



Review

The Emerging Role of Visible Light in Melanocyte Biology and Skin Pigmentary Disorders: Friend or Foe?

Xuanxuan He [†], Shanglin Jin [†], Xiaoxi Dai, Li Chen, Leihong Xiang ^{*} and Chengfeng Zhang ^{*}

Department of Dermatology, Huashan Hospital, Fudan University, Shanghai 200040, China; hxx20000819@163.com (X.H.); jinshanglin203@126.com (S.J.); ynkmykdx123@163.com (X.D.); cl_1063@163.com (L.C.)

^{*} Correspondence: flora_xiang@vip.163.com (L.X.); e3dangdang@hotmail.com (C.Z.)

[†] These authors contributed equally to this work.

Abstract: Electromagnetic radiation, notably visible light (VL), has complicated effects on human skin, particularly pigmentation, which have been largely overlooked. In this review, we discuss the photobiological mechanisms, pathological effects, clinical applications and therapeutic strategies of VL at varying wavelengths on melanocyte biology and skin pigmentary disorders. Different VL wavelengths may impose positive or negative effects, depending on their interactions with specific chromophores, photoaging, ROS production, circadian rhythm and other photon-mediated reactions. Further in vivo and in vitro studies are required to establish the pathologic mechanisms and application principles of VL in pigmentary disorders, as well as optimal photoprotection with coverage against VL wavelengths.

Keywords: visible light; pigmentary disorders; vitiligo; melasma; laser; LEDs; IPL



Citation: He, X.; Jin, S.; Dai, X.; Chen, L.; Xiang, L.; Zhang, C. The Emerging Role of Visible Light in Melanocyte Biology and Skin Pigmentary Disorders: Friend or Foe? *J. Clin. Med.* **2023**, *12*, 7488. <https://doi.org/10.3390/jcm12237488>

Academic Editors: Konstantinos Lasithiotakis and Alexander C. Katoulis

Received: 19 October 2023
Revised: 22 November 2023
Accepted: 1 December 2023
Published: 4 December 2023



Copyright: © 2023 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

1. Introduction

Visible light (VL) refers to the narrow spectrum of electromagnetic radiation (EMR) that human eyes can perceive, with a wavelength range of 400 to 700 nm, although some individuals can also sense wavelengths from 380 to 780 nm [1]. Different wavelengths of VL cause various sensations to the human eyes, exhibiting different colors, which is also the principle of further division (Figure 1). Sunlight is the major source of VL, which makes up approximately 50% of sunlight encountering the Earth's surface. Artificial sources of VL include flashlights, fluorescent lights, lasers, light-emitting diodes (LEDs), and other therapeutic devices [2]. In modern society, electronic devices such as computers and smartphones are also becoming increasingly prevalent as a source of radiation. Therefore, as the first barrier of the body, skin is bound to be affected by visible light exposure.

Skin pigmentary disorders, such as melasma and vitiligo, are characterized by hyperpigmented or depigmented lesions, and have a rapidly growing incidence worldwide. These disorders have serious impacts on people's appearance as well as mental health [3], making it an urgent task to unveil the mystery of their pathogenesis and optimized treatment. While previous studies have mainly focused on the effect of ultraviolet (UV) radiation, the effect of VL on the skin, especially melanocyte biology, has been overlooked or considered negligible. The objective of this review is to enhance the understanding of the pathogenesis and management of pigmentary disorders by exploring the respective effects of VL at different wavelengths on melanocyte biology and diverse phototherapies for pigmentary disorders, using various light sources within the VL spectrum.

VL can be classified into primarily four categories, according to the wavelength and color: these are blue light, green light, yellow light and red light, with the wavelength of 400–490 nm, 490–570 nm, 570–595 nm, and 630–770 nm, respectively. The longer the wavelength, the deeper the VL can penetrate into the skin. Therefore, red light can penetrate through the full thickness of the epidermis and dermis, reaching the subcutaneous layer, while blue light has less penetration. The sources of VL can be divided into three

categories: natural light, artificial light and electronic devices. UV refers to ultraviolet; IR to infrared radiation.

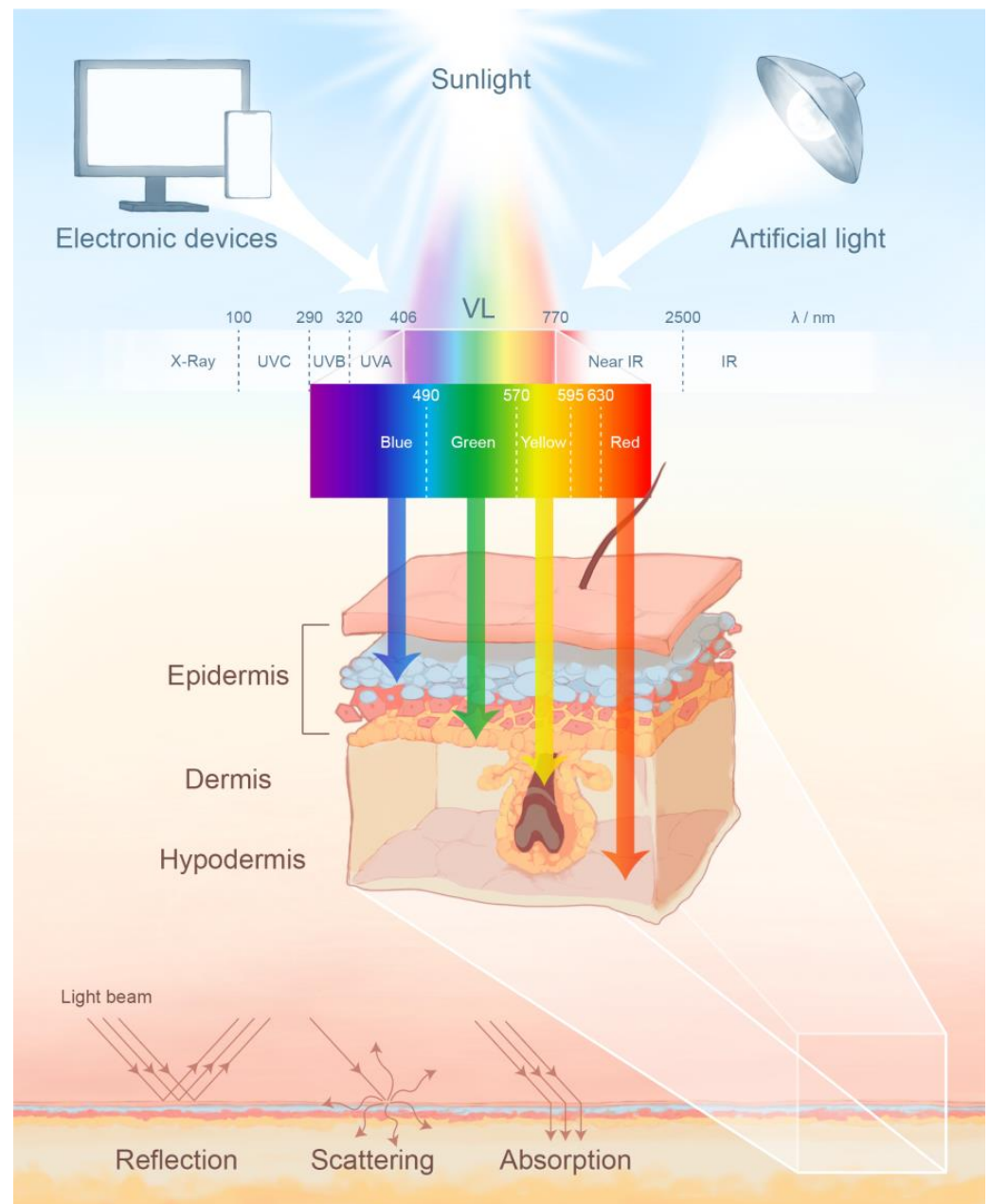


Figure 1. The classification, sources and penetration of visible light (VL).

2. The Photobiological Effects of Visible Light on Skin

It was previously believed that VL had minimal biological effects. However, a large number of researches in the past decades has made it evident that VL can exert significant impacts on various biological processes of skin, including chromophore activation, photoaging, oxidative stress, DNA damage and circadian rhythm, all of which may thus contribute more or less to skin pigmentation. It is to be note that different wavelengths of VL can have a synergistic or antagonistic effect on the same biological process.

2.1. The Activation of Chromophores by Visible Light

When a photon reaches the skin's surface, it can be reflected, scattered, or absorbed (Figure 1), but only absorbed light can cause photobiological changes by interacting with

chromophores [4]. Chromophores are photoreceptor molecules that can be activated and energized by photons to mediate biological effects [5]. Hemoglobin, cytochrome C oxidase (CCO), opsins (OPN) and melanin are the primary chromophores responsible for VL absorption, which is the premise and basis of the use of VL in laser therapy, intense pulsed therapy (IPL), and especially in low-level light therapy (LLLT) [6]. Chromophores absorb specific wavelengths of light, excite electrons to higher-energy states, and then activate second messengers such as ROS, Ca^{2+} , ATP, cAMP, and NO, followed by the modulation of subsequent cascades of signaling pathways related to fundamental activities such as migration and proliferation, protein synthesis and tissue repair, inflammation, anti-apoptosis and redox reactions [7].

The relation between chromophore activation by VL and pigmentation in melanocytes has also been proved [8]. One study demonstrated that shorter wavelengths of VL (415 nm, 50 J/cm²) can activate OPN3, which senses blue light and then activates CAMKII, CREB, ERK and p38, thus up-regulating MITF signaling and inducing potent and sustained hyperpigmentation [9]. However, the activation may be dose-dependent, for another study showed that, under much smaller doses, blue light (450 nm, 200 mJ/cm²) and green light (550 nm, 200 mJ/cm²) failed to initiate human OPN3, which can act as a negative regulator of melanogenesis when treated, by regulating the α -MSH-induced MC1R-mediated cAMP signaling [10]. Furthermore, Campiche et al. reported that LED blue light (450 nm, 60 J/cm²) resulted in changes in skin chromophores and signs of skin photoaging, including hyperpigmentation [11].

Different chromophores may exist in different layers of the skin. The depth of light radiation penetration is inversely proportional to the absorption rate and scattering rate, which are both inversely related to the wavelength [12]. Therefore, the longer the wavelength, the deeper the light can penetrate into the skin, meaning that VL can go even deeper than ultraviolet, and that VL in different wavelengths may activate chromophores present in different layers, resulting in diverse subsequent reactions.

Taken together, with deep penetration into the skin and multiple chromophore-mediated activities, VL has a profound and substantial impact on skin, including the epidermal melanin-unit system.

2.2. The Regulation of Skin Aging by Visible Light

Skin photoaging is a complex process that involves degenerative changes in the skin, such as mottled hyperpigmentation and decreased elasticity and laxity caused by exposure to sunlight. This process is caused by the dysfunction of the nucleus, mitochondria, and the extracellular matrix (ECM) resulting from DNA damage and ROS generation [13].

Considering that VL participates in inflammation, oxidative stress, and the production of matrix metalloproteinases (MMPs), which is thought to be one of the crucial factors participated in ECM degradation in the skin [14], it is closely involved in the regulation of skin aging. However, different types of VL affect skin aging by distinct mechanisms, primarily targeting fibroblasts [15]. For instance, blue and green light have been proved to precipitate photoaging by increasing ROS and MMP-1, decreasing collagen type I in human fibroblasts [16–19] and inducing the production of singlet oxygen, followed by nuclear DNA damage in epithelial cells [20]. Besides inducing oxidative stress, blue light has also demonstrated the ability to damage fibroblast mitochondrial functionality [21]. In contrast, yellow light has been shown to protect against skin aging by increasing collagen I in the dermis [22] and upregulating the expression of antioxidant enzymes, to reduce the generation of UVA-induced MMP-1, phosphorylated stress-activated protein kinase (pJNK) and ROS, in human fibroblasts [23]. Similarly, low red light has been shown to stimulate fibroblast growth, upregulating antioxidant-related genes in human fibroblasts, reducing ROS, down-regulating MMP and reversing collagen I degradation of skin cells, exerting anti-aging effects [15,24–26].

As mentioned above, dermal fibroblasts, as the major skin cell implicated in VL-induced skin photoaging, have been shown to engage in the signal crosstalk between dermal and epithelial cells related to pigmentary disorders, acting on melanocytes by releasing abundant proteins, cytokines and growth factors [27]. Therefore, pigmentation is closely related to photoaging, and some pigmentary disorders, such as melasma, have been considered as photoaging disorders [28]. One study also inferred that photooxidation may be involved in the mechanism of pigmentation after blue-light irradiation [21]. Hence, it is essential to enhance the comprehension of the mechanisms through which yellow and red light can potentially impede the photoaging process, while blue and green light may give rise to exacerbation.

2.3. The Disruption of the Skin Circadian Rhythm by Visible Light

It is now well established that circadian oscillators exist not only in the suprachiasmatic nucleus (SCN) of the hypothalamus, but also in peripheral tissues, including the skin [29]. Studies have demonstrated that VL can disrupt the normal circadian rhythm, negatively affecting skin homeostasis. Exposure to blue and green light emitted from computer screens significantly suppresses and delays nocturnal melatonin secretion, disrupting the sleep–wakefulness cycle [30–33]. In addition, red-light (631 nm) exposure was shown to potentially delay the circadian clock and the onset of sleep, inducing circadian resetting, despite small samples [34]. VL also regulates the skin's circadian rhythm by interacting directly with skin circadian clock genes, such as *PER1* and *BMAL1*, in peripheral pathways [35].

Circadian rhythm has intricate influences on physiological processes in the skin, including melanogenesis. Hardman et al. showed that silencing peripheral clock genes *BMAL1/PER1* in human skin and isolated melanocytes upregulated melanogenesis and thus increased melanin content [36]. A subsequent study further proved that the peripheral clock was crucial for well-organized melanin synthesis in normal melanocytes [37]. *BMAL1* also transcriptionally upregulates microphthalmia-associated transcription factor (MITF) and prevents UVB-induced DNA damage through enhanced melanin synthesis [38]. Since circadian disruption or altered clock genes can result in multiple skin diseases [39], considering its effect on melanin synthesis, it is highly possible that the circadian rhythm may participate in the process of skin pigment regulation, to some extent. And disruption of the circadian rhythm may accelerate skin aging, increase pigmentation and even induce pigmentary disorders [40].

3. Visible Light as an Enhancer of Skin Pigmentary Disorder

Different regions of the VL spectrum are more likely to contribute to distinct pigmentary disorders. While blue or green light is generally considered one of the significant causes of disorders of hyperpigmentation under most conditions, in some cases, yellow or red light may serve as a crucial facilitator for the development of hypopigmentary diseases (Figure 2).

Blue and green light may induce disorders of hyperpigmentation through several possible pathways: (a) direct interaction with OPN3; (b) photoaging aggravation; (c) circadian rhythm disruption; and (d) melanogenesis upregulation.

Yellow and red light impose possible stimulation on the onset and aggravation of vitiligo by affecting the viability and survival of melanocytes and the progression of melanin production. OPN3, opsin3; MITF, microphthalmia-associated transcription factor; TYR, tyrosinase; MC, melanocyte; ERK, extracellular regulated protein kinases; ROS, reactive oxygen species; MMP, matrix metalloproteinase; By figdraw.

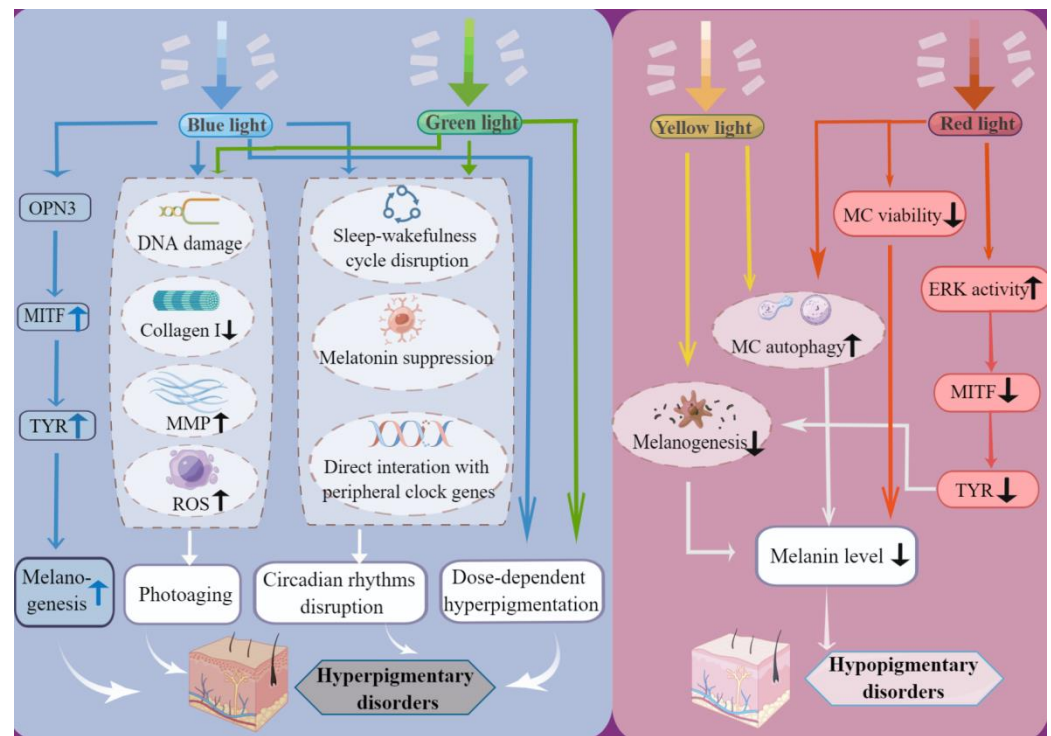


Figure 2. Pathologic effects of VL on pigmentary disorders.

3.1. Blue Light and Green Light Induce Hyperpigmentation

It is now well accepted that the short wavelength of VL plays a stimulatory role in hyperpigmentation. Multiple studies have shown that VL has synergistic effects with long-wavelength UVA1 on pigmentation [6,41,42]. VL-induced hyperpigmentation was found more potent and long-lasting than UVA1-induced hyperpigmentation in individuals with dark skins [6,43], even though the mechanism of hyperpigmentation caused by VL has been proved similar to that caused by UV [44]. Typically, three mechanisms are involved in the responsive reaction of melanocytes to VL, with increased melanin content: immediate pigment darkening (IPD), persistent pigment darkening (PPD), and delayed tanning (DT) [45]. IPD and PPD result from the oxidation of melanin precursors and the redistribution of melanin, while DT is associated with melanogenesis [46]. Randhawa et al. performed a series of ex vivo and clinical studies, and demonstrated that a single exposure to VL was ineffective in generating persistent pigmentation, while multiple exposure to VL induced PPD both in vivo and ex vivo [47]. In contrast, a classic study conducted by Mahmoud et al. demonstrated that a single dose of VL irradiation was enough to induce IPD and PPD in skin-type IV–VI individuals, and that higher doses (80–120 J/cm²) could induce DT [6]. Another study also revealed that both single high-dose (135 J/cm²) and repetitive (45 J/cm² over 5 consecutive days) exposure to blue light (450 nm) would result in IPD [48]. Those results indicate that VL, especially in the short wavelength, is able to induce apparent darkening via regulation of melanin redistribution, melanosome maturation or melanin synthesis.

While numerous studies have documented that irradiation with blue light and green light can induce a dose-dependent hyperpigmentation response [6,11,43,45,49–51], the skin phototype plays a crucial role in this process. For instance, it has been proved that blue light (453 nm, 18 J/cm²) can induce IPD in type I–III healthy skins [52], which is in line with the findings of Kleinpenning et al., who explored the clinical and histological effects of blue light (420 nm, 20 J/cm²) on normal skin types I–III. The authors observed transient melanogenesis and inexplicable vacuolization without melanocyte apoptosis [51]. Likewise, Moreiras et al. demonstrated that both blue light (450 nm) and green light (530 nm) induced melanin production in healthy human skin of types II and III ex vivo,

without any detectable increase in DNA damage or cell apoptosis, even under fairly high doses of exposure (140 J/cm^2) [53]. The same study evaluated melanin induction histologically in the epidermis of blue- and green-light-irradiated phototype I skin, although it was invisible, which is somewhat inconsistent with Mahmoud's findings that melanogenesis induced by blue light (495 nm) and green light (595 nm) at $8\text{--}480 \text{ J/cm}^2$ tends to occur in phototypes IV–VI skin, yet remains undetectable in lighter skin [6].

The clinical relevance of the capability of blue light and green light to induce pigmentation highlights the pathologic role of VL in photo-induced disorders of hyperpigmentation. It has been proposed that broad-spectrum sunscreens that do not adequately protect against VL fail to prevent worsening of post inflammatory hyperpigmentation (PIH) [54] and relapse of melasma [55,56]. A study involving 22 melasma patients showed that blue light induced melanogenesis both in the lesional and the neighboring skin, suggesting that blue light imposed a stimulatory effect on the onset and progression of melasma for the first time [57]. Although short-term exposure to blue light from electronic devices is not considered to exacerbate melasma [58], the low energy of artificial indoor VL is sufficient to induce hyperpigmentation in melasma patients [59]. Blue light emitted by the sun has also been proved to accelerate relapse of melasma [60].

3.2. Yellow Light and Red Light Induce or Aggravate Hypopigmentation

Yellow light inhibits melanogenesis and downregulates melanin content. In our former study, a dose-dependent inhibition of melanogenesis was observed, along with the induction of human epidermal melanocyte autophagy by yellow light irradiation (585 nm, $5\text{--}20 \text{ J/cm}^2$) [61]. On the other hand, 630 nm red light ($10\text{--}150 \text{ J/cm}^2$) was found to have no promotion effect on melanogenesis in a study by Duteil L et al. [43], and turned out to impose an inhibitory effect on melanin synthesis, both in vitro and in vivo. Additionally, 633 nm (96 J/cm^2) red light LED was reported to decrease melanin levels significantly after phototherapy of patients with acne vulgaris [62]. Similarly, 660 nm red light showed a depigmenting effect with downregulation of tyrosinase and MITF, due to increased ERK activity [63]. We previously revealed that red light (630 nm, $5\text{--}20 \text{ J/cm}^2$) might decrease cell viability and increase apoptosis of melanocytes [61], which might also lead to hypopigmentation.

Limited clinical studies could be found on the promoting effect of VL on hypopigmentary diseases. However, several case reports showed that yellow light and red light may potentially induce depigmentation. Lee's study showed that the melanin levels of the treated area in acne vulgaris patients slightly increased when treated with blue light (415 nm , 48 J/cm^2) LED for 20 min, twice a week, for a duration of four weeks, while they significantly decreased when treated with red light (633 nm , 96 J/cm^2) LED in the treated area [62]. What's more, a 41-year-old woman who had no previous history of vitiligo or halo nevus developed vitiligo patches on the treatment site after IPL treatment for rejuvenation [64]. Moreover, using a 585 nm pulsed dye laser to treat port-wine stains was reported to induce depigmented patches and cause the Koebner phenomenon in vitiligo patients [65,66], something which has also been reported after 755 nm laser (red light) treatment [67,68].

3.3. Measures to Protect against Cutaneous Damage by Visible Light

3.3.1. Exposure Reduction

As discussed previously, it is important to control light exposure, especially blue light from electronic devices and sunlight exposure. To prevent disruption of circadian rhythm by VL, reducing screen time by taking frequent breaks from long-period device usage and restricting artificial light at night are effective ways [69]. Simple but efficient physical methods for reducing sunlight exposure include seeking shade or staying indoors during peak hours, using a parasol and wearing photoprotective clothing and accessories, such as sunglasses or wide-brimmed hats [70], which have been proved to lower the chance of sunburn more significantly than photoprotection products on the market such

as sunscreen and antioxidants [71]. While, theoretically, using standard window glass such as reflective or tinted automobile windows and window films that filter out VL may also help in reducing VL exposure, most glasses are mainly used for UV reflection, with little effect on VL protection [72,73], and are awaiting more exploration from the VL-protection perspective.

3.3.2. Sunscreens

Over the past decade, sunscreens have undergone significant changes, due to the growing recognition that traditional broad-spectrum sunscreens with only organic or inorganic (ZnO and TiO₂) UV filters do not provide adequate protection against VL [54]. Tinted sunscreens have emerged as a promising solution, which combines UV filters with different concentrations and ratios of iron oxides and titanium dioxide to protect against VL for individuals of all skin types. The critical component of tinted sunscreens is Fe₂O₃, which has three different colors, depending on its oxidation state: yellow, red, or black [70]. Among them, yellow Fe₂O₃ offers the strongest protection against VL-induced hyperpigmentation [74]. Studies have demonstrated that iron-oxide-containing tinted sunscreen significantly lessens the development of VL-induced hyperpigmentation [75] and that the efficiency increases to over 93% with increasing iron oxide content [76]. It has also been proved to benefit individuals with hyperpigmentation disorders such as melasma and PIH [60,77–79]. Additionally, Fe₂O₃-containing products with multiple shades and tones can be used to cover pigmentary blemishes [78], making them a novel cosmetic-friendly strategy for full-spectrum photoprotection beyond the UV range, and with a profound influence on patients with pigmentary disorders.

3.3.3. Antioxidants

It has been estimated that 50% of ROS generation can be attributed to VL and infrared radiation [80], which may induce melanogenesis [81] and exacerbate pre-existing hyperpigmentation [82]. Antioxidants can mitigate the harm caused by VL through ROS neutralization or melanogenesis pathway regulations [80] (Table 1).

WH130, a kind of licorice extract, inhibits melanogenesis by suppressing tyrosinase activity, particularly when heated, making it a promising option for treating various disorders of hyperpigmentation, including brown spots, ephelides, and melasma [83]. French maritime pine bark extract (PBE), with its antioxidant property, has also proved to reduce VL-induced melanin synthesis *in vitro*, via inhibiting tyrosinase and other pigmentation-related mediators [84]. Furthermore, a clinical study comparing antioxidant-enriched sunscreens with tinted sunscreens showed that the former had comparable or even better efficacy than the latter [85]. Another study also demonstrated that topical antioxidants inhibited erythema and reduced pigmentation caused by VL and UVA1, suggesting that antioxidants may prevent the exacerbation of pigmentary disorders due to sunlight exposure [86].

On the other hand, multiple studies have provided theoretical evidence that antioxidants may attenuate hyperpigmentation by reducing oxidative damage and preventing VL-triggered photoaging. Sunscreens containing antioxidants have been shown to repair some clinical signs of photoaging [87]. Hydroxytyrosol from olive fruits prevents human keratinocytes and fibroblasts from blue-light-induced photoaging [18], while resveratrol can potentially scavenge ROS induced by blue light (415 nm) in fibroblasts [16]. Licochalcone A, a Nrf2 inducer, has been reported to reduce ROS formation *in vitro* and prevent intradermal carotenoid depletion *in vivo* [19]. Furthermore, polypodium leucotomos extract (PLE) may also offer protection against VL-induced photoaging. PLE treatment led to a significant decrease in VL-induced PPD and DT and a reduction in the markers for cellular damage [88]. PLE was also reported to prevent human dermal fibroblast damage and mitigate photoaging-related ECM degradation *in vivo* by reducing VL-induced MMP-1 [89,90]. Relatively, scientific and commercially available antioxidants against UV radiation are

much more abundant than those against VL [91]. Therefore, the potential of antioxidants targeting VL remains to be extensively explored.

Table 1. The hyperpigmentation attenuated functions of antioxidants targeting VL.

Antioxidants	Mechanisms	Origins	Objects
WH130	Inhibits melanogenesis by suppressing tyrosinase activity	Extract from Licorice (Wongam);	Murine melanoma B16F10 cells [83]
PBE	Reduces VL-induced melanin synthesis by reducing tyrosinase activity and decreasing ED1, and PPAR α , δ , and γ production	Extract from French maritime pine bark (<i>Pinus pinaster</i>)	Human melanocytes [84]
Resveratrol	Scavenges ROS induced by blue light (415 nm) LED in human fibroblast	Root extract from <i>Veratrum grandiflorum</i>	Human Skin Fibroblasts [16]
Hydroxytyrosol	Protects keratinocytes and fibroblasts from damage induced by blue light through preventing ROS formation, reducing MMP levels, preserving collagen type I production, and decreasing DNA damage	Extract from olive fruits	Human keratinocytes and fibroblasts [18]
Licochalcone A	Decreases VL-induced ROS formation in human fibroblast to a level equivalent to unirradiated fibroblast cells, or even below, in vitro, and prevents intradermal carotenoid depletion by VL irradiation in vivo	Root extract from Licorice (<i>Glycyrrhiza inflata</i>);	Human dermal fibroblasts and 10 healthy subjects with Fitzpatrick skin phototype II or III [19]
PLE	1. Decreases PPD and DT; 2. Decreases cyclooxygenase-2 and cell damage; 3. Prevents alterations in morphology, cell survival and cell cycle of human dermal fibroblasts and changes in the expression of MMP-1, CTSK, fibrillins 1 and 2 and elastin, caused by VL	Extract from <i>Polypodium leucotomos</i> ;	22 subjects with Fitzpatrick skin phototype IV–VI [88] Human dermal fibroblasts [90] 7 healthy subjects [89]
Carotenoid	Filters out high-energy blue-light rays	Diets	46 healthy subjects [92]
Flavonoid	Decreases photosensitivity of phospholipids to blue-light oxidative damage	Extract from green tea	Langmuir monolayers of 1,2-dipalmitoyl-sn-glycero-3-[phospho-rac-(1-glycerol) (sodium salt) (DPPG) [93]
Vitachelox	Protects human keratinocytes by reducing oxidative damage (protein carbonylation) induced by blue-light radiation.	A mixture of three natural extracts: grape (<i>Vitis vinifera</i>) seeds, green tea (<i>Camellia sinensis</i> green) leaves, and oak (<i>Quercus robur</i>)	Human keratinocytes [94]

4. Visible Light as a Therapeutic Option for Pigmentary Disorders

Visible-light therapy (VLT) is commonly used for various skin diseases, mostly as a second-line option. Likewise, it plays a primary or adjunctive role in the clinical management of pigmentary disorders. In the treatment of pigmentary disorders, there are three primary types of visible-light therapies utilized: laser, IPL, and LED therapy. Each type of light has unique features and mechanisms that cater to different skin conditions and disorders (Table 2).

Table 2. Differences between LED, Laser and IPL.

	Laser	IPL	LED
Intensity	High	High	Low
Pulse width	Unadjustable	Continuous and adjustable	Continuous and adjustable
Coherence	Coherent	Incoherent	Incoherent
Directionality	High/Single	Low/Multiple	Low/Multiple
Wavelength and Chromaticity	Monochromatic	500–1200 nm, Polychromatic	400–800 nm Polychromatic
Spot Size	Small point: <1 cm × 1 cm	Medium: 5 cm × 2 cm	Large: 30 cm × 30 cm
Mechanism	Selective Photothermolysis	Selective Photothermolysis	Photobiomodulation
Indications for pigmentary disorders	Benign epidermal pigmented lesions (ephelides, lentigo, PLH, café au lait macules, pigmented seborrheic keratoses. . .); Benign dermal pigmented lesions (CMN, nevus of Ota/Ito. . .); Mixed (epidermal/ dermal) pigmented lesions (Becker’s nevus, melasma, PIH); Tattoos; Vitiligo	Benign epidermal pigmented lesions (ephelides, lentigo, café au lait macules, . . .); mixed (epidermal/dermal) pigmented lesions (Becker’s nevus, melasma, PIH, poikiloderma of Civatte)	Melasma; Vitiligo
Adverse effects	Relatively common, mainly hyperpigmentation, sometimes scarring	Infrequent, sometimes erythema and hyperpigmentation	Rarely seen

4.1. Laser-Emitting Lights in the Visible Range

Visible lasers are a class of laser devices that emit light in the visible-spectrum region, containing pulsed dye laser (PDL), copper vapor laser, potassium titanyl phosphate laser (KTP), helium-neon (He-Ne), ruby laser, argon laser and krypton laser [95]. As a coherent light, laser has the advantages of high intensity, low divergence, and precise control over the amount and location of skin heating [96], which makes it an ideal method for treating skin diseases based on the principle of selective photothermolysis [97].

While PDL was initially designed for cutaneous vascular disorders, recent studies have shown that 595 nm and 607 nm PDL can also be used to treat benign epidermal pigmented lesions (EPLs) [98–104]. And 585 nm and 595 nm PDL have also been found to be effective in improving melasma lesions that exhibit increased vascularity, with or without the combination of other therapies [105–107].

Another type of laser that is highly specific for vascular lesions is copper vapor laser, emitting a dual wavelength comprising 10% 511 nm and 90% 578 nm, which is at the proximity of the absorption peak of hemoglobin [108]. Nonetheless, the dual-wavelength copper vapor laser shows great efficacy in eliminating congenital melanocytic nevi (CMN) [109], yet demonstrates less efficacy in treating melasma patients [108,110], except for those with pronounced vascular abnormality [111].

The KTP laser, also known as the (Q-switched) Nd:YAG double-frequency 532 nm laser, is another type of laser that has been proved effective in treating EPLs, such as ephelides or solar lentigines [112–115], physiological lip hyperpigmentation (PLH) [116,117], and even tattoos [118,119]. When combined with IPL, it has been successful in treating postoperative inflammatory hyperpigmentation [115].

The 633 nm He-Ne laser emitting red light is a popular choice for low-level light therapy (LLLT), and has been found to be effective for vitiligo. Yu et al. discovered that low-energy He-Ne lasers (632.8 nm) enhance melanocyte migration and proliferation, and even rescue damaged melanocytes, creating a positive microenvironment for repigmen-

tation [120]. The same group also investigated the molecular mechanism and biological effects of the low-energy He-Ne laser on pigment cells at different maturation stages. They found that the laser induced differentiation and mitochondrial biogenesis of primitive pigment cells through calcium-dependent mitochondrial retrograde signaling [121], as well as stimulating the differentiation of immature melanoblasts through enhanced pp125FAK expression and the melanogenesis of more mature melanoblasts [122]. Furthermore, they explored the role of the low-energy He-Ne laser in melanocytes, and demonstrated enhanced functional melanocyte proliferation via increased expression of $\alpha 2 \beta 1$ integrin and increased attachment to collagen IV [123]. These studies provide a solid theoretical basis for understanding how low-level laser therapy induces repigmentation in vitiligo. Clinical evidence also supports the application of the low-energy He-Ne laser in treating segmental-type vitiligo, with an effectiveness comparable to conventional therapies [120].

Interestingly, red light can also be used as an effective and safe modality for further depigmentation of vitiligo. The cosmetically disturbing remnants of normal pigmentation in patients with vitiligo whose skin has been almost depigmented on the whole can be removed by the Q-switched 694 nm Ruby laser (QSRL) [124]. The QSRL is particularly effective for treating benign pigmented diseases, such as tattoos, nevus of Ota and ephelides, due to its high absorption by melanin [125–130]. While QSRL was previously believed to be ineffective in treating melasma [131], recent studies with small sample sizes have demonstrated its efficacy [132–134]. Similarly, the Q-switched 755 nm Alexandrite laser (QSAL) can also be used to treat a variety of superficial and deep hyperpigmented diseases such as nevus of Ota/Ito, tattoos, café au lait macules and melasma [129,135–137]. QSRL and QSAL are considered the best choices as phototherapy for treating dermal pigmented lesions by Bogdan et al. [102]. Yet the newly developed picosecond laser with higher efficiency in pigment removal and less thermal damage is worthy of consideration [138,139].

Taken together, the visible laser can attenuate or eliminate hyperpigmentation to a certain extent, except for the He-Ne laser, which is usually used for depigmentation. Among them, the PDL and copper vapor laser are classic modalities for vascular lesions, with recently discovered use in benign EPLs and melasma with a vascular component. KTP can tackle both epidermal and dermal hyperpigmentation, with a better effect on the former. QSRL and QSAL are best applied in dermal hyperpigmentation, but also have solid efficacy on EPLs, with new findings relating to melasma treatment that may renew the conventional views.

4.2. Intense Pulsed Light (IPL)

IPL is a polychromatic and noncoherent light released by a high-energy tritium flash-lamp under high voltage, featuring high intensity, a relatively concentrated wavelength, and a wide and tunable pulse width. The IPL spectrum primarily falls in the range of 500–1200 nm, and can be selectively filtered by filters based on the skin type and lesions. IPL also works based on the principles of selective photothermolysis effects, as does laser [140]. Clinical studies have demonstrated that IPL is capable of effectively decreasing melanin production and accumulation at the cellular level, making it a suitable treatment option for various types of hyperpigmented skin conditions [141].

IPL has been proven effective in treating lentiginos, ephelides, poikiloderma of Civatte and other epidermal hyperpigmentation, as well as benign melanocytic nevi such as Becker's nevus [98,142–147]. However, it should be noted that the Q-switched laser still remains the preferred choice in light therapy for treating benign pigmented lesions. Furthermore, IPL is not a viable solution for tattoo removal, as it lacks the ability to perform Q-switching in incoherent light sources [148].

In the treatment of melasma, IPL has demonstrated superior efficacy when compared to laser treatment. In a split-face comparative study conducted by Hassan et al., IPL was observed to more effectively lighten epidermal melasma and melasma lesions with vascular alteration, in comparison to PDL [106]. Li et al. also demonstrated that IPL was ideal for melasma treatment with minimal and acceptable adverse events [149], which

is consistent with Yi's conclusion [150]. In addition to skin brightening, IPL has been popularly employed for skin rejuvenation, owing to its remarkable efficacy in addressing photoaging concerns [151].

4.3. Light-Emitting Diodes (LEDs)

LEDs emit incoherent light with a narrow spectrum and low intensity, which induces a mild effect on cells for regulating biological activity, rather than a thermal or exfoliative effect. This process is referred to as photomodulation or photobiomodulation (PBM), also known as LLLT [152]. A vast array of LED semiconductor materials has been available at lower wavelengths, and research over the past decade suggests that LED therapy is more suitable than laser therapy for LLLT, due to its mild output and convenient accessibility [153]. LED therapy using VL for pigmentary disorders has been a controversial approach, but recent studies have shed light on its potential application in melasma (Figure 3).

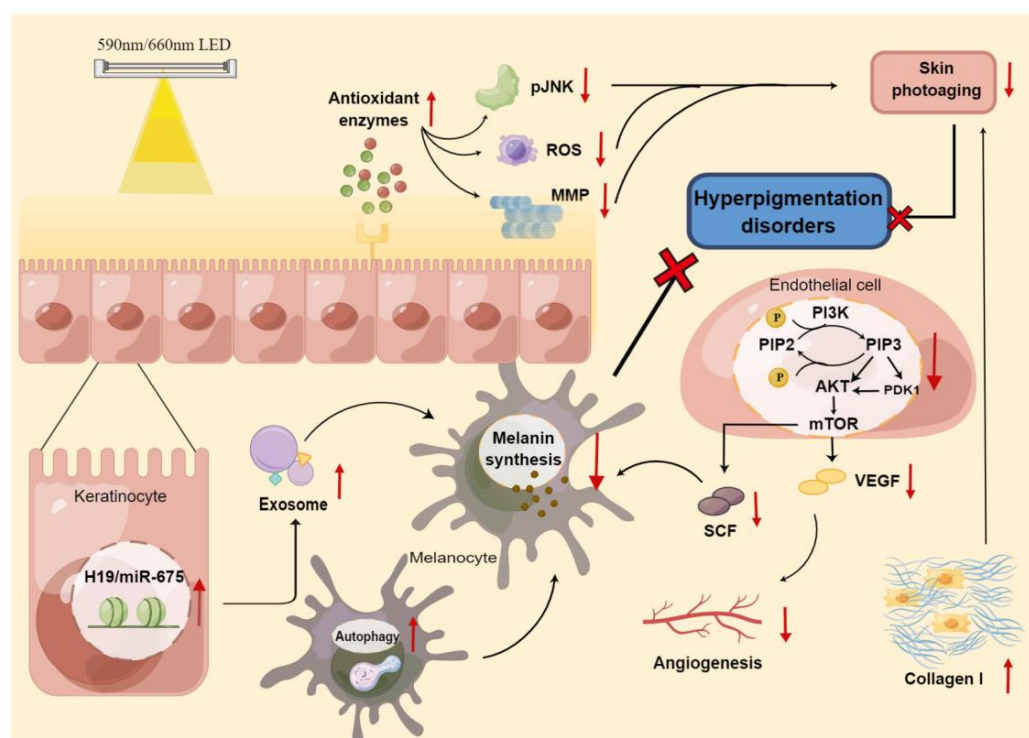


Figure 3. The mechanism of LED phototherapy for hyperpigmentation disorders.

As previously mentioned, 585 nm yellow LED light has been shown to inhibit melanogenesis in melanocytes by the inducing of autophagy [61]. This was further explored by our later study, which demonstrated that irradiation with 585 nm LEDs resulted in the containment of melanin synthesis by upregulating H19 and its exosomal miR-675 derived from keratinocytes in vitro [154]. Our subsequent study further demonstrated in vivo and in vitro that 590 nm yellow LED decreased the secretion of melanogenic factor and reduced the angiogenesis of the human microvascular endothelial cell (HMEC-1) by dampening the PI3K/AKT/mTOR signaling pathway, thus prominently attenuating erythema and hyperpigmentation in melasma [155]. This series of experimental data is consistent with Mpofana's study, in which 633 nm-LED combined with 830 nm-LED exposure significantly ameliorated melasma in patients with skin types V and VI [156]. In addition, our recent clinical trial focusing on melasma patients who underwent 590 nm LED treatment further proved that home-based 590 nm LEDs exhibited a similar efficacy and safety as in-hospital 1064 nm QSNY, with higher portability and lower cost [157].

From another point of view, phototherapy using LED to treat skin photoaging is increasingly prevalent. Lee et al. conducted a prospective split-face clinical study on LED phototherapy for skin rejuvenation, and indicated an altered enzymatic activity related to dermal matrix remodeling, as well as a reduction in melanin content after irradiation with 633 nm red LED [158]. Moreover, 590 nm-LED or 660 nm-LED therapy increased collagen and decreased MMP-1 activity in the dermis, with pigmentation reduction [22,23,25,159], suggesting a novel perspective for LED therapy to tackle melasma, which is now defined as a photoaging disorder [28].

While yellow and red LEDs are promising for treating disorders of hyperpigmentation, LED blue light has been applied for vitiligo repigmentation. Research indicates that blue LED, combined with *Buddleja officinalis*, can be used to treat vitiligo through induced melanin production by promoting melanogenic signaling, in addition to CREB/MITF/TYR pathways [160]. A retrospective study also demonstrated that 417 nm blue LED induced repigmentation in 30 patients with localized vitiligo, of varying ages and different skin types [161]. Despite the relatively small sample, these results encouraged the utilization of LED on melanin-deficiency skin diseases.

Of note, LED treatments have the special advantages of high safety and convenience with fewer side effects, yet well-designed studies with larger sample sizes and repeated measures of response are sorely lacking and highly required.

LED can be applied in disorders of hyperpigmentation treatment by directly affecting melanin production through various pathways or by alleviating the photoaging process, including antioxidant enzyme and collagen I production. Red cross refers to inhibition; Red arrows refer to upregulation or downregulation. ROS, reactive oxygen species; MMP, matrix metalloproteinase; PIP, phosphatidylinositol phosphate; PI3K, phosphatidylinositol 3-kinase; mTOR, mammalian target of rapamycin; SCF, stem cell factor; VEGF, vascular endothelial growth factor; By figdraw.

5. Perspectives

The threat posed by hyperpigmented or depigmented lesions to patients and society as a whole prompts deeper and more extensive research from angles beyond the traditional etiology and therapeutics. VL has thus emerged as a rapidly evolving field in photomedicine, and here we collated and clarified the detrimental and beneficial impacts of VL on melanocyte biology and pigmentary disorders, hoping to better instruct the prevention and treatment strategies of refractory pigmentary disorders in clinical practice. Despite this, studies attempting to determine the effects of VL on skin pigmentary disorders are still woefully inadequate and require further exploration, on multiple levels.

Firstly, the harmful effects of VL, including skin photoaging and circadian disruption, highlight the importance of proper protection against VL. As the evidence continues to mount, additional research on VL photoprotection is needed for sunscreens and antioxidants. Moreover, greater attention must be paid during medical treatments to take into account intensity, dose, exposed area, exposure duration, expose frequency, operation mode, and skin phototype-dependent differences in the pigmentary response of VL.

Secondly, despite some limited clinical evidence regarding the inducement of vitiligo by red or yellow light, the exact pathologic role of VL in depigmenting diseases has not been clearly determined. More fundamental and clinical studies are needed to clarify the precise role of VL in disorders of hypopigmentation.

Thirdly, in the last 5–8 years, the progress and development of pigment removal using QSRL has been slow, due to the lack of clinical and in vivo research. It is crucial to conduct studies with larger samples and more-strictly conducted procedures, to confirm the effectiveness of QSRL on melasma.

Last but not least, it is worth noting that LED, particularly yellow light, has exhibited great potential for melasma treatment, with the emerging fundamental and clinical studies. Considering its portability and economic applicability, LED yellow light is highly promising as a therapeutic alternative for treating melasma. There is a need for further clinical research,

though, to determine the specific benefits of LED in treating other pigmentary disorders besides melasma, and to thus open up new fields of investigation and markets for both skin darkening and skin lightening.

In conclusion, like a double-edged sword, VL plays distinct roles in the onset, progression, and treatment in skin pigmentary disorders under different parameters and modes, targeting different skin-phototype individuals. More basic and clinical studies are merited to explore the precise mechanisms of pigment metabolism through VL regulation, which may provide a scientific basis for more effective prevention and management of photo-aggravated pigmentary disorders.

Author Contributions: Conceptualization, C.Z. and L.X.; methodology, X.H.; software, X.D.; validation, S.J. and L.C.; formal analysis, S.J.; investigation, X.H.; resources, C.Z.; data curation, S.J.; writing—original draft preparation, X.H.; writing—review and editing, X.H. and S.J.; visualization, S.J.; supervision, L.X.; project administration, C.Z.; funding acquisition, C.Z. All authors have read and agreed to the published version of the manuscript.

Funding: This work was supported by the National Natural Science Foundation of China (No. 82173421), the Natural Science Foundation of Shanghai (No. 23ZR1408600) and the Medical innovation research special project of Shanghai Science and Technology Commission (No. 20Y11905700).

Conflicts of Interest: The authors declare no conflict of interest.

References

- Diffey, B.L. What is light? *Photodermatol. Photoimmunol. Photomed.* **2002**, *18*, 68–74. [[CrossRef](#)] [[PubMed](#)]
- Austin, E.; Geisler, A.N.; Nguyen, J.; Kohli, I.; Hamzavi, I.; Lim, H.W.; Jagdeo, J. Visible light. Part I: Properties and cutaneous effects of visible light. *J. Am. Acad. Dermatol.* **2021**, *84*, 1219–1231. [[CrossRef](#)] [[PubMed](#)]
- Ko, D.; Wang, R.F.; Ozog, D.; Lim, H.W.; Mohammad, T.F. Disorders of hyperpigmentation. Part II. Review of management and treatment options for hyperpigmentation. *J. Am. Acad. Dermatol.* **2023**, *88*, 291–320. [[CrossRef](#)]
- Zanolli, M. The modern paradigm of phototherapy. *Clin. Dermatol.* **2003**, *21*, 398–406. [[CrossRef](#)] [[PubMed](#)]
- Mahmoud, B.H.; Hexsel, C.L.; Hamzavi, I.H.; Lim, H.W. Effects of Visible Light on the Skin. *Photochem. Photobiol.* **2008**, *84*, 450–462. [[CrossRef](#)]
- Mahmoud, B.H.; Ruvolo, E.; Hexsel, C.L.; Liu, Y.; Owen, M.R.; Kollias, N.; Lim, H.W.; Hamzavi, I.H. Impact of long-wavelength UVA and visible light on melanocompetent skin. *J. Investig. Dermatol.* **2010**, *130*, 2092–2097. [[CrossRef](#)]
- de Freitas, L.F.; Hamblin, M.R. Proposed Mechanisms of Photobiomodulation or Low-Level Light Therapy. *IEEE J. Sel. Top. Quantum Electron.* **2016**, *22*, 348–364. [[CrossRef](#)]
- Setty, S.R. Opsin3-A Link to Visible Light-Induced Skin Pigmentation. *J. Investig. Dermatol.* **2018**, *138*, 13–15. [[CrossRef](#)]
- Regazzetti, C.; Sormani, L.; Debayle, D.; Bernerd, F.; Tulic, M.K.; De Donatis, G.M.; Chignon-Sicard, B.; Rocchi, S.; Passeron, T. Melanocytes Sense Blue Light and Regulate Pigmentation through Opsin-3. *Investig. Dermatol.* **2018**, *138*, 171–178. [[CrossRef](#)]
- Ozdeslik, R.N.; Olinski, L.E.; Trieu, M.M.; Oprian, D.D.; Oancea, E. Human nonvisual opsin 3 regulates pigmentation of epidermal melanocytes through functional interaction with melanocortin 1 receptor. *Proc. Natl. Acad. Sci. USA* **2019**, *116*, 11508–11517. [[CrossRef](#)]
- Campiche, R.; Curpen, S.J.; Lutchmanen-Kolanthan, V.; Gougeon, S.; Cherel, M.; Laurent, G.; Gempeler, M.; Schuetz, R. Pigmentation effects of blue light irradiation on skin and how to protect against them. *Int. J. Cosmet. Sci.* **2020**, *42*, 399–406. [[CrossRef](#)] [[PubMed](#)]
- Anderson, R.R.; Parrish, J.A. The Optics of Human Skin. *J. Investig. Dermatol.* **1981**, *77*, 13–19. [[CrossRef](#)] [[PubMed](#)]
- Wang, A.S.; Dreesen, O. Biomarkers of Cellular Senescence and Skin Aging. *Front. Genet.* **2018**, *9*, 247. [[CrossRef](#)] [[PubMed](#)]
- Liebel, F.; Kaur, S.; Ruvolo, E.; Kollias, N.; Southall, M.D. Irradiation of skin with visible light induces reactive oxygen species and matrix-degrading enzymes. *J. Investig. Dermatol.* **2012**, *132*, 1901–1907. [[CrossRef](#)] [[PubMed](#)]
- Pourang, A.; Tisack, A.; Ezekwe, N.; Torres, A.E.; Kohli, I.; Hamzavi, I.H.; Lim, H.W. Effects of visible light on mechanisms of skin photoaging. *Photodermatol. Photoimmunol. Photomed.* **2022**, *38*, 191–196. [[CrossRef](#)] [[PubMed](#)]
- Mamalis, A.; Koo, E.; Jagdeo, J. Resveratrol Prevents Reactive Oxygen Species-Induced Effects of Light-Emitting Diode-Generated Blue Light in Human Skin Fibroblasts. *Dermatol. Surg.* **2016**, *42*, 727–732. [[CrossRef](#)] [[PubMed](#)]
- Nakashima, Y.; Ohta, S.; Wolf, A.M. Blue light-induced oxidative stress in live skin. *Free. Radic. Biol. Med.* **2017**, *108*, 300–310. [[CrossRef](#)]
- Avola, R.; Graziano, A.C.E.; Pannuzzo, G.; Bonina, F.; Cardile, V. Hydroxytyrosol from olive fruits prevents blue-light-induced damage in human keratinocytes and fibroblasts. *J. Cell. Physiol.* **2019**, *234*, 9065–9076. [[CrossRef](#)]
- Mann, T.; Eggers, K.; Rippke, F.; Tesch, M.; Buerger, A.; Darvin, M.E.; Schanzer, S.; Meinke, M.C.; Lademann, J.; Kolbe, L. High-energy visible light at ambient doses and intensities induces oxidative stress of skin—Protective effects of the antioxidant and Nrf2 inducer Licochalcone A in vitro and in vivo. *Photodermatol. Photoimmunol. Photomed.* **2020**, *36*, 135–144. [[CrossRef](#)]

20. Chiarelli-Neto, O.; Ferreira, A.S.; Martins, W.K.; Pavani, C.; Severino, D.; Faião-Flores, F.; Maria-Engler, S.S.; Aliprandini, E.; Martinez, G.R.; Di Mascio, P.; et al. Melanin photosensitization and the effect of visible light on epithelial cells. *PLoS ONE* **2014**, *9*, e113266. [[CrossRef](#)]
21. Lorrio, S.; Rodríguez-Luna, A.; Delgado-Wicke, P.; Mascaraque, M.; Gallego, M.; Pérez-Davó, A.; González, S.; Juarranz, Á. Protective Effect of the Aqueous Extract of *Deschampsia antarctica* (EDAFENCE®) on Skin Cells against Blue Light Emitted from Digital Devices. *Int. J. Mol. Sci.* **2020**, *21*, 988. [[CrossRef](#)] [[PubMed](#)]
22. Lan CC, E.; Ho, P.Y.; Wu, C.S.; Yang, R.C.; Yu, H.S. LED 590 nm photomodulation reduces UVA-induced metalloproteinase-1 expression via upregulation of antioxidant enzyme catalase. *J. Dermatol. Sci.* **2015**, *78*, 125–132. [[CrossRef](#)] [[PubMed](#)]
23. Weiss, R.A.; McDaniel, D.H.; Geronemus, R.G.; Weiss, M.A. Clinical trial of a novel non-thermal LED array for reversal of photoaging: Clinical, histologic, and surface profilometric results. *Lasers Surg. Med.* **2005**, *36*, 85–91. [[CrossRef](#)] [[PubMed](#)]
24. You, H.R.; Kim, S.H.; Yun, S.J.; Lee, S.C.; Lee, J.B. Skin photorejuvenation effects of light-emitting diodes (LEDs): A comparative study of yellow and red LEDs in vitro and in vivo. *Clin. Exp. Dermatol.* **2016**, *41*, 798–805.
25. Barolet, D.; Roberge, C.J.; Auger, F.A.; Boucher, A.; Germain, L. Regulation of skin collagen metabolism in vitro using a pulsed 660 nm LED light source: Clinical correlation with a single-blinded study. *J. Investig. Dermatol.* **2009**, *129*, 2751–2759. [[CrossRef](#)]
26. Song, S.; Zhang, Y.; Fong, C.-C.; Tsang, C.-H.; Yang, Z.; Yang, M. cDNA microarray analysis of gene expression profiles in human fibroblast cells irradiated with red light. *J. Investig. Dermatol.* **2003**, *120*, 849–857. [[CrossRef](#)]
27. Wang, Y.; Viennet, C.; Robin, S.; Berthon, J.-Y.; He, L.; Humbert, P. Precise role of dermal fibroblasts on melanocyte pigmentation. *J. Dermatol. Sci.* **2017**, *88*, 159–166. [[CrossRef](#)]
28. Passeron, T.; Picardo, M. Melasma, a photoaging disorder. *Pigment. Cell Melanoma Res.* **2018**, *31*, 461–465. [[CrossRef](#)]
29. Lyons, A.B.; Moy, L.; Moy, R.; Tung, R. Circadian Rhythm and the Skin: A Review of the Literature. *J. Clin. Aesthetic Dermatol.* **2019**, *12*, 42–45.
30. Gooley, J.J.; Rajaratnam, S.M.W.; Brainard, G.C.; Kronauer, R.E.; Czeisler, C.A.; Lockley, S.W. Spectral responses of the human circadian system depend on the irradiance and duration of exposure to light. *Sci. Transl. Med.* **2010**, *2*, 31ra33. [[CrossRef](#)]
31. Cajochen, C.; Frey, S.; Anders, D.; Späti, J.; Bues, M.; Pross, A.; Mager, R.; Wirz-Justice, A.; Stefani, O. Evening exposure to a light-emitting diodes (LED)-backlit computer screen affects circadian physiology and cognitive performance. *J. Appl. Physiol.* **2011**, *110*, 1432–1438. [[CrossRef](#)] [[PubMed](#)]
32. Green, A.; Cohen-Zion, M.; Haim, A.; Dagan, Y. Evening light exposure to computer screens disrupts human sleep, biological rhythms, and attention abilities. *Chronobiol. Int.* **2017**, *34*, 855–865. [[CrossRef](#)] [[PubMed](#)]
33. Sletten, T.L.; Revell, V.L.; Middleton, B.; Lederle, K.A.; Skene, D.J. Age-related changes in acute and phase-advancing responses to monochromatic light. *J. Biol. Rhythm.* **2009**, *24*, 73–84. [[CrossRef](#)] [[PubMed](#)]
34. Ho Mien, I.; Chua EC, P.; Lau, P.; Tan, L.C.; Lee IT, G.; Yeo, S.C.; Tan, S.S.; Gooley, J.J. Effects of exposure to intermittent versus continuous red light on human circadian rhythms, melatonin suppression, and pupillary constriction. *PLoS ONE* **2014**, *9*, e96532. [[CrossRef](#)] [[PubMed](#)]
35. Dong, K.; Goyarts, E.C.; Pelle, E.; Trivero, J.; Pernodet, N. Blue light disrupts the circadian rhythm and create damage in skin cells. *Int. J. Cosmet. Sci.* **2019**, *41*, 558–562. [[CrossRef](#)] [[PubMed](#)]
36. Hardman, J.A.; Tobin, D.J.; Haslam, I.S.; Farjo, N.; Farjo, B.; Al-Nuaimi, Y.; Grimaldi, B.; Paus, R. The peripheral clock regulates human pigmentation. *J. Investig. Dermatol.* **2015**, *135*, 1053–1064. [[CrossRef](#)]
37. Poletini, M.O.; de Assis, L.V.M.; Moraes, M.N.; Castrucci, A.M.d.L. Estradiol differently affects melanin synthesis of malignant and normal melanocytes: A relationship with clock and clock-controlled genes. *Mol. Cell. Biochem.* **2016**, *421*, 29–39. [[CrossRef](#)]
38. Sarkar, S.; Porter, K.I.; Dakup, P.P.; Gajula, R.P.; Koritala, B.S.C.; Hylton, R.; Kemp, M.G.; Wakamatsu, K.; Gaddameedhi, S. Circadian clock protein BMAL1 regulates melanogenesis through MITF in melanoma cells. *Pigment. Cell Melanoma Res.* **2021**, *34*, 955–965. [[CrossRef](#)]
39. Duan, J.; Greenberg, E.N.; Karri, S.S.; Andersen, B. The circadian clock and diseases of the skin. *FEBS Lett.* **2021**, *595*, 2413–2436. [[CrossRef](#)]
40. Suiythimeatregor, O.; Yang, C.; Ma, Y.; Liu, W. Direct and Indirect Effects of Blue Light Exposure on Skin: A Review of Published Literature. *Ski. Pharmacol. Physiol.* **2022**, *35*, 305–318. [[CrossRef](#)]
41. Kohli, I.; Chaowattanapanit, S.; Mohammad, T.; Nicholson, C.; Fatima, S.; Jacobsen, G.; Kollias, N.; Lim, H.; Hamzavi, I. Synergistic effects of long-wavelength ultraviolet A1 and visible light on pigmentation and erythema. *Br. J. Dermatol.* **2018**, *178*, 1173–1180. [[CrossRef](#)]
42. Kohli, I.; Braunberger, T.L.; Nahhas, A.F.; Mirza, F.N.; Mokhtari, M.; Lyons, A.B.; Kollias, N.; Ruvolo, E.; Lim, H.W.; Hamzavi, I.H. Long-wavelength Ultraviolet A1 and Visible Light Photoprotection: A Multimodality Assessment of Dose and Response. *Photochem. Photobiol.* **2020**, *96*, 208–214. [[CrossRef](#)]
43. Duteil, L.; Cardot-Leccia, N.; Queille-Roussel, C.; Maubert, Y.; Harmelin, Y.; Boukari, F.; Ambrosetti, D.; Lacour, J.; Passeron, T. Differences in visible light-induced pigmentation according to wavelengths: A clinical and histological study in comparison with UVB exposure. *Pigment. Cell Melanoma Res.* **2014**, *27*, 822–826. [[CrossRef](#)] [[PubMed](#)]
44. Ramasubramaniam, R.; Roy, A.; Sharma, B.; Nagalakshmi, S. Are there mechanistic differences between ultraviolet and visible radiation induced skin pigmentation? *Photochem. Photobiol. Sci.* **2011**, *1*, 1887–1893. [[CrossRef](#)]
45. Sklar, L.R.; Almutawa, F.; Lim, H.W.; Hamzavi, I. Effects of ultraviolet radiation, visible light, and infrared radiation on erythema and pigmentation: A review. *Photochem. Photobiol. Sci.* **2013**, *12*, 54–64. [[CrossRef](#)] [[PubMed](#)]

46. Schutz, R. Blue Light and the Skin. *Curr. Probl. Dermatol.* **2021**, *55*, 354–373. [[PubMed](#)]
47. Randhawa, M.; Seo, I.; Liebel, F.; Southall, M.D.; Kollias, N.; Ruvolo, E. Visible Light Induces Melanogenesis in Human Skin through a Photoadaptive Response. *PLoS ONE* **2015**, *10*, e0130949. [[CrossRef](#)]
48. Schütz, R.; Vollhardt, J.; Rudolph, T. clinical effects of LED blue light on skin pigmentation and carotenoid depletion. *J. Investig. Dermatol.* **2019**, *5*, 130. [[CrossRef](#)]
49. Kleinpenning, M.M.; Otero, M.E.; Van Erp PE, J.; Gerritsen MJ, P.; Van De Kerkhof PC, M. Efficacy of blue light vs. red light in the treatment of psoriasis: A double-blind, randomized comparative study. *J. Eur. Acad. Dermatol. Venereol.* **2012**, *26*, 219–225. [[CrossRef](#)]
50. Jo, H.L.; Jung, Y.; Suh, B.; Cho, E.; Kim, K.; Kim, E. Clinical evaluation method for blue light (456 nm) protection of skin. *J. Cosmet. Dermatol.* **2020**, *19*, 2438–2443. [[CrossRef](#)]
51. Kleinpenning, M.M.; Smits, T.; Frunt, M.H.A.; van Erp, P.E.J.; van de Kerkhof, P.C.M.; Gerritsen, R.M.J.P. Clinical and histological effects of blue light on normal skin. *Photodermatol. Photoimmunol. Photomed.* **2010**, *26*, 16–21. [[CrossRef](#)] [[PubMed](#)]
52. Falcone, D.; Uzunbajakava, N.E.; van Abeelen, F.; Oversluizen, G.; Peppelman, M.; van Erp, P.E.J.; van de Kerkhof, P.C.M. Effects of blue light on inflammation and skin barrier recovery following acute perturbation. Pilot study results in healthy human subjects. *Photodermatol. Photoimmunol. Photomed.* **2018**, *34*, 184–193. [[CrossRef](#)] [[PubMed](#)]
53. Moreiras, H.; O'Connor, C.; Bell, M.; Tobin, D.J. Visible light and human skin pigmentation: The importance of skin phototype. *Exp. Dermatol.* **2021**, *30*, 1324–1331. [[CrossRef](#)] [[PubMed](#)]
54. Lyons, A.B.; Trullas, C.; Kohli, I.; Hamzavi, I.H.; Lim, H.W. Photoprotection beyond ultraviolet radiation: A review of tinted sunscreens. *J. Am. Acad. Dermatol.* **2021**, *84*, 1393–1397. [[CrossRef](#)] [[PubMed](#)]
55. Sheth, V.M.; Pandya, A.G. Melasma: A comprehensive update: Part I. *J. Am. Acad. Dermatol.* **2011**, *65*, 689–697. [[CrossRef](#)] [[PubMed](#)]
56. Rivas, S.; Pandya, A.G. Treatment of melasma with topical agents, peels and lasers: An evidence-based review. *Am. J. Clin. Dermatol.* **2013**, *14*, 359–376. [[CrossRef](#)] [[PubMed](#)]
57. Alcantara, G.P.; Esposito, A.C.C.; Olivatti, T.O.F.; Yoshida, M.M.; Miot, H.A. Evaluation of ex vivo melanogenic response to UVB, UVA, and visible light in facial melasma and unaffected adjacent skin. *An. Bras. Dermatol.* **2020**, *95*, 684–690. [[CrossRef](#)]
58. Duteil, L.; Queille-Roussel, C.; Lacour, J.-P.; Montaudié, H.; Passeron, T. Short-term exposure to blue light emitted by electronic devices does not worsen melasma. *J. Am. Acad. Dermatol.* **2020**, *83*, 913–914. [[CrossRef](#)]
59. Verallo-Rowell, V.M.; Pua, J.M.; Bautista, D. Visible light photopatch testing of common photocontactants in female filipino adults with and without melasma: A cross-sectional study. *J. Drugs Dermatol.* **2008**, *7*, 149–156.
60. Boukari, F.; Jourdan, E.; Fontas, E.; Montaudié, H.; Castela, E.; Lacour, J.-P.; Passeron, T. Prevention of melasma relapses with sunscreen combining protection against UV and short wavelengths of visible light: A prospective randomized comparative trial. *J. Am. Acad. Dermatol.* **2015**, *72*, 189–190.e1. [[CrossRef](#)]
61. Chen, L.; Xu, Z.; Jiang, M.; Zhang, C.; Wang, X.; Xiang, L. Light-emitting diode 585nm photomodulation inhibiting melanin synthesis and inducing autophagy in human melanocytes. *J. Dermatol. Sci.* **2018**, *89*, 11–18. [[CrossRef](#)] [[PubMed](#)]
62. Lee, S.Y.; You, C.E.; Park, M.Y. Blue and red light combination LED phototherapy for acne vulgaris in patients with skin phototype IV. *Lasers Surg. Med.* **2007**, *39*, 180–188. [[CrossRef](#)] [[PubMed](#)]
63. Oh, C.T.; Kwon, T.R.; Choi, E.J.; Kim, S.R.; Seok, J.; Mun, S.K.; Yoo, K.H.; Choi, Y.S.; Choi, S.Y.; Kim, B.J. Inhibitory effect of 660-nm LED on melanin synthesis in in vitro and in vivo. *Photodermatol. Photoimmunol. Photomed.* **2017**, *33*, 49–57. [[CrossRef](#)] [[PubMed](#)]
64. Shin, J.U.; Roh, M.R.; Lee, J.H. Vitiligo following intense pulsed light treatment. *J. Dermatol.* **2010**, *37*, 674–676. [[CrossRef](#)] [[PubMed](#)]
65. Sommer, S.; Sheehan-Dare, R.A. The Koebner phenomenon in vitiligo following treatment of a port-wine stain naevus by pulsed dye laser. *Br. J. Dermatol.* **1998**, *138*, 200–201. [[CrossRef](#)] [[PubMed](#)]
66. Mazzotta, F.; Bonifazi, E.; Greco, I. Koebner phenomenon in vitiligo after treatment with dye laser. *Eur. J. Pediatr. Dermatol.* **2006**, *3*, 169.
67. van Geel, N.; Speeckaert, R.; Mollet, I.; De Schepper, S.; De Wolf, J.; Tjin, E.P.; Luiten, B.M.; Lambert, J.; Brochez, L. In vivo vitiligo induction and therapy model: Double-blind, randomized clinical trial. *Pigment. Cell Melanoma Res.* **2012**, *25*, 57–65. [[CrossRef](#)]
68. Alkhalifah, A. A Case Report of Vitiligo Induced by Alexandrite Hair Removal Laser. *Case Rep. Dermatol.* **2021**, *13*, 521–524. [[CrossRef](#)]
69. Kumari, J.; Das, K.; Babaei, M.; Rokni, G.R.; Goldust, M. The impact of blue light and digital screens on the skin. *J. Cosmet. Dermatol.* **2023**, *22*, 1185–1190. [[CrossRef](#)]
70. Geisler, A.N.; Austin, E.; Nguyen, J.; Hamzavi, I.; Jagdeo, J.; Lim, H.W. Visible light. Part II: Photoprotection against visible and ultraviolet light. *J. Am. Acad. Dermatol.* **2021**, *84*, 1233–1244. [[CrossRef](#)]
71. Linos, E.; Keiser, E.; Fu, T.; Colditz, G.; Chen, S.; Tang, J.Y. Hat, shade, long sleeves, or sunscreen? Rethinking US sun protection messages based on their relative effectiveness. *Cancer Causes Control* **2011**, *22*, 1067–1071. [[CrossRef](#)] [[PubMed](#)]
72. Almutawa, F.; Vandal, R.; Wang, S.Q.; Lim, H.W. Current status of photoprotection by window glass, automobile glass, window films, and sunglasses. *Photodermatol. Photoimmunol. Photomed.* **2013**, *29*, 65–72. [[CrossRef](#)]
73. Tuchinda, C.; Srivannaboon, S.; Lim, H.W. Photoprotection by window glass, automobile glass, and sunglasses. *J. Am. Acad. Dermatol.* **2006**, *54*, 845–854. [[CrossRef](#)] [[PubMed](#)]

74. Ruvolo, E.; Fair, M.; Hutson, A.; Liebel, F. Photoprotection against visible light-induced pigmentation. *Int. J. Cosmet. Sci.* **2018**, *40*, 589–595. [[CrossRef](#)]
75. Martini, A.P.M.; Campos, P.M.B.G.M. Influence of visible light on cutaneous hyperchromias: Clinical efficacy of broad-spectrum sunscreens. *Photodermatol. Photoimmunol. Photomed.* **2018**, *34*, 241–248. [[CrossRef](#)] [[PubMed](#)]
76. Bernstein, E.F.; Sarkas, H.W.; Ba, P.B.; Bouche, D. Beyond sun protection factor: An approach to environmental protection with novel mineral coatings in a vehicle containing a blend of skincare ingredients. *J. Cosmet. Dermatol.* **2020**, *19*, 407–415. [[CrossRef](#)]
77. Castanedo-Cazares, J.P.; Hernandez-Blanco, D.; Carlos-Ortega, B.; Fuentes-Ahumada, C.; Torres-Álvarez, B. Near-visible light and UV photoprotection in the treatment of melasma: A double-blind randomized trial. *Photodermatol. Photoimmunol. Photomed.* **2014**, *30*, 35–42. [[CrossRef](#)]
78. Dumbuya, H.; Grimes, P.; Lynch, S.; Ji, K.; Brahmachary, M.; Zheng, Q.; Bouez, C.; Wangari-Talbot, J. Impact of Iron-Oxide Containing Formulations Against Visible Light-Induced Skin Pigmentation in Skin of Color Individuals. *J. Drugs Dermatol.* **2020**, *19*, 712–717. [[CrossRef](#)]
79. Moyal, D.; Seite, S. Prevention of melasma intensification with sunscreen. *J. Am. Acad. Dermatol.* **2020**, *6*, AB20. [[CrossRef](#)]
80. Lim, H.W.; Kohli, I.; Ruvolo, E.; Kolbe, L.; Hamzavi, I.H. Impact of visible light on skin health: The role of antioxidants and free radical quenchers in skin protection. *J. Am. Acad. Dermatol.* **2022**, *86*, S27–S37. [[CrossRef](#)]
81. Zastrow, L.; Lademann, J. Light—Instead of UV Protection: New Requirements for Skin Cancer Prevention. *Anticancer Res.* **2016**, *36*, 1389–1393. [[PubMed](#)]
82. Callender, V.D.; St Surin-Lord, S.; Davis, E.C.; Maclin, M. Postinflammatory hyperpigmentation: Etiologic and therapeutic considerations. *Am. J. Clin. Dermatol.* **2011**, *12*, 87–99. [[CrossRef](#)] [[PubMed](#)]
83. Kang, M.H.; Jang, G.Y.; Ji, Y.-J.; Lee, J.H.; Choi, S.J.; Hyun, T.K.; Kim, H.D. Antioxidant and Anti-Melanogenic Activities of Heat-Treated Licorice (*Wongam, Glycyrrhiza glabra* × *G. uralensis*) Extract. *Curr. Issues Mol. Biol.* **2021**, *43*, 1171–1187. [[CrossRef](#)] [[PubMed](#)]
84. Leis Ayres, E.; dos Santos Silva, J.; Eberlin, S.; Facchini, G.; Vasconcellos, C.; Da Costa, A. In vitro effect of pine bark extract on melanin synthesis, tyrosinase activity, production of endothelin-1, and PPAR in cultured melanocytes exposed to Ultraviolet, Infrared, and Visible light radiation. *J. Cosmet. Dermatol.* **2022**, *21*, 1234–1242. [[CrossRef](#)] [[PubMed](#)]
85. Ruvolo, E.; Boothby-Shoemaker, W.; Kumar, N.; Hamzavi, I.H.; Lim, H.W.; Kohli, I. Evaluation of efficacy of antioxidant-enriched sunscreen products against long wavelength ultraviolet A1 and visible light. *Int. J. Cosmet. Sci.* **2022**, *44*, 394–402. [[CrossRef](#)]
86. Lyons, A.B.; Zubair, R.; Kohli, I.; Nahhas, A.F.; Braunberger, T.L.; Mokhtari, M.; Ruvolo, E.; Lim, H.W.; Hamzavi, I.H. Mitigating Visible Light and Long Wavelength UVA1-induced Effects with Topical Antioxidants. *Photochem. Photobiol.* **2022**, *98*, 455–460. [[CrossRef](#)]
87. Kern, J.; Wood, E.; Almukhtar, R.; Angra, K.; Lipp, M.; Goldman, M. Evaluation of an SPF50 Sunscreen Containing Photolyase and Antioxidants for its Anti-Photoaging Properties and Photoprotection. *J. Drugs Dermatol.* **2022**, *21*, 517–520. [[CrossRef](#)]
88. Mohammad, T.F.; Kohli, I.; Nicholson, C.L.; Treyger, G.; Chaowattanapanit, S.; Nahhas, A.F.; Braunberger, T.L.; Lim, H.W.; Hamzavi, I.H. Oral Polypodium Leucotomos Extract and Its Impact on Visible Light-Induced Pigmentation in Human Subjects. *J. Drugs Dermatol.* **2019**, *18*, 1198–1203.
89. Truchuelo, M.T.; Jiménez, N.; Días, I.J.; Gallego-Rentero, M.; Alonso-Juarranz, M.; Gonzalez, S. A Pilot Study to Assess the Effects of an Oral Photo Protector of Botanical Origin against Visible and Infrared Radiations in Human Volunteers. *Dermatol. Dermatol. Dis.* **2019**, *6*, 2.
90. Zamarrón, A.; Llorio, S.; González, S.; Juarranz, Á. Fernblock Prevents Dermal Cell Damage Induced by Visible and Infrared A Radiation. *Int. J. Mol. Sci.* **2018**, *19*, 2250. [[CrossRef](#)]
91. Nahhas, A.F.; Abdel-Malek, Z.A.; Kohli, I.; Braunberger, T.L.; Lim, H.W.; Hamzavi, I.H. The potential role of antioxidants in mitigating skin hyperpigmentation resulting from ultraviolet and visible light-induced oxidative stress. *Photodermatol. Photoimmunol. Photomed.* **2019**, *35*, 420–428. [[CrossRef](#)] [[PubMed](#)]
92. Juturu, V.; Bowman, J.P.; Deshpande, J. Overall skin tone and skin-lightening-improving effects with oral supplementation of lutein and zeaxanthin isomers: A double-blind, placebo-controlled clinical trial. *Clin. Cosmet. Investig. Dermatol.* **2016**, *9*, 325–332. [[CrossRef](#)] [[PubMed](#)]
93. Pires, F.; Geraldo, V.P.; Antunes, A.; Marletta, A.; Oliveira, O.N., Jr.; Raposo, M. Effect of blue light irradiation on the stability of phospholipid molecules in the presence of epigallocatechin-3-gallate. *Colloids Surf. B Biointerfaces* **2019**, *177*, 50–57. [[CrossRef](#)] [[PubMed](#)]
94. Togni, S.; Maramaldi, G.; Cavagnino, A.; Corti, A.; Giacomelli, L. Vitachelox Protection of the Skin Against Blue Light-Induced Protein Carbonylation. *Cosmetics* **2019**, *3*, 49. [[CrossRef](#)]
95. Avci, P.; Gupta, A.; Sadasivam, M.; Vecchio, D.; Pam, Z.; Pam, N.; Hamblin, M.R. Low-level laser (light) therapy (LLLT) in skin: Stimulating, healing, restoring. *Semin. Cutan. Med. Surg.* **2013**, *32*, 41–52. [[PubMed](#)]
96. Ross, E.V. Laser versus intense pulsed light: Competing technologies in dermatology. *Lasers Surg. Med.* **2006**, *38*, 261–272. [[CrossRef](#)] [[PubMed](#)]
97. Anderson, R.R.; Parrish, J.A. Selective photothermolysis: Precise microsurgery by selective absorption of pulsed radiation. *Science* **1983**, *4569*, 524–527. [[CrossRef](#)] [[PubMed](#)]
98. Galeckas, K.J.; Collins, M.; Ross, E.V.; Uebelhoer, N.S. Split-face treatment of facial dyschromia: Pulsed dye laser with a compression handpiece versus intense pulsed light. *Dermatol. Surg.* **2008**, *34*, 672–680. [[CrossRef](#)]

99. Chern, P.L.; Domankevitz, Y.; Ross, E.V. Pulsed dye laser treatment of pigmented lesions: A randomized clinical pilot study comparison of 607- and 595-nm wavelength lasers. *Lasers Surg. Med.* **2010**, *42*, 865–869. [[CrossRef](#)]
100. Fateh, S.; Farnaghi, F.; Ehsani, A.H.; Noormohammadpour, P.; Seirafi, H. Efficacy and safety of long-pulse pulsed dye laser delivered with compression versus cryotherapy for treatment of solar lentigines. *Indian J. Dermatol.* **2011**, *56*, 48–51. [[CrossRef](#)]
101. Ghaninejadi, H.; Ehsani, A.; Edrisi, L.; Gholamali, F.; Akbari, Z.; Noormohammadpour, P. Solar Lentigines: Evaluating Pulsed Dye Laser (PDL) as an Effective Treatment Option. *J. Lasers Med. Sci.* **2013**, *4*, 33–38. [[PubMed](#)]
102. Allemanna, I.B.; Goldberg, D.J. (Eds.) Benign Pigmented Lesions. In *Basics in Dermatological Laser Applications*; Part of: Current Problems in Dermatology; S. Karger Publishing: Basel, Switzerland, 2011; pp. 81–96.
103. Abd El-Naby, N.; Mostafa Ali, M.; Hawwam, S.A.; Sarhan, N. The clinical and electron microscopic evaluation of the impact of pulsed dye laser techniques on solar lentigines (randomized clinical trial). *J. Dermatolog. Treat.* **2022**, *33*, 361–368. [[CrossRef](#)] [[PubMed](#)]
104. Labadie, J.G.; Kronic, A.L. Long pulsed dye laser with a back-to-back double-pulse technique and compression for the treatment of epidermal pigmented lesions. *Lasers Surg. Med.* **2019**, *51*, 136–140. [[CrossRef](#)]
105. Geddes, E.R.; Stout, A.B.; Friedman, P.M. Retrospective analysis of the treatment of melasma lesions exhibiting increased vascularity with the 595-nm pulsed dye laser combined with the 1927-nm fractional low-powered diode laser. *Lasers Surg. Med.* **2017**, *49*, 20–26. [[CrossRef](#)] [[PubMed](#)]
106. Hassan, A.M.; Elfar, N.N.; Rizk, O.M.; Eissa, N.Y. Pulsed dye laser versus intense pulsed light in melasma: A split-face comparative study. *J. Dermatolog. Treat.* **2018**, *29*, 725–732. [[CrossRef](#)] [[PubMed](#)]
107. Kong, S.H.; Suh, H.S.; Choi, Y.S. Treatment of Melasma with Pulsed-Dye Laser and 1,064-nm Q-Switched Nd:YAG Laser: A Split-Face Study. *Ann. Dermatol.* **2018**, *30*, 1. [[CrossRef](#)] [[PubMed](#)]
108. Ghorbel, H.H.; Boukari, F.; Fontas, E.; Montaudié, H.; Bahadoran, P.; Lacour, J.P.; Passeron, T. Copper Bromide Laser vs Triple-Combination Cream for the Treatment of Melasma: A Randomized Clinical Trial. *JAMA Dermatol.* **2015**, *151*, 791–792. [[CrossRef](#)] [[PubMed](#)]
109. Ponomarev, I.V.; Topchiy, S.B.; Pushkareva, A.E.; Klyuchareva, S.V.; Andrusenko, Y.N. Treatment of Congenital Melanocytic Nevi With a Dual-Wavelengths Copper Vapor Laser: A Case Series. *J. Lasers Med. Sci.* **2021**, *12*, e5. [[CrossRef](#)]
110. Eimpunth, S.; Wanitphakdeedecha, R.; Triwongwaranat, D.; Varothai, S.; Manuskiatti, W. Therapeutic outcome of melasma treatment by dual-wavelength (511 and 578 nm) laser in patients with skin phototypes III–V. *Clin. Exp. Dermatol.* **2014**, *39*, 292–297. [[CrossRef](#)]
111. Lee, H.I.; Lim, Y.Y.; Kim, B.J.; Kim, M.N.; Min, H.J.; Hwang, J.H.; Song, K.Y. Clinicopathologic efficacy of copper bromide plus/yellow laser (578 nm with 511 nm) for treatment of melasma in Asian patients. *Dermatol. Surg.* **2010**, *36*, 885–893. [[CrossRef](#)]
112. Guss, L.; Goldman, M.P.; Wu, D.C. Picosecond 532 nm Neodymium-Doped Yttrium Aluminium Garnet Laser for the Treatment of Solar Lentigines in Darker Skin Types: Safety and Efficacy. *Dermatol. Surg.* **2017**, *43*, 456–459. [[CrossRef](#)] [[PubMed](#)]
113. Negishi, K.; Akita, H.; Matsunaga, Y. Prospective study of removing solar lentigines in Asians using a novel dual-wavelength and dual-pulse width picosecond laser. *Lasers Surg. Med.* **2018**, *50*, 851–858. [[CrossRef](#)] [[PubMed](#)]
114. Vachiramon, V.; Iamsurang, W.; Triyankulsri, K. Q-switched double frequency Nd:YAG 532-nm nanosecond laser vs. double frequency Nd:YAG 532-nm picosecond laser for the treatment of solar lentigines in Asians. *Lasers Med. Sci.* **2018**, *33*, 1941–1947. [[CrossRef](#)] [[PubMed](#)]
115. Hom, D.B.; Ingraffea, A. Efficacy of Q-switched Nd:YAG Frequency Doubling 532nm Laser Combined with Photorejuvenation on Freckles. *Med. Aesthet. Beauty* **2020**, *21*, 41–45.
116. Kerkar, S.; Shilpa, K.; Revathi, T.N. Efficacy of 532-nm Q-switched Nd:YAG Laser in the Treatment of Lip Melanosis. *J. Cutan. Aesthet. Surg.* **2021**, *14*, 203–207.
117. Altalhab, S.; Aljamal, M.; Mubki, T.; AlNomair, N.; Algoblan, S.; Alalola, A.; Alissa, I.; Alissa, A. Q-switched 532 nm Nd:YAG laser therapy for physiological lip hyperpigmentation: Novel classification, efficacy, and safety. *J. Dermatol. Treat.* **2022**, *33*, 1324–1328. [[CrossRef](#)]
118. Alabdulrazzaq, H.; Brauer, J.A.; Bae, Y.-S.; Geronemus, R.G. Clearance of yellow tattoo ink with a novel 532-nm picosecond laser. *Lasers Surg. Med.* **2015**, *47*, 285–288. [[CrossRef](#)]
119. Bernstein, E.F.; Schomacker, K.T.; Basilavecchio, L.D.; Plugis, J.M.; Bhawalkar, J.D. A novel dual-wavelength, Nd:YAG, picosecond-domain laser safely and effectively removes multicolor tattoos. *Lasers Surg. Med.* **2015**, *47*, 542–548. [[CrossRef](#)]
120. Yu, H.-S.; Wu, C.-S.; Kao, Y.-H.; Chiou, M.-H.; Yu, C.-L. Helium-neon laser irradiation stimulates migration and proliferation in melanocytes and induces repigmentation in segmental-type vitiligo. *J. Investig. Dermatol.* **2003**, *120*, 56–64. [[CrossRef](#)]
121. Lan, C.-C.E.; Wu, S.-B.; Wu, C.-S.; Shen, Y.-C.; Chiang, T.-Y.; Wei, Y.-H.; Yu, H.-S. Induction of primitive pigment cell differentiation by visible light (helium–neon laser): A photoacceptor-specific response not replicable by UVB irradiation. *J. Mol. Med.* **2012**, *90*, 321–330. [[CrossRef](#)]
122. Lan, C.C.; Wu, C.S.; Chiou, M.H.; Hsieh, P.C.; Yu, H.S. Low-energy helium-neon laser induces locomotion of the immature melanoblasts and promotes melano genesis of the more differentiated melanoblasts: Recapitulation of vitiligo repigmentation in vitro. *J. Investig. Dermatol.* **2006**, *126*, 2119–2126. [[CrossRef](#)] [[PubMed](#)]
123. Lan, C.-C.; Wu, C.-S.; Chiou, M.-H.; Chiang, T.-Y.; Yu, H.-S. Low-energy helium–Neon laser induces melanocyte proliferation via interaction with type IV collagen: Visible light as a therapeutic option for vitiligo. *Br. J. Dermatol.* **2009**, *2*, 273–280. [[CrossRef](#)] [[PubMed](#)]

124. Boen, M.; Wilson, M.J.V.; Wu, D.C.; Goldman, M.P. Laser_Depigmentation_in_Extensive_Vitiligo.26. *Dermatol. Surg.* **2019**, *4*, 621–623. [[CrossRef](#)] [[PubMed](#)]
125. Taylor, C.R.; Anderson, R.R. Treatment of Benign Pigmented Epidermal Lesions by Q-Switched Ruby Laser. *Int. J. Dermatol.* **1993**, *32*, 908–912. [[CrossRef](#)] [[PubMed](#)]
126. Taylor, C.R.; Flotte, T.J.; Gange, R.W.; Anderson, R.R. Treatment of nevus of Ota by Q-switched ruby laser. *J. Am. Acad. Dermatol.* **1994**, *30 Pt 1*, 743–751. [[CrossRef](#)]
127. Watanabe, S.; Takahashi, H. Treatment of nevus of ota with the q-switched rubylaser. *N. Engl. J. Med.* **1994**, *26*, 1745–1750. [[CrossRef](#)]
128. Rathod, S.; Munshi, A.; Agarwal, J. Skin markings methods and guidelines: A reality in image guidance radiotherapy era. *South Asian J. Cancer* **2020**, *1*, 27–29. [[CrossRef](#)]
129. Ma, S.-Y.; Gong, Y.-Q.; Zhang, W.J.; Liang, B.-H.; Li, Y.-M.; Xie, Z.-M.; Zhu, H.-L. Split-face comparison of the efficacy of picosecond 532 nm Nd:YAG laser and Q-switched 755 nm Alexandrite laser for treatment of freckles. *J. Cosmet. Laser Ther.* **2022**, *24*, 22–27. [[CrossRef](#)]
130. Yamada-Kanazawa, S.; Jinnin, M.; Fukushima, S. Nevus of Ota on the auricle successfully treated with Q-switched ruby laser. *Drug Discov. Ther.* **2022**, *16*, 254–255. [[CrossRef](#)]
131. Taylor, C.R.; Anderson, R.R. Ineffective treatment of refractory melasma and postinflammatory hyperpigmentation by Q-switched ruby laser. *J. Dermatol. Surg. Oncol.* **1994**, *20*, 592–597. [[CrossRef](#)]
132. Jang, W.S.; Lee, C.K.; Kim, B.J.; Kim, M.N. Efficacy of 694-nm Q-Switched Ruby Fractional Laser Treatment of Melasma in Female Korean Patients. *Dermatol. Surg.* **2011**, *37*, 1133–1140. [[CrossRef](#)] [[PubMed](#)]
133. Hilton, S.; Heise, H.; Bühren, B.A.; Schrumpf, H.; Bölke, E.; Gerber, P.A. Treatment of melasma in Caucasian patients using a novel 694-nm Q-switched ruby fractional laser. *Eur. J. Med. Res.* **2013**, *18*, 43. [[CrossRef](#)] [[PubMed](#)]
134. Zhou, H.L.; Hu, B.; Zhang, C. Efficacy of 694-nm fractional Q-switched ruby laser (QSRL) combined with sonophoresis on levorotatory vitamin C for treatment of melasma in Chinese patients. *Lasers Med. Sci.* **2016**, *31*, 991–995. [[CrossRef](#)] [[PubMed](#)]
135. Li, Y.; Tong, X.; Yang, J.; Yang, L.; Tao, J.; Tu, Y. Q-switched alexandrite laser treatment of facial and labial lentigines associated with Peutz-Jeghers syndrome. *Photodermatol. Photoimmunol. Photomed.* **2012**, *28*, 196–199. [[CrossRef](#)] [[PubMed](#)]
136. Wattanakrai, P.; Mornchan, R.; Eimpunth, S. A randomized, split-face clinical trial of low-fluence Q-switched neodymium-doped yttrium aluminum garnet (1064 nm) laser versus low-fluence Q-switched alexandrite laser (755 nm) for the treatment of facial melasma. *Lasers Surg. Med.* **2014**, *46*, 531–537.
137. Zhang, B.; Chu, Y.; Xu, Z.; Sun, Y.; Li, L.; Han, X.; Wang, C.; Wei, L.; Liu, Y.; Ma, L. Treatment of Café-Au-Lait Spots Using Q-Switched Alexandrite Laser: Analysis of Clinical Characteristics of 471 Children in Mainland China. *Lasers Surg. Med.* **2019**, *51*, 694–700. [[CrossRef](#)] [[PubMed](#)]
138. Manuskiatti, W.; Yan, C.; Tantrapornpong, P.; Cembrano, K.A.; Techapichetvanich, T.; Wanitphakdeedecha, R. A Prospective, Split-Face, Randomized Study Comparing a 755-nm Picosecond Laser With and Without Diffractive Lens Array in the Treatment of Melasma in Asians. *Lasers Surg. Med.* **2021**, *53*, 95–103. [[CrossRef](#)]
139. Vachiramon, V.; Namasondhi, A.; Anuntrangsee, T.; Jurairattanaporn, N. Randomized, evaluator-blinded comparative study of a potassium titanyl phosphate (KTP) 532-nm picosecond laser and an alexandrite 755-nm picosecond laser for the treatment of solar lentigines in Asians. *J. Cosmet. Dermatol.* **2022**, *21*, 4370–4377. [[CrossRef](#)]
140. Raulin, C.; Greve, B.; Grema, H. IPL technology: A review. *Lasers Surg. Med.* **2003**, *32*, 78–87. [[CrossRef](#)]
141. Kim, J.; Lee, J.; Choi, H. Intense Pulsed Light Attenuates UV-Induced Hyperimmune Response and Pigmentation in Human Skin Cells. *Int. J. Mol. Sci.* **2021**, *22*, 3173. [[CrossRef](#)]
142. Bjerring, P.; Christiansen, K. Intense pulsed light source for treatment of small melanocytic nevi and solar lentigines. *J. Cutan. Laser Ther.* **2000**, *2*, 177–181. [[CrossRef](#)]
143. Kawada, A.; Shiraiishi, H.; Asai, M.; Kameyama, H.; Sangen, Y.; Aragane, Y.; Tezuka, T. Clinical improvement of solar lentigines and ephelides with an intense pulsed light source. *Dermatol. Surg.* **2002**, *28*, 504–508. [[PubMed](#)]
144. Sasaya, H.; Kawada, A.; Wada, T.; Hirao, A.; Oiso, N. Clinical effectiveness of intense pulsed light therapy for solar lentigines of the hands. *Dermatol. Ther.* **2011**, *24*, 584–586. [[CrossRef](#)] [[PubMed](#)]
145. Friedmann, D.P.; Peterson, J.D. Efficacy and safety of intense pulsed light with a KTP filter for the treatment of solar lentigines. *Lasers Surg. Med.* **2019**, *51*, 500–508. [[CrossRef](#)] [[PubMed](#)]
146. Amornpetkul, W.; Kanokrungrsee, S.; Kamanamool, N.; Udompataikul, M.; Rojhirunsakool, S. Comparison between the use of intense pulsed light and Q-switched neodymium-doped yttrium aluminum garnet laser for the treatment of axillary hyperpigmentation. *J. Cosmet. Dermatol.* **2021**, *20*, 2785–2793. [[CrossRef](#)] [[PubMed](#)]
147. Zhong, Y.; Huang, L.; Chen, Y.; Yan, T.; Yang, B.; Man, M.Q. The efficacy of intense pulsed light for Becker’s nevus: A retrospective analysis of 45 cases. *J. Cosmet. Dermatol.* **2021**, *20*, 466–471. [[CrossRef](#)] [[PubMed](#)]
148. Babilas, P.; Schreml, S.; Szeimies, R.-M.; Landthaler, M. Intense pulsed light (IPL): A review. *Lasers Surg. Med.* **2010**, *42*, 93–104. [[CrossRef](#)] [[PubMed](#)]
149. Li, Y.H.; Chen, J.Z.; Wei, H.C.; Wu, Y.; Liu, M.; Xu, Y.Y.; Dong, G.H.; Chen, H.D. Efficacy and Safety of Intense Pulsed Light in Treatment of Melasma in Chinese Patients. *Dermatol. Surg.* **2008**, *34*, 693–701.
150. Yi, J.; Hong, T.; Zeng, H.; Li, P.; Li, P.; Wang, S.; Chen, J.; Li, P.; Zhou, J. A Meta-analysis-Based Assessment of Intense Pulsed Light for Treatment of Melasma. *Aesthetic Plast. Surg.* **2020**, *44*, 947–952. [[CrossRef](#)]

151. Dierickx, C.C.; Anderson, R.R. Visible light treatment of photoaging. *Dermatol. Ther.* **2005**, *18*, 191–208. [[CrossRef](#)]
152. Weiss, R.A.; McDaniel, D.H.; Geronemus, R.G.; Margaret, A.W.; Karen, L.B.; Munavalli, G.M.; Bellew, S.G. Clinical experience with light-emitting diode (LED) photomodulation. *Dermatol. Surg.* **2005**, *31 Pt 2*, 1199–1205. [[CrossRef](#)]
153. Heiskanen, V.; Hamblin, M.R. Photobiomodulation: Lasers vs. light emitting diodes? *Photochem. Photobiol. Sci.* **2018**, *17*, 1003–1017. [[CrossRef](#)]
154. Jin, S.; Chen, L.; Xu, Z.; Xing, X.; Zhang, C.; Xiang, L. 585 nm light-emitting diodes inhibit melanogenesis through upregulating H19/miR-675 axis in LEDs-irradiated keratinocytes by paracrine effect. *J. Dermatol. Sci.* **2020**, *98*, 102–108. [[CrossRef](#)]
155. Dai, X.; Jin, S.; Xuan, Y.; Yang, Y.; Lu, X.; Wang, C.; Chen, L.; Xiang, L.; Zhang, C. 590 nm LED Irradiation Improved Erythema through Inhibiting Angiogenesis of Human Microvascular Endothelial Cells and Ameliorated Pigmentation in Melasma. *Cells* **2022**, *11*, 3949. [[CrossRef](#)]
156. Mpfana, N.; Abrahamse, H. The Management of Melasma on Skin Types V and VI Using Light Emitting Diode Treatment. *Photomed. Laser Surg.* **2018**, *36*, 522–529. [[CrossRef](#)]
157. Xuan, Y.J.; Dai, X.X.; Chen, L.; Xiang, L.H.; Jin, S.L.; Zhang, C.F. Efficacy and safety of home-based 590 nm light-emitting diodes and in-hospital 1064 nm Q-switched Nd:YAG laser in the treatment of facial melasma: A single-centre, prospective, randomized clinical trial. *J. Eur. Acad. Dermatol. Venereol.* **2023**, *ahead of print*. [[CrossRef](#)]
158. Lee, S.Y.; Park, K.-H.; Choi, J.-W.; Kwon, J.-K.; Lee, D.R.; Shin, M.S.; Lee, J.S.; You, C.E.; Park, M.Y. A prospective, randomized, placebo-controlled, double-blinded, and split-face clinical study on LED phototherapy for skin rejuvenation: Clinical, profilometric, histologic, ultrastructural, and biochemical evaluations and comparison of three different treatment settings. *J. Photochem. Photobiol. B* **2007**, *88*, 51–67.
159. A Weiss, R.; A Weiss, M.; Geronemus, R.G.; McDaniel, D.H. A novel non-thermal non-ablative full panel LED photomodulation device for reversal of photoaging: Digital microscopic and clinical results in various skin types. *J. Drugs Dermatol.* **2004**, *3*, 605–610.
160. Cho, H.; Kim, B.; Kim, O.S.; Kim, Y.; Yang, Y.; Song, J.; Liu, D.; Jeon, S.; Kim, O. Photochemical reaction to increase melanogenesis using *Buddleja officinalis* and blue light-emitting diode irradiation in B16F10. *Photodiagn. Photodyn. Ther.* **2021**, *35*, 102456. [[CrossRef](#)]
161. Lodi, G.; Del Re, C.; Nisticò, S.P.; Bennardo, L.; Cannarozzo, G.; Sannino, M. Blue light-emitting diodes for the treatment of localized vitiligo: A retrospective study. *J. Cosmet. Dermatol.* **2023**, *22*, 1273–1278. [[CrossRef](#)]

Disclaimer/Publisher's Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.