<u>Plant Gene Register</u>

Nucleotide Sequence of a cDNA Encoding the Ethylene-Forming Enzyme from Petunia Corollas¹

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An increase in the production of ethylene is associated with specific tissues during development and in response to abiotic and biotic stresses (10). In most tissues, the increase in ethylene synthesis is the result of increased ACC² synthase activity, which catalyzes the conversion of AdoMet to ACC. An exception to this, however, is tissues that exhibit an increase in ethylene production associated with senescence, including ripening fruit and senescing flowers. In these tissues, ethylene production is regulated by both ACC synthase and EFE, which oxidizes ACC to ethylene (10). The role of EFE in the regulation of ethylene biosynthesis in these tissues has led to considerable interest in the molecular mechanisms underlying EFE expression. Early progress on the purification and characterization of EFE was slow due to the lack of a suitable cell-free enzyme assay. A significant advancement was made when the expression of an antisense fruit ripeningrelated mRNA in transgenic tomatoes was found to inhibit ethylene production by blocking the conversion of ACC to ethylene (3). This led to the speculation that the ripeningrelated mRNA represented by this cDNA encoded the elusive EFE. It was noted by Hamilton et al. (3) that the deduced protein of pTOM13 shared significant homology with a flavanone-3-hydroxylase from Antirrhinum majus. This led Ververidis and John (8) to assay EFE in vitro as a hydroxylase in the presence of Fe²⁺ and ascorbate. Indeed, these researchers were able to establish authentic EFE activity in soluble plant extracts by this assay. Recently, the identity of pTOM13 as representative of a tomato ripening-related EFE was confirmed by expression of homologous transcripts and recovery of enzyme activity in Xenopus oocytes (7) and yeast (2).

In the flowers of petunia, pollination induces senescence of the corolla through the increased synthesis and action of

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ethylene. We are interested in the regulation of ethylene biosynthesis during flower senescence and report here the nucleotide sequence of a cDNA (pPHEFE) from senescing petunia corollas that represents an EFE mRNA. The sequence of pPHEFE is 1238 bp and contains an open reading frame of 319 amino acids (Fig. 1). pPHEFE shares greater than 77% nucleotide sequence homology with a fruit ripening-related EFE cDNA clone from tomato (4). The predicted protein of pPHEFE (Table I) shares extensive homology with other known EFEs (1, 5, 6, 9) and is greater than 90% identical to the tomato protein, which has been proven to encode EFE (2).

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² Abbreviations: ACC, 1-aminocyclopropane-1-carboxylic acid; EFE, ethylene-forming enzyme.

Figure 1. Nucleotide and deduced amino acid sequence of petunia pPHEFE.

1	GTTATTCACACACTATATACTCTAAAAACACATTCTTCATTCTGCAAGAAAGA	
	ME	2
61	GAACTTCCCAATTATCAGCTTGGACAAAGTGAATGGTGTTGAAAGAGCTGCCACTATGGA	
	N F P I I S L D K V N G V E R A A T M E	22
121	AATGATTAAGGATGCATGTGAAAACTGGGGCTTCTTTGAGTTGGTGAACCATGGAATCCC	
	MIKDACENWGFFELVNHGIP	42
181	ACGTGAAGTAATGGACACTGTGGAGAAGATGACAAAGGGTCATTACAAGAAGTGCATGGA	
	R E V M D T V E K M T K G H Y K K C M E	62
241	ACAAAGGTTCAAGGAATTGGTTGCCAGTAAAGCTCTTGAAGGTGTTCAAGCTGAGGTTAC	
	Q R F K E L V A S K A L E G V Q A E V T	82
301	TGATATGGATTGGGAAAGCACCTTTTTCTTGAAACATCTCCCCATTTCTAACATTTCTGA	
	D M D W E S T F F L K H L P I S N I S E	102
361	AGTCCCTGATCTTGATGAAGAATACAGGGAAGTTATGAGAGATTTTGCTAAAAGGTTAGA	
	V P D L D E E Y R E V M R D F A K R L E	122
421	GAAGCTGGCAGAGGAGCTTTTGGACTTGCTGTGTGAAAAATCTTGGCCTGGAAAAAGGTTA	
	K L A E E L L D L L C E N L G L E K G Y	142
481	CCTTAAAAATGCCTTTTATGGATCGAAAGGTCCAAACTTTGGGACTAAAGTGAGCAACTA	
	L K N A F Y G S K G P N F G T K V S N Y	162
541	TCCACCATGCCCAAAACCAGATTTAATCAAAGGACTACGTGCCCACACAGACGCTGGTGG	
	P P C P K P D L I K G L R A H T D A G G	182
601	AATAATCCTTCTGTTCCAAGATGATAAAGTAAGTGGCCTACAACTCCTCAAAGACGGCCA	
	IILLFODDKVSGLOLLKDGO	202
661	ATGGATCGATGTTCCTCCCATGCGCCATTCCATTGTGGTTAACCTCGGTGACCAACTCGA	
	WIDVPPMBHSTVVNLGDOLE	222
721	GETGATCACTAATGGAAAATACAAGAGTGTGATGCACAGAGTGATAGCCCAAAAAGACGG	
	V T T N G K Y K S V M H B V T A O K D G	242
781	GGCTCGAATGTCATTAGCTTCCTTTTACAATCCAGGAAGTGATGCAGTGATCTATCCAGC	
	A R M S L A S F Y N P G S D A V T Y P A	262
841	ACCAGCTCTTGTTGAGAAAGAAGAAGCAGAGAAAACAAAC	202
•	PALVEKEAEENKOVYPKEVE	282
901	TGATGATTACATGAAGTTATATGCTGGACTCAAGTTTCAAGGCTAAGGAGCCAAGATTTGA	202
	D D Y M K L Y A G L K F O A K E P R F F	302
961	TABATAATOTTOTOTOTOTAADOTABOTAADTAATOTAATO	002
		319
1021	GGAAGTTGGATAAAAATGATTTGTTATTAAAAAGTAGTACAAGACCTGTGACATATATTAT	519
1081	ATGTTTTCTTTAGTATAGTGTGATGATCAATCTATTTACAAGAGGGGGGTGTGTCCCCTACTATATA	
1141	ͲϹͲϪϹͲͲͲϹͲϹϹϹͲϪϹͳϪϹϿϿϿͲϹͲϪϹϔͲϪϪϹͲϹϽϹϹϹϹͲϪϪϪϹͲͲͲϹͲϪͲϪϮͲϪϿϪ	
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1201	CTGATAAAGCTTCTTTTTACACAAATATTAATATACCC	

Table 1. Characteristics of an EFE cDNA (pPHEFE) from Petunia

Organism Petunia hybrida L. 'Pink Flash'. Function: Encodes ACC oxidase (EFE), which catalyzes the conversion of ACC to ethylene. Clone Type; Designation: cDNA, full-length; pPHEFE Source: cDNA library in λ ZAP vector constructed using poly(A)⁺RNA isolated from senescing petunia corollas 4 to 5 d after pollination. Method of Identification: Library was screened with the cDNA pSR120 (9), which represents EFE mRNA from senescing carnation flower petals. Sequencing Strategy: Plasmid sequencing; restriction fragment and exonuclease III deletion subclones used to completely sequence both strands. **Expression Characteristics:** Transcript of approximately 1300 nucleotides; increase in mRNA abundance in senescing corolla tissue and pollinated pistil concomitant with increased ethylene production; increase in mRNA detected in corollas following ethylene exposure. Features of cDNA Structure: Nucleotide sequence is 77% identical to the tomato EFE cDNA pTOM13 (4). Structural Features of Deduced Protein: 319 amino acids (Mr 36,186); pl of 5.17; 92%, 78%, 76%, and 75% identical to predicted EFE proteins from tomato (4), apple (1), avocado (5, 6), and carnation (9), respectively. Antibodies: None available. Subcellular Location: Not determined. GenBank Accession No.: M90294