

Review

Human Intestinal Organoids and Microphysiological Systems for Modeling Radiotoxicity and Assessing Radioprotective Agents

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Simple Summary: Because of the limitations of current *in vivo* and *in vitro* models, this review considers a human relevant approach: modeling colorectal radiotoxicity through human-derived organoids and microfluidics. This system can offer a closer representation of the microenvironment. Co-culturing bacteria and patient derived tumor organoids under a radiotherapy setup, can enable an understanding of the interplay between radiotherapy, the gut microbiota, patient outcomes and assess radioprotective agents of interest. However, challenges in model development highlight the necessity for refinement, questioning their potential to bridge the gap between preclinical research and clinical applications in CRC treatment.



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Abstract: Radiotherapy is a commonly employed treatment for colorectal cancer, yet its radiotoxicity-related impact on healthy tissues raises significant health concerns. This highlights the need to use radioprotective agents to mitigate these side effects. This review presents the current landscape of human translational radiobiology, outlining the limitations of existing models and proposing engineering solutions. We delve into radiotherapy principles, encompassing mechanisms of radiation-induced cell death and its influence on normal and cancerous colorectal cells. Furthermore, we explore the engineering aspects of microphysiological systems to represent radiotherapy-induced gastrointestinal toxicity and how to include the gut microbiota to study its role in treatment failure and success. This review ultimately highlights the main challenges and future pathways in translational research for pelvic radiotherapy-induced toxicity. This is achieved by developing a humanized *in vitro* model that mimics radiotherapy treatment conditions. An *in vitro* model should provide in-depth analyses of host-gut microbiota interactions and a deeper understanding of the underlying biological mechanisms of radioprotective food supplements. Additionally, it would be of great value if these models could produce high-throughput data using patient-derived samples to address the lack of human representability to complete clinical trials and improve patients' quality of life.

Keywords: ionizing radiation; colorectal cancer; radioprotection; radiotoxicity; organoid; microbiota; organ-on-a-chip

1. Introduction

Colorectal cancer (CRC) is the third most prevalent cancer worldwide [1–3] with nearly 1 million deaths per year, according to the WHO [2]. As a part of treatment optimization,

radiotherapy is one of the most used treatments to manage CRC [4]. However, radiotherapy treatment is limited by its toxicity to healthy tissues and potential to induce resistance mechanisms in tumor cells. Therefore, there is growing interest in identifying and applying radioprotective agents that can enhance the effectiveness of radiotherapy while minimizing the toxicity to healthy tissues. The identification of such radioprotective agents is not straightforward since current research does not give a full understanding of their potential. Emerging studies have focused on using microphysiological systems with a radiotherapy setup that models the human cancer microenvironment [5] and that could be used to assess radioprotective candidates. This review explores the use of microphysiological systems in the context of radiotherapy treatment for colorectal cancer and their potential for assessing radioprotective candidates.

2. The Use of Radiotherapy and Its Targets

Since their discovery by Röntgen in 1895 and their first clinical use in 1896 [6], X-rays have been widely used in radiotherapy as an effective cancer treatment [7]. Through ionizing molecules, ionizing radiation transfers energy to targeted cells to generate lethal lesions [8]. Treatment relies on the therapeutic ratio that depends on the radio-responsiveness of the tumors [9,10]. This necessitates finding a balance between maximizing the radiation dose to kill cancer cells and minimizing the radiation dose to healthy cells. Various factors, such as linear energy transfer, total dose, fractionation rate, and radiosensitivity, must be considered to modulate the effectiveness of tumor cell killing in the selected treatment strategy [9]. The “5Rs” of radiotherapy, used to describe how the patient’s tumor responds to radiation [11,12], are also essential considerations. They are defined as (1) repair: the ability to repair DNA damage caused by radiation, (2) repopulation: the extent of cell proliferation varying between fractions, (3) redistribution: radiation mostly affects cells in mitosis, while S-phase cells are mostly resistant, (4) reoxygenation: radiation sensitivity increases in oxygen-rich environments, and (5) radiosensitivity: individual radiosensitivity and intrinsic tumor radiosensitivity determine the response to radiation.

Ionizing radiation ruptures chemical bonds when interacting with biological materials. In cells, the main components including proteins, lipids, and nucleic acids can be damaged [13]. DNA is a biological target for radiation because of energy transfer, with single-strand and double-strand breaks affecting tumor cell viability [14]. When double-strand breaks cannot be repaired due to reduced repair capacity, it leads to cell death [15]. Single-strand breaks can be repaired by cells, given the available antisense template. Cancer cells generally produce more double stranded DNA breaks and are less effective in repairing the damage caused by radiotherapy [15]. However, treatment resistance can occur. A consequence of DNA breaks can be an increase in the number of mutations within the DNA of cancer cells. This provokes more proliferation, (for example, alteration in NRAS, KRAS, APC [1]) resulting in tumor burden. Another adverse impact of radiotherapy is the creation of a microenvironment/hypoxia in favor of the tumor, allowing further growth and promoting tumor progression [1]. Concurrently, in the aftermath of irradiation, a phenomenon known as irradiation repopulation takes place, marked by the proliferation of resilient cancer cells. This process is closely intertwined with resistance to treatment, consequently promoting the advancement of cancer cells (Figure 1). This correlates with the importance of the 5Rs in setting up a treatment strategy that can affect cancer cells and avoid resistance. In addition, radiation stimulates the immune system via two main pro-inflammatory pathways: nuclear factor kappa-light-chain-enhancer of activated B cells (NF- κ B), and signal transducers and activators of transcription (STAT). NF- κ B regulates the expression of anti-apoptotic proteins, pro-inflammatory cytokines, and chemokines such as tumor necrosis factor alpha (TNF- α) and Interleukin-1 (IL1) in macrophages. STAT proteins also play a significant role in tumor development [16]. Among these proteins, STAT3 is associated with radioresistance. As the most important regulator of survivin (part of the apoptosis protein family), its inhibition showed to increase CRC apoptosis in vitro [2]. The STAT1 protein has dual functions, it can induce pro-apoptotic genes, such as *caspase 3*,

and promote radioresistant cancer cell phenotypes and tumor metastasis. Additionally, tumor cells and tumor-infiltrating lymphocytes produce cytokines and growth factors (e.g., TNF- α and IL6) in response to radiation and are mostly dose-dependent [17]. These pro-inflammatory responses after radiotherapy, involving the acquired immune system, could create a favorable environment for the tumor through the over-stimulation of inflammatory components, thereby enhancing cancer cell invasiveness [16–19], which causes unwanted side effects.

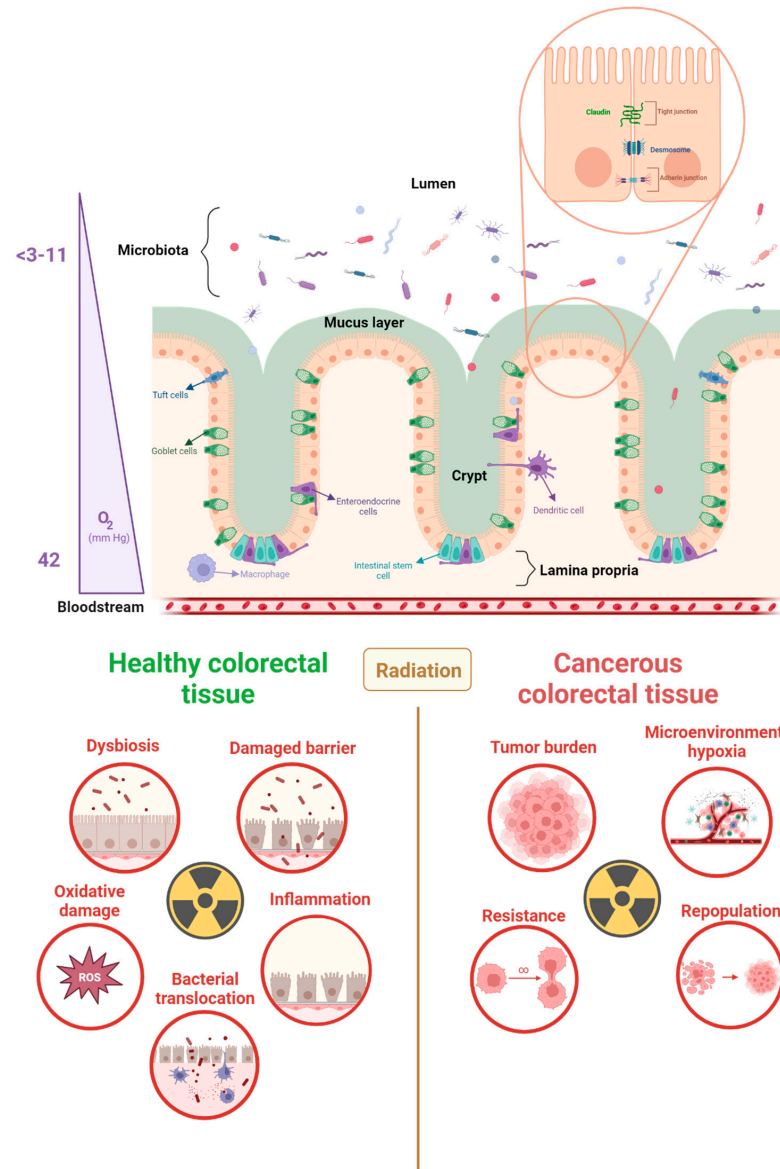


Figure 1. Large intestine morphology, radiotoxicity, and cancer radioresistance. The large intestine exhibits a unique physiology that facilitates its absorption and waste storage functions. The epithelial barrier, which separates the lumen and the lamina propria, is linked by junctions and is covered by a mucus layer. The oxygen gradient within the large intestine, ranging from 3 to 40 pO₂ (mmHg), creates an anaerobic environment in the lumen and an aerobic environment in the lamina propria. During radiotherapy, both healthy and cancerous colorectal tissue can be adversely affected by radiation. Radiotoxicity of healthy tissues is mediated by a series of interconnected events, while radioresistance in cancer tissues can result in an increase in tumor growth. This can be attributed to a facilitated microenvironment and tumor-enhancing mutations [20–23].

3. The Radiotoxicity of the Gastrointestinal Tract

Within the context of radiotherapy treatment, it is crucial to consider radiotoxicity, which encompasses the detrimental impact of radiation on healthy tissues. About 90% of pelvic cancer patients in Europe reported gastrointestinal problems during and/or after treatment, with 50% experiencing symptoms such as diarrhea, cramping, and nausea [19,24]. Because of these symptoms, patients need to break from therapy, reducing the effectiveness of the treatment.

3.1. The Physiology and Function of the Gastrointestinal Tract

The human digestive system is made of the mouth, pharynx, esophagus, stomach, and the small and large intestines. The gastrointestinal tract is a tubular structure composed of a lumen, mucosa, submucosa, muscularis, and serosa, with different microphysiological and cellular organization in the different parts of the digestive system [21,22]. The small intestine comprises three segments, namely, the duodenum, jejunum, and ileum, which are primarily involved in breaking down food, absorbing nutrients and water, and propelling food through the gastrointestinal tract. In contrast, the large intestine consists of the cecum, colon, rectum, and anal canal. It serves as an organ for absorption, the storage of waste, and the transportation of solid waste. Unlike the small intestine, which has villi and crypts, the epithelium of the large intestine comprises only crypts that allow for interactions with the gut microbiota, thereby creating a compartment for waste storage [21–23]. The bottom of the crypts is made of stem cells, that self-renew after 3 to 4 divisions before differentiating and moving up the crypt [25]. Owing to its unique microphysiology, the gastrointestinal tract undergoes renewal every five days [26]. The inner surface of the large intestine is composed of epithelium containing (1) epithelial cells that allow the absorption of nutrients, (2) chemosensory tuft cells that track the luminal gut content and contribute to the immune responses, and (3) goblet cells that secrete mucus to cover the surface of the epithelium (Figure 1). The mucus and intestinal epithelial cells serve as a barrier that prevents bacteria, located in the lumen, from infiltrating the body. Moreover, desmosomes, adherens junctions, and tight junctions allow physical protection and an interface for the immune system (e.g., resident macrophages, dendritic cells, neutrophils, lymphocytes, and monocytes) to induce adequate immunological responses [27] (Figure 1). Therefore, they play a crucial role in the barrier function and exhibit robust adhesion of the epithelial lining [28]. Indeed, the mucosal surface lining regulates its immune responses through mucosa-associated lymphoid tissue (MALT). More specifically in the gastrointestinal tract, gut-associated lymphoid tissues (GALT) induce adaptive immune responses (lymphocytes B and T) [29]. Finally, a specific aspect of the physiology of the gastrointestinal tract is a highly vascularized lamina propria of the intestinal mucosa. It contrasts with an anaerobic lumen inhabited by trillions of metabolically active microbes, creating a very specific tissue microenvironment. Among the microorganisms colonizing the gut, the anaerobic population mirrors the need for hypoxic conditions in the gut environment [30]. The combination of variations in blood flow, epithelial metabolism, and oxygen diffusion into the lumen leads to this particular physiology. Microbiota-produced metabolites, such as short-chain fatty acids (SCFAs), are used by colon epithelial cells and stimulate oxygen consumption via oxidative phosphorylation, sustaining and contributing to the hypoxic environment. By coordinating numerous elements of the intestinal epithelium's activity (Figure 2), SCFA producers appear to be crucial for preserving physiological hypoxia characterized by a PO_2 value below 10 mmHg (Figure 1) [30–32].

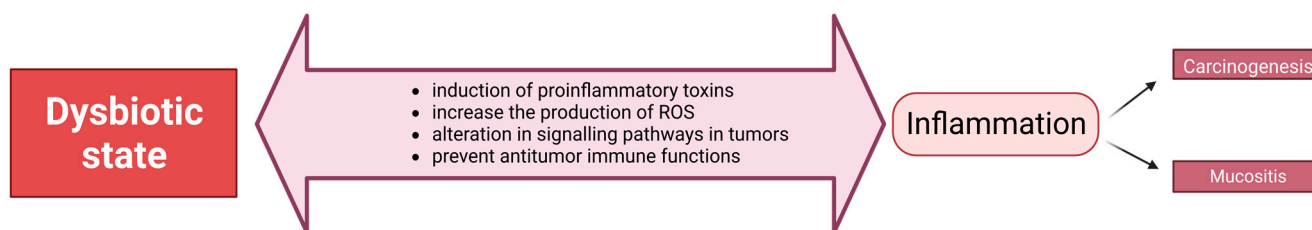


Figure 2. The effect of pathogenic bacteria in dysbiosis. An overgrowth of pathogenic bacteria in the lumen leads to dysbiosis. This has detrimental effects on health through various mechanisms, including the stimulation of an inflammatory state. The resulting mucositis and carcinogenesis can further propagate dysbiosis, leading to an increase in pathogenic bacterial presence in the gut [33]. ROS: reactive oxygen species.

3.2. Pelvic Radiation Disease

With the specific physiology we described, in the frame of cancer survival, the rise of research towards treatments and their side effects [34–39] has put forward the symptoms of pelvic radiation disease [40,41]. Due to the quick turnover of the gut epithelium, the stem cells present in the small intestine are highly radiosensitive [40,42–45], resulting in damage to the gut epithelium. There, DNA strand breaks provoke the activation of transcription factors, leading to the release of pro-inflammatory cytokines and chemokines [46]. The production of these mediators stimulates the recruitment of neutrophils, characterizing acute inflammation. This inflammatory process leads to the migration of monocytes, the activation of mast cells produces pro-inflammatory and pro-fibrotic mediators, such as the transforming growth factor beta 1 (TGF- β 1), cooperating with IL4 and IL13 cytokines [16,46]. In addition, as ionizing radiation affects the mucus layer and intestinal walls, bacterial translocation occurs and activates inflammatory responses as well. Bacterial toxins that pass through the intestinal wall and enter the bloodstream increase the risk of local infection and sepsis [47].

In parallel, the biological effects on neighboring cells that are not directly irradiated are known as bystander effects or non-targeted responses. They originate from lesions caused by the initial radiation and differ from the alterations that remain after repair. Bystander effects can be classified into two types. One type, called direct bystander effects, involves the release of molecular factors by cells through gap junction intercellular contact [48,49]. In contrast, another type, known as indirect bystander effects, occurs when cells that are further away encounter secreted factors through the bloodstream. The complex mechanisms underlying bystander effects involve various signaling pathways, including oxidative stress, DNA damage, inflammation, apoptosis, and pyroptosis. These pathways activate different molecular factors such as cytokines, growth factors, and reactive oxygen species (ROS). They initiate a series of events leading to biological alterations in neighboring cells. The innate immune system is triggered by damage-associated molecular patterns (DAMPs) and pathogen-associated molecular patterns (PAMPs). These signals primarily recruit resident macrophages and dendritic cells. The cytosolic multiprotein complex, known as the inflammasome, recognizes PAMPs and DAMPs through pathogen recognition receptors (PRRs) and facilitates the proteolytic cleavage of inflammatory proteins in response to endothelial cell proliferation [50]. Furthermore, lipopolysaccharides (LPSs), present on the outer membrane of all Gram-negative bacteria, induce inflammatory responses through Toll-like receptor 4 (TLR4) recognition. This activates the NF- κ B pathway resulting in inflammation [51]. However, when LPSs are sensed intracellularly, it stimulates pyroptosis. This pro-inflammatory programmed cell death releases inflammatory components (cytokines, DAMPs, and PAMPs) [52]. Linked through the AIM2 inflammasome, epithelial cell death is mediated, and intestinal radio-sensitivity is regulated through caspase-1 mediation [53]. Under such biological effects, healthy tissues are, together with cancer cells, negatively impacted by the progression and chronicity of radiation injury to the intestinal

wall. This can also be facilitated by excessive and chronic ROS and reactive nitrogen species (RNS) generation [46].

Bystander effects have significant implications for radiotherapy as they have the potential to influence treatment efficacy and increase the risk of adverse effects on normal tissues and an elevated risk of secondary malignancies. However, these effects may enhance the elimination of tumor cells by amplifying the extent of tumor damage. Hence, a comprehensive understanding of bystander effects' mechanisms and the development of strategies to regulate them may lead to better therapeutic outcomes for radiotherapy [13,54,55].

4. The Complex Relationship between the Gut Microbiota and the Gastrointestinal Tract

The human gut microbiota is known as the microorganisms that live collectively in the intestinal lumen and mucosal surfaces along the intestinal tract and interact with the different host cells. Studies have shown that the composition of the gut microbiota (notably defined with sequencing methods listing 9,879,896 genes [56]) influences the response to radiotherapy and gastrointestinal toxicity [33]. The different taxa differ in their capacity to modulate innate and systemic immune responses [49,57]. In addition, the members of the gut microbiota possess various physiological capacities required by the host for its functioning, including the biosynthesis of vitamins, steroid hormones, and neurotransmitters [33]; metabolism of xenobiotics [58]; modulation of intestinal epithelial cell turnover [59,60]; and immunomodulatory functions [61–65]. An imbalance of the proper functioning of the gut microbiota can occur if specific members of the gut microbiota are altered (increase in pathogenic members or decrease in protective commensals by ionizing radiation). Such modifications have been linked to several disorders, like colorectal cancer and inflammatory bowel disease [35,66–68]. This so-called dysbiosis state highlights the distinction between the impact of commensal microbiota, which plays a role in enteric immunity by shaping the development of the immune system and its responses, [68,69] and pathogenic bacteria. If not controlled by the commensal microbiota, the latter can take over and induce detrimental effects (Figure 2). Examples of this control include the production of inhibitory substances (secondary bile acids) or the consumption of limited common resources (carbohydrates), leading to the starvation of competing pathogens [70]. By stimulating inflammation, certain bacterial species, such as *Fusobacterium nucleatum*, can promote CRC initiation and progression (Figure 2) [33,71,72]. Another study modelled the interaction between a gut microbiota bacterium and the host through intestinal colonization with the microinjection of the non-pathogenic strain *Escherichia coli* ECOR2 into the lumen of human intestinal organoids (HIOs) [73]. The observed stimuli given by the bacterial contact demonstrated an increase in barrier functions (induction of mucus secretion, increased cell junction expression, measure of bacterial translocation), antimicrobial defense (secretion of antimicrobial peptides) and tissue maturation (analysis of principle component to determine tissue maturation status) [73]. Moreover, pathogenic *Clostridium difficile* strain VPI 10463 being microinjected in HIOs, showed to damage the epithelium with impaired barrier function [74].

Those studies reported the impacts that bacteria can have on the host. In the event of radiotherapy, the disruption of the gut microbiota and intestinal tissue can be escalated by each other. Indeed, exposure to radiation results in inflammation leading to gut microbiota dysbiosis that can also cause and influence inflammation in return [75]. This is connected to post-irradiation tissue damage and the pathogenic impact of specific bacteria in the microbiota [42,76]. Dysbiosis of the gut microbiota has been shown to make patients more susceptible to radiation damage and plays a role in aggravating intestinal inflammation (Figure 1) [42,75,77].

4.1. The Influence of the Gut Microbiota on Colorectal Cancer

CRC has direct contact with the gut microbiota as it originates from growth on the inner lining of the large intestine. The build-up of numerous independent genetic changes,

such as mutations of the *APC*, *TP53*, *KRAS*, *PIK3CA*, *FBXW7*, *SMAD4*, *TCF7L2*, and *NRAS* genes, leads to cancer development. These genes affect cell differentiation, proliferation, and apoptosis in cancer [78]. Aberrant proliferation results in the formation of polyps that can become malignant with the invasion of cancer cells into the submucosa [79,80]. The immune responses of the gut microbiota and host immune responses are linked through the inflammasome, as mentioned before in the radiotoxicity section. Recent research has highlighted the significance of the “inflammasome-microbiota axis” in several disease scenarios, including CRC [81], as the metabolism of the gut microbiota can have both positive and negative effects. In the case of dysbiosis, pathogenic strains can become prevalent over commensal strains. Pathogens such as *Escherichia coli* can, for example, produce B2-colicin which can induce DNA damage [82,83]. With this role in carcinogenesis, some strains have also been shown to overstimulate immune responses, such as *Bacteroides fragilis* via T helper 17 cells and *Peptostreptococcus anaerobius*, which notably modulates tumor-associated macrophages. The latter strain also interacts with colon cells via TLR2 and TLR4 [71].

4.2. Interactions between Bacteria and the Tumor Microenvironment

Solid tumors are sophisticated organs to which numerous other cells are attracted and diverted for their own benefit. The tumor microenvironment (TME) is a cellular environment created by the interactions between malignant and unaltered cells via the lymphatic and circulatory systems to affect the progression of cancer [84]. Its extracellular matrix is composed of various proteins including collagen, fibronectin, elastin, and laminin. This matrix serves as a physical scaffold for cells and can also contain cytokines and growth factors, such as proangiogenic factors including vascular endothelial growth factor (VEGF) and TGF- β . Together, these components not only provide physical support but also promote tumor progression [85].

In the TME, healthy cells play a crucial role in all stages of carcinogenesis by allowing and promoting unchecked cell proliferation [86]. Cancer-associated fibroblasts (CAFs) were notably linked to colorectal cancer recurrence, by producing growth factors, cytokines, and extracellular matrix [87]. Due to the vast heterogeneity of cell types (e.g., T cells, macrophages, CAFs [84]), this microenvironment has a huge impact on how tumors react to treatment [88]. As change in the gut microbiota's balance is systematically observed in patients with colorectal cancers, it has the potential to alter the TME and create a favorable environment for tumor growth [89]. Bacteria that live inside tumors have been found to be tumor type-specific, indicating a connection with tumor progression by increasing mutagenesis, regulating oncogenic pathways, and modulating the host immune system [90]. The latter is notably witnessed in CRC tissues, with *Fusobacterium nucleatum* that provides a pro-inflammatory environment by activating the NF- κ B pathway [91]. It also modifies the TME to evade anticancer immune responses by binding to fatty-acid-binding protein 2 (Fap2), an adhesin that binds to natural killer cells and other tumor-infiltrating lymphocytes via T-cell immunoglobulin (TIGIT) receptors [92].

5. Using Food Supplements to Mitigate Radiotoxicity

As radiation toxicity is a major concern for cancer patients undergoing radiotherapy, it is leading researchers to focus on the prevention and mitigation of its effects, notably using radioprotectors. There are currently only two radioprotective agents approved by the Food and Drug Administration: palifermin (Kepivance[®]) and amifostine (Ethyol[®]). The former is an artificial keratinocyte growth factor used to diminish severe oral mucositis. One of the potential concerns of its use regards its mitogen impact on epithelial cells which could promote tumorigenesis [3]. On the other hand, amifostine is an inactive phosphorylated aminothioliol prodrug. It is dephosphorylated by alkaline phosphatase to its active and free radical-scavenging sulfhydryl metabolite: WR-1065 [93]. It can selectively protect healthy tissues, such as the intestine, but may cause side effects, such as nausea and diarrhea, as well as allergic reactions [94]. In addition, its potential to also protect malignant tissues

from radiotherapy still needs clarification [95]. To avoid these side effects and reduce injury to healthy intestinal tissues, researchers have investigated the use of bacterial food supplements instead, as safer radioprotectors including probiotics and prebiotics [96,97]. They have been the primary focus of previous in vitro and in vivo research on radioprotection mechanisms, with antimicrobial, barrier-enhancing, and immunomodulatory capacities being the most extensively studied functions [57,71,96]. Intestinal cells treated with specific probiotics or prebiotics before and/or after radiation, have already suggested the potential of their action [97–99], but remain to be confirmed by dedicated clinical trials. In 2021, Tripathy et al. reviewed the impact of several probiotic strains on colorectal cancerous cell lines. For instance, *Lacticaseibacillus rhamnosus* GG was tested on the LT-97, HT-29, HCT-116, and Caco-2 cancer cell lines. This model probiotic strain is shown to stimulate apoptotic pathways in malignant cells, halt their proliferation, enhance the release of interleukin-8 (a chemokine that attracts immune cells including neutrophils), and modulate SCFA production [28]. As the product of microbial fermentation, the latter offers protective benefits to healthy cells by enhancing barrier function through tightening junctions and alleviating metabolic stress [28]. Colonocytes absorb these molecules and upregulate the expression of apoptosis-activating proteins in CRC cells [28]. They are essential as they restrict cancer tissue growth by modulating cell differentiation and proliferation through the regulation of cyclin expression at cell cycle checkpoints [100]. The retinoblastoma protein, a tumor suppressor, notably targets D-type cyclin as a substrate. Therefore, this protein enables cells in the G0 phase to progress to the G1 phase and regulates its progression. Moreover, A-type cyclin is also important as it participates in DNA synthesis but also helps prevent its overabundance by facilitating entry into the S phase, completion of the S phase, and entry into the M phase [100]. This highlights that proper microbial fermentation through commensal microorganisms, producing SCFAs, is essential.

The use of specific probiotics and prebiotics to complement CRC treatment has also been studied in clinical trials, showing evidence of their efficacy on the quality of life of patients experiencing fewer symptoms and lowered inflammatory marker concentrations, such as TNF- α levels [101,102]. Clinical trials recapitulated by Shuwen et al. showed that SCFAs, from microbial fermentation, have been used as therapeutic treatment against CRC cells [5]. One positive impact is the exertion of a regulatory influence on the immune response by promoting the production of anti-inflammatory cytokines [103]. In 2016, Mansouri-Tehrani et al. reported the impact of probiotics alone, coupled with honey, and placebo controlled in 67 patients with pelvic cancer receiving radiotherapy, who were exposed to a total dose between 40 and 50 Gy. Species including *L. casei*, *L. rhamnosus*, and *Bifidobacterium breve* were combined and administered orally before and during radiotherapy. The Food and Drug Administration and Isfahan University of Medical Sciences evaluated and authorized the use of probiotics and honey in this study [104]. Another clinical trial by Delia et al. regrouped 490 sigmoid, rectal, and cervical cancer patients following postoperative radiotherapy treatment with radiation doses within the range of 60–70 Gy with a combined solution, administered orally, of the following probiotics: *L. casei*, *L. acidophilus*, *L. rhamnosus*, *L. bulgaricus*, *B. breve*, *B. longum*, and *S. thermophiles* [104]. Patients in these clinical trials showed a reduced incidence of radiation-induced diarrhea and improved quality of life.

In parallel, studies testing the use of prebiotics showed their impact on colon cancer cells. Arun et al. reported that the fermentation (by probiotics) of the plantain inflorescence dietary fiber produces SCFAs and induces apoptosis of HT29 cells [105]. Moreover, Nowacka-Jechalke et al. studied the role of polysaccharides (prebiotic) from the mushroom *Cantharellus cibarius* in preventing and treating colon cancer [106]. Those metabolites act on inhibiting the proliferation of colon cancer cells and on stimulating *Lactobacillus* strains growth. While the existing literature provides the results of the efficacy of probiotics and prebiotics as adjuncts to radiotherapy, there is still missing data on the beneficial effects because of the low number of clinical trials investigating their use in this context. Furthermore, a significant proportion of the studies to date have focused on using food supplements in

the case of chemotherapy rather than radiotherapy treatment. Finally, the heterogeneity of studies with respect to radiation doses, and radiation administration frequency limits the general application of the findings.

6. Modeling Colorectal Radiotoxicity with Human Gut In Vitro Models

6.1. Radiobiology Models: Pros and Cons

Before reaching human clinical trials, animal testing was abundantly used in radiobiology. Animal research models, regulated by law, government policies, and ethical guidelines, provide a representation of the entire organism through its regulatory processes and responses [107]. Through genetic manipulation and disease induction, in vivo animal models allow the study of radiosensitivity as well as radioresistance [108]. This enables the modeling of conditions of radiotherapy treatment with disease reproduction and the study of the role of specific genes, such as ATM or p53, in radiobiology [109,110], or testing of treatment enhancing strategies such as food supplements [98,111]. In 1959, Russell and Burch introduced the 3Rs, defining the concepts of replacement, reduction, and refinement [112,113]. The goals are to find alternatives, reduce the number of animals used, and reduce the harshness of animal testing. Working with animals to study the interaction of gut microbiota and the gastrointestinal tract also presents a major issue. With its low degree of translation in terms of gut microbiota composition, it provides poor human relevance [114]. Alternatively, in vitro models using human cells can be used to get closer to the human reference. Cell culture plates deliver high-throughput screening, allow genetic manipulation, have the potential to provide a personalized approach, and are easy to handle [113,115]. For radiobiology, in vitro models have been used to complement in vivo preclinical studies with human relevance to test novel irradiation treatments, such as FLASH irradiation [116], optimize the conditions of exposure [117], and evaluate the potential of radiosensitizers [118] or prebiotic supplements [119]. However, cell culture models lack cell–cell interactions, do not give control of in vivo cell morphology, do not give a gut microbiota representation nor microenvironment management, creating a gap with in vivo representation [115,120].

6.2. Providing Human Relevance and Representation of Microorganisms–Host Interaction with Microphysiological Systems

Owing to these limitations, interest has grown in 3D models coupled with the use of microphysiological systems, which represent the complexity of living tissues more accurately [120,121]. In human-specific studies, the use of organoids, 3D organ-like models, allows for the adoption of a translational human approach [122]. To represent large intestine tissues, colorectal organoids contain differentiated enterocytes, goblet cells, and tuft cells, and can recapitulate developing crypts and mucus layers [123]. They can be co-cultured with immune cells to give a better microenvironment representation [124]. To mimic the responses of patients to treatment-induced toxicity [125–129], patient-derived tumor organoids (PDTOs) [130] can be used to provide an improved model to study the effects of irradiation on colorectal tissue and gut microbiota (Figure 3). With radiotherapy and PDTOs being an increasing focus of studies since 2018 [131], multiple models derived from rectal cancer patients undergoing radiotherapy have been implemented to understand resistance mechanisms to treatment, predict treatment outcomes, and evaluate new adjuvant treatment strategies, including food supplements [128,129,132–134]. However, organoids have a wide range of sizes and shapes, making it challenging to keep cells in stable positions in these structures for prolonged examination and to sample for analysis. Furthermore, limitations are posed with PDTOs being cultured with different protocols and in different conditions, creating variability in the existing studies [135]. Tissue–tissue interactions and multiscale architectures are absent in many systems, and cells are not typically exposed to physiological mechanical cues, such as fluid shear stress, tension, and compression. The latter affects how organoids grow and function in both health and

disease, suggesting that supplying these elements via microfluidic technology may increase representativeness and human relevance [121].

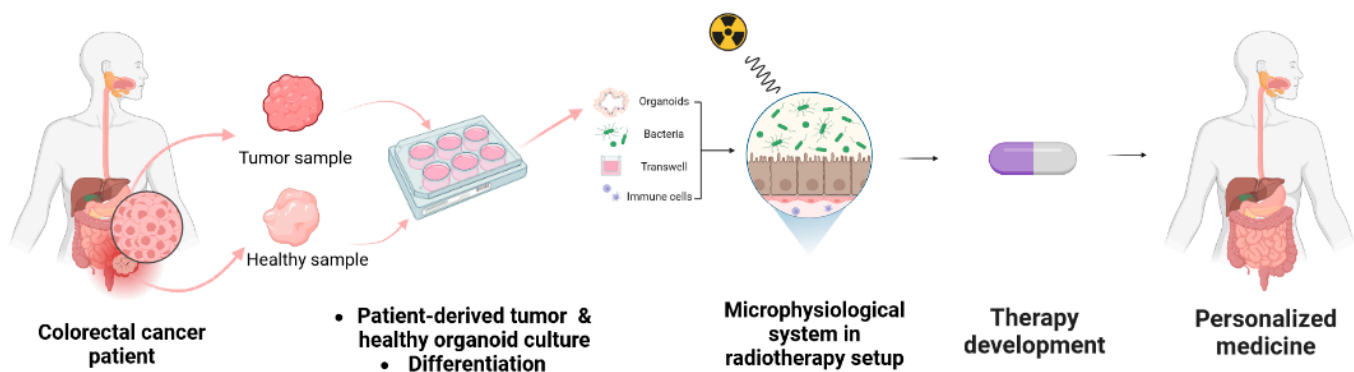


Figure 3. Set up of personalized medicine. Incorporating patient-derived samples into microphysiological systems under conditions mimicking radiotherapy enables the representation of treatment conditions and the development of novel therapies [86].

6.3. Implementing Microfluidic Technology

Radiobiology requires complex models to allow representation at both tissue and organ levels. Various cell types, their interactions, and the release of cytokines, growth factors, and chemokines are necessary to model radiation-induced toxicity, tumor radioresistance, and the interaction with the microenvironment [136]. Microfluidics allows for culturing in continuously perfused chambers and provides physiological models. They represent the function of targeted tissues, recreate the interface between different tissues, and provide cell–cell interactions. This technology can be used to maintain tissue structure and function [121]. Its features allow control of the microenvironment with the laminar flow of the culture medium and biocompatibility between the cultures and the material used [137]. Channels, valves, reservoirs, membranes, and other microscale fluid-handling compartments constitute a microfluidic platform. This enables integrated, automated, parallel, and miniaturized biochemical analyses in a consistent manner [115]. It allows representation of *in vitro* radiotherapy [5], the co-culture of epithelial cells with bacteria [138,139], the modeling of cancer [140,141], drug discovery with toxicity tests [142], and the study of the barrier function [143].

Such systems can be coupled with PDTs to establish microphysiological systems. The ideal gut microphysiological model, described in Table 1 (first column), mimics the structure, function, and physiology of the human intestine and its interaction with microorganisms from the gut microbiota. The use of organoids to form monolayers provides *in vitro* intestinal epithelium representation and access to both the luminal and basal sides. Recreating the oxygen gradient is necessary to allow co-culture of oxygen-sensitive microorganisms and primary colorectal epithelial cells. To simulate the lumen environment described previously, e.g., *Bifidobacterium* and *Lactobacillaceae* species (frequently employed as food supplements or probiotics as described above) require anaerobic conditions on the apical side. While the basolateral side requires aerobic conditions to mimic the oxygenated blood-rich lamina propria environment that contains resident immune cells. The hardware design allows fluid pumping to create apical and basolateral flow to model shear stress, respectively, remove spent medium, promote oxygenation, and supply oxygen needed for the epithelial cells [139]. To support cell attachment, a specific scaffold material that remains intact, compatible with radiation, and that does not absorb compounds is essential. However, considering the sensitivity of the host cells to living bacteria and their metabolites remains crucial. In 2011, Sato et al. published the first description of the long-term culture of epithelial organoids made from human colorectal and neoplastic tissues [144]. Another specificity that *in vitro* microphysiological systems can provide is the TME, as described previously. Growing PDTs from a patient's colorectal cancer sample coupled

with a microfluidic system (Figure 3) allows for the representation of colorectal cancer cell subtypes and their microenvironments [86,123]. This technology is particularly useful for precision medicine by enabling the use of patient samples (Figure 3) [122]. These cancer organoid (i.e., tumoroid) models from primary patient tumor cells can allow the study of treatments and their efficiency [145], by reproducing the contact of the epithelium with microorganisms. This raises the question of whether combining a microphysiological system with a radiotherapy setup can provide representative treatment conditions. Indeed, radiotherapy treatment presents challenges in its establishment with the timing, irradiation dose, dose rate, and fractionation. Such a representation would allow the study of the effect of radiation on colorectal cancer tissue and new personalized treatment strategies such as the use of food supplements [136].

Table 1. Overview of gut microphysiological models compared to the ideal human relevant model.

Models	Ideal Gut Microphysiological Model †	GuMI [146,147]	EACC [148]	HuMix Model [149] HMI Module [150]	Gut Chip [151,152]	AOI Chip [153]	Nicole C. Roy [154]	HoxBan [155]
Cell types	Epithelial cells, goblet cells, innate immune cells, adaptive immune cells, bacterial cells	Primary colon epithelial cells	Human primary jejunal enteroids (4 donors)	Caco-2 cells; CCD-18Co	Caco-2 cells/primary cells	Caco-2BBE cells	Caco-2 cells	Caco-2 cells
Cell architecture	Monolayer with villus shape	Monolayer	Monolayer	Monolayer	Microfluidic chip	Microfluidic chip	Monolayer	Monolayer
Barrier function	Mucus, TEER measurements close to the in vivo: 300–400 Ω cm^2 in the large intestine [156]	Yes, TEER measurements significantly higher than 300 Ω cm^2	Yes, only expressed in fold changes in TEER. Increase after bacteria exposure	TEER measurements, 1000 Ω cm^2	TEER measurements > 2500 Ω cm^2	TEER measurements: 5–10 k Ω cm^2	60% decrease (even in control) in first 1 h, then recover.	No values, cells attached to glass slide
Contact of bacteria and host cells	Direct contact or separate by mucus layer	Yes	Yes	No, separated by membrane	Yes	Yes	Yes	Yes
Anaerobic conditions (oxygen level)	Yes, ideally gradient anaerobic conditions	Anaerobic, low oxygen content	Yes	0.1% O_2 ; oxygen optode sensors measurement	Yes	Yes	Yes	Yes
Co-culture maintaining time	Yes, ideally one week or more *	5 days	8–24 h	Up to 2 days	Up to 5 days	Up to 7 days	Half a day	Up to 1 day and a half
Static or flow	Flow, in both apical and basal sides	Flow, 600 $\mu\text{L}/\text{h}$	Static	Flow, 1500 $\mu\text{L}/\text{h}$ [0.416 $\mu\text{L}/\text{s}$]	Flow, 30 $\mu\text{L}/\text{h}$ [0.05 $\mu\text{L}/\text{min}$]	Flow, 50 $\mu\text{L}/\text{h}$	Static	Static
Immune cells	Yes, both innate and adaptive immune cells	dendritic cells, macrophages, CD4+ T cells	No	CD4+ T cells in third layer	PBMCs	No	No	No
Bacterial species	Oxygen sensitive strain	<i>Faecalibacterium prausnitzii</i> , <i>Eubacterium rectale</i> , <i>Bacteroides thetaiotaomicron</i>	<i>B. thetaiotaomicron</i> and <i>Blautia</i> sp.	<i>Lactocaseibacillus rhamnosus</i> GG <i>Bacteroides caccae</i>	<i>E. coli</i> , probiotic mix (<i>Lactobacillus acidophilus</i> , <i>Lactobacillus plantarum</i> , <i>Lactobacillus paracasei</i> , <i>Bifidobacterium breve</i> , <i>B. longum</i> , <i>B. infantis</i>)	<i>Bifidobacterium adolescentis</i> and <i>Eubacterium hallii</i>	<i>F. prausnitzii</i> DSM17677	<i>F. prausnitzii</i>
Hardware material	Biocompatible, intact while running	Monolithic polysulfone (low absorption of hydrophobic compounds)	Not specified	polycarbonate (PC) (low absorption of hydrophobic compounds)	polydimethylsiloxane (PDMS) (high absorption of hydrophobic compounds)	PDMS (high absorption of hydrophobic compounds)	Not specified	Glass for cell attachment and agar for bacteria (low absorption of hydrophobic compounds)
Stress applied ($\mu\text{m}/\text{s}$)	Fluid shear stress: ~0–2.5 μPa [157]	Shear stress: 0–11 μPa	Not applied	Not specified	Not specified	Shear stress: 3–10 μPa	Not applied	Not applied

† Based on the gastrointestinal tract physiology. * One week of co-culturing is the target because differentiated cells are used in the model and cannot model the physiological replacement of dead cells by stem cells. TEER = trans-epithelial electrical resistance; PBMC = peripheral blood mononuclear cells.

6.4. Radiotherapy Setup for the Evaluation of Novel Treatments

The choice of cell type and the *in vitro* microenvironment is crucial for achieving an ideal model that closely mimics *in vivo* tissue. Using PDOs instead of immortalized cancer cell lines, can preserve tissue characteristics that are otherwise lost. In addition, accurately controlling oxygen levels allows for the aforementioned gradient of the colorectal cancer environment to be replicated. To show the human relevance of tumoroids, Vlachogiannis et al. compared PDOs from 71 colorectal cancer patients to parental tissue biopsy, showing significant similarities in the pattern of tumor expression such as the responsiveness to treatment [145]. Current models (Table 1) including both human immortalized cell lines and microbial cells enable maintaining this kind of co-culture for up to seven days using biocompatible hardware materials. The latter also need to be suitable for the type of study of interest. In radiobiology studies, the hardware must be resistant and compatible with ionizing irradiation. Previously used in human gut models, (Table 1) polydimethylsiloxane (PDMS) has many useful physical properties. However, it showed that it absorbs hydrophobic compounds such as pharmaceutical compounds (ciprofloxacin and paclitaxel) [158]. This can have a detrimental impact on the biological functioning and measurements by modifying the microenvironment composition by altering the concentrations of essential components of the cell culture (amino acids, growth factors, vitamins, etc.). Other materials such as monolithic polysulfone, that have been shown to be inert, seem to be more suitable [146].

To study the impact of radiotherapy on the gut and its microbiota, a model that provides oxygen control with both apical anaerobic conditions for bacteria, and basolateral aerobic conditions for epithelial and immune cells has been developed [147]. That complete system could enable the generation of high-content data on the testing of various food supplements type and concentration to prevent radiation induced toxicity (Figure 3). Further studies on microphysiological systems would provide more data to support their implementation by pharmaceuticals industries.

7. Conclusions

As radiotherapy is a pillar of CRC treatment, counteracting its radiotoxicity remains a priority. Research is increasingly exploring ways to do so through *in vivo*, *in vitro*, and clinical studies. A central point of focus within this field revolves around investigating the radioprotective mechanisms of food supplements. Evidence of their beneficial impact on health by balancing the gut microbiota, enhancing barrier function, and immunomodulation has been observed. Human derived organoids have emerged as innovative 3D tools for studying human biology and disease modeling to address the limitations of existing *in vitro* cell lines and animal radiobiology models in terms of human relevance. Coupling them with microfluidics to design microphysiological systems can provide a representation of cellular components and microenvironments. To assess radioprotective candidates, such as food supplements, there is a need to provide human-relevant bacteria–host co-culture representation under radiotherapy treatment conditions. This model of gut radiotoxicity offers a deeper mechanistic insight. Given the significant role of the microorganisms of the gut microbiota in modulating systemic immune responses that affect patient outcomes, combining tumor organoids with such a model exposed to radiotherapy presents a promising opportunity. However, the development of these models also comes with limitations. The establishment of organoid protocols lacks standardization, which may lead to variability in study outcomes. Furthermore, microphysiological models of the human gut have yet to implement the immune system and the gut microbiota. Also, by only featuring differentiated cells, the replacement of dead cells by stem cells and the bystander effect are not modeled. Still, using the described model, research would provide more human-relevant data, which could guide further translation in human patients by performing clinical trials.

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References

1. *Colorectal Cancer Statistics | How Common Is Colorectal Cancer?* American Cancer Society: Kennesaw, GA, USA, 2023.
2. *Globocan 2018 Latest Global Cancer Data—IAARC*; Union for International Cancer Control: Geneva, Switzerland, 2018.
3. Vendrely, V.; Rivin Del Campo, E.; Modesto, A.; Jolnerowski, M.; Meillan, N.; Chiavassa, S.; Serre, A.A.; Gérard, J.P.; Créhanges, G.; Huguet, F.; et al. Rectal cancer radiotherapy. *Cancer/Radiothérapie* **2022**, *26*, 272–278. [[CrossRef](#)] [[PubMed](#)]
4. Roeder, F.; Meldolesi, E.; Gerum, S.; Valentini, V.; Rödel, C. Recent advances in (chemo-)radiation therapy for rectal cancer: A comprehensive review. *Radiat. Oncol.* **2020**, *15*, 262. [[CrossRef](#)] [[PubMed](#)]
5. Chermat, R.; Ziaee, M.; Mak, D.Y.; Refet-Mollof, E.; Rodier, F.; Wong, P.; Carrier, J.F.; Kamio, Y.; Gervais, T. Radiotherapy on-chip: Microfluidics for translational radiation oncology. *Lab A Chip* **2022**, *22*, 2065–2079. [[CrossRef](#)] [[PubMed](#)]
6. Grubbé, E.H. Priority in the Therapeutic Use of X-rays. *Radiology* **1933**, *21*, 156–162. [[CrossRef](#)]
7. Bernier, J.; Hall, E.J.; Giaccia, A. Radiation oncology: A century of achievements. *Nat. Rev. Cancer* **2004**, *4*, 737–747. [[CrossRef](#)] [[PubMed](#)]
8. Schaeue, D.; McBride, W.H. Opportunities and challenges of radiotherapy for treating cancer. *Nat. Rev. Clin. Oncol.* **2015**, *12*, 527–540. [[CrossRef](#)] [[PubMed](#)]
9. Baskar, R.; Lee, K.A.; Yeo, R.; Yeoh, K.W. Cancer and Radiation Therapy: Current Advances and Future Directions. *Int. J. Med. Sci.* **2012**, *9*, 193–199. [[CrossRef](#)]
10. Minniti, G.; Goldsmith, C.; Brada, M. Radiotherapy. *Handb. Clin. Neurol.* **2012**, *104*, 215–228. [[CrossRef](#)]
11. Dagenet, E.; Khalifa, J.; Tolédano, A.; Borchiellini, D.; Pointreau, Y.; Rodriguez-Lafrasse, C.; Chargari, C.; Magné, N. To exploit the 5 ‘R’ of radiobiology and unleash the 3 ‘E’ of immunoeediting: ‘RE’-inventing the radiotherapy-immunotherapy combination. *Ther. Adv. Med. Oncol.* **2020**, *12*. [[CrossRef](#)]
12. Thoms, J.; Bristow, R.G. DNA Repair Targeting and Radiotherapy: A Focus on the Therapeutic Ratio. *Semin. Radiat. Oncol.* **2010**, *20*, 217–222. [[CrossRef](#)]
13. Prise, K.M.; O’Sullivan, J.M. Radiation-induced bystander signalling in cancer therapy. *Nat. Rev. Cancer* **2009**, *9*, 351–360. [[CrossRef](#)] [[PubMed](#)]
14. Kargiotis, O.; Geka, A.; Rao, J.S.; Kyritsis, A.P. Effects of irradiation on tumor cell survival, invasion and angiogenesis. *J. Neuro-Oncol.* **2010**, *100*, 323–338. [[CrossRef](#)] [[PubMed](#)]
15. Baskar, R.; Dai, J.; Wenlong, N.; Yeo, R.; Yeoh, K.W. Biological response of cancer cells to radiation treatment. *Front. Mol. Biosci.* **2014**, *1*, 24. [[CrossRef](#)] [[PubMed](#)]
16. Di Maggio, F.M.; Minafra, L.; Forte, G.I.; Cammarata, F.P.; Lio, D.; Messa, C.; Gilardi, M.C.; Bravatà, V. Portrait of inflammatory response to ionizing radiation treatment. *J. Inflamm.* **2015**, *12*, 14. [[CrossRef](#)] [[PubMed](#)]
17. Shan, Y.X.; Jin, S.Z.; Liu, X.D.; Liu, Y.; Liu, S.Z. Ionizing radiation stimulates secretion of pro-inflammatory cytokines: Dose-response relationship, mechanisms and implications. *Radiat. Environ. Biophys.* **2007**, *46*, 21–29. [[CrossRef](#)]
18. Rodríguez-Ruiz, M.E.; Vitale, I.; Harrington, K.J.; Melero, I.; Galluzzi, L. Immunological impact of cell death signaling driven by radiation on the tumor microenvironment. *Nat. Immunol.* **2019**, *21*, 120–134. [[CrossRef](#)]
19. Nicholas, S.; Chen, L.; Choflet, A.; Fader, A.; Guss, Z.; Hazell, S.; Song, D.Y.; Tran, P.T.; Viswanathan, A.N. Pelvic Radiation and Normal Tissue Toxicity. *Semin. Radiat. Oncol.* **2017**, *27*, 358–369. [[CrossRef](#)]
20. Espey, M.G. Role of oxygen gradients in shaping redox relationships between the human intestine and its microbiota. *Free. Radic. Biol. Med.* **2013**, *55*, 130–140. [[CrossRef](#)]
21. Possenti, L.; Mecchi, L.; Rossoni, A.; Sangalli, V.; Bersini, S.; Cicchetti, A.; Costantino, M.L.; Candrian, C.; Arrigoni, C.; Rancati, T.; et al. Radiobiological Studies of Microvascular Damage through In Vitro Models: A Methodological Perspective. *Cancers* **2021**, *13*, 1182. [[CrossRef](#)]
22. Eaton, S.E.; Kaczmarek, J.; Mahmood, D.; McDiarmid, A.M.; Norarfan, A.N.; Scott, E.G.; Then, C.K.; Tsui, H.Y.; Kiltie, A.E. Exploiting dietary fibre and the gut microbiota in pelvic radiotherapy patients. *Br. J. Cancer* **2022**, *127*, 2087–2098. [[CrossRef](#)]
23. Macpherson, A.J.; Harris, N.L. Interactions between commensal intestinal bacteria and the immune system. *Nat. Rev. Immunol.* **2004**, *4*, 478–485. [[CrossRef](#)] [[PubMed](#)]

24. Wedlake, L.J. Nutritional strategies to prevent gastrointestinal toxicity during pelvic radiotherapy. *Proc. Nutr. Soc.* **2018**, *77*, 357–368. [[CrossRef](#)]
25. Barker, N.; van Es, J.H.; Kuipers, J.; Kujala, P.; van den Born, M.; Cozijnsen, M.; Haegebarth, A.; Korving, J.; Begthel, H.; Peters, P.J.; et al. Identification of stem cells in small intestine and colon by marker gene Lgr5. *Nature* **2007**, *449*, 1003–1007. [[CrossRef](#)] [[PubMed](#)]
26. Radtke, F.; Clevers, H. Self-Renewal and Cancer of the Gut: Two Sides of a Coin. *Science* **2005**, *307*, 1904–1909. [[CrossRef](#)] [[PubMed](#)]
27. Goto, Y.; Ivanov, I.I. Intestinal epithelial cells as mediators of the commensal–host immune crosstalk. *Immunol. Cell Biol.* **2013**, *91*, 204–214. [[CrossRef](#)] [[PubMed](#)]
28. Tripathy, A.; Dash, J.; Kancharla, S.; Kolli, P.; Mahajan, D.; Senapati, S.; Jena, M.K. Probiotics: A Promising Candidate for Management of Colorectal Cancer. *Cancers* **2021**, *13*, 3178. [[CrossRef](#)]
29. Silva-Sanchez, A.; Randall, T.D. Chapter 2—Anatomical Uniqueness of the Mucosal Immune System (GALT, NALT, iBALT) for the Induction and Regulation of Mucosal Immunity and Tolerance. In *Mucosal Vaccines, 2nd*; Kiyono, H., Pascual, D.W., Eds.; Academic Press: Cambridge, MA, USA, 2020; pp. 21–54.
30. Zheng, L.; Kelly, C.J.; Colgan, S.P. Physiologic hypoxia and oxygen homeostasis in the healthy intestine. A review in the theme: Cellular responses to hypoxia. *Am. J. Physiol.—Cell Physiol.* **2015**, *309*, C350–C360. [[CrossRef](#)]
31. Konjar, Š.; Pavšič, M.; Veldhoen, M. Regulation of Oxygen Homeostasis at the Intestinal Epithelial Barrier Site. *Int. J. Mol. Sci.* **2021**, *22*, 9170. [[CrossRef](#)]
32. Pral, L.P.; Fachi, J.L.; Corrêa, R.O.; Colonna, M.; Vinolo, M.A.R. Hypoxia and HIF-1 as key regulators of gut microbiota and host interactions. *Trends Immunol.* **2021**, *42*, 604–621. [[CrossRef](#)]
33. Lynch, S.V.; Pedersen, O. The Human Intestinal Microbiome in Health and Disease. *N. Engl. J. Med.* **2016**, *375*, 2369–2379. [[CrossRef](#)]
34. Andreyev, J. Gastrointestinal complications of pelvic radiotherapy: Are they of any importance? *Gut* **2005**, *54*, 1051–1054. [[CrossRef](#)] [[PubMed](#)]
35. Manichanh, C.; Varela, E.; Martinez, C.; Antolin, M.; Llopis, M.; Doré, J.; Giralt, J.; Guarner, F.; Malagelada, J.R. The gut microbiota predispose to the pathophysiology of acute postradiotherapy diarrhea. *Off. J. Am. Coll. Gastroenterol.* **2008**, *103*, 1754–1761. [[CrossRef](#)] [[PubMed](#)]
36. Wang, L.; Wang, X.; Zhang, G.; Ma, Y.; Zhang, Q.; Li, Z.; Ran, J.; Hou, X.; Geng, Y.; Yang, Z.; et al. The impact of pelvic radiotherapy on the gut microbiome and its role in radiation-induced diarrhoea: A systematic review. *Radiat. Oncol.* **2021**, *16*, 187. [[CrossRef](#)]
37. Nam, Y.D.; Kim, H.J.; Seo, J.G.; Kang, S.W.; Bae, J.W. Impact of Pelvic Radiotherapy on Gut Microbiota of Gynecological Cancer Patients Revealed by Massive Pyrosequencing. *PLoS ONE* **2013**, *8*, e82659. [[CrossRef](#)] [[PubMed](#)]
38. Fernandes, A.; Oliveira, A.; Guedes, C.; Fernandes, R.; Soares, R.; Barata, P. Ionizing Radiation from Radiopharmaceuticals and the Human Gut Microbiota: An Ex Vivo Approach. *Int. J. Mol. Sci.* **2022**, *23*, 10809. [[CrossRef](#)] [[PubMed](#)]
39. Jalili-Firoozinezhad, S.; Prantil-Baun, R.; Jiang, A.; Potla, R.; Mammoto, T.; Weaver, J.C.; Ferrante, T.C.; Kim, H.J.; Cabral, J.M.S.; Levy, O.; et al. Modeling radiation injury-induced cell death and countermeasure drug responses in a human Gut-on-a-Chip. *Cell Death Dis.* **2018**, *9*, 223. [[CrossRef](#)]
40. Andreyev, H.J.N.; Wotherspoon, A.; Denham, J.W.; Hauer-Jensen, M. Defining pelvic-radiation disease for the survivorship era. *Lancet Oncol.* **2010**, *11*, 310–312. [[CrossRef](#)]
41. Andreyev, H.J.N.; Wotherspoon, A.; Denham, J.W.; Hauer-Jensen, M. “Pelvic radiation disease”: New understanding and new solutions for a new disease in the era of cancer survivorship. *Scand. J. Gastroenterol.* **2011**, *46*, 389–397. [[CrossRef](#)]
42. Gerassy-Vainberg, S.; Blatt, A.; Danin-Poleg, Y.; Gershovich, K.; Sabo, E.; Nevelsky, A.; Daniel, S.; Dahan, A.; Ziv, O.; Dheer, R.; et al. Radiation induces proinflammatory dysbiosis: Transmission of inflammatory susceptibility by host cytokine induction. *Gut* **2018**, *67*, 97–107. [[CrossRef](#)]
43. Ahlberg, K.; Ekman, T.; Gaston-Johansson, F. The experience of fatigue, other symptoms and global quality of life during radiotherapy for uterine cancer. *Int. J. Nurs. Stud.* **2005**, *42*, 377–386. [[CrossRef](#)]
44. Kirsch, D.G.; Diehn, M.; Kesarwala, A.H.; Maity, A.; Morgan, M.A.; Schwarz, J.K.; Bristow, R.; Demaria, S.; Eke, I.; Griffin, R.J.; et al. The Future of Radiobiology. *JNCI J. Natl. Cancer Inst.* **2018**, *110*, 329–340. [[CrossRef](#)] [[PubMed](#)]
45. Huh, J.W.; Tanksley, J.; Chino, J.; Willett, C.G.; Dewhirst, M.W. Long-term consequences of pelvic irradiation: Toxicities, challenges, and therapeutic opportunities with pharmacologic mitigators. *Clin. Cancer Res.* **2020**, *26*, 3079–3090. [[CrossRef](#)] [[PubMed](#)]
46. François, A.; Milliat, F.; Guipaud, O.; Benderitter, M. Inflammation and immunity in radiation damage to the gut mucosa. *BioMed Res. Int.* **2013**, *2013*, 123241. [[CrossRef](#)] [[PubMed](#)]
47. Kim, T.-R.; Yoon, J.-H.; Kim, Y.-C.; Yook, Y.-H.; Kim, I.G.; Kim, Y.-S.; Lee, H.; Paik, S.-G. LPS-induced CD53 expression: A protection mechanism against oxidative and radiation stress. *Mol. Cells* **2004**, *17*, 125–131.
48. López, M.; Martín, M. Medical management of the acute radiation syndrome. *Rep. Pract. Oncol. Radiother.* **2011**, *16*, 138–146. [[CrossRef](#)]
49. Schluter, J.; Peled, J.U.; Taylor, B.P.; Markey, K.A.; Smith, M.; Taur, Y.; Niehus, R.; Staffas, A.; Dai, A.; Fontana, E.; et al. The gut microbiota is associated with immune cell dynamics in humans. *Nature* **2020**, *588*, 303–307. [[CrossRef](#)] [[PubMed](#)]
50. Man, S.M. Inflammasomes in the gastrointestinal tract: Infection, cancer and gut microbiota homeostasis. *Nat. Rev. Gastroenterol. Hepatol.* **2018**, *15*, 721–737. [[CrossRef](#)]

51. Rathinam, V.A.K.; Zhao, Y.; Shao, F. Innate immunity to intracellular LPS. *Nat. Immunol.* **2019**, *20*, 527–533. [[CrossRef](#)]
52. Wright, S.S.; Vasudevan, S.O.; Rathinam, V.A. Mechanisms and Consequences of Noncanonical Inflammasome-Mediated Pyroptosis. *J. Mol. Biology* **2022**, *434*, 167245. [[CrossRef](#)]
53. Hu, B.; Jin, C.; Li, H.B.; Tong, J.; Ouyang, X.; Cetinbas, N.M.; Zhu, S.; Strowig, T.; Lam, F.C.; Zhao, C.; et al. The DNA-sensing AIM2 inflammasome controls radiation-induced cell death and tissue injury. *Science* **2016**, *354*, 765–768. [[CrossRef](#)]
54. Ward, J.F. The radiation-induced lesions which trigger the bystander effect. *Mutat. Res./Fundam. Mol. Mech. Mutagen.* **2002**, *499*, 151–154. [[CrossRef](#)] [[PubMed](#)]
55. Butterworth, K.T.; McMahon, S.J.; Hounsell, A.R.; O’Sullivan, J.M.; Prise, K.M. Bystander Signalling: Exploring Clinical Relevance Through New Approaches and New Models. *Clin. Oncol.* **2013**, *25*, 586–592. [[CrossRef](#)]
56. Thursby, E.; Juge, N. Introduction to the human gut microbiota. *Biochem. J.* **2017**, *474*, 1823–1836. [[CrossRef](#)]
57. Yang, W.; Cong, Y. Gut microbiota-derived metabolites in the regulation of host immune responses and immune-related inflammatory diseases. *Cell. Mol. Immunol.* **2021**, *18*, 866–877. [[CrossRef](#)]
58. Krul, C.; Humblot, C.E.; Philippe, C.; Vermeulen, M.; Van Nuenen, M.; Havenaar, R.; Rabot, S. Metabolism of sinigrin (2-propenyl glucosinolate) by the human colonic microflora in a dynamic in vitro large-intestinal model. *Carcinogenesis* **2002**, *23*, 1009–1016. [[CrossRef](#)]
59. Leshner, S.; Walburg, H.E.; Sacher, G.A. Generation cycle in the duodenal crypt cells of germ-free and conventional mice. *Nature* **1964**, *202*, 884–886. [[CrossRef](#)]
60. Savage, D.C.; Siegel, J.E.; Snellen, J.E.; Whitt, D.D. Transit time of epithelial cells in the small intestines of germfree mice and ex-germfree mice associated with indigenous microorganisms. *Appl. Environ. Microbiol.* **1981**, *42*, 996–1001. [[CrossRef](#)]
61. Macpherson, A.J.; Uhr, T. Induction of Protective IgA by Intestinal Dendritic Cells Carrying Commensal Bacteria. *Science* **2004**, *303*, 1662–1665. [[CrossRef](#)]
62. Lee, Y.K.; Mazmanian, S.K. Has the microbiota played a critical role in the evolution of the adaptive immune system? *Science* **2010**, *330*, 1768–1773. [[CrossRef](#)] [[PubMed](#)]
63. Noverr, M.C.; Huffnagle, G.B. Does the microbiota regulate immune responses outside the gut? *Trends Microbiol.* **2004**, *12*, 562–568. [[CrossRef](#)] [[PubMed](#)]
64. Mazmanian, S.K.; Cui, H.L.; Tzianabos, A.O.; Kasper, D.L. An Immunomodulatory Molecule of Symbiotic Bacteria Directs Maturation of the Host Immune System. *Cell* **2005**, *122*, 107–118. [[CrossRef](#)]
65. Bauer, H.; Horowitz, R.E.; Levenson, S.M.; Popper, H. The Response of the Lymphatic Tissue to the Microbial Flora. *Studies on Germfree Mice. Am. J. Pathol.* **1963**, *42*, 471.
66. Fernandes, D.C.R.; Andreyev, H.J.N. Gastrointestinal Toxicity of Pelvic Radiotherapy: Are We Letting Women Down? *Clin. Oncol.* **2021**, *33*, 591–601. [[CrossRef](#)]
67. Liu, J.; Liu, C.; Yue, J. Radiotherapy and the gut microbiome: Facts and fiction. *Radiat. Oncol.* **2021**, *16*, 9. [[CrossRef](#)]
68. Lozupone, C.A.; Stombaugh, J.I.; Gordon, J.I.; Jansson, J.K.; Knight, R. Diversity, stability and resilience of the human gut microbiota. *Nature* **2012**, *489*, 220–230. [[CrossRef](#)] [[PubMed](#)]
69. Shiao, S.L.; Kershaw, K.M.; Limon, J.J.; You, S.; Yoon, J.; Ko, E.Y.; Guarnerio, J.; Potdar, A.A.; McGovern, D.P.B.; Bose, S.; et al. Commensal bacteria and fungi differentially regulate tumor responses to radiation therapy. *Cancer Cell* **2021**, *39*, 1202–1213.e1206. [[CrossRef](#)]
70. Rolhion, N.; Chassaing, B. When pathogenic bacteria meet the intestinal microbiota. *Philos. Trans. R. Soc. B Biol. Sci.* **2016**, *371*, 20150504. [[CrossRef](#)]
71. Cheng, W.Y.; Wu, C.Y.; Yu, J. The role of gut microbiota in cancer treatment: Friend or foe? *Gut* **2020**, *69*, 1867–1876. [[CrossRef](#)]
72. Helmink, B.A.; Khan, M.W.; Hermann, A.; Gopalakrishnan, V.; Wargo, J.A. The microbiome, cancer, and cancer therapy. *Nature* **2019**, *25*, 377–388. [[CrossRef](#)] [[PubMed](#)]
73. Hill, D.R.; Huang, S.; Nagy, M.S.; Yadagiri, V.K.; Fields, C.; Mukherjee, D.; Bons, B.; Dedhia, P.H.; Chin, A.M.; Tsai, Y.-H.; et al. Bacterial colonization stimulates a complex physiological response in the immature human intestinal epithelium. *eLife* **2017**, *6*, e29132. [[CrossRef](#)] [[PubMed](#)]
74. Leslie, J.L.; Huang, S.; Opp, J.S.; Nagy, M.S.; Kobayashi, M.; Young, V.B.; Spence, J.R. Persistence and Toxin Production by *Clostridium difficile* within Human Intestinal Organoids Result in Disruption of Epithelial Paracellular Barrier Function. *Infect. Immun.* **2015**, *83*, 138–145. [[CrossRef](#)] [[PubMed](#)]
75. Segers, C.; Mysara, M.; Claesen, J.; Baatout, S.; Leys, N.; Lebeer, S.; Verslegers, M.; Mastroleo, F. Intestinal mucositis precedes dysbiosis in a mouse model for pelvic irradiation. *ISME Commun.* **2021**, *1*, 24. [[CrossRef](#)] [[PubMed](#)]
76. Fernandes, A.; Oliveira, A.; Soares, R.; Barata, P. The effects of ionizing radiation on gut microbiota, a systematic review. *Nutrients* **2021**, *13*, 3025. [[CrossRef](#)] [[PubMed](#)]
77. Hovdenak, N.; Fajardo, L.F.; Hauer-Jensen, M. Acute radiation proctitis: A sequential clinicopathologic study during pelvic radiotherapy. *Int. J. Radiat. Oncol. Biol. Phys.* **2000**, *48*, 1111–1117. [[CrossRef](#)]
78. Muzny, D.M.; Bainbridge, M.N.; Chang, K.; Dinh, H.H.; Drummond, J.A.; Fowler, G.; Kovar, C.L.; Lewis, L.R.; Morgan, M.B.; Newsham, I.F.; et al. Comprehensive molecular characterization of human colon and rectal cancer. *Nature* **2012**, *487*, 330–337. [[CrossRef](#)]
79. Aarons, C.B.; Shanmugan, S.; Bleier, J.I.S. Management of malignant colon polyps: Current status and controversies. *World J. Gastroenterol.* **2014**, *20*, 16178. [[CrossRef](#)]

80. Bujanda, L. Malignant colorectal polyps. *World J. Gastroenterol.* **2010**, *16*, 3103. [[CrossRef](#)]
81. Privitera, G.; Rana, N.; Scaldaferrri, F.; Armuzzi, A.; Pizarro, T.T. Novel Insights Into the Interactions Between the Gut Microbiome, Inflammasomes, and Gasdermins During Colorectal Cancer. *Front. Cell. Infect. Microbiol.* **2022**, *11*, 806680. [[CrossRef](#)]
82. Dalmasso, G.; Cougnoux, A.; Delmas, J.; Darfeuille-Michaud, A.; Bonnet, R. The bacterial genotoxin colibactin promotes colon tumor growth by modifying the tumor microenvironment. *Gut Microbes* **2015**, *5*, 675–680. [[CrossRef](#)]
83. Buc, E.; Dubois, D.; Sauvanet, P.; Raisch, J.; Delmas, J.; Darfeuille-Michaud, A.; Pezet, D.; Bonnet, R. High Prevalence of Mucosa-Associated *E. coli* Producing Cyclomodulin and Genotoxin in Colon Cancer. *PLoS ONE* **2013**, *8*, e56964. [[CrossRef](#)]
84. Balkwill, F.R.; Capasso, M.; Hagemann, T. The tumor microenvironment at a glance. *J. Cell Sci.* **2012**, *125*, 5591–5596. [[CrossRef](#)] [[PubMed](#)]
85. Anderson, N.M.; Simon, M.C. The tumor microenvironment. *Curr. Biol.* **2020**, *30*, R921–R925. [[CrossRef](#)]
86. Hanahan, D.; Coussens Lisa, M. Accessories to the Crime: Functions of Cells Recruited to the Tumor Microenvironment. *Cancer Cell* **2012**, *21*, 309–322. [[CrossRef](#)]
87. Roma-Rodrigues, C.; Mendes, R.; Baptista, P.V.; Fernandes, A.R. Targeting Tumor Microenvironment for Cancer Therapy. *Int. J. Mol. Sci.* **2019**, *20*, 840. [[CrossRef](#)]
88. Neal, J.T.; Li, X.; Zhu, J.; Giangarra, V.; Grzeskowiak, C.L.; Ju, J.; Liu, I.H.; Chiou, S.H.; Salahudeen, A.A.; Smith, A.R.; et al. Organoid Modeling of the Tumor Immune Microenvironment. *Cell* **2018**, *175*, 1972–1988.e1916. [[CrossRef](#)]
89. Wang, L.; Peng, F.; Peng, C.; Du, J.-R. Gut Microbiota in Tumor Microenvironment: A Critical Regulator in Cancer Initiation and Development as Potential Targets for Chinese Medicine. *Am. J. Chin. Med.* **2021**, *49*, 609–626. [[CrossRef](#)]
90. Wong-Rolle, A.; Wei, H.K.; Zhao, C.; Jin, C. Unexpected guests in the tumor microenvironment: Microbiome in cancer. *Protein Cell* **2021**, *12*, 426–435. [[CrossRef](#)]
91. Yachida, S.; Mizutani, S.; Shiroma, H.; Shiba, S.; Nakajima, T.; Sakamoto, T.; Watanabe, H.; Masuda, K.; Nishimoto, Y.; Kubo, M.; et al. Metagenomic and metabolomic analyses reveal distinct stage-specific phenotypes of the gut microbiota in colorectal cancer. *Nat. Med.* **2019**, *25*, 968–976. [[CrossRef](#)]
92. Brennan, C.A.; Garrett, W.S. *Fusobacterium nucleatum*—Symbiont, opportunist and oncobacterium. *Nat. Rev. Microbiol.* **2019**, *17*, 156–166. [[CrossRef](#)] [[PubMed](#)]
93. Singh, V.K.; Seed, T.M. The efficacy and safety of amifostine for the acute radiation syndrome. *Expert Opin. Drug Saf.* **2019**, *18*, 1077–1090. [[CrossRef](#)] [[PubMed](#)]
94. Zivkovic Radojevic, M.; Milosavljevic, N.; Miladinovic, T.B.; Janković, S.; Folic, M. Review of compounds that exhibit radioprotective and/or mitigatory effects after application of diagnostic or therapeutic ionizing radiation. *Int. J. Radiat. Biol.* **2022**, *99*, 594–603. [[CrossRef](#)] [[PubMed](#)]
95. Koukourakis, M.I.; Giatromanolaki, A.; Zois, C.E.; Kalamida, D.; Pouliliou, S.; Karagounis, I.V.; Yeh, T.L.; Abboud, M.I.; Claridge, T.D.; Schofield, C.J.; et al. Normal tissue radioprotection by amifostine via Warburg-type effects. *Sci. Rep.* **2016**, *6*, 30986. [[CrossRef](#)]
96. Segers, C.; Mastroleo, F.; Verslegers, M.; Baatout, S.; Leys, N.; Lebeer, S. Food Supplements to Mitigate Detrimental Effects of Pelvic Radiotherapy. *Microorganisms* **2019**, *7*, 97. [[CrossRef](#)]
97. Xin, J.Y.; Wang, J.; Ding, Q.Q.; Chen, W.; Xu, X.K.; Wei, X.T.; Lv, Y.H.; Wei, Y.P.; Feng, Y.; Zu, X.P. Potential role of gut microbiota and its metabolites in radiation-induced intestinal damage. *Ecotoxicol. Environ. Saf.* **2022**, *248*, 114341. [[CrossRef](#)] [[PubMed](#)]
98. Segers, C.; Mysara, M.; Coolkens, A.; Wouters, S.; Baatout, S.; Leys, N.; Lebeer, S.; Verslegers, M.; Mastroleo, F. *Limnospira indica* PCC 8005 Supplementation Prevents Pelvic Irradiation-Induced Dysbiosis but Not Acute Inflammation in Mice. *Antioxidants* **2023**, *12*, 572. [[CrossRef](#)]
99. Roselli, M.; Finamore, A.; Britti, M.S.; Mengheri, E. Probiotic bacteria *Bifidobacterium animalis* MB5 and *Lactobacillus rhamnosus* GG protect intestinal Caco-2 cells from the inflammation-associated response induced by enterotoxigenic *Escherichia coli* K88. *Br. J. Nutr.* **2006**, *95*, 1177–1184. [[CrossRef](#)]
100. Johnson, D.G.; Walker, C.L. Cyclins and cell cycle checkpoints. *Annu. Rev. Pharmacol. Toxicol.* **1999**, *39*, 295–312. [[CrossRef](#)]
101. Golkhalkhali, B.; Rajandram, R.; Paliany, A.S.; Ho, G.F.; Wan Ishak, W.Z.; Johari, C.S.; Chin, K.F. Strain-specific probiotic (microbial cell preparation) and omega-3 fatty acid in modulating quality of life and inflammatory markers in colorectal cancer patients: A randomized controlled trial. *Asia-Pac. J. Clin. Oncol.* **2018**, *14*, 179–191. [[CrossRef](#)]
102. Amitay, E.L.; Carr, P.R.; Gies, A.; Laetsch, D.C.; Brenner, H. Probiotic/Synbiotic Treatment and Postoperative Complications in Colorectal Cancer Patients: Systematic Review and Meta-analysis of Randomized Controlled Trials. *Clin. Transl. Gastroenterol.* **2020**, *11*, e00268. [[CrossRef](#)]
103. Shuwen, H.; Miao, D.; Quan, Q.; Wei, W.; Zhongshan, Z.; Chun, Z.; Xi, Y. Protective effect of the “food-microorganism-SCFAs” axis on colorectal cancer: From basic research to practical application. *J. Cancer Res. Clin. Oncol.* **2019**, *145*, 2169–2197. [[CrossRef](#)] [[PubMed](#)]
104. Mansouri-Tehrani, H.s.; Rabbani Khorasgani, M.; Roayaei, M. Effects of Probiotics with or without Honey on Radiation-induced Diarrhea. *Internatuinal J. Radiat. Res.* **2016**, *14*, 205–213. [[CrossRef](#)]
105. Arun, K.B.; Madhavan, A.; Reshmitha, T.R.; Thomas, S.; Nisha, P. Short chain fatty acids enriched fermentation metabolites of soluble dietary fibre from *Musa paradisiaca* drives HT29 colon cancer cells to apoptosis. *PLoS ONE* **2019**, *14*, e0216604. [[CrossRef](#)]

106. Nowacka-Jechalke, N.; Nowak, R.; Juda, M.; Malm, A.; Lemieszek, M.; Rzeski, W.; Kaczyński, Z. New biological activity of the polysaccharide fraction from *Cantharellus cibarius* and its structural characterization. *Food Chem.* **2018**, *268*, 355–361. [[CrossRef](#)] [[PubMed](#)]
107. Greek, R.; Menache, A. Systematic Reviews of Animal Models: Methodology versus Epistemology. *Int. J. Med. Sci.* **2013**, *10*, 206. [[CrossRef](#)] [[PubMed](#)]
108. Nagle, P.W.; Coppes, R.P. Current and Future Perspectives of the Use of Organoids in Radiobiology. *Cells* **2020**, *9*, 2649. [[CrossRef](#)] [[PubMed](#)]
109. Moding, E.J.; Lee, C.L.; Castle, K.D.; Oh, P.; Mao, L.; Zha, S.; Min, H.D.; Ma, Y.; Das, S.; Kirsch, D.G. Atm deletion with dual recombinase technology preferentially radiosensitizes tumor endothelium. *J. Clin. Investig.* **2014**, *124*, 3325–3338. [[CrossRef](#)] [[PubMed](#)]
110. Kemp, C.J.; Wheldon, T.; Balmain, A. p53-deficient mice are extremely susceptible to radiation-induced tumorigenesis. *Nat. Genet.* **1994**, *8*, 66–69. [[CrossRef](#)]
111. Segers, C.; Mysara, M.; Coolkens, A.; Baatout, S.; Leys, N.; Lebeer, S.; Verslegers, M.; Mastroleone, F. *Limnospira indica* PCC 8005 or *Lacticaseibacillus rhamnosus* GG Dietary Supplementation Modulate the Gut Microbiome in Mice. *Appl. Microbiol.* **2022**, *2*, 636–650. [[CrossRef](#)]
112. Russell, W.M.S.; Burch, R.L. *The Principles of Humane Experimental Technique*; Methuen Publishing: London, UK, 1959.
113. Petri, Richard, J. *Eine Kleine Modification Des Koch'Schen Plattenverfahrens (English Translation, Braus, 2020)*; Free Download, Borrow, and Streaming: Internet Archive; Internet Archive: San Francisco, CA, USA, 2020.
114. Uribe-Herranz, M.; Rafail, S.; Beghi, S.; Gil-de-Gómez, L.; Verginadis, I.; Bittinger, K.; Pustynnikov, S.; Pierini, S.; Perales-Linares, R.; Blair, I.A.; et al. Gut microbiota modulate dendritic cell antigen presentation and radiotherapy-induced antitumor immune response. *J. Clin. Investig.* **2020**, *130*, 466–479. [[CrossRef](#)]
115. Azizipour, N.; Avazpour, R.; Rosenzweig, D.H.; Sawan, M.; Ajji, A. Evolution of Biochip Technology: A Review from Lab-on-a-Chip to Organ-on-a-Chip. *Micromachines* **2020**, *11*, 599. [[CrossRef](#)]
116. Adrian, G.; Ruan, J.L.; Paillas, S.; Cooper, C.R.; Petersson, K. In vitro assays for investigating the FLASH effect. *Expert Rev. Mol. Med.* **2022**, *24*, e10. [[CrossRef](#)]
117. Mackonis, E.C.; Suchowerska, N.; Naseri, P.; McKenzie, D.R. Optimisation of exposure conditions for in vitro radiobiology experiments. *Australas. Phys. Eng. Sci. Med.* **2012**, *35*, 151–157. [[CrossRef](#)]
118. Liu, Q.; Wang, M.; Kern, A.M.; Khaled, S.; Han, J.; Yeap, B.Y.; Hong, T.S.; Settleman, J.; Benes, C.H.; Held, K.D.; et al. Adapting a drug screening platform to discover associations of molecular targeted radiosensitizers with genomic biomarkers. *Mol. Cancer Res.* **2015**, *13*, 713–720. [[CrossRef](#)] [[PubMed](#)]
119. Nissen, L.; Valerii, M.C.; Spisni, E.; Casciano, F.; Gianotti, A. Multiunit in vitro colon model for the evaluation of prebiotic potential of a fiber plus d-limonene food supplement. *Foods* **2021**, *10*, 2371. [[CrossRef](#)] [[PubMed](#)]
120. Swaters, D.; van Veen, A.; van Meurs, W.; Turner, J.E.; Ritskes-Hoitinga, M. A History of Regulatory Animal Testing: What Can We Learn? *Altern. Lab. Anim.* **2022**, *50*, 322–329. [[CrossRef](#)]
121. Bhatia, S.N.; Ingber, D.E. Microfluidic organs-on-chips. *Nat. Biotechnol.* **2014**, *32*, 760–772. [[CrossRef](#)] [[PubMed](#)]
122. Kim, J.; Koo, B.K.; Knoblich, J.A. Human organoids: Model systems for human biology and medicine. *Nat. Rev. Mol. Cell Biol.* **2020**, *21*, 571–584. [[CrossRef](#)]
123. Van de Wetering, M.; Francies, H.E.; Francis, J.M.; Bounova, G.; Iorio, F.; Pronk, A.; van Houdt, W.; van Gorp, J.; Taylor-Weiner, A.; Kester, L.; et al. Prospective Derivation of a Living Organoid Biobank of Colorectal Cancer Patients. *Cell* **2015**, *161*, 933–945. [[CrossRef](#)] [[PubMed](#)]
124. Rathje, F.; Klingler, S.; Aberger, F. Organoids for Modeling (Colorectal) Cancer in a Dish. *Cancers* **2022**, *14*, 5416. [[CrossRef](#)]
125. Engel, R.M.; Jardé, T.; Oliva, K.; Kerr, G.; Chan, W.H.; Hlavca, S.; Nickless, D.; Archer, S.K.; Yap, R.; Ranchod, P.; et al. Modeling colorectal cancer: A bio-resource of 50 patient-derived organoid lines. *J. Gastroenterol. Hepatol.* **2022**, *37*, 898–907. [[CrossRef](#)]
126. Ooft, S.N.; Weeber, F.; Dijkstra, K.K.; McLean, C.M.; Kaing, S.; van Werkhoven, E.; Schipper, L.; Hoes, L.; Vis, D.J.; van de Haar, J.; et al. Patient-derived organoids can predict response to chemotherapy in metastatic colorectal cancer patients. *Sci. Transl. Med.* **2019**, *11*, eaay2574. [[CrossRef](#)] [[PubMed](#)]
127. Narasimhan, V.; Wright, J.A.; Churchill, M.; Wang, T.; Rosati, R.; Lannagan, T.R.M.; Vrbanac, L.; Richardson, A.B.; Kobayashi, H.; Price, T.; et al. Medium-throughput drug screening of patient-derived organoids from colorectal peritoneal metastases to direct personalized therapy. *Clin. Cancer Res. Off. J. Am. Assoc. Cancer Res.* **2020**, *26*, 3662. [[CrossRef](#)] [[PubMed](#)]
128. Ganesh, K.; Wu, C.; O'Rourke, K.P.; Szeglin, B.C.; Zheng, Y.; Sauv e, C.E.G.; Adileh, M.; Wasserman, I.; Marco, M.R.; Kim, A.S.; et al. A rectal cancer organoid platform to study individual responses to chemoradiation. *Nat. Med.* **2019**, *25*, 1607–1614. [[CrossRef](#)]
129. Yao, Y.; Xu, X.; Yang, L.; Zhu, J.; Wan, J.; Shen, L.; Xia, F.; Fu, G.; Deng, Y.; Pan, M.; et al. Patient-Derived Organoids Predict Chemoradiation Responses of Locally Advanced Rectal Cancer. *Cell Stem. Cell* **2020**, *26*, 17–26.e16. [[CrossRef](#)]
130. Long, A.G.; Lundsmith, E.T.; Hamilton, K.E. Inflammation and Colorectal Cancer. *Curr. Color. Cancer Rep.* **2017**, *13*, 341–351. [[CrossRef](#)] [[PubMed](#)]
131. Wang, Y.; Li, Y.; Sheng, Z.; Deng, W.; Yuan, H.; Wang, S.; Liu, Y. Advances of Patient-Derived Organoids in Personalized Radiotherapy. *Front. Oncol.* **2022**, *12*, 888416. [[CrossRef](#)]
132. Janakiraman, H.; Zhu, Y.; Becker, S.A.; Wang, C.; Cross, A.; Curl, E.; Lewin, D.; Hoffman, B.J.; Warren, G.W.; Hill, E.G.; et al. Modeling rectal cancer to advance neoadjuvant precision therapy. *Int. J. Cancer* **2020**, *147*, 1405–1418. [[CrossRef](#)]

133. Tirado, F.R.; Bhanja, P.; Castro-Nallar, E.; Olea, X.D.; Salamanca, C.; Saha, S. Radiation-induced toxicity in rectal epithelial stem cell contributes to acute radiation injury in rectum. *Stem Cell Res. Ther.* **2021**, *12*, 63. [[CrossRef](#)]
134. Park, M.; Kwon, J.; Shin, H.J.; Moon, S.M.; Kim, S.B.; Shin, U.S.; Han, Y.H.; Kim, Y. Butyrate enhances the efficacy of radiotherapy via FOXO3A in colorectal cancer patient derived organoids. *Int. J. Oncol.* **2020**, *57*, 1307–1318. [[CrossRef](#)]
135. Lau, H.C.H.; Kranenburg, O.; Xiao, H.; Yu, J. Organoid models of gastrointestinal cancers in basic and translational research. *Nat. Rev. Gastroenterol. Hepatol.* **2020**, *17*, 203–222. [[CrossRef](#)]
136. Lacombe, J.; Phillips, S.L.; Zenhausem, F. Microfluidics as a new tool in radiation biology. *Cancer Lett.* **2016**, *371*, 292–300. [[CrossRef](#)]
137. Whitesides, G.M. The origins and the future of microfluidics. *Nature* **2006**, *442*, 368–373. [[CrossRef](#)]
138. Kim, J.; Hegde, M.; Jayaraman, A. Co-culture of epithelial cells and bacteria for investigating host–pathogen interactions. *Lab A Chip* **2010**, *10*, 43–50. [[CrossRef](#)] [[PubMed](#)]
139. Zhang, J.; Hernandez-Gordillo, V.; Trapecar, M.; Wright, C.; Taketani, M.; Schneider, K.; Chen, W.L.K.; Stas, E.; Breault, D.T.; Carrier, R.L.; et al. Co-culture of primary human colon monolayer with human gut bacteria. *Nat. Protoc.* **2021**, *16*, 3874–3900. [[CrossRef](#)] [[PubMed](#)]
140. Wlodkowic, D.; Cooper, J.M. Tumors on chips: Oncology meets microfluidics. *Curr. Opin. Chem. Biol.* **2010**, *14*, 556–567. [[CrossRef](#)] [[PubMed](#)]
141. Mitchell, M.J.; Jain, R.K.; Langer, R. Engineering and physical sciences in oncology: Challenges and opportunities. *Nat. Rev. Cancer* **2017**, *17*, 659–675. [[CrossRef](#)] [[PubMed](#)]
142. Pocock, K.; Delon, L.; Bala, V.; Rao, S.; Priest, C.; Prestidge, C.; Thierry, B. Intestine-on-a-Chip Microfluidic Model for Efficient in Vitro Screening of Oral Chemotherapeutic Uptake. *ACS Biomater. Sci. Eng.* **2017**, *3*, 951–959. [[CrossRef](#)] [[PubMed](#)]
143. Odijk, M.; van der Meer, A.D.; Levner, D.; Kim, H.J.; van der Helm, M.W.; Segerink, L.I.; Frimat, J.-P.; Hamilton, G.A.; Ingber, D.E.; van den Berg, A. Measuring direct current trans-epithelial electrical resistance in organ-on-a-chip microsystems. *Lab A Chip* **2015**, *15*, 745–752. [[CrossRef](#)] [[PubMed](#)]
144. Sato, T.; Stange, D.E.; Ferrante, M.; Vries, R.G.J.; van Es, J.H.; van den Brink, S.; van Houdt, W.J.; Pronk, A.; van Gorp, J.; Siersema, P.D.; et al. Long-term Expansion of Epithelial Organoids From Human Colon, Adenoma, Adenocarcinoma, and Barrett’s Epithelium. *Gastroenterology* **2011**, *141*, 1762–1772. [[CrossRef](#)]
145. Vlachogiannis, G.; Hedayat, S.; Vatsiou, A.; Jamin, Y.; Fernández-Mateos, J.; Khan, K.; Lampis, A.; Eason, K.; Huntingford, I.; Burke, R.; et al. Patient-derived organoids model treatment response of metastatic gastrointestinal cancers. *Science* **2018**, *359*, 920–926. [[CrossRef](#)]
146. Zhang, J.; Huang, Y.J.; Yoon, J.Y.; Kemmitt, J.; Wright, C.; Schneider, K.; Sphabmixay, P.; Hernandez-Gordillo, V.; Holcomb, S.J.; Bhushan, B.; et al. Primary Human Colonic Mucosal Barrier Crosstalk with Super Oxygen-Sensitive Faecalibacterium prausnitzii in Continuous Culture. *Med* **2021**, *2*, 74–98.e79. [[CrossRef](#)]
147. Zhang, J.; Huang, Y.; Trapecar, M.; Wright, C.; Schneider, K.; Kemmit, J.; Hernandez-Gordillo, V.; Griffith, L.; Alm, E.; Trumper, D.; et al. An immune-competent human gut microphysiological system enables inflammation-modulation of Faecalibacterium prausnitzii. *NPJ Biofilms Microbiomes* **2023**, preprint. [[CrossRef](#)]
148. Fofanova, T.; Stewart, C.; Auchtung, J.; Wilson, R.; Britton, R.; Grande-Allen, K.; Estes, M.; Petrosino, J. A novel human enteroid-anaerobe co-culture system to study microbial-host interaction under physiological hypoxia. *BioRxiv* **2019**, 555755. [[CrossRef](#)]
149. Shah, P.; Fritz, J.V.; Glaab, E.; Desai, M.S.; Greenhalgh, K.; Frchet, A.; Niegowska, M.; Estes, M.; Jäger, C.; Seguin-Devaux, C.; et al. A microfluidics-based in vitro model of the gastrointestinal human–microbe interface. *Nat. Commun.* **2016**, *7*, 11535. [[CrossRef](#)] [[PubMed](#)]
150. Marzorati, M.; Vanhoecke, B.; De Ryck, T.; Sadaghian Sadabad, M.; Pinheiro, I.; Possemiers, S.; Van Den Abbeele, P.; Derycke, L.; Bracke, M.; Pieters, J.; et al. The HMI™ module: A new tool to study the Host-Microbiota Interaction in the human gastrointestinal tract in vitro. *BMC Microbiol.* **2014**, *14*, 133. [[CrossRef](#)] [[PubMed](#)]
151. Kim, H.J.; Li, H.; Collins, J.J.; Ingber, D.E. Contributions of microbiome and mechanical deformation to intestinal bacterial overgrowth and inflammation in a human gut-on-a-chip. *Proc. Natl. Acad. Sci. USA* **2016**, *113*, E7–E15. [[CrossRef](#)] [[PubMed](#)]
152. Jalili-Firoozinezhad, S.; Gazzaniga, F.S.; Calamari, E.L.; Camacho, D.M.; Fadel, C.W.; Bein, A.; Swenor, B.; Nestor, B.; Cronce, M.J.; Tovaglieri, A.; et al. A complex human gut microbiome cultured in an anaerobic intestine-on-a-chip. *Nat. Biomed. Eng.* **2019**, *3*, 520–531. [[CrossRef](#)] [[PubMed](#)]
153. Shin, W.; Wu, A.; Massidda, M.W.; Foster, C.; Thomas, N.; Lee, D.W.; Koh, H.; Ju, Y.; Kim, J.; Kim, H.J. A robust longitudinal co-culture of obligate anaerobic gut microbiome with human intestinal epithelium in an anoxic-oxic interface-on-a-chip. *Front. Bioeng. Biotechnol.* **2019**, *7*, 13. [[CrossRef](#)]
154. Ulluwishewa, D.; Anderson, R.C.; Young, W.; McNabb, W.C.; van Baarlen, P.; Moughan, P.J.; Wells, J.M.; Roy, N.C. Live Faecalibacterium prausnitzii in an apical anaerobic model of the intestinal epithelial barrier. *Cell. Microbiol.* **2015**, *17*, 226–240. [[CrossRef](#)]
155. Sadaghian Sadabad, M.; von Martels, J.Z.; Khan, M.T.; Blokzijl, T.; Paglia, G.; Dijkstra, G.; Harmsen, H.J.; Faber, K.N. A simple coculture system shows mutualism between anaerobic faecalibacteria and epithelial Caco-2 cells. *Sci. Rep.* **2015**, *5*, 17906. [[CrossRef](#)]

156. Srinivasan, B.; Kolli, A.R.; Esch, M.B.; Abaci, H.E.; Shuler, M.L.; Hickman, J.J. TEER measurement techniques for in vitro barrier model systems. *J. Lab. Autom.* **2015**, *20*, 107–126. [[CrossRef](#)] [[PubMed](#)]
157. Delon, L.C.; Guo, Z.; Oszmiana, A.; Chien, C.C.; Gibson, R.; Prestidge, C.; Thierry, B. A systematic investigation of the effect of the fluid shear stress on Caco-2 cells towards the optimization of epithelial organ-on-chip models. *Biomaterials* **2019**, *225*, 119521. [[CrossRef](#)] [[PubMed](#)]
158. Toepke, M.W.; Beebe, D.J. PDMS absorption of small molecules and consequences in microfluidic applications. *Lab A Chip* **2006**, *6*, 1484–1486. [[CrossRef](#)] [[PubMed](#)]

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