REVIEW

OPEN ACCESS Check for updates

Tavlor & Francis

Taylor & Francis Group

The impact of aging-induced gut microbiome dysbiosis on dendritic cells and lung diseases

Jonaid Ahmad Malik, Mohammad Adeel Zafar, Taruna Lamba, Sidhanta Nanda, Mohammad Affan Khan, and Javed Naim Agrewala

Department of Biomedical Engineering, Indian Institute of Ropar, Rupnagar, Punjab, India

ABSTRACT

Aging is an inevitable natural process that impacts every individual, and understanding its effect on the gut microbiome and dendritic cell (DC) functionality in elderly subjects is crucial. DCs are vital antigen-presenting cells (APCs) that orchestrate the immune response, maintaining immune tolerance to self-antigens and bridging innate and adaptive immunity. With aging, there is a shift toward nonspecific innate immunity, resulting in a decline in adaptive immune responses. This alteration raises significant concerns about managing the health of an elderly population. However, the precise impact of aging and microbiome changes on DC function and their implications in lung-associated diseases remain relatively understudied. To illuminate this subject, we will discuss recent advancements in understanding the connections between aging, gut dysbiosis, DCs, and lung diseases. Emphasizing the key concepts linking age-related gut microbiome changes and DC functions, we will focus on their relevance to overall health and immune response in elderly individuals. This article aims to improve our understanding of the intricate relationship between aging, gut microbiome, and DCs, potentially benefiting the management of age-associated diseases and promoting healthy aging.

ARTICLE HISTORY

Received 18 August 2023 Revised 31 October 2023 Accepted 28 November 2023

KEYWORDS

Aging; gut microbiome; dendritic cells; lung diseases

Introduction

The microbiome consists of diverse microorganisms colonizing the skin, mucosal compartments, and the gut¹. The microbiome has been showing active involvement in the vital functions of the human body, such as immunity, circadian rhythmicity, metabolism, and nutritional responses². The human immune system is a complex network present in all tissues of the human body. The immune system plays a vital role in the host's defense against harmful exogenous and endogenous molecules for maintaining homeostasis. From an ecological perspective, the commensal microorganisms and mammals co-evolved towards homeostasis and beneficial relationships³. It is important to maintain the proper functioning of host immunity to prevent overexploitation of resources by commensals while maintaining immune tolerance against innocuous stimulation for healthy and beneficial relationships⁴. Environmental incursions such as diet, antibiotics, or changes in geography can alter the gut microbiome leading to impairment in the human-microbiome relationship. Diseases like rheumatoid arthritis (RA), metabolic syndrome, celiac diseases, malignancies, inflammatory bowel disease (IBD), and neurodegenerative disorders occur due to alterations in the microbiome population⁵. The cross-talk between the immune system and gut microbiome is dynamic, context-dependent, and complex⁵.

Recent findings have demonstrated that the agemediated inflamed microenvironment enhances autoimmune and inflammatory responses with decreased protective immune responses⁶. Further, there is a decline in immune responses to infection, vaccination, phagocytic activity, antigen capturing capacity, and presentation⁷. With the advancement of age, it has been suggested that there is the possibility of gaining nonspecific innate immunity with a weakening in adaptive immune responses⁸. The professional APCs called dendritic cells (DCs) are vital regulators of immune responses against infectious agents⁸. DCs have the property of both

CONTACT Javed Naim Agrewala iggrewala@iitrpr.ac.in Department of Biomedical Engineering, Indian Institute of Ropar, Rupnagar, Punjab 140001, India

^{© 2023} The Author(s). Published with license by Taylor & Francis Group, LLC.

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited. The terms on which this article has been published allow the posting of the Accepted Manuscript in a repository by the author(s) or with their consent.

the activation and the induction of tolerance to antigens (self/innocuous)^{8,9}. The DC maturation and activation depend on antigen uptake, processing, and the delivery of the pathogen's danger signals⁸. The differentiation and activation of naive T cells can only be done by DCs¹⁰. The DCs express the optimum levels of MHCs and costimulatory molecules to activate naive T cells¹¹. However, the tolerogenic DCs display a low range of costimulatory molecules and proinflammatory cytokines in contrast to immunogenic DCs. Tolerogenic DCs express high amounts of inhibitory molecules like CTLA-4, Lag-3, Tim-3, and PDL-1⁸. The DCs maintain tolerogenic behavior by inducing anergy and clonal deletion of T-cells and the generation of Tregs¹². Additionally, they are crucial in maintaining peripheral tolerance against self-antigens¹³.

Immunosenescence is an age-associated immune system dysfunction characterized by alterations in several aspects of the immunity, such as loss of adaptive immune diversity and thymic involution. The major characteristics of immunosenescence are the loss of the ability to recognize antigens, a decrease in memory T cells, and persistent low-grade inflammation called inflammaging. The other features of immunosenescence are phenotypical alterations in several immune cell types. Several viruses, such as Epstein-Barr and human cytomegalovirus, influence the immune system, resulting in immunosenescence⁸. Agingaffects differently various subsets of DCs (plasmacytoid DCs, myeloid DCs, follicular DCs). Plasmacytoid DCs (pDCs) are known for their role in initiating an immune response to viral infection and are reported to be majorly affected by aging^{14–} ¹⁶. The gut microbiota plays a significant role in

¹⁰. The gut microbiota plays a significant role in inflammation by producing inflammatory mediators¹⁷. Although there is not much evidence about the role of DCs in inflammaging, different subsets of DCs present in the gut have been reported to express pattern recognition receptors and respond to microbial products to produce inflammatory cytokines and induction of inflammatory cells^{18–21}.

DCs initiate adaptive immunity and process antigens in the draining lymph nodes (DLNs). In mouse models of respiratory virus infections, the DCs in the lung demonstrated decreased migration to DLNs, resulting in decline in T-cell responses because of the age-dependent increase in prostaglandin D2 in lungs^{8,22}. It is reported that during aging, there is an increased production of autoantibodies against self-antigens, and the DCs from the elderly revealed increased reactivity toward human DNA, causing the increase in the production of inflammatory markers and T cell proliferation²³. The state of chronic inflammation during aging might be an underlying cause of various diseases and deaths that are associated with aging. Although the mechanisms that mediate chronic inflammation associated with aging are not well understood²³. Future investigation should focus on understanding the chronic inflammatory states in elderly subjects.

The gut microbiome has been observed to be increasingly involved in the immune system's development, maturation, and maintenance²⁴. The microbiome induces peripheral tolerance by the induction of Tregs, tolerogenic DCs, and IgAsecreting B-cells⁵. Advancement in age is associated with alteration in the microbiome, provoking several autoimmune diseases²⁵. Chronic inflammation and immune dysregulation persist with the loss of Firmicutes and Bacteroides and an increase in the Proteobacteria population in the gut²⁶. It is still unclear how gut dysbiosis and loss of DC tolerance occur with the advancement of age. With a focus on the latest advancements, this article sheds light on the intricate interplay between aging, gut dysbiosis, and DCs, revealing their significant impact on overall health and immune responses.

Alteration in the microbiome with aging

Microbiome alteration occurs throughout human life and plays an important role in health and wellbeing²⁷. Microbiome changes with age, antibiotics use and the prevalence of diseases. DCs balance the activation and inhibition of immune responses, a unique feature of their functionality. With aging, a gradual loss of DCs tolerance is demonstrated by low expression of costimulatory molecules and proinflammatory cytokines, the deterioration in phagocytic activity, and the inability to induce Tregs, leading to increased chances of autoimmune and inflammatory disorders²⁸. The main reason behind the DC tolerance is the enhanced proinflammatory responses and activation of NF- κ B²⁸. However, the mechanism regarding age-associated DC tolerance is not clear. The studies are required to find out the reasons behind this phenomenon. Contradictory observations have been reported on the differences in the microbiome between young and old subjects, especially the presence of *Bifidobacterium*, *Ruminococcus*, and *Bacteroides*²⁷.

In addition to alterations in the microbiome, dietary factors play a pivotal role in maintaining the homeostasis of the immune system. Research has yielded conflicting findings regarding the impact of specific dietary components, such as carbohydrates, proteins, fatty acids, and phytochemicals, on the aging immune system. These components can exhibit both detrimental and beneficial effects on immune function. When it comes to fatty acids, their influence on pro-inflammatory markers in DCs hinges primarily on the type of fatty acid employed^{29,30}. Some studies have suggested that the elderly residing in Mediterranean (MED) regions often enjoy good health, which is frequently attributed to their adherence to MED diets. Further, it has been reported that a MED diet could potentially enhance immune responses in elderly individuals, particularly by positively affecting DC function²⁹. The study, involving 120 elderly subjects over the age of 60, revealed that intervention with the MED diet had the potential to mitigate age-associated increase in the secretion of resistin, a marker associated with aging²⁹. These findings emphasize the essential role of diet in immune system function and the importance of identifying suitable dietary choices for the elderly to prevent undesirable immune reactions. Further investigation into the potential beneficial effects of the MED diet in larger cohorts across various regions, especially among the elderly, is warranted.

Reports indicating the significant impact of diet on gut microbiota suggest that elderly individuals (aged >65) often exhibit altered dietary patterns, leading to decreased levels of *Firmicutes* and *Bifidobacterium*, as well as an increased abundance of Clostridium and *Bacteroides* ^{31,32}. However, it is important to note that these findings may not universally apply across different regions worldwide due to the substantial heterogeneity in dietary habits. This summary aims to outline the role of diets that promote gut health. For instance, the MED diet, primarily characterized by high consumption of fruits, vegetables, whole grains, and legumes, and reduced intake of meat, fish and lactose-rich products has been associated with beneficial effects on gut microbiota. It enhances gut diversity by increasing Bacteroides and Firmicutes while reducing *Clostridium*³³. An oriental diet rich in soy protein has also been reported to have a positive impact on the gut flora, particularly by promoting Bacteroides, Proteobacteria. Bifidobacterium and Enterococcus, while reducing the presence of Firmicutes and Lactococcus³⁴. Furthermore, recent studies have highlighted the influence of specific dietary components, such as fibers, saturated fats and polyphenols on gut microbiota composition. Fiber-rich foods, for have been found enrich instance, to Bifidobacterium and Lactobacillus while reducing pathogenic colonies³⁵. On the other hand, saturated fats tend to increase Firmicutes and Bacteroides³⁶. Proteobacteria but decrease Polyphenols, on the other hand, have been associated with a positive impact on Bifidobacterium abundance while decreasing Firmicutes and Clostridium ^{37.}

A study in Italy reported that with aging, microbiota а different population of Bacteroidaceae, Lachnospiraceae, and Ruminococcaceae families dominate, which usually decreases as age advances³⁸. A study from China reports a negative correlation between aging and the persistence of Faecalibacterium, Roseburia, and Coprococcus genera³⁹. The human body is densely populated with Archaea, Eukarya, Bacteria, and viruses, in which four bacterial phyla of Firmicutes, Proteobacteria, Bacteroides, and Actinobacteria contribute to 98% of the microorganisms²⁶.

The microbiota produces short-chain fatty acids (SCFA) from the undigestible fibers⁴⁰. The secondary bile acids (BAs) modulated by the gut microbiome play an essential role in host metabolism and energy balance, primarily through their interactions with nuclear receptors and G proteincoupled receptors (GPCRs). It has been reported that BAs also exert an influence on the composition of the gut microbiome. Moreover, emerging evidence suggests that BAs are of critical importance in the regulation of immune responses via their interactions with nuclear receptors and GPCRs. Notably, studies have proposed that the restoration of gut BAs can alleviate experimental autoimmune uveitis (EAU) in animal models, primarily through the suppression of NFκB-associated inflammatory cytokines in DCs⁴¹. Furthermore, investigations have demonstrated that DCs treated with BAs exhibit reduced levels of IL-12 and TNF-a in response to bacterial antigens and stimulation. Additionally, BAs induce the differentiation of IL-12 hypo-producing DCs from monocytes through the TGR5/cAMP signaling pathway⁴². These findings collectively suggest that the loss of BAs may lead to reduced DC tolerance and contribute to developing autoimmune and inflammatory diseases. Notably, the restoration of BAs has shown promising results, underscoring the requirement for optimal BA levels in maintaining proper immune system function⁴¹.

In addition to SCFAs, the gut microbiota abundantly produces various other metabolites, such as LPS, butyrate, propionate, secondary bile acids (e.g., deoxycholic acid), and protein metabolites (e.g., p-cresol sulfate, spermidine, spermine). These metabolites have been shown to impact the immune compartment, particularly DCs. Butyrate, propionate and deoxycholic acid have been demonstrated to inhibit DC development by downregulating the expression of costimulatory molecules and pro-inflammatory cytokines, reducing chemokine expression, and promoting the induction of Tr1 and Tregs⁴¹⁻⁴⁵. The LPS produced by the gut microbiota exhibits pleiotropic effects on the immune compartment. Gut-derived LPS has been reported to enhance the migratory capability of DCs, while p-cresol sulfate has been found to migration during impede DC airway inflammation^{46,47}. Another metabolite, spermidine, exerts an immune-suppressive effect by enhancing the activation of indoleamine 2,3-dioxygenase (IDO), p-Src, and FOXO3, while inhibiting NF- κ B activation when co-cultured with DCs^{48,49}.

Microbiota plays an essential role in food digestion throughout the alimentary canal. In addition, the digestive slurry gets mixed with microbiota for synthesizing, absorbing, and extracting metabolites and nutrients⁴⁰. The functions of SCFA are diverse such as regulating immune cells (activation of CD8⁺ T and differentiation of CD4⁺ T cells), controlling microbial functions, maintaining intestinal integrity, microbial energy source, and combating pathogens²⁶.

The studies on the microbiome can be classified into two categories i) alterations in gut microbiota composition with age; ii) changes in the microbiome along with aging disorders (Table 1). From these two categories, the major finding is literaturebased recognition of specific groups of taxa that demonstrate changes with aging, whether healthy or unhealthy⁵⁰.

Recent studies reported the relative differences in the population of Clostridium cluster XIVa, Escherichia, Shigella, Blautia, Faecalibacterium, Ruminococcaceae, Lachnospiraceae, and Erysipelotrichaceae in aged subjects of more than 100 years²⁶. Some investigations suggested that the gut gets enriched with Proteobacteria and Bacteroidetes and drops in Lactobacilli and Bifidobacteria as age advances. Understanding these age-related changes may inspire targeted interventions for healthy aging and disease prevention. It was revealed that the Bifidobacteria adhered more to the intestinal mucosa in infants and young adults than in old subjects⁵¹. Such studies reveal that the aged subjects have less ability to adhere and colonize Bifidobacteria. The intestinal mucosa may lose adhesion property (decay in adhesion proteins), which might be the reason behind the failure in colonization. The Bifidobacteria, which are more sensitive to changes in the intestinal mucosa adhesion property (decreased affinity), may be lost, although the exact mechanism is yet to be identified. Bifidobacteria use lipoteichoic acids (LTA) for adhesion to the intestinal mucosa⁵². The different Bifidobacteria groups might express LTA to different levels, and other microorganisms may compete with it for adhesion to the intestinal mucosa. Probiotic Bifidobacterium having good adhesive characteristics might help elderly subjects with their colonization in the gut. Akkermansia, Christensenellaceae, Oscillospira, and *Bifidobacterium* are health-associated bacteria³⁸.

Christensenellaceae and *Oscillospira* have been suggested to control leanness and decrease inflammatory disorders in elderly subjects⁵³. *Akkermansia muciniphila* has been demonstrated

Changes in the microo	Changes in the microbiome along with the aging disorder	Disorder	Country Ref	kererences
Increase	Decrease			
An increase in the population of <i>Lactobacillus</i> and Pathobionts An increase in the population of <i>Anaerotruncus</i> , <i>Coprobacillus</i> , and <i>Parabacteroides</i>	– Major SCFA-secreting bacteria and main gut microbiota	Reduced bone mass density Frailty	Ireland	50
Parabacterolaes, Anderotruncus, Loprobacillus Pathobionts	SCFA producer Faecalibacterium prausnitzii		United Kingdom	
Pathobionts, Ruminococcus, Coprobacillus	Prevotella copri, SCFA secreters, Coprococcus eutactus Alpha diversity		Korea United States	
Atopobiaceae, <i>Ruminococcus torques</i> <i>Ruminococcus</i>	Eubacterium, Gemella, Azospira. ruminatium Christensenellaceae, Barnesiellaceae		China Italy	
Pathobionts, Ruminococcus, Lactobacillus, Blautia	Prevotella, Odoribacter, Christensenellaceae, Barnesiella, Butyricimonas, Lachnospira, alpha diversity	Cognitive decline	United Kingdom	
NA NA	<i>Akkermasia</i> and Lentisphaerae <i>Akkermasia</i>		United States	
Lactobacillus and Pathobionts	Faecalibacterium, Roseburia, and Prevotella	Chronic kidney disease and frailty	Italy	
Coprobacillus, Eggerthella, Anaerotruncus, Megasphaera NA	Cetobacterium, Faecalibacterium, Lachnospiraceae, Prevotella Bilophila positive. Faecalibacterium prausnitzii	Reduced physical activity	United States Sweden	
Lactobacillus and Pathobionts	Roseburia, Faecalibacterium, and Prevotella	Chronic kidney disease and frailty	Italy	
Clostridioides difficile	Oscillospira	Cardiometabolic disease	Japan	
Pathobionts	Prevotella, core SCFA producers, Bifidobacterium, Odoribacter, Victivalis	Parkinson disease	Germany	
NA Collinsella Rifidoharterium Daransaiotella	Koseburia, Bindobacterium, and Lactobacilius Aktermancia: Eneralibarterium: Drevotella	Reduced bone mass density Obseity and metabolic synchrome	China Ireland	
Pathobionts	Butyrivibrio, core SCFA producers, and Adlercreutzia equolifaciens	Alzheimer disease	United States	
	Oscillospira, Christensenellaceae, Ruminococcaceae, Lachnospiraceae	Visceral fat deposition	United Kingdom	
	SCFA secreting bacteria, Bifidobacterium adolescentis, Butyrivibrio other SCFA secreters	Migraine	5	
Xenobiotic degradation-pathway-genes and Pathobionts	NA	Comorbidities (among long-living individuals)	China	
NA	Butyrivibrio crossotus, Alistipes sp., Bacteroides sp., Prevotella stercorea, Akkermansia muciniphila	Mortality (among centenarians)	China	
Pathobionts	Akkermansia	Comorbidity	United States	
NA		Progeria	Spain	

Table 1. Changes in the microbiome of aged subjects related to the aging disorder.

GUT MICROBES 😔 5

to control metabolic and inflammatory diseases, protect epithelial integrity, and support SCFAsecreting bacteria⁵⁴. SCFA and lactate produced by *Bifidobacterium* help reduce inflammatory microbes²⁶. A study on 371 subjects comprising newborn babies and centenarians revealed an aging-associated increase of *Oscillospira* compared to middle-aged adults and children. Some beneficial bacteria are lost while advancing with age progression⁵⁵. The microbiome alteration occurs in old age despite the influence of external environmental factors such as medications, sedentary lifestyles, diet, exercise, inter-individual variation, and geographical locations.

Influence of gut-dysbiosis on DCs

The DCs play an important role in tolerance by acting as a connecting bridge between adaptive and innate immunity⁷. The tolerogenic DCs control the Tregs and effector T cell responses⁵⁶. The function of DCs, particularly regulatory, gets impaired as age advances and with the loss of beneficial gut microbiota⁷. The unique characteristic of DCs is the induction of Th1 response in the presence of infection or Tregs in the absence of any infections. The CD103⁺ DCs in mice regulate the balance between Th1 cells and Tregs via the p38-MAPK signaling pathway during their differentiation from naïve T cells⁵⁷. The DCs may change the T cell phenotype by alteration of danger signals, recruitment of inflammatory cytokines, or changed con-The gut microbes use ditioning. LTA/ polysaccharides or DNA to cross-talk with the intestinal mucosa and produce antibacterial effects against pathogenic microorganisms by conjugated linoleic acid and bacteriocins⁵⁸.

Microorganisms can modulate the function of DCs by interacting with the toll-like receptors (TLRs) expressed by DCs⁵⁹. The different microorganisms network differently with DCs through distinct TLRs⁶⁰. The DCs (CD10⁺ CD11b⁺) expressing TLR5 cross-talk with bacterial flagellin to induce Th17 cells⁵⁹. The production and development of the Th17 cell anti-microbial peptide RegIII γ immune response depend on activating the DCs that produce IL-23 and IL-6¹⁸. The DCs can also be activated via TLR7 agonist to generate CD8⁺ T cell responses *in vivo*^{61,62}. The gut

microbiota co-evolved with the host, which renders the APCs to protect from infectious agents while maintaining self-tolerance with gut microorganisms. The best example of tolerance is that the DCs of the spleen produce low levels of IL-10, whereas the DCs of Peyer's patches secrete more IL-10 under similar conditions⁶³. There is a pivotal role of gut microorganisms in regulating the development and maturation of APCs, particularly DCs. Research studies revealed that in the germ-free (GF) mice, the DCs population decreased but not in the systemic circulation. Interestingly, gut colonization with Escherichia coli in the GF mice recruited the DCs to the intestines⁶⁴. DCs expressing CXCR1 and CD70 get activated through microbe-derived ATP, which induces Th17 cells⁶⁵.

The microbial metabolites influence the DC's development by altering bone marrow (BM) hematopoiesis, which changes the type/phenotype of DCs in the airways and lungs. The DC number in lymph nodes and spleen remains unchanged in germ-free mice compared to specific pathogenfree mice (SPFM)⁶⁶. The studies conclude that microbiota dysbiosis does not impact the steadystate generation of DCs. This does not suggest that the gut microbiota does not affect DCs⁶⁶. The treatment of SCFAs derived from microbiota has demonstrated the generation of DC progenitors and influences their development⁶⁶. From the findings, it is evident that gut dysbiosis has a negative influence on the DCs functions. The maintenance of DC tolerance is crucial for homeostasis. It is important to find microbial species and their metabolites that can recover the loss of DC tolerance. As the microbiome is diverse with many organisms, sophisticated methods should be inculcated to identify the species responsible for DC tolerance and maintenance. In the case of lung disorders such as COPD, infections, and cystic fibrosis, the play a crucial role. Saccharomyces, DCs Lactobacillus, and Bifidobacterium are the most commonly used probiotics as supplements in infectious diseases, IBD, and colon cancer. Probiotics act as immunomodulatory agents that activate the pathways in defense various respiratory disorders⁶⁷

The development of probiotic formulations will help in managing old-aged subjects suffering from lung disorders. The involvement of microbial metabolites and their functionality in various lung disorders is paramount such as SCFA metabolites have proven beneficial effects in treating several lung diseases. The susceptibility to lung disorders due to gut dysbiosis leads to a diminished production of SCFAs. With the advancement in research findings, SCFA producers could be formulated as probiotic supplements for managing lung disorders. Future investigations should focus on whether SCFA could be formulated for sustained release in lung disorders. The probiotics have shown promising results in stimulating the DC regulatory functions by targeting specific pathogen recognition receptors (PRR) and signaling pathways⁶⁸. Such results are not only for immune functionality but can be interventional in inflammatory bowel disease⁶⁸. Such studies show promise in potentially restoring the impaired tolerogenic behavior of DCs due to aging and gut dysbiosis. Given that aging and gut dysbiosis can trigger autoimmunity, further investigation into probiotic treatments is warranted, particularly in the context of autoimmune and lung disorders.

DCs dysfunction with age

Aging leads to inflammation, autoimmunity, immunodeficiency, infection susceptibility, and weak/non-response to vaccines¹⁵. This suggests that the immune response against infectious agents decreases during aging, and immune reactivity against endogenous molecules becomes more prominent because of increased inflammatory response. Further, the decrease in immune tolerance and loss of tissue integrity gives rise to new antigens and autoimmune immune reactivity through molecular mimicry⁶⁹. The continuous impairment in the functions of immune cells in aged subjects compared to young subjects are the major reason for morbidity and mortality.

With significant advancements in molecular and cellular mechanisms in science, there is still much to unravel concerning the changes associated with aging-mediated immune dysfunction and chronic inflammation. A recent study reported that age-related impairment of DC function leads to immune dysfunction, such as a loss of DC toler-ance and increased chronic inflammation⁷⁰. DCs express pathogen-sensing receptors like Toll-like

receptors (TLRs), C-type lectin receptors (CLRs), and NOD-like receptors (NLRs). A comparison between young and elderly populations revealed a decrease in the expression of TLR1, TLR2, and TLR8 in classical DCs (cDCs) among the elderly, while TLR2 expression remained unchanged. Additionally, TLR7 was downregulated, with no change observed in TLR9 expression in plasmacytoid DCs (pDCs) in aged individuals⁷¹. Another group discovered that both TLR7 and TLR9 expressions were decreased in pDCs, leading to a selective influence on the declining percentage of pDCs during healthy aging, while no association was found with any alteration in myeloid DCs (mDCs)⁷². Interestingly, a study involving children showed that the number of pDCs decreased by 2.5-fold in the first 10 years of life, while mDCs remained unaffected⁷³. These findings suggest that specific types of cells undergo alterations during aging, warranting further investigation to understand this phenomenon in greater depth⁷³. However, there have been contrasting results published regarding pDCs and aging. While some studies have demonstrated a decline in the number of pDCs with age^{72,74,75}, other investigations claim no change in pDC levels⁷⁶. It is essential to conduct more comprehensive research to unravel the intricacies of age-mediated immune dysfunction and its impact on DCs. Further studies should focus on understanding the underlying mechanisms responsible for the observed changes in different types of DCs during the aging process, potentially leading to new therapeutic interventions targeting agerelated immune dysfunction and inflammation.

The maintenance of self-tolerance is a crucial function of DCs. These cells are consistently exposed to antigens produced from damaged tissues and dead cells. Although the DCs take up these self-antigens, they remain inactive and express low levels of costimulatory markers. Consequently, they fail to present the antigens to T cells²⁸. Presenting self-antigens to T cells without costimulatory signals results in T cell anergy, a state of unresponsiveness. However, during inflammation or tissue injury, the DCs may become activated and present self-antigens along with costimulatory signals to T cells, leading to autoimmunity. Aging contributes to impaired function, loss of homeostasis, and increased susceptibility to death. Aging

reduces DC tolerance, leading to gut dysbiosis and inflammation. This impacts mental well-being and heart health and causes tissue damage. Lung and autoimmune disorders also become more common with aging and gut dysbiosis. Understanding and addressing these changes is vital for healthy aging. The elevated basal level of NF- κ B in the DCs of the elderly indicates that these DCs are in an activated state²⁸. Similarly, we observed an increased level of NF- κ B in the DCs of old mice compared to young mice⁷. After apoptosis, self-DNA is released and phagocytosed by DCs. In the elderly, DCs have a reduced capacity to uptake apoptotic cells, leading to defective clearance and immune responses instead of tolerance against self-antigens²⁸.

During viral pneumonia, DCs fail to migrate into aged lungs and from lungs to lymph nodes, rendering them unable to prime naïve T cells against the influenza virus⁷⁷. Reports suggest agerelated defects in DCs presenting antigens via MHCI to prime CD8⁺ T cells, with diminished production of IL-1 β and inflammasome activation⁷⁸. The impact of antibiotics on lung microbiota is not well-studied, although it has been established that antibiotics alter the microbiome and lung functions in cystic fibrosis⁷⁹. Some studies indicate that antibiotics increase the death rate in animals infected with S. pneumoniae and Influenza A virus⁸⁰. The influence of antibiotics and age-related gut dysbiosis on DC function requires thorough examination, and appropriate probiotic preparations should be investigated in disease models. Supplementation of depleted beneficial bacteria with age through diet is essential for maintaining homeostasis. Recent research has shown that bacteriotherapy holds promise against frailty and unhealthy aging⁸¹. External factors such as drug therapy, exercise, and diet play crucial roles in healthy aging and increasing life expectancy⁸¹. Moreover, the intestinal microbiota is considered a major factor in the anti-aging process⁸¹. Future studies should focus on elucidating the mechanisms involved in the anti-aging process for a better understanding. The elderly population is an important part of society and is more susceptible to various diseases. Therefore, the scientific community needs to investigate better therapies that can improve the quality of life during the later stages of life.

The modulation in the function of DCs in the aged population

Dendritic Cells (DCs) in the infected aged community have been found to weakly induce Tregs⁸². Aged individuals are more susceptible to infections caused by Chlamydia, pneumonia and influenza⁸³. The TLR function of pDCs and mDCs becomes impaired with age more than 65 years compared to younger individuals of age between 21-30 years ⁷¹. This impairment is associated with a poor Ab response against influenza viruses⁸³. Moreover, aged individuals are more prone to chronic obstructive pulmonary diseases such as bronchitis, emphysema, and asthma. Reduced DC tolerance in the airways allows for the invasion of pathogenic agents and inflammation, increasing susceptibility to lung-associated diseases. Unstimulated DCs from aged subjects exhibit increased expression of cytokines and costimulatory molecules, resulting in heightened epithelial permeability²⁸. As a consequence, the functionality of other cells is compromised, leading to the activation of T cells without any infection and similar reactions may occur in the skin and gut. Infections on the airway, gut, and skin surface are more common in aged subjects due to the loss of integrity of cell-to-cell junctions⁸⁴.

The gut microbiome synthesizes Short-Chain Fatty Acids (SCFA), such as butyrate, acetate, and propionate, which are crucial metabolites for normal body homeostasis. The percentage of SCFA declines in aged subjects compared to young subjects⁸⁵. SCFA induces Tregs, prevents DC activation, and maintains DC tolerance in the gut⁸⁶. DCs sense SCFA metabolites through special receptors like GPCRs (GPR109A, GPR41, and GPR43), which have different affinities and specificity toward particular metabolites⁸⁶. Several immune cells, including Tregs, neutrophils, and macrophages, express GPR43 and GPR109A, which play roles in regulating microbiome homeostasis by binding to butyrate and nicotinic acid, acting as anti-inflammatory molecules⁸⁷. Studies also suggest that propionate and GPR41, a receptor for SCFA, play a crucial role in generating DC and macrophage precursors⁸⁸.

Infections caused by *Clostridium difficile* and *Helicobacter pylori* in the gut are dangerous in

aged subjects, leading to hospitalization⁸⁹. As people age advances, microbiome alteration occurs in parallel, with an increase in *Enterobacteriaceae*, Gram-negative, and other pathogenic bacteria in the gut⁸⁹. The LPS secreted by Gram-negative bacteria activates APCs like DCs and macrophages, leading to inflammation⁷⁰. The increase in LPS-secreting Gram-negative bacteria is one of the reasons behind inflammation in aged subjects. Therefore, appropriate probiotics should be supplemented in elderly subjects to balance the healthy gut microbiome population and avoid inflammatory reactions.

Infections such as SARS-CoV-2 and influenza impair APCs, TLRs, cytokines, and T cells, resulting in poor immune responses⁹⁰. Age-mediated gut dysbiosis weakens the function of DCs and T cells, making old-aged subjects susceptible to lung infections. The supplement of probiotics regulates immune responses via modulation of the TLR signaling pathway, thereby regulating NK cells, DCs, and Th1 and Th2 immune responses⁹⁰. Probiotics have demonstrated preventive effects on upper respiratory infections, improved outcomes, and reduced disease duration and severity in children and adults⁹¹.

Numerous probiotics have been investigated in clinical trials in elderly subjects to improve immune responses against influenza vaccination and infections. L. plantarum (CECT7315/7316) has shown an immuno-stimulating effect and might improve immune responses against the influenza virus in old subjects⁹². Additionally, two randomized clinical trials have revealed that daily administration of specific probiotic preparations enhances specific Ab responses against influenza vaccination in old-aged subjects (70 years old), suggesting potential beneficial effects for managing lung infections⁹³. However, evidence suggests that not all probiotic preparations may improve protection against respiratory infections⁹⁰. Some investigations have shown no significant effect of probiotics in decreasing respiratory infections⁹⁴. For instance, a study focused on 737 healthy old reported that administration subjects of Lactobacillus casei for 176 days did not improve respiratory symptoms or show positive immune responses in vaccinated individuals compared to controls⁹⁵. Therefore, it is crucial to identify

bacterial species that can reverse regulatory DC functions and prevent the occurrence of diseases in aged subjects.

Molecular mechanisms in DC dysfunction during aging

Molecular mechanisms underlying DC dysfunction during aging are not fully understood, but NF-KB signaling plays a crucial role. NF-KB, the master regulator of inflammation, is implicated in various diseases, including autoimmunity and inflammatory disorders^{96,97}. Overexpression of NF-κB subunit RelA/p65 induces a senescent phenotype in cells, while low expression delays age-associated diseases in *in vivo* models (Figure 1(a)^{98,99}. Our group reported that age and gut dysbiosis lead to increased p65 phosphorylation in murine models, and Lactobacillus planetarium supplementation can prevent overt NF-KB activation, which is associated with DC maturation⁷. Other studies also demonstrate increased basal activation of NF-KB/ PI3K in monocyte derived dendritic cells (MODC) and circulatory DCs^{23,71}. The exact cause of DC dysfunction, whether external or intrinsic, remains unclear. Age-associated increases in prostaglandins and pro-inflammatory cytokines in circulation can trigger DC maturation and cytokine secretion even in the absence of disease^{100,101}. Additionally, an increase in bone marrow fat levels in old age may contribute to cytokine release, further promoting DC maturation and activation^{100,101}.

Epigenetic modifications, including DNA methylation and chromatin alterations, can also impair DCs during the aging process. In older individuals, inhibitory H3K9 histone proteins exhibit an increased binding affinity to IFN type I and III promoters in myeloid DCs. This leads to a reduction in IFNs production, particularly in the context of viral infections such as influenza¹⁰². Further investigation is essential to delve into intrinsic alterations within DCs in elderly subjects. This exploration should encompass the examination of factors like miRNAs, epigenetic markers, and various signaling pathways (such as mTOR/PI3K/PTEN/Hedgehog/ NrF2/Wnt) to gain a comprehensive understanding of molecular-level changes.

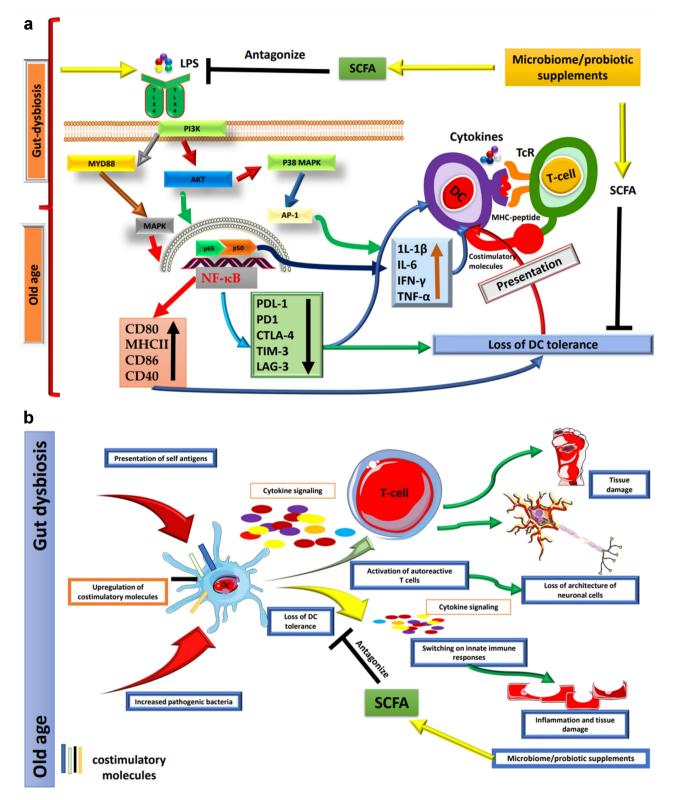


Figure 1. (a) Age-related changes in the gut microbiota affect DCs. As individuals progress through the aging process, significant alterations occur within the gut microbiome. These changes are characterized by an augmented presence of Gram-negative bacteria, which, in turn, leads to an increased secretion of LPS. The LPS exerts its influence via TLR4 on DCs, thus initiating a complex signaling cascade. This cascade involves critical components such as MYD88, AKT, PI3K, and MAPK pathways, ultimately culminating in the activation of the transcription factor p65 and a subsequent elevation in the basal activity of NF-kB. The heightened activation of NF-kB within DCs results in a noteworthy upregulation of proinflammatory cytokines and the augmentation of essential costimulatory molecules, including CD80, CD86, MHCII, and CD40. Concurrently, this activation leads to the downregulation of immune checkpoint proteins. These activated DCs then proceed to present antigens bound to MHC molecules and engage in interactions with T cells

Microbial metabolites influence DCs through various pathways, altering bone marrow hematopoiesis and influencing DC development in different body parts⁸⁸. Probiotics, like the VSL3 preparation containing specific bacterial strains, have shown promising results in ulcerative colitis patients¹⁰³. The protective activity was mediated through inhibition of the PI3K/Akt and NF-κB axis, leading to reduced pro-inflammatory cytokines and increased anti-inflammatory cytokines in colonic tissue¹⁰⁴.

Probiotics have demonstrated the potential to restore DC functionality. The VSL3 probiotic preparation showed protective effects against wildtype colitis but not in TLR9 knockout mice, indicating that gut microbiota acts through specific TLR receptors on DCs or other APCs¹⁰⁵. Bacterial metabolites and pathogen-associated molecular patterns (PAMPs) often act via TLR pathways, which are interlinked with inflammatory signaling cascades. Dysbiosis in the gut increases inflammatory signaling, resulting in the loss of immune tolerance in DCs. SCFA metabolites, on the other hand, can antagonize TLR4 signaling, offering probable future therapies for various inflammatory conditions. Research should now focus on elucidating the pathways of microbiome metabolites and evaluating their therapeutic potential against lungassociated diseases.

Influence of gut microbiota dysbiosis on the health of aged subjects

Hippocrates (460–370 BCE), the father of modern medicine, proposed that "All Diseases Begin in the Gut," a concept still relevant today¹⁰⁶. Gut dysbiosis, a natural occurrence in old subjects, can lead to various serious consequences, including loss of DC tolerance, susceptibility to infections, reduced bone mass index, neurodegenerative diseases, autoimmune diseases, and inflammatory diseases²⁶.

Clinical studies using fecal samples have investigated gut dysbiosis in young (28-46 years) and old (>65 years) subjects. In old subjects, the phylum Firmicutes was dominant at 40%, and phylum Bacteroidetes was at 57%, whereas in young subjects, Firmicutes was 51% dominant, and Bacteroidetes was 41%¹⁰⁶. Another investigation reported a decline in the Firmicutes/Bacteroidetes ratio from 10.9 to 0.6 between young (25-45 years) and old (70-90 years) subjects¹⁰⁷. The ratio of Firmicutes and Bacteroidetes was higher (9-fold) in old mice/rats than in young animals, with the old animals demonstrating anxiety behaviors and cognitive impairment¹⁰⁸. The gut-brain axis, linking the gut microbiome to brain functioning, has gained attention, revealing links between the gut microbiome and various body parts in physiological and pathological aspects. Evidence suggests that gut microbiome dysbiosis

through the TCR, thus facilitating immune responses. This shift in the equilibrium of immune signaling makes DCs more inclined to mount vigorous inflammatory responses. Consequently, these activated DCs evolve into proficient antigen captors and presenters to T cells, thereby promoting immune responses against potential threats. However, this heightened state of immune activation also brings about a potential loss of DC tolerance, rendering the immune system more susceptible to autoimmune reactions and the development of chronic inflammatory diseases. The maintenance of a harmonious and balanced immune response is paramount for healthy aging, as the dysregulation of DC function can significantly contribute to a spectrum of age-related pathologies. One promising approach to ameliorate the adverse impacts of gut dysbiosis on DCs involves the supplementation of SCFAs. SCFAs are microbial metabolites generated through the fermentation of dietary fibers within the gut. These compounds have been demonstrated to antagonize TLR inflammatory pathways, thereby offering a potential means to safeguard DC tolerance and sustain a more balanced and harmonious immune response. (b) Aging induces various changes in the human body. Throughout the aging process, the human body undergoes a multitude of transformations, encompassing significant shifts within the gut microbiome. These alterations wield a profound influence on the immune system, with a particular emphasis on DCs, pivotal regulators of immune tolerance. DCs play a critical role in the recognition and presentation of antigens to T cells, thus regulating immune responses. The sustained activation of DCs in the elderly populace can precipitate adverse consequences, given that these cells progressively lose their ability to uphold immune tolerance toward self-antigens. Consequently, this loss of tolerance manifests as chronic inflammation, autoimmunity, and tissue damage. The unrestrained expression of costimulatory molecules and the presentation of self-antigens by DCs to autoreactive T cells usher in a disarray of immune responses within aging individuals, fostering inflammation, disruption of neuronal cell architecture, and tissue damage associated autoimmune diseases. It is worth noting that age-related maladies can potentially be ameliorated through the judicious supplementation of probiotics and SCFAs. Probiotics, being beneficial microorganisms, can reinstate equilibrium within the gut microbiome and foster the tolerogenic behavior of DCs. By modulating the functions of DCs, probiotics hold the promise of mitigating inflammation and enhancing immune tolerance, thereby opening a promising avenue for the management of age-related health conditions. The artwork used in this figure was adapted from Servier Medical Art (http:// servier.com/Powerpoint-image-bank). Servier Medical Art by Servier is licensed under aCreative Commons Attribution 3.0 Unported License.

can influence brain functions and increase the risk of neurological diseases like Alzheimer's disease, substance use disorders (SUDs), COPD, and Parkinson's disease^{109,110}.

Aging leads to changes in the gut microbiome, increasing Gram-negative bacteria and LPS secretion. This activates DCs through the TLR4 pathway, leading to NF-kB activation and proinflammatory cytokine upregulation. DCs become more efficient at presenting antigens to T cells but may lose tolerance, promoting autoimmune reactions and inflammation. SCFA supplementation could help maintain a balanced immune response and mitigate the effects of gut dysbiosis on DCs (Figure 1A). Gut dysbiosis is also being studied in the context of inflammatory bowel disease (IBD). Although pathogenic bacteria were initially proposed as the cause of IBD, current research suggests that gut dysbiosis may play a major role in its pathogenesis¹¹¹. The rising incidence of cancer in older adults may be linked to gut dysbiosis. By 2035, there is expected to be a 60% increase in cancer cases in older adults, with more than 14 million cases¹¹². Some bacteria, such as Bifidobacterium pseudopodium, Lactobacillus johnsonii, and Olsenella species, are being explored for their potential in treating colorectal cancer (CRC). Probiotics are also under investigation in clinical trials for managing CRC¹¹³. Probiotics and immune checkpoint inhibitors are being used to enhance therapeutic potential against various types of cancers.

Aging impacts the gut microbiome and DCs, thereby affecting immune tolerance. Persistent DC activation leads to chronic inflammation and autoimmune diseases. Probiotics and SCFAs offer potential solutions to restore gut balance and promote DC tolerance, managing age-related conditions (Figure 1B). Age-associated lung diseases can be managed with probiotics, exercise, proper diet, and bacterial-associated metabolites. Healthy aging is essential to prevent age-associated diseases, as the elderly population often relies on various therapeutics. Lifestyle factors such as diet, exercise, and water intake play a significant role in disease prevention in old age. Importantly, managing ageassociated diseases requires a proactive approach to maintain a healthy lifestyle and prevent the occurrence of these conditions.

Role of aging in the gut-lung axis

During aging, physiological changes in the lung lead to a decline in lung function capacity and homeostasis. Lung-associated diseases such as tuberculosis, cystic fibrosis, pneumonia, chronic obstructive pulmonary disease (COPD), and pulmonary fibrosis become more prevalent with age¹¹⁴. The occurrence of lung disease during aging is influenced by immunological, cellular, and physiological changes. Understanding ageassociated alterations in lung physiology is essential for comprehending the healthy aging process¹¹⁴. Healthy lung status is considered a predictor of aging, with subjects having healthy lungs living longer compared to those with poor lung functionality who are more prone to diseases such as cardiovascular diseases, diabetes, and cognitive decline⁶⁴.

The nose, gut, and throat microbiota develop coordinately at the young age¹¹⁵. The gastrointestinal tract (GIT) and respiratory tract share a common entry of microbes in the oral cavity, leading to significant overlap between the lung and gut ecosystem¹¹⁶. The Human Microbiome Project data revealed that 45% of the microbiome population from stools overlaps with the oral cavity¹¹⁷. Aging results in a natural decline in lung functions due to genetic and environmental factors. Several hallmarks contribute to changes in the lung aging axis, including genomic instability, loss of proteostasis, epigenetic changes, telomere attrition, mitochondrial dysfunction, stem cell exhaustion, altered intracellular communication, deregulated nutrient sensing, and dysregulation of the extracellular matrix (ECM)¹¹⁸. These changes lead to physiological and structural alterations in the lung. Major changes during aging include abnormalities in the structure of cilia, mucins, and antioxidants in the lining of the epithelium, as well as reduced lung muscle strength in the diaphragm, resulting in a weakening in lung functionality and increased susceptibility to pulmonary diseases¹¹⁴. In aged lungs, the size increases due to ECM alterations, leading to increased expiratory lung volume (ELV) and decreased elastic recoil¹¹⁹. Previous studies have indicated that gut dysbiosis is linked to acute and chronic lung-associated

diseases¹¹⁸. Recent research reported that 10–18% of SARS-CoV-2 subjects experienced gastrointestinal symptoms, and older people were at an increased risk of developing severe illness¹²⁰. In old-aged subjects, increased epithelium permeability in the lungs and gut suggests impairment in tissue functions¹²¹.

The presence of high bacterial diversity is a hallmark of a healthy gut microbiota¹²². Reports indicate that subjects with chronic diseases have an increased abundance of harmful bacteria such as Escherichia and Clostridium species, along with a decrease in commensal bacteria¹²³. The mechanisms behind these bacterial population changes are not well understood and require further investigation. For asthma subjects, metagenomic analysis of stool samples revealed decreased microbiome diversity compared to healthy subjects in the Kingdom¹²⁴. Additionally, United SCFAproducing bacteria like Coprococcus eutactus and Faecalibacterium prausnitzii were reduced, while Eggerthella lenta and Clostridium species increased in asthma subjects¹²⁴.

In lung diseases like cystic fibrosis (CF), gut dysbiosis and intestinal inflammation are frequently reported due to antibiotic use in both young and old subjects¹²⁵. Antibiotic use can disrupt the gut microbiome and lead to various ailments. The gutlung axis has been demonstrated in CF with pathogenic colonization in the lungs and a decrease in the beneficial microbiome population. Metagenomics that analysis revealed in CF subjects, Propionibacterium acnes, Clostridium difficile, and Staphylococcus spp were more abundant in stools than in healthy subjects, while populations of beneficial bacteria like Roseburia, Anaerostipes, Blautia, Pseudobutyrivibrio, Faecalibacterium, Subdoligranulum, Streptococcus, Dorea, and Coprococcus species were depleted¹²⁵. Considering the microbiota while prescribing antibiotics in clinics is crucial, and certain diets can also impact the lung microbiome, with some spiced foods having negative effects. Major causes of diseases are lack of exercise and improper diet intake. Maintaining a healthy lifestyle is essential to support the body's natural defenses and promote overall well-being.

Fecal Microbial Transplantation (FMT) has been shown to increase the transcriptional activity of

NF-κB in the lungs with lung injury in mouse models, causing an increase in TNF-α, IL-1β, and IL-6 expression, as reported in some studies¹²⁶. However, the transplantation of fecal bacteria prevents the transcriptional activity of NF-κB and decreases the secretion of proinflammatory cytokines¹²⁷. Some studies have demonstrated that FMT restores the gut microbiome diversity and abundance in the mouse model with pneumonia, prevents tissue damage and inflammation, and maintains the balance between Tregs and Th17 cells^{126,127}.

The diet and exercise are very important in maintaining homeostasis of body functions. Diet has been found to influence the gut microbiome. The nutrition rich in fibers, like the MED diet, promotes a healthy gut microbiome and releases butyrate¹²⁸. The benefits of a good diet and a healthy gut microbiome help improve lung functions in diseased conditions such as COPD, asthma, or chronic inflammatory conditions¹²⁸. Some clinical studies have shown a significant and independent correlation between low exercise and low adherence to MED diets and changed pulmonary functional patterns¹²⁹.

Even though probiotics were quite a success, a certain drawback has emerged, e.g., transferring antibiotic-resistant genes to other microorganisms in the gut to produce harmful metabolites. Hence, exploring other alternatives like prebiotics, synbiotics (a mixture of probiotics and prebiotics), and fecal FMT becomes necessary. Applying prebiotics (fructans, oligosaccharides, fructooligosaccharides, galactooligosaccharides, lactose) to enhance gut diversity has recently been explored in altering the immune compartment and their implications for various diseases¹³⁰. Using prebiotics, synbiotics, and FMT to enrich gut species (e.g., Bifidobacterium) responsible for DC health may be an interesting approach to overcoming the problem of DCs associated with aging.

Role DCs in age-associated lung diseases

The epithelial lining covering the nasal passages and airways acts as a physical barrier and regulates immune responses against pathogens. DCs and airway epithelial cells (AECs) closely interact,

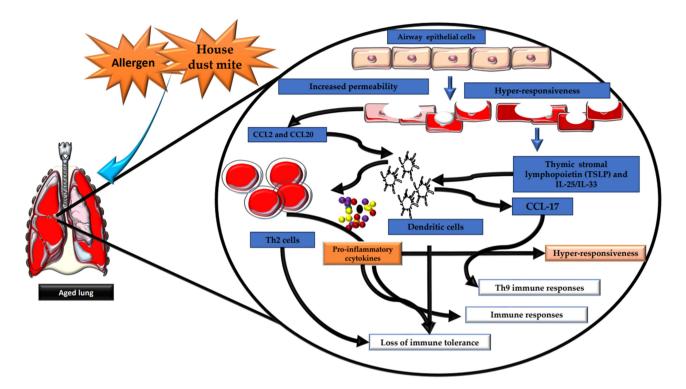


Figure 2. Aging induces significant changes in the structural and functional integrity of the airways. Aging is linked to heightened airway epithelium permeability, rendering it more susceptible to environmental challenges and allergens. This increased vulnerability can induce elevated inflammation and immune reactions within the airways. Airway epithelial cells (AECs) play a pivotal role in sensing and responding to allergens and environmental stimuli. Following allergen exposure, AECs release cytokines, including IL-25, IL-33, and TSLP, which activate DCs. DCs present antigens to T cells, thus initiating and modulating immune responses. In the context of aging, DC activation by AEC-released cytokines can lead to the generation of proinflammatory cytokines, contributing to chronic airway inflammation. Moreover, AECs release chemokines such as CCL2 and CCL20, further activating DCs and promoting the recruitment of inflammatory cells to the airways. This sequence of events can result in the production of CCL17 by DCs, associated with the recruitment and activation of Th2 cells and Th9 cells involved in allergic responses. Dysregulated immune responses in aging airways may lead to diminished immune tolerance, increasing susceptibility to lung tissue damage and heightened antigen hyperresponsiveness, which manifest to asthma or chronic obstructive pulmonary disease (COPD) in older individuals.The artwork used in this figure was adapted from Servier Medical Art (http://servier.com/Powerpoint-image-bank). Servier Medical Art by Servier is licensed under aCreative Commons Attribution 3.0 Unported License.

influencing each other's functions¹³¹. DCs release interferons during viral infections, enhancing MHCI expression on AECs to combat virusinfected cells¹³². Immune cells infiltrate infection sites through tight epithelial junctions, facilitated by DC-released pro-inflammatory cytokines¹³³. AECs also impact DC function, expressing PRR for allergens and initiating pathogen-host interactions¹³⁴.

LPS-mediated TLR4 signaling on AECs induces lung DC migration to mediastinal nodes¹³⁴. House dust mite (HDM) in mouse models triggers AECs to release CCL2 and CCL20, attracting immature DCs/monocytes to the lungs¹³¹. The AEC-DC interaction is vital for Th2 immune responses against allergens¹³⁵. AECs release TSLP and IL- 25/IL-33 upon allergen exposure, acting on DCs through OX40-OX40L cross-talk to elicit Th2 immune responses¹³⁵. IL-25 induces DCs to release CCL-17, contributing to Th9-mediated immune responses during allergic inflammation¹³⁶.

Aging increases airway epithelium permeability, heightening susceptibility to environmental triggers and allergens, leading to inflammation and immune responses. AECs detect and respond to allergens by releasing cytokines that activate DCs. In the elderly, activated DCs produce proinflammatory cytokines, contributing to chronic airway inflammation. Dysregulated immune responses in aging airways result in a loss of immune tolerance, increasing susceptibility to lung tissue damage and hyperresponsiveness to antigens, potentially causing

asthma or COPD (Figure 2(a)). Aging affects AEC and DC functions. In healthy-aged subjects, nasal epithelium exhibits reduced ciliary beat frequency and microtubular disarrangements¹³⁷. Older adults have difficulty clearing teflon particles from small airways, taking over 21 days, leading to respiratory symptoms¹³⁸. Age-related changes in the microbiome influence metabolite concentrations that affect immune cells, leading to increased inflammation. Understanding how these metabolites regulate DC function to maintain immune tolerance is crucial. In aged subjects, DCs often exhibit inflammatory responses to microbial metabolites¹³¹. Further research should investigate whether the DCs of aged subjects become immune to gut microbial metabolites or the microbiome, leading to a potential reversal of their immune system functions.

Microbiome interventions in elderly care

Our recent study revealed that DCs from old mice (DC^{old}) mice and DCs from dysbiotic mice (DC^{dys}) exhibited diminished tolerance compared to DC from young mice (DC^{young}). Specifically, DC^{old} and DC^{dys} displayed reduced capacity to induce Treg generation and regulate CD4 T cell activation. We observed a notable decrease in the prevalence of the Lactobacillus genus in the gut of aged mice. To explore potential therapeutic approaches, we introduced Lactobacillus plantarum into the gut of elderly mice. This intervention successfully restored the tolerogenic function of DCs by modifying inflammatory and metabolic pathways. This novel finding sheds light on the impact of agerelated gut dysbiosis on DC tolerance. Moreover, it offers promising insights into potential therapeutic strategies for addressing age-associated disorders through the use of *Lactobacillus plantarum*⁷.

A microbial mixture called IRT5, comprising *Bifidobacterium iridium, Streptococcus thermophilus, L. casei, L. reuteri*, and *L. acidophilus*, has shown promising results in experimental models of various diseases. In the context of rheumatoid arthritis, atopic dermatitis, and IBD, the administration of IRT5 increased the levels of IL-10, TGF- β , and IDO in DCs, leading to the induction of Tregs¹³⁹. Additionally, IRT5 was found to be effective in reducing body trembling and weight loss in the myasthenia gravis model by blocking T cell-

dependent B cell Ab responses against acetylcholine receptors (AchR)¹⁴⁰. Furthermore, IRT5 supplementation resulted in decreased levels of AChRspecific IgG Abs, lymphocyte proliferation, and pro-inflammatory cytokines IL-17, IL-6, TNF-a and IFN-y. DCs treated with IRT5 showed reduced production of pro-inflammatory cytokines in AChR-specific lymphocytes, accompanied by upregulation of arginase-1, RA-producing gene aldh1a2, IL-10 and TGF-B, indicating enhanced DC tolerance and induction of Tregs, which demonstrated protective effects^{60,140}. Probiotics are currently under investigation for various diseases and have shown promising outcomes in preclinical and clinical studies. Diets rich in Lactobacillus have been associated with a reduced incidence of colon cancer¹⁴¹. Moreover, our laboratory's research demonstrated that replenishing Lactobacillus plantarum restored tolerogenic behavior in DCs of aged and dysbiotic mice⁷. Probiotics are also gaining interest for their potential role in altering tumor apoptosis and proliferation, offering promising alternatives to radiotherapy and chemotherapy¹⁴².

Conclusion

DCs play a critical role in maintaining self-antigen tolerance, and their dysfunction significantly impacts the health of elderly individuals. With advancing age, the gut microbiome undergoes changes that profoundly affect the immune system, particularly impairing DC function. This agemediated dysbiosis creates an inflamed microenvironment, increasing the risk of autoimmune and inflammatory responses. The immune system undergoes a series of changes during aging, shifting the balance toward innate immune responses rather than adaptive ones. Intestinal DCs are particularly affected by these aging-related alterations. Cross-talk between the gut microbiome and intestinal DCs is crucial in maintaining tolerogenic behavior. Studies are exploring probiotic preparations and immune checkpoint inhibitors for cancer management, holding promise for future therapeutic approaches. Further research is needed to address better age-related disorders like COPD, cystic fibrosis, lung infections, Alzheimer's disease, and Parkinson's disease.

Although we have highlighted the impact of aging and gut dysbiosis on the immune system, especially focusing on DCs and the gutlung axis, much remains to be explored. Unraveling the underlying mechanisms responsible for the loss of DC tolerance, both intrinsic and extrinsic, warrants investigation in preclinical and clinical models. Probiotics have emerged as significant players and have demonstrated promising results in preclinical and clinical studies. Future research should concentrate on developing optimal compositions of beneficial microbes that could aid elderly subjects in managing pathophysiological conditions. Strategies for monitoring gut health could prove valuable in detecting and addressing health alterations in elderly individuals at an early stage. Given the vulnerability of older subjects to infections and chronic diseases like age-related cognitive impairments, addressing their healthcare needs becomes a crucial aspect in managing their quality of life.

Future prospects of early diagnosis of diseases and delaying aging by exploiting gut microbiota

Aging exerts a profound influence on the gut microbiota composition, consequently impacting immune system function. To comprehensively understand age-related changes in the gut microbiome, systematic studies are imperative. Unraveling the complicated relationship between specific gut microbes and their association with disease susceptibility or prevention could provide valuable insights for remedial interventions. Of particular interest is exploring the correlation between specific gut microbes and their role in disease susceptibility.

Numerous distinct hallmarks, encompassing immunosenescence, genomic instability, inflammaging, altered intracellular communication, epigenetic modifications, and telomere attrition, collectively contribute to the deterioration of immune cell tolerance¹⁴³. This complex cascade culminates in the establishment of a chronic inflammatory state, which plays a pivotal role in the process of lung aging (Figure 3). While the

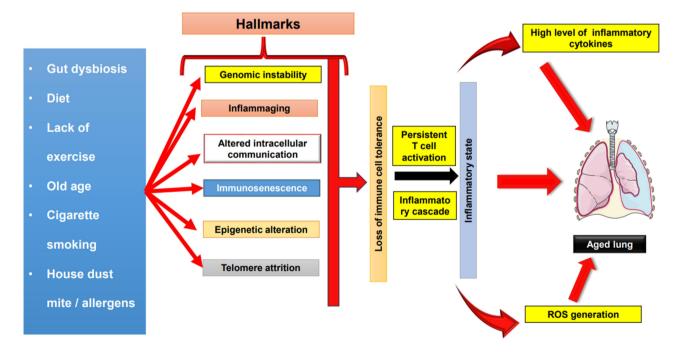


Figure 3. Role of inflammation and other hallmarks in driving lung aging. Numerous factors, including gut dysbiosis, aging, cigarette smoking, exposure to allergens, dietary patterns, and sedentary lifestyles, collectively contribute to a spectrum of immunological phenomena. These encompass immunosenescence, genomic instability, inflammaging, alterations in intracellular communication, epigenetic modifications, and telomere attrition. This intricate web of influences invariably results in the erosion of immune cell tolerance. The culmination of these hallmarks further translates into the loss of tolerance among DCs subsequently giving rise to sustained T cell activation. Consequently, triggers an augmented production of pro-inflammatory cytokines and ROS, ultimately fostering an inflammatory state. The consequences of this chronic inflammation, in the context of the respiratory system, precipitate the phenomenon known as lung aging. The figure is party adapted¹⁴³. The artwork used in this figure was adapted from Servier Medical Art (http://servier.com/Powerpoint-image-bank). Servier Medical Art by Servier is licensed under aCreative Commons Attribution 3.0 Unported Licen.

phenomenon of lung aging is multifaceted and characterized by various contributing factors, the underlying mechanism remains enigmatic. Future investigations are imperative to elucidate the precise roles played by DCs and other immune cells in mediating this inflammatory state and their involvement in the overall process of lung aging.

The presence or absence of certain bacteria may serve as indicative markers of disease protection or vulnerability, enabling early disease diagnosis through gut microbiome analysis. However, the cultivation of non-culturable bacteria remains a significant challenge, underscoring the need for research to identify the metabolites secreted by these microbes for potential disease treatment.

Moreover, comparative analysis of the gut microbiota in young and older adults could unveil microbes responsible for maintaining youthfulness. This intriguing concept presents the possibility of using these microbes or their products as therapeutics to delay or reverse aging and promote youthfulness, though unconventional, holding promise for future research and therapeutic interventions. Such investigations have the potential to revolutionize the field of aging and open new avenues for age-related disease management.

Acknowledgments

We are thankful to IIT Ropar for providing facilities and to the Ministry of Human Resources Development, India, for funding the project and fellowship support to JAM for pursuing his PhD.

Disclosure statement

No potential conflict of interest was reported by the authors.

Funding

We are thankful to the Council of Scientific and Industrial Research and DST-SPARC for funding.

References

 Baquero F, Nombela C. The microbiome as a human organ. Clin Microbiol Infect. 2012;18(Suppl 4):2–4. doi:10.1111/j.1469-0691.2012.03916.x.

- 2. Lynch JB, Hsiao EY. Microbiomes as sources of emergent host phenotypes. Sci. 2019;365(6460):1405–1409. doi:10.1126/science.aay0240.
- Dethlefsen L, McFall-Ngai M, Relman DA. An ecological and evolutionary perspective on human-microbe mutualism and disease. Nature. 2007;449 (7164):811–818. doi:10.1038/nature06245.
- Chu H, Mazmanian SK. Innate immune recognition of the microbiota promotes host-microbial symbiosis. Nat Immunol. 2013;14(7):668–675. doi:10.1038/ni.2635.
- Zheng D, Liwinski T, Elinav E. Interaction between microbiota and immunity in health and disease. Cell Res. 2020;30(6):492–506. doi:10.1038/s41422-020-0332-7.
- 6. Kim C, Fang F, Weyand CM, Goronzy JJ. The life cycle of a T cell after vaccination – where does immune ageing strike? Clin Exp Immunol. 2016;187(1):71–81. doi:10.1111/cei.12829.
- Bashir H, Singh S, Singh RP, Agrewala JN, Kumar R. Age-mediated gut microbiota dysbiosis promotes the loss of dendritic cells tolerance. Aging Cell. 2023;22(6): doi:10.1111/ACEL.13838.
- Lee K-A, Flores RR, Jang IH, Saathoff A, Robbins PDIS. Immune Senescence, Immunosenescence and Aging. Front Aging. 2022;3:61. doi:10.3389/fragi.2022.900028.
- Coquerelle C, Moser M. DC subsets in positive and negative regulation of immunity. Immunol Rev. 2010;234(1):317–334. doi:10.1111/j.0105-2896.2009. 00887.x.
- Trombetta ES, Mellman I. Cell biology of antigen processing in vitro and in vivo. Annu Rev Immunol. 2004;23(1):975–1028. doi:10.1146/annurev.immunol. 22.012703.104538.
- Hackstein H, Thomson AW. Dendritic cells: emerging pharmacological targets of immunosuppressive drugs. Nat Rev Immunol. 2004;4(1):24–35. doi:10.1038/ nri1256.
- Horton C, Shanmugarajah K, Fairchild PJ. Harnessing the properties of dendritic cells in the pursuit of immunological tolerance. Biomed J. 2017;40(2):80–93. doi:10. 1016/j.bj.2017.01.002.
- Hawiger D, Inaba K, Dorsett Y, Guo M, Mahnke K, Rivera M, Ravetch JV, Steinman RM, Nussenzweig MC. Dendritic cells induce peripheral T cell unresponsiveness under steady state conditions in vivo. J Exp Med. 2001;194(6):769–780. doi:10.1084/jem.194.6.769.
- Lande R, Gilliet M. Plasmacytoid dendritic cells: key players in the initiation and regulation of immune responses. Ann N Y Acad Sci. 2010;1183(1):89–103. doi:10.1111/j.1749-6632.2009.05152.x.
- Agrawal A, Gupta S. Impact of aging on dendritic cell functions in humans. Ageing Res Rev. 2011;10(3):336. doi:10.1016/j.arr.2010.06.004.
- Agrawal A, Agrawal S, Tay J, Gupta S. Biology of dendritic cells in aging. J Clin Immunol. 2008;28 (1):14–20. doi:10.1007/s10875-007-9127-6.

- Franceschi C, Garagnani P, Parini P, Giuliani C, Santoro A. Inflammaging: a new immune-metabolic viewpoint for age-related diseases. Nat Rev Endocrinol. 2018;14(10):576–590. doi:10.1038/s41574-018-0059-4.
- Kinnebrew MA, Buffie C, Diehl G, Zenewicz L, Leiner I, Hohl T, Flavell R, Littman D, Pamer E. Interleukin 23 production by intestinal CD103(+)CD11b(+) dendritic cells in response to bacterial flagellin enhances mucosal innate immune defense. Immunity. 2012;36 (2):276–287. doi:10.1016/j.immuni.2011.12.011.
- Fujimoto K, Karuppuchamy T, Takemura N, Shimohigoshi M, Machida T, Haseda Y, Aoshi T, Ishii KJ, Akira S, Uematsu S, et al. A new subset of CD103+CD8α+ dendritic cells in the small Intestine Expresses TLR3, TLR7, and TLR9 and induces Th1 response and CTL activity. J Immunol. 2011;186 (11):6287-6295. doi:10.4049/jimmunol.1004036.
- 20. Liu H, Chen F, Wu W, Cao AT, Xue X, Yao S, Evans-Marin HL, Li Y-Q, Cong Y. TLR5 mediates CD172α+ intestinal lamina propria dendritic cell induction of Th17 cells. Sci Rep. 2016;6(1): doi:10.1038/srep22040.
- Persson EK, Uronen-Hansson H, Semmrich M, Rivollier A, Hägerbrand K, Marsal J, Gudjonsson S, Håkansson U, Reizis B, Kotarsky K, et al. IRF4 transcription-factor-dependent CD103(+)CD11b(+) dendritic cells drive mucosal T helper 17 cell differentiation. Immunity. 2013;38(5):958–969. doi:10. 1016/j.immuni.2013.03.009.
- 22. Zhao J, Zhao J, Legge K, Perlman S. Age-related increases in PGD2 expression impair respiratory DC migration, resulting in diminished T cell responses upon respiratory virus infection in mice. J Clin Invest. 2011;121(12):4921–4930. doi:10.1172/JCI59777.
- Agrawal A, Tay J, Ton S, Agrawal S, Gupta S. Increased reactivity of dendritic cells from aged subjects to self-antigen, the human DNA. J Immunol. 2009;182 (2):1138. doi:10.4049/jimmunol.182.2.1138.
- 24. Honda K, Littman DR. The microbiota in adaptive immune homeostasis and disease. Nat. 2016;535 (7610):75–84. doi:10.1038/nature18848.
- Bosco N, Noti M. The aging gut microbiome and its impact on host immunity. Genes Immun. 2021;22(5– 6):289–303. doi:10.1038/s41435-021-00126-8.
- Ragonnaud E, Biragyn A. Gut microbiota as the key controllers of "healthy" aging of elderly people. *Immun Ageing*. 2021;18(1):1–11. doi:10.1186/s12979-020-00213-w.
- O'Toole PW, Claesson MJ. Gut microbiota: changes throughout the lifespan from infancy to elderly. Int Dairy J. 2010;20(4):281–291. doi:10.1016/j.idairyj. 2009.11.010.
- Agrawal A, Agrawal S, Gupta S. Role of dendritic cells in inflammation and loss of tolerance in the elderly. Front Immunol. 2017;8:896. doi:10.3389/fimmu.2017.00896.
- 29. Clements SJ, Maijo M, Ivory K, Nicoletti C, Carding SR. Age-associated decline in dendritic cell function and

the impact of Mediterranean diet intervention in elderly subjects. Front Nutr. 2017;4:294058. doi:10. 3389/fnut.2017.00065.

- Calder PC. Lipid-laden dendritic cells fail to function. Cell Res. 2010;20(10):1089–1091. doi:10.1038/cr.2010.124.
- 31. De Clercq NC, Frissen MN, Davids M, Groen AK, Nieuwdorp M. Weight gain after fecal microbiota transplantation in a patient with recurrent underweight following clinical recovery from anorexia nervosa. Psychother Psychosom. 2019;88(1):58-60. doi:10. 1159/000495044.
- 32. Claesson MJ, Cusack S, O'Sullivan O, Greene-Diniz R, de Weerd H, Flannery E, Marchesi JR, Falush D, Dinan T, Fitzgerald G, et al. Composition, variability, and temporal stability of the intestinal microbiota of the elderly. Proc Natl Acad Sci U S A. 2011;108 Suppl 1:4586–4591. doi:10.1073/pnas.1000097107.
- Tosti V, Bertozzi B, Fontana L. Health benefits of the Mediterranean diet: metabolic and molecular mechanisms. J Gerontol A Biol Sci Med Sci. 2018;73 (3):318–326. doi:10.1093/gerona/glx227.
- 34. Tamura K, Sasaki H, Shiga K, Miyakawa H, Shibata S. The timing effects of soy protein intake on mice gut microbiota. Nutrients. 2019;12(1):87. doi:10.3390/ nu12010087.
- Ticinesi A, Tana C, Nouvenne A, Prati B, Lauretani F, Meschi T. Gut microbiota, cognitive frailty and dementia in older individuals: a systematic review. Clin Interv Aging. 2018;13:1497–1511. doi:10.2147/CIA.S139163.
- 36. Everitt AV, Hilmer SN, Brand-Miller JC, Jamieson HA, Truswell AS, Sharma AP, Mason RS, Morris BJ, Couteur DGL. Dietary approaches that delay age-related diseases. Clin Interv Aging. 2006;1 (1):11–31. doi:10.2147/ciia.2006.1.1.11.
- Larrosa M, Luceri C, Vivoli E, Pagliuca C, Lodovici M, Moneti G, Dolara P. Polyphenol metabolites from colonic microbiota exert anti-inflammatory activity on different inflammation models. Mol Nutr Food Res. 2009;53(8):1044–1054. doi:10.1002/mnfr.200800446.
- Biagi E, Franceschi C, Rampelli S, Severgnini M, Ostan R, Turroni S, Consolandi C, Quercia S, Scurti M, Monti D, et al. Gut Microbiota and Extreme Longevity. Curr Biol. 2016;26(11):1480–1485. doi:10. 1016/j.cub.2016.04.016.
- 39. Wang N, Li R, Lin H, Fu C, Wang X, Zhang Y, Su M, Huang P, Qian J, Jiang F, et al. Enriched taxa were found among the gut microbiota of centenarians in East China. PLoS ONE. 2019;14(10):e0222763. doi:10. 1371/journal.pone.0222763.
- Brestoff JR, Artis D. Commensal bacteria at the interface of host metabolism and the immune system. Nat Immunol. 2013;14(7):676. doi:10.1038/ni.2640.
- 41. Hu J, Wang C, Huang X, Yi S, Pan S, Zhang Y, Yuan G, Cao Q, Ye X, Li H, et al. Gut microbiota-mediated secondary bile acids regulate dendritic cells to attenuate autoimmune uveitis through TGR5 signaling. Cell Rep. 2021;36(12):109726. doi:10.1016/j.celrep.2021.109726.

- 42. Ichikawa R, Takayama T, Yoneno K, Kamada N, Kitazume MT, Higuchi H, Matsuoka K, Watanabe M, Itoh H, Kanai T, et al. Bile acids induce monocyte differentiation toward interleukin-12 hypo-producing dendritic cells via a TGR5-dependent pathway. Immunology. 2012;136(2):153. doi:10.1111/j.1365-2567.2012.03554.x.
- 43. Singh N, Thangaraju M, Prasad PD, Martin PM, Lambert NA, Boettger T, Offermanns S, Ganapathy V. Blockade of dendritic cell development by bacterial fermentation products butyrate and propionate through a transporter (Slc5a8)-dependent inhibition of histone deacetylases. J Biol Chem. 2010;285 (36):27601–27608. doi:10.1074/jbc.M110.102947.
- 44. Sä MD, Parolini O, Böhmig GA, Kelemen P, Krieger P-M, Neumüller J, Knarr K, Kammlander W, Hörl WH, Diakos C, et al. Bacterial metabolite interference with maturation of human monocyte-derived dendritic cells. J Leukoc Biol. 2002;71(2):238–246. doi:10.1189/jlb.71.2.238.
- 45. Kaisar MMM, Pelgrom LR, van der Ham AJ, Yazdanbakhsh M, Everts B. Butyrate conditions human dendritic cells to prime type 1 regulatory T cells via both histone deacetylase inhibition and G protein-coupled receptor 109A signaling. Front Immunol. 2017;8. doi:10.3389/fimmu.2017.01429.
- 46. Wypych TP, Pattaroni C, Perdijk O, Yap C, Trompette A, Anderson D, Creek DJ, Harris NL, Marsland BJ. Microbial metabolism of L-tyrosine protects against allergic airway inflammation. Nat Immunol. 2021;22(3):279–286. doi:10.1038/s41590-020-00856-3.
- 47. Ichinohe T, Pang IK, Kumamoto Y, Peaper DR, Ho JH, Murray TS, Iwasaki A. Microbiota regulates immune defense against respiratory tract influenza a virus infection. Proc Natl Acad Sci U S A. 2011;108 (13):5354–5359. doi:10.1073/pnas.1019378108.
- 48. Li G, Ding H, Yu X, Meng Y, Li J, Guo Q, Zhou H, Shen N. Spermidine suppresses inflammatory DC function by activating the FOXO3 pathway and counteracts autoimmunity. iScience. 2020;23(1):100807. doi:10. 1016/j.isci.2019.100807.
- 49. Mondanelli G, Bianchi R, Pallotta MT, Orabona C, Albini E, Iacono A, Belladonna ML, Vacca C, Fallarino F, Macchiarulo A, et al. A relay pathway between arginine and tryptophan metabolism confers immunosuppressive properties on dendritic cells. Immunity. 2017;46(2):233–244. doi:10.1016/j.immuni. 2017.01.005.
- Ghosh TS, Shanahan F, O'Toole PW. The gut microbiome as a modulator of healthy ageing. Nat Rev Gastroenterol Hepatol. 2022;19(9):565. doi:10.1038/ s41575-022-00605-x.
- 51. Ouwehand AC, Isolauri E, Kirjavainen PV, Salminen SJ. Adhesion of four Bifidobacterium strains to human intestinal mucus from subjects in different

age groups. FEMS Microbiol Lett. 1999;172(1):61-64. doi:10.1111/j.1574-6968.1999.tb13450.x.

- 52. Op den Camp HJ, Oosterhof A, Veerkamp JH. Interaction of bifidobacterial lipoteichoic acid with human intestinal epithelial cells. Infect Immun. 1985;47(1):332-334. doi:10.1128/iai.47.1.332-334.1985.
- 53. Waters JL, Ley RE. The human gut bacteria Christensenellaceae are widespread, heritable, and associated with health. BMC Biol. 2019;17(1): doi:10.1186/ s12915-019-0699-4.
- 54. Bodogai M, O'Connell J, Kim K, Kim Y, Moritoh K, Chen C, Gusev F, Vaughan K, Shulzhenko N, Mattison JA, et al. Commensal bacteria contribute to insulin resistance in aging by activating innate B1a cells. Sci Transl Med. 2018;10(467). doi:10.1126/sci translmed.aat4271.
- 55. Xu C, Zhu H, Qiu P. Aging progression of human gut microbiota. BMC Microbiol. 2019;19(1): doi:10.1186/ s12866-019-1616-2.
- Hasegawa H, Matsumoto T. Mechanisms of tolerance induction by dendritic cells in vivo. Front Immunol. 2018;9. doi:10.3389/fimmu.2018.00350.
- 57. Huang G, Wang Y, Chi H. Control of T cell fates and immune tolerance by p38α signaling in mucosal CD103 + dendritic cells. J Immunol. 2013;191(2):650–659. doi:10.4049/jimmunol.1300398.
- O'Shea EF, Cotter PD, Stanton C, Ross RP, Hill C. Production of bioactive substances by intestinal bacteria as a basis for explaining probiotic mechanisms: bacteriocins and conjugated linoleic acid. Int J Food Microbiol. 2012;152(3):189–205. doi:10.1016/j.ijfoodmi cro.2011.05.025.
- Stagg AJ. Intestinal dendritic cells in health and gut inflammation. Front Immunol. 2018;9:2883. doi:10. 3389/fimmu.2018.02883.
- Owen JL, Mohamadzadeh M. Microbial activation of gut dendritic cells and the control of mucosal immunity. J Interf Cytokine Res. 2013;33(11):619. doi:10.1089/jir.2013.0046.
- Malik JA, Kaur G, Agrewala JN. Revolutionizing medicine with toll-like receptors: a path to strengthening cellular immunity. Int J Biol Macromol. 2023;253:127252. doi:10.1016/J.IJBIOMAC.2023.127252.
- 62. Cerovic V, Houston SA, Westlund J, Utriainen L, Davison ES, Scott CL, Bain CC, Joeris T, Agace WW, Kroczek RA, et al. Lymph-borne CD8α+ dendritic cells are uniquely able to cross-prime CD8+ T cells with antigen acquired from intestinal epithelial cells. Mucosal Immunol. 2015;8(1):38–48. doi:10.1038/mi. 2014.40.
- Iwasaki A, Kelsall BL. Freshly isolated Peyer's patch, but not spleen, dendritic cells produce interleukin 10 and induce the differentiation of T helper type 2 cells. The Journal Of Experimental Medicine. 1999;190 (2):229–240. doi:10.1084/jem.190.2.229.

- 64. Williams AM, Probert CSJ, Stepankova R, Tlaskalova-Hogenova H, Phillips A, Bland PW. Effects of microflora on the neonatal development of gut mucosal T cells and myeloid cells in the mouse. Immunology. 2006;119(4):470. doi:10.1111/j.1365-2567.2006.02458.x.
- 65. Atarashi K, Nishimura J, Shima T, Umesaki Y, Yamamoto M, Onoue M, Yagita H, Ishii N, Evans R, Honda K, et al. ATP drives lamina propria TH17 cell differentiation. Nature. 2008;455(7214):808-812. doi:10.1038/nature07240.
- 66. Wilson KR, Gressier E, McConville MJ, Bedoui S. Microbial metabolites in the maturation and activation of dendritic cells and their relevance for respiratory immunity. Front Immunol. 2022;13. doi:10.3389/ fimmu.2022.897462.
- Mortaz E, Adcock IM, Folkerts G, Barnes PJ, Paul Vos A, Garssen J. Probiotics in the management of lung diseases. Mediators Inflamm. 2013;2013:1–10. doi:10.1155/2013/751068.
- Foligne B, Zoumpopoulou G, Dewulf J, Ben Younes A, Chareyre F, Sirard J-C, Pot B, Grangette C. A key role of dendritic cells in probiotic functionality. PLoS ONE. 2007;2(3):e313. doi:10.1371/journal.pone.0000313.
- Ginaldi L, De Martinis M, Monti D, Franceschi C. The immune system in the elderly: activation-induced and damage-induced apoptosis. Immunol Res. 2004;30 (1):081–094. doi:10.1385/IR:30:1:081.
- Agrawal A, Agrawal S, Cao J-N, Su H, Osann K, Gupta S. Altered innate immune functioning of dendritic cells in elderly humans: a role of phosphoinositide 3-kinase-signaling pathway. J Immunol. 2007;178 (11):6912–6922. doi:10.4049/jimmunol.178.11.6912.
- 71. Panda A, Qian F, Mohanty S, van Duin D, Newman FK, Zhang L, Chen S, Towle V, Belshe RB, Fikrig E, et al. Age-associated decrease in TLR function in primary human dendritic cells predicts influenza vaccine response. J Immunol. 2010;184(5):2518–2527. doi:10. 4049/jimmunol.0901022.
- 72. Jing Y, Shaheen E, Drake RR, Chen N, Gravenstein S, Deng Y. Aging is associated with a numerical and functional decline in plasmacytoid dendritic cells, whereas myeloid dendritic cells are relatively unaltered in human peripheral blood. Hum Immunol. 2009;70 (10):777. doi:10.1016/j.humimm.2009.07.005.
- Teig N, Moses D, Gieseler S, Schauer U. Age-related changes in human blood dendritic cell subpopulations. Scand J Immunol. 2002;55(5):453–457. doi:10.1046/j. 1365-3083.2002.01068.x.
- 74. Shodell M, Siegal FP. Circulating, interferon-producing plasmacytoid dendritic cells decline during human ageing. Scand J Immunol. 2002;56(5):518–521. doi:10. 1046/j.1365-3083.2002.01148.x.
- Pérez-Cabezas B, Naranjo-Gómez M, Fernández MA, Grífols JR, Pujol-Borrell R, Borràs FE. Reduced numbers of plasmacytoid dendritic cells in aged blood donors. Exp Gerontol. 2007;42(10):1033–1038. doi:10. 1016/j.exger.2007.05.010.

- 76. Della Bella S, Bierti L, Presicce P, Arienti R, Valenti M, Saresella M, Vergani C, Villa ML. Peripheral blood dendritic cells and monocytes are differently regulated in the elderly. Clin Immunol. 2007;122(2):220–228. doi:10.1016/j.clim.2006.09.012.
- 77. Valkenburg SA, Venturi V, Dang THY, Bird NL, Doherty PC, Turner SJ, Davenport MP, Kedzierska K. Correction: early priming minimizes the age-related immune Compromise of CD8 + T cell diversity and function. PLoS Pathog. 2012;8(3): doi:10.1371/annota tion/e142f9de-7f30-4759-bda1-a651e86d5ba6.
- Stout-Delgado HW, Vaughan SE, Shirali AC, Jaramillo RJ, Harrod KS. Impaired NLRP3 inflammasome function in elderly mice during influenza infection is rescued by treatment with nigericin. J Immunol. 2012;188(6):2815–2824. doi:10.4049/jim munol.1103051.
- 79. Inam Z, Felton E, Burrell A, Chaney H, Sami I, Koumbourlis AC, Freishtat RJ, Zemanick ET, Crandall KA, Hahn A, et al. Impact of antibiotics on the lung microbiome and lung function in children with cystic fibrosis 1 year after hospitalization for an initial pulmonary exacerbation. Open Forum Infect Dis. 2022;9(9). doi:10.1093/ofid/ofac466.
- 80. Schuijt TJ, Lankelma JM, Scicluna BP, de Sousa e Melo F, Roelofs JJTH, de Boer JD, Hoogendijk AJ, de Beer R, de Vos A, Belzer C, et al. The gut microbiota plays a protective role in the host defence against pneumococcal pneumonia. Gut. 2016;65(4):575–583. doi:10. 1136/gutjnl-2015-309728.
- Du Y, Gao Y, Zeng B, Fan X, Yang D, Yang M. Effects of anti-aging interventions on intestinal microbiota. Gut Microbes. 2021;13(1): doi:10.1080/19490976.2021. 1994835.
- 82. Agrawal S, Ganguly S, Tran A, Sundaram P, Agrawal A. Retinoic acid treated human dendritic cells induce T regulatory cells via the expression of CD141 and GARP which is impaired with age. Aging (Albany NY). 2016;8(6):1223–1235. doi:10.18632/aging.100973.
- Busse PJ, Mathur SK. Age-related changes in immune function: effect on airway inflammation. J Allergy Clin Immunol. 2010;126(4):690–699. doi:10.1016/j.jaci. 2010.08.011.
- 84. MOUTON CP, BAZALDUA OV, PIERCE B, ESPINO DV. Common infections in older adults. Am Fam Physician. 2001;63:257–268.
- 85. Biagi E, Nylund L, Candela M, Ostan R, Bucci L, Pini E, Nikküla J, Monti D, Satokari R, Franceschi C, et al. Correction: through Ageing, and beyond: gut microbiota and inflammatory status in seniors and centenarians. PLoS ONE. 2010;5(6). doi:10.1371/anno tation/df45912f-d15c-44ab-8312-e7ec0607604d.
- 86. Nastasi C, Candela M, Bonefeld CM, Geisler C, Hansen M, Krejsgaard T, Biagi E, Andersen MH, Brigidi P, Ødum N, et al. The effect of short-chain fatty acids on human monocyte-derived dendritic cells. Sci Rep. 2015;5(1). doi:10.1038/srep16148.

- 87. Singh N, Gurav A, Sivaprakasam S, Brady E, Padia R, Shi H, Thangaraju M, Prasad P, Manicassamy S, Munn D, et al. Activation of the receptor (Gpr109a) for niacin and the commensal metabolite butyrate suppresses colonic inflammation and carcinogenesis. Immunity. 2014;40(1):128. doi:10.1016/j.immuni.2013. 12.007.
- Trompette A, Gollwitzer ES, Yadava K, Sichelstiel AK, Sprenger N, Ngom-Bru C, Blanchard C, Junt T, Nicod LP, Harris NL, et al. Gut microbiota metabolism of dietary fiber influences allergic airway disease and hematopoiesis. Nat Med. 2014;20(2):159–166. doi:10. 1038/nm.3444.
- Kumar M, Babaei P, Ji B, Nielsen J. Human gut microbiota and healthy aging: recent developments and future prospective. Nutr Heal Aging. 2016;4(1):3–16. doi:10.3233/NHA-150002.
- Shahbazi R, Yasavoli-Sharahi H, Alsadi N, Ismail N, Matar C. Probiotics in treatment of viral respiratory infections and neuroinflammatory disorders. Molecules. 2020;25(21):4891. doi:10.3390/molecules25214891.
- Hardy H, Harris J, Lyon E, Beal J, Foey AD. Probiotics, prebiotics and immunomodulation of gut mucosal defences: homeostasis and immunopathology. Nutrients. 2013;5(6):1869–1912. doi:10.3390/nu5061869.
- Bosch M, Méndez M, Pérez M, Farran A, Fuentes MC, Cuñé J. Lactobacillus plantarum CECT7315 and CECT7316 stimulate immunoglobulin production after influenza vaccination in elderly. Nutr Hosp. 2012;27 (2):504–509. doi:10.1590/S0212-16112012000200023.
- 93. Boge T, Rémigy M, Vaudaine S, Tanguy J, Bourdet-Sicard R, van der Werf S. A probiotic fermented dairy drink improves antibody response to influenza vaccination in the elderly in two randomised controlled trials. Vaccine. 2009;27(41):5677–5684. doi:10.1016/j.vaccine. 2009.06.094.
- 94. Kanauchi O, Andoh A, AbuBakar S, Yamamoto N. Probiotics and paraprobiotics in viral infection: clinical application and effects on the innate and acquired immune systems. Curr Pharm Des. 2018;24(6):710. doi:10.2174/1381612824666180116163411.
- 95. Van Puyenbroeck K, Hens N, Coenen S, Michiels B, Beunckens C, Molenberghs G, Van Royen P, Verhoeven V. Efficacy of daily intake of Lactobacillus casei Shirota on respiratory symptoms and influenza vaccination immune response: a randomized, double-blind, placebo-controlled trial in healthy elderly nursing home residents. Am J Clin Nutr. 2012;95 (5):1165–1171. doi:10.3945/ajcn.111.026831.
- 96. Barnabei L, Laplantine E, Mbongo W, Rieux-Laucat F, Weil R. NF-κB: at the borders of autoimmunity and inflammation. Front Immunol. 2021;12. doi:10.3389/ fimmu.2021.716469.
- Adler AS, Sinha S, Kawahara TLA, Zhang JY, Segal E, Chang HY. Motif module map reveals enforcement of aging by continual NF-κB activity. Genes Dev. 2007;21 (24):000.1–000. doi:10.1101/gad.1588507.

- 98. Flores RR, Clauson CL, Cho J, Lee B-C, McGowan SJ, Baker DJ, Niedernhofer LJ, Robbins PD. Expansion of myeloid-derived suppressor cells with aging in the bone marrow of mice through a NF-κB-dependent mechanism. Aging Cell. 2017;16(3):480–487. doi:10.1111/acel.12571.
- Seitz C, Deng H, Hinata K, Lin Q, Khavari P. Nuclear factor κB subunits induce epithelial cell growth arrest. Cancer Res. 2000;60(15):4085–92.
- Tilg H, Moschen AR. Adipocytokines: mediators linking adipose tissue, inflammation and immunity. Nat Rev Immunol. 2006;6(10):772–783. doi:10.1038/nri1937.
- 101. Tuljapurkar SR, McGuire TR, Brusnahan SK, Jackson JD, Garvin KL, Kessinger MA, Lane JT, O' Kane BJ, Sharp JG. Changes in human bone marrow fat content associated with changes in hematopoietic stem cell numbers and cytokine levels with aging. J Anat. 2011;219(5):574–581. doi:10.1111/j.1469-7580. 2011.01423.x.
- 102. Prakash S, Agrawal S, Cao JN, Gupta S, Agrawal A. Impaired secretion of interferons by dendritic cells from aged subjects to influenza: role of histone modifications. Age (Dordr). 2013;35(5):1785–1797. doi:10.1007/s11357-012-9477-8.
- 103. Bibiloni R, Fedorak RN, Tannock GW, Madsen KL, Gionchetti P, Campieri M, De Simone C, Sartor RB. VSL#3 probiotic-mixture induces remission in patients with active ulcerative colitis. Am J Gastroenterol. 2005;100(7):1539–1546. doi:10.1111/j.1572-0241.2005. 41794.x.
- 104. Dai C, Zheng C-Q, Meng F-J, Zhou Z, Sang L-X, Jiang M. VSL#3 probiotics exerts the antiinflammatory activity via PI3k/Akt and NF-κB pathway in rat model of DSS-induced colitis. Mol Cell Biochem. 2013;374(1-2):1-11. doi:10.1007/s11010-012-1488-3.
- 105. Rachmilewitz D, Katakura K, Karmeli F, Hayashi T, Reinus C, Rudensky B, Akira S, Takeda K, Lee J, Takabayashi K, et al. Toll-like receptor 9 signaling mediates the anti-inflammatory effects of Probiotics in murine experimental colitis. Gastroenterology. 2004;126(2):520–528. doi:10.1053/j.gastro.2003.11.019.
- 106. Alsegiani A, Shah Z. The influence of gut microbiota alteration on age-related neuroinflammation and cognitive decline. Neural Regen Res. 2022;17(11):2407. doi:10.4103/1673-5374.335837.
- 107. Mariat D, Firmesse O, Levenez F, Guimarăes VD, Sokol H, Doré J, Corthier G, Furet J-P. The Firmicutes/Bacteroidetes ratio of the human microbiota changes with age. BMC Microbiol. 2009;9 (1):123. doi:10.1186/1471-2180-9-123.
- 108. Li Y, Ning L, Yin Y, Wang R, Zhang Z, Hao L, Wang B, Zhao X, Yang X, Yin L, et al. Age-related shifts in gut microbiota contribute to cognitive decline in aged rats. Aging (Albany NY). 2020;12(9):7801. doi:10.18632/ aging.103093.
- Meckel KR, Kiraly DD. A potential role for the gut microbiome in substance use disorders. Psychopharmacol (Berl). 2019;236(5):1513. doi:10.1007/s00213-019-05232-0.

- 110. Wang HX, Wang YPC. Gut microbiota-brain axis. Med J (Engl). 2016;129(19):2373–2380. doi:10.4103/0366-6999.190667.
- 111. Degruttola AK, Low D, Mizoguchi A, Mizoguchi E. Current understanding of dysbiosis in disease in human and animal models. Inflamm Bowel Dis. 2016;22(5):1137. doi:10.1097/MIB.000000000000750.
- 112. Pilleron S, Sarfati D, Janssen-Heijnen M, Vignat J, Ferlay J, Bray F, Soerjomataram I. Global cancer incidence in older adults, 2012 and 2035: A population-based study. Int J Cancer. 2019;144 (1):49–58. doi:10.1002/ijc.31664.
- 113. Wu J, Wang, S, Zheng, B, Qiu, X, Wang, H, Chen, L. Modulation of gut microbiota to enhance effect of checkpoint inhibitor immunotherapy. Front Immunol. 2021;12:2554. doi:10.3389/fimmu.2021.669150.
- 114. Bowdish DME. The aging lung: is lung health good health for older adults? Chest. 2019;155(2):391-400. doi:10.1016/j.chest.2018.09.003.
- 115. Grier A, McDavid A, Wang B, Qiu X, Java J, Bandyopadhyay S, Yang H, Holden-Wiltse J, Kessler HA, Gill AL, et al. Neonatal gut and respiratory microbiota: coordinated development through time and space. Microbiome. 2018;6(1):1–19. doi:10.1186/ s40168-018-0566-5.
- 116. Mathieu E, MacPherson CW, Belvis J, Mathieu O, Robert V, Saint-Criq V, Langella P, Tompkins TA, Thomas M. Oral Primo-Colonizing Bacteria Modulate Inflammation and Gene Expression in Bronchial Epithelial Cells. Microorganisms. 2020;8(8):1–19. doi:10.3390/microorganisms8081094.
- 117. Segata N, Haake S, Mannon P, Lemon KP, Waldron L, Gevers D, Huttenhower C, Izard J. Composition of the adult digestive tract bacterial microbiome based on seven mouth surfaces, tonsils, throat and stool samples. Genome Biol. 2012;13(6):1–18. doi:10.1186/ gb-2012-13-6-r42.
- 118. Saint-Criq V, Lugo-Villarino G, Thomas M. Dysbiosis, malnutrition and enhanced gut-lung axis contribute to age-related respiratory diseases. Ageing Res Rev. 2021;66:101235. doi:10.1016/j.arr.2020.101235.
- 119. Cho SJ, Stout-Delgado HW. Aging and lung disease. Annu Rev Physiol. 2020;82(1):433-459. doi:10.1146/ annurev-physiol-021119-034610.
- 120. Khan N, Vidyarthi A, Nadeem S, Negi S, Nair G, Agrewala JN. Alteration in the gut microbiota provokes susceptibility to tuberculosis. Front Immunol. 2016;7:218883. doi:10.3389/fimmu.2016.00529.
- 121. Cerqueira César Machado M, Pinheiro da Silva F. Intestinal barrier dysfunction in human pathology and aging. Curr Pharm Des. 2016;22(30):4645–4650. doi:10. 2174/1381612822666160510125331.
- 122. Molinero N, Antón-Fernández A, Hernández F, Ávila J, Bartolomé B, Moreno-Arribas M Victoria. (). Gut Microbiota, an Additional Hallmark of Human Aging and Neurodegeneration. Neuroscience. 2023;518:141– 161. 10.1016/j.neuroscience.2023.02.014

- 123. Marsland BJ, Trompette A, Gollwitzer ES. The gut-lung axis in respiratory disease. Ann Am Thorac Soc. 2015;12 Suppl 2:S150–S156. doi:10.1513/AnnalsATS. 201503-133AW.
- 124. Wang Q, Li F, Liang B, Liang Y, Chen S, Mo X, Ju Y, Zhao H, Jia H, Spector TD, et al. A metagenome-wide association study of gut microbiota in asthma in UK adults. BMC Microbiol. 2018;18(1). doi:10.1186/ s12866-018-1257-x.
- 125. Vernocchi P, Del Chierico F, Russo A, Majo F, Rossitto M, Valerio M, Casadei L, La Storia A, De Filippis F, Rizzo C, et al. Gut microbiota signatures in cystic fibrosis: loss of host CFTR function drives the microbiota enterophenotype. PLoS ONE. 2018;13(12): e0208171. doi:10.1371/journal.pone.0208171.
- 126. Song W, Yue Y, Zhang Q. Imbalance of gut microbiota is involved in the development of chronic obstructive pulmonary disease: a review. Biomed Pharmacother. 2023;165:115150. doi:10.1016/j.biopha.2023.115150.
- 127. Wen L, Shi, L, Kong, X-L, Li, K-Y, Li, H, Jiang, D-X, Zhang, F, Zhou, Z-G. Gut microbiota protected against pseudomonas aeruginosa pneumonia via restoring Treg/Th17 balance and metabolism. Front Cell Infect Microbiol. 2022;12:856633. doi:10.3389/fcimb.2022. 856633.
- 128. Santo CE, Caseiro C, Martins MJ, Monteiro R, Brandão I. Gut microbiota, in the halfway between nutrition and lung function. Nutrients. 2021;13 (5):1716. doi:10.3390/nu13051716.
- 129. Gutiérrez-Carrasquilla L, Sánchez E, Hernández M, Polanco D, Salas-Salvadó J, Betriu À, Gaeta A, Carmona P, Purroy F, Pamplona R, et al. Effects of Mediterranean diet and physical activity on pulmonary function: a cross-Sectional analysis in the ILERVAS project. Nutrients. 2019;11(2):329. doi:10.3390/ nu11020329.
- 130. Davani-Davari D, Negahdaripour M, Karimzadeh I, Seifan M, Mohkam M, Masoumi S, Berenjian A, Ghasemi Y. Prebiotics: definition, types, sources, mechanisms, and clinical applications. Foods (Basel, Switzerland). 2019;8(3):92. doi:10.3390/foods8030092.
- 131. Agrawal A. Dendritic cell-airway epithelial cell cross-talk changes with age and contributes to chronic lung inflammatory diseases in the elderly. Int J Mol Sci. 2017;18(6):1206. doi:10.3390/ijms18061206.
- Ank N, Paludan SR. Type III IFNs: new layers of complexity in innate antiviral immunity. Biofactors. 2009;35(1):82–87. doi:10.1002/biof.19.
- 133. Martin LD, Rochelle LG, Fischer BM, Krunkosky TM, Adler KB. Airway epithelium as an effector of inflammation: molecular regulation of secondary mediators. Eur Respir J. 1997;10(9):2139–2146. doi:10.1183/ 09031936.97.10092139.
- 134. Hammad H, Chieppa M, Perros F, Willart MA, Germain RN, Lambrecht BN. House dust mite allergen induces asthma via Toll-like receptor 4 triggering of

airway structural cells. Nat Med. 2009;15(4):410-416. doi:10.1038/nm.1946.

- Lambrecht BN, Hammad H. Allergens and the airway epithelium response: gateway to allergic sensitization. J Allergy Clin Immunol. 2014;134(3):499–507. doi:10. 1016/j.jaci.2014.06.036.
- 136. Claudio E, Tassi I, Wang H, Tang W, Ha H-L, Siebenlist U. Cutting edge: IL-25 targets dendritic cells to attract IL-9-producing T cells in acute allergic lung inflammation. J Immunol. 2015;195(8):3525-3529. doi:10.4049/jimmunol.1500436.
- 137. Ho JC, CHAN K, HU W, LAM W, ZHENG L, TIPOE G, SUN J, LEUNG R, TSANG K. The effect of aging on nasal mucociliary clearance, beat frequency, and ultrastructure of respiratory cilia. *Am J Respir Crit Care Med.* 2012;163 (4):983–988. doi:10.1164/ajrccm.163.4.9909121.
- 138. Svartengren M. Long-term clearance from small airways decreases with age. Eur Respir J. 2005;26 (4):609–615. doi:10.1183/09031936.05.00002105.
- 139. Kwon HK, Lee C-G, So J-S, Chae C-S, Hwang J-S, Sahoo A, Nam JH, Rhee JH, Hwang K-C, Im S-H, et al. Generation of regulatory dendritic cells and CD4

+Foxp3+ T cells by probiotics administration suppresses immune disorders. Proc Natl Acad Sci U S A. 2010;107(5):2159–2164. doi:10.1073/pnas.0904055107.

- 140. Chae CS, Kwon HK, Hwang JS, Kim JE, Im SH, Platten M. Prophylactic effect of Probiotics on the development of experimental autoimmune myasthenia gravis. PLoS ONE. 2012;7(12):52119. doi:10.1371/jour nal.pone.0052119.
- 141. Goldin BR, Gorbach SL. Effect of Lactobacillus acidophilus dietary supplements on 1,2-dimethylhydrazine dihydrochloride-induced intestinal cancer in rats. J Natl Cancer Inst. 1980;64(2):263–265. doi:10.1093/ jnci/64.2.263.
- 142. Górska A, Przystupski D, Niemczura MJ, Kulbacka J. Probiotic bacteria: a promising tool in cancer prevention and therapy. Curr Microbiol. 2019;76(8):939. doi:10.1007/s00284-019-01679-8.
- 143. Guimarães GR, Almeida PP, de Oliveira Santos L, Rodrigues LP, de Carvalho JL, Boroni M. Hallmarks of aging in macrophages: consequences to skin inflammaging. Cells. 2021;10(6):1323. doi:10.3390/ cells10061323.