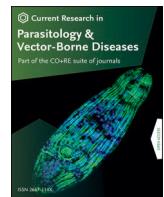




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Cryptosporidium and Giardia in cats and dogs: What is the real zoonotic risk?

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ABSTRACT

Due to the close bond between humans and companion animals, a thorough understanding of the diversity of *Cryptosporidium* species and *Giardia* assemblages in cats and dogs is essential to determine the potential zoonotic risks. Analysis of molecular studies shows that *C. felis* and *C. canis* are the main species infecting cats and dogs, respectively. These species are largely host-specific, as despite intense association with humans, prevalence of *C. felis* and *C. canis* in humans is low and predominantly in immunocompromised individuals and low-income countries. There have been reports of *C. parvum* in cats and dogs and two reports of *C. hominis* in dogs. In most studies conducted to date, however, the prevalence of zoonotic species was low and may be associated with coprophagy and/or spillback, but this remains to be determined. Results of subtyping studies suggest that for *C. felis* and *C. canis*, some zoonotic transmission may occur but host-adapted subtypes also exist. *Giardia duodenalis* assemblages C and D are commonly reported in dogs, with assemblages F and A most common in cats. Assemblages C, D and F are largely host-specific as there are only a handful of reports of them in humans. Reports of assemblage A and B in cats and dogs may be due to coprophagy or spillback from owners. Despite the extent of pet ownership and the close contact between humans and companion animals worldwide, the overall risk of zoonotic transmission from cats and dogs to humans is uncertain but thought to be low due to *C. canis*, *C. felis* and *G. duodenalis* assemblages C, D and F being predominantly host-specific, the relatively low prevalence of *C. parvum* (and *C. hominis*) in cats and dogs (which may be due to mechanical carriage), and low oo/cyst shedding. Carefully designed epidemiological studies of cats and dogs and their owners using subtyping tools are essential to better quantify the extent of spillover and spillback of *Cryptosporidium* and *Giardia* between pets and their owners.

1. Introduction

Humans and their companion animals, particularly cats and dogs, have had a long and close association over many centuries and provide a wide variety of benefits to their owners and foster carers including increased physical activity, companionship and improvements in mental health (Martins et al., 2023; Zablan et al., 2023). Cat and dog ownership also increases connections with family members and the wider community (Wood et al., 2015), with higher wellbeing and trust, reduced anxiety and reduced screen time particularly in adolescents (Char-maraman et al., 2022; Koyasu et al., 2023).

Pet owner behaviours such as sleeping with pets, allowing pets to lick faces and hands, insufficient/inrequent hand-washing, and feeding raw

meat to cats and dogs, can promote the transmission of a variety of zoonotic pathogens from pets to humans. These include *Salmonella* (Davies et al., 2019), antimicrobial resistant bacteria (Jin et al., 2023), ectoparasites and associated zoonotic pathogens (Tahir et al., 2019; Saleh et al., 2021; Mateo et al., 2023), *Toxocara* (Patterson, 2023) and other helminths (Overgaauw et al., 2020; Mateo et al., 2023), and enteric protozoa such as *Cryptosporidium* and *Giardia* (Cai et al., 2021; Meng et al., 2021).

Cryptosporidium oocysts and *Giardia* cysts are immediately infectious when shed in faeces and are environmentally robust (Cai et al., 2021; Ryan et al., 2021a). Transmission of *Cryptosporidium* and *Giardia* in cats and dogs is via faecal-oral contamination through coprophagy or ingestion of contaminated food or water. Although frequently

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asymptomatic in adult cats and dogs, *Cryptosporidium* and *Giardia* can both cause watery diarrhoea, abdominal pain and vomiting in both humans and young pets (Cai et al., 2021; Li et al., 2021a; Ryan et al., 2021a). Fenbendazole and metronidazole are commonly used to treat clinical giardiasis in cats and dogs, but have poor efficacy (Kaufmann et al., 2022). There is no registered treatment available for cryptosporidiosis in cats and dogs (ESCCAP, 2018). Mixed infections with *Giardia* and *Cryptosporidium* and other enteropathogens are common in both dogs and cats, particularly in dogs (Hamnes et al., 2007; Pallant et al., 2015; Xu et al., 2016; Gil et al., 2017; Sommer et al., 2018; López-Arias et al., 2019; Nagamori et al., 2020; Mateo et al., 2023; Murnik et al., 2023).

The 18S ribosomal RNA (rRNA) is the most commonly used locus for identifying *Cryptosporidium* to species level, while the glycoprotein 60 gene (*gp60*) is most widely used for subtyping (Ryan et al., 2021a; Yang et al., 2021). Over 49 *Cryptosporidium* species have been identified to date, with *C. hominis* and *C. parvum* responsible for ~95% of human infections (Ryan et al., 2021a).

Human infections with *Giardia* are caused by *Giardia duodenalis*, a multi-species complex, consisting of eight assemblages. Assemblages A and B predominantly infect humans, assemblages C and D infect mostly dogs, assemblage F infects mainly cats, assemblage G infects rodents and assemblage H infects pinnipeds (Cai et al., 2021; Ryan et al., 2021a). The most commonly used typing loci for *Giardia* are the betagiardin (*bg*), glutamate dehydrogenase (*gdh*), triosephosphate isomerase (*tpi*) genes, and the 18S rRNA locus (Cai et al., 2021; Ryan et al., 2021a). Multilocus sequence typing (MLST) of the *bg*, *gdh* and *tpi* genes has identified three sub-assemblages within assemblage A: sub-assemblage AI (predominantly in livestock, companion animals and some humans); AII (predominantly in humans); and AIII (in wild ruminants) (Cai et al., 2021). In contrast, sub-assemblages within assemblage B, cannot be reliably determined with the currently used loci (Cai et al., 2021).

Due to the close relationship between humans and companion animals, a thorough understanding of the diversity of *Cryptosporidium* species and *Giardia* assemblages in cats and dogs is essential to determine their zoonotic risk and public health implications. This review analyses *Cryptosporidium* species and *Giardia* assemblages reported in cats and dogs globally, and discusses the zoonotic risk posed by the *Cryptosporidium* species and *Giardia* assemblages identified. The majority of the > 350 publications on *Cryptosporidium* and *Giardia* in cats and dogs have been epidemiological surveys, with molecular tools used in more recent years, enabling researchers to type the pathogens to species and subtype level. This review centers on the findings and discussions emerging from these molecular studies.

2. *Cryptosporidium* in cats and dogs

Generally, the overall prevalence of *Cryptosporidium* in cats and dogs globally is < 10%. However, some studies using microscopy and immunological-based detection methods have reported higher prevalences (> 20%) (e.g. Cirak and Bauer, 2004; Hamnes et al., 2007; Ferreira et al., 2017). A recent systematic review of the prevalence of *Cryptosporidium* in cats reported the mean prevalence by microscopy, coproantigens and molecular tools to be 4.2%, 8.2%, and 5.0%, respectively, with an overall mean prevalence of 6% (Meng et al., 2021). In dogs, the overall mean prevalence has been reported at 8% using microscopic detection, 7% using coproantigen detection and 6% using molecular diagnostic methods (Taghipour et al., 2020). For a more complete overview of all studies including microscopy-based analyses, refer to Li et al. (2021a).

Cryptosporidium felis and *C. canis* are the main species infecting cats and dogs globally, respectively (Lucio-Forster et al., 2010; Li et al., 2021a; Meng et al., 2021; Ryan et al., 2021b) (Tables 1 and 2). *Cryptosporidium felis* and *C. canis* are also among the top five *Cryptosporidium* species infecting humans, predominantly in low-income countries (Yang et al., 2021). In cats, in addition to *C. felis*, there have also been some

reports of *C. parvum*, *C. canis*, *C. muris*, *C. ryanae*, and rat genotypes III and IV. In dogs, *C. hominis*, *C. parvum*, *C. andersoni*, *C. muris*, *C. scrofarum*, *C. meleagridis*, *C. ubiquitum*, and rat genotypes III and IV have been reported occasionally, in addition to *C. canis*. Many of these species are zoonotic and are listed in Tables 1 and 2. *Cryptosporidium ryanae* and rat genotype III were reported in one and six cats, respectively, in Australia (Yang et al., 2015), most likely due to coprophagy, and rat genotype IV was reported in a single cat in China (Li et al., 2019a). The zoonotic status of *C. andersoni* is uncertain (Yang et al., 2021) and it has not been included in Table 2, but there has been a single report of *C. andersoni* (0.1%; 1/676) in a shelter dog in the UK (Rosanowski et al., 2018). Young age, presence of diarrhoea, and living in pet shops and shelters are risk factors associated with *Cryptosporidium* infection in cats and dogs (Li et al., 2021a; Mateo et al., 2023).

There have been several reports of *C. parvum* in cats involving mostly single reports in individual cats from various countries (Table 1). There have also been a few reports of *C. muris* and rat genotypes III and IV in cats, which may be due to mechanical carriage from ingesting rodents or an accidental infection (Table 1). *Cryptosporidium muris* oocysts from a cat have been shown to be infective to laboratory mice (Pavlásek and Ryan, 2007), indicating that they may retain viability following passage through a cat. Another study indicated that *C. muris* was infective to cats, as DNA sequencing of paraffin-embedded sections of intestinal and stomach biopsies from a cat with persistent diarrhoea detected *C. muris* in the stomach and *C. felis* in the intestine (FitzGerald et al., 2011). A follow-up study 12 months later on the same cat, detected *C. muris*-like oocysts from endoscopic biopsies but *C. felis* oocysts were not identified (FitzGerald et al., 2011). The identification of *C. muris* and other rodent genotypes in cats is likely due to frequent contact between cats and rodents, with the study by FitzGerald et al. (2011) showing *C. muris* can infect the gastric mucosa of cats. A previous cross-transmission study also reported that cats were “highly susceptible” to *C. muris*, whereas dogs had low susceptibility (Iseki et al., 1989). There has been one report of *C. canis* in one cat, which is likely due to mechanical carriage (Table 1).

There have only been two reports of *C. hominis* in dogs; one in a wild dog in Sydney catchments in Australia (Ng et al., 2011) and the other in a shelter dog faecal sample in Spain (Gil et al., 2017). The parasite was identified by PCR and sequencing at the 18S locus, but in both cases, amplification at the *gp60* locus was not successful (Ng et al., 2011; Gil et al., 2017), suggesting that oocyst numbers were low and may have been present due to coprophagy of human faeces. There have been several reports of *C. parvum* in dogs, with the highest prevalence (10%; 5/50) reported in household dogs in Egypt (Gharieb et al., 2018), and in two puppies with diarrhoea in Egypt (10%; 2/20) (El-Madawy et al., 2010). Another study in Thailand reported *C. parvum* in 6.4% (7/109) of dogs (31.8% of the 22 typed samples) (Tangtrongsup et al., 2017) (Table 2). In the majority of other studies, the prevalence of *C. parvum* has been low (0.3–2.9%) (Table 2).

Only a few studies have subtyped the *C. parvum* detected in dogs. Among them, subtypes IIaA15G2R1 and IIaA17G2R1 have been detected in dogs in the UK (Smith et al., 2010; Rosanowski et al., 2018), and subtype IIaA15G2R1 in dogs in Germany (Murnik et al., 2022). These are common subtypes in cattle and humans (Feng and Xiao, 2017; Holzhausen et al., 2019; Santín, 2020; Guo et al., 2021, 2022; Yang et al., 2021). In the study by Smith et al. (2010), IIaA17G2R1 was detected in the single farm dog sampled, and was the dominant subtype detected in cattle on the farm. Epidemiological analysis indicates that dogs are at a higher risk of being infected with *C. parvum* from cattle when living in areas on or near cattle farms (Moreira et al., 2018). In the study of shelter dogs in the UK (Rosanowski et al., 2018), the authors reported that “some of the dogs arriving at the shelter, may have been roaming as strays prior to submission, with the potential that the environment was more contaminated than for owned dogs”. The study in Germany, which identified IIaA15G2R1 in two dogs (Murnik et al., 2022), was conducted in Saxony, and IIaA15G2R1 was the most

Table 1Zoonotic *Cryptosporidium* and *Giardia* species and assemblages identified in cats globally.

Species/Assemblage	Hosts	Prevalence	Country	Reference
<i>Cryptosporidium</i>				
<i>C. felis</i>	Domestic cats	1.3% (2/162)	Australia	Sargent et al. (1998)
	Domestic cat	—	Australia	Morgan et al. (1998)
	Shelter & domestic cats	1.7% (18/1063)	Australia	Palmer et al. (2008a, b)
	Various	2.3% (8/345)	Australia	Yang et al. (2015)
	Domestic cats	1.7% (5/298)	Austria	Hinney et al. (2015)
	Shelter cats	1.5% (2/130)	Austria	Joachim et al. (2023)
	<i>Cryptosporidium</i> -positive domestic cats	100% (7/7)	Brazil	Thomaz et al. (2007)
	Domestic cats	6.1% (3/49)	Brazil	Alves et al. (2018)
	Domestic cats	5.4% (3/55)	Brazil	de Oliveira et al. (2021)
	Various	1.9% (1/52)	China	Li et al. (2015)
	Domestic cats	3.8% (6/160)	China	Xu et al. (2016)
	Domestic cats	2.3% (8/346)	China	Li et al. (2019b)
	Wide range	5.0% (21/418)	China	Li et al. (2019a)
	Domestic cats	0.6% (1/171)	China	Wang et al. (2021)
	Domestic cat	10.9% (5/46)	Colombia	Santín et al. (2006)
	Domestic cats	100% (1/1)	Czech Republic	Hajdušek et al. (2004)
	Domestic & stray cats	4.3% (11/255)	Czech Republic, Poland, Slovakia	Kváč et al. (2017)
	Domestic cats	2.8% (9/317)	Denmark	Enemark et al. (2020)
	Domestic cats	0.7% (1/134)	Egypt	Elmahallawy et al. (2023)
	Domestic & shelter cats	1.5% (4/264)	Greece	Kostopoulou et al. (2017)
	Domestic dogs	1.4% (2/140)	Iran	Ranjbar et al. (2018)
	Domestic dogs	0.6% (2/300)	Iran	Homayouni et al. (2019)
	Stray cats	5.3% (7/132)	Iran	Karimi et al. (2023)
	Shelter cats	1.5% (1/65)	Italy	Gil et al. (2017)
	Domestic dogs	0.1% (1/705)	Italy	Simonato et al. (2017)
	Domestic cats	1.8% (19/1079)	Japan	Yamamoto et al. (2009)
	Domestic cats	12.7% (7/55)	Japan	Yoshiuchi et al. (2010)
	Domestic & pet shop cats	1.4% (4/286)	Japan	Ito et al. (2017)
	Domestic cats	1.0% (1/101)	Poland	Piekara-Stepińska et al. (2021)
	Shelter cats	0.6% (1/158)	South Korea	Kwak and Seo (2020)
	Domestic cats	2.9% (1/34)	Spain	de Lucio et al. (2017)
	Temple cats	2.5% (2/80)	Thailand	Koompapong et al. (2014)
	Domestic & shelter cats	1.5% (1/66)	Thailand	Tangtrongsup et al. (2020)
	Stray cats	3.0% (12/399)	Turkey	Köseoglu et al. (2022)
	Cat colony cats	100% (18/18)	USA	Fayer et al. (2006)
	Domestic cats	4.8% (12/250)	USA	Ballweber et al. (2009)
<i>C. parvum</i>	Domestic cats	2.0% (1/49)	Brazil	Alves et al. (2018)
	Domestic cat	100% (1/1)	Chile	Neira et al. (2010)
	Various	1.9% (1/52)	China	Li et al. (2015)
	Wide range	0.7% (3/418)	China	Li et al. (2019a)
	Domestic cats	14.3% (1/7)	Costa Rica	Scorza et al. (2011)
	Domestic cats	3.7% (5/134)	Egypt	Elmahallawy et al. (2023)
	Domestic cats	5.3% (1/19)	Germany	Sotiriadou et al. (2013)
	Domestic & shelter cats	1.5% (1/66)	Thailand	Tangtrongsup et al. (2020)
<i>C. canis</i>	Domestic cats	3.0% (1/133)	Iran	Karimi et al. (2023)
<i>C. muris</i>	Domestic cat	100% (1/1)	Australia	FitzGerald et al. (2011)
	Various	0.3% (1/345)	Australia	Yang et al. (2015)
	Wide range	0.3% (1/418)	China	Li et al. (2019a)
	Domestic cat	2.2% (1/46)	Colombia	Santín et al. (2006)
	Domestic cat	100% (1/1)	Czech Republic	Pavlásek and Ryan (2007)
<i>Giardia</i>				
Assemblage A	<i>Giardia</i> -positive cats	33.3% (6/18)	Australia	Read et al. (2004)
	Various	0.3% (1/345)	Australia	Yang et al. (2015)
	Domestic cats	1.3% (4/298)	Austria	Hinney et al. (2015)
	Shelter cats	1.5% (2/130)	Austria	Joachim et al. (2023)
	<i>Giardia</i> -positive domestic cats	42.1% (8/19)	Brazil	Souza et al. (2007)
	Domestic cats	100% (1/1)	Brazil	Volotão et al. (2007)
	<i>Giardia</i> -positive domestic cats	61.5% (8/13)	Canada	McDowall et al. (2011)
	Various	3.6% (8/219)	Canada	Hoopes et al. (2015)
	Stray cats	7.8% (8/102)	China	Zheng et al. (2015)
	Domestic cats	1.2% (2/160)	China	Xu et al. (2016)
	Stray cats	2.8% (3/104)	China	Pan et al. (2018)
	Wide range	0.3% (1/418)	China	Li et al. (2019a)
	Domestic cats	0.3% (1/346)	China	Li et al. (2019b)
	Domestic cats	28.6% (2/7)	Costa Rica	Scorza et al. (2011)
	<i>Giardia</i> -positive domestic cats	5.5% (1/18)	Czech Republic	Lecová et al. (2020)
	Domestic cats	2.5% (8/317)	Denmark	Enemark et al. (2020)
	<i>Giardia</i> -positive cats	43.0% (68/158)	Europe	Sprong et al. (2009)
	Domestic cats	10.5% (2/19)	Germany	Sotiriadou et al. (2013)
	Domestic cats	1.4% (2/145)	Germany	Sommer et al. (2018)
	Domestic & shelter cats	2.3% (6/264)	Greece	Kostopoulou et al. (2017)
	Kennel cat	100% (1/1)	Italy	Berrilli et al. (2004)
	Stray & domestic cats	37.0% (10/27)	Italy	Papini et al. (2007)

(continued on next page)

Table 1 (continued)

Species/Assemblage	Hosts	Prevalence	Country	Reference
Assemblage B	Domestic cats	1.7% (3/181)	Italy	Paoletti et al. (2011)
	Domestic cats	9.0% (14/156)	Italy	Zanzani et al. (2014)
	Sheltered cats	1.5% (1/65)	Italy	Gil et al. (2017)
	Stray cats	27.8% (37/133)	Italy	Guadano Procesi et al. (2022)
	<i>Giardia</i> -positive cats	100% (3/3)	Italy, Croatia	Cacciò et al. (2008)
	Cats from cat café's and petshops	0.6% (2/321)	Japan	Suzuki et al. (2011)
	<i>Giardia</i> -positive domestic cat	100% (1/1)	Mexico	Ponce-Macotela et al. (2002)
	Various	1.0% (2/200)	Mexico	Veyna-Salazar et al. (2023)
	Domestic cats	1.6% (1/60)	Netherlands	Overgaauw et al. (2009)
	Domestic cats	3.0% (1/33)	Poland	Piekarska et al. (2016)
	Household & kennel cats	9.0% (2/22)	Portugal	Ferreira et al. (2011)
	<i>Giardia</i> -positive cats	27.7% (5/18)	Sweden	Lebbad et al. (2010)
	Domestic & shelter cats	1.5% (1/66)	Thailand	Tangtrongsup et al. (2020)
	Domestic cats	3.3% (4/122)	UK	Krumrie et al. (2022)
	Domestic cats	2.4% (6/250)	USA	Vasilopoulos et al. (2007)
	<i>Giardia</i> -positive domestic cats	32.4% (12/37)	USA	Saleh et al. (2019)
	<i>Giardia</i> -positive cats	11.1% (2/18)	Australia	Read et al. (2004)
	<i>Giardia</i> -positive domestic cats	7.7% (1/13)	Canada	McDowall et al. (2011)
Assemblage C	Domestic cats	3.7% (6/160)	China	Xu et al. (2016)
	<i>Giardia</i> -positive cats	1.9% (3/158)	Europe	Sprong et al. (2009)
	Domestic & shelter cats	0.4% (1/264)	Greece	Kostopoulou et al. (2017)
	Stray cats	0.7% (1/132)	Iran	Karimi et al. (2023)
	Symptomatic cats	29.4% (30/102)	Turkey	Sursal et al. (2020)
	Domestic cats	8.0% (8/100)	Turkey	Önder et al. (2021)
	Domestic cats	3.3% (4/122)	UK	Krumrie et al. (2022)
	<i>Giardia</i> -positive cats	11.1% (2/18)	Australia	Read et al. (2004)
	Stray cats	1.0% (1/102) ^b	China	Zheng et al. (2015)
	Domestic cats	1.3% (2/160)	China	Xu et al. (2016)
Assemblage D	Domestic cats	9.0% (1/11)	China	Liu et al. (2017)
	<i>Giardia</i> -positive cats	3.2% (5/158)	Europe	Sprong et al. (2009)
	Domestic & shelter cats	0.4% (1/264)	Greece	Kostopoulou et al. (2017)
	Domestic cats	1.4% (2/146)	Italy	Mancianti et al. (2015)
	Domestic & shelter cats	6.1% (4/66)	Thailand	Tangtrongsup et al. (2020)
	<i>Giardia</i> -positive cats	38.8% (7/18)	Australia	Read et al. (2004)
	Shelter & domestic cats	0.1% (1/1063)	Australia	Palmer et al. (2008a, b)
	Domestic cats	0.6% (1/160)	China	Xu et al. (2016)
	<i>Giardia</i> -positive cats	1.9% (3/158)	Europe	Sprong et al. (2009)
	Domestic cats	1.9% (3/156)	Italy	Zanzani et al. (2014)
Assemblage E	Domestic cats	80.0% (8/10)	Romania	Adriana et al. (2016)
	Domestic & shelter cats	1.5% (1/66)	Thailand	Tangtrongsup et al. (2020)
	<i>Giardia</i> -positive cats	5.5% (1/18)	Australia	Read et al. (2004)
	<i>Giardia</i> -positive cats	1.3% (2/158)	Europe	Sprong et al. (2009)
	<i>Giardia</i> -positive cats	5.5% (1/18)	Sweden	Lebbad et al. (2010)
	Shelter & domestic cats	0.7% (7/1063)	Australia	Palmer et al. (2008a, b)
	Various	2.6% (9/345)	Australia	Yang et al. (2015)
	Domestic cats	3.3% (10/298)	Austria	Hinney et al. (2015)
	Shelter cats	7.7% (10/130)	Austria	Joachim et al. (2023)
	<i>Giardia</i> -positive domestic cats	57.9% (11/19)	Brazil	Souza et al. (2007)
Assemblage F	Various	1.9% (1/52)	China	Li et al. (2015)
	Stray cats	1.0% (1/102)	China	Zheng et al. (2015)
	Domestic cats	4.4% (7/160)	China	Xu et al. (2016)
	Domestic cats	9.0% (1/11)	China	Liu et al. (2017)
	Stray cats	2.8% (3/104)	China	Pan et al. (2018)
	Domestic cats	2.6% (9/346)	China	Li et al. (2019b)
	Wide range	3.3% (14/418)	China	Li et al. (2019a)
	Domestic cats	1.2% (2/171)	China	Wang et al. (2021)
	Domestic cats	6.3% (1/16)	China	Wu et al. (2022)
	Domestic cats	6.5% (3/46)	Columbia	Santín et al. (2006)
	<i>Giardia</i> -positive domestic cats	88.8% (16/18)	Czech Republic	Lecová et al. (2020)
	Domestic & stray cats	6.3% (16/255)	Czech Republic, Poland, Slovakia	Kváč et al. (2017)
	Domestic cats	0.6% (2/317)	Denmark	Enemark et al. (2020)
	<i>Giardia</i> -positive cats	48.7% (77/158)	Europe	Sprong et al. (2009)
	Domestic cats	9.6% (14/145)	Germany	Sommer et al. (2018)
	Domestic & shelter cats	2.3% (6/264) ^a	Greece	Kostopoulou et al. (2017)
	Domestic cats	1.0% (3/300)	Iran	Homayouni et al. (2019)
	Stray cats	0.7% (1/132)	Iran	Karimi et al. (2023)
	<i>Giardia</i> -positive cat	100% (1/1)	Italy	Lalle et al. (2005a)
	Domestic cats	2.8% (5/181)	Italy	Paoletti et al. (2011)
	Domestic cats	6.2% (9/146)	Italy	Mancianti et al. (2015)
	Shelter cats	1.5% (1/65)	Italy	Gil et al. (2017)
	<i>Giardia</i> -positive cat	100% (1/1)	Japan	Itagaki et al. (2005)
	Domestic cats	1.8% (1/55)	Japan	Yoshiuchi et al. (2010)
	Cats from cat café's and petshops	6.2% (20/321)	Japan	Suzuki et al. (2011)
	Domestic cats	12.1% (4/33)	Poland	Piekarska et al. (2016)
	Shelter cats	3.8% (6/158)	South Korea	Kwak and Seo (2020)
	Domestic cats	5.9% (2/34)	Spain	de Lucio et al. (2017)

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Table 1 (continued)

Species/Assemblage	Hosts	Prevalence	Country	Reference
Mixed F and A	<i>Giardia</i> -positive cats	66.6% (12/18)	Sweden	Lebbad et al. (2010)
	Domestic cats	31.1% (38/122)	UK	Krumrie et al. (2022)
	Cat colony cats	44.4% (8/18)	USA	Fayer et al. (2006)
	Domestic cats	4.4% (11/250)	USA	Vasilopoulos et al. (2007)
	<i>Giardia</i> -positive cat	100% (1/1)	USA	Miska et al. (2009)
	<i>Giardia</i> -positive domestic cats	56.7% (21/37)	USA	Saleh et al. (2019)
	<i>Giardia</i> -positive domestic cats	30.8% (4/13)	Canada	McDowall et al. (2011)
	Domestic & shelter cats	1.5% (4/264)	Greece	Kostopoulou et al. (2017)
	<i>Giardia</i> -positive domestic cats	10.8% (4/37)	USA	Saleh et al. (2019)
	Stray cats	A + B (<i>n</i> = 2)	Italy	Guadano Procesi et al. (2022)

^a 4 samples typed as assemblage A at 18S locus.

prevalent subtype in calves in the same area in a previous study (Holzhausen et al., 2019). In another study, *C. parvum* subtype IIcA5G3b was identified in two street dogs in Nigeria (Ayinmode et al., 2018b). The *C. parvum* IIc subtype family almost exclusively infects humans (Yang et al., 2021), suggesting that its presence in dogs may have been due to coprophagy of human faeces by the dogs.

There have been two reports of *C. muris* in dogs (Table 2). Oocysts of *C. muris* directly isolated from the stomach of infected mice have previously been orally inoculated in dogs with very few oocysts (7–12) detected between 7 and 12 days post-infection, and a few *C. muris* life-cycle stages were identified in the gastric mucosa of one dog (Aydin and Ozkul, 1996). These findings suggest that dogs may not be a natural host of *C. muris*, which is consistent with a study by Iseki et al. (1989). In one study in Texas, *Cryptosporidium* antigens were detected in 70% (49/70) of kennel dogs (Lupo et al., 2008), with six of the positives being amplified at the 18S locus and identified as *C. muris* and “*C. muris*-like”. Oocysts were also identified in these six samples (Lupo et al., 2008). The actual prevalence of *C. muris* in the study remains unknown, but the reported 70% prevalence is likely an over-estimate, as there have been numerous reports of false positives by *Cryptosporidium* enzyme antigen tests, especially considering the fact that most coproantigen kits are not expected to detect *C. muris* (CDC, 1999; Doing et al., 1999; Katanik et al., 2001).

There has also been a single report of *C. scrofarum* in one shelter dog in Greece (Kostopoulou et al., 2017), *C. meleagridis* in a domestic dog in the Czech Republic, and *C. ubiquitum* and rat genotype IV in individual dogs in China (Li et al., 2019a). These are all likely due to coprophagy.

Cryptosporidium oocyst shedding from cats and dogs appears to be low (Asahi et al., 1991; Cox et al., 2005; Hamnes et al., 2007; Labana et al., 2018; Smith et al., 2020). For example, oocyst concentrations ranging from 0 to 17 and 0–5000 oocysts per gram of faeces (OPG) have been reported in cats and dogs, respectively, using direct immunofluorescence microscopy (DFA) (Cox et al., 2005). A more recent larger study of 869 dogs reported median *Cryptosporidium* oocyst excretion of 3.8×10^3 OPG using DFA (Smith et al., 2020). In comparison, livestock, particularly cattle, shed much higher numbers of oocysts, with calves shedding $> 5.5 \times 10^6$ oocysts per day and $> 3 \times 10^{10}$ oocysts over a 6-day period (Nydam et al., 2001). Infected lambs can shed up to 3.7×10^9 OPG (Yang et al., 2014), while humans can shed up to 10^8 oocysts in a single bowel movement (Goodgame et al., 1993).

3. Giardia in cats and dogs

The prevalence of *Giardia* in cats (~1.3–44.4%) and dogs (~1–57.9%) varies widely but is generally < 10% (Feng and Xiao, 2011; Bouzid et al., 2015; López-Arias et al., 2019; Nagamori et al., 2020; Cai et al., 2021; Sui et al., 2022) (Tables 1 and 2). In one meta-analysis, the global prevalence of *Giardia* was calculated at 2.61% (112,513/4,309,451) in dogs and 2.33% (5807/248,195) in cats (Bouzid et al., 2015). In China, the overall prevalence of *Giardia* in dogs has been estimated at 12.3% by molecular testing (Zhao et al., 2022). Factors that influence the prevalence of *Giardia* include age, presence of diarrhoea,

socioeconomic situation, and origin of the samples (e.g. household pets versus kennel/shelter dogs/cats) and the number of doses of anthelmintic administered each year (Bugg et al., 1999; Bouzid et al., 2015; Cai et al., 2021; French et al., 2023; Murnik et al., 2023).

The highest prevalence of *Giardia* in dogs and cats has been reported from young animals (< 6 months) (Bouzid et al., 2015), in dogs from low- to middle-income areas, and from kennel/shelter dogs, where the potential for transmission is greater due to high densities of animals, continuous environmental contamination and frequent introduction of new animals. For example, the overall prevalence of *Giardia* in dogs was 44.4% (16/36) (Kuthyar et al., 2021) in Argentina, and 41.1% (69/168) in shelter dogs in Italy (Agresti et al., 2021). In another study from Spain, the highest prevalence was detected in breeding (45.8%; 11/24) and shelter dogs (40.4%; 88/218) (Adell-Aledon et al., 2018). In China, a recent study reported a prevalence of 40.3% (60/149) in shelter dogs compared to only 3.0% (6/199) in household dogs (Liao et al., 2020). Similarly in Portugal, 47.0% (23/49) of kennel dogs were *Giardia*-positive, but household dogs (*n* = 97) were negative (Ferreira et al., 2011). In cats, in one study in Italy, an overall prevalence of 37% (10/27) was reported for stray and domestic cats and all were assemblage A (Papini et al., 2007), and in the USA, 44.4% of colony cats were positive for *Giardia* (Fayer et al., 2006). However, another study reported similar prevalences of *Giardia* in shelter/breeding cats and household cats (Sommer et al., 2018).

Microscopy-based screening studies generally under-report the prevalence of *Giardia* due to sporadic shedding of cysts and the poor sensitivity of flotation assays and microscopy (Thompson, 2004). Many of the studies conducted the initial screens for *Giardia* by microscopy and then genotyped the positive samples, which can also underestimate the true prevalence. For example, one study detected *Giardia* in 57.9% (165/285) of kennel dogs by qPCR analysis of the 18S rRNA gene, compared to only 14.6% (39/268) detected by microscopy (Simonato et al., 2015). However, of the qPCR-positive samples, only 64% (106/165) were confirmed by nested PCR, and of these, only 74.5% (79/106) produced sequences (Simonato et al., 2015), emphasising the importance of sequencing to confirm positives.

Assemblages C and D are the most commonly reported *Giardia* in dogs, while assemblages F and A are most common in cats (Tables 1 and 2) (Ramírez-Ocampo et al., 2017; Cai et al., 2021; Zhao et al., 2022). In addition to assemblages C and D, assemblages A, B, E and F have also been reported in dogs. Similarly, assemblages B, C, D and occasionally E have also been reported in cats (Tables 1 and 2). Although assemblages C and D dominate in dogs, assemblages A and B (which infect humans) have also been detected. The detection of assemblages A and B in dogs and cats is thought to be mainly due to coprophagy and/or spill-back from human contact (Feng and Xiao, 2011; Cai et al., 2021), but this remains to be conclusively determined.

Despite the presence of host-adapted assemblages, assemblages A and B are common in cats and dogs (Tables 1 and 2). Between them, studies conducted in China revealed that assemblage A was the dominant assemblage in stray cats and dogs (Pan et al., 2018), and in police and farm dogs (92.6% of typed isolates; 25/27) (Li et al., 2013).

Table 2Zoonotic *Cryptosporidium* and *Giardia* species and assemblages identified in dogs globally.

Species/Assemblage	Hosts	Prevalence	Country	Reference
<i>Cryptosporidium</i>				
<i>C. canis</i>	Shelter & domestic dogs	0.3% (4/1400)	Australia	Palmer et al. (2008a,b)
	Wild dogs	20.5% (9/44)	Australia	Ng et al. (2011)
	Wild dogs	3.1% (1/32)	Australia	Nolan et al. (2013)
	Wild dogs	100% (2/2)	Australia	Zahedi et al. (2018)
	<i>Cryptosporidium</i> -positive dogs	100% (5/5)	Australia, USA	Morgan et al. (2000b)
	<i>Cryptosporidium</i> -positive domestic dogs	100% (9/9)	Brazil	Thomaz et al. (2007)
	Domestic dogs	3.1% (4/128)	Brazil	Alves et al. (2018)
	Domestic dogs	4.7% (3/64)	Brazil	de Oliveira et al. (2021)
	Domestic, shelter & pet shop dogs	1.9% (4/209)	Canada	Uehlinger et al. (2013)
	Various	1.7% (5/294)	Canada	Julien et al. (2019)
	Domestic dogs	0.3% (3/860)	Canada	Smith et al. (2020)
	Various	3.8% (29/770)	China	Jian et al. (2014)
	Domestic dogs	1.6% (5/315)	China	Gu et al. (2015)
	Domestic dogs	8.0% (39/485)	China	Xu et al. (2016)
	Dogs with diarrhoea	4.1% (20/485)	China	Yu et al. (2018)
	Various	2.9% (19/651)	China	Liao et al. (2020)
	Domestic dogs	4.6% (12/262)	China	Wang et al. (2021)
	Domestic dogs	1.6% (10/604)	China	Cao et al. (2022)
	Domestic dogs	1.8% (4/218)	Egypt	Elmahallawy et al. (2023)
	Stray & domestic dogs	5.0% (15/300)	Egypt	Khalifa et al. (2023)
	Domestic dogs	2.0% (3/150)	Finland	Rimhanen-Finne et al. (2007)
	Domestic dogs	2.6% (3/116)	France	Osmar et al. (2015)
	Domestic dogs	9.4% (33/349)	Germany	Murnik et al. (2022)
	Domestic & shelter dogs	0.2% (2/807)	Greece	Kostopoulou et al. (2017)
	Domestic dogs	0.6% (2/315)	Iran	Homayouni et al. (2019)
	Kennel & domestic dogs	0.4% (1/240)	Italy	Giangaspero et al. (2006)
	Kennel dogs	1.0% (3/285)	Italy	Simonato et al. (2015)
	Domestic dogs	0.1% (1/705)	Italy	Simonato et al. (2017)
	Domestic dogs	9.3% (13/140)	Japan	Abe et al. (2002)
	Domestic dogs	0.1% (1/906)	Japan	Yamamoto et al. (2009)
	Domestic dogs	3.9% (3/77)	Japan	Yoshiuchi et al. (2010)
	Kennel dogs	21.1% (66/314)	Japan	Itoh et al. (2019)
	Domestic dog	100% (1/1)	Nigeria	Ayinmode et al. (2018a)
	Domestic dogs	1.1% (3/264)	Poland	Piekara-Stepińska et al. (2021)
	Domestic dogs	3.6% (2/55)	Spain	de Lucio et al. (2017)
	Shelter dogs	2.6% (5/194)	Spain	Gil et al. (2017)
	Domestic dogs	1.6% (4/252)	Spain	Mateo et al. (2023)
	Temple dogs	2.1% (2/95)	Thailand	Koompapong et al. (2014)
	Domestic dogs	13.8% (15/109)	Thailand	Tangtrongsup et al. (2017)
	Domestic & shelter dogs	2.6% (8/301)	Thailand	Tangtrongsup et al. (2020)
	Monastery dogs	0.7% (4/540)	Thailand	Khine et al. (2021)
	Shelter dogs	4.2% (28/676)	UK	Rosanowski et al. (2018)
	Domestic dogs	0.8% (1/129)	USA	Wang et al. (2012)
	Domestic dogs	4.8% (4/84)	USA	Scorza and Lappin (2017)
	Domestic dogs	33.3% (1/3)	Vietnam	Iwashita et al. (2021)
<i>C. hominis</i>	Wild dogs	2.3% (1/44)	Australia	Ng et al. (2011)
	Shelter dogs	0.5% (1/194)	Italy	Gil et al. (2017)
<i>C. parvum</i>	Domestic dogs	1.6% (2/128)	Brazil	Alves et al. (2018)
	Domestic dogs	3.1% (2/64)	Brazil	de Oliveira et al. (2021)
	Various	0.3% (2/651)	China	Liao et al. (2020)
	Domestic dogs	1.7% (1/58)	Costa Rica	Scorza et al. (2011)
	Domestic dogs	50.0% (1/2)	Czech Republic	Hajdušek et al. (2004)
	Stray dogs	10.0% (2/20)	Egypt	El-Madawy et al. (2010)
	Domestic dogs	10.0% (5/50)	Egypt	Gharieb et al. (2018)
	Domestic dogs	1.2% (1/81)	Germany	Sotiriadou et al. (2013)
	Domestic dogs	0.6% (2/349)	Germany	Murnik et al. (2022)
	Kennel & privately-owned dogs	2.9% (7/240)	Italy	Giangaspero et al. (2006)
	Domestic dogs	1.4% (10/705)	Italy	Simonato et al. (2017)
	Street dogs	1.5% (3/203)	Nigeria	Ayinmode et al. (2018b)
	Domestic dogs	0.8% (2/264)	Poland	Piekara-Stepińska et al. (2021)
	Domestic dogs	0.4% (1/252)	Spain	Mateo et al. (2023)
	Domestic dogs	6.4% (7/109)	Thailand	Tangtrongsup et al. (2017)
	Domestic & shelter dogs	0.3% (1/301)	Thailand	Tangtrongsup et al. (2020)
	Farm dog	100% (1/1)	UK	Smith et al. (2010)
	Shelter dogs	0.3% (2/676)	UK	Rosanowski et al. (2018)
<i>C. muris</i>	Wide range	0.2% (1/601)	China	Li et al. (2019b)
	Street dogs	1.0% (2/203)	Nigeria	Ayinmode et al. (2018b)
	Kennel dogs	8.6% (6/70)	USA	Lupo et al. (2008)
<i>C. scrofarum</i>	Domestic & shelter dogs	0.1% (1/807)	Greece	Kostopoulou et al. (2017)
<i>C. meleagridis</i>	Domestic dogs	50.0% (1/2)	Czech Republic	Hajdušek et al. (2004)
<i>C. ubiquitum</i>	Wide range	0.4% (1/267)	China	Li et al. (2015)

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Table 2 (continued)

Species/Assemblage	Hosts	Prevalence	Country	Reference
<i>Giardia</i>				
Assemblage A				
	Domestic dogs	13.8% (5/36)	Argentina	Kuthyar et al. (2021)
	Shelter & domestic dogs	0.07% (1/1400)	Australia	Palmer et al. (2008a, b)
	Wild dogs	6.8% (3/44)	Australia	Ng et al. (2011)
	Wild dogs	3.1% (1/32)	Australia	Nolan et al. (2013)
	Various	3.5% (40/1159)	Belgium	Claerebout et al. (2009)
	Stray dogs	36.8% (7/19)	Brazil	Volotão et al. (2007)
	<i>Giardia</i> -positive stray dogs	100% (60/60)	Brazil	Fantinatti et al. (2018)
	Dogs from a remote indigenous community	8.4% (13/155)	Canada	Himsworth et al. (2010)
	Dogs in indigenous communities	13.8% (32/231)	Canada	Schurer et al. (2012)
	Domestic, shelter & pet shop dogs	1.4% (3/209)	Canada	Uehlinger et al. (2013)
	Domestic dogs	2.4% (5/209)	China	Li et al. (2012)
	Police & farm dogs	12.2% (25/205)	China	Li et al. (2013)
	Domestic dogs	3.2% (7/216)	China	Zheng et al. (2014)
	<i>Giardia</i> -positive domestic dogs	40.0% (8/20)	China	Tan et al. (2016)
	Domestic dogs	4.7% (23/485)	China	Xu et al. (2016)
	Stray dogs	4.9% (26/527)	China	Pan et al. (2018)
	Domestic dogs	0.2% (1/604)	China	Cao et al. (2022)
	Domestic dogs	18.2% (2/11) ^b	Côte d'Ivoire	Berrilli et al. (2012)
	<i>Giardia</i> -positive domestic & kennel dogs	2.7% (2/96)	Croatia	Beck et al. (2012)
	Domestic dogs	10.2% (10/98)	Cuba	Puebla et al. (2017)
	Stray & domestic dogs	0.6% (2/300)	Egypt	Khalifa et al. (2023)
	<i>Giardia</i> -positive dog samples	22.8% (137/600)	Europe	Sprong et al. (2009)
	<i>Giardia</i> -positive domestic dogs	60.0% (33/55)	Germany	Leonhard et al. (2007)
	Domestic dogs	6.2% (5/81)	Germany	Sotiriadou et al. (2013)
	<i>Giardia</i> -positive domestic dogs	19.5% (24/123)	Germany	Pallant et al. (2015)
	Domestic dogs	0.5% (2/376)	Germany	Sommer et al. (2018)
	Domestic dogs	6.5% (2/31)	Germany	Rehbein et al. (2019)
	Domestic dogs	2.3% (9/386)	Germany	Murnik et al. (2023)
	Domestic & shelter dogs	0.9% (7/807) ^a	Greece	Kostopoulou et al. (2017)
	Domestic dogs	4.9% (5/101)	India	Traub et al. (2004)
	Domestic dogs	0.5% (1/212)	India	Utaaker et al. (2018)
	Domestic dogs	0.3% (1/315)	Iran	Homayouni et al. (2019)
	<i>Giardia</i> -positive domestic dogs	11.8% (2/17)	Italy	Berrilli et al. (2004)
	Domestic & kennel dogs	0.8% (2/240)	Italy	Paolletti et al. (2008)
	Domestic dogs	6.3% (8/127)	Italy	Scaramozzino et al. (2009)
	Stray dogs	64.3% (9/14)	Italy	Marangi et al. (2010)
	Domestic dogs	0.3% (2/655)	Italy	Pipia et al. (2014)
	Sheltered dogs	3.6% (7/194)	Italy	Gil et al. (2017)
	Shelter dogs	9.5% (16/168)	Italy	Agresti et al. (2021)
	Domestic dogs	19.5% (44/225)	Jamaica	Lee et al. (2017)
	<i>Giardia</i> -positive domestic dogs	66.6% (14/21)	Japan	Itagaki et al. (2005)
	Shelter dogs	1.0% (2/202)	Korea	Shin et al. (2015)
	<i>Giardia</i> -positive domestic dogs	100% (2/2)	Mexico	Ponce-Macotela et al. (2002)
	<i>Giardia</i> -positive domestic dogs	100% (11/11)	Mexico	Elgio-Garcia et al. (2005)
	Puppies	100% (5/5)	Mexico	Lalle et al. (2005b)
	Various	1.1% (2/189)	Netherlands	Uiterwijk et al. (2020)
	Domestic dogs	3.5% (2/58)	Nicaragua	Roegner et al. (2019)
	Domestic dogs	1.4% (5/348)	Spain	Adell-Aledon et al. (2018)
	Domestic dogs	0.4% (1/252)	Spain	Mateo et al. (2023)
	<i>Giardia</i> -positive dog samples	3.6% (1/28)	Sweden	Lebbad et al. (2010)
	Temple dogs	2.2% (5/229)	Thailand	Inpankaew et al. (2007)
	Temple dogs	14.4% (33/229)	Thailand	Traub et al. (2009)
	Monastery dogs	0.4% (2/540)	Thailand	Khine et al. (2021)
	Domestic dogs	0.7% (1/152)	The Netherlands	Overgaauw et al. (2009)
	<i>Giardia</i> -positive shelter dogs	2.4% (1/41)	UK	Upjohn et al. (2010)
	Domestic dogs	1.9% (1/52)	UK	Krumrie et al. (2022)
	<i>Giardia</i> -positive domestic dogs	11.1% (1/9)	Unknown	Read et al. (2004)
	<i>Giardia</i> -positive domestic dogs	28.0% (36/128)	USA	Covacin et al. (2011)
	Domestic dogs	13.3% (16/120)	USA	Munoz and Mayer (2016)
Assemblage B	Domestic dog	100% (1/1)	Argentina	Minvielle et al. (2008)
	Various	0.3% (4/1159)	Belgium	Claerebout et al. (2009)
	Domestic dogs	3.1% (1/32)	Brazil	Colli et al. (2015)
	Domestic dogs	2.2% (2/94)	Cambodia	Inpankaew et al. (2014)
	Domestic dogs	1.9% (6/315)	China	Gu et al. (2015)
	Domestic dogs	0.2% (1/485)	China	Xu et al. (2016)
	<i>Giardia</i> -positive domestic & kennel dogs	23.3% (17/96)	Croatia	Beck et al. (2012)
	<i>Giardia</i> -positive dog samples	8.8% (53/600)	Europe	Sprong et al. (2009)
	<i>Giardia</i> -positive domestic dogs	6.5% (8/123)	Germany	Pallant et al. (2015)
	Domestic & shelter dogs	5.7% (46/807)	Greece	Kostopoulou et al. (2017)
	Domestic dogs	2.0% (2/101)	India	Traub et al. (2004)
	Domestic dogs	0.5% (1/212)	India	Utaaker et al. (2018)
	Kennel dogs	0.3% (1/318)	Italy	Simonato et al. (2015)
	Sheltered dogs	4.1% (8/194)	Italy	Gil et al. (2017)
	Domestic dogs	0.1% (1/705)	Italy	Simonato et al. (2017)
	Shelter dogs	3.6% (6/168)	Italy	Agresti et al. (2021)

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Table 2 (continued)

Species/Assemblage	Hosts	Prevalence	Country	Reference
Assemblage C	Domestic dogs	8.6% (5/58)	Nicaragua	Roegner et al. (2019)
	Domestic dogs	0.8% (1/128)	Poland	Piekarska et al. (2016)
	Household & kennel dogs	1.6% (2/126)	Portugal	Ferreira et al. (2011)
	Domestic dogs	2.3% (8/348)	Spain	Adell-Aledor et al. (2018)
Assemblage C	Temple dogs	3.9% (9/229)	Thailand	Traub et al. (2009)
	Domestic dogs	10.8% (51/473)	Turkey	Gultekin et al. (2017)
	<i>Giardia</i> -positive domestic dogs	22.2% (2/9)	Unknown	Read et al. (2004)
	<i>Giardia</i> -positive domestic dogs	40.6% (52/128)	USA	Covacini et al. (2011)
	<i>Giardia</i> -positive domestic dogs	90.9% (10/11)	Australia	Monis et al. (1998, 2003)
	Shelter & domestic dogs	2.9% (41/1400)	Australia	Palmer et al. (2008a, b)
	Wild dogs	11.4% (5/44)	Australia	Ng et al. (2011)
	Shelter dogs	1.4% (1/70)	Austria	Joachim et al. (2023)
	Various	2.2% (26/1159)	Belgium	Claerebout et al. (2009)
	<i>Giardia</i> -positive domestic dogs	29.9% (7/27)	Brazil	Souza et al. (2007)
Assemblage C	Domestic dogs	3.1% (1/32)	Brazil	Colli et al. (2015)
	Domestic dogs	1.4% (2/147)	Brazil	Paim Arruda Trevisan et al. (2020)
	<i>Giardia</i> -positive domestic dogs	16.6% (2/12)	Brazil	Silva et al. (2022)
	Domestic dogs	4.3% (4/94)	Cambodia	Inpankaew et al. (2014)
	<i>Giardia</i> -positive domestic dogs	30.6% (23/75)	Canada	McDowall et al. (2011)
	Domestic, shelter & pet shop dogs	4.8% (10/209)	Canada	Uehlinger et al. (2013)
	Various	0.7% (2/294)	Canada	Julien et al. (2019)
	Domestic dogs	1.6% (14/860)	Canada	Smith et al. (2020)
	Police & farm dogs	1.0% (2/205)	China	Li et al. (2013)
	Domestic dogs	0.5% (1/216)	China	Zheng et al. (2014)
	Wide range	2.6% (7/267)	China	Li et al. (2015)
	<i>Giardia</i> -positive domestic dogs	25.0% (5/20)	China	Tan et al. (2016)
	Domestic dogs	5.4% (26/485)	China	Xu et al. (2016)
	Domestic dogs	15.8% (3/19)	China	Liu et al. (2017)
	Stray dogs	5.7% (9/159)	China	Zhang et al. (2017)
	Stray dogs	3.4% (18/527)	China	Pan et al. (2018)
	Wide range	4.5% (27/601)	China	Li et al. (2019b)
	Various	0.8% (5/651)	China	Liao et al. (2020)
Assemblage C	Domestic dogs	1.5% (4/262)	China	Wang et al. (2021)
	Domestic dogs	2.3% (14/604)	China	Cao et al. (2022)
	Domestic dogs	2.9% (13/448)	China	Sui et al. (2022)
	Domestic dogs	3.8% (4/105)	China	Wu et al. (2022)
	Domestic dogs	3.0% (3/100)	Colombia	Hernandez et al. (2021)
	Domestic dogs	9.1% (1/11)	Côte d'Ivoire	Berrilli et al. (2012)
	<i>Giardia</i> -positive domestic & kennel dogs	36.9% (27/73)	Croatia	Beck et al. (2012)
	<i>Giardia</i> -positive domestic dogs	38.8% (21/54)	Czech Republic	Lecová et al. (2020)
	Domestic & shelter dogs	3.0% (50/1645)	Eastern European countries	Sommer et al. (2015)
	Domestic dogs	0.5% (1/218)	Egypt	Elmahallawy et al. (2023)
	<i>Giardia</i> -positive dog samples	31.8% (191/600)	Europe	Sprong et al. (2009)
	Domestic dogs	2.0% (3/150)	Finland	Rimhanen-Finne et al. (2007)
	<i>Giardia</i> -positive dogs	76.9% (30/39)	France	Kaufmann et al. (2022)
	<i>Giardia</i> -positive domestic dogs	35.9% (33/92)	Germany	Barutzki et al. (2007)
	<i>Giardia</i> -positive domestic dogs	36.4% (20/55) ^b	Germany	Leonhard et al. (2007)
	<i>Giardia</i> -positive domestic dogs	42.3% (52/123)	Germany	Pallant et al. (2015)
	Domestic dogs	10.1% (38/376)	Germany	Sommer et al. (2018)
	Domestic dogs	3.2% (1/31)	Germany	Rehbein et al. (2019)
	Domestic dogs	11.2% (43/386)	Germany	Murnik et al. (2023)
Assemblage C	Domestic & shelter dogs	5.7% (46/807)	Greece	Kostopoulou et al. (2017)
	Domestic dogs	2.2% (5/229)	Hungary	Szénási et al. (2007)
	Domestic dogs	4.2% (9/212)	India	Utaaker et al. (2018)
	Domestic dogs	0.9% (3/315)	Iran	Homayouni et al. (2019)
	<i>Giardia</i> -positive domestic dogs	64.7% (11/17)	Italy	Berrilli et al. (2004)
	Domestic & kennel dogs	1.2% (3/240)	Italy	Paoletti et al. (2008)
	Domestic dogs	11.0% (14/127)	Italy	Scaramozzino et al. (2009)
	Domestic dogs	2.6% (17/655)	Italy	Pipia et al. (2014)
	Domestic dogs	2.4% (6/253)	Italy	Zanzani et al. (2014)
	Domestic dogs	3.0% (15/502)	Italy	Paoletti et al. (2015)
Assemblage C	Kennel dogs	17.2% (49/285)	Italy	Simonato et al. (2015)
	Domestic dogs	1.3% (9/705)	Italy	Simonato et al. (2017)
	Stray dogs	2.3% (6/262)	Italy	Liberato et al. (2018)
	Dogs with primary chronic enteropathy	2.2% (1/47)	Italy	Perrucci et al. (2020)
	Shelter dogs	1.2% (2/168)	Italy	Agresti et al. (2021)
	<i>Giardia</i> -positive domestic dogs	4.8% (1/21)	Japan	Itagaki et al. (2005)
	Pet shop puppies	0.5% (9/1794)	Japan	Itoh et al. (2011)
	Shelter dogs	0.5% (1/202)	Korea	Shin et al. (2015)
	Domestic dogs	1.4% (9/640)	Korea	Kim et al. (2019)
	Various	9.0% (17/189)	Netherlands	Uiterwijk et al. (2020)
Assemblage C	Domestic dogs	2.0% (2/100)	Nicaragua	Lebbad et al. (2008)
	Domestic dogs	5.2% (3/58)	Nicaragua	Roegner et al. (2019)
	Domestic dogs	1.5% (9/605)	Peru	Cooper et al. (2010)
	Domestic & shelter dogs	0.7% (1/148)	Poland	Solarczyk and Majewska (2010)
	Sled dogs	8.0% (2/25)	Poland	Bajer et al. (2011)

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Table 2 (continued)

Species/Assemblage	Hosts	Prevalence	Country	Reference
	Domestic dogs	14.0% (18/128)	Poland	Piekarska et al. (2016)
	Household & kennel dogs	11.9% (15/126)	Portugal	Ferreira et al. (2011)
	Domestic dogs	3.7% (3/80)	Portugal	Pereira et al. (2021)
	Domestic dogs	20.5% (8/39)	Romania	Adriana et al. (2016)
	Domestic dogs	5.4% (3/55)	Spain	de Lucio et al. (2017)
	Domestic dogs	0.3% (2/348)	Spain	Adell-Aledon et al. (2018)
	Domestic dogs	1.6% (4/252)	Spain	Mateo et al. (2023)
	<i>Giardia</i> -positive dog samples	28.6% (8/28)	Sweden	Lebbad et al. (2010)
	Domestic dogs	2.4% (1/42)	Taiwan	Liang et al. (2012)
	Stray dogs	5.9% (7/118)	Taiwan	Tseng et al. (2014)
	Temple dogs	0.4% (1/229)	Thailand	Inpankaew et al. (2007)
	Temple dogs	2.2% (5/229)	Thailand	Traub et al. (2009)
	Domestic dogs	7.3% (8/109)	Thailand	Tangtrongsup et al. (2017)
	Domestic & shelter dogs	0.3% (1/301)	Thailand	Tangtrongsup et al. (2020)
	Monastery dogs	1.3% (7/540)	Thailand	Khine et al. (2021)
	Stray dogs	3.8% (4/104)	Trinidad and Tobago	Mark-Carew et al. (2013)
	<i>Giardia</i> -positive shelter dogs	24.4% (10/41)	UK	Upjohn et al. (2010)
	Domestic dogs	19.2% (10/52)	UK	Krumrie et al. (2022)
	<i>Giardia</i> -positive domestic dogs	44.4% (4/9)	Unknown	Read et al. (2004)
	<i>Giardia</i> -positive domestic dogs	100% (15/15)	USA	Sulaiman et al. (2003)
	<i>Giardia</i> -positive dog samples	33.3% (1/3)	USA	Miska et al. (2009)
	<i>Giardia</i> -positive domestic dogs	14.8% (19/128)	USA	Covacin et al. (2011)
	Domestic dogs	0.8% (1/129)	USA	Wang et al. (2012)
	Shelter dogs	0.1% (1/672) ^b	USA	Johansen et al. (2014)
	Domestic dogs	4.8% (10/209)	Vietnam	Nguyen et al. (2018)
Assemblage D	<i>Giardia</i> -positive domestic dogs	9.1% (10/11)	Australia	Monis et al. (1998, 2003)
	Shelter & domestic dogs	3.1% (44/1400)	Australia	Palmer et al. (2008a, b)
	Various	4.2% (49/1159)	Belgium	Claerebout et al. (2009)
	<i>Giardia</i> -positive domestic dogs	74.1% (20/27)	Brazil	Souza et al. (2007)
	Kennel dogs	16.5% (16/97)	Brazil	Fava et al. (2016)
	Domestic dogs	4.3% (4/94)	Cambodia	Inpankaew et al. (2014)
	<i>Giardia</i> -positive domestic dogs	68.0% (51/75)	Canada	McDowall et al. (2011)
	Domestic, shelter & pet shop dogs	9.1% (19/209)	Canada	Uehlinger et al. (2013)
	Various	1.0% (3/294)	Canada	Julien et al. (2019)
	Domestic dogs	1.5% (13/860)	Canada	Smith et al. (2020)
	Domestic dogs	8.6% (18/209)	China	Li et al. (2012)
	Domestic dogs	0.5% (1/216)	China	Zheng et al. (2014)
	Domestic dogs	1.3% (4/315)	China	Gu et al. (2015)
	<i>Giardia</i> -positive domestic dogs	35.0% (7/20)	China	Tan et al. (2016)
	Domestic dogs	11.9% (58/485)	China	Xu et al. (2016)
	Domestic dogs	15.8% (3/19)	China	Liu et al. (2017)
	Stray dogs	5.7% (9/159)	China	Zhang et al. (2017)
	Stray dogs	2.5% (13/527)	China	Pan et al. (2018)
	Dogs with diarrhoea	3.1% (15/485)	China	Yu et al. (2018)
	Wide range	4.3% (26/601)	China	Li et al. (2019b)
	Various	2.1% (14/651)	China	Liao et al. (2020)
	Domestic dogs	12.2% (32/262)	China	Wang et al. (2021)
	Domestic dogs	1.2% (7/604)	China	Cao et al. (2022)
	Domestic dogs	3.1% (14/448)	China	Sui et al. (2022)
	Domestic dogs	1.0% (1/105)	China	Wu et al. (2022)
	Domestic dogs	3.0% (3/100)	Columbia	Hernandez et al. (2021)
	Domestic dogs	3.4% (2/58)	Costa Rica	Scorza et al. (2011)
	Domestic dogs	18.2% (2/11)	Côte d'Ivoire	Berrilli et al. (2012)
	<i>Giardia</i> -positive domestic & kennel dogs	36.9% (27/73)	Croatia	Beck et al. (2012)
	<i>Giardia</i> -positive domestic dogs	59.2% (32/54)	Czech Republic	Lecová et al. (2020)
	Domestic & shelter dogs	4.1% (68/1645)	Eastern European countries	Sommer et al. (2015)
	Domestic dogs	0.4% (4/986)	Egypt	Abdelaziz and Sorour (2021)
	Domestic dogs	1.4% (3/218)	Egypt	Elmahallawy et al. (2023)
	Stray & domestic dogs	1.6% (5/300)	Egypt	Khalifa et al. (2023)
	<i>Giardia</i> -positive dog samples	35.8% (215/600)	Europe	Sprong et al. (2009)
	Domestic dogs	2.6% (4/150)	Finland	Rimhanen-Finne et al. (2007)
	<i>Giardia</i> -positive dogs	23.1% (9/39)	France	Kaufmann et al. (2022)
	<i>Giardia</i> -positive domestic dogs	54.3% (50/92)	Germany	Barutzki et al. (2007)
	<i>Giardia</i> -positive domestic dogs	3.6% (2/55)	Germany	Leonhard et al. (2007)
	<i>Giardia</i> -positive domestic dogs	56.1% (69/123)	Germany	Pallani et al. (2015)
	Domestic dogs	12.5% (47/376)	Germany	Sommer et al. (2018)
	Domestic dogs	6.5% (2/31)	Germany	Rehbein et al. (2019)
	Domestic dogs	10.4% (40/386)	Germany	Murnik et al. (2023)
	Domestic & shelter dogs	2.7% (22/807)	Greece	Kostopoulou et al. (2017)
	Domestic dogs	6.5% (15/229)	Hungary	Szénási et al. (2007)
	Domestic dogs	3.3% (7/212)	India	Utaaker et al. (2018)
	Domestic dogs	0.6% (2/315)	Iran	Homayouni et al. (2019)
	<i>Giardia</i> -positive domestic dogs	5.8% (1/17)	Italy	Berrilli et al. (2004)
	Domestic & kennel dogs	10.4% (25/240)	Italy	Paoletti et al. (2008)
	Domestic dogs	3.2% (4/127)	Italy	Scaramozzino et al. (2009)
	Domestic dogs	3.5% (23/655)	Italy	Pipia et al. (2014)

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Table 2 (continued)

Species/Assemblage	Hosts	Prevalence	Country	Reference
Mixed C & D	Domestic dogs	2.0% (5/253)	Italy	Zanzani et al. (2014)
	Domestic dogs	0.4% (2/502)	Italy	Paoletti et al. (2015)
	Domestic dogs	10.2% (29/285)	Italy	Simonato et al. (2015)
	Domestic dogs	1.6% (11/705)	Italy	Simonato et al. (2017)
Assemblage E	Stray dogs	7.2% (19/262)	Italy	Liberato et al. (2018)
	Dogs with primary chronic enteropathy	10.6% (5/47)	Italy	Perrucci et al. (2020)
	<i>Giardia</i> -positive domestic dogs	100% (6/6)	Italy	Ciuca et al. (2021)
	<i>Giardia</i> -positive domestic dogs	100% (4/4)	Japan	Abe et al. (2003)
	<i>Giardia</i> -positive domestic dogs	28.6% (6/21)	Japan	Itagaki et al. (2005)
	Domestic dogs	2.6% (2/77)	Japan	Yoshiuchi et al. (2010)
	Pet shop puppies	1.1% (20/1794)	Japan	Itoh et al. (2011)
	Domestic dogs	1.4% (9/640)	Korea	Kim et al. (2019)
	Various	25.9% (49/189)	Netherlands	Uiterwijk et al. (2020)
	Domestic dogs	5.0% (5/100)	Nicaragua	Lebbad et al. (2008)
Assemblage F	Domestic dogs	5.2% (3/58)	Nicaragua	Roegner et al. (2019)
	Domestic dogs	5.3% (32/605)	Peru	Cooper et al. (2010)
	Domestic & shelter dogs	0.7% (1/148)	Poland	Solarczyk and Majewska (2010)
	Domestic dogs	1.6% (2/128)	Poland	Piekarska et al. (2016)
	Household & kennel dogs	9.5% (12/126)	Portugal	Ferreira et al. (2011)
	Domestic dogs	13.7% (11/80)	Portugal	Pereira et al. (2021)
	Domestic dogs	27.6% (29/39)	Romania	Adriana et al. (2016)
	Domestic dogs	3.7% (13/348)	Spain	Adell-Aledon et al. (2018)
	Domestic dogs	1.6% (4/252)	Spain	Mateo et al. (2023)
	<i>Giardia</i> -positive dog samples	50.0% (14/28)	Sweden	Lebbad et al. (2010)
Assemblage G	Domestic dogs	7.1% (3/42)	Taiwan	Liang et al. (2012)
	Stray dogs	3.4% (4/118)	Taiwan	Tseng et al. (2014)
	Temple dogs	1.3% (3/229)	Thailand	Inpankaew et al. (2007)
	Temple dogs	5.7% (13/229)	Thailand	Traub et al. (2009)
	Domestic dogs	11.0% (12/109)	Thailand	Tangtrongsup et al. (2017)
	Domestic & shelter dogs	6.0% (18/301)	Thailand	Tangtrongsup et al. (2020)
	Monastery dogs	1.3% (7/540)	Thailand	Khine et al. (2021)
	Stray dogs	20.2% (21/104)	Trinidad and Tobago	Mark-Carew et al. (2013)
	<i>Giardia</i> -positive shelter dogs	70.7% (29/41)	UK	Upjohn et al. (2010)
	<i>Giardia</i> -positive domestic dogs	22.2% (2/9)	Unknown	Read et al. (2004)
Assemblage H	<i>Giardia</i> -positive dog samples	66.6% (2/3)	USA	Miska et al. (2009)
	<i>Giardia</i> -positive domestic dogs	16.4% (21/128)	USA	Covacini et al. (2011)
	Domestic dogs	1.5% (2/129)	USA	Wang et al. (2012)
	Domestic dogs	0.8% (1/120)	USA	Munoz and Mayer (2016)
	Domestic dogs	4.8% (10/209)	Vietnam	Nguyen et al. (2018)
	Domestic dogs	2.7% (1/36)	Argentina	Kuthyar et al. (2021)
	Shelter & domestic dogs	0.1% (2/1400)	Australia	Palmer et al. (2008a, b)
	Domestic, shelter & pet shop dogs	0.5% (1/209)	Canada	Uehlinger et al. (2013)
	Domestic & stray dogs	8.5% (80/940)	China	Qi et al. (2016)
	Wide range	0.8% (5/601)	China	Li et al. (2019b)
Assemblage I	Domestic dogs	1.1% (5/448)	China	Sui et al. (2022)
	<i>Giardia</i> -positive domestic dogs	1.8% (1/54)	Czech Republic	Lecová et al. (2020)
	<i>Giardia</i> -positive dogs	2.6% (1/39)	France	Kaufmann et al. (2022)
	<i>Giardia</i> -positive domestic dogs	8.7% (8/92)	Germany	Barutzki et al. (2007)
	Domestic dogs	3.1% (12/386) ^b	Germany	Murnik et al. (2023)
	Domestic & shelter dogs	1.9% (15/807)	Greece	Kostopoulou et al. (2017)
	Domestic dogs	0.4% (1/229)	Hungary	Szénási et al. (2007)
	<i>Giardia</i> -positive domestic dogs	5.8% (1/17)	Italy	Berrilli et al. (2004)
	Domestic dogs	0.6% (3/502)	Italy	Paoletti et al. (2015)
	Various	7.4% (14/189)	Netherlands	Uiterwijk et al. (2020)
Assemblage J	Domestic dogs	1.0% (1/100)	Nicaragua	Lebbad et al. (2008)
	Domestic dogs	4.3% (26/605)	Peru	Cooper et al. (2010)
	Domestic dogs	0.8% (1/128)	Poland	Piekarska et al. (2016)
	Domestic dogs	5.0% (4/80)	Portugal	Pereira et al. (2021)
	Domestic dogs	2.6% (1/39)	Romania	Adriana et al. (2016)
	Domestic dogs	0.4% (1/252)	Spain	Mateo et al. (2023)
	<i>Giardia</i> -positive dog samples	17.8% (5/28)	Sweden	Lebbad et al. (2010)
	Domestic dogs	0.9% (1/109)	Thailand	Tangtrongsup et al. (2017)
	<i>Giardia</i> -positive shelter dogs	2.4% (1/41)	UK	Upjohn et al. (2010)
	Domestic dogs	0.8% (1/129)	USA	Wang et al. (2012)
Assemblage K	Shelter dogs	24.4% (164/672)	USA	Johansen et al. (2014)
	Domestic dogs	2.3% (7/300)	USA	Hascall et al. (2016)
	Domestic dogs	10.7% (9/84)	USA	Scorza and Lappin (2017)
	Wild dogs	4.5% (2/44)	Australia	Ng et al. (2011)
	Domestic dogs	1.4% (2/143)	Brazil	Harvey et al. (2023)
	Domestic, shelter & pet shop dogs	0.5% (1/209)	Canada	Uehlinger et al. (2013)
	Various	0.3% (1/294)	Canada	Julien et al. (2019)
	Wide range	1.9% (5/267)	China	Li et al. (2015)
	<i>Giardia</i> -positive dog samples	0.8% (5/600)	Europe	Sprong et al. (2009)
	Domestic dogs	0.6% (1/150)	Finland	Rimhanen-Finne et al. (2007)
Assemblage L	Domestic dogs	0.5% (2/376)	Germany	Sommer et al. (2018)
	Domestic dogs	0.5% (1/212)	India	Utaaker et al. (2018)

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Table 2 (continued)

Species/Assemblage	Hosts	Prevalence	Country	Reference
Assemblage F	Domestic dogs	2.6% (1/39)	Romania	Adriana et al. (2016)
	Stray dogs	1.0% (1/104)	Trinidad and Tobago	Mark-Carew et al. (2013)
	<i>Giardia</i> -positive domestic dogs	16.6% (2/12)	Brazil	Silva et al. (2022)
	Dogs with diarrhoea	0.2% (1/485)	China	Yu et al. (2018)
	Various	0.1% (1/651)	China	Liao et al. (2020)
	<i>Giardia</i> -positive domestic dogs	4.9% (6/123)	Germany	Pallant et al. (2015)
Other mixtures	Domestic dogs	0.5% (2/376)	Germany	Sommer et al. (2018)
	Domestic dogs	15.4% (8/52)	UK	Krumrie et al. (2022)
	Domestic dogs	C + A (1/36)	Argentina	Kuthyar et al. (2021)
	Wild dogs	A + E (3/44)	Australia	Ng et al. (2011)
	Domestic dogs	A + C (1/147)	Brazil	Paim Arruda Trevisan et al. (2020)
	Domestic, shelter & pet shop dogs	A + C (1/209)	Canada	Uehlinger et al. (2013)
		B + D (1/209)		
		D + E (1/209)		
	Domestic dogs	5.1% (11/216) ^b	China	Zheng et al. (2014)
	Domestic dogs	A + B (4/105)	China	Wu et al. (2022)
		A + D (1/105)		
		A + B + D (2/105)		
	<i>Giardia</i> -positive domestic dogs	A + D (1/92)	Germany	Barutzki et al. (2007)
	Domestic dogs	A + B (1/31)	Germany	Rehbein et al. (2019)
	<i>Giardia</i> -positive domestic dogs	A + C (2/17)	Italy	Berrilli et al. (2004)
	Temple dogs	A + B (3/229) A + D (1/229)	Thailand	Inpankaew et al. (2007)
	Domestic dogs	A + B (38/473)	Turkey	Gultekin et al. (2017)

^a Also contained mixtures with C and D.^b Various mixtures.

Assemblage A was also the dominant assemblage in dogs in Japan (66.6%; 14/21 typed isolates) (Itagaki et al., 2005) and in Germany (60%; 33/55 typed isolates) (Leonhard et al., 2007). Assemblage A was the only assemblage identified in slum and stray dogs in Brazil (Volotão et al., 2007; Fantinatti et al., 2018), domestic dogs in Jamaica (Lee et al., 2017), and dogs in indigenous communities in Saskatchewan, Canada (32/32 typed isolates) (Schurer et al., 2012). Subtyping of assemblage A in cats and dogs has identified mostly sub-assemblage AI, which is mainly confined to animal hosts (Lalle et al., 2005b; Souza et al., 2007; Volotão et al., 2007; Marangi et al., 2010; Zheng et al., 2015; Saleh et al., 2019; Mateo et al., 2023). However, some studies have identified sub-assemblage AII in cats and dogs (e.g. Claerebout et al., 2009; Traub et al., 2004; Gil et al., 2017; Ito et al., 2017; Agresti et al., 2021; Guadano Procesi et al., 2022), which is mainly human-associated and even sub-assemblage AIII (Claerebout et al., 2009; Guadano Procesi et al., 2022), which is wild ruminant-associated (Klotz et al., 2022). Differences in sub-assemblage A assignment may be related to the typing locus used, as a meta-analysis revealed that subtyping assemblage A at multiple loci produced divergent results, with sub-assemblage AI dominating amongst assemblage A identified in dogs, when the *tpi* locus was used, and sub-assemblage AII dominating at the *bg* locus (Cai et al., 2021). Therefore, the identity of A sub-assemblages in cats and dogs is unclear.

Assemblage B has been reported less frequently in cats and dogs than assemblage A (Feng and Xiao, 2011; Cai et al., 2021) (Tables 1 and 2). However, assemblage B was identified as the dominant assemblage in these animals in some studies. For example, in Spain, assemblage B was detected in 42.1% (8/19) of typed *Giardia*-positive dogs, followed by assemblage A (7/19), with assemblages C and D only detected in 4 samples (Gil et al., 2017). A study in the USA, also reported assemblage B as the dominant assemblage (41% of typed samples) in household dogs, followed by assemblage A (28% of typed samples) (Covacini et al., 2011). In Turkey, assemblage B was reported in all 89 typed *Giardia*-positive dogs, but 42.7% (38/89) of these were mixed infections with assemblage A (Gultekin et al., 2017). Moreover, also in Turkey, assemblage B was the only assemblage detected in 30 typed *Giardia*-positive symptomatic cats (Sursal et al., 2020). Interestingly, in a study conducted in Romania, assemblage D was the only assemblage detected in 80% (8/10) of cats screened (Adriana et al., 2016). In that study, seven of the cats positive for assemblage D resided in the same house, and

although dogs were not present, all cats had outdoor access (Adriana et al., 2016).

Many of these studies, particularly earlier studies, have relied on one locus for identification of *Giardia* assemblages in cats and dogs. Due to discordance in genotyping results between loci, however, it is possible for *Giardia* isolates from cats and dogs to be typed as ‘zoonotic’ at one locus but ‘host-specific’ at another (Cacciò and Ryan, 2008), which makes interpretation difficult. In many studies, where multiple loci were used, there was significant discordance between loci (e.g. Covacini et al., 2011; Beck et al., 2012; Johansen et al., 2014; Pallant et al., 2015; Kostopoulou et al., 2017), with one study reporting that the 18S locus preferentially amplified assemblages B, D and C, whereas the *bg* locus preferentially amplified assemblages A and B (Covacini et al., 2011).

Giardia cyst excretion is highly variable and sporadic and relatively few studies have examined cyst excretion in dogs and cats. In DFA analysis, cyst concentrations ranging from 0 to 7143 and 0–6061 (median 835) cysts per gram of faeces (CPG) have been reported in cats and dogs, respectively (Cox et al., 2005). Another study of *Giardia* cyst shedding in 104 dogs and 21 cats from Germany, reported that 92.3% (96/104) of dogs and 76.2% (16/21) of cats, shed low numbers of cysts (< 50 cysts per slide) (Sommer et al., 2018). In Canada, a study of 869 dog faecal samples from urban parks reported median *Giardia* cyst excretion rates of 1.0×10^4 CPG (Smith et al., 2020). Similarly, studies of *Giardia*-positive dogs in Italy (Ciucca et al., 2021) and France (Kaufmann et al., 2022) reported a mean of 2×10^4 CPG (2.7×10^3 – 4.8×10^4) and a geometric mean excretion of 2.1×10^5 CPG, respectively. A more recent study of *Giardia* in dogs and cats in Egypt using qPCR reported that 56% (47/84) of the *Giardia* positives from dogs and 90.7% (39/43) of the *Giardia* positives from cats had high qPCR CT values indicating “light infections” (Elmahallawy et al., 2023). In comparison, cattle can shed $> 1 \times 10^6$ *Giardia* cysts per day (Nydam et al., 2001), while infected humans can shed 10^8 – 10^9 cysts per day (Danciger and Lopez, 1975; Rendtorff, 1979).

4. Zoonotic risk of cryptosporidiosis and giardiasis from cats and dogs

The dominant *Cryptosporidium* species and *Giardia* assemblages infecting cats are *C. felis* and assemblages F and A, compared with *C. canis* and assemblages C and D in dogs (Tables 1 and 2). Although

Table 3Reports of *Cryptosporidium felis*, *C. canis* and *Giardia* assemblages C, D and F in humans globally.

Species/ Assemblage	No. of cases	Comment	Country	Reference
<i>Cryptosporidium</i>				
<i>C. felis</i>	2	HIV+ patients	Algeria	Semmani et al. (2023)
	1	21-year-old female with diarrhoea	Australia	Ebner et al. (2015)
	5	HIV+ patients	Brazil	Lucca et al. (2009)
	4	School children	Brazil	Pacheco et al. (2022)
	2	Sporadic cases	Canada	Guy et al. (2021)
	2	Hospitalised children (median 36 months-old)	China	Feng et al. (2012)
	8	1 HIV+, 4 with diarrhoea, 2 with non-gastrointestinal illnesses	China	Liu et al. (2020)
	3	Kindergarten children aged 2–6 years	China	Wang et al. (2022b)
	1	Average age 5 years	Colombia	Higuera et al. (2020)
	2	1 child and 1 HIV+ patient	Colombia	Uran-Velasquez et al. (2022)
	1	<i>C. felis</i> identified in owner's cat	Colombia	Potes-Morales and Crespo-Ortiz (2023)
	5	HIV+/AIDS patients	Ethiopia	Adamu et al. (2014)
	6	HIV+ patients	France	Guyot et al. (2001)
	15	10 HIV+ patients, 3 transplant patients	France	ANOFEEL Cryptosporidium National Network (2010)
	10	Immunodeficient individuals	France	Costa et al. (2018)
	6	40% immunodeficient patients	France	Costa et al. (2020)
	2	AIDS patients	Hatai	RaccourtBrasseur et al. (2006)
	1	HIV+ patients	India	Muthusamy et al. (2006)
	3	Patients with diarrhoea; many had underlying clinical conditions	India	Khalil et al. (2017)
	1	HIV+ patients	Jamaica	Gatei et al. (2008)
	2	Children aged < 5 years	Kenya	Gatei et al. (2006)
	1	HIV+/AIDS patients	Malaysia	Lim et al. (2011)
	4	Children aged 3–14 years	Mozambique	Muadica et al. (2021)
	1	HIV+/AIDS patients	Nigeria	Ukwah et al. (2017)
	1	All children; not associated with diarrhoea	Peru	Xiao et al. (2001)
	24	3.3% of typed positives from HIV+/AIDS patients	Peru	Cama et al. (2003)
	9	HIV+ patients	Peru	Cama et al. (2006)
	6	HIV+ patients	Peru	Cama et al. (2007)
	4	Children aged < 3 years	Portugal	Cama et al. (2008)
	1	HIV+ patients	Portugal	Alves et al. (2001)
	4	HIV+ patients	Portugal	Matos et al. (2004)
	1	HIV+ patient	Spain	Cieloszyk et al. (2012)
	1	–	Spain	Abal-Fabeira et al. (2014)
	1	37-year-old immunocompetent female with a range of underlying health issues and used inhaled steroids, pet cat also had <i>C. felis</i>	Sweden	Beser et al. (2015)
	4	Sporadic cases - one had travelled to Asia	Sweden	Lebbad et al. (2021)
	3	HIV+ patients	Switzerland	Morgan et al. (2000a)
	3	HIV+ patients	Thailand	Gatei et al. (2002)
	1	HIV+ patients	Thailand	Tiangtip and Jongwutiwas (2002)
	1	HIV+/AIDS patients	Thailand	Srisuphanunt et al. (2011)
	7	HIV+ patients	Thailand	Sannella et al. (2019)
	1	Primary school children	Thailand	Sutthikornchai et al. (2021)
	4	One HIV+ patient, one had a severe underlying condition, one aged 1 year	UK	Pedraza-Díaz et al. (2001)
	6	2 were known to be immunocompromised	UK	Leoni et al. (2006)
	4	0.01% (4/8000) of typed cases	UK	Chalmers et al. (2009b)
	38	0.3% (38/14,469) of typed cases, 6/38 were immunocompromised	UK	Elwin et al. (2012b)
	1	HIV+ patients	USA	Pieniazek et al. (1999)
	3	Rural Nebraska; age range 7 months to 79 years	USA	Loeck et al. (2020)
	10	New York City residents. 1.8% (10/547) of typed samples	USA	Alderisio et al. (2023)
	1	Patients with diarrhoea	Zambia	Mulunda et al. (2020)
	2	One patient infected while on holiday in India and second patient had "contact with a kitten with diarrhoea 3 weeks before disease onset"	Sweden	Insulander et al. (2013)
<i>C. canis</i>	1	School children (10% prevalence)	Angola	Dacal et al. (2018)
	1	HIV+ patients	Brazil	Lucca et al. (2009)
	5	Symptomatic children attending hospital	Cambodia	Moore et al. (2016)
	1	Hospitalised children (median 36 months-old)	China	Feng et al. (2012)
	2	Both with non-gastrointestinal illnesses	China	Liu et al. (2020)
	18	40% (12/30) of dog owners and 24% of (6/25) dog shelter workers	Egypt	Khalifa et al. (2023)
	2	HIV+/AIDS patients	Ethiopia	Adamu et al. (2014)
	4	3 HIV+ patients	France	ANOFEEL Cryptosporidium National Network (2010)
	3	40% immunodeficient patients	France	Costa et al. (2020)
	1	HIV+ patients	Jamaica	Gatei et al. (2008)
	1	Critically ill child in hospital with additional co-infections	Jordan	Hijjawi et al. (2010)
	3	Children aged < 5 years	Kenya	Gatei et al. (2006)
	2	Children (24- and 14 months old)	New Zealand	Learnmonth et al. (2004)
	1	Children aged 6 months to 6 years	Nigeria	Molloy et al. (2010)
	2	All children; not associated with diarrhoea	Peru	Xiao et al. (2001)
	36	4% of typed samples from HIV+/AIDS patients	Peru	Cama et al. (2003)
	12	HIV+ patients	Peru	Cama et al. (2006)

(continued on next page)

Table 3 (continued)

Species/ Assemblage	No. of cases	Comment	Country	Reference
	6	HIV+ patients	Peru	Cama et al. (2007)
	2	32-month-old female, 6.5-year-old brother (and household dog)	Peru	Xiao et al. (2007)
	4	Children aged < 3 years	Peru	Cama et al. (2008)
	2	HIV+ patients	Thailand	Gatei et al. (2002)
	1	HIV+/AIDS patients	Thailand	Srisuphanunt et al. (2011)
	12	HIV+ patients	Thailand	Sannella et al. (2019)
	1	Child aged 2 years	UK	Pedraza-Díaz et al. (2001)
	1	Child aged 1 year; had returned from Africa	UK	Leoni et al. (2006)
	1	0.01% (1/8000) of typed cases	UK	Chalmers et al. (2009a)
	1	0.007% (1/14,469) of typed cases	UK	Elwin et al. (2012b)
	1	HIV+ patients	USA	Pieniazek et al. (1999)
	1	Rural Nebraska; age range 7 months to 79 years	USA	Loeck et al. (2020)
	3	New York City residents. 0.5% (3/547) of typed samples	USA	Alderisio et al. (2023)
	1	HIV+ patients	Venezuela	Certad et al. (2006)
	2	1 human diarrheal and 1 non-diarrheal sample	Vietnam	Iwashita et al. (2021)
Giardia				
Assemblage C	5	Children attending a day-care centre	Brazil	Durigan et al. (2014)
	6	Immunocompromised patients	Brazil	Jeske et al. (2022)
	16	Typing only conducted at 1 locus	China	Liu et al. (2014)
	3	HIV+/AIDS patients	China	Jiang et al. (2023)
	1	Adult immunocompromised male with cancer and diarrhoea	Egypt	Soliman et al. (2011)
	1	44-year-old female with chronic diarrhoea; was not a pet owner	Slovakia	Štrkolcová et al. (2015)
Assemblage D	2	Travel returnees from Southeastern Asia	Germany	Broglia et al. (2013)
	1	Rural village, lack of water and sanitation. Only typed at 1 locus	Argentina	Candela et al. (2021)
	1	Immunocompromised patients	Brazil	Jeske et al. (2022)
Assemblage F	1	10-month-old child	Brazil	Silva et al. (2022)
	3	Children. Mixed A and F	Ethiopia	Gelanew et al. (2007)
	3	Children living with poor sanitation	Slovakia	Pipiková et al. (2022)
	1	Primary school children	Thailand	Sutthikornchai et al. (2021)

C. felis, *C. canis*, and assemblages C, D and F are considered to be of limited zoonotic potential, this risk is not negligible as there have been numerous reports of *C. felis* and *C. canis* in humans, as well as a few reports of assemblages C, D and F in humans (Table 3). The prevalence of *C. felis* and *C. canis* in humans is usually low (1–3% of cryptosporidiosis cases); however, a few studies have reported higher prevalences; 5–40% for *C. canis* and 7.1–37.5% for *C. felis* cryptosporidiosis cases, mostly in children or HIV-positive individuals (Moore et al., 2016; Dacal et al., 2018; Sannella et al., 2019; Uran-Velasquez et al., 2022; Wang et al., 2022b; Khalifa et al., 2023).

Cryptosporidium canis and *C. felis* are however, largely host-specific, as the prevalence of *C. felis* and *C. canis* in humans is usually low, despite the close contact between humans and companion animals. For example, in the USA, ~14% of households own a cat and ~22.2% of households own a dog, equating to 111.6 million households (APPA, 2022). Across Europe, 26% of households own one or more cats and 25% own one or more dogs, equating to ~127.2 million cats and 104.4 million dogs (FEDIAF, 2023). In Australia, 48% of Australian households have dogs and 33% of households have cats across both urban and rural areas (Animal Medicines Australia). The majority of reports of *C. felis*, *C. canis* and assemblages C, D and F in humans have been in immunologically immature children or immunocompromised adults and predominantly in low-income countries (Table 3). Therefore, the zoonotic risk from *C. felis*, *C. canis* and assemblages C, D and F in immunocompetent humans is probably low.

There have been several reports suggesting possible zoonotic transmission between owners and pets using genotyping analysis of human and animal samples from the same household or workplace (Xiao et al., 2007; Beser et al., 2015). For example, *C. felis* was characterised in a 37-year-old immunocompetent woman and her cat in Sweden (Beser et al., 2015) and *C. canis* was identified in a 2.5 year-old child, her 6.5 year-old brother and their dog in Peru (Xiao et al., 2007). In a recent study in Egypt, *C. canis* was identified in 12 domestic dog owners and 6 dog shelter workers (Khalifa et al., 2023).

The recent development of gp60 subtyping tools for *C. felis* and *C. canis* has facilitated studies of zoonotic transmission of these two pathogens (Rojas-Lopez et al., 2020; Jiang et al., 2021). Results of these

studies have identified both zoonotic and human-adapted subtypes in *C. felis*. Therefore, subtypes XIXc, XIXd and XIXe have been reported only in humans, while the remaining two subtypes (XIXa and XIXb) have been reported in both humans and cats (Jiang et al., 2020; Rojas-Lopez et al., 2020; Guy et al., 2021; Li et al., 2021b; Joachim et al., 2023). In total, nine *C. canis* subtype families (XXa to XXi) have been identified (Jiang et al., 2021; Murnik et al., 2022; Wang et al., 2022a; Elmahallawy et al., 2023; Mateo et al., 2023). The limited typing to date in *C. canis* has supported potential zoonotic transmission (Jiang et al., 2021), including re-analysis of samples from a suspected case of zoonotic transmission of *C. canis* in a household (Xiao et al., 2007), with *C. canis* subtype XXa4 identified from the 2.5 year-old female index case, her 6.5-year-old brother and their dog (Jiang et al., 2021). Similarly, subtyping has confirmed zoonotic transmission of *C. felis* between two human cases and their cats in Sweden (Rojas-Lopez et al., 2020).

Cats and particularly dogs can also be infected with *C. parvum*, which is a more significant public health risk. However, in most studies where *C. parvum* was detected, with a few exceptions, the occurrence rate was low and it is unknown if dogs and cats are mechanically carrying *C. parvum* due to coprophagy of human or cattle faeces or actually infected. It is possible that cats and particularly dogs from rural locations close to ruminants, may have a higher prevalence of *C. parvum*. Zoonotic transmission of *C. parvum* from cats and dogs has been suggested (Neira et al., 2010; Gharieb et al., 2018). For example, *C. parvum* was detected in children and their pet dogs in Egypt (Gharieb et al., 2018) and in a pregnant immunocompetent woman and her cat (Neira et al., 2010). The direction of transmission, i.e. spillover from pets to humans or spillback from humans to pets, in these studies is unknown. In contrast, a study in Spain reported no evidence supporting zoonotic transmission between owners and their pet cats and dogs (de Lucio et al., 2017). Similarly, in households in Peru, only *Giardia* assemblages A and B were identified in humans, while assemblages C and D were identified in their dogs, with the exception of one dog with a mixed assemblage B/D infection (Cooper et al., 2010). There have only been two reports of *C. hominis* in dogs and in both cases, PCR analysis of a second locus was unsuccessful, indicating very low oocyst numbers (Ng et al., 2011; Gil et al., 2017).

As with *Cryptosporidium*, several studies have suggested possible

transmission of *Giardia* from pets to owners (Traub et al., 2004; Palmer et al., 2008b; Marangi et al., 2010; Quadros et al., 2016; Rehbein et al., 2019; Kuthyar et al., 2021; Harvey et al., 2023; Khalifa et al., 2023), particularly in socially-deprived areas. For example, in a low socio-economic community in southern Italy, sub-assemblage AI was identified in nine of 14 dogs and six of 14 children (Marangi et al., 2010). In Argentina, assemblage A was detected in 12 humans and five pet dogs (Kuthyar et al., 2021). In one study in Brazil, assemblage E was the only assemblage detected in dogs, and assemblages A, B and E were recovered from children from the same rural community (Harvey et al., 2023). Another study of *Giardia*-positive samples from humans, dogs and cats in Brazil, identified assemblage F in one human and assemblages C, D and F in dogs and F in one cat (Silva et al., 2022). These studies are suggestive of zoonotic transmission but have not been confirmed by finding identical multilocus subtypes in both humans and their companion animals. Furthermore, it would also be necessary to demonstrate that the animals acquired the infection prior to human infection (Cai et al., 2021). Other studies have found no evidence of transmission between companion animals and their owners (Hopkins et al., 1997; Inpankaew et al., 2014; Wu et al., 2022). For example, one study examined *Giardia* assemblages in dogs and humans in rural Cambodia and found the occurrence of different assemblages between the two host species, leading to the conclusion that the risk of zoonotic transmission was low (Inpankaew et al., 2014). Another recent study in Egypt detected assemblages A and D in domestic and shelter dogs but assemblage A only in their owners and dog shelter workers (Khalifa et al., 2023).

Many of the typing studies conducted to date on *Giardia* in cats and dogs are problematic due to (i) high levels of mixed infections with multiple *Giardia* assemblages, (ii) the use of only one locus or lack of agreement between multiple loci, and (iii) the high levels of allelic sequence heterozygosity (ASH) in assemblages C and D, which make assemblage assignment unreliable (Morrison et al., 2007; Franzén et al., 2009; Kooyman et al., 2019; Veldhuis et al., 2022; Klotz et al., 2023). Therefore, it is difficult to draw firm conclusions on the zoonotic potential of *Giardia* infections in companion animals. It has been suggested that in dogs, two main transmission cycles exist, with assemblage C and D predominating, particularly in kennel/shelter dogs, and assemblage A and occasionally assemblage B transmission between dogs and humans in households (Cai et al., 2021). In agreement with this, a case-control study in the UK, reported a significant correlation between dog ownership and assemblage A infection in humans (Minetti et al., 2015). Others argue that overall, humans and pets are preferentially infected with sub-assemblages AII and AI, respectively (Feng and Xiao, 2011).

5. Conclusions

Our understanding of the zoonotic transmission of *C. felis* and *C. canis* from cats and dogs to humans will be improved by the recent development of molecular typing tools, with preliminary evidence supporting some level of zoonotic transmission (Rojas-Lopez et al., 2020; Jiang et al., 2020, 2021; Guy et al., 2021; Li et al., 2021a). Most molecular-based household transmission studies to date, however, have focussed on middle to high-income countries, where the prevalence of *C. felis* and *C. canis* in humans is low. Relatively few studies have been conducted in dogs and cats in low-income countries, where the prevalence of *C. felis* and *C. canis* is much higher in humans. Overall, the risk of zoonotic transmission from cats and dogs to humans is uncertain but likely to be low due to the relatively low zoonotic risk from *C. canis*, *C. felis* and assemblages C, D and F, the overall low prevalence of *C. parvum* (and *C. hominis*) in cats and dogs, and low oocyst shedding (Lucio-Forster et al., 2010; de Lucio et al., 2016; Rehbein et al., 2019; Li et al., 2021a; Cai et al., 2021). This would be even more the case for *G. duodenalis* in cats and dogs due to differences in host preferences of this parasite at the assemblage and sub-assemblage levels. Future epidemiological studies should concentrate on applying the most recently developed *Cryptosporidium* and *Giardia* multi-locus sub-typing

tools, as well as whole genome sequencing, to longitudinal *Giardia*-positive samples from cats and dogs and their owners, before a clearer understanding of the extent of spillover and/or spillback of *Cryptosporidium* and *Giardia* between pets and their owners can be determined.

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CRedit authorship contribution statement

Amanda D. Barbosa: Writing – original draft, Writing – review & editing. **Siobhon Egan:** Writing – review & editing. **Yaoyu Feng:** Writing – review & editing, Funding acquisition. **Lihua Xiao:** Writing – review & editing, Funding acquisition. **Una Ryan:** Writing – original draft, Writing – review & editing, Project administration, Funding acquisition, All authors read and approved the final manuscript.

Declaration of competing interests

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper. Given their role as Guest Editor, Amanda D. Barbosa had no involvement in the peer review of this article and has no access to information regarding its peer review. Full responsibility for the editorial process for this article was delegated to Dr Frank Katzer (Co-Editor) and Professor Aneta Kostadinova (Editor-in-Chief).

Data availability

The data supporting the conclusions of this article are included within the article.

References

- Abal-Fabeiro, J.L., Maside, X., Llovo, J., Bello, X., Torres, M., Treviño, M., et al., 2014. High-throughput genotyping assay for the large-scale genetic characterization of *Cryptosporidium* parasites from human and bovine samples. *Parasitology* 141, 491–500. <https://doi.org/10.1017/S0031182013001807>.
- Abdelaziz, A.R., Sorour, S.S.G., 2021. Prevalence and molecular characterization of *Giardia duodenalis* assemblage D of dogs in Egypt, and its zoonotic implication. *Microbes Infect. Chemother.* 1, e1268. <https://doi.org/10.54034/mic.e1268>.
- Abe, N., Sawano, Y., Yamada, K., Kimata, I., Iseki, M., 2002. *Cryptosporidium* infection in dogs in Osaka, Japan. *Vet. Parasitol.* 108, 185–193.
- Abe, N., Kimata, I., Iseki, M., 2003. Identification of genotypes of *Giardia intestinalis* isolates from dogs in Japan by direct sequencing of the PCR amplified glutamate dehydrogenase gene. *J. Vet. Med. Sci.* 65, 29–33.
- Adamu, H., Petros, B., Zhang, G., Kassa, H., Amer, S., Ye, J., et al., 2014. Distribution and clinical manifestations of *Cryptosporidium* species and subtypes in HIV/AIDS patients in Ethiopia. *PLoS Negl. Trop. Dis.* 8, e2831.
- Adell-Aledon, M., Köster, P.C., de Lucio, A., Puente, P., Hernández-de-Mingo, M., Sánchez-Thevenet, P., Dea-Ayuela, M.A., Carmena, D., 2018. Occurrence and molecular epidemiology of *Giardia duodenalis* infection in dog populations in eastern Spain. *BMC Vet. Res.* 14, 26.
- Adriana, G., Zsuzsa, K., Mirabela Oana, D., Mircea, G.C., Viorica, M., 2016. *Giardia duodenalis* genotypes in domestic and wild animals from Romania identified by PCR-RFLP targeting the *gdh* gene. *Vet. Parasitol.* 217, 71–75.
- Agresti, A., Berrilli, F., Maestrini, M., Guadano Procesi, I., Loretti, E., Vonci, N., Perrucci, S., 2021. Prevalence, risk factors and genotypes of *Giardia duodenalis* in sheltered dogs in Tuscany (Central Italy). *Pathogens* 11, 12.
- Alderisio, K.A., Mergen, K., Moessner, H., Madison-Antenucci, S., 2023. Identification and evaluation of *Cryptosporidium* species from New York City cases of cryptosporidiosis (2015 to 2018): A watershed perspective. *Microbiol. Spectr.* 11, e039212.
- Alves, M., Matos, O., Pereira, Da Fonseca, I., Delgado, E., Lourenco, A.M., Antunes, F., 2001. Multilocus genotyping of *Cryptosporidium* isolates from human HIV-infected and animal hosts. *J. Eukaryot. Microbiol.* 48, 17S–18S.

- Alves, M.E.M., Martins, F.D.C., Bräunig, P., Pivoto, F.L., Sangioni, L.A., Vogel, F.S.F., 2018. Molecular detection of *Cryptosporidium* spp. and the occurrence of intestinal parasites in fecal samples of naturally infected dogs and cats. *Parasitol. Res.* 117, 3033–3038.
- Animal Medicines Australia, 2022. Pets in Australia: A national survey of pets and people. <https://animalmedicinesaustralia.org.au/report/pets-in-australia-a-national-survey-of-pets-and-people-2/>. (Accessed 28 August 2023).
- ANOFEL Cryptosporidium National Network, 2010. Laboratory-based surveillance for *Cryptosporidium* in France, 2006–2009. *Euro Surveill.* 15, 19642.
- APPA, 2022. 2021–2022 APPA National Pet Owners Survey. American Pet Products Manufacturers Association (APPA). http://www.americanpetproducts.org/press_industrytrends.asp. (Accessed 27 September 2022).
- Asahi, H., Koyama, T., Arai, H., Funakoshi, Y., Yamaura, H., Shirasaka, R., Okutomi, K., 1991. Biological nature of *Cryptosporidium* sp. isolated from a cat. *Parasitol. Res.* 77, 237–240. <https://doi.org/10.1007/BF00930864>.
- Aydin, Y., Ozkul, I.A., 1996. Infectivity of *Cryptosporidium muris* directly isolated from the murine stomach for various laboratory animals. *Vet. Parasitol.* 66, 257–262. [https://doi.org/10.1016/s0304-4017\(96\)01007-2](https://doi.org/10.1016/s0304-4017(96)01007-2).
- Ayinmode, A.B., Oliveira, B.C.M., Obebe, O.C., Dada-Adgebola, H.O., Ayede, A.I., Widmer, G., 2018a. Genotypic characterization of *Cryptosporidium* species in humans and peri-domestic animals in Ekiti and Oyo States, Nigeria. *J. Parasitol.* 104, 639–644.
- Ayinmode, A.B., Obebe, O.O., Falohun, O.O., 2018b. Molecular detection of *Cryptosporidium* species in street-sampled dog faeces in Ibadan, Nigeria. *Vet. Parasitol. Reg. Stud. Reports.* 14, 54–58.
- Bajer, A., Bednarska, M., Rodo, A., 2011. Risk factors and control of intestinal parasite infections in sled dogs in Poland. *Vet. Parasitol.* 175, 343–350.
- Ballweber, L.R., Panuska, C., Huston, C.L., Vasiliopoulos, R., Pharr, G.T., Mackin, A., 2009. Prevalence of and risk factors associated with shedding of *Cryptosporidium felis* in domestic cats of Mississippi and Alabama. *Vet. Parasitol.* 160, 306–310.
- Barutzki, D., Thompson, R.C., Wielinga, C., Parka, U., Schaper, R., 2007. Observations on *Giardia* infection in dogs from veterinary clinics in Germany. *Parasitol. Res.* 101 (Suppl. 1), 153–156.
- Beck, R., Sprong, H., Pozio, E., Cacciò, S.M., 2012. Genotyping *Giardia duodenalis* isolates from dogs: Lessons from a multilocus sequence typing study. *Vector Borne Zoonotic Dis.* 12, 206–213.
- Berrilli, F., Di Cave, D., De Liberato, C., Franco, A., Scaramozzino, P., Orechia, P., 2004. Genotype characterisation of *Giardia duodenalis* isolates from domestic and farm animals by SSU-rRNA gene sequencing. *Vet. Parasitol.* 122, 193–199.
- Berrilli, F., D'Alfonso, R., Giangaspero, A., Marangi, M., Brandonisio, O., Kaboré, Y., et al., 2012. *Giardia duodenalis* genotypes and *Cryptosporidium* species in humans and domestic animals in Côte d'Ivoire: Occurrence and evidence for environmental contamination. *Trans. R. Soc. Trop. Med. Hyg.* 106, 191–195.
- Beser, J., Toresson, L., Eitrem, R., Troell, K., Winiecka-Krusnell, J., Lebbad, M., 2015. Possible zoonotic transmission of *Cryptosporidium felis* in a household. *Infect. Ecol. Epidemiol.* 5, 28463.
- Bouzid, M., Halai, K., Jeffreys, D., Hunter, P.R., 2015. The prevalence of *Giardia* infection in dogs and cats, a systematic review and meta-analysis of prevalence studies from stool samples. *Vet. Parasitol.* 207, 181–202.
- Brogli, A., Weitzel, T., Harms, G., Cacciò, S.M., Nöckler, K., 2013. Molecular typing of *Giardia duodenalis* isolates from German travellers. *Parasitol. Res.* 112, 3449–3456. <https://doi.org/10.1007/s00436-013-3524-y>.
- Bugg, R.J., Robertson, I.D., Elliot, A.D., Thompson, R.C., 1999. Gastrointestinal parasites of urban dogs in Perth, Western Australia. *Vet. J.* 157, 295–301.
- Cacciò, S.M., Ryan, U., 2008. Molecular epidemiology of giardiasis. *Mol. Biochem. Parasitol.* 160, 75–80.
- Cacciò, S.M., Beck, R., Lalle, M., Marincola, A., Pozio, E., 2008. Multilocus genotyping of *Giardia duodenalis* reveals striking differences between assemblages A and B. *Int. J. Parasitol.* 38, 1523–1531.
- Cai, W., Ryan, U., Xiao, L., Feng, Y., 2021. Zoonotic giardiasis: An update. *Parasitol. Res.* 120, 4199–4218.
- Cama, V.A., Bern, C., Sulaiman, I.M., Gilman, R.H., Ticona, E., Vivar, A., et al., 2003. *Cryptosporidium* species and genotypes in HIV-positive patients in Lima, Peru. *J. Eukaryot. Microbiol.* 50 (Suppl. 1), 531–533.
- Cama, V., Gilman, R.H., Vivar, A., Ticona, E., Ortega, Y., Bern, C., Xiao, L., 2006. Mixed *Cryptosporidium* infections and HIV. *Emerg. Infect. Dis.* 12, 1025–1028.
- Cama, V.A., Ross, J.M., Crawford, S., Kawai, V., Chavez-Valdez, R., Vargas, D., et al., 2007. Differences in clinical manifestations among *Cryptosporidium* species and subtypes in HIV-infected persons. *J. Infect. Dis.* 196, 684–691.
- Cama, V.A., Bern, C., Roberts, J., Cabrera, L., Sterling, C.R., Ortega, Y., et al., 2008. *Cryptosporidium* species and subtypes and clinical manifestations in children, Peru. *Emerg. Infect. Dis.* 14, 1567–1574.
- Candela, E., Goizueta, C., Periago, M.V., Muñoz-Antoli, C., 2021. Prevalence of intestinal parasites and molecular characterization of *Giardia intestinalis*, *Blastocystis* spp. and *Entamoeba histolytica* in the village of Fortín Mbororé (Puerto Iguazú, Misiones, Argentina). *Parasites Vectors* 14, 510. <https://doi.org/10.1186/s13071-021-04968-z>.
- Cao, Y., Fang, C., Deng, J., Yu, F., Ma, D., Chuai, L., Wang, T., Qi, M., Li, J., 2022. Molecular characterization of *Cryptosporidium* spp. and *Giardia duodenalis* in pet dogs in Xinjiang, China. *Parasitol. Res.* 121, 1429–1435.
- CDC, 1999. Centers for Disease Control and Prevention: False-positive laboratory tests for *Cryptosporidium* involving an enzyme-linked immunosorbent assay - United States. *MMWR Morb. Mortal. Wkly. Rep.* 48, 4–8.
- Certad, G., Ngouanesavanh, T., Hernan, A., Rojas, E., Contreras, R., Pocaterra, L., et al., 2006. First molecular data on cryptosporidiosis in Venezuela. *J. Eukaryot. Microbiol.* 53, S30–S32.
- Chalmers, R.M., Robinson, G., Elwin, K., Hadfield, S.J., Xiao, L., Ryan, U., et al., 2009a. *Cryptosporidium* sp. rabbit genotype, a newly identified human pathogen. *Emerg. Infect. Dis.* 15, 829–830.
- Chalmers, R.M., Elwin, K., Thomas, A.L., Guy, E.C., Mason, B., 2009b. Long-term *Cryptosporidium* typing reveals the aetiology and species-specific epidemiology of human cryptosporidiosis in England and Wales, 2000 to 2003. *Euro Surveill.* 14, 19086.
- Charmaraman, L., Cobas, S., Weed, J., Gu, Q., Kiel, E., Chin, H., et al., 2022. From regulating emotions to less lonely screen time: Parents' qualitative perspectives of the benefits and challenges of adolescent pet companionship. *Behav. Sci.* 12, 143.
- Cieloszyk, J., Goñi, P., García, A., Remacha, M.A., Sánchez, E., Clavel, A., 2012. Two cases of zoonotic cryptosporidiosis in Spain by the unusual species *Cryptosporidium ubiquitum* and *Cryptosporidium felis*. *Enferm. Infect. Microbiol. Clín.* 30, 549–551.
- Cirak, V.Y., Bauer, C., 2004. Comparison of conventional coproscopic methods and commercial coproantigen ELISA kits for the detection of *Giardia* and *Cryptosporidium* infections in dogs and cats. *Berl. Münchener Tierärztliche Wochenschr.* 117, 410–413.
- Ciuca, L., Pepe, P., Bosco, A., Caccio, S.M., Maurelli, M.P., Sannella, A.R., et al., 2021. Effectiveness of fenbendazole and metronidazole against *Giardia* infection in dogs monitored for 50-days in home-conditions. *Front. Vet. Sci.* 8, 626424.
- Claerebout, E., Casaert, S., Dalemans, A.C., De Wilde, N., Levecke, B., Vercruyse, J., et al., 2009. *Giardia* and other intestinal parasites in different dog populations in Northern Belgium. *Vet. Parasitol.* 161, 41–46.
- Colli, C.M., Bezagio, R.C., Nishi, L., Bignotto, T.S., Ferreira, É.C., Falavigna-Guilherme, A.L., et al., 2015. Identical assemblage of *Giardia duodenalis* in humans, animals and vegetables in an urban area in southern Brazil indicates a relationship among them. *PLoS One* 10, e0118065.
- Cooper, M.A., Sterling, C.R., Gilman, R.H., Cama, V., Ortega, Y., Adam, R.D., 2010. Molecular analysis of household transmission of *Giardia lamblia* in a region of high endemicity in Peru. *J. Infect. Dis.* 202, 1713–1721.
- Costa, D., Razakandraine, R., Sautour, M., Valot, S., Basmaciyan, L., Gargala, G., et al., 2018. Human cryptosporidiosis in immunodeficient patients in France (2015–2017). *Exp. Parasitol.* 192, 108–112.
- Costa, D., Razakandraine, R., Valot, S., Vannier, M., Sautour, M., Basmaciyan, L., et al., 2020. Epidemiology of cryptosporidiosis in France from 2017 to 2019. *Microorganisms* 8, 1358.
- Covacinc, A., Aucoin, D.P., Elliot, A., Thompson, R.C.A., 2011. Genotypic characterisation of *Giardia* from domestic dogs in the USA. *Vet. Parasitol.* 177, 28–32.
- Cox, P., Griffith, M., Angles, M., Deere, D., Ferguson, C., 2005. Concentrations of pathogens and indicators in animal feces in the Sydney watershed. *Appl. Environ. Microbiol.* 71, 5929–5934.
- Dacal, E., Saugar, J.M., de Lucio, A., Hernandez-de-Mingo, M., Robinson, E., Koster, P.C., et al., 2018. Prevalence and molecular characterization of *Strongyloides stercoralis*, *Giardia duodenalis*, *Cryptosporidium* spp., and *Blastocystis* spp. isolates in school children in Cuban, Western Angola. *Parasites Vectors* 11, 67.
- Danciger, M., Lopez, M., 1975. Numbers of *Giardia* in the feces of infected children. *Am. J. Trop. Med. Hyg.* 24, 237–242.
- Davies, R.H., Lawes, J.R., Wales, A.D., 2019. Raw diets for dogs and cats: A review, with particular reference to microbiological hazards. *J. Small Anim. Pract.* 60, 329–339.
- de Lucio, A., Amor-Aramendia, A., Bailo, B., Saugar, J.M., Anegagrie, M., Arroyo, A., et al., 2016. Prevalence and genetic diversity of *Giardia duodenalis* and *Cryptosporidium* spp. among school children in a rural area of the Amhara region, north-west Ethiopia. *PLoS One* 11, e0159992. <https://doi.org/10.1371/journal.pone.0159992>.
- de Lucio, A., Bailo, B., Aguilera, M., Cardona, G.A., Fernández-Crespo, J.C., Carmena, D., 2017. No molecular epidemiological evidence supporting household transmission of zoonotic *Giardia duodenalis* and *Cryptosporidium* spp. from pet dogs and cats in the province of Álava, northern Spain. *Acta Trop.* 170, 48–56.
- de Oliveira, A.G.L., Sudré, A.P., Bergamo do Bomfim, T.C., Santos, H.L.C., 2021. Molecular characterization of *Cryptosporidium* spp. in dogs and cats in the city of Rio de Janeiro, Brazil, reveals potentially zoonotic species and genotype. In: *PLoS One*, 16, e0255087.
- Doing, K.M., Hamm, J.L., Jellison, J.A., Marquis, J.A., Kingsbury, C., 1999. False-positive results obtained with the Alexon ProSpecT *Cryptosporidium* enzyme immunoassay. *J. Clin. Microbiol.* 37, 1582–1583.
- Durigan, M., Abreu, A.G., Zucchi, M.I., Franco, R.M., de Souza, A.P., 2014. Genetic diversity of *Giardia duodenalis*: multilocus genotyping reveals zoonotic potential between clinical and environmental sources in a metropolitan region of Brazil. *PLoS One* 9, e115489.
- Ebner, J., Koehler, A.V., Robertson, G., Bradbury, R.S., Jex, A.R., Haydon, S.R., et al., 2015. Genetic analysis of *Giardia* and *Cryptosporidium* from people in Northern Australia using PCR-based tools. *Infect. Genet. Evol.* 36, 389–395.
- El-Madawy, R.S., Khalifa, N.O., Khater, H.F., 2010. Detection of cryptosporidial infection among Egyptian stray dogs by using *Cryptosporidium parvum* outer wall protein gene. *Bulg. J. Vet. Med.* 13, 104–110.
- Elgio-Garcia, L., Cortes-Campos, A., Jimenez-Cardoso, E., 2005. Genotype of *Giardia intestinalis* isolates from children and dogs and its relationship to host origin. *Parasitol. Res.* 97, 1–6.
- Elmahallawy, E.K., Gareh, A., Abu-Oakail, A., Köster, P.C., Dashti, A., Asseri, J., et al., 2023. Molecular characteristics and zoonotic potential of enteric protists in domestic dogs and cats in Egypt. *Front. Vet. Sci.* 10, 1229151.
- Elwin, K., Hadfield, S.J., Robinson, G., Chalmers, R.M., 2012. The epidemiology of sporadic human infections with unusual cryptosporidia detected during routine typing in England and Wales, 2000–2008. *Epidemiol. Infect.* 140, 673–683.

- Enemark, H.L., Starostka, T.P., Larsen, B., Takeuchi-Storm, N., Thamsborg, S.M., 2020. *Giardia* and *Cryptosporidium* infections in Danish cats: Risk factors and zoonotic potential. *Parasitol. Res.* 119, 2275–2286.
- ESCCAP, 2018. Control of intestinal protozoa in dogs and cats. In: ESCCAP Guideline 06, 2nd edition. European Specialist Counsel Companion Animal Parasites. <https://www.esccap.org/guidelines/g16>.
- Fantinatti, M., Caseca, A.C., Bello, A.R., Fernandes, O., Da-Cruz, A.M., 2018. The presence of *Giardia lamblia* assemblage A in dogs suggests an anthropozoonotic cycle of the parasite in Rio de Janeiro, Brazil. *Infect. Genet. Evol.* 65, 265–269.
- Fava, N.M., Soares, R.M., Scalvia, L.A., Cunha, M.J., Faria, E.S., Cury, M.C., 2016. Molecular typing of canine *Giardia duodenalis* isolates from Minas Gerais, Brazil. *Exp. Parasitol.* 161, 1–5. <https://doi.org/10.1016/j.exppara.2015.12.003>.
- Fayer, R., Santín, M., Trout, J.M., Dubey, J.P., 2006. Detection of *Cryptosporidium felis* and *Giardia duodenalis* Assemblage F in a cat colony. *Vet. Parasitol.* 140, 44–53.
- FEDIAF, 2023. European Pet Food Industry Federation: Annual Report 2023. https://ueuropeanpetfood.org/wp-content/uploads/2023/06/FEDIAF_Annual-Report_2023_Facts-Figures.pdf. (Accessed 27 September 2023).
- Feng, Y., Xiao, L., 2011. Zoonotic potential and molecular epidemiology of *Giardia* species and giardiasis. *Clin. Microbiol. Rev.* 24, 110–140.
- Feng, Y., Xiao, L., 2017. Molecular epidemiology of cryptosporidiosis in China. *Front. Microbiol.* 8, 1701.
- Feng, Y., Wang, L., Duan, L., Gomez-Puerta, L.A., Zhang, L., Zhao, X., et al., 2012. Extended outbreak of cryptosporidiosis in a pediatric hospital, China. *Emerg. Infect. Dis.* 18, 312–314.
- Ferreira, F.S., Pereira-Baltazar, P., Parreira, R., Padre, L., Vilhena, M., Távora Tavira, L., et al., 2011. Intestinal parasites in dogs and cats from the district of Évora, Portugal. *Vet. Parasitol.* 179, 242–245.
- Ferreira, A., Alho, A.M., Otero, D., Gomes, L., Nijssse, R., Overgaauw, P.A.M., et al., 2017. Urban dog parks as sources of canine parasites: contamination rates and pet owner behaviours in Lisbon, Portugal. *J. Environ. Public Health* 2017, 5984086.
- FitzGerald, L., Bennett, M., Ng, J., Nicholls, P., James, F., Elliott, A., et al., 2011. Morphological and molecular characterisation of a mixed *Cryptosporidium muris*/ *Cryptosporidium felis* infection in a cat. *Vet. Parasitol.* 17, 160–164.
- Franzén, O., Jerlström-Hultqvist, J., Castro, E., Sherwood, E., Ankarklev, J., Reiner, D.S., et al., 2009. Draft genome sequencing of *Giardia intestinalis* assemblage B isolate GS: Is human giardiasis caused by two different species? *PLoS Pathog.* 5, e1000560.
- French, S.K., Kotwa, J.D., Singh, B., Greer, T., Pearl, D.L., Elsemore, D.A., et al., 2023. Factors associated with *Giardia* infection in dogs in southern Ontario, Canada. *Vet. Parasitol. Reg. Stud. Reports* 41, 100870.
- Gatei, W., Suputtamongkol, Y., Waywa, D., Ashford, R.W., Bailey, J.W., Greensill, J., et al., 2002. Zoonotic species of *Cryptosporidium* are as prevalent as the anthropopotic in HIV-infected patients in Thailand. *Ann. Trop. Med. Parasitol.* 96, 797–802.
- Gatei, W., Kamwati, S.K., Mbae, C., Waruru, A., Hart, C.A., Wamae, C.N., et al., 2006. Cryptosporidiosis: Prevalence, genotype analysis, and symptoms associated with infections in children in Kenya. *Am. J. Trop. Med. Hyg.* 75, 78–82.
- Gatei, W., Barrett, D., Lindo, J.F., Eldemire-Shearer, D., Cama, V., Xiao, L., 2008. Unique *Cryptosporidium* population in HIV-infected persons, Jamaica. *Emerg. Infect. Dis.* 14, 841–843.
- Gelanew, T., Lalle, M., Hailu, A., Pozio, E., Caccio, S.M., 2007. Molecular characterization of human isolates of *Giardia duodenalis* from Ethiopia. *Acta Trop.* 102, 92–99.
- Gharieb, R.M.A., Merwad, A.M.A., Saleh, A.A., Abd El-Ghany, A.M., 2018. Molecular screening and genotyping of *Cryptosporidium* species in household dogs and in contact children in Egypt: Risk factor analysis and zoonotic importance. *Vector Borne Zoonotic Dis.* 18, 424–432.
- Giangaspero, A., Iorio, R., Paoletti, B., Traversa, D., Capelli, G., 2006. Molecular evidence for *Cryptosporidium* infection in dogs in Central Italy. *Parasitol. Res.* 99, 297–299.
- Gil, H., Cano, L., de Lucio, A., Bailo, B., de Mingo, M.H., Cardona, G.A., et al., 2017. Detection and molecular diversity of *Giardia duodenalis* and *Cryptosporidium* spp. in sheltered dogs and cats in northern Spain. *Infect. Genet. Evol.* 50, 62–69.
- Goodgame, R.W., Genta, R.M., White, A.C., Chappell, C.L., 1993. Intensity of infection in AIDS-associated cryptosporidiosis. *J. Infect. Dis.* 167, 704–709.
- Gu, Y.F., Wang, K., Liu, D.Y., Mei, N., Chen, C., Chen, T., et al., 2015. Molecular detection of *Giardia lamblia* and *Cryptosporidium* species in pet dogs. *Zhongguo Ji Sheng Chong Xue Yu Ji Sheng Chong Bing Za Zhi* 33, 362–367.
- Guadano Procesi, I., Carnio, A., Berrilli, F., Montalbano, Di Filippo, M., Scarito, A., et al., 2022. *Giardia duodenalis* in colony stray cats from Italy. *Zoonoses Public Health* 69, 46–54.
- Gultekin, M., Ural, K., Aysul, N., Ayan, A., Balikci, C., Akyildiz, G., 2017. Prevalence and molecular characterization of *Giardia duodenalis* in dogs in Aydin, Turkey. *Int. J. Environ. Health Res.* 27, 161–168.
- Guo, Y., Li, N., Ryan, U., Feng, Y., Xiao, L., 2021. Small ruminants and zoonotic cryptosporidiosis. *Parasitol. Res.* 120, 4189–4198.
- Guo, Y., Ryan, U., Feng, Y., Xiao, L., 2022. Emergence of zoonotic *Cryptosporidium parvum* in China. *Trends Parasitol.* 38, 335–343.
- Guy, R.A., Yanta, C.A., Muchaal, P.K., Rankin, M.A., Thivierge, K., Lau, R., et al., 2021. Molecular characterization of *Cryptosporidium* isolates from humans in Ontario, Canada. *Parasites Vectors* 14, 69.
- Guyot, K., Follet-Dumoulin, A., Lelievre, E., Sarfati, C., Rabodonirina, M., Nevez, G., et al., 2001. Molecular characterization of *Cryptosporidium* isolates obtained from humans in France. *J. Clin. Microbiol.* 39, 3472–3480.
- Hajdušek, O., Ditrich, O., Šlapeta, J., 2004. Molecular identification of *Cryptosporidium* spp. in animal and human hosts from the Czech Republic. *Vet. Parasitol.* 122, 183–192.
- Hamnes, I.S., Gjerde, B.K., Robertson, L.J., 2007. A longitudinal study on the occurrence of *Cryptosporidium* and *Giardia* in dogs during their first year of life. *Acta Vet. Scand.* 49, 22.
- Harvey, T.V., Carvalho, J.P.D.S., Aquino, M.C.C., Oliveira, B.C.M., Barros, L.D., Fehlberg, H.F., et al., 2023. Giardiasis in children and dogs, and the first report of assemblage E in dogs from northeastern Brazil. *Rev. Bras. Parasitol. Vet.* 32, e012222.
- Hascall, K.L., Kass, P.H., Saksen, J., Ahlmann, A., Scorza, A.V., Lappin, M.R., Marks, S.L., 2016. Prevalence of enteropathogens in dogs attending 3 regional dog parks in Northern California. *J. Vet. Intern. Med.* 30, 1838–1845. <https://doi.org/10.1111/jvim.14603>.
- Hernandez, P.C., Morales de la Pava, L., Chaparro-Olaya, J., Lopez-Osorio, S., Lopez-Arias, A., Chaparro-Gutierrez, J.J., 2021. Multilocus genotyping of *Giardia intestinalis* in pet dogs of Medellin, Colombia. *Vet. Parasitol. Reg. Stud. Reports* 23, 100520.
- Higuera, A., Villamizar, X., Herrera, G., Giraldo, J.C., Vasquez-A, L.R., Urbano, P., et al., 2020. Molecular detection and genotyping of intestinal protozoa from different biogeographical regions of Colombia. *PeerJ* 8, e8554.
- Hijjawi, N., Ng, J., Yang, R., Atoum, M.F., Ryan, U., 2010. Identification of rare and novel *Cryptosporidium gp60* subtypes in human isolates from Jordan. *Exp. Parasitol.* 125, 161–164.
- Himsworth, C.G., Skinner, S., Chaban, B., Jenkins, E., Wagner, B.A., Harms, N.J., et al., 2010. Multiple zoonotic pathogens identified in canine feces collected from a remote Canadian indigenous community. *Am. J. Trop. Med. Hyg.* 83, 338–341.
- Hinney, B., Ederer, C., Stengl, C., Wilding, K., Štrkolcová, G., Harl, J., et al., 2015. Enteric protozoa of cats and their zoonotic potential - a field study from Austria. *Parasitol. Res.* 114, 2003–2006.
- Holzhausen, I., Lendner, M., Göhring, F., Steinböck, I., Daugschies, A., 2019. Distribution of *Cryptosporidium parvum* gp60 subtypes in calf herds of Saxony, Germany. *Parasitol. Res.* 118, 1549–1558.
- Homayouni, M.M., Razavi, S.M., Shaddel, M., Asadpour, M., 2019. Prevalence and molecular characterization of *Cryptosporidium* spp. and *Giardia intestinalis* in household dogs and cats from Shiraz, southwestern Iran. *Vet. Ital.* 55, 311–318.
- Hoopes, J., Hill, J.E., Polley, L., Fernando, C., Wagner, B., Schurer, J., et al., 2015. Enteric parasites of free-roaming, owned, and rural cats in prairie regions of Canada. *Can. Vet. J.* 56, 495–501.
- Hopkins, R.M., Meloni, B.P., Groth, D.M., Wetherall, J.D., Reynoldson, J.A., Thompson, R.C.A., 1997. Ribosomal RNA sequencing reveals differences between the genotypes of *Giardia* isolates recovered from humans and dogs living in the same locality. *J. Parasitol.* 83, 44–51.
- Inpankaew, T., Traub, R., Thompson, R.C., Sukrana, Y., 2007. Canine parasitic zoonoses in Bangkok temples. *Southeast Asian J. Trop. Med. Publ. Health* 38, 247–255.
- Inpankaew, T., Schär, F., Odermann, P., Dalsgaard, A., Chimnoi, W., Khieu, V., et al., 2014. Low risk for transmission of zoonotic *Giardia duodenalis* from dogs to humans in rural Cambodia. *Parasites Vectors* 7, 412.
- Insulander, M., Silverlas, C., Lebbad, M., Karlsson, L., Mattsson, J.G., Svennungsson, B., 2013. Molecular epidemiology and clinical manifestations of human cryptosporidiosis in Sweden. *Epidemiol. Infect.* 141, 1009–1020.
- Iseki, M., Maekawa, T., Moriya, K., Uni, S., Takada, S., 1989. Infectivity of *Cryptosporidium muris* (strain RN 66) in various laboratory animals. *Parasitol. Res.* 75, 218–222.
- Itagaki, T., Kinoshita, S., Aoki, M., Itoh, N., Saeki, H., Sato, N., et al., 2005. Genotyping of *Giardia intestinalis* from domestic and wild animals in Japan using glutamate dehydrogenase gene sequencing. *Vet. Parasitol.* 133, 283–287.
- Ito, Y., Itoh, N., Iijima, Y., Kimura, Y., 2017. Molecular prevalence of *Cryptosporidium* species among household cats and pet shop kittens in Japan. *JFMS Open Rep.* 3, 2055116917730719.
- Itoh, N., Itagaki, T., Kawabata, T., Konaka, T., Muraoaka, N., Saeki, H., et al., 2011. Prevalence of intestinal parasites and genotyping of *Giardia intestinalis* in pet shop puppies in east Japan. *Vet. Parasitol.* 176, 74–78.
- Itoh, N., Tanaka, H., Iijima, Y., Kameshima, S., Kimura, Y., 2019. Molecular prevalence of *Cryptosporidium* spp. in breeding kennel dogs. *Kor. J. Parasitol.* 57, 197–200.
- Iwashita, H., Takemura, T., Tokizawa, A., Sugamoto, T., Thiem, V.D., Nguyen, T.H., et al., 2021. Molecular epidemiology of *Cryptosporidium* spp. in an agricultural area of northern Vietnam: A community survey. *Parasitol. Int.* 83, 102341.
- Jeske, S.T., Macedo, M.R.P., Bianchi, T., Leon, I.F., Pinheiro, N.B., Borsuk, S., Villela, M., 2022. Molecular characterization of *Giardia lamblia* and risk factors for giardiasis among immunocompromised patients in southern Brazil. *Braz. J. Biol.* 82, e265055. <https://doi.org/10.1590/1519-6984.265055>.
- Jian, F., Qi, M., He, X., Wang, R., Zhang, S., Dong, H., et al., 2014. Occurrence and molecular characterization of *Cryptosporidium* in dogs in Henan province, China. *BMC Vet. Res.* 10, 26.
- Jiang, W., Roellig, D.M., Lebbad, M., Beser, J., Troell, K., Guo, Y., et al., 2020. Subtype distribution of zoonotic pathogen *Cryptosporidium felis* in humans and animals in several countries. *Emerg. Microb. Infect.* 9, 2446–2454.
- Jiang, W., Roellig, D.M., Guo, Y., Li, N., Feng, Y., Xiao, L., 2021. Development of a subtyping tool for zoonotic pathogen *Cryptosporidium canis*. *J. Clin. Microbiol.* 59, e02474–20.
- Jiang, Y., Liu, L., Yuan, Z., Liu, A., Cao, J., Shen, Y., 2023. Molecular identification and genetic characteristics of *Cryptosporidium* spp., *Giardia duodenalis*, and *Enterocytozoon bieneusi* in human immunodeficiency virus/acquired immunodeficiency syndrome patients in Shanghai, China. *Parasites Vectors* 16, 53. <https://doi.org/10.1186/s13071-023-05666-8>.
- Jin, M., Osman, M., Green, B.A., Yang, Y., Ahuja, A., Lu, Z., et al., 2023. Evidence for the transmission of antimicrobial resistant bacteria between humans and companion animals: A scoping review. *One Health* 17, 100593.

- Joachim, A., Auersperg, V., Drüe, J., Wiedermann, S., Hinney, B., Spergser, J., 2023. Parasites and zoonotic bacteria in the feces of cats and dogs from animal shelters in Carinthia, Austria. *Vet. Res.* 164, 105022.
- Johansen, K.M., Castro, N.S., Lancaster, K.E., Madrid, E., Havas, A., Simms, J., et al., 2014. Characterization of *Giardia lamblia* genotypes in dogs from Tucson, Arizona using SSU-rRNA and β-giardin sequences. *Parasitol. Res.* 113, 387–390.
- Julien, D.A., Sergeant, J.M., Guy, R.A., Shapiro, K., Imai, R.K., Bunce, A., et al., 2019. Prevalence and genetic characterization of *Giardia* spp. and *Cryptosporidium* spp. in dogs in Iqaluit, Nunavut, Canada. *Zoonoses Public Health* 66, 813–825.
- Karimi, P., Shafagh-Sisi, S., Meamar, A.R., Razmjou, E., 2023. Molecular identification of *Cryptosporidium*, *Giardia*, and *Blastocystis* from stray and household cats and cat owners in Tehran, Iran. *Sci. Rep.* 13, 1554.
- Katanik, M.T., Schneider, S.K., Rosenblatt, J.E., Hall, G.S., Procop, G.W., 2001. Evaluation of ColorPAC *Giardia/Cryptosporidium* rapid assay and ProSpecT *Giardia/Cryptosporidium* microplate assay for detection of *Giardia* and *Cryptosporidium* in fecal specimens. *J. Clin. Microbiol.* 39, 4523–4525.
- Kaufmann, H., Zenner, L., Benabed, S., Poirel, M.T., Bourgoign, G., 2022. Lack of efficacy of fenbendazole against *Giardia duodenalis* in a naturally infected population of dogs in France. *Parasite* 29, 49.
- Khalil, M.M., Fouad, E.A., Kamel, N.O., Auda, H.M., El-Bahy, M.M., Ramadan, R.M., 2023. Dogs as a source for the spreading of enteric parasites including zoonotic ones in Giza Province, Egypt. *Res. Vet. Sci.* 161, 122–131.
- Khalil, S., Mirdha, B.R., Panda, A., Singh, Y., Makharla, G., Paul, J., 2017. *Cryptosporidium* species subtypes and associated clinical manifestations in Indian patients. *Gastroenterol. Hepatol. Bed. Bench.* 10, 311–318.
- Khine, N.O., Chimnoi, W., Kamyngkird, K., Kengradomkij, C., Saetiew, N., Simking, P., et al., 2021. Molecular detection of *Giardia duodenalis* and *Cryptosporidium* spp. from stray dogs residing in monasteries in Bangkok, Thailand. *Parasitol. Int.* 83, 102337.
- Kim, H.Y., Lee, H., Lee, S.H., Seo, M.G., Yi, S., Kim, J.W., et al., 2019. Multilocus genotyping and risk factor analysis of *Giardia duodenalis* in dogs in Korea. *Acta Trop.* 199, 105113.
- Klotz, C., Sannella, A.R., Weisz, F., Chaudhry, U., Sroka, J., Túmová, P., et al., 2022. Extensive testing of a multi-locus sequence typing scheme for *Giardia duodenalis* assemblage A confirms its good discriminatory power. *Parasites Vectors* 15, 489. <https://doi.org/10.1186/s13071-022-05615-x>.
- Klotz, C., Schmid, M.W., Winter, K., Ignatius, R., Weisz, F., Saghaut, C.S., et al., 2023. Highly contiguous genomes of human clinical isolates of *Giardia duodenalis* reveal assemblage- and sub-assemblage-specific presence-absence variation in protein-coding genes. *Microb. Genom.* 9, mgen000963.
- Koopapong, K., Mori, H., Thammasonthijareern, N., Prasertbun, R., Pintong, A.R., Popruk, S., et al., 2014. Molecular identification of *Cryptosporidium* spp. in seagulls, pigeons, dogs, and cats in Thailand. *Parasite* 21, 52.
- Kooyman, F.N.J., Wagenaar, J.A., Zomer, A., 2019. Whole-genome sequencing of dog-specific assemblages C and D of *Giardia duodenalis* from single and pooled cysts indicates host-associated genes. *Microb. Genom.* 5, e000302.
- Köseoglu, A.E., Can, H., Karakavuk, M., Güvendi, M., Değirmenci Dökşaya, A., Manyatsı, P.B., et al., 2022. Molecular prevalence and subtyping of *Cryptosporidium* spp. in fecal samples collected from stray cats in İzmir, Turkey. *BMC Vet. Res.* 18, 89.
- Kostopoulou, D., Claerebout, E., Arvanitis, D., Ligda, P., Voutzourakis, N., Casaert, S., et al., 2017. Abundance, zoonotic potential and risk factors of intestinal parasitism amongst dog and cat populations: The scenario of Crete, Greece. *Parasites Vectors* 10, 43.
- Koyasu, H., Ogasawara, S., Kikusui, T., Nagasawa, M., 2023. Ownership of dogs and cats leads to higher levels of well-being and general trust through family involvement in late adolescence. *Front. Vet. Sci.* 10, 1220265.
- Krumrie, S., Capewell, P., McDonald, M., Dunbar, D., Panarese, R., Katzer, F., et al., 2022. Molecular characterisation of *Giardia duodenalis* from human and companion animal sources in the United Kingdom using an improved triosephosphate isomerase molecular marker. *Curr. Res. Parasitol. Vector Borne Dis.* 2, 100105.
- Kuthyar, S., Kowalewski, M.M., Seabolt, M., Roellig, D.M., Gillespie, T.R., 2021. Molecular characterization of *Giardia duodenalis* and evidence for cross-species transmission in Northern Argentina. *Transbound. Emerg. Dis.* 69, 2209–2218.
- Kváč, M., Hofmannová, L., Ortega, Y., Holubová, N., Horčícková, M., Kicia, M., et al., 2017. Stray cats are more frequently infected with zoonotic protists than pet cats. *Folia Parasitol.* 64, 2017.034.
- Kwak, D., Seo, M.G., 2020. Genetic analysis of zoonotic gastrointestinal protozoa and microsporidia in shelter cats in South Korea. *Pathogens* 9, 894.
- Labana, R.V., Dungca, J.Z., Nissapattorn, V., 2018. Community-based surveillance of *Cryptosporidium* in the indigenous community of Boliwong, Philippines, from April to December 2017. *Epidemiol. Health* 40, e2018047.
- Lalle, M., Pozio, E., Capelli, G., Bruschi, F., Crotti, D., Cacciò, S.M., 2005a. Genetic heterogeneity at the beta-giardin locus among human and animal isolates of *Giardia duodenalis* and identification of potentially zoonotic subgenotypes. *Int. J. Parasitol.* 35, 207–213.
- Lalle, M., Jimenez-Cardosa, E., Caccio, S.M., Pozio, E., 2005b. Genotyping of *Giardia duodenalis* from humans and dogs from Mexico using a beta-giardin nested polymerase chain reaction assay. *J. Parasitol.* 91, 203–205.
- Learnmonth, J.J., Ionas, G., Ebbett, K.A., Kwan, E.S., 2004. Genetic characterization and transmission cycles of *Cryptosporidium* species isolated from humans in New Zealand. *Appl. Environ. Microbiol.* 70, 3973–3978.
- Lebbad, M., Ankarklev, J., Tellez, A., Leiva, B., Andersson, J.O., Svärd, S., 2008. Dominance of *Giardia* assemblage B in Leon, Nicaragua. *Acta Trop.* 106, 44–53.
- Lebbad, M., Mattsson, J.G., Christensson, B., Ljungström, B., Backhans, A., Andersson, J. O., et al., 2010. From mouse to moose: Multilocus genotyping of *Giardia* isolates from various animal species. *Vet. Parasitol.* 168, 231–239.
- Lebbad, M., Winiecka-Krusnell, J., Stensvold, C.R., Beser, J., 2021. High diversity of *Cryptosporidium* species and subtypes identified in cryptosporidiosis acquired in Sweden and abroad. *Pathogens* 10, 523.
- Lecová, L., Hammerbauerová, I., Túmová, P., Nohýnková, E., 2020. Companion animals as a potential source of *Giardia intestinalis* infection in humans in the Czech Republic - a pilot study. *Vet. Parasitol. Reg. Stud. Reports* 21, 100431.
- Lee, M.F., Cadogan, P., Eytle, S., Copeland, S., Walochnik, J., Lindo, J.F., 2017. Molecular epidemiology and multilocus sequence analysis of potentially zoonotic *Giardia* spp. from humans and dogs in Jamaica. *Parasitol. Res.* 116, 409–414.
- Leonhard, S., Pfister, K., Beelitz, P., Wielinga, C., Thompson, R.C.A., 2007. The molecular characterisation of *Giardia* from dogs in southern Germany. *Vet. Parasitol.* 150, 33–38.
- Leoni, F., Amar, C., Nichols, G., Pedraza Díaz, S., McLauchlin, J., 2006. Genetic analysis of *Cryptosporidium* from 2414 humans with diarrhoea in England between 1985 and 2000. *J. Med. Microbiol.* 55, 703–707.
- Li, J., Zhang, P., Wang, P., Alsarakibi, M., Zhu, H., Liu, Y., et al., 2012. Genotype identification and prevalence of *Giardia duodenalis* in pet dogs of Guangzhou, southern China. *Vet. Parasitol.* 188, 368–371.
- Li, W., Liu, C., Yu, Y., Li, J., Gong, P., Song, M., et al., 2013. Molecular characterization of *Giardia duodenalis* isolates from police and farm dogs in China. *Exp. Parasitol.* 135, 223–226.
- Li, W., Li, Y., Song, M., Lu, Y., Yang, J., Tao, W., et al., 2015. Prevalence and genetic characteristics of *Cryptosporidium*, *Enterocytozoon bieneusi* and *Giardia duodenalis* in cats and dogs in Heilongjiang Province, China. *Vet. Parasitol.* 208, 125–134.
- Li, J., Dan, X., Zhu, K., Li, N., Guo, Y., Zheng, Z., et al., 2019a. Genetic characterization of *Cryptosporidium* spp. and *Giardia duodenalis* in dogs and cats in Guangdong, China. *Parasites Vectors* 12, 571.
- Li, W., Liu, X., Gu, Y., Liu, J., Luo, J., 2019b. Prevalence of *Cryptosporidium*, *Giardia*, *Blastocystis*, and trichomonads in domestic cats in east China. *J. Vet. Med. Sci.* 81, 890–896.
- Li, J., Ryan, U., Guo, Y., Feng, Y., Xiao, L., 2021a. Advances in molecular epidemiology of cryptosporidiosis in dogs and cats. *Int. J. Parasitol.* 51, 787–795.
- Li, J., Yang, F., Liang, R., Guo, S., Guo, Y., Li, N., Feng, Y., et al., 2021b. Subtype Characterization and zoonotic potential of *Cryptosporidium felis* in cats in Guangdong and Shanghai, China. *Pathogens* 10, 89.
- Liang, C., Tsaihong, J.C., Cheng, Y., Peng, S., 2012. Occurrence and genotype of *Giardia* cysts isolated from faecal samples of children and dogs and from drinking water samples in an aboriginal area of central Taiwan. *Exp. Parasitol.* 131, 204–209.
- Liao, S., Lin, X., Sun, Y., Qi, N., Lv, M., Wu, C., et al., 2020. Occurrence and genotypes of *Cryptosporidium* spp., *Giardia duodenalis*, and *Blastocystis* sp. in household, shelter, breeding, and pet market dogs in Guangzhou, southern China. *Sci. Rep.* 10, 17736.
- Liberato, C., Berrilli, F., Odorizi, L., Scarcella, R., Barni, M., Amoruso, C., et al., 2018. Parasites in stray dogs from Italy: Prevalence, risk factors and management concerns. *Acta Parasitol.* 63, 27–32.
- Lim, Y.A., Iqbal, A., Surin, J., Sim, B.L., Jex, A.R., Nolan, M.J., et al., 2011. First genetic classification of *Cryptosporidium* and *Giardia* from HIV/AIDS patients in Malaysia. *Infect. Genet. Evol.* 11, 968–974.
- Liu, H., Shen, Y., Yin, J., Yuan, Z., Jiang, Y., Xu, Y., et al., 2014. Prevalence and genetic characterization of *Cryptosporidium*, *Enterocytozoon*, *Giardia* and *Cyclospora* in diarrhoeal outpatients in China. *BMC Infect. Dis.* 14, 25.
- Liu, H., Shen, Y., Liu, A., Yin, J., Yuan, Z., Jiang, Y., et al., 2017. Occurrence and multilocus genotyping of *Giardia duodenalis* in pets and zoo animals in Shanghai, China. *J. Infect. Dev. Ctries.* 11, 479–486.
- Liu, A., Gong, B., Liu, X., Shen, Y., Wu, Y., Zhang, W., et al., 2020. A retrospective epidemiological analysis of human *Cryptosporidium* infection in China during the past three decades (1987–2018). *PLoS Negl. Trop. Dis.* 14, e0008146.
- Loeck, B.K., Pedati, C., Iwen, P.C., McCutchen, E., Roellig, D.M., Hlavsa, M.C., et al., 2020. Genotyping and subtyping *Cryptosporidium* to identify risk factors and transmission patterns - Nebraska, 2015–2017. *MMWR Morb. Mortal. Wkly. Rep.* 69, 335–338.
- López-Arias, Á., Villar, D., López-Osorio, S., Calle-Vélez, D., Chaparro-Gutiérrez, J.J., 2019. *Giardia* is the most prevalent parasitic infection in dogs and cats with diarrhea in the city of Medellín, Colombia. *Vet. Parasitol. Reg. Stud. Reports* 18, 100335. <https://doi.org/10.1016/j.vprsr.2019.100335>.
- Lucca, P., De Gaspari, E.N., Bozzoli, L.M., Funada, M.R., Silva, S.O., Iuliano, W., Soares, R.M., 2009. Molecular characterization of *Cryptosporidium* spp. from HIV-infected patients from an urban area of Brazil. *Rev. Inst. Med. Trop. São Paulo* 51, 341–343. <https://doi.org/10.1590/s0036-46652009000600006>.
- Lucio-Forster, A., Griffiths, J.K., Cama, V.A., Xiao, L., Bowman, D.D., 2010. Minimal zoonotic risk of cryptosporidiosis from pet dogs and cats. *Trends Parasitol.* 26, 174–179. <https://doi.org/10.1016/j.pt.2010.01.004>.
- Lupo, P.J., Langer-Curry, R.C., Robinson, M., OkhuySEN, P.C., Chappell, C.L., 2008. *Cryptosporidium muris* in a Texas canine population. *Am. J. Trop. Med. Hyg.* 78, 917–921.
- Mancianti, F., Nardoni, S., Mugnaini, L., Zambernardi, L., Guerrini, A., Gazzola, V., et al., 2015. A retrospective molecular study of select intestinal protozoa in healthy pet cats from Italy. *J. Feline Med. Surg.* 17, 163–167.
- Marangi, M., Berrilli, F., Otranto, D., Giangaspero, A., 2010. Genotyping of *Giardia duodenalis* among children and dogs in a closed socially deprived community from Italy. *Zoonoses Public Health* 57, e54–e58.
- Mark-Carew, M.P., Adesiyun, A.A., Basu, A., Georges, K.A., Pierre, T., Tilitz, S., et al., 2013. Characterization of *Giardia duodenalis* infections in dogs in Trinidad and Tobago. *Vet. Parasitol.* 196, 199–202.
- Martins, C.F., Soares, J.P., Cortinhas, A., Silva, L., Cardoso, L., Pires, M.A., Mota, M.P., 2023. Pet's influence on humans' daily physical activity and mental health: A meta-analysis. *Front. Public Health* 11, 1196199.

- Mateo, M., Montoya, A., Bailo, B., Köster, P.C., Dashti, A., Hernández-Castro, C., et al., 2023. Prevalence and public health relevance of enteric parasites in domestic dogs and cats in the region of Madrid (Spain) with an emphasis on *Giardia duodenalis* and *Cryptosporidium* sp. Vet. Med. Sci. 9, 2542–2558. <https://doi.org/10.1002/vms3.1270>.
- Matos, O., Alves, M., Xiao, L., Cama, V., Antunes, F., 2004. *Cryptosporidium felis* and *C. meleagridis* in persons with HIV, Portugal. Emerg. Infect. Dis. 10, 2256–2257.
- McDowall, R.M., Peregrine, A.S., Leonard, E.K., Lacombe, C., Lake, M., Rebelo, A.R., Cai, H.Y., 2011. Evaluation of the zoonotic potential of *Giardia duodenalis* in fecal samples from dogs and cats in Ontario. Can. Vet. J. 52, 1329–1333.
- Meng, X.Z., Li, M.Y., Lyu, C., Qin, Y.F., Zhao, Z.Y., Yang, X.B., et al., 2021. The global prevalence and risk factors of *Cryptosporidium* infection among cats during 1988–2021: A systematic review and meta-analysis. Microb. Pathog. 158, 105096.
- Minetti, C., Lamden, K., Durband, C., Cheesbrough, J., Platt, K., Charlett, A., et al., 2015. Case-control study of risk factors for sporadic giardiasis and parasite assemblages in North-West England. J. Clin. Microbiol. 53, 3133–3140.
- Minivelle, M.C., Molina, N.B., Polverino, D., Basualdo, J.A., 2008. First genotyping of *Giardia lamblia* from human and animal feces in Argentina, South America. Mem. Inst. Oswaldo Cruz 103, 98–103.
- Miska, K.B., Jenkins, M.C., Trout, J.M., Santín, M., Fayer, R., 2009. Detection and comparison of *Giardia* virus (GLV) from different assemblages of *Giardia duodenalis*. J. Parasitol. 95, 1197–1200.
- Molloy, S.F., Kirwan, P., Asaolu, S.O., Holland, C.V., Nichols, R.A.B., Connolly, L., Smith, H.V., 2010. Identification of a high diversity of *Cryptosporidium* species genotypes and subtypes in a pediatric population in Nigeria. Am. J. Trop. Med. Hyg. 82, 608–613.
- Monis, P.T., Andrews, R.H., Mayrhofer, G., Mackrill, J., Kulda, J., Isaac-Renton, J.L., Ey, P.L., 1998. Novel lineages of *Giardia intestinalis* identified by genetic analysis of organisms isolated from dogs in Australia. Parasitology 116, 7–19.
- Monis, P.T., Andrews, R.H., Mayrhofer, G., Ey, P.L., 2003. Genetic diversity within the morphological species *Giardia intestinalis* and its relationship to host origin. Infect. Genet. Evol. 3, 29–38.
- Moore, C.E., Elwin, K., Seng, C., Mao, S., Suy, K., Kumar, V., et al., 2016. Molecular characterization of *Cryptosporidium* species and *Giardia duodenalis* from symptomatic Cambodian children. PLoS Negl. Trop. Dis. 10, e0004822.
- Moreira, A.S., Baptista, C.T., Brasil, C.L., Valente, J.S.S., Bruhn, F.R.P., Pereira, D.I.B., 2018. Risk factors and infection due to *Cryptosporidium* spp. in dogs and cats in southern Rio Grande do Sul. Rev. Bras. Parasitol. Vet. 27, 112–117.
- Morgan, U.M., Sargent, K.D., Elliot, A., Thompson, R.C., 1998. *Cryptosporidium* in cats - additional evidence for *C. felis*. Vet. J. 156, 159–161.
- Morgan, U., Weber, R., Xiao, L., Sulaiman, I., Thompson, R.C., Ndhiritu, W., et al., 2000a. Molecular characterization of *Cryptosporidium* isolates obtained from human immunodeficiency virus-infected individuals living in Switzerland, Kenya, and the United States. J. Clin. Microbiol. 38, 1180–1183.
- Morgan, U.M., Xiao, L., Monis, P., Fall, A., Irwin, P.J., Fayer, R., et al., 2000b. *Cryptosporidium* spp. in domestic dogs: the “dog” genotype. Appl. Environ. Microbiol. 66, 2220–2223.
- Morrison, H.G., McArthur, A.G., Gillin, F.D., Aley, S.B., Adam, R.D., Olsen, G.J., et al., 2007. Genomic minimalism in the early diverging intestinal parasite *Giardia lamblia*. Science 317, 1921–1926.
- Muadica, A.S., Koster, P.C., Dashti, A., Bailo, B., Hernández-de-Mingo, M., Balasegaram, S., Carmena, D., 2021. Molecular diversity of *Giardia duodenalis*, *Cryptosporidium* spp. and *Blastocystis* sp. in symptomatic and asymptomatic school children in Zambezia province (Mozambique). Pathogens 10, 255.
- Mulunda, N.R., Hayashida, K., Yamagishi, J., Sianongo, S., Munsaka, G., Sugimoto, C., Mutengo, M.M., 2020. Molecular characterization of *Cryptosporidium* spp. from patients with diarrhoea in Lusaka, Zambia. Parasite 27, 53.
- Munoz, J., Mayer, D.C., 2016. *Toxoplasma gondii* and *Giardia duodenalis* infections in domestic dogs in New York City public parks. Vet. J. 211, 97–99.
- Murnik, L.C., Daugschies, A., Delling, C., 2022. *Cryptosporidium* infection in young dogs from Germany. Parasitol. Res. 121, 2985–2993.
- Murnik, L.C., Daugschies, A., Delling, C., 2023. Gastrointestinal parasites in young dogs and risk factors associated with infection. Parasitol. Res. 122, 585–596.
- Muthusamy, D., Rao, S.S., Ramani, S., Monica, B., Banerjee, I., Abraham, O.C., et al., 2006. Multilocus genotyping of *Cryptosporidium* sp. isolates from human immunodeficiency virus-infected individuals in South India. J. Clin. Microbiol. 44, 632–634.
- Nagamori, Y., Payton, M.E., Looper, E., Apple, H., Johnson, E.M., 2020. Retrospective survey of parasitism identified in feces of client-owned cats in North America from 2007 through 2018. Vet. Parasitol. 277, 109008.
- Neira, P., Munoz, N., Rosales, J., 2010. *Cryptosporidium parvum* infection in a pregnant immunocompetent woman with occupational risk. Rev. Chilena Infectol. 27, 345–349.
- Ng, J., Yang, R., Whiffin, V., Cox, P., Ryan, U., 2011. Identification of zoonotic *Cryptosporidium* and *Giardia* genotypes infecting animals in Sydney's water catchments. Exp. Parasitol. 128, 138–144.
- Nguyen, S.T., Fukuda, Y., Nguyen, D.T., Dao, H.T., Le, D.Q., Bui, K.L., et al., 2018. Prevalence, genotyping and risk factors of *Giardia duodenalis* from dogs in Vietnam. J. Vet. Med. Sci. 80, 92–97.
- Nolan, M.J., Jex, A.R., Koehler, A.V., Haydon, S.R., Stevens, M.A., Gasser, R.B., 2013. Molecular-based investigation of *Cryptosporidium* and *Giardia* from animals in water catchments in southeastern Australia. Water Res. 47, 1726–1740.
- Nydam, D.V., Wade, S.E., Schafaf, S.L., Mohammed, H.O., 2001. Number of *Cryptosporidium parvum* oocysts or *Giardia* spp. cysts shed by dairy calves after natural infection. Am. J. Vet. Res. 62, 1612–1615.
- Önder, Z., Yetişmiş, G., Pekmezci, D., Delibaşı Kökü, N., Pekmezci, G.Z., Çiloğlu, A., et al., 2021. Investigation of zoonotic *Cryptosporidium* and *Giardia intestinalis* species and genotypes in cats (*Felis catus*). Turk. Parazitoloji Derg. 45, 252–256.
- Osman, M., Bories, J., El Safadi, D., Poirel, M.T., Gantois, N., Benamrouz-Vanneste, S., et al., 2015. Prevalence and genetic diversity of the intestinal parasites *Blastocystis* sp. and *Cryptosporidium* spp. in household dogs in France and evaluation of zoonotic transmission risk. Vet. Parasitol. 214, 167–170.
- Overgaauw, P.A., van Zutphen, L., Hoek, D., Yaya, F.O., Roelfsema, J., Pinelli, E., et al., 2009. Zoonotic parasites in fecal samples and fur from dogs and cats in the Netherlands. Vet. Parasitol. 163, 115–122.
- Overgaauw, P.A.M., Vinke, C.M., Hagen, M.A.E.V., Lipman, L.J.A., 2020. A One Health perspective on the human-companion animal relationship with emphasis on zoonotic aspects. Int. J. Environ. Res. Publ. Health 17, 3789.
- Pacheco, F.T.F., Freitas, H.F., Silva, R.K.N.R., Carvalho, S.S., Martins, A.S., Menezes, J.F., et al., 2022. *Cryptosporidium* diagnosis in different groups of children and characterization of parasite species. Rev. Soc. Bras. Med. Trop. 55, e00412022.
- Paim Arruda Trevisan, Y., Do Bom Parto Ferreira de Almeida, A., Nakazato, L., Dos Anjos Pacheco, T., Iglesias de Souza, J., Henrique Canei, D., et al., 2020. Frequency of *Giardia duodenalis* infection and its genetic variability in dogs in Cuiabá, Midwest Brazil. J. Infect. Dev. Ctries. 14, 1431–1436.
- Pallant, L., Barutzki, D., Schaper, R., Thompson, R.C.A., 2015. The epidemiology of infections with *Giardia* species and genotypes in well cared for dogs and cats in Germany. Parasites Vectors 8, 2.
- Palmer, C.S., Thompson, R.C., Traub, R.J., Rees, R., Robertson, I.D., 2008a. National study of the gastrointestinal parasites of dogs and cats in Australia. Vet. Parasitol. 151, 181–190.
- Palmer, C.S., Traub, R.J., Robertson, I.D., Devlin, G., Rees, R., Thompson, R.C., 2008b. Determining the zoonotic significance of *Giardia* and *Cryptosporidium* in Australian dogs and cats. Vet. Parasitol. 154, 142–147.
- Pan, W., Wang, M., Abdullahi, A.Y., Fu, Y., Yan, X., Yang, F., et al., 2018. Prevalence and genotypes of *Giardia lamblia* from stray dogs and cats in Guangdong, China. Vet. Parasitol. Reg. Stud. Reports 13, 30–34.
- Paoletti, B., Iorio, R., Capelli, G., Sparagano, O.A., Giangaspero, A., 2008. Epidemiological scenario of giardiosis in dogs from central Italy. Ann. N. Y. Acad. Sci. 1149, 371–374.
- Paoletti, B., Otranto, D., Weigl, S., Giangaspero, A., Di Cesare, A., Traversa, D., 2011. Prevalence and genetic characterization of *Giardia* and *Cryptosporidium* in cats from Italy. Res. Vet. Sci. 91, 397–399.
- Paoletti, B., Traversa, D., Iorio, R., De Berardinis, A., Bartolini, R., Salini, R., Di Cesare, A., 2015. Zoonotic parasites in feces and fur of stray and private dogs from Italy. Parasitol. Res. 114, 2135–2141. <https://doi.org/10.1007/s00436-015-4402-6>.
- Papini, R., Cardini, G., Paoletti, B., Giangaspero, A., 2007. Detection of *Giardia* assemblage A in cats in Florence, Italy. Parasitol. Res. 100, 653–656.
- Patterson, J., 2023. Toxocarosis in humans: How much of a problem is it in the UK? Drug Therapeut. Bull. 61, 7–11.
- Pavlásek, I., Ryan, U., 2007. The first finding of a natural infection of *Cryptosporidium muris* in a cat. Vet. Parasitol. 144, 349–352.
- Pedraza-Díaz, S., Amar, C., Iversen, A.M., Stanley, P.J., McLauchlin, J., 2001. Unusual *Cryptosporidium* species recovered from human faeces: First description of *Cryptosporidium felis* and *Cryptosporidium* “dog type” from patients in England. J. Med. Microbiol. 50, 293–296.
- Pereira, A., Teixeira, J., Sousa, S., Parreira, R., Campino, L., Meireles, J., Maia, C., 2021. *Giardia duodenalis* infection in dogs from the metropolitan area of Lisbon, Portugal: Prevalence, genotyping and associated risk factors. J. Parasit. Dis. 45, 372–379.
- Perrucci, S., Berrilli, F., Procopio, C., Di Filippo, M.M., Pierini, A., Marchetti, V., 2020. *Giardia duodenalis* infection in dogs affected by primary chronic enteropathy. Open Vet. J. 10, 74–79.
- Piekarska-Stepińska, A., Piekarska, J., Gorczykowski, M., 2021. *Cryptosporidium* spp. in dogs and cats in Poland. Ann. Agric. Environ. Med. 28, 345–347.
- Piekarska, J., Bajzert, J., Gorczykowski, M., Kantyka, M., Podkowik, M., 2016. Molecular identification of *Giardia duodenalis* isolates from domestic dogs and cats in Wrocław, Poland. Ann. Agric. Environ. Med. 23, 410–415.
- Pieniazek, N.J., Bornay-Llinares, F.J., Slemenda, S.B., da Silva, A.J., Moura, I.N., Arrowood, M.J., et al., 1999. New *Cryptosporidium* genotypes in HIV-infected persons. Emerg. Infect. Dis. 5, 444–449.
- Pipia, A.P., Varcasia, A., Tamponi, C., Sanna, G., Soda, M., Paoletti, B., et al., 2014. Canine giardiosis in Sardinia Island, Italy: Prevalence, molecular characterization, and risk factors. J. Infect. Dev. Ctries. 8, 655–660.
- Pipková, J., Papajová, I., Majláthová, V., Soltys, J., Bystrianska, J., Schusterová, I., Vargová, V., 2022. First report on *Giardia duodenalis* assemblage F in Slovakian children living in poor environmental conditions. J. Microbiol. Immunol. Infect. 53, 148–156. <https://doi.org/10.1016/j.jmii.2018.04.007>.
- Ponce-Macotela, M., Martínez-Gordillo, M.N., Bermúdez-Cruz, R.M., Salazar-Schettino, P.M., Ortega-Pierres, G., Ey, P.L., 2002. Unusual prevalence of the *Giardia intestinalis* A-II subtype amongst isolates from humans and domestic animals in Mexico. Int. J. Parasitol. 32, 1201–1202.
- Potes-Morales, C., Crespo-Ortiz, M.D.P., 2023. Molecular diagnosis of intestinal protozoa in young adults and their pets in Colombia, South America. PLoS One 18, e0283824.
- Puebla, L.E.J., Nunez, F.A., Rivero, L.R., Hernandez, Y.R., Millan, I.A., Muller, N., 2017. Prevalence of intestinal parasites and molecular characterization of *Giardia duodenalis* from dogs in La Habana, Cuba. Vet. Parasitol. Reg. Stud. Reports 8, 107–112.
- Qi, M., Dong, H., Wang, R., Li, J., Zhao, J., Zhang, L., Luo, J., 2016. Infection rate and genetic diversity of *Giardia duodenalis* in pet and stray dogs in Henan Province, China. Parasitol. Int. 65, 159–162.

- Quadros, R.M., Weiss, P.H., Marques, S.M., Milette, L.C., 2016. Potential cross-contamination of similar *Giardia duodenalis* assemblage in children and pet dogs in southern Brazil, as determined by PCR-RFLP. Rev. Inst. Med. Trop. São Paulo 58, 66.
- Raccurt, C.P., Brasseur, P., Verdier, R.I., Li, X., Eyma, E., Stockman, C.P., et al., 2006. Human cryptosporidiosis and *Cryptosporidium* spp. in Haiti. Trop. Med. Int. Health 11, 929–934.
- Ramírez-Ocampo, S., Cotte-Alzate, J.D., Escobedo, Á.A., Rodríguez-Morales, A.J., 2017. Prevalence of zoonotic and non-zoonotic genotypes of *Giardia intestinalis* in cats: A systematic review and meta-analysis. Infekz. Med. 25, 326–338.
- Ranjbar, R., Mirhendi, H., Izadi, M., Behrouz, B., Mohammadi Manesh, R., 2018. Molecular identification of *Cryptosporidium* spp. in Iranian dogs using seminested PCR: A first report. Vector Borne Zoonotic Dis. 18, 96–100.
- Read, C.M., Monis, P.T., Thompson, R.C., 2004. Discrimination of all genotypes of *Giardia duodenalis* at the glutamate dehydrogenase locus using PCR-RFLP. Infect. Genet. Evol. 4, 125–130.
- Rehbein, S., Klotz, C., Ignatius, R., Müller, E., Aebsicher, A., Kohn, B., 2019. *Giardia duodenalis* in small animals and their owners in Germany: A pilot study. Zoonoses Public Health 66, 117–124.
- Rendtorff, R.C., 1979. The experimental transmission of *Giardia lamblia* among volunteer subjects. In: Waterborne Transmission of Giardiasis. Proceeding of a Symposium, Cincinnati, USA.
- Rimhanen-Finne, R., Enemark, H.L., Kolehmainen, J., Toropainen, P., Hanninen, M.L., 2007. Evaluation of immunofluorescence microscopy and enzyme-linked immunosorbent assay in detection of *Cryptosporidium* and *Giardia* infections in asymptomatic dogs. Vet. Parasitol. 145, 345–348.
- Roegner, A.F., Daniels, M.E., Smith, W.A., Gottdenker, N., Schwartz, L.M., Liu, J., et al., 2019. *Giardia* infection and *Trypanosoma cruzi* exposure in dogs in the Bosawas biosphere reserve, Nicaragua. EcoHealth 16, 512–522.
- Rojas-Lopez, L., Elwin, K., Chalmers, R.M., Enemark, H.L., Beser, J., Troell, K., 2020. Development of a gp60-subtyping method for *Cryptosporidium felis*. Parasites Vectors 13, 39.
- Rosanowski, S.M., Banica, M., Ellis, E., Farrow, E., Harwood, C., Jordan, B., et al., 2018. The molecular characterisation of *Cryptosporidium* species in relinquished dogs in Great Britain: A novel zoonotic risk? Parasitol. Res. 117, 1663–1667.
- Ryan, U., Feng, Y., Fayer, R., Xiao, L., 2021a. Taxonomy and molecular epidemiology of *Cryptosporidium* and *Giardia* - a 50-year perspective (1971–2021). Int. J. Parasitol. 51, 1099–1119.
- Ryan, U., Zahedi, A., Feng, Y., Xiao, L., 2021b. An update on zoonotic *Cryptosporidium* species and genotypes in humans. Animals 11, 3307.
- Saleh, M.N., Lindsay, D.S., Leib, M.S., Zajac, A.M., 2019. *Giardia duodenalis* assemblages in cats from Virginia, USA. Vet. Parasitol. Reg. Stud. Reports 15, 100257.
- Saleh, M.N., Allen, K.E., Lineberry, M.W., Little, S.E., Reichard, M.V., 2021. Ticks infesting dogs and cats in North America: Biology, geographic distribution, and pathogen transmission. Vet. Parasitol. 294, 109392.
- Sannella, A.R., Suputtamongkol, Y., Wongswat, E., Cacciò, S.M., 2019. A retrospective molecular study of *Cryptosporidium* species and genotypes in HIV-infected patients from Thailand. Parasites Vectors 12, 91.
- Santín, M., 2020. *Cryptosporidium* and *Giardia* in ruminants. Vet. Clin. North. Am. Food Anim. Pract. 36, 223–238.
- Santín, M., Trout, J.M., Vecino, J.A., Dubey, J.P., Fayer, R., 2006. *Cryptosporidium*, *Giardia* and *Enterocytozoon bieneusi* in cats from Bogota (Colombia) and genotyping of isolates. Vet. Parasitol. 141, 334–339.
- Sargent, K.D., Morgan, U.M., Elliot, A., Thompson, R.C., 1998. Morphological and genetic characterisation of *Cryptosporidium* oocysts from domestic cats. Vet. Parasitol. 77, 221–227.
- Scaramozzino, P., Di Cave, D., Berrilli, F., D'Orazi, C., Spaziani, A., Mazzanti, S., et al., 2009. A study of the prevalence and genotypes of *Giardia duodenalis* infecting kennelled dogs. Vet. J. 182, 231–234.
- Schurer, J.M., Hill, J.E., Fernando, C., Jenkins, E.J., 2012. Sentinel surveillance for zoonotic parasites in companion animals in indigenous communities of Saskatchewan. Am. J. Trop. Med. Hyg. 87, 495–498.
- Scorza, A.V., Lappin, M.R., 2017. Prevalence of selected zoonotic and vector-borne agents in dogs and cats on the pine ridge reservation. Vet. Sci. 4, 43.
- Scorza, A.V., Duncan, C., Miles, L., Lappin, M.R., 2011. Prevalence of selected zoonotic and vector-borne agents in dogs and cats in Costa Rica. Vet. Parasitol. 183, 178–183.
- Semani, M., Costa, D., Achour, N., Cherchar, M., Ziane, H., Mouhajir, A., et al., 2023. Occurrence and molecular characterization of *Cryptosporidium* infection in HIV/AIDS patients in Algeria. Viruses 15, 362.
- Shin, J.C., Reyes, A.W., Kim, S.H., Kim, S., Park, H.J., Seo, K.W., Song, K.H., 2015. Molecular detection of *Giardia intestinalis* from stray dogs in animal shelters of Gyeongsangbuk-do (Province) and Daejeon, Korea. Kor. J. Parasitol. 53, 477–481.
- Silva, A.C.D.S., Martins, F.D.C., Ladeia, W.A., Kakimori, M.T.A., Lucas, J.I., Sasse, J.P., et al., 2022. First report of *Giardia duodenalis* assemblage F in humans and dogs in southern Brazil. Comp. Immunol. Microbiol. Infect. Dis. 89, 101878.
- Simonato, G., Frangipane di Regalbono, A., Cassini, R., Traversa, D., Beraldo, P., Tessarin, C., Pietrobelli, M., 2015. Copromicroscopic and molecular investigations on intestinal parasites in kenneled dogs. Parasitol. Res. 114, 1963–1970.
- Simonato, G., Frangipane di Regalbono, A., Cassini, R., Traversa, D., Tessarin, C., DiCesare, A., Pietrobelli, M., 2017. Molecular detection of *Giardia duodenalis* and *Cryptosporidium* spp. in canine faecal samples contaminating public areas in northern Italy. Parasitol. Res. 116, 3411–3418.
- Smith, R.P., Chalmers, R.M., Mueller-Doblies, D., Clifton-Hadley, F.A., Elwin, K., et al., 2010. Investigation of farms linked to human patients with cryptosporidiosis in England and Wales. Prev. Vet. Med. 94, 9–17.
- Smith, A.F., Neumann, N., Banting, G., Klein, C., Liccioli, S., Massolo, A., 2020. Molecular characterization of *Giardia* spp. and *Cryptosporidium* spp. from dogs and coyotes in an urban landscape suggests infrequent occurrence of zoonotic genotypes. Vet. Parasitol. 281, 109115.
- Solarczyk, P., Majewska, A.C., 2010. A survey of the prevalence and genotypes of *Giardia duodenalis* infecting household and sheltered dogs. Parasitol. Res. 106, 1015–1019.
- Soliman, R.H., Fuentes, I., Rubio, J.M., 2011. Identification of a novel assemblage B subgenotype and a zoonotic assemblage C in human isolates of *Giardia intestinalis* in Egypt. Parasitol. Int. 60, 507–511.
- Sommer, M.F., Beck, R., Ionita, M., Stefanovska, J., Vasić, A., Zdravković, N., Hamel, D., et al., 2015. Multilocus sequence typing of canine *Giardia duodenalis* from south-eastern European countries. Parasitol. Res. 114, 2165–2174.
- Sommer, M.F., Rupp, P., Pietsch, M., Kaspar, A., Beelitz, P., 2018. *Giardia* in a selected population of dogs and cats in Germany - diagnostics, coinfections and assemblages. Vet. Parasitol. 249, 49–56.
- Sotiriadou, I., Pantchev, N., Gassmann, D., Karanis, P., 2013. Molecular identification of *Giardia* and *Cryptosporidium* from dogs and cats. Parasite 20, 8.
- Souza, S.L., Gennari, S.M., Richtzenhain, L.J., Pena, H.F., Funada, M.R., Cortez, A., et al., 2007. Molecular identification of *Giardia duodenalis* isolates from humans, dogs, cats and cattle from the state of São Paulo, Brazil, by sequence analysis of fragments of glutamate dehydrogenase (gdh) coding gene. Vet. Parasitol. 149, 258–264.
- Sprong, H., Cacciò, S.M., van der Giessen, J.W., ZOOPNET network and partners, 2009. Identification of zoonotic genotypes of *Giardia duodenalis*. PLoS Negl. Trop. Dis. 3, e558.
- Srisuphanunt, M., Saksirisampant, W., Karanis, P., 2011. Prevalence and genotyping of *Cryptosporidium* isolated from HIV/AIDS patients in urban areas of Thailand. Ann. Trop. Med. Parasitol. 105, 463–468.
- Štrkolcová, G., Maďar, M., Hinney, B., Goldová, M., Mojžišová, J., Halánková, M., 2015. Dog's genotype of *Giardia duodenalis* in human: First evidence in Europe. Acta Parasitol. 60, 796–799.
- Sui, Y., Zhang, X., Wang, H., Yu, F., Zheng, L., Guo, Y., et al., 2022. Prevalence and genetic diversity of *Giardia duodenalis* in pet dogs from Zhengzhou, central China and the association between gut microbiota and fecal characteristics during infection. One Health 14, 100401.
- Sulaiman, I.M., Fayer, R., Bern, C., Gilman, R.H., Trout, J.M., Schantz, P.M., et al., 2003. Triosephosphate isomerase gene characterization and potential zoonotic transmission of *Giardia duodenalis*. Emerg. Infect. Dis. 9, 1444–1452.
- Sursal, N., Simsek, E., Yıldız, K., 2020. Feline giardiasis in Turkey: Prevalence and genetic and haplotype diversity of *Giardia duodenalis* based on the β-giardin gene sequence in symptomatic cats. J. Parasitol. 106, 699–706.
- Sutthikornchai, C., Popruk, S., Mahittikorn, A., Arthan, D., Soonthornworasiri, N., Parathakonkun, C., et al., 2021. Molecular detection of *Cryptosporidium* spp., *Giardia duodenalis*, and *Enterocytozoon bieneusi* in school children at the Thai-Myanmar border. Parasitol. Res. 120, 2887–2895.
- Suzuki, J., Murata, R., Kobayashi, S., Sadamasu, K., Kai, A., Takeuchi, T., 2011. Risk of human infection with *Giardia duodenalis* from cats in Japan and genotyping of the isolates to assess the route of infection in cats. Parasitology 138, 493–500.
- Szénási, Z., Marton, S., Kucsera, I., Tánczos, B., Horváth, K., Orosz, E., et al., 2007. Preliminary investigation of the prevalence and genotype distribution of *Giardia intestinalis* in dogs in Hungary. Parasitol. Res. 101, 145–152.
- Taghipour, A., Olfatifar, M., Bahadory, S., Godfrey, S.S., Abdoli, A., Khatami, A., Javamard, E., Shahrivar, F., 2020. The global prevalence of *Cryptosporidium* infection in dogs: A systematic review and meta-analysis. Vet. Parasitol. 281, 109093.
- Tahir, D., Davoust, B., Parola, P., 2019. Vector-borne nematode diseases in pets and humans in the Mediterranean Basin: An update. Vet. World 12, 1630–1643.
- Tan, L., Wu, S., Abdullahi, A.Y., Yu, X., Hu, W., Song, M., et al., 2016. PCR-RFLP method to detect zoonotic and host-specific *Giardia duodenalis* assemblages in dog fecal samples. Parasitol. Res. 115, 2045–2050.
- Tangtrongsup, S., Scorza, A.V., Reif, J.S., Ballweber, L.R., Lappin, M.R., Salman, M.D., 2017. Prevalence and multilocus genotyping analysis of *Cryptosporidium* and *Giardia* isolates from dogs in Chiang Mai, Thailand. Vet. Sci. 4, 26.
- Tangtrongsup, S., Scorza, A.V., Reif, J.S., Ballweber, L.R., Lappin, M.R., Salman, M.D., 2020. Seasonal distributions and other risk factors for *Giardia duodenalis* and *Cryptosporidium* spp. infections in dogs and cats in Chiang Mai, Thailand. Prev. Vet. Med. 174, 104820.
- Thomaz, A., Meireles, M.V., Soares, R.M., Pena, H.F., Gennari, S.M., 2007. Molecular identification of *Cryptosporidium* spp. from fecal samples of felines, canines and bovines in the state of São Paulo, Brazil. Vet. Parasitol. 150, 291–296.
- Thompson, R.C.A., 2004. Zoonotic significance and molecular epidemiology of *Giardia* and giardiasis. Vet. Parasitol. 126, 15–35.
- Tiangtip, R., Jongwutiwes, S., 2002. Molecular analysis of *Cryptosporidium* species isolated from HIV-infected patients in Thailand. Trop. Med. Int. Health 7, 357–364.
- Traub, R.J., Monis, P.T., Robertson, I., Irwin, P., Mencke, N., Thompson, R.C., 2004. Epidemiological and molecular evidence supports the zoonotic transmission of *Giardia* among humans and dogs living in the same community. Parasitology 128, 253–262.
- Traub, R.J., Inpankaew, T., Reid, S.A., Sutthikornchai, C., Sukthana, Y., Robertson, I.D., Thompson, R.C., 2009. Transmission cycles of *Giardia duodenalis* in dogs and humans in Temple communities in Bangkok - a critical evaluation of its prevalence using three diagnostic tests in the field in the absence of a gold standard. Acta Trop. 111, 125–132.
- Tseng, Y.C., Ho, G.D., Chen, T.T.W., Huang, B.F., Cheng, P.C., Chen, J.L., Peng, S.Y., 2014. Prevalence and genotype of *Giardia duodenalis* from faecal samples of stray dogs in Hualien city of eastern Taiwan. Trop. Biomed. 31, 305–311.
- Uehlinger, F.D., Greenwood, S.J., McClure, J.T., Conboy, G., O'Handley, R., Barkema, H. W., 2013. Zoonotic potential of *Giardia duodenalis* and *Cryptosporidium* spp. and

- prevalence of intestinal parasites in young dogs from different populations on Prince Edward Island, Canada. *Vet. Parasitol.* 196, 509–514.
- Uiterwijk, M., Mughini-Gras, L., Nijssse, R., Wagenaar, J.A., Ploeger, H.W., Kooyman, F.N.J., 2020. *Giardia duodenalis* multi-locus genotypes in dogs with different levels of synanthropism and clinical signs. *Parasites Vectors* 13, 605.
- Ukwah, B.N., Ezeonu, I.M., Ezeonu, C.T., Roellig, D., Xiao, L., 2017. *Cryptosporidium* species and subtypes in diarrheal children and HIV-infected persons in Ebonyi and Nsukka, Nigeria. *J. Infect. Dev. Ctries.* 11, 173–179.
- Upjohn, M., Cobb, C., Monger, J., Geurden, T., Claerebout, E., Fox, M., 2010. Prevalence, molecular typing and risk factor analysis for *Giardia duodenalis* infections in dogs in a central London rescue shelter. *Vet. Parasitol.* 172, 341–346.
- Uran-Velasquez, J., Alzate, J.F., Farfan-Garcia, A.E., Gomez-Duarte, O.G., Martinez-Rosado, L.L., Dominguez-Hernandez, D.D., et al., 2022. Multilocus sequence typing helps understand the genetic diversity of *Cryptosporidium hominis* and *Cryptosporidium parvum* isolated from Colombian patients. *PLoS One* 17, e0270995.
- Utaaker, K.S., Tysnes, K.R., Kroness, M.M., Robertson, L.J., 2018. Not just a walk in the park: Occurrence of intestinal parasites in dogs roaming recreational parks in Chandigarh, Northern India. *Vet. Parasitol. Reg. Stud. Reports* 14, 176–180.
- Vasilopoulos, R.J., Rickard, L.G., Mackin, A.J., Pharr, G.T., Huston, C.L., 2007. Genotypic analysis of *Giardia duodenalis* in domestic cats. *J. Vet. Intern. Med.* 21, 352–355.
- Veldhuis, F.L., Nijssse, R., Wagenaar, J.A., Arkesteijn, G., Kooyman, F.N.J., 2022. Variation in haplotypes in single cysts of assemblages C and D, but not of assemblage E of *Giardia duodenalis*. *BMC Microbiol.* 22, 166.
- Veyna-Salazar, N.P., Cantó-Alarcón, G.J., Olvera-Ramírez, A.M., Ruiz-López, F.J., Bernal-Reynaga, R., Bárcenas-Reyes, J., Durán-Aguilar, M., 2023. Occurrence of *Giardia duodenalis* in cats from Querétaro and the risk to public health. *Animals* 13, 1098.
- Volotão, A.C., Costa-Macedo, L.M., Haddad, F.S.M., Brandao, A., Peralta, J.M., Fernandes, O., 2007. Genotyping of *Giardia duodenalis* from human and animal samples from Brazil using beta-giardin gene: A phylogenetic analysis. *Acta Trop.* 102, 10–19.
- Wang, A., Ruch-Gallie, R., Scorza, V., Lin, P., Lappin, M.R., 2012. Prevalence of *Giardia* and *Cryptosporidium* species in dog park attending dogs compared to non-dog park attending dogs in one region of Colorado. *Vet. Parasitol.* 184, 335–340.
- Wang, Y.G., Zou, Y., Yu, Z.Z., Chen, D., Gui, B.Z., Yang, J.F., et al., 2021. Molecular investigation of zoonotic intestinal protozoa in pet dogs and cats in Yunnan Province, Southwestern China. *Pathogens* 10, 1107.
- Wang, W., Wei, Y., Cao, S., Wu, W., Zhao, W., Guo, Y., et al., 2022a. Divergent *Cryptosporidium* species and host-adapted *Cryptosporidium canis* subtypes in farmed minks, raccoon dogs and foxes in Shandong, China. *Front. Cell. Infect. Microbiol.* 12, 980917.
- Wang, T., Wei, Z., Zhang, Y., Zhang, Q., Zhang, L., Yu, F., et al., 2022b. Molecular detection and genetic characterization of *Cryptosporidium* in kindergarten children in Southern Xinjiang, China. *Infect. Genet. Evol.* 103, 105339.
- Wood, L., Martin, K., Christian, H., Nathan, A., Lauritsen, C., Houghton, S., et al., 2015. The pet factor-companion animals as a conduit for getting to know people, friendship formation and social support. *PLoS One* 10, e0122085.
- Wu, Y., Yao, L., Chen, H., Zhang, W., Jiang, Y., Yang, F., et al., 2022. *Giardia duodenalis* in patients with diarrhea and various animals in northeastern China: Prevalence and multilocus genetic characterization. *Parasites Vectors* 15, 165.
- Xiao, L., Bern, C., Limor, J., Sulaiman, I., Roberts, J., Checkley, W., et al., 2001. Identification of 5 types of *Cryptosporidium* parasites in children in Lima, Peru. *J. Infect. Dis.* 183, 492–497.
- Xiao, L., Cama, V.A., Cabrera, L., Ortega, Y., Pearson, J., Gilman, R.H., 2007. Possible transmission of *Cryptosporidium canis* among children and a dog in a household. *J. Clin. Microbiol.* 45, 2014–2016.
- Xu, H., Jin, Y., Wu, W., Li, P., Wang, L., Li, N., Feng, Y., Xiao, L., 2016. Genotypes of *Cryptosporidium* spp., *Enterocytozoon bieneusi* and *Giardia duodenalis* in dogs and cats in Shanghai, China. *Parasites Vectors* 9, 121.
- Yamamoto, N., Kon, M., Saito, T., Maeno, N., Koyama, M., Sunaoshi, K., et al., 2009. Prevalence of intestinal canine and feline parasites in Saitama Prefecture, Japan. *Kansenshogaku Zasshi* 83, 223–228.
- Yang, R., Jacobson, C., Gardner, G., Carmichael, I., Campbell, A.J., Ng-Hublin, J., Ryan, U., 2014. Longitudinal prevalence, oocyst shedding and molecular characterisation of *Cryptosporidium* species in sheep across four states in Australia. *Vet. Parasitol.* 200, 50–58.
- Yang, R., Ying, J.L., Monis, P., Ryan, U., 2015. Molecular characterisation of *Cryptosporidium* and *Giardia* in cats (*Felis catus*) in Western Australia. *Exp. Parasitol.* 155, 13–18.
- Yang, X., Guo, Y., Xiao, L., Feng, Y., 2021. Molecular epidemiology of human cryptosporidiosis in low- and middle-income countries. *Clin. Microbiol. Rev.* 34, e00087-19.
- Yoshiuchi, R., Matsubayashi, M., Kimata, I., Furuya, M., Tani, H., Sasaki, K., 2010. Survey and molecular characterization of *Cryptosporidium* and *Giardia* spp. in owned companion animal, dogs and cats, in Japan. *Vet. Parasitol.* 174, 313–316.
- Yu, Z., Ruan, Y., Zhou, M., Chen, S., Zhang, Y., Wang, L., et al., 2018. Prevalence of intestinal parasites in companion dogs with diarrhea in Beijing, China, and genetic characteristics of *Giardia* and *Cryptosporidium* species. *Parasitol. Res.* 117, 35–43.
- Zablan, K., Melvin, G., Hayley, A., 2023. Older adult companion animal-owner wellbeing during the COVID-19 pandemic: A qualitative exploration. *Anthrozoös* 36, 237–256.
- Zahedi, A., Gofton, A.W., Greay, T., Monis, P., Oskam, C., Ball, A., et al., 2018. Profiling the diversity of *Cryptosporidium* species and genotypes in wastewater treatment plants in Australia using next generation sequencing. *Sci. Total Environ.* 644, 635–648.
- Zanzani, S.A., Gazzonis, A.L., Scarpa, P., Berrilli, F., Manfredi, M.T., 2014. Intestinal parasites of owned dogs and cats from metropolitan and micropolitan areas: Prevalence, zoonotic risks, and pet owner awareness in northern Italy. *BioMed Res. Int.* 2014, 696508.
- Zhang, Y., Zhong, Z., Deng, L., Wang, M., Li, W., Gong, C., et al., 2017. Detection and multilocus genotyping of *Giardia duodenalis* in dogs in Sichuan Province, China. *Parasite* 24, 31.
- Zhao, Z.Y., Li, M.H., Lyu, C., Meng, X.Z., Qin, Y.F., Yang, X.B., et al., 2022. Prevalence of *Giardia duodenalis* among dogs in China from 2001 to 2021: A systematic review and meta-analysis. *Foodb. Pathog. Dis.* 19, 179–191.
- Zheng, G., Alsarakibi, M., Liu, Y., Hu, W., Luo, Q., Tan, L., Li, G., 2014. Genotyping of *Giardia duodenalis* isolates from dogs in Guangdong, China based on multi-locus sequence. *Kor. J. Parasitol.* 52, 299–304.
- Zheng, G., Hu, W., Liu, Y., Luo, Q., Tan, L., Li, G., 2015. Occurrence and molecular identification of *Giardia duodenalis* from stray cats in Guangzhou, southern China. *Kor. J. Parasitol.* 53, 119–124.