

Invasive fusariosis

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SUMMARY Invasive fusariosis is a serious invasive fungal disease, affecting immunocompetent and, more frequently, immunocompromised patients. Localized disease is the typical clinical form in immunocompetent patients. Immunocompromised hosts at elevated risk of developing invasive fusariosis are patients with acute leukemia receiving chemotherapeutic regimens for remission induction, and those undergoing allogeneic hematopoietic cell transplant. In this setting, the infection is usually disseminated with positive blood cultures, multiple painful metastatic skin lesions, and lung involvement. Currently available antifungal agents have poor *in vitro* activity against *Fusarium* species, but a clear-cut correlation between *in vitro* activity and clinical effectiveness does not exist. The outcome of invasive fusariosis is largely dependent on the resolution of immunosuppression, especially neutrophil recovery in neutropenic patients.

KEYWORDS *Fusarium*, fusariosis, immunocompromised, fungal infection

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INTRODUCTION

Fusarium species are ubiquitous fungi, widely distributed in the air, soil, and water, including seawater and community and hospital water distribution systems (1, 2). *Fusarium* is considered one of the five most important plant pathogens, with outbreaks of disease in cereals, resulting in significant agricultural losses (3). *Fusarium* species also cause disease in animals, especially aquatic animals such as dolphins, seahorses, and turtles (4). In humans, the most frequent diseases caused by *Fusarium* species are superficial: onychomycosis and keratitis (5, 6). Onychomycosis caused by *Fusarium* species is increasingly reported worldwide. It is difficult to treat superficial mycosis, with negative health consequences such as pain and impaired quality of life. In addition, in severely immunocompromised patients, onychomycosis may serve as a portal of entry for invasive disease, either locally invasive such as cellulitis and lymphangitis, or disseminated disease (7). *Fusarium* is the most frequent agent of fungal keratitis. Trauma and the use of contact lenses are common predisposing factors. Severe cases may evolve into corneal perforation and endophthalmitis (8).

In addition to onychomycosis and keratitis, *Fusarium* species cause invasive disease, which may be localized or, more frequently, disseminated. The latter occurs almost exclusively in severely immunosuppressed patients, particularly patients with acute myeloid leukemia (AML), acute lymphoid leukemia (ALL), and hematopoietic cell transplant (HCT) recipients (9). Finally, *Fusarium* species may cause allergic sinusitis, allergic bronchopulmonary disease, and mycotoxicosis (10–15).

PATHOGENESIS OF INVASIVE FUSARIOSIS

The main portal of entry of *Fusarium* species in cases of invasive disease is the airways, followed by the skin at sites of breakdown. As with invasive aspergillosis, after the inhalation of conidia, hyphae are formed in the alveoli, resulting in inflammation and bronchial dissemination. Subsequently, hyphae invade blood vessels, causing thrombosis and tissue infarction. The innate and adaptative immune responses are important in containing infections by *Fusarium* species and other filamentous fungi (16), with alveolar macrophages and neutrophils playing a major role in preventing hyphal formation and angioinvasion (17). Interferon-gamma and granulocyte colony-stimulating factor can enhance the phagocytic activity of monocytes and neutrophils (18, 19).

A remarkable difference between invasive fusariosis and aspergillosis is the frequent occurrence of positive blood cultures and metastatic skin lesions in the former (20). *Fusarium* microconidia produce yeast-like structures (adventitious sporulation). These small structures called aleuroconidia invade blood vessels causing fungemia and dissemination to various organs including the skin (21).

Animal models of invasive fusariosis in neutropenic mice showed a very high fungal burden, with no cell infiltration, whereas infection in non-neutropenic mice was characterized by necrosis, hemorrhage, macrophagic and neutrophilic infiltration, and a lower fungal burden (22). In addition to neutropenia, the inoculum size is an important predictor of outcome. In an animal model of non-neutropenic mice, intratracheal inoculation of 1×10^6 microconidia resulted in pulmonary infection, with a neutrophilic infiltrate and alveolar hemorrhage but no deaths. By contrast, the inoculation of 1×10^8 microconidia resulted in the rapid death—within 24 hours—of all mice (23).

FUSARIUM CHARACTERISTICS AND TAXONOMY

More than 300 phylogenetically different species of *Fusarium* grouped in more than 20 species complexes have been described, most of which are found in the environment (24). Most medically important *Fusarium* species belong to seven species complexes: *Fusarium solani* species complex (FSSC), *Fusarium oxysporum* species complex (FOSC), *Fusarium fujikuroi* species complex (FFSC), *Fusarium incarnatum-equiseti* species complex (FIESC), *Fusarium chlamidosporum* species complex (FCSC), *Fusarium dimerum* species complex (FDSC), and *Fusarium sporotrichoides* species complex (FSAMSC) (Table 1) (25).

TABLE 1 The most common *Fusarium* species complexes causing disease in humans and their respective species

<i>Fusarium solani</i> species complex	<i>Fusarium oxysporum</i> species complex	<i>Fusarium fujikuroi</i> species complex	<i>Fusarium incarnatum-equiseti</i> species complex	<i>Fusarium chlamidosporum</i> species complex	<i>Fusarium dimerum</i> species complex	<i>Fusarium sporotrichoides</i> species complex
<i>F. falciforme</i>	<i>F. oxysporum</i>	<i>F. acutatum</i>	<i>F. incarnatum</i>	<i>F. chlamydosporum</i>	<i>F. dimerum</i>	<i>F. aermeniicum</i>
<i>F. keratoplasticum</i>	Unnamed	<i>F. anthophilum</i>	<i>F. equiseti</i>		<i>F. delphinoides</i>	<i>F. brachygibbosum</i>
<i>F. lichenicola</i>		<i>F. andiyazi</i>	Unnamed		<i>F. penzigii</i>	<i>F. langsethiae</i>
<i>F. petroliphilum</i>		<i>F. fujikuroi</i>				<i>F. sporotrichioides</i>
<i>F. pseudensiforme</i>		<i>F. nygamai</i>				
		<i>F. proliferatum</i>				
		<i>F. verticillioides</i>				

Species belonging to FSSC account for approximately 50% of cases of invasive fusariosis (especially *F. falciforme* and *F. keratoplasticum*) with 20% of infections caused by FOFC. A few studies suggest geographic clustering of *Fusarium* species causing invasive disease. For example, most cases reported in Brazil belonged to the FSSC and FOFC species complexes (26, 27), while FFSC (mostly *F. verticillioides*, and *F. proliferatum*) were the most common agents reported from Europe (28). By contrast, a study evaluating 127 isolates from 26 countries (including isolates from the environment and from cases of superficial infection) did not show a clustering of species in a particular region of the globe (25).

Fusarium species grow easily and rapidly on different media without cycloheximide. On potato dextrose agar, the colonies have velvety to cottony surfaces and may present diverse colors: pink, yellow, red, gray, or white (Fig. 1). A distinguishing characteristic of the genus *Fusarium* is the production of hyaline banana-shaped macroconidia from phialides, with several transverse septa and foot cells at the base (Fig. 2). Ovoid microconidia may also be present, sometimes arranged in the apex of a conidiophore. In direct examination of biological materials, the hyphae are irregular, hyaline, septate branched, with swollen cells. Hyaline, thick-walled chlamydospores may be present, intercalary or in terminal position. Species identification requires molecular methods (29–32) or matrix-assisted laser desorption/ionization flight time (MALDI-TOF) (33–38).



FIG 1 Culture of *Fusarium* species on potato dextrose agar after 7 days showing colony with white cottony margins and velvety center with shades of gray. (Courtesy of Dr. Geovanni Breda, reproduced with permission.)

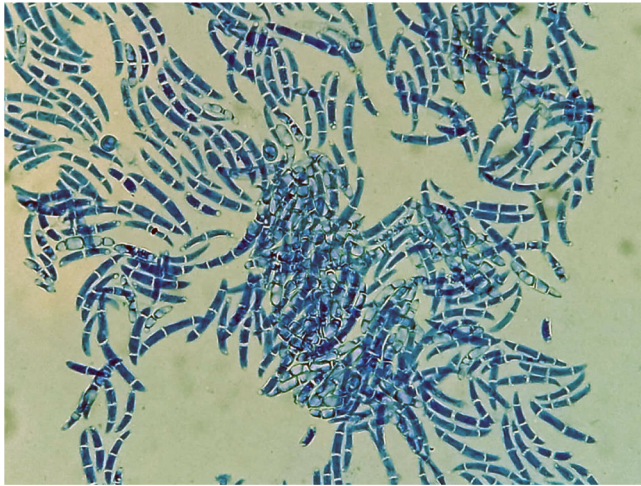


FIG 2 Microscopic morphology ($\times 400$) showing hyaline septate hyphae with banana-shaped macroconidia. (Courtesy of Dr. Giovanni Breda, reproduced with permission.)

EPIDEMIOLOGY AND CLINICAL SPECTRUM OF INVASIVE FUSARIOSIS

The clinical spectrum of invasive fusariosis is broad, and the clinical form depends on the portal of entry and the immune status of the host (Table 2). Immunocompromised patients are more likely to present with disseminated disease and immunocompetent patients usually have localized disease. In nonimmunocompromised patients with localized disease, skin breakdown may be present. This is the case of skin and soft tissue infection in burn patients (39–41) or after trauma (42), including combat-related injuries (43); skin ulcers in the setting of vascular insufficiency such as diabetes mellitus (44–47), venous insufficiency (48) and sickle cell disease (49); osteomyelitis after trauma (50–53) or surgery (54); arthritis after trauma (55, 56); endophthalmitis following eye injury (57), ocular surgery (58–60), or complicating keratitis (61, 62); peritonitis associated with peritoneal dialysis (63–66); and endocarditis following cardiac surgery (67, 68). *Fusarium* species may also be occasional agents of eumycetoma (69, 70). Immunocompetent patients may also develop sinusitis (71, 72), pneumonia (73, 74), fungemia (75), and rarely disseminated disease (76).

In immunocompromised patients, the most frequent clinical form of invasive fusariosis is that of disseminated disease with pneumonia, multiple skin lesions, and positive blood cultures. Most cases of invasive fusariosis in such patients occur in the context of remission induction of de novo or relapsed acute leukemia, or after allogeneic HCT (20). Occasional cases of disseminated disease have been reported in patients without classical immunosuppression, such as in a patient with acute respiratory distress syndrome and extracorporeal membrane oxygenation (77), and following multiple traumas (78, 79). Immunocompromised patients may also develop localized disease, including brain abscesses (80–82), endocarditis (83, 84), osteomyelitis (85, 86), sinusitis (87), and pneumonia (88).

Epidemiology of invasive fusariosis in patients with hematologic malignancies

The first case series of disseminated invasive fusariosis was reported in 1988. Nine cases of invasive fusariosis occurred over a 10-year period at a cancer center in the United States. Eight of these patients had hematologic malignancies, and four presented with positive blood cultures and metastatic skin lesions (89). In 1999, 43 cases of invasive fusariosis from the same institution were reported, with detailed characteristics of 54 additional published cases. The most frequent underlying disease was AML (53%), followed by ALL (16%) and most leukemic patients (83%) had relapsed disease. Fusariosis

TABLE 2 Clinical spectrum of invasive fusariosis

Immunocompetent patients	Immunocompromised patients
Skin and soft tissue infection	Disseminated disease
Osteomyelitis	Pneumonia
Arthritis	Fungemia
Endophthalmitis	Sinusitis
Peritonitis	Brain abscess
Endocarditis	Endocarditis
Eumycetoma	Osteomyelitis
Sinusitis	Arthritis
Pneumonia	Endophthalmitis
Fungemia	
Disseminated disease	

occurred after HCT in 12 patients (28%) and the diagnoses were made after engraftment in seven of those. Neutropenia was present in 84% of the 43 patients and 32% were receiving corticosteroids at the diagnosis of fusariosis (20).

In a multicenter retrospective study of 84 cases of invasive fusariosis in hematologic patients, AML and ALL were the most frequent underlying diseases (35% and 21%, respectively), followed by chronic myeloid leukemia (15%, all allogeneic HCT recipients). Fusariosis occurred in the context of HCT in 39% (33 cases, 29 of which were after allogeneic HCT). Among 79 patients with hematologic malignancies, 67 had active cancer at diagnosis of fusariosis and 40 were receiving treatment for relapsed disease. Neutropenia and corticosteroid therapy were present in 83% and 46%, respectively. The median duration of neutropenia before the diagnosis of fusariosis was 16 days (range 2–93) (90).

Detailed epidemiologic information regarding invasive fusariosis in HCT recipients was provided in a multicenter retrospective study of 61 cases (54 allogeneic and 7 autologous) (Table 3). The types of allogeneic HCT reflected the usual population of patients during the study period (1985–2001), with 48% of cases after receipt of HLA-matched related, 31% matched unrelated, and 10% mismatched related. The overall incidence of invasive fusariosis was 5.97 cases per 1,000 HCT, with a wide variation across centers (5.00–11.33 cases per 1,000 HCT), and the type of HCT: 4.21–5.0 after HLA matched related, 2.28 after HLA-compatible matched unrelated, 20.19 among mismatched related, and 1.4–2.0 among autologous HCTs (91).

Between 2007 and 2009, we conducted a prospective multicenter (eight sites) evaluation of the incidence of invasive fusariosis in Brazil among HCT recipients and patients with AML or myelodysplasia (MDS) receiving chemotherapy for remission induction. The 1-year cumulative incidence of invasive fusariosis among 237 patients with AML or MDS, 378 allogeneic HCT recipients, and 322 autologous HCT recipients was 5.2%, 3.8%, and 0.6% respectively (92).

In a recent prospective study evaluating the epidemiology of invasive fungal diseases (IFD) in four Brazilian centers between 2015 and 2016, the incidence of invasive fusariosis was 4.3% and 2%, respectively, in patients with AML and autologous HCT recipients, for an overall incidence of 1.6%. No case of fusariosis were diagnosed among allogeneic HCT recipients (93).

Two single-center studies confirmed the high frequency of invasive fusariosis in Brazil. The first was a retrospective study evaluating the etiologic agents of invasive mold disease (IMD) in patients with hematologic malignancies between 2004 and 2006. Among 29 cases of IMD, invasive fusariosis was the second most frequent (6 cases, 20.7%) (94). In the other study, 94 cases of IFD were diagnosed in 664 hematologic patients and 316 HCT recipients in a 10-year period. Invasive fusariosis was the second most frequent IFD (17 cases, 18.1%). The incidence of invasive fusariosis in patients with AML and allogeneic HCT recipients was 3.1%, for an overall incidence of 1.7% (95).

TABLE 3 Characteristics of 54 cases of invasive fusariosis in allogeneic hematopoietic cell transplant recipients

Time (days) of diagnosis after transplant	Neutropenia	Receipt of corticosteroids	GVHD ^a	Disseminated disease
0–30 (<i>n</i> = 20)	17 (85%)	12 (50%)	4 (20%)	18 (90%)
31–60 (<i>n</i> = 16)	5 (31%)	12 (75%)	14 (87%)	12 (75%)
100–365 (<i>n</i> = 10)	3 (30%)	5 (50%)	5 (50%)	4 (40%)
>365 (<i>n</i> = 8)	0	3 (38%)	8 (100%)	6 (75%)

^aGVHD = graft versus host disease.

In Italy, a retrospective multicenter study reported all cases of mold infections diagnosed in hematologic patients in 14 hospitals between 1988 and 1997. Six cases of invasive fusariosis were diagnosed in 2 of the 14 hospitals (one case in one hospital and five in another). The incidence of invasive fusariosis in patients with acute leukemia was 0.06% (0.08% in AML and no case in ALL) (96). In a subsequent study from the same group, 15 cases of invasive fusariosis were diagnosed among 11,802 patients with hematologic malignancies treated in 18 centers (0.1%), with 13 cases among 3012 patients with AML (0.4%), one case in 1173 patients with ALL (0.08%), and one case in 3457 patients with non-Hodgkin's lymphoma (0.03%) (97). In another study, only three cases of invasive fusariosis developed among 1,249 allogeneic HCT (0.2%) (98).

A surveillance study evaluated the epidemiology of IFD in HCT recipients from 23 transplant centers in the United States. The investigators identified invasive fusariosis in 53 of 1514 HCTs (3.5%). The 1-year cumulative incidence of IFD caused by rare molds (including *Fusarium* species) was <0.3% (99).

A retrospective single-center study in a hospital in Israel described the characteristics of 87 cases of non-*Aspergillus* mold infections occurring in patients with hematologic malignancies and in allogeneic HCT recipients. Invasive fusariosis was the leading infection, accounting for 35% of cases, followed by mucormycosis (25%). As reported in other studies, most cases of fusariosis occurred in patients with acute myeloid leukemia and disseminated disease was the most frequent clinical presentation (100).

In Spain, a retrospective multicenter study evaluated the incidence and epidemiology of invasive fusariosis in neutropenic and non-neutropenic patients. Between 2000 and 2015, all cases of invasive fusariosis diagnosed in 18 centers were reviewed. A total of 58 cases were diagnosed, 44 in neutropenic and 14 in non-neutropenic patients. Most cases occurred in patients with hematologic malignancies (79%). The incidence of invasive fusariosis increased from 0.40 cases per 100,000 admissions during 2000–2009 to 0.79 cases per 100,000 admissions during 2010–2015 ($P < 0.01$), for an overall incidence of 0.55 cases per 100,000 admissions (101).

Investigators assessed the risk factors for invasive fusariosis in a multicenter prospective cohort of 237 patients with AML or MDS receiving induction remission chemotherapy and in 663 HCT recipients. There were eight cases of invasive fusariosis in the AML/MDS cohort (3.4%). The only significant variable associated with invasive fusariosis was active smoking, with a hazard ratio (HR) of 9.11 [95% confidence interval (CI) 2.04–40.71]. In all, 17 cases (2.6%) developed in the HCT cohort, two among 318 autologous (0.6%) and 15 among 345 allogeneic HCTs (4.3%). Variables associated with invasive fusariosis in the early post-transplant period of allogeneic HCT (until day +40) were AML as underlying disease (HR: 4.38, 95% CI: 1.39–13.81), one of the eight centers (HR: 5.15, 95% CI: 1.66–15.97), and receipt of anti-thymocyte globulin in the conditioning regimen (HR: 22.17, 95% CI: 4.86–101.34). Factors associated with invasive fusariosis occurring after day +40 post-allogeneic HCT were a history of IMD before HCT (HR: 10.65, 95% CI: 1.19–95.39), non-myeloablative conditioning regimen (HR: 35.08, 95% CI: 3.90–315.27), and grade III/IV acute graft versus host disease (GVHD; HR: 16.50, 95% CI: 2.67–102.28). Cytomegalovirus (CMV) reactivation was also associated with invasive fusariosis (HR 5.99), but the P value was marginally significant ($P = 0.05$) (102).

Other studies evaluated risk factors for non-*Aspergillus* IMD following allogeneic HCT. Using data from the CIBMTR (Center for International Blood and Marrow Transplant Research) database, 52 cases of invasive fusariosis were identified between 1995 and 2008. Variables associated with invasive fusariosis occurring in the first-year post-transplant were umbilical cord blood as a stem cell source, with a relative risk (RR) of 3.11 (95% CI: 1.14–6.81) and prior CMV infection (RR: 2.72, 95% CI 1.24–5.97) (103).

As pointed out, most cases of invasive fusariosis occur in patients with hematologic malignancies, particularly in patients with acute leukemia. In the largest series of invasive fusariosis, 215 of 233 cases (92%) occurred in patients with hematologic diseases, and 150 of the 215 (69.8%) patients had AML, ALL, or MDS as an underlying disease. Other hematologic diseases included aplastic anemia, non-Hodgkin's lymphoma, multiple myeloma, and myelofibrosis. In most of these cases, invasive fusariosis occurred in the context of HCT (9).

Patients receiving ibrutinib and other Bruton kinase inhibitors are at increased risk of developing IDF, especially invasive aspergillosis (104). Recently, two cases of invasive fusariosis occurred in patients with chronic lymphocytic leukemia (CLL) receiving ibrutinib (105, 106). The first patient started ibrutinib as the fourth line of therapy and developed disseminated fusariosis 6 weeks after ibrutinib initiation. The second patient received ibrutinib as the sixth line of therapy and developed fusarial sinusitis after 4 years of ibrutinib.

Recently, chimeric antigen receptor T (CAR-T) cell therapy has been approved for the treatment of various hematologic malignancies including ALL, lymphoma, and multiple myeloma (107). Patients receiving CAR-T cell therapy are at increased risk of infection for various reasons including the cumulative immunosuppression associated with the underlying disease and prior therapies, the lymphodepleting chemotherapy, lymphopenia, hypogammaglobulinemia, and prolonged neutropenia (108). Bacteria and viruses account for most infections with only a minority caused by fungi (109). A case of invasive fusariosis was reported in a patient with refractory ALL who received CAR-T cell therapy, with a skin nodule and sinusitis (110).

Epidemiology of invasive fusariosis in other immunosuppressed patients

In contrast with the higher frequency of invasive fusariosis in patients with hematologic malignancies, including HCT recipients, invasive fusariosis is uncommon in solid organ transplant (SOT) recipients. For example, in a prospective survey of IFD in SOT recipients from 15 centers in the United States, 1,208 cases of IFD were diagnosed in a 5-year period, with only six cases of invasive fusariosis (111).

Cases of invasive fusariosis (either localized or disseminated) have been reported after liver (88, 112–117), kidney (118–124), lung (125–128), and multi-organ transplant (85, 129). Most cases of invasive fusariosis in liver transplant recipients occurred early after re-transplantation or rejection in the context of severe immunosuppression, with disseminated disease. By contrast, most cases in renal transplant recipients occurred years after transplant, with skin and subcutaneous nodules that evolved over weeks to months.

Sporadic cases of invasive fusariosis have been reported in other immunosuppressive conditions including patients with solid tumors (9, 130), chronic granulomatous disease (81, 131), AIDS (132, 133), hemophagocytic lymphohistiocytosis (134), chronic corticosteroid exposure (86), end-stage renal disease (84), primary immunodeficiency syndromes (135, 136) and, more recently, COVID-19 (137, 138).

Nosocomial acquisition of fusariosis, outbreaks, and pseudo-outbreaks

Since *Fusarium* species are widely encountered in the environment, invasive fusariosis may be acquired in the community. However, except for cases of localized disease associated with trauma in which the disease is community-acquired, it is difficult to identify whether the patient acquired fusariosis in the community or the hospital.

In the hospital, patients may acquire invasive fusariosis by airborne transmission, as shown in an outbreak in the hematology unit of a Brazilian hospital. Molecular typing was performed in 104 *Fusarium* species isolates recovered from the air of the unit and 15 isolates recovered from blood cultures. Genotypic relatedness was present in five isolates from the blood and seven from the air, belonging to FSSC, and in one FFSC bloodstream isolates and in one isolate recovered from the air of the same room occupied by the patient. A reduction in the incidence of invasive fusariosis coincided with the installation of water filters at the exit of faucets and showers in patients' rooms (139).

Fusarium species are frequently recovered from hospital water systems worldwide (140–142). In a prospective study conducted in a hospital in the United States, *Fusarium* species were present in the hospital water tanks and water-related structures such as shower heads, drains, and aerators. In addition, aerosolization of *Fusarium* species occurred after running the showers. Molecular methods of patients' and environmental isolates showed close relatedness, indicating the nosocomial source of invasive fusariosis (2).

In another study, an outbreak of 10 cases of invasive fusariosis diagnosed in a 2-year period in a children's cancer hospital was investigated. *Fusarium* species grew from the water of six patients' rooms, and from the air and other environmental sources in three rooms. Molecular typing showed relatedness between all *Fusarium oxysporum* isolates from the environment and two patients (143).

An outbreak of seven cases of fungemia due to *Fusarium verticillioides* (FFSC) was reported in a hospital in Greece. None of the patients had hematologic disease. An environmental source was not found, and the outbreak was resolved after the implementation of infection control measures consisting of intensive disinfection of patients' medication preparation and storage rooms (144). In another study, seven cases of catheter-related fungemia due to FOSC were diagnosed in a 5-month period at a pediatric cancer center. No environmental source was found, and the outbreak was controlled after the implementation of a multidisciplinary central line insertion care bundle (145).

An increase in the incidence of invasive fusariosis was observed in the hematology unit of a Brazilian hospital. Between 2001 and 2004, no cases of invasive fusariosis were diagnosed. In 2005, there were two cases, with an incidence of 2.47 cases per 1,000 admissions. The incidence increased to 4.95 in 2006, 16.78 in 2007, and 13.6 cases in 2008. A distinguishing feature of this outbreak was that in 17 of 20 cases (85%), a cutaneous portal of entry was present, either onychomycosis or interdigital intertrigo (7). In a review of skin lesions, 259 cases of invasive fusariosis were reported, and a cutaneous portal of entry was present in 11% of cases only (146). This prompted an environmental investigation, with the hypothesis that the hospital water was the source of infection. *Fusarium* species grew from 14 air samples, 44 swab samples of water-related structures, and 10 water samples. Molecular typing of these environmental isolates and 98 clinical isolates (55 from the hematologic patients and 43 from patients with superficial infections diagnosed in the dermatology outpatient clinic). Most clinical isolates belonged to the FSSC while most environmental isolates belonged to the FOSC. Furthermore, the predominant FSSC strains in patients were rarely found in the environment (147). The incidence of invasive fusariosis in the unit reduced in subsequent years without any intervention in the environment.

Interestingly, the incidence of superficial infections diagnosed in the dermatology outpatient unit caused by *Fusarium* species increased in the same period, suggesting that hematologic patients who developed invasive fusariosis were admitted with skin lesions that were overlooked at admission (7). Subsequently, a prospective study was conducted to investigate the frequency of skin lesions with positive culture for *Fusarium* species on admission of high-risk hematologic patients. Among 61 patients screened, alterations in the skin and/or nails were present in 32 patients (mostly interdigital intertrigo and onychomycosis) and 4 of these 32 patients had *Fusarium* species recovered

from their lesions. The presence of fusarial intertrigo or onychomycosis on admission was associated with the subsequent development of invasive fusariosis (148).

Pseudo-outbreaks of fusariosis have also been reported; three were associated with contamination of bronchoscopes by FOSC (149) and FSSC (150, 151). In a third pseudo-outbreak, sterile containers used to store and transport biologic materials for culture were contaminated by FFSC (152).

CLINICAL MANIFESTATIONS OF INVASIVE FUSARIOSIS

Clinical presentation of invasive fusariosis

The four most frequent clinical presentations of invasive fusariosis in immunosuppressed patients are as follows: (a) disseminated disease, (b) pneumonia, (c) fungemia, and (d) cellulitis or lymphangitis at sites of skin breakdown. Disseminated disease is the most frequent clinical presentation, and manifests as persistent or recurrent fever in the context of febrile neutropenia (20), with the concomitant or subsequent appearance of skin lesions (90), and involvement of other organs such as lungs, sinuses, and central nervous system. Blood cultures are frequently positive. Less frequently, the disease presents with persistent or recurrent fever and pneumonia (153).

Skin lesions

The skin is a frequent organ involved in invasive fusariosis, either as a primary infection or by hematogenous spread. The characteristics of skin involvement by *Fusarium* species were described in a study that evaluated 43 new cases of invasive fusariosis and 216 published cases. Among 232 immunocompromised and 27 immunocompetent patients, skin involvement was present in 70% of patients and was more frequent in immunocompromised patients (72% versus 52%) (146). Among 14 immunocompetent patients with skin lesions, 13 presented with localized infection, usually with a history of recent skin breakdown because of trauma, or onychomycosis. All patients with skin lesions associated with onychomycosis presented as cellulitis. By contrast, various patterns of lesions were present in patients with a history of trauma, including necrotic lesions, cellulitis, ulcers, and subcutaneous abscesses. Among 167 immunocompromised patients with skin lesions, 20 (12%) presented with localized lesions. Cellulitis at the site of preexisting onychomycosis (Fig. 3 and 4) was present in 8 of the 20 patients with localized skin lesions. Various patterns of skin lesions occurred in the other patients including necrotic lesions, abscesses, ulcers, and papular lesions. The most frequent skin lesions in immunocompromised patients were multiple disseminated painful erythematous papular or nodular lesions, with or without central necrosis (ecthyma gangrenosum-like) (Fig. 5). The necrosis is the result of invasion of the blood vessels of the dermis by hyphae, with subsequent thrombosis. Myalgia may occur in the context of disseminated metastatic skin lesions (20).

In addition to metastatic skin lesions, immunocompromised patients may present with localized skin involvement that subsequently disseminates to other organs. Among 14 patients with invasive fusariosis with a cutaneous portal of entry, the most frequent lesion was periungueal cellulitis with preexisting onychomycosis (6 cases), followed by interdigital intertrigo (6 patients) with or without lymphangitis (Fig. 6 to 8) (7).

Pneumonia

Lung involvement in fusariosis has many common features with aspergillosis, including the spectrum of clinical forms, similar clinical presentation, images, and fungal biomarkers (154). In a literature review of 357 cases of fusariosis diagnosed in immunocompetent and immunocompromised patients, pneumonia was reported in 152 cases (42%) and was more frequent in immunocompromised patients (46% versus 17.5%) (153). Among seven cases of pneumonia in immunocompromised patients identified in that study, lung involvement was part of disseminated disease in four. Bilateral lung involvement



FIG 3 Periungueal cellulitis. (Courtesy of Dr. Marcia Matos, reproduced with permission.)



FIG 4 Periungueal cellulitis with tissue destruction. (Courtesy of Dr. Marcia Matos, reproduced with permission.)



FIG 5 Nodular skin lesion with an area of central necrosis, with a typical appearance of ecthyma gangrenosum. (Courtesy of Dr. Marcia Matos, reproduced with permission.)

was present in five cases. More recently, another case of pneumonia occurred in an immunocompetent patient, with a right lung cavitory lesion (74).

In the literature review of 357 cases of invasive fusariosis, 24 cases occurred in SOT recipients. Pneumonia was reported in 10 cases, all in lung transplant recipients. Images



FIG 6 Interdigital intertrigo with cellulitis. (Courtesy of Dr. Marcia Matos, reproduced with permission.)



FIG 7 Interdigital intertrigo with cellulitis. (Courtesy of Dr. Hugo Morales, reproduced with permission.)



FIG 8 Interdigital intertrigo with cellulitis. (Courtesy of Dr. Marcia Matos, reproduced with permission.)

in the lungs included ground-glass infiltrates, nodules, alveolar infiltrates, bronchiectasis, and pleural effusion (153).

Lung involvement is frequent in patients with hematologic malignancies and HCT recipients: 54% and 84% in a series of 84 (90) and 43 cases, respectively (20). In a series of 233 cases (92% with hematologic diseases), lung involvement occurred in 49% of cases, with a higher frequency among neutropenic patients (55% versus 32% in non-neutropenic patients). Interestingly, patients with a cutaneous portal of entry were more likely to have bilateral lung involvement (88%) compared with patients with a presumed airborne portal of entry (68%), suggesting hematogenous spread to the lungs in patients with a cutaneous portal of entry (9).

Imaging of pulmonary fusariosis in patients with hematologic malignancies was characterized in 20 patients. Pulmonary symptoms were present in 95% of cases, and the most frequent manifestation was shortness of breath (14 patients). Chest CT scans of 11 patients were available for review. Nodules in nine (82%) patients, with sizes ranging from 0.3 to 2.7 cm, and a lung mass was present in six patients (size range: 3.0–6.7 cm). Mass or nodule was present in 9 of the 11 patients. No patient presented with a halo sign or tree-in-bud infiltrates (155).

Another study characterized the pattern of lung images in neutropenic patients with invasive fusariosis. Among nine cases, lung infiltrates were present in eight. The most frequent patterns of the image were ground-glass opacities and/or centrilobular micronodules and peribronchial consolidations with air bronchogram (5 cases each), followed by macronodules (4 cases). No patient presented with a halo sign. Compared with 11 cases of invasive aspergillosis, patients with invasive fusariosis were more likely to have small airway involvement and less likely to have macronodules with a halo sign (156).

In another study, 26 cases of invasive fusariosis were compared with 36 cases of invasive aspergillosis. Lung involvement was more frequent in aspergillosis (88.9% versus 50%, $P = 0.001$). The most frequent pattern of the image in cases of invasive fusariosis were macronodules (8 cases, 61.5%) (Fig. 9) and centrilobular micronodules and ground-glass infiltrates (7 cases each). The halo sign was present in three cases (Fig. 10). The only significant difference between cases of fusariosis and aspergillosis was a higher proportion of a halo sign in aspergillosis (62.5% versus 23.1%, $P = 0.02$) (154).

Sinusitis

Sinusitis is a frequent manifestation of invasive fusariosis, either occurring as a localized disease or, more commonly, as part of a disseminated disease. In a review of 294 cases of invasive fusariosis, sinusitis was reported in 54 cases (18%), with only two cases occurring in immunocompetent individuals (157). These patients presented with chronic infection. Among immunocompromised patients, in 70% sinusitis was part of disseminated disease. In a series of 233 cases of invasive fusariosis (92% occurring in patients with hematologic diseases), sinusitis was reported in 72 cases (31%).

Sinusitis may present as a radiologic finding with sinus opacities in a febrile neutropenic patient, or with nasal discharge and obstruction, with or without necrosis of mucosal surfaces and periorbital and nasal cellulitis. In patients with suspected fusarial sinusitis, nasal endoscopy with biopsy may yield the diagnosis (157).

Fungemia

Fungemia is frequent in invasive fusariosis, alone or (more frequently) as part of a disseminated disease. Indeed, *Fusarium* species are the leading agents of fungemia caused by molds in patients with hematologic malignancies (157). Among 84 patients with hematologic disease and a diagnosis of invasive fusariosis, fungemia was reported in 46 (55%), with 37 cases as part of disseminated disease and nine without apparent involvement of other organs (90). Occasional cases of catheter-related fungemia have been reported, including immunocompetent individuals (75, 112, 145). Typically, fungemia is the sole clinical manifestation of infection, the patient is in good general clinical conditions, and catheter removal plus a short course of antifungal therapy results in control of the disease.

Disseminated fusariosis

Disseminated disease is by far the most frequent clinical presentation of invasive fusariosis in severely immunocompromised patients such as those with profound neutropenia. In a series of 233 cases of invasive fusariosis, disseminated disease was present in 72% (9). In another study, disseminated disease occurred in 39 of 58 patients



FIG 9 Chest computed tomography showing macronodules in both lungs.

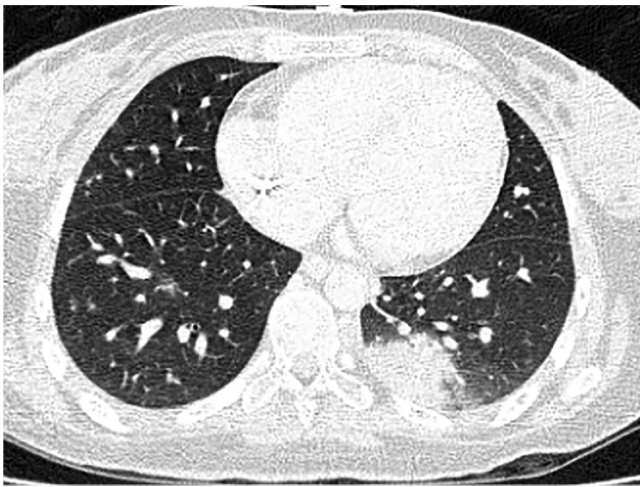


FIG 10 Chest computed tomography showing a large nodule with a halo sign in the left lung.

with invasive fusariosis (67.2%), with a higher incidence in neutropenic patients (79.5% versus 28.6% in non-neutropenic patients (101).

The typical presentation of disseminated fusariosis is that of the sudden appearance of various painful skin lesions in a persistently febrile neutropenic patient. Myalgia is frequent and the patients usually present with a toxic appearance (Fig. 11 and 12). In addition to lungs and sinus involvement, other organs affected include the liver, spleen (Fig. 13), eyes (endophthalmitis) (Fig. 14 to 16), and joints (55, 158).

DIAGNOSIS OF INVASIVE FUSARIOSIS

Apart from the clinical presentation, the diagnosis of invasive fusariosis relies on direct examination, culture, and/or histopathology of different biologic materials. In a series of 84 hematologic patients with invasive fusariosis, culture alone was the source of diagnosis in 65 patients (77%), and culture + histopathology in the remaining 19 patients. The most frequent sources of diagnosis by culture were the blood and the skin (90). In another study of 233 cases of invasive fusariosis, the diagnosis was made by culture alone in 138 cases (59%), culture plus histopathology in 83, and histopathology alone in 3 cases. The most frequent sources of diagnosis were the skin (100 cases) and the blood (85 cases) (9).

In a neutropenic patient who presents with typical skin lesions of invasive fusariosis (erythematous painful nodules), the fastest way of establishing a preliminary diagnosis of invasive fusariosis is by performing the direct examination of a fragment of skin biopsy.



FIG 11 Disseminated skin lesions and toxemic appearance. Lesions at various stages of evolution: papular and nodular lesions with and without central necrosis. (Courtesy of Dr. Marilza Campos Magalhães, reproduced with permission.)

In this context, a direct examination showing septate acute branching hyaline hyphae is highly suggestive of invasive fusariosis and should prompt the immediate start of appropriate antifungal therapy. The biopsy must be deep enough to identify hyphae invading blood vessels of the dermis in the histopathologic examination of the skin. In addition, one fragment of the biopsy should be placed in formalin for histopathology and another in sterile saline for direct examination and culture. This combination of culture and histopathology is crucial to establish a confirmatory diagnosis of invasive fusariosis because in tissue various hyaline fungi have the same picture. Therefore, in the absence of culture showing the growth of *Fusarium* species, the histopathologic diagnosis should be of hyalohyphomycosis, unless the fungus is identified in paraffin-embedded tissue by *in situ* hybridization (159) or real-time quantitative PCR (160).

As mentioned before, fungemia is a frequent manifestation of invasive fusariosis. Two studies evaluated the performance of fungal media in growing *Fusarium* species. In the first study, the authors compared the performance of selective fungal medium with that of standard aerobic media. For lower inocula, fungal growth was detected faster in the fungal bottle (161). The other study evaluated the performance of fungal media versus bacterial media with concomitant bacterial and fungal infection and concluded that blood culture bottles with fungal media should be preferred for optimal fungal growth (162).

Until recently, species identification was only achieved by molecular methods, generally available only in reference laboratories (29–32). More recently, MALDI-TOF has become available in routine laboratories and has been an efficient method for the early identification of fungi at the species level, including *Fusarium* (33–38). In one study



FIG 12 Disseminated skin lesions and toxemic appearance. Lesions at various stages of evolution: papular and nodular lesions with and without central necrosis. (Courtesy of Dr. Marilza Campos Magalhães, reproduced with permission.)

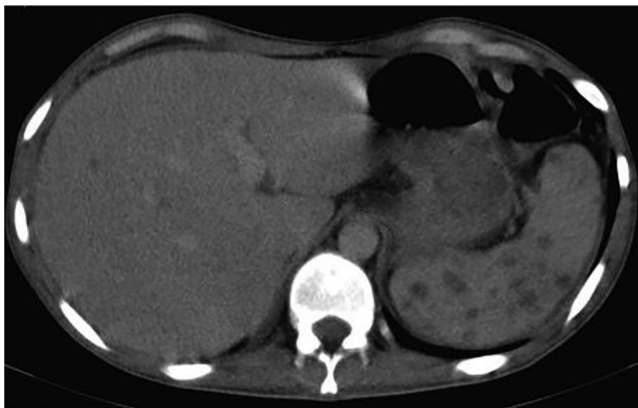


FIG 13 Nodules in the spleen.

evaluating 289 *Fusarium* isolates, MALDI-TOF correctly identified all species complexes and 82.8% of isolates at the species level (34).

Patients with invasive fusariosis may have a positive *Aspergillus* galactomannan (GMI) test in the serum. In a study, 11 patients with invasive fusariosis caused by distinct species had at least two GMI tests performed per week. Nine of these 11 patients had repeated positive GMI results, with an index ranging from 0.5 to 7.7, in the absence of isolation of *Aspergillus* species in the culture of bronchial secretions or of other respiratory specimens (163). In another study, 18 hematologic patients with invasive fusariosis and at least one GMI test performed within 2 days before or after the diagnosis were evaluated. In total, 15 (83%) had at least one positive GMI test, with sensitivity and specificity of 83% and 67%, respectively. The test was positive before the diagnosis of invasive fusariosis in 11 of the 15 cases (73%), at a median of 10 days (range 3–39) (164).



FIG 14 Endophthalmitis with periorbital cellulitis. (Courtesy of Dr. Clara Rosemberg, reproduced with permission.)



FIG 15 Endophthalmitis with periorbital cellulitis. (Courtesy of Dr. Clara Rosemberg, reproduced with permission.)

In another study reporting the characteristics of 65 cases of invasive fusariosis diagnosed in a multicenter prospective surveillance study, 10 had positive GMI tests. Three of these 10 patients had positive cultures for *Aspergillus*, suggesting that the positive tests represented mixed infection (165). More recently, investigators compared the cases of invasive aspergillosis to those with invasive fusariosis diagnosed in a Brazilian center. All patients were followed with serial (2–3×/week) serum GMI, with a median of 12 tests among 35 patients with aspergillosis and 13 tests among 26 patients with fusariosis.



FIG 16 Endophthalmitis with periorbital cellulitis. (Courtesy of Dr. Clara Roseberg, reproduced with permission.)

Serum GMI was positive in 89% of patients with aspergillosis and 73% of patients with fusariosis. The authors did not observe differences in the median number of positive tests, value of the first positive, or the peak GMI (154). Taken together, one should be cautious in interpreting a positive GMI in high-risk hematologic patients cared in areas with high incidence of invasive fusariosis.

Patients with invasive fusariosis may also have a positive 1,3-beta-D-glucan (BDG) test in the serum. In a study, serum samples of 13 patients with invasive fusariosis were tested for BDG. In all, 12 patients (92%) had at least one positive BDG test in the serum. Interestingly, the test was positive before the diagnosis of fusariosis in 11 patients, at a median of 10 days. However, given the high sensitivity and false-positive rates of the test (166), the positive predictive value was 7% only, considering two positive tests, suggesting that BDG is more useful to rule out rather than to confirm the diagnosis of invasive fusariosis (167). In another study, 81 blood samples from 15 patients with invasive fusariosis were tested for BDG. The rate of positivity was 58.3% (168).

In a study using a quantitative PCR assay as fungal biomarkers for the earlier diagnosis of invasive hematogenous fusariosis, the investigators detected *Fusarium* species in the blood in 14 of 15 patients with invasive fusariosis, at a median of 6 days before the diagnosis was confirmed by positive cultures or biopsy. The test was negative in all control samples, including patients with other IFD or those without IFD (168).

MANAGEMENT OF INVASIVE FUSARIOSIS

The management of invasive fusariosis depends on the immune status of the host and the form of the disease. We consider surgical debridement with or without systemic antifungal therapy in most cases of disease limited to the skin and soft tissues. By contrast, we always apply systemic antifungal therapy for infections in deeper organs and/or homogeneously disseminated with or without surgical debridement.

Antifungal susceptibility

Fusarium species typically exhibit high minimum inhibitory concentrations (MICs) to most antifungal agents, with higher MICs against azoles such as voriconazole, posaconazole, and isavuconazole, compared with amphotericin B. Fluconazole and the echinocandins have no activity against *Fusarium* species. In general, *Fusarium solani* and *Fusarium verticillioides* (FFSC) have higher MICs for azoles compared with other species.

Investigators evaluated 1,150 *Fusarium* species isolates belonging to various species complexes and established epidemiologic cutoff values (the highest MIC that would categorize an isolate as wild type, that is, without known mechanisms of resistance) (169). Among 608 FSSC isolates, the epidemiologic cutoff values ($\mu\text{g}/\text{mL}$) for amphotericin, voriconazole, and posaconazole were 4, 16, and 32, respectively. For FOSC, the epidemiologic cutoff for amphotericin B, voriconazole, and posaconazole was 4, 8 and 8 $\mu\text{g}/\text{mL}$, respectively (170).

Among 14 *Fusarium* species isolates tested, the MIC of isavuconazole was 16 $\mu\text{g}/\text{mL}$ in one isolate, and >16 $\mu\text{g}/\text{mL}$ in the remaining 13 isolates (171). In another study, the MIC₅₀ of 14 isolates was >8 $\mu\text{g}/\text{mL}$ for isavuconazole (172).

The *in vitro* activity of olorofim, an antifungal agent under development, was evaluated against 45 FOSC and 16 FSSC clinical isolates. Olorofim exhibited good *in vitro* activity for FOSC. When a 50% inhibition was considered, the MIC ranges were between 0.03 and 0.5 $\mu\text{g}/\text{mL}$ and 0.06 and >4 $\mu\text{g}/\text{mL}$ when a 100% inhibition was the endpoint. For FSSC, the activity was lower, with MIC ranges of 0.25–1 $\mu\text{g}/\text{mL}$ and 1– >4 $\mu\text{g}/\text{mL}$ at 50% and 100% inhibition, respectively. The authors concluded that since olorofim is a new class of agent with a novel mechanism of action, the endpoint for *in vitro* activity that correlates with *in vivo* activity is yet to be determined (173).

The activity of manogepix, a new antifungal drug with a new mechanism of action, was tested against 49 FOSC and 19 FSSC isolates and exhibited good *in vitro* activity. For FOSC, the MIC₅₀ range was ≤ 0.015 to 0.125 $\mu\text{g}/\text{mL}$, and for FSSC the MIC₅₀ range was ≤ 0.015 to 0.25 $\mu\text{g}/\text{mL}$. The same FSSC isolates exhibited MIC ranges of 0.25 to 2 $\mu\text{g}/\text{mL}$ for amphotericin B, 4 to >16 $\mu\text{g}/\text{mL}$ for posaconazole, 2 to >16 $\mu\text{g}/\text{mL}$ to voriconazole, and >16 $\mu\text{g}/\text{mL}$ for isavuconazole (174). Table 4 summarizes the susceptibility of *Fusarium* species to different antifungal agents.

Antifungal susceptibility tests are meant to help clinicians choose the most appropriate antimicrobial agent to treat their patients. An important question when we evaluate the results of antifungal susceptibility tests in invasive fusariosis is the apparent lack of correlation between *in vitro* data and clinical outcomes. This is illustrated by the high MICs exhibited by different *Fusarium* species against voriconazole and the good clinical response to this agent (9, 176). A multicenter study evaluated the correlation between MIC and outcome in 88 patients with invasive fusariosis (74 with hematologic disease). The most frequent treatment was voriconazole monotherapy (30.7%), followed by liposomal amphotericin B plus voriconazole (26.1%). A correlation between MIC and outcomes (survival or death) was not observed (177). The results of this study reflect on the recommendations of recently published global guidelines for the management of rare mold infections: strong recommendations for epidemiologic purposes but weak recommendations for the choice of primary therapy (178).

Prophylaxis

Mold-active prophylaxis with agents that may have activity against *Fusarium* species is considered standard of care in high-risk patients such as HCT recipients with GVHD and AML patients receiving intensive induction remission chemotherapy (179). Primary prophylaxis specifically for invasive fusariosis was evaluated in one study. In a previous publication, high-risk hematologic patients with superficial skin lesions in the feet (onychomycosis and/or interdigital intertrigo) at hospital admission with positive culture for *Fusarium* species were at an increased risk for invasive fusariosis (148). Subsequently, in a non-randomized trial, anti-mold prophylaxis (voriconazole or posaconazole) was given to 20 episodes at elevated risk (neutropenia or graft versus host disease) and compared with 219 episodes where fluconazole or no prophylaxis was given. Overall, anti-mold prophylaxis did not decrease the incidence of invasive fusariosis: 5.9% without versus 5% with anti-mold prophylaxis. However, 4 of 5 patients with superficial skin lesions with positive cultures for *Fusarium* species who did not receive anti-mold prophylaxis developed invasive fusariosis versus none of the six with anti-mold prophylaxis ($P = 0.01$)(180). Based on these data, primary anti-mold prophylaxis

TABLE 4 Antifungal susceptibility of *Fusarium* species to different antifungal agents

Drug	Reference	Method	No. isolates	MIC range (µg/mL)	MIC ₅₀ (µg/mL) ^a	MIC ₉₀ (µg/mL)	Mode (µg/mL)	GM (µg/mL)
<i>Fusarium solani</i> species complex								
Amphotericin B	(169)	CLSI	608	≤0.25 to 16	-	-	2	-
Itraconazole	(169)	CLSI	608	0.5 to ≥16	-	-	16	-
Posaconazole	(169)	CLSI	608	1 to ≥16	-	-	8	-
Voriconazole	(169)	CLSI	608	0.5 to ≥16	-	-	8	-
Isavuconazole	(175)	EUCAST	22	4 to ≥16	>16	>16	-	14.02
Olorofim	(173)	CLSI	16	1 to >4	>4	>4	>4	>4
Manogepix	(174)	CLSI	19	≤0.015*	-	-	-	≤0.015**
<i>Fusarium oxysporum</i> species complex								
Amphotericin B	(169)	CLSI	226	≤0.25 to 16	-	-	2	-
Itraconazole	(169)	CLSI	226	1 to ≥16	-	-	16	-
Posaconazole	(169)	CLSI	226	0.5–16	-	-	2	-
Voriconazole	(169)	CLSI	226	0.5 to ≥16	-	-	4	-
Isavuconazole	(175)	EUCAST	17	2 to ≥16	8	>16	-	9.41
Olorofim	(173)	CLSI	45	0.06 to >4	0.5	4	0.25	0.515
Manogepix	(174)	CLSI	49	≤0.015 to 0.03*	-	-	-	≤0.015**
<i>Fusarium fujikuroi</i> species complex								
Amphotericin B	(169)	CLSI	151	0.5–16	-	-	2	-
Itraconazole	(169)	CLSI	151	1 to ≥16	-	-	16	-
Posaconazole	(169)	CLSI	151	≤0.25 to ≥16	-	-	0.5	-
Voriconazole	(169)	CLSI	151	0.5 to ≥16	-	-	2	-
Isavuconazole	(175)	EUCAST	31	4 to ≥16	>16	>16	-	13.68

^aMIC = minimum inhibitory concentration; GM = geometric mean; *, minimal effective concentration; geometric mean MEC/MIC.

is recommended in high-risk hematologic patients who present on admission with superficial skin lesions with positive cultures for *Fusarium* species (178).

Secondary prophylaxis for patients who had a history of invasive fusariosis and underwent subsequent periods at risk (GVHD or neutropenia) was evaluated in a multicenter retrospective study of forty patients. Relapse of invasive fusariosis occurred in two of eight patients (25%) not receiving secondary prophylaxis and in 3 of 32 (9.4%) on prophylaxis. Among patients with a history of disseminated fusariosis, relapse occurred in two of two (100%) patients who were not on secondary prophylaxis and in 3 of 26 (11.5%) who were receiving secondary prophylaxis ($P = 0.03$) (181). Therefore, we recommend that patients with prior invasive fusariosis who will undergo additional immunosuppressive therapies receive secondary prophylaxis (mold-active azole or a lipid formulation of amphotericin B).

In addition to antifungal prophylaxis, measures to reduce patient exposure to *Fusarium* should be attempted, including the treatment of high-risk neutropenic patients in rooms with HEPA filter and positive pressure, and avoiding contact with reservoirs of *Fusarium*, including cleaning showers prior to use by high-risk patients and avoiding contact with contaminated tap water (2).

Prognostic factors

As with other IFDs, recovery of immunosuppression is an important prognostic factor. Prognostic factors in invasive fusariosis were evaluated in 84 patients with hematologic diseases. Multivariate analysis revealed two factors negatively impacting survival: persistent neutropenia (HR: 5.43) and receipt of corticosteroids (HR: 2.18). The 90-day probability of survival was 67% when both factors were absent and zero with both factors. Survival was 30% in patients recovering from neutropenia but receiving corticosteroids, and 4% in persistently neutropenic patients without corticosteroids (90). In another study, among 54 allogeneic HCT recipients with invasive fusariosis, univariate predictors of death were acute GVHD (HR: 2.05) and persistent neutropenia (HR: 3.64). By

multivariate analysis, only persistent neutropenia was significant (HR: 3.65) (91), a finding also reported by others (9, 99, 101).

Primary therapy

There are no randomized studies evaluating different treatment regimens for the treatment of invasive fusariosis. The largest series of invasive fusariosis ever published involved 44 centers from 11 countries in a retrospective study of 236 patients diagnosed between 1985 and 2011. Among the 206 patients who received treatment, the most frequent agent was deoxycholate amphotericin B (110 patients), followed by voriconazole (38 patients), and a lipid formulation of amphotericin B (liposomal 20, lipid complex 8, and colloidal dispersion 6). Combination therapy was given to 21 patients, mainly voriconazole plus amphotericin B. The 90-day probability of survival was not significantly different among patients receiving voriconazole or lipid amphotericin B (53% and 48%, respectively). By contrast, the 90-day probability of survival of patients receiving deoxycholate amphotericin B was poor (27%). There was no difference in the outcome of patients receiving monotherapy or combination therapy. Improved outcome was observed between patients treated between 2001 and 2010 and those treated before 2000 (9). Based on these results, recently published guidelines recommend either voriconazole (6 mg/kg twice daily on day 1, followed by 4 mg/kg twice daily subsequently) or a lipid formulation of amphotericin B (liposomal amphotericin B—3 mg/kg daily; amphotericin B lipid complex—5 mg/kg daily) as primary therapy. Combination therapy can also be considered, with a potential for early step-down to monotherapy (178).

The treatment of invasive fusariosis may be challenging because of the poor penetration of antifungal agents in infected tissues, such as endophthalmitis and arthritis. For endophthalmitis, we recommend systemic and intravitreal antifungal, sometimes with vitrectomy (158).

Evaluation of response to treatment

Assessing response to treatment relies on physical examination and laboratory studies. Signs of progression of fusariosis include the appearance of new skin lesions, signs of infection in new organs as well as persistent fungemia and elevated serum GMI and/or BDG. In patients with extensive disease, positron-emission tomography/computed tomography (PET/CT) can assist in response assessment (182–184).

Adjunctive therapies

Patients with necrotic lesions that are prone may benefit from surgical debridement of necrotic tissue (185). For the occasional cases of catheter-related fungemia, catheter removal and a short course of antifungal treatment result in a cure of infection (186).

The use of granulocyte transfusions as adjuvant treatment was evaluated in 11 neutropenic patients with invasive fusariosis. Clinical response was observed in 10 patients. The authors performed a literature review of 23 published cases, with a response rate of 30% (187). It is important to note that granulocyte transfusions represent a transient measure to allow time for neutrophil recovery.

Other ancillary measures include the use of granulocyte or granulocyte-monocyte colony-stimulating factors (G-CSF and GM-CSF), and interferon-gamma (16). More recently, the checkpoint inhibitor nivolumab was used as adjuvant treatment of patient with AML who developed invasive fusariosis in the lungs that subsequently disseminated to the liver and spleen, with marked improvement after four doses (188).

TABLE 5 Approach to the diagnosis and management of invasive fusariosis in high-risk hematologic patients^a

Action
Identify patients at elevated risk
Acute leukemia receiving induction chemotherapy for newly diagnosed or relapsed disease with profound (<100/mm ³) neutropenia; active smoking
Allogeneic HCT recipient
Pre-engraftment period: profound (<100/mm ³) neutropenia, cord blood HCT, ATG in the conditioning regimen
Post-engraftment period: receipt of corticosteroids or other immunosuppressive agents for the treatment of severe GVHD, CMV reactivation
The presence of skin breakdowns at sites of onychomycosis and/or interdigital intertrigo should increase the alertness
Consider the diagnosis of invasive fusariosis if
Skin lesions or unexplained myalgia
New pulmonary infiltrates
Sinusitis
Endophthalmitis
Positive blood culture for mold
Diagnostic workup
Prompt biopsy of a skin lesion, with direct examination, culture, and histopathology
Treat immediately if
Presence of hyaline hyphae on the direct exam of a fragment of skin biopsy
Positive blood culture for a mold

^aHCT = hematopoietic cell transplantation; ATG = anti-thymocyte globulin; GVHD = graft versus host disease; CMV = cytomegalovirus.

Approach to the diagnosis and treatment of invasive fusariosis in hematologic patients

The first step in the approach to the diagnosis of invasive fusariosis is to identify the typical scenario/patient at risk: patients with AML or ALL receiving induction chemotherapy for newly diagnosed or relapsed disease, and allogeneic HCT recipients with profound (<100/mm³) neutropenia or, in non-neutropenic HCT recipients, receipt of corticosteroids or other immunosuppressive agents for the treatment of severe GVHD. The presence of skin breakdowns (onychomycosis and/or interdigital intertrigo) should increase alertness, as well as the presence of risk factors such as active smoking in AML, and receipt of anti-thymocyte globulin, cord blood as a source of stem cells or CMV reactivation in allogeneic HCT. In these scenarios, clinicians should strongly consider the diagnosis of invasive fusariosis in the presence of skin lesions, new pulmonary infiltrates, sinusitis, endophthalmitis, or a positive blood culture for mold or elevation of serially obtained serum markers of IFD. In the presence of skin lesions, it is important to promptly obtain a biopsy with direct examination, culture, and histopathology; direct examination is the fastest way of achieving a presumptive diagnosis of invasive fusariosis. Anti-mold-active antifungal therapy should be immediately started if direct examination shows hyaline hyphae, or in the presence of positive blood culture for a mold and/or increasing serum markers of IFD (Table 5).

CONCLUSIONS

Invasive fusariosis is a serious IFD, affecting both immunocompetent and, more frequently, immunocompromised patients. In immunocompetent individuals, the disease is usually localized. Immunocompromised patients more prone to develop invasive fusariosis are patients with acute leukemia receiving chemotherapeutic regimens for induction remission and allogeneic HCT recipients. The disease is usually disseminated with multiple painful metastatic skin lesions, positive blood cultures, and lung involvement. Currently available antifungal agents have poor *in vitro* activity

against *Fusarium* species, but a clear-cut correlation between *in vitro* activity and clinical effectiveness does not exist. The outcome of invasive fusariosis is largely dependent on the recovery of immunosuppression, especially neutrophil recovery in neutropenic patients.

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REFERENCES

- Kadaifciler DG, Demirel R. 2018. Fungal contaminants in man-made water systems connected to municipal water. *J Water Health* 16:244–252. <https://doi.org/10.2166/wh.2018.272>
- Anaissie EJ, Kuchar RT, Rex JH, Francesconi A, Kasai M, Müller FC, Lozano - Chiu M, Summerbell RC, Dignani MC, Chanock SJ, Walsh TJ. 2001. Fusariosis associated with pathogenic *Fusarium* species colonization of a hospital water system: a new paradigm for the epidemiology of opportunistic mold infections. *CLIN INFECT DIS* 33:1871–1878. <https://doi.org/10.1086/324501>
- Dean R, Van Kan JAL, Pretorius ZA, Hammond-Kosack KE, Di Pietro A, Spanu PD, Rudd JJ, Dickman M, Kahmann R, Ellis J, Foster GD. 2012. The top 10 fungal pathogens in molecular plant pathology. *Mol Plant Pathol* 13:414–430. <https://doi.org/10.1111/j.1364-3703.2011.00783.x>
- Sáenz V, Alvarez-Moreno C, Pape PL, Restrepo S, Guarro J, Ramírez AMC. 2020. A one health perspective to recognize *Fusarium* as important in clinical practice. *JoF* 6:235. <https://doi.org/10.3390/jof6040235>
- Uemura EVG, Barbosa M dos S, Simionatto S, Al-Harrasi A, Al-Hatmi AMS, Rossato L. 2022. Onychomycosis caused by *Fusarium* species. *JoF* 8:360. <https://doi.org/10.3390/jof8040360>
- Thomas PA, Kalliamurthy J. 2013. Mycotic keratitis: epidemiology, diagnosis and management. *Clin Microbiol Infect* 19:210–220. <https://doi.org/10.1111/1469-0691.12126>
- Nucci M, Varon AG, Garnica M, Akiti T, Barreiros G, Trope BM, Nouér SA. 2013. Increased incidence of invasive fusariosis with cutaneous portal of entry, Brazil. *Emerg Infect Dis* 19:1567–1572. <https://doi.org/10.3201/eid1910.120847>
- Walther G, Stasch S, Kaerger K, Hamprecht A, Roth M, Cornely OA, Geerling G, Mackenzie CR, Kurzai O, von Lilienfeld-Toal M. 2017. *Fusarium* keratitis in Germany. *J Clin Microbiol* 55:2983–2995. <https://doi.org/10.1128/JCM.00649-17>
- Nucci M, Marr KA, Vehreschild M, de Souza CA, Velasco E, Cappellano P, Carlesse F, Queiroz-Telles F, Sheppard DC, Kindo A, Cesaro S, Hamerschlag N, Solza C, Heinz WJ, Schaller M, Atalla A, Arikian-Akdagli S, Bertz H, Galvão Castro C, Herbrecht R, Hoenigl M, Härter G, Hermansen NEU, Josting A, Pagano L, Salles MJC, Mossad SB, Ogunc D, Pasqualotto AC, Araujo V, Troke PF, Lortholary O, Cornely OA, Anaissie E. 2014. Improvement in the outcome of invasive fusariosis in the last decade. *Clin Microbiol Infect* 20:580–585. <https://doi.org/10.1111/1469-0691.12409>
- Wickern GM. 1993. *Fusarium* allergic fungal sinusitis. *J Allergy Clin Immunol* 92:624–625. [https://doi.org/10.1016/0091-6749\(93\)90087-v](https://doi.org/10.1016/0091-6749(93)90087-v)
- Qiu J, Xu J, Shi J. 2019. *Fusarium* toxins in Chinese wheat since the 1980s. *Toxins (Basel)* 11:248. <https://doi.org/10.3390/toxins11050248>
- Backman KS, Roberts M, Patterson R. 1995. Allergic bronchopulmonary mycosis caused by *Fusarium* vasinfectum. *Am J Respir Crit Care Med* 152:1379–1381. <https://doi.org/10.1164/ajrccm.152.4.7551398>
- Saini SK, Boas SR, Jerath A, Roberts M, Greenberger PA. 1998. Allergic bronchopulmonary mycosis to *Fusarium* vasinfectum in a child. *Ann Allergy Asthma Immunol* 80:377–380. [https://doi.org/10.1016/S1081-1206\(10\)62986-9](https://doi.org/10.1016/S1081-1206(10)62986-9)
- Ramirez RM, Jacobs RL. 2014. Hypersensitivity pneumonitis by *Fusarium* vasinfectum in a home environment. *J Allergy Clin Immunol Pract* 2:483–484. <https://doi.org/10.1016/j.jaip.2014.04.002>
- Dickson SD, Tankersley MS. 2015. Fatal hypersensitivity pneumonitis from exposure to *Fusarium* vasinfectum in a home environment: a case report. *Int Arch Allergy Immunol* 166:150–153. <https://doi.org/10.1159/000377631>
- Antachopoulos C, Katragkou A, Roilides E. 2012. Immunotherapy against invasive mold infections. *Immunotherapy* 4:107–120. <https://doi.org/10.2217/imt.11.159>
- Shoham S, Levitz SM. 2005. The immune response to fungal infections. *Br J Haematol* 129:569–582. <https://doi.org/10.1111/j.1365-2141.2005.05397.x>
- Gaviria JM, van Burik JA, Dale DC, Root RK, Liles WC. 1999. Comparison of interferon-gamma, granulocyte colony-stimulating factor, and granulocyte-macrophage colony-stimulating factor for priming leukocyte-mediated hyphal damage of opportunistic fungal pathogens. *J Infect Dis* 179:1038–1041. <https://doi.org/10.1086/314679>
- Winn RM, Gil-Lamaignere C, Maloukou A, Roilides E, Network E. 2003. Interactions of human phagocytes with moulds *Fusarium* spp. and *verticillium nigrescens* possessing different pathogenicity. *Med Mycol* 41:503–509. <https://doi.org/10.1080/1369378030001615394>
- Boutati EI, Anaissie EJ. 1997. *Fusarium*, a significant emerging pathogen in patients with hematologic malignancy: ten years' experience at a cancer center and implications for management. *Blood* 90:999–1008.
- Khan Z, Ahmad S, Alfouzan W, Joseph L, Varghese S. 2019. Demonstration of adventitious Sporulation in *Fusarium Petroliphilum*

- Onychomycosis. *Mycopathologia* 184:303–308. <https://doi.org/10.1007/s11046-019-0318-5>
22. Legrand C, Anaissie E, Hashem R, Nelson P, Bodey GP, Ro J. 1991. Experimental fusarial hyalohyphomycosis in a murine model. *J Infect Dis* 164:944–948. <https://doi.org/10.1093/infdis/164.5.944>
 23. Costa MI, Vilugron Rodrigues FA, Veiga FF, Jarros IC, Kischkel B, Negri M, Alexandrino Becker TC, Svidzinski TIE. 2019. Effects of intratracheal *Fusarium solani* inoculation in immunocompetent mice. *Microb Pathog* 128:317–322. <https://doi.org/10.1016/j.micpath.2019.01.020>
 24. Al-Hatmi AMS, Meis JF, de Hoog GS, Heitman J. 2016. *Fusarium*: molecular diversity and intrinsic drug resistance. *PLoS Pathog* 12:e1005464. <https://doi.org/10.1371/journal.ppat.1005464>
 25. Al-Hatmi AM, Hagen F, Menken SB, Meis JF, de Hoog GS. 2016. Global molecular epidemiology and genetic diversity of *Fusarium*, a significant emerging group of human opportunists from. *Emerg Microbes Infect* 5:e124. <https://doi.org/10.1038/emi.2016.126>
 26. Herkert PF, Al-Hatmi AMS, de Oliveira Salvador GL, Muro MD, Pinheiro RL, Nucci M, Queiroz-Telles F, de Hoog GS, Meis JF. 2019. Molecular characterization and antifungal susceptibility of clinical *Fusarium* species from Brazil. *Front Microbiol* 10:737. <https://doi.org/10.3389/fmicb.2019.00737>
 27. da Rosa PD, Aquino V, Fuentesfria AM, Goldani LZ. 2021. Diversity of *Fusarium* species causing invasive and disseminated infections. *J Mycol Med* 31:101137. <https://doi.org/10.1016/j.mycmed.2021.101137>
 28. Tortorano AM, Prigitano A, Esposto MC, Arsic Arsenijevic V, Kolarovic J, Ivanovic D, Paripovic L, Klingspor L, Nordøy I, Hamal P, Arkan Akdagli S, Ossi C, Grancini A, Cavanna C, Lo Cascio G, Scarparo C, Candoni A, Caira M, Drogari Apiranthitou M, ECMM Working Group. 2014. European confederation of medical mycology (ECMM) epidemiological survey on invasive infections due to *Fusarium* species in Europe. *Eur J Clin Microbiol Infect Dis* 33:1623–1630. <https://doi.org/10.1007/s10096-014-2111-1>
 29. de Souza M, Matsuzawa T, Sakai K, Muraosa Y, Lyra L, Busso-Lopes AF, Levin ASS, Schreiber AZ, Mikami Y, Gonoï T, Kamei K, Moretti ML, Trabasso P. 2017. Comparison of DNA microarray, loop-mediated isothermal amplification (LAMP) and real-time PCR with DNA sequencing for identification of *Fusarium* Spp. obtained from patients with hematologic malignancies. *Mycopathologia* 182:625–632. <https://doi.org/10.1007/s11046-017-0129-5>
 30. Alastruey-Izquierdo A, Alcazar-Fuoli L, Rivero-Menéndez O, Ayats J, Castro C, García-Rodríguez J, Goterris-Bonet L, Ibáñez-Martínez E, Linares-Sicilia MJ, Martín-Gomez MT, Martín-Mazuelos E, Pelaez T, Peman J, Rezusta A, Rojo S, Tejero R, Anza DV, Viñuelas J, Zapico MS, Cuenca-Estrella M, the FILPOP2 Project from GEMICOMED (SEIMC) and REIPI. 2018. Molecular identification and susceptibility testing of molds isolated in a prospective surveillance of triazole resistance in Spain (FILPOP2 study). *Antimicrob Agents Chemother* 62:e00358-18. <https://doi.org/10.1128/AAC.00358-18>
 31. Gaviria-Rivera A, Giraldo-López A, Santa-Cardona C, Cano-Restrepo L. 2018. Molecular identification of clinical isolates of *Fusarium* in Colombia. *Rev Salud Publica (Bogota)* 20:94–102. <https://doi.org/10.15446/rsap.V20n1.51923>
 32. Thomas B, Contet Audonneau N, Machouart M, Debourgogne A. 2019. Molecular identification of *Fusarium* species complexes: which gene and which database to choose in clinical practice. *J Mycol Med* 29:56–58. <https://doi.org/10.1016/j.mycmed.2019.01.003>
 33. De Carolis E, Posteraro B, Lass-Flörl C, Vella A, Florio AR, Torelli R, Girmenia C, Colozza C, Tortorano AM, Sanguinetti M, Fadda G. 2012. Species identification of *Aspergillus*, *Fusarium* and *Mucorales* with direct surface analysis by matrix-assisted laser desorption ionization time-of-flight mass spectrometry. *Clin Microbiol Infect* 18:475–484. <https://doi.org/10.1111/j.1469-0691.2011.03599.x>
 34. Triest D, Stubbe D, De Cremer K, Piérard D, Normand A-C, Piarroux R, Detandt M, Hendrickx M. 2015. Use of matrix-assisted laser desorption ionization-time of flight mass spectrometry for identification of molds of the *Fusarium* genus. *J Clin Microbiol* 53:465–476. <https://doi.org/10.1128/JCM.02213-14>
 35. Sanguinetti M, Posteraro B. 2017. Identification of molds by matrix-assisted laser desorption ionization-time of flight mass spectrometry. *J Clin Microbiol* 55:369–379. <https://doi.org/10.1128/JCM.01640-16>
 36. Walsh TJ, McCarthy MW. 2019. The expanding use of matrix-assisted laser desorption/ionization-time of flight mass spectrometry in the diagnosis of patients with mycotic diseases. *Expert Rev Mol Diagn* 19:241–248. <https://doi.org/10.1080/14737159.2019.1574572>
 37. Normand A-C, Blaize M, Imbert S, Packeu A, Becker P, Fekkar A, Stubbe D, Piarroux R, Hanson KE. 2021. Identification of molds with matrix-assisted laser desorption ionization-time of flight mass spectrometry: performance of the newly developed MSI-2 application in comparison with the bruker filamentous fungi database and MSI-1. *J Clin Microbiol* 59:e0129921. <https://doi.org/10.1128/JCM.01299-21>
 38. Barker KR, Kus JV, Normand A-C, Gharabaghi F, McTaggart L, Rotstein C, Richardson SE, Campigotto A, Tadros M. 2022. A practical workflow for the identification of *Aspergillus*, *Fusarium*, and *Mucorales* by MALDI-TOF MS: database, medium, and incubation optimization. *J Clin Microbiol* 60:e0103222. <https://doi.org/10.1128/jcm.01032-22>
 39. Wheeler MS, McGinnis MR, Schell WA, Walker DH. 1981. *Fusarium* infection in burned patients. *Am J Clin Pathol* 75:304–311. <https://doi.org/10.1093/ajcp/75.3.304>
 40. Latenser BA. 2003. *Fusarium* infections in burn patients: a case report and review of the literature. *J Burn Care Rehabil* 24:285–288. <https://doi.org/10.1097/01.BCR.0000085845.20730.AB>
 41. Rosanova MT, Brizuela M, Villasboas M, Guarracino F, Alvarez V, Santos P, Finquelievich J. 2016. *Fusarium* spp infections in a pediatric burn unit: nine years of experience. *Braz J Infect Dis* 20:389–392. <https://doi.org/10.1016/j.bjid.2016.04.004>
 42. de Oliveira Ramos Silva A, Barata ESR, Ferraz TLL, Batista FP, Medeiros ACR, Ferraz CE, Inacio CP. 2021. Cutaneous fusariosis in immunocompetent farmer. *Mycopathologia* 186:465–467. <https://doi.org/10.1007/s11046-021-00548-y>
 43. Warkentien T, Rodriguez C, Lloyd B, Wells J, Weintrob A, Li P, Seillier-Moiseiwitsch F, Fleming M, Tribble DR, Infectious disease clinical research program trauma infectious disease outcomes study G. 2012. Invasive mold infections following combat-related injuries. *Clin Infect Dis* 55:1441–1449. <https://doi.org/10.1093/cid/cis749>
 44. van Dijk E, van den Berg WH, Landwehr AJ. 1980. *Fusarium solani* infection of a hypertensive leg ulcer in a diabetic. *Mykosen* 23:603–606. <https://doi.org/10.1111/j.1439-0507.1980.tb02568.x>
 45. Taj-Aldeen SJ, Gene J, Al Bozom I, Buzina W, Cano JF, Guarro J. 2006. Gangrenous necrosis of the diabetic foot caused by *Fusarium acutatum*. *Med Mycol* 44:547–552. <https://doi.org/10.1080/13693780500543246>
 46. Dutta P, Premkumar A, Chakrabarti A, Shah VN, Behera A, De D, Rudramurthy SM, Bhansali A. 2013. *Fusarium* calciforme infection of foot in a patient with type 2 diabetes mellitus: a case report and review of the literature. *Mycopathologia* 176:225–232. <https://doi.org/10.1007/s11046-013-9646-z>
 47. João AL, Lencastre A, Dutra E, Pessoa E Costa T, Formiga A, Neves J. 2021. *Fusarium* spp.-an emerging pathogen in chronic diabetic ulcer: case report and review of the literature. *Int J Low Extrem Wounds* 20:67–72. <https://doi.org/10.1177/1534734619879030>
 48. Mansur AT, Artunkal S, Ener B. 2011. *Fusarium* oxysporum infection of stasis ulcer: eradication with measures aimed to improve stasis. *Mycoses* 54:e205–7. <https://doi.org/10.1111/j.1439-0507.2009.01800.x>
 49. Samrah S, Sweidan A, Aleshawi A, Ayesh M. 2020. *Fusarium*-induced cellulitis in an immunocompetent patient with sickle cell disease: a case report. *J Investig Med High Impact Case Rep* 8:2324709620934303. <https://doi.org/10.1177/2324709620934303>
 50. Bourguignon RL, Walsh AF, Flynn JC, Baro C, Spinos E. 1976. *Fusarium* species osteomyelitis. Case report. *J Bone Joint Surg Am* 58:722–723.
 51. Nuovo MA, Simmonds JE, Chacho MS, McKittrick J. 1988. *Fusarium solani* osteomyelitis with probable nosocomial spread. *Am J Clin Pathol* 90:738–741. <https://doi.org/10.1093/ajcp/90.6.738>
 52. Smith M, McGinnis MR. 2005. *Fusarium* sporodochia on cutaneous wounds. *Med Mycol* 43:83–86. <https://doi.org/10.1080-13693780410001712089>
 53. Sierra-Hoffman M, Paltiyevich-Gibson S, Carpenter JL, Hurley DL. 2005. *Fusarium* osteomyelitis: case report and review of the literature. *Scand J Infect Dis* 37:237–240. <https://doi.org/10.1080/00365540410021036>
 54. Page JC, Friedlander G, Dockery GL. 1982. Postoperative *Fusarium* osteomyelitis. *J Foot Surg* 21:174–176.
 55. Jakle C, Leek JC, Olson DA, Robbins DL. 1983. Septic arthritis due to *Fusarium solani*. *J Rheumatol* 10:151–153.

56. Graddon JD, Lerman A, Lutwick LI. 1990. Septic arthritis due to *Fusarium moniliforme*. Rev Infect Dis 12:716–717. <https://doi.org/10.1093/clinids/12.4.716>
57. Rowsey JJ, Acers TE, Smith DL, Mohr JA, Newsom DL, Rodriguez J. 1979. *Fusarium oxysporum* endophthalmitis. Arch Ophthalmol 97:103–105. <https://doi.org/10.1001/archophth.1979.01020010043010>
58. Pflugfelder SC, Flynn HW, Zwickey TA, Forster RK, Tsiligianni A, Culbertson WW, Mandelbaum S. 1988. Exogenous fungal endophthalmitis. Ophthalmology 95:19–30. [https://doi.org/10.1016/S0161-6420\(88\)33229-X](https://doi.org/10.1016/S0161-6420(88)33229-X)
59. Buchta V, Feuermannová A, Váša M, Bašková L, Kutová R, Kubátová A, Vejsová M. 2014. Outbreak of fungal endophthalmitis due to *Fusarium oxysporum* following cataract surgery. Mycopathologia 177:115–121. <https://doi.org/10.1007/s11046-013-9721-5>
60. Güngel H, Eren MH, Pinarci EY, Altan C, Baylanççek DO, Kara N, Gürsel T, Yegenoglu Y, Susever S. 2011. An outbreak of *Fusarium solani* endophthalmitis after cataract surgery in an eye training and research hospital in Istanbul. Mycoses 54:e767–74. <https://doi.org/10.1111/j.1439-0507.2011.02019.x>
61. Dursun D, Fernandez V, Miller D, Alfonso EC. 2003. Advanced *Fusarium* keratitis progressing to endophthalmitis. Cornea 22:300–303. <https://doi.org/10.1097/00003226-200305000-00004>
62. Rosenberg KD, Flynn HW, Alfonso EC, Miller D. 2006. *Fusarium* endophthalmitis following keratitis associated with contact lenses. Ophthalmic Surg Lasers Imaging 37:310–313. <https://doi.org/10.3928/15428877-20060701-08>
63. Kerr CM, Perfect JR, Craven PC, Jorgensen JH, Drutz DJ, Shelburne JD, Gallis HA, Gutman RA. 1983. Fungal peritonitis in patients on continuous ambulatory peritoneal dialysis. Ann Intern Med 99:334–336. <https://doi.org/10.7326/0003-4819-99-3-334>
64. Rippon JW, Larson RA, Rosenthal DM, Clayman J. 1988. Disseminated cutaneous and peritoneal hyalohyphomycosis caused by *Fusarium* species: three cases and review of the literature. Mycopathologia 101:105–111. <https://doi.org/10.1007/BF00452895>
65. Chiaradia V, Schinella D, Pascoli L, Tesio F, Santini GF. 1990. *Fusarium* peritonitis in peritoneal dialysis: report of two cases. Microbiologica 13:77–78.
66. Flynn JT, Meislich D, Kaiser BA, Polinsky MS, Baluarte HJ. 1996. *Fusarium* peritonitis in a child on peritoneal dialysis: case report and review of the literature. Perit Dial Int 16:52–57.
67. Jorens PG, Van Den Heuvel PA, Van Cauwelaert PA, Parizel GA, Mertens AN. 1990. *Fusarium* endocarditis involving aortic valve following coronary artery surgery. Eur Heart J 11:476–478. <https://doi.org/10.1093/oxfordjournals.eurheartj.a059732>
68. Camin AM, Michelet C, Langanay T, de Place C, Chevrier S, Guého E, Guiguen C. 1999. Endocarditis due to *Fusarium dimerum* four years after coronary artery bypass grafting. Clin Infect Dis 28:150. <https://doi.org/10.1086/517184>
69. Das L, Dahiya D, Gupta K, Prakash M, Malhotra B, Rastogi A, Choudhary H, Rudramurthy SM, Dutta P. 2021. Eumycetoma of the foot due to *Fusarium solani* in a person with diabetes mellitus: report of a case and review of literature. Mycopathologia 186:277–288. <https://doi.org/10.1007/s11046-020-00524-y>
70. Correia C, Ferreira J, Soares-de-Almeida L, Filipe P. 2022. An unusual cause of eumycetoma - *Fusarium solani* keratoplasticum. Actas Dermosifiliogr 113:899. <https://doi.org/10.1016/j.ad.2021.04.013>
71. Kurien M, Anandi V, Raman R, Brahmadathan K. 1992. Maxillary sinus *Fusariosis* in immunocompetent hosts. J Laryngol Otol 106:733–736. <https://doi.org/10.1017/s0022215100120729>
72. Macêdo DPC, Neves RP, Fontan J, Souza-Motta CM, Lima D. 2008. A case of invasive rhinosinusitis by *Fusarium verticillioides* (saccardo) nirenberg in an apparently immunocompetent patient. Med Mycol 46:499–503. <https://doi.org/10.1080/13693780701861462>
73. Gorman SR, Magiorakos A-P, Zimmerman SK, Craven DE. 2006. *Fusarium oxysporum* pneumonia in an immunocompetent host. South Med J 99:613–616. <https://doi.org/10.1097/01.smj.0000217160.63313.63>
74. Chae SY, Park HM, Oh TH, Lee JE, Lee H-J, Jeong WG, Kim Y-H. 2020. *Fusarium* species causing invasive fungal pneumonia in an immunocompetent patient: a case report. J Int Med Res 48:300060520976475. <https://doi.org/10.1177/0300060520976475>
75. Dananché C, Cassier P, Sautour M, Gautheron N, Wegrzyn J, Perraud M, Biennu A-L, Nicolle M-C, Boibieux A, Vanhems P. 2015. Fungaemia caused by *Fusarium proliferatum* in a patient without definite immunodeficiency. Mycopathologia 179:135–140. <https://doi.org/10.1007/s11046-014-9817-6>
76. Abramowsky CR, Quinn D, Bradford WD, Conant NF. 1974. Systemic infection by *Fusarium* in a burned child. the emergence of a saprophytic strain. J Pediatr 84:561–564. [https://doi.org/10.1016/s0022-3476\(74\)80681-5](https://doi.org/10.1016/s0022-3476(74)80681-5)
77. Sander A, Beyer U, Amberg R. 1998. Systemic *Fusarium oxysporum* infection in an immunocompetent patient with an adult respiratory distress syndrome (ARDS) and extracorporeal membrane oxygenation (ECMO). Mycoses 41:109–111. <https://doi.org/10.1111/j.1439-0507.1998.tb00310.x>
78. Testerman GM, Steagald MK, Colquitt LA, Maki A. 2008. Disseminated *Fusarium* infection in a multiple trauma patient. South Med J 101:320–323. <https://doi.org/10.1097/SMJ.0b013e318164e392>
79. Kang Y, Li L, Zhu J, Zhao Y, Zhang Q. 2013. Identification of *Fusarium* from a patient with fungemia after multiple organ injury. Mycopathologia 176:151–155. <https://doi.org/10.1007/s11046-013-9664-x>
80. Garcia RR, Min Z, Narasimhan S, Bhanot N. 2015. *Fusarium* brain abscess: case report and literature review. Mycoses 58:22–26. <https://doi.org/10.1111/myc.12271>
81. Okura Y, Kawamura N, Okano M, Toita N, Takezaki S, Yamada M, Kobayashi I, Ariga T. 2015. *Fusarium falciforme* infection in a patient with chronic granulomatous disease: unique long-term course of epidural abscess. Pediatr Int 57:e4–e6. <https://doi.org/10.1111/ped.12458>
82. Chen Y-J, Chou C-L, Lai K-J, Lin Y-JL. 2017. *Fusarium* brain abscess in a patient with diabetes mellitus and liver cirrhosis. Acta Neurol Taiwan 26(3):128–132.
83. Kassar O, Charfi M, Trabelsi H, Hammami R, Elloumi M. 2016. *Fusarium solani* endocarditis in an acute leukemia patient. Med Mal Infect 46:57–59. <https://doi.org/10.1016/j.medmal.2015.11.004>
84. Ghosh S, Phillips A, Ghosh S, Singh A. 2018. Native valve endocarditis, *Fusarium* and end-stage renal disease. BMJ Case Rep 2018:bcr-2017. <https://doi.org/10.1136/bcr-2017-223290>
85. Nambiar P, Cober E, Johnson L, Brizendine K. 2018. Fatal *Fusarium* infection manifesting as osteomyelitis following previous treatment with amphotericin B in a multi-visceral transplant: case report and review of *Fusarium* infections in solid organ transplantation. Transpl Infect Dis 20:e12872. <https://doi.org/10.1111/tid.12872>
86. Edupuganti S, Roupael N, Mehta A, Eaton M, Heller JG, Bressler A, Brandt M, O'Donnell K. 2011. *Fusarium falciforme* vertebral abscess and osteomyelitis: case report and molecular classification. J Clin Microbiol 49:2350–2353. <https://doi.org/10.1128/JCM.02547-10>
87. Dallé Rosa P, Ramirez-Castrillon M, Valente P, Meneghello Fuentesria A, Van Diepeningen AD, Goldani LZ. 2018. *Fusarium riograndense* sp. nov., a new species in the *Fusarium solani* species complex causing fungal rhinosinusitis. J Mycol Med 28:29–35. <https://doi.org/10.1016/j.mycmed.2018.01.004>
88. Yamasmith E, Chongtrakool P, Chayakulkeeree M. 2020. Isolated pulmonary fusariosis caused by neocosmospora pseudensiformis in a liver transplant recipient: a case report and review of the literature. Transpl Infect Dis 22:e13344. <https://doi.org/10.1111/tid.13344>
89. Anaissie E, Kantarjian H, Ro J, Hopfer R, Rolston K, Fainstein V, Bodey G. 1988. The emerging role of *Fusarium* infections in patients with cancer. Medicine (Baltimore) 67:77–83. <https://doi.org/10.1097/00005792-198803000-00001>
90. Nucci M, Anaissie EJ, Queiroz-Telles F, Martins CA, Trabasso P, Solza C, Mangini C, Simões BP, Colombo AL, Vaz J, Levy CE, Costa S, Moreira VA, Oliveira JS, Paraguay N, Duboc G, Voltarelli JC, Maiolino A, Pasquini R, Souza CA. 2003. Outcome predictors of 84 patients with hematologic malignancies and *Fusarium* infection. Cancer 98:315–319. <https://doi.org/10.1002/cncr.11510>
91. Nucci M, Marr KA, Queiroz-Telles F, Martins CA, Trabasso P, Costa S, Voltarelli JC, Colombo AL, Imhof A, Pasquini R, Maiolino A, Souza CA, Anaissie E. 2004. *Fusarium* infection in hematopoietic stem cell transplant recipients. Clin Infect Dis 38:1237–1242. <https://doi.org/10.1086/383319>

92. Nucci M, Garnica M, Gloria AB, Lehugeur DS, Dias VCH, Palma LC, Cappellano P, Fertrin KY, Carlesse F, Simões B, Bergamasco MD, Cunha CA, Seber A, Ribeiro MPD, Queiroz-Telles F, Lee MLM, Chauffaille ML, Silla L, de Souza CA, Colombo AL. 2013. Invasive fungal diseases in haematopoietic cell transplant recipients and in patients with acute myeloid leukaemia or myelodysplasia in Brazil. *Clin Microbiol Infect* 19:745–751. <https://doi.org/10.1111/1469-0691.12002>
93. Souza L, Nouér SA, Morales H, Simões B, Solza C, Queiroz-Telles F, Nucci M. 2021. Epidemiology of invasive fungal disease in haematologic patients. *Mycoses* 64:252–256. <https://doi.org/10.1111/myc.13205>
94. Aquino VR, Verçosa EB, Falhauber G, Lunardi LW, Silla L, Pasqualotto AC. 2010. Distribution of filamentous fungi causing invasive fungal disease at the haematological unit. *Braz J Infect Dis* 14:277–280. <https://doi.org/10.1590/S1413-86702010000300013>
95. Bergamasco MD, Pereira CAP, Arrais-Rodrigues C, Ferreira DB, Baiocchi O, Kerbauy F, Nucci M, Colombo AL. 2021. Epidemiology of invasive fungal diseases in patients with hematologic malignancies and hematopoietic cell transplantation recipients managed with an antifungal diagnostic driven approach. *J Fungi (Basel)* 7:588. <https://doi.org/10.3390/jof7080588>
96. Girmenia C, Pagano L, Corvatta L, Mele L, del Favero A, Martino P. 2000. The epidemiology of fusariosis in patients with haematological diseases. *gimema infection programme*. *Br J Haematol* 111:272–276. <https://doi.org/10.1046/j.1365-2141.2000.02312.x>
97. Pagano L, Caira M, Candoni A, Offidani M, Fianchi L, Martino B, Pastore D, Picardi M, Bonini A, Chierichini A, Fanci R, Caramatti C, Invernizzi R, Mattei D, Mitra ME, Melillo L, Aversa F, Van Lint MT, Falucci P, Valentini CG, Girmenia C, Nosari A. 2006. The epidemiology of fungal infections in patients with hematologic malignancies: the SEIFEM-2004 study. *Haematologica* 91:1068–1075.
98. Pagano L, Caira M, Nosari A, Van Lint MT, Candoni A, Offidani M, Aloisi T, Irrera G, Bonini A, Picardi M, Caramatti C, Invernizzi R, Mattei D, Melillo L, de Waure C, Reddiconto G, Fianchi L, Valentini CG, Girmenia C, Leone G, Aversa F. 2007. Fungal infections in recipients of hematopoietic stem cell transplants: Results of the SEIFEM B-2004 study—sorveglianza epidemiologica infezioni fungine nelle emopatie maligne. *Clin Infect Dis* 45:1161–1170. <https://doi.org/10.1086/522189>
99. Kontoyiannis DP, Marr KA, Park BJ, Alexander BD, Anaissie EJ, Walsh TJ, Ito J, Andes DR, Baddley JW, Brown JM, Brumble LM, Freifeld AG, Hadley S, Herwaldt LA, Kauffman CA, Knapp K, Lyon GM, Morrison VA, Papanicolaou G, Patterson TF, Perl TM, Schuster MG, Walker R, Wannemuehler KA, Wingard JR, Chiller TM, Pappas PG. 2010. 2001–2006: Overview of the transplant-associated infection surveillance network (TRANSNET) database. *Clin infect dis* 50:1091–1100. <https://doi.org/10.1086/651263>
100. Hardak E, Fuchs E, Geffen Y, Zuckerman T, Oren I. 2020. Clinical spectrum, diagnosis and outcome of rare fungal infections in patients with hematological malignancies: experience of 15-year period from a single tertiary medical center. *Mycopathologia* 185:347–355. <https://doi.org/10.1007/s11046-020-00436-x>
101. Perez-Nadales E, Alastruey-Izquierdo A, Linares-Sicilia MJ, Soto-Debran JC, Abdala E, Garcia-Rodriguez J, Montejó M, Muñoz P, Lleti MS, Rezusta A, de Pipaon MRP, Yanez L, Merino E, Campos-Herrero MI, Costa-Mateo JM, Fortun J, Garcia-Lozano T, Garcia-Vidal C, Fernandez-Ruiz M, Sanchez-Reus F, Castro-Mendez C, Guerrero-Lozano I, Soler-Palacin P, Aguado JM, Martinez-Martinez L, Torre-Cisneros J, Nucci M, Spanish Fusariosis Study G. 2021. Invasive fusariosis in nonneutropenic patients Spain, 2000–2015. *Emerg Infect Dis* 27:26–35.
102. Garnica M, da Cunha MO, Portugal R, Maiolino A, Colombo AL, Nucci M. 2015. Risk factors for invasive fusariosis in patients with acute myeloid leukemia and in hematopoietic cell transplant recipients. *Clin Infect Dis* 60:875–880. <https://doi.org/10.1093/cid/ciu947>
103. Riches ML, Trifilio S, Chen M, Ahn KW, Langston A, Lazarus HM, Marks DI, Martino R, Maziarz RT, Papanicolaou GA, Wingard JR, Young J-AH, Bennett CL. 2016. Risk factors and impact of non-aspergillus mold infections following allogeneic HCT: a CIBMTR infection and immune reconstitution analysis. *Bone Marrow Transplant* 51:277–282. <https://doi.org/10.1038/bmt.2015.263>
104. Varughese T, Taur Y, Cohen N, Palomba ML, Seo SK, Hohl TM, Redelman-Sidi G. 2018. Serious infections in patients receiving Ibrutinib for treatment of lymphoid cancer. *Clin Infect Dis* 67:687–692. <https://doi.org/10.1093/cid/ciy175>
105. Chan TSY, Au-Yeung R, Chim C-S, Wong SCY, Kwong Y-L. 2017. Disseminated *Fusarium* infection after Ibrutinib therapy in chronic lymphocytic leukaemia. *Ann Hematol* 96:871–872. <https://doi.org/10.1007/s00277-017-2944-7>
106. Anastasopoulou A, DiPippo AJ, Kontoyiannis DP. 2020. Non-aspergillus invasive mould infections in patients treated with Ibrutinib. *Mycoses* 63:787–793. <https://doi.org/10.1111/myc.13120>
107. Sterner RC, Sterner RM. 2021. CAR-T cell therapy: current limitations and potential strategies. *Blood Cancer J* 11:69. <https://doi.org/10.1038/s41408-021-00459-7>
108. Gudiol C, Lewis RE, Strati P, Kontoyiannis D. 2021. Chimeric antigen receptor T-cell therapy for the treatment of lymphoid malignancies: is there an excess risk for infection?. *Lancet Haematol* 8:e216–e228. [https://doi.org/10.1016/S2352-3026\(20\)30376-8](https://doi.org/10.1016/S2352-3026(20)30376-8)
109. Bupha-Intr O, Haeusler G, Chee L, Thursky K, Slavin M, Teh B. 2021. CAR-T cell therapy and infection: a review. *Expert Rev Anti Infect Ther* 19:749–758. <https://doi.org/10.1080/14787210.2021.1855143>
110. Haidar G, Dorritie K, Farah R, Bogdanovich T, Nguyen MH, Samanta P. 2020. Invasive mold infections after chimeric antigen receptor-modified T-cell therapy: a case series review of the literature, and implications for prophylaxis. *Clin Infect Dis* 71:672–676. <https://doi.org/10.1093/cid/ciz1127>
111. Park BJ, Pappas PG, Wannemuehler KA, Alexander BD, Anaissie EJ, Andes DR, Baddley JW, Brown JM, Brumble LM, Freifeld AG, Hadley S, Herwaldt L, Ito J, Kauffman CA, Lyon GM, Marr KA, Morrison VA, Papanicolaou G, Patterson TF, Perl TM, Schuster MG, Walker R, Wingard JR, Walsh TJ, Kontoyiannis DP. 2011. Invasive non-aspergillus mold infections in transplant recipients, United States, 2001–2006. *Emerg Infect Dis* 17:1855–1864. <https://doi.org/10.3201/eid1710.110087>
112. Sganga G, Bianco G, Fiori B, Nure E, Spanu T, Liroi MC, Frongillo F, Agnes S. 2013. Surveillance of bacterial and fungal infections in the postoperative period following liver transplantation: a series from 2005–2011. *Transplant Proc* 45:2718–2721. <https://doi.org/10.1016/j.transproceed.2013.08.010>
113. Halpern M, Balbi E, Carius L, Roma J, Gonzalez AC, Agoglia L, Covelo M, Araujo A, Guedes C, Alves J, Enne M, Martinho JM, Pacheco L. 2010. Cellulitis and nodular skin lesions due to *Fusarium* spp in liver transplant: case report. *Transplant Proc* 42:599–600. <https://doi.org/10.1016/j.transproceed.2010.01.004>
114. Lodato F, Tamé MR, Montagnani M, Sambri V, Liguori G, Azzaroli F, Costigliola P, Grazi G, Roda E, Mazzella G. 2006. Systemic fungemia and hepatic localizations of *fusarium* solani in a liver transplanted patient: an emerging fungal agent. *Liver Transpl* 12:1711–1714. <https://doi.org/10.1002/lt.20899>
115. Tascini C, Urbani L, Doria R, Catalano G, Leonildi A, Filipponi F, Menichetti F. 2009. Breakthrough *Fusarium* spp fungemia during caspofungin therapy in an ABO-incompatible orthotopic liver transplant patient. *J Chemother* 21:236–238. <https://doi.org/10.1179/joc.2009.21.2.236>
116. Cocchi S, Codeluppi M, Venturelli C, Bedini A, Grottola A, Gennari W, Cavrini F, Di Benedetto F, De Ruvo N, Rumpianesi F, Gerunda GE, Guaraldi G. 2011. *Fusarium* verticillioides fungemia in a liver transplantation patient: successful treatment with voriconazole. *Diagn Microbiol Infect Dis* 71:438–441. <https://doi.org/10.1016/j.diagmicrobio.2011.08.024>
117. Stropnický P, Heß K, Becker T, Braun F. 2022. Disseminated cerebral fusariosis in a liver-transplant patient: a case report and review of the literature. *Z Gastroenterol* 60:1231–1234. <https://doi.org/10.1055/a-1535-2981>
118. Cocuroccia B, Gaido J, Gubinelli E, Annessi G, Girolomoni G. 2003. Localized cutaneous hyalohyphomycosis caused by a *Fusarium* species infection in a renal transplant patient. *J Clin Microbiol* 41:905–907. <https://doi.org/10.1128/JCM.41.2.905-907.2003>
119. Girardi M, Glusac EJ, Imaeda S. 1999. Subcutaneous *Fusarium* foot abscess in a renal transplant patient. *Cutis* 63:267–270.
120. Young CN, Meyers AM. 1979. Opportunistic fungal infection by *Fusarium oxysporum* in a renal transplant patient. *Sabouraudia* 17:219–223.
121. Keskar VS, Wanjare S, Jamale TE, Mahajan D, Jawale SY, Fernandes G, Suryawanshi R, Hase NK. 2014. Subcutaneous hyalohyphomycosis caused by *Fusarium* in a kidney transplant recipient. *Ren Fail* 36:1129–1132. <https://doi.org/10.3109/0886022X.2014.926756>

122. Mohanty NK, Sahu S. 2014. *Fusarium solani* infection in a kidney transplant recipient. Indian J Nephrol 24:312–314. <https://doi.org/10.4103/0971-4065.133014>
123. Rekha A, Kindo AJ, Ravi A. 2008. *Fusarium solani* in the post-transplant patient: an unusual fungus. Int J Low Extrem Wounds 7:38–40. <https://doi.org/10.1177/1534734607313879>
124. Guarro J, Nucci M, Akiti T, Gené J, Barreiro MD, Gonçalves RT. 2000. Fungemia due to *Fusarium sacchari* in an immunosuppressed patient. J Clin Microbiol 38:419–421. <https://doi.org/10.1128/JCM.38.1.419-421.2000>
125. Arney KL, Tiernan R, Judson M. 1997. Primary pulmonary involvement of *Fusarium solani* in a lung transplant recipient. Chest 112:1128–1130. <https://doi.org/10.1378/chest.112.4.1128>
126. Carneiro HA, Coleman JJ, Restrepo A, Mylonakis E. 2011. *Fusarium* infection in lung transplant patients: report of 6 cases and review of the literature. Medicine (Baltimore) 90:69–80. <https://doi.org/10.1097/MD.0b013e318207612d>
127. Herbrecht R, Kessler R, Kravanja C, Meyer MH, Waller J, Letscher-Bru V. 2004. Successful treatment of *Fusarium proliferatum* pneumonia with posaconazole in a lung transplant recipient. J Heart Lung Transplant 23:1451–1454. <https://doi.org/10.1016/j.healun.2003.09.033>
128. Terasaki JM, Shah SK, Schnadig VJ, Valentine VG. 2014. Airway complication contributing to disseminated fusariosis after lung transplantation. Transpl Infect Dis 16:621–624. <https://doi.org/10.1111/tid.12240>
129. Sampathkumar P, Paya CV. 2001. *Fusarium* infection after solid-organ transplantation. Clin Infect Dis 32:1237–1240. <https://doi.org/10.1086/319753>
130. Benish M, Elitzur S, Arad-Cohen N, Barg AA, Ben-Harosh M, Bielorai B, Fischer S, Gilad G, Levy I, Rosenfeld-Keidar H, Shachor-Meyouhas Y, Soen-Grisaru G, Weinreb S, Nirel R, Elhasid R. 2022. Invasive fusariosis in pediatric hematology/oncology and stem cell transplant patients: a report from the Israeli society of pediatric hematology-oncology. J Fungi (Basel) 8:387. <https://doi.org/10.3390/jof8040387>
131. Mansoor D, Roobzahany NA, Mazinany H, Samimagam A. 2003. Chronic *Fusarium* infection in an adult patient with undiagnosed chronic granulomatous disease. Clin Infect Dis 37:e107–8. <https://doi.org/10.1086/377608>
132. Medaglia AA, Marco-Hernández J, de Ossó Acuña JT, Hermida Lama E, Martínez-Rebollar M, Caballero M, Rodríguez-Carunchio L, García F. 2018. *Fusarium* keratoplastom infection in an HIV-infected patient. Int J STD AIDS 29:1039–1042. <https://doi.org/10.1177/0956462418761259>
133. Esnakula AK, Summers I, Naab TJ. 2013. Fatal disseminated *Fusarium* infection in a human immunodeficiency virus positive patient. Case Rep Infect Dis 2013:379320. <https://doi.org/10.1155/2013/379320>
134. Albisetti M, Lauener RP, Güngör T, Schär G, Niggli FK, Nadal D. 2004. Disseminated *Fusarium oxysporum* infection in hemophagocytic lymphohistiocytosis. Infection 32:364–366. <https://doi.org/10.1007/s15010-004-3135-8>
135. Mellouli F, Ksouri H, Barbouche R, Maamer M, Hamed LB, Hmida S, Hassen AB, Béjaoui M. 2010. Successful treatment of *Fusarium solani* ecthyma gangrenosum in a patient affected by leukocyte adhesion deficiency type 1 with granulocytes transfusions. BMC Dermatol 10:10. <https://doi.org/10.1186/1471-5945-10-10>
136. Abbara S, Freeman AF, Cohen JF, Leclerc-Mercier S, Sanchez L, Schlatter J, Cisternino S, Parker R, Cowen EW, Rouzaud C, Bougnoux ME, Lantier F, Lionakis MS, Lortholary O. 2023. Primary invasive cutaneous fusariosis in patients with STAT3 hyper-IgE syndrome. J Clin Immunol 43:647–652. <https://doi.org/10.1007/s10875-022-01404-4>
137. Poignon C, Blaize M, Vezinet C, Lampros A, Monsel A, Fekkar A. 2020. Invasive pulmonary fusariosis in an immunocompetent critically ill patient with severe COVID-19. Clin Microbiol Infect 26:1582–1584. <https://doi.org/10.1016/j.cmi.2020.06.026>
138. Gangneux JP, Dannaoui E, Fekkar A, Luyt CE, Botterel F, de Prost N, Tadie JM, Reizine F, Houze S, Timsit JF, Iriart X, Riu-Poulenc B, Sendid B, Nseir S, Persat F, Wallet F, Le Pape P, Canet E, Novara A, Manai M, Cateau E, Thille AW, Brun S, Cohen Y, Alanio A, Megarbane B, Cornet M, Terzi N, Lamhaut L, Sabourin E, Desoubreux G, Ehrmann S, Hennequin C, Voirit G, Nevez G, Aubron C, Letscher-Bru V, Meziani F, Blaize M, Mayaux J, Monsel A, Boquel F, Robert-Gangneux F, Le Tulzo Y, Seguin P, Guegan H, Autier B, Lesouhaitier M, Pelletier R, Belaz S, Bonnal C, Berry A, Leroy J, François N, Richard J-C, Paulus S, Argaud L, Dupont D, Menotti J, Morio F, Soulié M, Schwebel C, Garnaud C, Guitard J, Le Gal S, Quinio D, Morcet J, Laviolle B, Zahar JR, Bougnoux ME. 2022. Fungal infections in mechanically ventilated patients with COVID-19 during the first wave: the French multicentre MYCOVID study. Lancet Respir Med 10:180–190. <https://doi.org/10.2139/ssrn.3858565>
139. Moretti ML, Busso-Lopes AF, Tararam CA, Moraes R, Muraosa Y, Mikami Y, Gonoï T, Taguchi H, Lyra L, Reichert-Lima F, Trabasso P, de Hoog GS, Al-Hatmi AMS, Schreiber AZ, Kamei K. 2018. Airborne transmission of invasive fusariosis in patients with hematologic malignancies. PLoS One 13:e0196426. <https://doi.org/10.1371/journal.pone.0196426>
140. Edel-Hermann V, Sautour M, Gautheron N, Laurent J, Aho S, Bonnin A, Sixt N, Hartemann P, Dalle F, Steinberg C. 2016. A clonal lineage of *Fusarium oxysporum* circulates in the tap water of different French hospitals. Appl Environ Microbiol 82:6483–6489. <https://doi.org/10.1128/AEM.01939-16>
141. Balmas V, Fancellu F, Sanna S, Scherm B, Migheli Q, Malbrán I. 2021. Water distribution systems in sardinian hospitals host invasive clonal lineages of the *Fusarium oxysporum* and *Fusarium solani* species complexes. Mycologia 113:725–733. <https://doi.org/10.1080/00275514.2021.1905497>
142. Steinberg C, Laurent J, Edel-Hermann V, Barbezant M, Sixt N, Dalle F, Aho S, Bonnin A, Hartemann P, Sautour M. 2015. Adaptation of *Fusarium oxysporum* and *Fusarium dimerum* to the specific aquatic environment provided by the water systems of hospitals. Water Res 76:53–65. <https://doi.org/10.1016/j.watres.2015.02.036>
143. Litvinov N, da Silva MTN, van der Heijden IM, Graça MG, Marques de Oliveira L, Fu L, Giudice M, Zilda de Aquino M, Odone-Filho V, Marques HH, Costa SF, Levin AS. 2015. An outbreak of invasive fusariosis in a children's cancer hospital. Clin Microbiol Infect 21:268. <https://doi.org/10.1016/j.cmi.2014.09.004>
144. Georgiadou SP, Velegraki A, Arabatzis M, Neonakis I, Chatzipanagiotou S, Dalekos GN, Petinaki E. 2014. Cluster of *Fusarium verticillioides* bloodstream infections among immunocompetent patients in an internal medicine department after reconstruction works in Larissa, central Greece. J Hosp Infect 86:267–271. <https://doi.org/10.1016/j.jhin.2014.01.011>
145. Carlesse F, Amaral AP, Gonçalves SS, Xafranski H, Lee ML, Zecchin V, Petrilli AS, Al-Hatmi AM, Hagen F, Meis JF, Colombo AL. 2017. Outbreak of *Fusarium oxysporum* infections in children with cancer: an experience with 7 episodes of catheter-related fungemia. Antimicrob Resist Infect Control 6:93. <https://doi.org/10.1186/s13756-017-0247-3>
146. Nucci M, Anaissie E. 2002. Cutaneous infection by *Fusarium* species in healthy and immunocompromised hosts: implications for diagnosis and management. Clin Infect Dis 35:909–920. <https://doi.org/10.1086/342328>
147. Scheel CM, Hurst SF, Barreiros G, Akiti T, Nucci M, Balajee SA. 2013. Molecular analyses of *Fusarium* isolates recovered from a cluster of invasive mold infections in a Brazilian hospital. BMC Infect Dis 13:49. <https://doi.org/10.1186/1471-2334-13-49>
148. Varon AG, Nouer SA, Barreiros G, Trope BM, Magalhães F, Akiti T, Garnica M, Nucci M. 2014. Superficial skin lesions positive for *Fusarium* are associated with subsequent development of invasive fusariosis. J Infect 68:85–89. <https://doi.org/10.1016/j.jinf.2013.08.011>
149. Barton E, Borman A, Johnson E, Sherlock J, Giles A. 2016. Pseudo-outbreak of *Fusarium oxysporum* associated with bronchoscopy. J Hosp Infect 94:197–198. <https://doi.org/10.1016/j.jhin.2016.06.016>
150. Schaffer K, Fitzgerald SF, Commene M, Maguiness A, Fenelon LE. 2008. A pseudo-outbreak of *Fusarium solani* in an intensive care unit associated with bronchoscopy. J Hosp Infect 69:400–402. <https://doi.org/10.1016/j.jhin.2008.03.008>
151. Levy L, Block C, Schwartz C, Gross I, Cohen M, Fridlender ZG, Moses AE, Berkman N, Benenson S. 2016. Cluster of *Fusarium solani* Isolations in a bronchoscopy unit. Clin Microbiol Infect 22:e5–e6. <https://doi.org/10.1016/j.cmi.2015.09.017>
152. Grigis A, Farina C, Symoens F, Nolard N, Goglio A. 2000. Nosocomial pseudo-outbreak of *Fusarium verticillioides* associated with sterile plastic containers. Infect Control Hosp Epidemiol 21:50–52. <https://doi.org/10.1086/501699>
153. Nucci F, Nouér SA, Capone D, Anaissie E, Nucci M. 2015. Fusariosis. Semin Respir Crit Care Med 36:706–714. <https://doi.org/10.1055/s-0035-1562897>
154. Nucci F, Nouér SA, Capone D, Nucci M. 2018. Invasive mould disease in haematologic patients: comparison between fusariosis and aspergilliosis. Clin Microbiol Infect 24:1105. <https://doi.org/10.1016/j.cmi.2018.05.006>

155. Marom EM, Holmes AM, Bruzzi JF, Truong MT, O'Sullivan PJ, Kontoyianis DP. 2008. Imaging of pulmonary fusariosis in patients with hematologic malignancies. *AJR Am J Roentgenol* 190:1605–1609. <https://doi.org/10.2214/AJR.07.3278>
156. Sassi C, Stanzani M, Lewis RE, Vianelli N, Tarsi A, Poerio A, Cavo M, Battista G. 2017. Radiologic findings of *Fusarium* pneumonia in neutropenic patients. *Mycoses* 60:73–78. <https://doi.org/10.1111/myc.12538>
157. Nucci M, Anaissie E. 2006. Emerging fungi. *Infect Dis Clin North Am* 20:563–579.
158. Rizzello I, Castagnetti F, Toschi PG, Bertaccini P, Primavera L, Paolucci M, Faccioli L, Spinardi L, Lewis RE, Cavo M, Stanzani M. 2018. Successful treatment of bilateral endogenous *Fusarium solani* endophthalmitis in a patient with acute lymphocytic leukaemia. *Mycoses* 61:53–60. <https://doi.org/10.1111/myc.12697>
159. Hayden RT, Isotalo PA, Parrett T, Wolk DM, Qian X, Roberts GD, Lloyd RV. 2003. In situ hybridization for the differentiation of *Aspergillus*, *Fusarium*, and *Pseudallescheria* species in tissue section. *Diagn Mol Pathol* 12:21–26. <https://doi.org/10.1097/00019606-200303000-00003>
160. Salehi E, Hedayati MT, Zoll J, Rafati H, Ghasemi M, Doroudinia A, Abastabar M, Tolooe A, Snelders E, van der Lee HA, Rijs A, Verweij PE, Seyedmousavi S, Melchers WJG, Warnock DW. 2016. Discrimination of aspergillosis, mucormycosis, fusariosis, and scedosporiosis in formalin-fixed paraffin-embedded tissue specimens by use of multiple real-time quantitative PCR assays. *J Clin Microbiol* 54:2798–2803. <https://doi.org/10.1128/JCM.01185-16>
161. Hennequin C, Ranaivoarimalala C, Chouaki T, Tazerout M, Ancelle T, Cabaud JJ, Raccour CP. 2002. Comparison of aerobic standard medium with specific fungal medium for detecting *Fusarium* spp in blood cultures. *Eur J Clin Microbiol Infect Dis* 21:748–750. <https://doi.org/10.1007/s10096-002-0812-3>
162. Oz Y, Onder S, Alpaslan E, Durmaz G. 2020. Does concomitant bacteraemia hide the fungi in blood cultures? an in vitro study. *J Med Microbiol* 69:944–948. <https://doi.org/10.1099/jmm.0.001210>
163. Tortorano AM, Esposito MC, Prigitano A, Grancini A, Ossi C, Cavanna C, Cascio GL. 2012. Cross-reactivity of *Fusarium* spp. in the *Aspergillus* galactomannan enzyme-linked immunosorbent assay. *J Clin Microbiol* 50:1051–1053. <https://doi.org/10.1128/JCM.05946-11>
164. Nucci M, Carlesse F, Cappellano P, Varon AG, Seber A, Garnica M, Nouér SA, Colombo AL. 2014. Earlier diagnosis of invasive fusariosis with *Aspergillus* serum galactomannan testing. *PLoS One* 9:e87784. <https://doi.org/10.1371/journal.pone.0087784>
165. Horn DL, Freifeld AG, Schuster MG, Azie NE, Franks B, Kauffman CA. 2014. Treatment and outcomes of invasive fusariosis: review of 65 cases from the PATH Alliance(R) registry. *Mycoses* 57:652–658. <https://doi.org/10.1111/myc.12212>
166. Mikulska M, Balletto E, Castagnola E, Mularoni A. 2021. Beta-D-glucan in patients with haematological malignancies. *J Fungi (Basel)* 7:1046. <https://doi.org/10.3390/jof7121046>
167. Nucci M, Barreiros G, Reis H, Paixão M, Akiti T, Nouér SA. 2019. Performance of 1,3-beta-D-glucan in the diagnosis and monitoring of invasive fusariosis. *Mycoses* 62:570–575. <https://doi.org/10.1111/myc.12918>
168. Dellièrè S, Guitard J, Sabou M, Angebault C, Moniot M, Cornu M, Hamane S, Bougnoux M-E, Imbert S, Pasquier G, Botterel F, Garcia-Hermoso D, Alanio A. 2022. Detection of circulating DNA for the diagnosis of invasive fusariosis: retrospective analysis of 15 proven cases. *Med Mycol* 60:myac049. <https://doi.org/10.1093/mmy/myac049>
169. Espinel-Ingroff A, Colombo AL, Cordoba S, Dufresne PJ, Fuller J, Ghannoum M, Gonzalez GM, Guarro J, Kidd SE, Meis JF, Melhem T, Pelaez T, Pfaller MA, Szesz MW, Takahaschi JP, Tortorano AM, Wiederhold NP, Turnidge J. 2016. International evaluation of MIC distributions and epidemiological cutoff value (ECV) definitions for *fusarium* species identified by molecular methods for the CLSI broth microdilution method. *Antimicrob Agents Chemother* 60:1079–1084. <https://doi.org/10.1128/AAC.02456-15>
170. Pfaller MA, Carvalhaes CG, Rhomberg P, Messer SA, Castanheira M. 2021. Antifungal susceptibilities of opportunistic filamentous fungal pathogens from the Asia and western Pacific region: data from the SENTRY antifungal surveillance program. *J Antibiot (Tokyo)* 74:519–527. <https://doi.org/10.1038/s41429-021-00431-4>
171. Jørgensen KM, Astvad KMT, Hare RK, Arendrup MC. 2019. EUCAST susceptibility testing of isavuconazole: MIC data for contemporary clinical mold and yeast isolates. *Antimicrob Agents Chemother* 63:e00073-19. <https://doi.org/10.1128/AAC.00073-19>
172. Messer SA, Carvalhaes CG, Castanheira M, Pfaller MA. 2020. In vitro activity of isavuconazole versus opportunistic filamentous fungal pathogens from the SENTRY antifungal surveillance program, 2017–2018. *Diagn Microbiol Infect Dis* 97:115007. <https://doi.org/10.1016/j.diagmicrobio.2020.115007>
173. Badali H, Cañete-Gibas C, Patterson H, Sanders C, Mermella B, Garcia V, Mele J, Fan H, Wiederhold NP. 2021. In vitro activity of olorofim against clinical isolates of the *Fusarium oxysporum* and *Fusarium solani* species complexes. *Mycoses* 64:748–752. <https://doi.org/10.1111/myc.13273>
174. Badali H, Patterson HP, Sanders CJ, Mermella B, Gibas CFC, Ibrahim AS, Shaw KJ, Wiederhold NP. 2021. Manogepix, the active moiety of the investigational agent fosmanogepix, demonstrates *in vitro* activity against members of the *Fusarium oxysporum* and *Fusarium solani* species complexes. *Antimicrob Agents Chemother* 65:e02343-20. <https://doi.org/10.1128/AAC.02343-20>
175. Broutin A, Bigot J, Senghor Y, Moreno-Sabater A, Guitard J, Hennequin C. 2020. In vitro susceptibility of *Fusarium* to isavuconazole. *Antimicrob Agents Chemother* 64:e01621-19. <https://doi.org/10.1128/AAC.01621-19>
176. Lortholary O, Obenga G, Biswas P, Caillot D, Chachaty E, Bienvenu A-L, Cornet M, Greene J, Herbrecht R, Lacroix C, Grenouillet F, Raad I, Sitbon K, Troke P, French Mycoses Study Group. 2010. International retrospective analysis of 73 cases of invasive fusariosis treated with voriconazole. *Antimicrob Agents Chemother* 54:4446–4450. <https://doi.org/10.1128/AAC.00286-10>
177. Nucci M, Jenks J, Thompson GR, Hoenigl M, Dos Santos MC, Forghieri F, Rico JC, Bonuomo V, López-Soria L, Lass-Flörl C, Candoni A, Garcia-Vidal C, Cattaneo C, Buil J, Rabagliati R, Roiz MP, Gudiol C, Fracchiolla N, Campos-Herrero MI, Delia M, Farina F, Fortun J, Nadali G, Sastre E, Colombo AL, Pérez Nadales E, Alastruey-Izquierdo A, Pagano L. 2021. Do high mics predict the outcome in invasive fusariosis? *J Antimicrob Chemother* 76:1063–1069. <https://doi.org/10.1093/jac/dkaa516>
178. Hoenigl M, Salmanton-García J, Walsh TJ, Nucci M, Neoh CF, Jenks JD, Lackner M, Sprute R, Al-Hatmi AMS, Bassetti M, Carlesse F, Freiburger T, Koehler P, Lehrnbecher T, Kumar A, Prattes J, Richardson M, Revankar S, Slavin MA, Stemler J, Spiess B, Taj-Aldeen SJ, Warris A, Woo PCY, Young J-AH, Albus K, Arenz D, Arsic-Arsenijevic V, Bouchara J-P, Chinniah TR, Chowdhary A, de Hoog GS, Dimopoulos G, Duarte RF, Hamal P, Meis JF, Mfinanga S, Queiroz-Telles F, Patterson TF, Rahav G, Rogers TR, Rotstein C, Wahyuningsih R, Seidel D, Cornely OA. 2021. Global guideline for the diagnosis and management of rare mould infections: an initiative of the European confederation of medical mycology in cooperation with the international society for human and animal mycology and the American society for microbiology. *Lancet Infect Dis* 21:e246–e257. [https://doi.org/10.1016/S1473-3099\(20\)30784-2](https://doi.org/10.1016/S1473-3099(20)30784-2)
179. Maertens JA, Girmenia C, Brüggemann RJ, Duarte RF, Kibbler CC, Ljungman P, Racil Z, Ribaud P, Slavin MA, Cornely OA, Peter Donnelly J, Cordonnier C, European Conference on Infections in Leukaemia (ECIL), a joint venture of the European Group for Blood and Marrow Transplantation (EBMT), the European Organization for Research and Treatment of Cancer (EORTC), the Immunocompromised Host Society (ICHS) and. 2018. European guidelines for primary antifungal prophylaxis in adult haematology patients: summary of the updated recommendations from the European conference on infections in leukaemia. *J Antimicrob Chemother* 73:3221–3230. <https://doi.org/10.1093/jac/dky286>
180. Varon AG, Nouér SA, Barreiros G, Trope BM, Akiti T, Nucci M. 2016. Antimold prophylaxis may reduce the risk of invasive fusariosis in hematologic patients with superficial skin lesions with positive culture for *Fusarium*. *Antimicrob Agents Chemother* 60:7290–7294. <https://doi.org/10.1128/AAC.00636-16>
181. Nucci M, Shoham S, Abdala E, Hamerschlag N, Rico JC, Forghieri F, Nouér SA, Cappellano P, Solza C, Gonzaga Y, Nadali G, Nucci F, Colombo AL, Albuquerque AM, Queiroz-Telles Filho F, Lima CBL, Arrais-Rodrigues C, Rocha V, Marty FM. 2019. Outcomes of patients with invasive fusariosis who undergo further immunosuppressive treatments, is there a role for secondary prophylaxis? *Mycoses* 62:413–417. <https://doi.org/10.1111/myc.12901>
182. Seban RD, Bonardel G, Guernou M, Lussato D, Queneau M. 2017. The use of FDG PET-CT imaging for the assessment of early antifungal treatment response in disseminated fusariosis. *Clin Nucl Med* 42:569–570. <https://doi.org/10.1097/RLU.0000000000001682>

183. Schirmer MR, Carneiro MP, Machado LS, Chaves A da S, Lopes F. 2018. Fluorine-18-fluorodeoxyglucose PET/CT in hematopoietic stem cell transplant patients with fusariosis: initial findings of a case series review. *Nucl Med Commun* 39:545–552. <https://doi.org/10.1097/MNM.0000000000000834>
184. Longhitano A, Alipour R, Khot A, Bajel A, Antippa P, Slavin M, Thursky K. 2021. The role of 18F-fluorodeoxyglucose positron emission tomography/computed tomography (FDG PET/CT) in assessment of complex invasive fungal disease and opportunistic co-infections in patients with acute leukemia prior to allogeneic hematopoietic cell transplant. *Transpl Infect Dis* 23:e13547. <https://doi.org/10.1111/tid.13547>
185. Cesaro S, Marinello S, Alessia B, Alaggio R, Rossi L, Toffolutti T, Putti MC, Gamba P. 2010. Successful treatment of disseminated fusariosis in a child with acute myelogenous leukaemia with medical and surgical approach. *Mycoses* 53:181–185. <https://doi.org/10.1111/j.1439-0507.2008.01674.x>
186. Velasco E, Martins CA, Nucci M. 1995. Successful treatment of catheter-related fusarial infection in immunocompromised children. *Eur J Clin Microbiol Infect Dis* 14:697–699. <https://doi.org/10.1007/BF01690877>
187. Kadri SS, Remy KE, Strich JR, Gea-Banacloche J, Leitman SF. 2015. Role of granulocyte transfusions in invasive fusariosis: systematic review and single-center experience. *Transfusion* 55:2076–2085. <https://doi.org/10.1111/trf.13099>
188. Khatamzas E, Mellinghoff SC, Thelen M, Schlößer HA, Kunz WG, Buerkle C, Dichtl K, Ormanns S, von Bergwelt-Baildon M. 2022. Nivolumab induces long-term remission in a patient with fusariosis. *Eur J Cancer* 173:91–94. <https://doi.org/10.1016/j.jejca.2022.06.035>

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