-Score ($P = 0.42$). Same for OxPL.
					• High-Lp(a) pts showed increased Ca-Score progression [309 (142–483) vs. 93 (56–296) AU/year; $P = 0.015$], faster hemodynamic progression on echo (0.23 \pm 0.20 vs. 0.14 \pm 0.20 m/s/year; $P = 0.019$), and increased risk for aortic valve replacement and death (HR: 1.87; 95% CI: 1.13–3.08; $P = 0.014$), during follow-up.
All values in mean ± ARIC, Atherosclerosi Coronary artery dise Kringle IV-2 repeat; LI Peripheral arterial dis jet velocity; yrs, Yean	sD or median (IQR); #, Info is Risk In Communities stu ase; CAVD, Calcific aortic DLc, Low density lipoprote ease; PRECISE, Polyvasculal s	rmation obtained after cc by: aRR, Adjusted relative valve disease; CAVS, Calc in cholesterol; Lp(a), Lipoj R Evaluation for Cognitive	nrtacting the authors, ¹⁸ FN risk; ATX, Autotaxin; ¹⁸ FN ific aotric valve stenosis; (protein(a); LPA, Lipoprote s Impairment and vaScular	IaF PET, 18F-Sodium Fluoride Positron Emis J. Agatston units, AV, Aortic valve; AVA, Ao. C.P. C-reactive protein; CT, Computed ton C.R) creactive protein; CT, Computer, n/a, Nu in(a) protein coding gene; n, Number; n/a, Nc Events study; Pts; Patients; SD, Standard dev Events study; Pts; Patients; SD, Standard dev	sion Tomography; 95% CI, 95% Confidence interval: aHR, Adjusted hazard ratio; aOR, Adjusted odds ratio; prit: valve area; AVC, Aortic valve calcification; AVS, Aortic valve stenosis; Ca-Score, Calcium score; CAD, nography; CVD, Cardiovascular disease; Echo, Echocardiography; FH, Familial hypercholestenolemia; KIV-2, st available; OxPL-apo(a)/-apoB, Oxidized phospholipids bound to apolipoprotetin(a)/apolipoprotein B; PAD; iation; SNPs, Single nucleotide polymorphisms; TBR _{mean} , Mean tissue-to-background ratio; V _{peak} ; peak aortic



Figure 2 (A–*C*) Forest plots showing the pooled (A) standardised mean difference (MD) in lipoprotein(a) between patients with aortic valve stenosis and those without, (B) MD only for studies reporting in nmol/L (only the subcohort of individuals with measurements in nmol/L was used from the study by Wodaje *et al.*) and (*C*) standardised MD for patients with severe against those with milder stenosis. Random effects model was applied with the size of each marker corresponding to its relative study weight. AVS, Aortic valve stenosis; Lp(a), Lipoprotein(a); SD, Standard deviation; CI, Confidence interval; MD, Mean difference (SMD, Standardised MD); I², Higgins' and Thompson's I² statistic.

prevalent in White participants, as compared to Black, although the latter displayed higher median Lp(a) levels.^{10,40}

The effect of Lp(a) on CAVD was also confirmed for patients with type I diabetes mellitus (T1DM), as shown by a study with 1860 T1DM patients (median age: 48 years), which reported a significantly increased risk of AVC for elevated-Lp(a) patients (adjusted RR: 2.03; 95% CI: 1.03–4.03, when Lp(a) > 120 mg/dL).³⁴ Among subjects with bicuspid aortic valve (BAV) of younger median age (48 years), Lp(a) levels were also found significantly elevated in both the AVC and stenosis subgroups.⁴⁴ Finally, this relationship also holds for heterozygous FH patients, as shown by Vongpromek

et *al.* who found elevated Lp(a) to increase the risk of AVC (adjusted OR per 10-mg/dL increase: 1.11; 95% Cl 1.01–1.20; P = 0.03) in a sample of 129 FH subjects with a median age of 51 years.⁴⁶ On the contrary, Hovland et *al.* showed no association of Lp(a) with AVS in a smaller sample of 64 FH patients.²⁵

3.4 Quantitative synthesis

After excluding outlying and influential studies, the meta-analysis of 11 studies^{7,12,18–21,24,36,37,45,49} with 26 191 subjects, showed significantly

Α		Odds ratio per risk allele			
Study	Total N	(rs10455872)	OR	95%-CI	Weight
Helgadottir et al. (Stockholm cohort)	1694		1.19	[0.90; 1.57]	3.9%
Chen et al. (GERA)	42353		1.26	[1.15; 1.39]	16.8%
Junco-Vicente et al.	578		1.31	[0.79; 2.18]	1.3%
Perrot et al. (QUEBEC-CAVS)	2026		1.32	[1.07; 1.63]	6.1%
Trenkwalder et al. (GeneCAST)	12882		1.37	[1.24; 1.52]	15.8%
Helgadottir et al. (Iceland cohort)	351799		1.40	[1.24; 1.58]	13.5%
Helgadottir et al. (HUNT)	25781		1.48	[1.28; 1.71]	10.6%
Cairns et al. (CCHS, CGPS combined)	77133	<u> </u>	1.53	[1.21; 1.94]	5.1%
Helgadottir et al. (UK Biobank)	408658		1.54	[1.38; 1.71]	15.0%
Helgadottir et al. (MDCS)	28722		1.55	[1.28; 1.88]	7.1%
Arsenault et al. (MHI Biobank)	763		1.57	[1.10; 2.25]	2.4%
Cairns et al. (EPIC-Norfolk)	14634		- 1.96	[1.37; 2.81]	2.4%
Random effects model		•	1.42	[1.34: 1.50]	100.0%
Heterogeneity: $l^2 = 29\%$					
		0.5 1 2			
В					
-		Odds ratio per risk allele			
Study	Total N	(rs3798220)	OR	95%-CI V	Veight
Cairns et al. (CCHS, CGPS combined)	77158		1.00	[0.53; 1.90]	5.6%
Perrot et al. (QUEBEC-CAVS)	2026		1.12	[0.78; 1.60]	17.7%

Figure 3 (A and B) Forest plots showing the pooled odds ratio of AVS for two LPA SNPs: (A) rs10455872 minor allele G and (B) rs3798220 minor allele C. Random effects model was used with the size of each marker corresponding to its relative study weight. OR, Odds ratio; Cl, Confidence interval; N, Number of participants; I², Higgins' and Thompson's I² statistic.

0.5

1

2

42353

11152

higher Lp(a) levels for AVS patients, by a standardised mean difference of 0.34 (95% CI: 0.19–0.48; P < 0.001; Figure 2A), with a low risk of publication bias. The sensitivity analysis of five studies (5858 subjects), reporting Lp(a) levels in nmol/L, 12,19,20,36,49 confirmed the results, showing higher Lp(a) levels for AVS patients by a pooled mean difference of 22.63 nmol/L (95% Cl: 9.98–35.27; P = 0.008; Figure 2B). No significant difference in Lp(a) levels was observed between severe and milder AVS cases (standardised mean difference 0.21; 95% CI: -0.12 to 0.54; P = 0.130; Figure 2C). Meta-regression identified both age and sex as significant predictors of the Lp(a) difference between AVS and non-AVS patients, with lower Lp(a) differences associated with older age (β -0.02; 95% CI: -0.035 to -0.006; P = 0.012) and higher percentages of female subjects (β -0.017; 95% CI: -0.03 to -0.004; P = 0.017). Both LPA SNPs were significantly associated with the risk of AVS, with pooled odds ratios 1.42 (95% CI: 1.34-1.50; P < 0.001; Figure 3A) for rs10455872 (minor allele G) and 1.27 (95% CI: 1.09–1.48; P = 0.002; Figure 3B) for rs3798220 (minor allele C). While age was inversely associated with the effect size of rs10455872 (β -0.013; 95% CI: -0.021 to -0.005; P = 0.008), no similar association was found for rs3798220, or sex regarding both SNPs (P > 0.05).

Chen et al. (GERA)

Cairns et al. (UK Biobank)

Random effects model

Heterogeneity: $I^2 = 0\%$

High-Lp(a) patients were found to progress faster than their low-Lp(a) counterparts to AVS, displaying a higher annualised peak aortic velocity change by 0.09 m/s/year (95% CI: 0.09-0.09; P < 0.001; Figure 4A).^{15,51}

Moreover, the risk of serious adverse outcomes, including death, was higher for individuals with elevated Lp(a) (pooled hazard ratio 1.39; 95% Cl: 1.01–1.90; P = 0.042; Figure 4B),^{15,35,49–51} an outcome that was retained after excluding influential studies (pooled HR: 1.56; 95% Cl: 1.11-2.18; P = 0.025). No association was found between the adverse event risk and age (P = 0.84) or sex (P = 0.86), across studies. Supplementary material online, Appendix S5 offers a detailed view of the meta-analysis results.

1.26 [1.04; 1.53] 61.1%

1.63 [1.11; 2.39] 15.7%

1.27 [1.09; 1.48] 100.0%

4. Discussion

4.1 Summary of findings in clinical context

Elevated Lp(a) has already been characterised as a potential causal risk factor for atherosclerotic burden and cardiovascular disease, with the most recent European Society of Cardiology (ESC)/European Atherosclerosis Society guidelines on dyslipidaemias suggesting its measurement at least once in the lifetime.⁸ However, despite the sporadic data linking Lp(a) with CAVD, there is still a lack of systematic approaches that summarise and quantify this relationship,⁸ with no definite thresholds to direct mitigation strategies, as reflected in the recent 2021 ESC/European Association of Cardio-Thoracic Surgery guidelines for the management of valvular



Figure 4 (A and B) Forest plots showing the (A) pooled mean difference (MD) in annualised peak aortic velocity change (measured in m/s/year), and (B) the pooled hazard ratio for serious adverse events (death, aortic valve replacement or stenosis-related hospitalisation), between patients with low and those with high lipoprotein(a). Random effects model was applied, with the size of each marker corresponding to its relative study weight. Lp(a), Lipoprotein(a); SD, Standard deviation; CI, Confidence interval; MD, Mean difference; HR, Hazard ratio; *N*, Number of participants; I², Higgins' and Thompson's I² statistic.

diseases.^{5,61} Extending previous attempts to summarise existing data, $^{62-66}$ this work focuses directly on the effect of Lp(a) circulating levels and Lp(a)-associated gene SNPs on CAVD, encompassing the most recent evidence. Moreover, it provides new insights that complement previous metanalytic approaches, 67 by systematically standardising Lp(a) differences between CAVD patients and healthy controls and by offering additional collective evidence regarding the Lp(a)-related AVS progression acceleration and risk of serious adverse events.

Most studies confirm a link between elevated Lp(a) and increased incidence of aortic valve disease.^{21,50} There are also data supporting a dose-dependent effect.^{14,27,30,31,47,48} Genetic factors may contribute to this relationship, with the LPA SNPs rs10455872 (allele G) and rs3798220 (allele C) displaying the most prominent effects. Interestingly, such genetic substrates seem to increase the risk of AVS, not only by affecting the Lp(a) levels but also independently and, in fact, in a dose-dependent manner.⁹ The multifactorially determined number of KIV₂ repeats is also associated with AVS risk, but only through determining the size of apo(a) and, consequently, the levels of Lp(a).^{23,44} Furthermore, pooling the results from available studies showed that Lp(a) not only impacts CAVD onset, but it also accelerates hemodynamic deterioration and results in significantly more deleterious outcomes, including death.^{15,50,51} This association seems to be mediated by the Lp(a)-driven AVC, since more vivid micro-calcification measured with ¹⁸FNaF uptake in PET scanning was observed for patients with no visible AVC but elevated Lp(a).^{19,51} Interestingly, those patients progressed faster to visible calcification and stenosis.⁵¹ In contrast to such evidence supporting the acceleration of CAVD progression by Lp(a), Kaiser et al. did not find a similar relationship in a sample of 922 individuals, yet with a considerable drop-out rate of about two-thirds.²⁸ Given the impact of Lp(a) on CAVD onset and, possibly, progression, future treatment strategies might not only have a place early in life, when Lp(a) displays its original insult, but also later in order to diminish its possible effect on progression. However, more evidence is needed to determine the clinical benefit of such policies, along with the time of intervention and the extent of level reduction that is required.

As implied by our meta-regression analyses, older populations present lesser differences in Lp(a) levels between stenosis cases and controls. Although diagnostic bias, due to early atherosclerotic manifestations in high-Lp(a) subjects leading to faster diagnoses, cannot be excluded, this risk is limited since most studies, spanning across the whole range of ages, are age-matched or do not display significant age differences, ^{7,21,36,37} and, furthermore, the within-study compared groups of most attempts, show similar percentages of comorbidities or predisposing risk factors. ^{21,36,37} However, this paradox of wider Lp(a) differences between CAVD and healthy subjects at younger ages, might also be explained by the hypothesis that elevated Lp(a) assumes its role and serves as an initiating insult early enough, while other age-related degenerative mechanisms also come into play in older ages, diminishing the already 'exhausted' role of Lp(a) and finally accounting for the majority of CAVD cases. ^{16,27}

Similarly, Lp(a) seems to play a more limited role in populations with a higher proportion of females. This variation might, in part, be attributed to other sex-related factors, such as the BAV incidence which is typically higher in males.⁶⁸ Although some studies appear with similar or considerably low BAV rates,^{7,21} or even display an insignificant impact of BAV in sensitivity analysis,⁴⁵ for those not reporting the BAV prevalence and distribution between compared groups, this factor could be confounding and, thus, the results should be interpreted with caution. Additionally, this variation might also be explained by the described sex-related pathophysiological differences in the development of AVS. For the same degree of hemodynamic stenosis, male subjects appear with a higher degree of aortic

cusp calcification, contrary to females presenting with increased fibrosis.^{69,70} This tendency toward a more calcific, than fibrotic, pattern in men might pathophysiologically involve the action of Lp(a),⁷¹ resulting in its more frequent appearance in elevated-Lp(a) male subgroups. However, calcification in the cardiovascular system is not only a male feature and its attribution solely, or even predominantly, to Lp(a) is not a solid assumption, as other sex-related factors, like testosterone, are known to affect calcification in clinical and experimental animal studies.^{72,73} Such contradictory evidence calls for a deeper investigation into whether and how gender interferes with the effect of Lp(a) on CAVD.

4.2 Heterogeneity in study design

The considerable heterogeneity observed in the design of studies offers valuable insights regarding potential sources of variations in outcomes. At first, only 10 studies reported the molar concentration of Lp(a), ^{12,19,20,23,25,34,36,42,47,49} while most of the rest quantified its mass. Since the mass of Lp(a) is heavily influenced by the highly variable apo(a) isoform size,^{5,74} mass concentrations are not linearly correlated with their molar counterparts among different individuals or populations. Therefore, direct comparisons between studies of different measurement types, or even between studies employing the same apo(a)-sensitive estimating method on heterogeneous populations, are prone to bias.⁷⁵ Substantial variations are also encountered in the definition of outcomes, concerning both AVC, which is estimated with computed tomography scanning, echo testing,²² relevant International Classification of Diseases (ICD) codes on medical records²⁹ or PET scan for micro-calcification,^{19,26} and AVS, as-sessed mostly with echo,^{7,12,16,18,24,25,37,39,44} but also with cardiac magnetic resonance imaging,⁴⁵ medical records with relevant ICD codes, and clinical events associated with AVS.^{9,30,32,49,50} Finally, variations exist in Lp(a) level reporting (arithmetic, log-transformed⁴⁷ or even geometric³⁶ means or medians), but also in the adopted Lp(a) cut-offs for risk assessment, defined as 1-SD increase,^{11,33} 10-mg/dL increase¹⁶ or in a percentile-based manner,^{27,31} apart from the more typical 30 or 50 mg/dL thresholds.

4.3 Underlying molecular mechanisms

A central role in controlling the circulatory levels of Lp(a) plays its size, determined by its apo(a) isoform with a highly variable number of KIV₂ repeats.⁵ Fewer KIV₂ repeats reduce the apo(a) size and lead to higher Lp(a) concentrations.^{23,31,32} Furthermore, many genetic factors and, most importantly, SNPs in the LPA locus, such as rs10455872 and rs3798220, affect Lp(a) concentration and increase Lp(a)-related CAVD risk.^{20,29,31-33,41} Ancestry- and race/ethnicity-based studies have revealed, not only the highly variable pattern of SNPs and KIV₂ repeats among different subgroups, but also that such variations can have diverging effects in different populations.^{74,76} Although our understanding of the Lp(a) detrimental effect on the aortic valve is not complete, converging data indicate a pleiotropic mechanism of action.³ Apart from its typical atherogenic property, originating from its lipid-carrying nature, Lp(a) seems to exert its primary effect through the delivery of oxidised phospholipids (OxPL) directly to valve leaflets.^{62,74} When stress is induced to valvular endothelium, hydrophilic Lp(a) molecules infiltrate endothelial cells, attract and act on monocytes, smooth muscle and interstitial cells, and trigger pro-inflammatory and pro-calcifying reactions, mainly through OxPL, which is converted to lysophosphatidic acid through ATX.^{12,15,16,19,32,39,51,62}

4.4 Treatment strategies

Several treatment alternatives for lowering circulating Lp(a) levels have been proposed. While statins seem to slightly raise Lp(a) levels, ^{8,74} niacin, cholesteryl ester transfer protein inhibitors (i.e. Anacetrapid) and mipomersen have been shown to reduce Lp(a) levels by 20-30%.⁷⁷ The same figure for monoclonal antibodies against the PCSK9 seems even higher. In the FOURIER (Further Cardiovascular Outcomes Research with PCSK9 Inhibition in Subjects with Elevated Risk) trial, evolocumab significantly reduced Lp(a) levels and reduced the atherosclerotic cardiovascular

risk.⁷⁸ Similar data were obtained from the ODYSSEY OUTCOMES trial for alirocumab.⁷⁹ Recent evidence suggests an apo(a) size-related manner in achieving Lp(a) level reduction, with larger molecules leading to higher reductions (and further 3% reduction for each additional kringle).⁸⁰ Despite their Lp(a) lowering effect, none of these approaches have proved their clinical benefit specifically for CAVD, so far.^{81,82} Elucidating answers are expected from the ongoing prospective, double-blind, randomised phase II clinical trial 'Effect of PCSK9 InhibitorS On Calcific Aortic Valve DiseasE' (NCT04968509), the results of which have not been published so far. Other approaches shift the interest from lowering Lp(a) to targeting its CAVD-inducing mechanism. An OxPL neutralising antibody, E06-single chain variable fragment, has also been proposed, showing a significant reduction in aortic valve pressure gradient in mice, yet with no clinical data from human studies.⁶²

Finally, the recently developed concept of inhibiting apo(a) messenger RNA production with antisense oligonucleotides comes with promising preliminary results. Pelacarsen (TQJ230), the first developed drug of this category, achieved up to 80% reduction of Lp(a), when used weekly.⁸³ The ongoing HORIZON clinical trial (NCT04023552), which explores the effect of pelacarsen on cardiovascular events, will further enlighten this field. Furthermore, the GalNAc-conjugated siRNAs olpasiran and SLN360, which reduce Lp(a) by directly targeting the *LPA* gene, have also denoted optimistic data.⁸⁴ Both drugs demonstrated a safe profile,^{85,86} while large prospective, double-blind, clinical trials (NCT04270760, NCT05537571) have already been approved and are expected to yield results by 2024.

4.5 Study limitations

The findings of our work can be better understood within its limitations. Diagnostic bias between groups of individual studies, mainly owed to incomplete reporting of hosted data, sporadically encountered in some of them, cannot be excluded. Although this risk seems limited due to the design and data balance of most included studies, the outcomes of this analysis should be interpreted with caution minding this factor. Although the multi-level heterogeneity imposed challenges in summarising the individual study outcomes and, primarily, in pooling estimates, we opted for wider inclusion criteria and included even diverging study designs, so as to provide a holistic view of the available evidence. Accordingly, pooling a standardised mean difference to account for different measurement units and other variations, produced a result that strongly captures the direction of association, but can hardly be physically interpreted. To address this, we repeated the analysis for the more coherent subdivision of studies that reported Lp(a) levels in nmol/L, and obtained a more meaningful mean difference of 22.63 nmol/L for between-group Lp(a) levels, yet based on a smaller sample than the original pool of more than 25 000 subjects.

5. Conclusions

The meta-analysis of existing evidence implies an active role for Lp(a) in the initiation and progression of CAVD, with increased mortality and risk for serious adverse outcomes associated with higher Lp(a) levels. Systematic review of the literature revealed that Lp(a) induces additional risk for T1DM, BAV and heterozygous FH populations, while its levels and effect vary across different race/ethnic groups. Moreover, the quantitative analysis showed a more potent association for male-prevalent populations, as well as for younger individuals, suggesting an early and distinct role of Lp(a) in the initiation of the disease, when other risk factors and degenerative lesions are absent. Of note, genetic variations of LPA gene loci are also implicated in the risk of AVS. Further research could shed light upon populations at risk and pave the way for acknowledging more actively the role of Lp(a) in the disease, along with establishing treatment strategies.

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Supplementary material

Supplementary material is available at Cardiovascular Research online.

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Statement of authors' contribution

All authors fulfil the authorship criteria according to ICMJE recommendations. In particular:

Panteleimon Pantelidis, Evangelos Oikonomou and Stamatios Lampsas substantially contributed to the conception and design of the work, as well as to the acquisition, analysis and interpretation of data. They also equally contributed to drafting the work. Georgios E. Zakynthinos, Antonios Lysandrou, Panagiotis Theofilis and Michael Andrew Vavuranakis substantially contributed to the acquisition and analysis of data for the work, while they also contributed to drafting the work. Konstantinos Kalogeras and Efstratios Katsianos substantially contributed to the conception and design of the work, as well as to the interpretation of data. They also revised it critically for important intellectual content. Gerasimos Siasos, Alexios S. Antonopoulos, Dimitris Tousoulis and Manolis Vavouranakis substantially contributed to the conception and design of the work, and they also revised it critically for important intellectual content. Moreover, all authors approved the final version to be published and all authors agreed to be accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved. In addition to being accountable for the parts of the work, each author is able to identify which co-authors are responsible for specific other parts of the work. In addition, all authors have confidence in the integrity of the contributions of their co-authors.

References

- Lindman BR, Clavel M-A, Mathieu P, lung B, Lancellotti P, Otto CM, Pibarot P. Calcific aortic stenosis. Nat Rev Dis Primer 2016;2:16006.
- 2. Levine GN. Cardiology secrets. 5th ed. Philadelphia, PA: Elsevier; 2018. p269-276.
- Trinder M, Zekavat SM, Uddin MM, Pampana A, Natarajan P. Apolipoprotein B is an insufficient explanation for the risk of coronary disease associated with lipoprotein(a). *Cardiovasc* Res 2021;**117**:1245–1247.
- 4. Chan K-L. Lipoprotein(a) and aortic stenosis. Heart 2022;108:9-10.
- Tsimikas S. A test in context: lipoprotein(a): diagnosis, prognosis, controversies, and emerging therapies. J Am Coll Cardiol 2017;69:692–711.
- Kronenberg F. Human genetics and the causal role of lipoprotein(a) for various diseases. Cardiovasc Drugs Ther 2016;30:87–100.
- Wilkinson MJ, Ma GS, Yeang C, Ang L, Strachan M, DeMaria AN, Tsimikas S, Cotter B. The prevalence of lipoprotein(a) measurement and degree of elevation among 2710 patients with calcific aortic valve stenosis in an academic echocardiography laboratory setting. *Angiology* 2017;**68**:795–798.
- Kronenberg F, Mora S, Stroes ESG, Ference BA, Arsenault BJ, Berglund L, Dweck MR, Koschinsky M, Lambert G, Mach F, McNeal CJ, Moriarty PM, Natarajan P, Nordestgaard BG, Parhofer KG, Virani SS, von Eckardstein A, Watts GF, Stock JK, Ray KK, Tokgözoğlu LS, Catapano AL. Lipoprotein(a) in atherosclerotic cardiovascular disease and aortic stenosis: a European atherosclerosis society consensus statement. *Eur Heart J* 2022;43:3925–3946.
- Arsenault BJ, Boekholdt SM, Dubé M-P, Rhéaume E, Wareham NJ, Khaw K-T, Sandhu MS, Tardif J-C. Lipoprotein(a) levels, genotype, and incident aortic valve stenosis: a prospective Mendelian randomization study and replication in a case-control cohort. *Circ Cardiovasc Genet* 2014;**7**:304–310.
- Boakye E, Dardari Z, Obisesan OH, Osei AD, Wang FM, Honda Y, Dzaye O, Osuji N, Carr JJ, Howard-Claudio CM, Wagenknecht L, Konety S, Coresh J, Matsushita K, Blaha MJ, Whelton SP. Sex-and race-specific burden of aortic valve calcification among older adults without overt coronary heart disease: the atherosclerosis risk in communities study. *Atherosclerosis* 2022;**355**:68–75.
- Bortnick AE, Bartz TM, Ix JH, Chonchol M, Reiner A, Cushman M, Owens D, Barasch E, Siscovick DS, Gottdiener JS, Kizer JR. Association of inflammatory, lipid and mineral markers with cardiac calcification in older adults. *Heart Br Card Soc* 2016;**102**:1826–1834.

- Bourgeois R, Devillers R, Perrot N, Després A-A, Boulanger M-C, Mitchell PL, Guertin J, Couture P, Boffa MB, Scipione CA, Pibarot P, Koschinsky ML, Mathieu P, Arsenault BJ. Interaction of autotaxin with lipoprotein(a) in patients with calcific aortic valve stenosis. JACC Basic Transl Sci 2020;5:888–897.
- Bozbas H, Yildirir A, Atar I, Pirat B, Eroglu S, Aydinalp A, Ozin B, Muderrisoglu H. Effects of serum levels of novel atherosclerotic risk factors on aortic valve calcification. J Heart Valve Dis 2007;16:387–393.
- Cao J, Steffen BT, Guan W, Budoff M, Michos ED, Kizer JR, Post WS, Tsai MY. Evaluation of lipoprotein(a) electrophoretic and immunoassay methods in discriminating risk of calcific aortic valve disease and incident coronary heart disease: the multi-ethnic study of atherosclerosis. *Clin Chem* 2017;63:1705–1713.
- Capoulade R, Chan KL, Yeang C, Mathieu P, Bossé Y, Dumesnil JG, Tam JW, Teo KK, Mahmut A, Yang X, Witztum JL, Arsenault BJ, Després J-P, Pibarot P, Tsimikas S. Oxidized phospholipids, lipoprotein(a), and progression of calcific aortic valve stenosis. J Am Coll Cardiol 2015;66:1236–1246.
- Capoulade R, Yeang C, Chan KL, Pibarot P, Tsimikas S. Association of mild to moderate aortic valve stenosis progression with higher lipoprotein(a) and oxidized phospholipid levels: secondary analysis of a randomized clinical trial. JAMA Cardiol 2018; 3:1212.
- Capoulade R, Torzewski M, Mayr M, Chan K-L, Mathieu P, Bossé Y, Dumesnil JG, Tam J, Teo KK, Burnap SA, Schmid J, Gobel N, Franke UFW, Sanchez A, Witztum JL, Yang X, Yeang C, Arsenault B, Després J-P, Pibarot P, Tsimikas S. ApoCIII-Lp(a) complexes in conjunction with lp(a)-OxPL predict rapid progression of aortic stenosis. *Heart* 2020;**106**:738–745.
- Chen J, Lyu L, Shen J, Pan Y, Jing J, Wang Y-J, Wei T. Epidemiological study of calcified aortic valve stenosis in a Chinese community population. *Postgrad Med J* 2022. doi: 10.1136/pmj-2022-141721
- 19. Després A-A, Perrot N, Poulin A, Tastet L, Shen M, Chen HY, Bourgeois R, Trottier M, Tessier M, Guimond J, Nadeau M, Engert JC, Thériault S, Bossé Y, Witztum JL, Couture P, Mathieu P, Dweck MR, Tsimikas S, Thanassoulis G, Pibarot P, Clavel M-A, Arsenault BJ. Lipoprotein(a), oxidized phospholipids, and aortic valve microcalcification assessed by 18F-sodium fluoride positron emission tomography and computed tomography. *CJC Open* 2019;**1**:131–140.
- Dong H, Cong H, Wang J, Jiang Y, Liu C, Zhang Y, Zhu Y, Wang Q. Correlations between lipoprotein(a) gene polymorphisms and calcific aortic valve disease and coronary heart disease in Han Chinese. *J Int Med Res* 2020;48:300060520965353.
- Glader CA, Birgander LS, Söderberg S, Ildgruben HP, Saikku P, Waldenström A, Dahlén GH. Lipoprotein(a), chlamydia pneumoniae, leptin and tissue plasminogen activator as risk markers for valvular aortic stenosis. *Eur Heart J* 2003;24:198–208.
- Gotoh T, Kuroda T, Yamasawa M, Nishinaga M, Mitsuhashi T, Seino Y, Nagoh N, Kayaba K, Yamada S, Matsuo H. Correlation between lipoprotein(a) and aortic valve sclerosis assessed by echocardiography (the JMS cardiac echo and cohort study). *Am J Cardiol* 1995;**76**: 928–932.
- 23. Gudbjartsson DF, Thorgeirsson G, Sulem P, Helgadottir A, Gylfason A, Saemundsdottir J, Bjornsson E, Norddahl GL, Jonasdottir A, Jonasdottir A, Eggertsson HP, Gretarsdottir S, Thorleifsson G, Indridason OS, Palsson R, Jonasson F, Jonsdottir I, Eyjolfsson GI, Sigurdardottir O, Olafsson I, Danielsen R, Matthiasson SE, Kristmundsdottir S, Halldorsson BV, Hreidarsson AB, Valdimarsson EM, Gudnason T, Benediktsson R, Steinthorsdottir V, Thorsteinsdottir U, Holm H, Stefansson K. Lipoprotein(a) concentration and risks of cardiovascular disease and diabetes. J Am Coll Cardiol 2019;**74**:2982–2994.
- Hojo Y, Kumakura H, Kanai H, Iwasaki T, Ichikawa S, Kurabayashi M. Lipoprotein(a) is a risk factor for aortic and mitral valvular stenosis in peripheral arterial disease. *Eur Heart J Cardiovasc Imaging* 2016;**17**:492–497.
- Hovland A, Narverud I, Lie Øyri LK, Bogsrud MP, Aagnes I, Ueland T, Mulder M, Leijten F, Langslet G, Wium C, Svilaas A, Arnesen KE, Roeters van Lennep J, Aukrust P, Halvorsen B, Retterstøl K, Holven KB. Subjects with familial hypercholesterolemia have lower aortic valve area and higher levels of inflammatory biomarkers. J Clin Lipidol 2021;15:134–141.
- Kaiser Y, Nurmohamed NS, Kroon J, Verberne HJ, Tzolos E, Dweck MR, Somsen AG, Arsenault BJ, Stroes ESG, Zheng KH, Boekholdt SM. Lipoprotein(a) has no major impact on calcification activity in patients with mild to moderate aortic valve stenosis. *Heart Br Card Soc* 2022;**108**:61–66.
- Kaiser Y, Singh SS, Zheng KH, Verbeek R, Kavousi M, Pinto S-J, Vernooij MW, Sijbrands EJG, Boekholdt SM, de Rijke YB, Stroes ESG, Bos D. Lipoprotein(a) is robustly associated with aortic valve calcium. *Heart Br Card Soc* 2021;**107**:1422–1428.
- Kaiser Y, van der Toorn JE, Singh SS, Zheng KH, Kavousi M, Sijbrands EJG, Stroes ESG, Vernooij MW, de Rijke YB, Boekholdt SM, Bos D. Lipoprotein(a) is associated with the onset but not the progression of aortic valve calcification. *Eur Heart J* 2022;43:3960–3967.
- Kaltoft M, Sigvardsen PE, Afzal S, Langsted A, Fuchs A, Kühl JT, Køber L, Kamstrup PR, Kofoed KF, Nordestgaard BG. Elevated lipoprotein(a) in mitral and aortic valve calcification and disease: the Copenhagen general population study. *Atherosclerosis* 2022;**349**:166–174.
- Kaltoft M, Langsted A, Afzal S, Kamstrup PR, Nordestgaard BG. Lipoprotein(a) and body mass compound the risk of calcific aortic valve disease. J Am Coll Cardiol 2022;79:545–558.
- Kamstrup PR, Tybjærg-Hansen A, Nordestgaard BG. Elevated lipoprotein(a) and risk of aortic valve stenosis in the general population. J Am Coll Cardiol 2014;63:470–477.
- Kamstrup PR, Hung M-Y, Witztum JL, Tsimikas S, Nordestgaard BG. Oxidized phospholipids and risk of calcific aortic valve disease: the Copenhagen general population study. Arterioscler Thromb Vasc Biol 2017;37:1570–1578.
- 33. Langsted A, Varbo A, Kamstrup PR, Nordestgaard BG. Elevated lipoprotein(a) does not cause low-grade inflammation despite causal association with aortic valve stenosis and

myocardial infarction: a study of 100,578 individuals from the general population. J Clin Endocrinol Metab 2015; **100**:2690–2699.

- 34. Littmann K, Wodaje T, Alvarsson M, Bottai M, Eriksson M, Parini P, Brinck J. The association of lipoprotein(a) plasma levels with prevalence of cardiovascular disease and metabolic control status in patients with type 1 diabetes. *Diabetes Care* 2020;43:1851–1858.
- Liu S-L, Rozi R, Shi H-W, Gao Y, Guo Y-L, Tang Y-D, Li J-J, Wu N-Q. Association of serum lipoprotein(a) level with the severity and prognosis of calcific aortic valve stenosis: a Chinese cohort study. J Geriatr Cardiol JGC 2020;17:133–140.
- 36. Ljungberg J, Holmgren A, Bergdahl IA, Hultdin J, Norberg M, Näslund U, Johansson B, Söderberg S. Lipoprotein(a) and the apolipoprotein B/A1 ratio independently associate with surgery for aortic stenosis only in patients with concomitant coronary artery disease. J Am Heart Assoc 2017;6:e007160.
- Mahabadi AA, Kahlert P, Kahlert HA, Dykun I, Balcer B, Forsting M, Heusch G, Rassaf T. Comparison of lipoprotein(a)-levels in patients ≥70 years of age with versus without aortic valve stenosis. Am J Cardiol 2018;122:645–649.
- Makshood M, Joshi PH, Kanaya AM, Ayers C, Budoff M, Tsai MY, Blaha M, Michos ED, Post WS. Lipoprotein (a) and aortic valve calcium in South Asians compared to other race/ethnic groups. *Atherosclerosis* 2020;**313**:14–19.
- 39. Nsaibia MJ, Mahmut A, Boulanger M-C, Arsenault BJ, Bouchareb R, Simard S, Witztum JL, Clavel M-A, Pibarot P, Bossé Y, Tsimikas S, Mathieu P. Autotaxin interacts with lipoprotein(a) and oxidized phospholipids in predicting the risk of calcific aortic valve stenosis in patients with coronary artery disease. J Intern Med 2016;280:509–517.
- 40. Obisesan OH, Kou M, Wang FM, Boakye E, Honda Y, Uddin SMI, Dzaye O, Osei AD, Orimoloye OA, Howard-Claudio CM, Coresh J, Blumenthal RS, Hoogeveen RC, Budoff MJ, Matsushita K, Ballantyne CM, Blaha MJ. Lipoprotein(a) and subclinical vascular and valvular calcification on cardiac computed tomography: the atherosclerosis risk in communities study. J Am Heart Assoc 2022;11:e024870.
- Ozkan O, Yildiz B. Lipoprotein(a) gene polymorphism increases a risk factor for aortic valve calcification. J Cardiovasc Dev Dis 2019;6:31.
- 42. Simony SB, Mortensen MB, Langsted A, Afzal S, Kamstrup PR, Nordestgaard BG. Sex differences of lipoprotein(a) levels and associated risk of morbidity and mortality by age: the Copenhagen general population study. *Atherosclerosis* 2022;**355**:76–82.
- Stewart BF, Siscovick D, Lind BK, Gardin JM, Gottdiener JS, Smith VE, Kitzman DW, Otto CM. Clinical factors associated with calcific aortic valve disease. Cardiovascular health study. J Am Coll Cardiol 1997;29:630–634.
- Sticchi E, Giusti B, Cordisco A, Gori AM, Sereni A, Sofi F, Mori F, Colonna S, Fugazzaro MP, Pepe G, Nistri S, Marcucci R. Role of lipoprotein (a) and LPA KIV2 repeat polymorphism in bicuspid aortic valve stenosis and calcification: a proof of concept study. *Intern Emerg Med* 2019;**14**:45–50.
- 45. Vassiliou VS, Flynn PD, Raphael CE, Newsome S, Khan T, Ali A, Halliday B, Studer Bruengger A, Malley T, Sharma P, Selvendran S, Aggarwal N, Sri A, Berry H, Donovan J, Lam W, Auger D, Cook SA, Pennell DJ, Prasad SK. Lipoprotein(a) in patients with aortic stenosis: insights from cardiovascular magnetic resonance. *PLoS One* 2017;**12**:e0181077.
- 46. Vongpromek R, Bos S, Ten Kate G-JR, Yahya R, Verhoeven AJM, de Feyter PJ, Kronenberg F, Roeters van Lennep JE, Sijbrands EJG, Mulder MT. Lipoprotein(a) levels are associated with aortic valve calcification in asymptomatic patients with familial hypercholesterolaemia. J Intern Med 2015;278:166–173.
- Wang W-G, He Y-F, Chen Y-L, Zhao F-M, Song Y-Q, Zhang H, Ma Y-H, Guan X, Zhang W-Y, Chen X-L, Liu C, Cong H-L. Proprotein convertase subtilisin/kexin type 9 levels and aortic valve calcification: a prospective, cross sectional study. J Int Med Res 2016;44: 865–874.
- Wang Z, Li M, Liu N. The nonlinear correlation between lipoprotein (a) and the prevalence of aortic valve calcification in patients with new-onset acute myocardial infarction. *Acta Cardiol* 2022;**77**:950–959.
- Wodaje T, Littmann K, Häbel H, Bottai M, Bäck M, Parini P, Brinck J. Plasma lipoprotein(a) measured in routine clinical care and the association with incident calcified aortic valve stenosis during a 14-year observational period. *Atherosclerosis* 2022;**349**:175–182.
- Zheng KH, Arsenault BJ, Kaiser Y, Khaw K-T, Wareham NJ, Stroes ESG, Boekholdt SM. Apob/apoA-I ratio and Ip(a) associations with aortic valve stenosis incidence: insights from the EPIC-norfolk prospective population study. J Am Heart Assoc 2019;8:e013020.
- 51. Zheng KH, Tsimikas S, Pawade T, Kroon J, Jenkins WSA, Doris MK, White AC, Timmers NKLM, Hjortnaes J, Rogers MA, Aikawa E, Arsenault BJ, Witztum JL, Newby DE, Koschinsky ML, Fayad ZA, Stroes ESG, Boekholdt SM, Dweck MR. Lipoprotein(a) and oxidized phospholipids promote valve calcification in patients with aortic stenosis. *J Am Coll Cardiol* 2019;**73**:2150–2162.
- 52. Trenkwalder T, Nelson CP, Musameh MD, Mordi IR, Kessler T, Pellegrini C, Debiec R, Rheude T, Lazovic V, Zeng L, Martinsson A, Gustav Smith J, Gådin JR, Franco-Cereceda A, Eriksson P, Nielsen JB, Graham SE, Willer CJ, Hveem K, Kastrati A, Braund PS, Palmer CNA, Aracil A, Husser O, Koenig W, Schunkert H, Lang CC, Hengstenberg C, Samani NJ. Effects of the coronary artery disease associated LPA and 9p21 loci on risk of aortic valve stenosis. Int J Cardiol 2019;**276**:212–217.
- 53. Perrot N, Thériault S, Dina C, Chen HY, Boekholdt SM, Rigade S, Després A-A, Poulin A, Capoulade R, Le Tourneau T, Messika-Zeitoun D, Trottier M, Tessier M, Guimond J, Nadeau M, Engert JC, Khaw K-T, Wareham NJ, Dweck MR, Mathieu P, Pibarot P, Schott J-J, Thanassoulis G, Clavel M-A, Bossé Y, Arsenault BJ. Genetic variation in LPA, calcific aortic valve stenosis in patients undergoing cardiac surgery, and familial risk of aortic valve micro-calcification. JAMA Cardiol 2019;4:620–627.

- Cairns BJ, Coffey S, Travis RC, Prendergast B, Green J, Engert JC, Lathrop M, Thanassoulis G, Replicated CRA. Genome-wide significant association of aortic stenosis with a genetic variant for lipoprotein(a): meta-analysis of published and novel data. *Circulation* 2017;**135**: 1181–1183.
- 55. Helgadottir A, Thorleifsson G, Gretarsdottir S, Stefansson OA, Tragante V, Thorolfsdottir RB, Jonsdottir I, Bjornsson T, Steinthorsdottir V, Verweij N, Nielsen JB, Zhou W, Folkersen L, Martinsson A, Heydarpour M, Prakash S, Oskarsson G, Gudbjartsson T, Geirsson A, Olafsson I, Sigurdsson EL, Almgren P, Melander O, Franco-Cereceda A, Hamsten A, Fritsche L, Lin M, Yang B, Hornsby W, Guo D, Brummett CM, Abecasis G, Mathis M, Milewicz D, Body SC, Eriksson P, Willer CJ, Hveem K, Newton-Cheh C, Smith JG, Danielsen R, Thorgeirsson G, Thorsteinsdottir U, Gudbjartsson DF, Holm H, Stefansson K. Genome-wide analysis yields new loci associating with aortic valve stenosis. *Nat Commun* 2018;9:987.
- Junco-Vicente A, Solache-Berrocal G, Del Río-García Á, Rolle-Sóñora V, Areces S, Morís C, Martín M, Rodríguez I. IL6 Gene polymorphism association with calcific aortic valve stenosis and influence on serum levels of interleukin-6. *Front Cardiovasc Med* 2022;**9**:989539.
- Chen HY, Dufresne L, Burr H, Ambikkumar A, Yasui N, Luk K, Ranatunga DK, Whitmer RA, Lathrop M, Engert JC, Thanassoulis G. Association of LPA variants with aortic stenosis: a large-scale study using diagnostic and procedural codes from electronic health records. JAMA Cardiol 2018;3:18–23.
- 58. Thanassoulis G, Campbell CY, Owens DS, Smith JG, Smith AV, Peloso GM, Kerr KF, Pechlivanis S, Budoff MJ, Harris TB, Malhotra R, O'Brien KD, Kamstrup PR, Nordestgaard BG, Tybjaerg-Hansen A, Allison MA, Aspelund T, Criqui MH, Heckbert SR, Hwang S-J, Liu Y, Sjogren M, van der Pals J, Kälsch H, Mühleisen TW, Nöthen MM, Cupples LA, Caslake M, Di Angelantonio E, Danesh J, Rotter JI, Sigurdsson S, Wong Q, Erbel R, Kathiresan S, Melander O, Gudnason V, O'Donnell CJ, Post WS, CHARGE Extracoronary Calcium Working Group. Genetic associations with valvular calcification and aortic stenosis. N Engl J Med 2013;368:503–512.
- Cardoso-Saldaña G, Fragoso JM, Lale-Farjat S, Torres-Tamayo M, Posadas-Romero C, Vargas-Alarcón G, Posadas-Sánchez R. The rs10455872-G allele of the LPA gene is associated with high lipoprotein(a) levels and increased aortic valve calcium in a Mexican adult population. *Genet Mol Biol* 2019;42:519–525.
- 60. Emdin CA, Khera AV, Natarajan P, Klarin D, Won H-H, Peloso GM, Stitziel NO, Nomura A, Zekavat SM, Bick AG, Gupta N, Asselta R, Duga S, Merlini PA, Correa A, Kessler T, Wilson JG, Bown MJ, Hall AS, Braund PS, Samani NJ, Schunkert H, Marrugat J, Elosua R, McPherson R, Farrall M, Watkins H, Willer C, Abecasis GR, Felix JF, Vasan RS, Lander E, Rader DJ, Danesh J, Ardissino D, Gabriel S, Saleheen D, Kathiresan S. Phenotypic characterization of genetically lowered human lipoprotein(a) levels. J Am Coll Cardiol 2016;68:2761–2772.
- 61. Vahanian A, Beyersdorf F, Praz F, Milojevic M, Baldus S, Bauersachs J, Capodanno D, Conradi L, De Bonis M, De Paulis R, Delgado V, Freemantle N, Gilard M, Haugaa KH, Jeppsson A, Jüni P, Pierard L, Prendergast BD, Sádaba JR, Tribouilloy C, Wojakowski W, ESC/EACTS Scientific Document Group. 2021 ESC/EACTS guidelines for the management of valvular heart disease. *Eur Heart J* 2022;**43**:561–632.
- Hu J, Lei H, Liu L, Xu D. Lipoprotein(a), a lethal player in calcific aortic valve disease. Front Cell Dev Biol 2022;10:812368.
- Guddeti RR, Patil S, Ahmed A, Sharma A, Aboeata A, Lavie CJ, Alla VM. Lipoprotein(a) and calcific aortic valve stenosis: a systematic review. Prog Cardiovasc Dis 2020;63:496–502.
- Hsieh G, Rizk T, Berman AN, Biery DW, Blankstein R. The current landscape of lipoprotein(a) in calcific aortic valvular disease. *Curr Opin Cardiol* 2021;36:542–548.
- Bhatia HS, Wilkinson MJ. Lipoprotein(a): evidence for role as a causal risk factor in cardiovascular disease and emerging therapies. J Clin Med 2022;11:6040.
- Santangelo G, Faggiano A, Bernardi N, Carugo S, Giammanco A, Faggiano P. Lipoprotein(a) and aortic valve stenosis: a casual or causal association? *Nutr Metab Cardiovasc Dis* 2022;**32**: 309–317.
- Liu Q, Yu Y, Xi R, Li J, Lai R, Wang T, Fan Y, Zhang Z, Xu H, Ju J. Association between lipoprotein(a) and calcific aortic valve disease: a systematic review and meta-analysis. Front Cardiovasc Med 2022;9:877140.
- Tutar E, Ekici F, Atalay S, Nacar N. The prevalence of bicuspid aortic valve in newborns by echocardiographic screening. Am Heart J 2005;150:513–515.
- Summerhill VI, Moschetta D, Orekhov AN, Poggio P, Myasoedova VA. Sex-specific features of calcific aortic valve disease. *Int J Mol Sci* 2020;21:5620.
- Simard L, Côté N, Dagenais F, Mathieu P, Couture C, Trahan S, Bossé Y, Mohammadi S, Pagé S, Joubert P, Clavel M-A. Sex-related discordance between aortic valve calcification and hemodynamic severity of aortic stenosis: is valvular fibrosis the explanation? *Circ Res* 2017;**120**:681–691.
- Alushi B, Curini L, Christopher MR, Grubitzch H, Landmesser U, Amedei A, Lauten A. Calcific aortic valve disease-natural history and future therapeutic strategies. Front Pharmacol 2020;11:685.
- McRobb L, Handelsman DJ, Heather AK. Androgen-induced progression of arterial calcification in apolipoprotein E-null mice is uncoupled from plaque growth and lipid levels. *Endocrinology* 2009;**150**:841–848.
- Christian RC, Dumesic DA, Behrenbeck T, Oberg AL, Sheedy PF, Fitzpatrick LA. Prevalence and predictors of coronary artery calcification in women with polycystic ovary syndrome. *J Clin Endocrinol Metab* 2003;88:2562–2568.
- 74. Tsimikas S, Fazio S, Ferdinand KC, Ginsberg HN, Koschinsky ML, Marcovina SM, Moriarty PM, Rader DJ, Remaley AT, Reyes-Soffer G, Santos RD, Thanassoulis G, Witztum JL, Danthi S, Olive M, Liu L. NHLBI Working group recommendations to reduce

lipoprotein(a)-mediated risk of cardiovascular disease and aortic stenosis. J Am Coll Cardiol 2018;**71**:177–192.

- Tsimikas S, Fazio S, Viney NJ, Xia S, Witztum JL, Marcovina SM. Relationship of lipoprotein(a) molar concentrations and mass according to lipoprotein(a) thresholds and apolipoprotein(a) isoform size. J Clin Lipidol 2018;**12**:1313–1323.
- Lee S-R, Prasad A, Choi Y-S, Xing C, Clopton P, Witztum JL, Tsimikas S. LPA Gene, ethnicity, and cardiovascular events. *Circulation* 2017;**135**:251–263.
- 77. AIM-HIGH Investigators, Boden WE, Probstfield JL, Anderson T, Chaitman BR, Desvignes-Nickens P, Koprowicz K, McBride R, Teo K, Weintraub W. Niacin in patients with low HDL cholesterol levels receiving intensive statin therapy. N Engl J Med 2011; 365:2255–2267.
- 78. O'Donoghue ML, Fazio S, Giugliano RP, Stroes ESG, Kanevsky E, Gouni-Berthold I, Im K, Lira Pineda A, Wasserman SM, Češka R, Ezhov MV, Jukema JW, Jensen HK, Tokgözoğlu SL, Mach F, Huber K, Sever PS, Keech AC, Pedersen TR, Sabatine MS. Lipoprotein(a), PCSK9 inhibition, and cardiovascular risk: insights from the FOURIER trial. *Circulation* 2019;**139**: 1483–1492.
- 79. Szarek M, Bittner VA, Aylward P, Baccara-Dinet M, Bhatt DL, Diaz R, Fras Z, Goodman SG, Halvorsen S, Harrington RA, Jukema JW, Moriarty PM, Pordy R, Ray KK, Sinnaeve P, Tsimikas S, Vogel R, White HD, Zahger D, Zeiher AM, Steg PG, Schwartz GG, for the ODYSSEY OUTCOMES Investigators. Lipoprotein(a) lowering by alirocumab reduces the total burden of cardiovascular events independent of low-density lipoprotein cholesterol lowering: ODYSSEY OUTCOMES trial. *Eur Heart J* 2020;**41**:4245–4255.
- Blanchard V, Chemello K, Hollstein T, Hong-Fong CC, Schumann F, Grenkowitz T, Nativel B, Coassin S, Croyal M, Kassner U, Lamina C, Steinhagen-Thiessen E, Lambert G. The size of apolipoprotein (a) is an independent determinant of the reduction in lipoprotein (a) induced by PCSK9 inhibitors. *Cardiovasc Res* 2022;**118**:2103–2111.

- Cannon CP, Shah S, Dansky HM, Davidson M, Brinton EA, Gotto AM, Stepanavage M, Liu SX, Gibbons P, Ashraf TB, Zafarino J, Mitchel Y, Barter P, Determining the Efficacy and Tolerability Investigators. Safety of anacetrapib in patients with or at high risk for coronary heart disease. N Engl | Med 2010;363:2406–2415.
- Nicholls SJ, Brewer HB, Kastelein JJP, Krueger KA, Wang M-D, Shao M, Hu B, McErlean E, Nissen SE. Effects of the CETP inhibitor evacetrapib administered as monotherapy or in combination with statins on HDL and LDL cholesterol: a randomized controlled trial. JAMA 2011;306:2099–2109.
- Tsimikas S, Karwatowska-Prokopczuk E, Gouni-Berthold I, Tardif J-C, Baum SJ, Steinhagen-Thiessen E, Shapiro MD, Stroes ES, Moriarty PM, Nordestgaard BG, Xia S, Guerriero J, Viney NJ, O'Dea L, Witztum JL. Lipoprotein(a) reduction in persons with cardiovascular disease. N Engl J Med 2020;382:244–255.
- Sohn W, Winkle P, Neutel J, Wu Y, Jabari F, Terrio C, Varrieur T, Wang J, Hellawell J. Pharmacokinetics, pharmacodynamics, and tolerability of olpasiran in healthy Japanese and non-Japanese participants: results from a phase I, single-dose, open-label study. *Clin Ther* 2022;44:1237–1247.
- Koren MJ, Moriarty PM, Baum SJ, Neutel J, Hernandez-Illas M, Weintraub HS, Florio M, Kassahun H, Melquist S, Varrieur T, Haldar SM, Sohn W, Wang H, Elliott-Davey M, Rock BM, Pei T, Homann O, Hellawell J, Watts GF. Preclinical development and phase 1 trial of a novel siRNA targeting lipoprotein(a). *Nat Med* 2022;**28**:96–103.
- Nissen SE, Wolski K, Balog C, Swerdlow DI, Scrimgeour AC, Rambaran C, Wilson RJ, Boyce M, Ray KK, Cho L, Watts GF, Koren M, Turner T, Stroes ES, Melgaard C, Campion GV. Single ascending dose study of a short interfering RNA targeting lipoprotein(a) production in individuals with elevated plasma lipoprotein(a) levels. JAMA 2022;**327**:1679.