

Deacetylation of Histones and Non-histone Proteins in Inflammatory Diseases and Cancer Therapeutic Potential of Histone Deacetylase Inhibitors



Ezgi Man^{1,2} and Serap Evran^{1,*}

¹Department of Biochemistry, Faculty of Science, Ege University, 35100, İzmir, Türkiye; ²EGE SCIENCE PRO Scientific Research Inc., Ege University, IdeEGE Technology Development Zone, 35100, Bornova-Izmir, Türkiye

ARTICLE HISTORY

Received: May 31, 2023 Revised: July 18, 2023 Accepted: August 26, 2023

DOI: 10.2174/0113892029265046231011100327



Abstract: Epigenetic changes play an important role in the pathophysiology of autoimmune diseases such as allergic asthma, multiple sclerosis, lung diseases, diabetes, cystic fibrosis, atherosclerosis, rheumatoid arthritis, and COVID-19. There are three main classes of epigenetic alterations: post-translational modifications of histone proteins, control by non-coding RNA and DNA methylation. Since histone modifications can directly affect chromatin structure and accessibility, they can regulate gene expression levels. Abnormal expression and activity of histone deacetylases (HDACs) have been reported in immune mediated diseases. Increased acetylated levels of lysine residues have been suggested to be related to the overexpression of inflammatory genes. This review focuses on the effect of HDAC modifications on histone and non–histone proteins in autoimmune diseases. Furthermore, we discuss the potential therapeutic effect of HDAC inhibitors (HDACi) used in these diseases.

Keywords: Autoimmune diseases, epigenetics, histone protein modifications, non-histone protein modifications, HDACs, HDAC inhibitors, lysine deacetylation, histone deacetylase.

1. INTRODUCTION

Post-translational modifications (PTMs) include methylation, glycosylation, ubiquitination, acetylation, phosphorylation and nitrosylation [1]. PTMs alter protein functions by regulating their stability and activity [2]. Epigenetic modifications involve many different cellular signalling pathways and contribute to various human diseases' pathogenesis [3]. Reversible lysine acetylation of histones is one of the important mechanisms for controlling gene expression, inflammation, cell development and differentiation [4]. Lysine acetylation levels are mediated by two enzyme groups: histone deacetylases (HDACs) and histone acetyltransferases (HATs) (Fig. 1) [5]. Furthermore, HDACs and HATs also target many non-histone substrates. This suggests that lysine-acetylation is also critical in the cell proteome and protein function beyond chromatin accessibility mediated gene regulation [6]. Accumulating evidence shows that abnormal activities of HDACs and HATs activities play a crucial role in inflammatory diseases [7]. HDAC catalysis removes acetyl groups from lysine residues on histone protein tails. Mammalian HDACs family consists of 18 members, and they are divided into Class I (HDAC1, HDAC2, HDAC3 and HDAC8), Class IIa (HDAC4, HDAC5, HDAC7 and HDAC9), Class IIb (HDAC6 and HDAC10), Class III (SIRT1-7) and Class IV (HDAC11) (Fig. 2) [8]. Classes I, II

and IV have highly conserved domains and their deacetylation activity is Zn²⁺ dependent, while Class III enzyme subgroup deacetylase activity is NAD⁺ dependent [9]. HDACi can be used to inhibit HDAC activity in diseases such as cancer, immune diseases, neurodegenerative diseases, diabetes and cystic fibrosis [10]. HDAC inhibitors can be categorized according to their synthetic or natural composition, subclass-class specificity, and chemical types of structures. Generally, they are divided into two classes: HDAC-pan inhibitors and HDAC-specific inhibitors [11]. HDAC inhibitors are grouped into four main subgroups based on their chemical composition: hydroxamates, benzamides, cyclic tetrapeptides and short-chain fatty acids. HDAC inhibitors are considered novel epigenetic drugs, and their therapeutic potential is widely tested in various disease models [12].

2. CYSTIC FIBROSIS

Cystic fibrosis is a genetic disorder caused by mutations in the cystic fibrosis transmembrane conductance regulator (CFTR) gene [13]. CTFR protein is responsible for the transport of chloride and bicarbonate ions, and mutations that impair its function favor lung infection by opportunistic pathogens, *Pseudomonas aeruginosa* being the major one [14]. Excessive inflammatory response to *P. aeruginosa* plays a critical role in lung damage [15-17]. Hence, antiinflammatory agents are considered as a treatment option [18, 19]. HDAC inhibitors are among those agents with promising results for reducing the inflammation in cystic fibrosis. Suberoylanilide hydroxamic acid (SAHA), a pan-HDAC inhibitor, was shown to modulate the inflammation

^{*}Address correspondence to this author at the Department of Biochemistry, Faculty of Science, Ege University, 35100, İzmir, Türkiye; Tel: +90 232 311 23 04; E-mail: serap.evran@ege.edu.tr

Deacetylation of Histones and Non-histone Proteins in Inflammatory Diseases

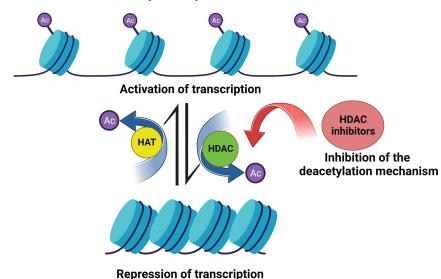


Fig. (1). Histone acetylation and deacetylation regulation mechanism. Histone acetyl transferases (HATs) catalyze the transfer of acetyl groups, histone deacetylases (HDACs) remove the acetyl groups from the lysine residues. (Created with BioRender.com). (*A higher resolution / colour version of this figure is available in the electronic copy of the article*).

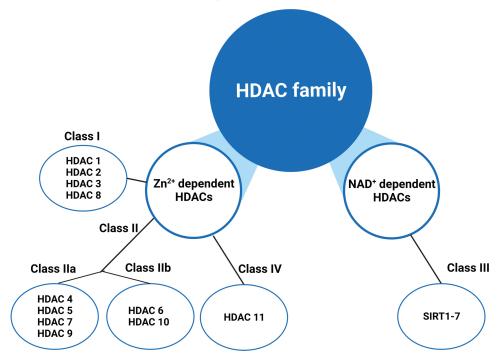


Fig. (2). Classification of histone deacetylase (HDAC) family (Created with BioRender.com). (A higher resolution / colour version of this figure is available in the electronic copy of the article).

caused by *P. aeruginosa* lipopolysaccharide [20]. In this study, treatment of *Cftr+/+* or *Cftr-/-* mice with SAHA resulted in changes in interleukin-6 (IL-6) levels, nuclear factor kappa B (NF κ B)-mediated signaling and neutrophil chemotaxis/activation. In another study [21], the efficiency of HDAC6 inhibitor, which was identified by screening patents and research papers was evaluated in the mouse model of *P. aeruginosa* acute and chronic respiratory infection. It was shown that inhibition of HDAC activity resulted in the reduction of several inflammatory interleukins, chemokines, growth factors, and interferon gamma (IFN- γ). Another interesting study revealed how the quorum sensing molecule 2-aminoacetophenone secreted by *P. aeruginosa* could af-

fect HDAC1-dependent chromatin modification [22]. In this study, 2-aminoacetophenone was found to be responsible for HDAC1-mediated deacetylation of histone 3 at lysine 18 (H3K18) at the promoter sites of the autophagy gene *Beclin*1 and the lipid biosynthesis gene *Scd*1, resulting in decreased expression of these genes. It was concluded that *P. aeruginosa* was able to reduce the macrophage activity by modulating membrane lipids and autophagy. In addition to their anti-inflammatory effects, HDAC inhibitors were also found to have repairing activity on loss-of-function CTFR mutants. The pan-HDAC inhibitor SAHA was shown to restore the surface channel activity of CTFR phenylalanine 508 deletion (F508 del) variant [23]. In the same study, si-

lencing of both HDAC1 and HDAC7 was found to enhance the stability of CTFR mutant. In particular, silencing of HDAC7 was found to be more effective in terms of restoring channel activity. The authors proposed that HDAC inhibitors could exert their effect through altering the transcriptional level of CFTR-related genes or by altering posttranslational acetylation levels of their non-histone substrates. In a similar study [24], HDAC inhibitors were shown to restore the transport function of the F508del variant of CTFR. In addition, a similar effect on the other CTFR variants were observed, albeit with different degrees of response to HDAC inhibitors. The authors concluded that HDAC inhibitors rescued CTRF trafficking through downregulation of HDAC7 and abrogation of the maladaptive stress response (MSR). In another study, depletion of HDAC6 in the cystic fibrosis mouse model was found to restore the aggressive inflammatory response back to to wild-type profiles [25].

3. RHEUMATOID ARTHRITIS

Rheumatoid arthritis is a chronic autoimmune disease with both environmental and genetic basis [26]. Cytokines [27] and chemokines [28] are mainly responsible for inflammation, which results in joint damage. Previous studies showed [29] that the inflammation mechanism involves multiple signal transduction pathways regulated through epigenetic mechanisms. For instance, HDAC3 was shown to be involved in type-I interferon (IFN) production and regulation of inflammatory gene expression [30]. Another study [31] showed that the overexpressed HDAC2 in rheumatoid arthritis synovial tissue regulated the signalling pathway of the inflammatory mediator chemokine CC motif ligand 7 (CCL7). Similarly, HDAC6 was shown to be overexpressed in the synovium tissues of the mouse model and activated the nuclear factor-kappaB (NF-kappaB) signalling pathway by deacetylating its non-histone protein substrate myeloid differentiation primary response 88 (MyD88) [32]. Similarly, another study revealed that non-histone proteins were among the substrates of HDAC6 [33]. It was shown that inhibition of HDAC6 resulted in hyperacetylation of cytoskeletal proteins tubulin and cortactin. In addition, it decreased the production of interleukin-6 (IL-6) and the matrix metalloproteinases MMP1 and MMP3, thereby suppressing inflammation. Those studies proposed HDAC inhibitors as anti-inflammatory agents for treating rheumatoid arthritis [34]. For instance, the selective HDAC6 inhibitor CKD-506 was shown to prevent experimental arthritis in a murine model [35]. M-134, another HDAC6-selective inhibitor, was shown to reduce the level of chemokine (C-X-C motif) ligand 10 (IP-10), interleukin-17A (IL-17A), and tumour necrosis factor-alpha (TNF- α) expression. Moreover, a combination of M-134 and the drug tofacitinib enhanced the expression of different cytokines, adhesion factors and chemokines involved in immune cell migration and chemoattraction [36]. The selective HDAC1-inhibitor TTA03-107 was shown to suppress the production of inflammatory cytokines and reduce the severity of autoimmune arthritis [37]. As reviewed elsewhere [38], a combination of HDAC, inosine monophosphate dehydrogenase (IMPDH), mammalian target of rapamycin (mTOR), and Janus kinase (JAK) inhibitors could be promising to reduce the inflammation caused by increased cytokine levels. However, it should not be overlooked that inhibition of some HDAC isoenzymes may not yield the desired anti-inflammatory effects [39]. For instance, the inflammatory stimuli were shown to suppress HDAC5 expression. Moreover, the silencing of HDAC5 increased the levels of different chemokines and cytokines [40].

4. ATHEROSCLEROSIS

Atherosclerosis is a chronic inflammatory disease characterized by plaque formation in the walls of arteries and leading to cardiovascular disease and stroke [41]. Several studies revealed the connection between inflammation and atherosclerosis [42-45]. The impact of epigenetic mechanisms, such as DNA methylation, histone methylation, and acetylation of histone and non-histone proteins, on the polarization of macrophages was revealed [46]. HDAC isoenzymes are involved in endothelial dysfunction through different mechanisms [47]. In particular, inflammatory factors are activated by HDACs, via histone acetylation [48, 49]. A remarkable issue is that not all HDAC isoenzymes show disease-inducing effects [50]. For instance, HDAC7, associated with myocyte enhancer factor-2 (MEF2), was shown to protect endothelial integrity by downregulating matrix metalloproteinase MMP10 gene transcription [51]. In contrast, other isoenzymes such as HDAC3 [52], HDAC6 [53] and HDAC9 [54] were shown to contribute to the development of atherosclerosis. Hence, HDAC inhibitors were proven to be effective in reducing inflammation [55].

5. COVID-19

Excessive inflammatory response and cytokine storm play critical roles in pathogenesis and severity of the coronavirus disease 2019 (COVID-19) [56-58]. Hence, the effectiveness of anti-inflammatory drugs was intensively investigated [59-62]. Considering the inflammatory roles and the link between COVID-19 and epigenetic mechanisms [63-66], HDACs and HDAC inhibitors were also under investigation. For instance, several HDAC6 inhibitors were tested on the in vitro models of immune and epithelial cells by mimicking the cellular status after viral infection [67]. The authors showed that pro-inflammatory cytokines and interferon pathway genes were downregulated. In addition, the HDAC6 selective inhibitor ITF3756 was capable of upregulating the genes responsible for T-cell memory phenotypes. A screening study of the clinically approved HDAC inhibitors showed that romidepsin, panobinostat, givinostat hydrochloride monohydrate, CAY10603, and sirtinol were able to inhibit the cellular entry of COVID-19 [68]. A further study showed that the HDAC inhibitor panobinostat suppressed the expression of angiotensin-converting enzyme 2 (ACE2) receptor in the gastric adenocarcinoma cell line [69]. In another study, a similar result was obtained and valproic acid was shown to reduce the expression of angiotensin-converting enzyme 2 (ACE2) and neuropilin-1 (NRP1) receptors [70]. HDAC inhibitors were also proposed as neuroprotective agents against COVID-19 infection, mainly through downregulation of proinflammatory cytokines [71].

6. ASTHMA

Asthma is a chronic disease defined by airway inflammation, hyperresponsiveness, increased mucus secretion and remodelling [72]. Asthmatic inflammation is classified into 4 different groups; paucigranulacytic, eosinophilic, neutrophilic and mixed granulocytic. Airway inflammation has similar symptoms, like breathlessness, cough, wheezing, chest tightness, and dyspnoea [73]. Numerous asthma cases have reported increased interleukin-4 (IL-4) and interleukin-5 (IL-5 levels, eosinophils-mediated infiltration and activated mast cells. Glucocorticoids are used as the main therapy agent for asthma [74]. However, they cause undesired side effects. In addition, mixed granulocytic type of airway inflammation is unresponsive to standard/high-dose glucocorticoid treatment [75]. Therefore, alternative treatment approaches are needed [76]. HDAC2 enzyme activity alters chromatin structure and regulates inflammatory, antiinflammatory gene expression in airway inflammation [77]. Various human asthma and murine models have been reported to decrease HDAC2 expression and specific enzyme activity [78]. Decreased HDAC2 expression level is associated with activation of NFkB signalling. Bruton's tyrosine kinase (BTK) is expressed in both innate and adaptive immune cells such as neutrophils, B cells and macrophages. An earlier study showed that inhibition of BTK by Inrutinib was effective in mouse models of eosinophilic and neutrophilic airway inflammation [79]. Another study combined dexamethasone (corticosteroid) therapy and BTK inhibitor ibrutinib to test their therapeutic effects in cockroach allergen extract (CE)-induced mixed granulocytic inflammation mice model. Corticosteroids were found to downregulate the pulmonary inflammation-related gene expressions, such as tumour necrosis factor-alpha (TNF- α), interleukin-8 (IL-8), granulocyte-macrophage colony-stimulating factor (GM-CSF), inducible nitric oxide synthase (iNOS), interleukin- $1\beta(IL-1\beta)$ and monocyte chemoattractant protein 1 (MCP1). Decreased HDAC2 expression was related to increased inflammatory cytokines [80]. In this study, BTK inhibition by Ibrutinib reestablished HDAC2 expression level and reduced inflammatory cytokines and NFkB expression. This study suggested that regulation of HDAC2 expression level by BTK inhibition might be an alternative approach to obtaining sensitivity to corticosteroids in granulocytic asthma [81]. Numerous human health investigations have studied curcumin's (CUR) biological potential. The studies showed that curcumin has anti-oxidant/inflammatory and antiallergic properties, and it functions as a natural HDAC-pan inhibitor [82]. Butyrate is an HDAC inhibitor, and it suppresses IL-8 expression. In another study, sodium butyrate (SoB) and CUR were used for modulating structural changes in the mouse model of asthma. HDAC1 and HDAC3 were extensively related to allergic-induced asthma [83]. The research authors concluded that SoB and CURmediated inhibition could effectively restore structural changes in airways, also suppress HDAC1 and NF-kB. In conclusion, the therapeutic properties of HDACi have offered alternative treatments for different human diseases. There is a need for new research for a better understanding of inhibitor/pathway relations [84].

7. NEUROINFLAMMATION

Recent findings indicate that neuroinflammation plays a crucial role in a range of neurological conditions, encompassing central nervous system (CNS) traumas, depression, and neurodegenerative illnesses such as Alzheimer's and Parkinson's diseases [85]. Neurological disorders' severity can be mitigated by inhibiting neuroinflammation [86]. HDACs play a pivotal role in modulating immune responses and inflammatory processes. HDACi have emerged as a prominent area of interest in investigating anti-inflammatory pharmaceuticals [87]. Earlier studies showed that in situations of brain injury and neurodegenerative disease, it is typical to observe an overexpression of HDAC1 and HDAC2 in microglia. The phenomenon induces polarization of microglia towards M1 macrophage. It results in the release of a considerable quantity of inflammatory mediators, which may ensue from the deacetylation of signal transducer and activator of transcription (STAT1/3), thereby intensifying the activation of the NF- κ B signalling cascade [88]. The activation of the NF- κ B signalling pathway ultimately results in the activation of microglia, thereby intensifying neuroinflammation and increasing neuronal damage. The same study has demonstrated that the application of HDAC inhibitor SAHA can impede the M1-polarization of microglia, reduce neuroinflammation dependent on HDAC1/2, and protect neuronal cells [89]. In another study, paninhibitory valproic acid (VPA) has been used to regulate STAT1/NF-kB and JAK2 (Janus Kinase 3)/STAT3 signal pathways to control microglial function and suppress spinal neuroinflammation in neuropathic pain [90]. HDACII inhibitory Tubastatin A has been used in cerebral ischemia; it has increased regulatory T cell (Treg) immunosuppressive ability and regulated interleukin-10 (IL-10) expression levels [87]. Histone deacetylases (HDACs) regulate gene expression by deacetylating histories and related proteins [86]. Additionally, HDACs have been found to directly deacetylate molecules involved in inflammatory signalling pathways, regulate the activation of glial cells in the central nervous system, and promote neuronal survival [91]. Further clarification is needed regarding the precise mechanisms HDAC regulates neuroinflammation [92].

8. CANCER

HDACs play several roles in cancer cell metabolism and they regulate cell cycle, apoptosis, DNA-damage response, metastasis, angiogenesis, autophagy [93-95]. Hence, dysregulation of HDACs results in cancer initiation and progression [96]. As reviewed elsewhere [97], HDAC8 is overexpressed in different types of cancers and the level of overexpression is correlated with the advanced stage of breast cancer and neuroblastoma. Similarly, HDAC2, HDAC3 and HDAC6 are overexpressed in lung cancer [97]. human cholangiocarcinoma [98], and colon cancer [99], respectively. In addition, the overexpressed class I HDACs have been shown to promote drug resistance in glioma cells [100]. Because of the crucial roles of HDACs in cancer, HDAC inhibitors (Table 1) have been proposed as anticancer agents [101-105]. For instance, the class I HDAC inhibitor valproic acid has been shown to enhance the effectiveness of chemotherapy agents in human melanoma cells [106]. Similarly, the class I HDAC inhibitor domatinostat

has been shown to sensitize pancreatic cancer cells to chemotherapy by exerting its effect on the transcription factor FOXM1 [107]. Likewise, the potential of HDAC inhibitors to overcome immunotherapy resistance has been revealed [108]. There is also a growing interest in dual HDAC inhibitors targeting both HDAC and another cancer target, such as phosphoinositide 3-kinases [109], microtubule polymerization [110], bromodomain and extra-terminal [111]. As reviewed elsewhere [112], HDAC-based dual drugs have been proposed to be superior to single-targeted drugs in terms of therapeutic efficiency. Despite the great potential of HDAC inhibitors, it should not be overlooked that not all HDAC isozenzymes are related to cancer progression. A remarkable study has shown that pan-HDAC inhibitor promotes breast cancer metastasis due to the inhibition of HDAC4 [113]. Another study has revealed the tumor suppressive role of HDAC10 in cervical cancer [114].

Table 1. HDACi classification.

HDAC Inhibitors	Types of HDAC Inhibitors	
MS-275 (Entinostat)	Benzamide	
Apicidin Depsipeptide Trapoxin A	Cyclic peptide	
Valproic acid Butyrate	Short chain fatty acid	
Tubacin Belinostat Vorinostat (SAHA)	Hydroxamate	

CONCLUSION

This mini-review summarized an overview of the latest literature on utilising HDAC inhibitors as pharmacological agents for the modulation of autoimmunity and inflammation. The summary of this review and outcomes from numerous investigations on autoimmune and autoinflammatory disorders clearly suggest that HDAC inhibitors have significant therapeutic potential in controlling the symptoms of immune-mediated diseases. Developing isoform-specific HDAC inhibitors is essential for effectively treating autoimmune disorders while overcoming adverse effects. In conclusion, a better understanding of the molecular consequences of HDAC inhibition is required to develop alternative treatment strategies for autoimmune diseases. Corepressor complexes consist of a variety of proteins that play a role in the repression of transcription. These proteins include DNA-binding proteins, histone deacetylases (HDACs), and components involved in the structural organization of chromatin. The role of corepressor function is crucial in controlling an extensive range of biological processes, including development, differentiation, and signal transduction. HDAC1, HDAC2, and HDAC3 generally function as a corepressor complex in transcriptional regulation. HDACs acting on both histone and non-histone proteins are attractive drug targets in a wide range of diseases. Hence, there is much interest in the discovery of HDAC inhibitors. However, the major limitation is that all of the FDA-approved drugs are pan-inibitors with no HDAC isoenzyme selectivity. Considering that each HDAC isoenzyme may have counter effects on the disease mechanism of interest, the design of isoenzyme-specific inhibitors is critical to prevent off-target effects and toxicity. Another issue is that HDACs are not only effective on histone proteins but also non-histone proteins. Although the number of studies on distinct biological functions of HDACs increases by year, there are still unknowns about the non-histone substrates, as well as the interaction partners of HDAC isoezymes. As more structural and mechanistic information is gathered, the therapeutic potential of HDACs is expected to be increased in the future. The interest in combination therapy approaches, as well as in dual-inhbitor design is encouraging efforts for the field of HDAC inhibitor research.

LIST OF ABBREVIATIONS

ACE2	=	Angiotensin-converting Enzyme 2
BTK	=	Bruton's Tyrosine Kinase
CFTR	=	Cystic Fibrosis Transmembrane Conduct- ance Regulator
CNS	=	Central Nervous System
COVID-19	=	Coronavirus Disease 2019
CUR	=	Curcumin's
GM-CSF	=	Granulocyte-macrophage Colony-stimula- ting factor
HATs	=	Histone Acetyltransferases
HDACi	=	HDAC Inhibitors
HDACs	=	Histone Deacetylases
IFN-γ	=	Interferon gamma
IL-6	=	Interleukin-6
IMPDH	=	Inosine Monophosphate Dehydrogenase
iNOS	=	inducible Nitric Oxide Synthase
JAK	=	Janus kinase
MCP1	=	Monocyte Chemoattractant Protein 1
MEF2	=	Myocyte Enhancer Factor-2
MSR	=	Maladaptive Stress Response
mTOR	=	mammalian Target of Rapamycin
ΝΓκΒ	=	Nuclear Factor Kappa B
NRP1	=	Neuropilin-1
PTMs	=	Post-translational Modifications
SAHA	=	Suberoylanilide Hydroxamic Acid
SoB	=	Sodium Butyrate
TNF-α	=	Tumour Necrosis Factor-alpha
VPA	=	Valproic Acid

CONSENT FOR PUBLICATION

Not applicable.

FUNDING

None.

CONFLICT OF INTEREST

The authors declare no conflict of interest, financial or otherwise.

ACKNOWLEDGEMENTS

Declared none.

REFERENCES

- Lee, J.M.; Hammarén, H.M.; Savitski, M.M.; Baek, S.H. Control [1] of protein stability by post-translational modifications. Nat. Commun., 2023, 14(1), 201. http://dx.doi.org/10.1038/s41467-023-35795-8 PMID: 36639369
- [2] Hermann, J.; Schurgers, L.; Jankowski, V. Identification and characterization of post-translational modifications: Clinical implications. Mol. Aspects Med., 2022, 86, 101066. http://dx.doi.org/10.1016/j.mam.2022.101066 PMID: 35033366
- Ao, C.; Gao, L.; Yu, L. Research progress in predicting DNA [3] methylation modifications and the relation with human diseases. Curr. Med. Chem., 2022, 29(5), 822-836. http://dx.doi.org/10.2174/0929867328666210917115733 PMID: 34533438
- Li, P.; Ge, J.; Li, H. Lysine acetyltransferases and lysine deacety-[4] lases as targets for cardiovascular disease. Nat. Rev. Cardiol., 2020, 17(2), 96-115.

http://dx.doi.org/10.1038/s41569-019-0235-9 PMID: 31350538 Yang, X.J.; Seto, E. Lysine acetylation: codified crosstalk with [5]

other posttranslational modifications. Mol. Cell, 2008, 31(4), 449-461.

http://dx.doi.org/10.1016/j.molcel.2008.07.002 PMID: 18722172

- [6] Wang, Z.A.; Cole, P.A. The chemical biology of reversible lysine post-translational modifications. Cell Chem. Biol., 2020, 27(8), 953-969. http://dx.doi.org/10.1016/j.chembiol.2020.07.002 PMID:
- 32698016 Li, Y.; Zhou, M.; Lv, X.; Song, L.; Zhang, D.; He, Y.; Wang, M.; [7] Zhao, X.; Yuan, X.; Shi, G.; Wang, D. Reduced activity of HDAC3 and increased acetylation of histones H3 in peripheral blood mononuclear cells of patients with rheumatoid arthritis. J. Immunol. Res., 2018, 2018, 1-10. http://dx.doi.org/10.1155/2018/7313515 PMID: 30402512
 - Witt, O.; Deubzer, H.E.; Milde, T.; Oehme, I. HDAC family:
- [8] What are the cancer relevant targets? Cancer Lett., 2009, 277(1), 8-21.
- http://dx.doi.org/10.1016/j.canlet.2008.08.016 PMID: 18824292 [9] Ganai, S.A. Characterizing binding intensity and energetic features of histone deacetylase inhibitor pracinostat towards class I HDAC isozymes through futuristic drug designing strategy. In Silico Pharmacol., 2021, 9(1), 18.

http://dx.doi.org/10.1007/s40203-021-00077-y PMID: 33628709

- [10] Bondarev, A.D.; Attwood, M.M.; Jonsson, J.; Chubarev, V.N.; Tarasov, V.V.; Schiöth, H.B. Recent developments of HDAC inhibitors: Emerging indications and novel molecules. Br. J. Clin. Pharmacol., 2021, 87(12), 4577-4597. http://dx.doi.org/10.1111/bcp.14889 PMID: 33971031
- [11] Ho, T.C.S.; Chan, A.H.Y.; Ganesan, A. Thirty years of HDAC inhibitors: 2020 insight and hindsight. J. Med. Chem., 2020, 63(21), 12460-12484. http://dx.doi.org/10.1021/acs.jmedchem.0c00830 PMID: 32608981
- Gatla, H.; Muniraj, N.; Thevkar, P.; Yavvari, S.; Sukhavasi, S.; [12] Makena, M. Regulation of chemokines and cytokines by histone

deacetylases and an update on histone decetylase inhibitors in human diseases. Int. J. Mol. Sci., 2019, 20(5), 1110. http://dx.doi.org/10.3390/ijms20051110 PMID: 30841513

- [13] Ribeiro, C.M.P.; Higgs, M.G.; Muhlebach, M.S.; Wolfgang, M.C.; Borgatti, M.; Lampronti, I.; Cabrini, G. Revisiting Host-Pathogen Interactions in Cystic Fibrosis Lungs in the Era of CFTR Modulators. Int. J. Mol. Sci., 2023, 24(5), 5010. http://dx.doi.org/10.3390/ijms24055010 PMID: 36902441
 - Malhotra, S.; Hayes, D., Jr; Wozniak, D.J. Cystic fibrosis and
- [14] Pseudomonas aeruginosa: the host-microbe interface. Clin. Microbiol. Rev., 2019, 32(3), e00138-18. http://dx.doi.org/10.1128/CMR.00138-18 PMID: 31142499
- [15] Phuong, M.S.; Hernandez, R.E.; Wolter, D.J.; Hoffman, L.R.; Sad, S. Impairment in inflammasome signaling by the chronic Pseudomonas aeruginosa isolates from cystic fibrosis patients results in an increase in inflammatory response. Cell Death Dis., 2021, 12(3), 241.

http://dx.doi.org/10.1038/s41419-021-03526-w PMID: 33664232

- [16] Mateu-Borrás, M.; González-Alsina, A.; Doménech-Sánchez, A.; Querol-García, J.; Fernández, F.J.; Vega, M.C.; Albertí, S. Pseudomonas aeruginosa adaptation in cystic fibrosis patients increases C5a levels and promotes neutrophil recruitment. Virulence, 2022, 13(1), 215-224. http://dx.doi.org/10.1080/21505594.2022.2028484 PMID: 35094639
- [17] Petrocheilou, A.; Moudaki, A.; Kaditis, A.G. Inflammation and Infection in Cystic Fibrosis: Update for the Clinician. Children (Basel), 2022, 9(12), 1898. http://dx.doi.org/10.3390/children9121898 PMID: 36553341
- Chmiel, J.F.; Konstan, M.W.; Elborn, J.S. Antibiotic and anti-[18] inflammatory therapies for cystic fibrosis. Cold Spring Harb. Perspect. Med., 2013, 3(10), a009779.
- http://dx.doi.org/10.1101/cshperspect.a009779 PMID: 23880054
- [19] Ribeiro, C.M.P.; McElvaney, N.G.; Cabrini, G. Editorial: Novel anti-inflammatory approaches for cystic fibrosis lung disease: Identification of molecular targets and design of innovative therapies. Front. Pharmacol., 2021, 12, 794854. http://dx.doi.org/10.3389/fphar.2021.794854 PMID: 34867428

[20] Bodas, M.; Mazur, S.; Min, T.; Vij, N. Inhibition of histone-

deacetylase activity rescues inflammatory cystic fibrosis lung disease by modulating innate and adaptive immune responses. Respir. Res., 2018, 19(1), 2.

http://dx.doi.org/10.1186/s12931-017-0705-8 PMID: 29301535

- [21] Brindisi, M.; Barone, S.; Rossi, A.; Cassese, E.; Del Gaudio, N.; Feliz Morel, Á.J.; Filocamo, G.; Alberico, A.; De Fino, I.; Gugliandolo, D.; Babaei, M.; Bove, G.; Croce, M.; Montesano, C.; Altucci, L.; Bragonzi, A.; Summa, V. Efficacy of selective histone deacetylase 6 inhibition in mouse models of Pseudomonas aeruginosa infection: A new glimpse for reducing inflammation and infection in cystic fibrosis. Eur. J. Pharmacol., 2022, 936, 175349. http://dx.doi.org/10.1016/j.ejphar.2022.175349 PMID: 36309047
- Chakraborty, A.; Kabashi, A.; Wilk, S.; Rahme, L.G. Quorum-[22] sensing signaling molecule 2-aminoacetophenone mediates the persistence of Pseudomonas aeruginosa in macrophages by interference with autophagy through epigenetic regulation of lipid biosynthesis. MBio, 2023, 14(2), e00159-23. http://dx.doi.org/10.1128/mbio.00159-23 PMID: 37010415
- [23] Hutt, D.M.; Herman, D.; Rodrigues, A.P.C.; Noel, S.; Pilewski, J.M.; Matteson, J.; Hoch, B.; Kellner, W.; Kelly, J.W.; Schmidt, A.; Thomas, P.J.; Matsumura, Y.; Skach, W.R.; Gentzsch, M.; Riordan, J.R.; Sorscher, E.J.; Okiyoneda, T.; Yates, J.R., III; Lukacs, G.L.; Frizzell, R.A.; Manning, G.; Gottesfeld, J.M.; Balch, W.E. Reduced histone deacetylase 7 activity restores function to misfolded CFTR in cystic fibrosis. Nat. Chem. Biol., 2010, 6(1), 25-33.

http://dx.doi.org/10.1038/nchembio.275 PMID: 19966789

[24] Anglès, F.; Hutt, D.M.; Balch, W.E. HDAC inhibitors rescue multiple disease-causing CFTR variants. Hum. Mol. Genet., 2019, 28(12), 1982-2000.

http://dx.doi.org/10.1093/hmg/ddz026 PMID: 30753450

[25] Rosenjack, J.; Hodges, C.A.; Darrah, R.J.; Kelley, T.J. HDAC6 depletion improves cystic fibrosis mouse airway responses to bacterial challenge. Sci. Rep., 2019, 9(1), 10282.

http://dx.doi.org/10.1038/s41598-019-46555-4 PMID: 31311988

[26] Lin, Y.J.; Anzaghe, M.; Schülke, S. Update on the pathomechanism, diagnosis, and treatment options for rheumatoid arthritis. *Cells*, **2020**, *9*(4), 880.

http://dx.doi.org/10.3390/cells9040880 PMID: 32260219

- [27] Kondo, N.; Kuroda, T.; Kobayashi, D. Cytokine networks in the pathogenesis of rheumatoid arthritis. *Int. J. Mol. Sci.*, 2021, 22(20), 10922.
- http://dx.doi.org/10.3390/ijms222010922 PMID: 34681582
- [28] Elemam, N.M.; Hannawi, S.; Maghazachi, A.A. Role of chemokines and chemokine receptors in rheumatoid arthritis. *Immuno-Targets Ther.*, 2020, 9, 43-56. http://dx.doi.org/10.2147/ITT.S243636 PMID: 32211348
- [29] Ding, Q.; Hu, W.; Wang, R.; Yang, Q.; Zhu, M.; Li, M.; Cai, J.; Rose, P.; Mao, J.; Zhu, Y.Z. Signaling pathways in rheumatoid arthritis: implications for targeted therapy. *Signal Transduct. Target. Ther.*, **2023**, 8(1), 68. http://dx.doi.org/10.1038/s41392-023-01331-9 PMID: 36797236
- [30] Angiolilli, C.; Kabala, P.A.; Grabiec, A.M.; Van Baarsen, I.M.;
 Ferguson, B.S.; García, S.; Malvar Fernandez, B.; McKinsey, T.A.; Tak, P.P.; Fossati, G.; Mascagni, P.; Baeten, D.L.;
 Reedquist, K.A. Histone deacetylase 3 regulates the inflammatory gene expression programme of rheumatoid arthritis fibroblast-like synoviocytes. *Ann. Rheum. Dis.*, **2017**, *76*(1), 277-285.
 http://dx.doi.org/10.1136/annrheumdis-2015-209064 PMID: 27457515
- [31] Mao, D.; Jiang, H.; Zhang, F.; Yang, H.; Fang, X.; Zhang, Q.; Zhao, G. HDAC2 exacerbates rheumatoid arthritis progression via the IL-17-CCL7 signaling pathway. *Environ. Toxicol.*, 2023, 38(7), 1743-1755. http://dx.doi.org/10.1002/tox.23802 PMID: 37021908
- [32] Li, M.; Hu, W.; Wang, R.; Li, Z.; Yu, Y.; Zhuo, Y.; Zhang, Y.; Wang, Z.; Qiu, Y.; Chen, K.; Ding, Q.; Qi, W.; Zhu, M.; Zhu, Y. Sp1 S-Sulfhydration Induced by Hydrogen Sulfide Inhibits Inflammation via HDAC6/MyD88/NF-κB Signaling Pathway in Adjuvant-Induced Arthritis. *Antioxidants*, **2022**, *11*(4), 732. http://dx.doi.org/10.3390/antiox11040732 PMID: 35453416
- [33] Park, J.K.; Shon, S.; Yoo, H.J.; Suh, D.H.; Bae, D.; Shin, J.; Jun, J.H.; Ha, N.; Song, H.; Choi, Y.I.; Pap, T.; Song, Y.W. Inhibition of histone deacetylase 6 suppresses inflammatory responses and invasiveness of fibroblast-like-synoviocytes in inflammatory arthritis. *Arthritis Res. Ther.*, **2021**, *23*(1), 177. http://dx.doi.org/10.1186/s13075-021-02561-4 PMID: 34225810
- [34] Zhu, M.; Ding, Q.; Lin, Z.; Fu, R.; Zhang, F.; Li, Z.; Zhang, M.; Zhu, Y. New targets and strategies for rheumatoid arthritis: from signal transduction to epigenetic aspect. *Biomolecules*, 2023, 13(5), 766.

http://dx.doi.org/10.3390/biom13050766 PMID: 37238636

- [35] Park, J.K.; Jang, Y.J.; Oh, B.R.; Shin, J.; Bae, D.; Ha, N.; Choi, Y.; Youn, G.S.; Park, J.; Lee, E.Y.; Lee, E.B.; Song, Y.W. Therapeutic potential of CKD-506, a novel selective histone deacetylase 6 inhibitor, in a murine model of rheumatoid arthritis. *Arthritis Res. Ther.*, **2020**, *22*(1), 176. http://dx.doi.org/10.1186/s13075-020-02258-0 PMID: 32711562
- Bae, D.; Choi, Y.; Lee, J.; Ha, N.; Suh, D.; Baek, J.; Park, J.; Son, W. M-134, a novel HDAC6-selective inhibitor, markedly improved arthritic severity in a rodent model of rheumatoid arthritis when combined with tofacitinib. *Pharmacol. Rep.*, 2021, 73(1), 185-201.
- http://dx.doi.org/10.1007/s43440-020-00188-x PMID: 33188511 [37] Zhe, W.; Hoshina, N.; Itoh, Y.; Tojo, T.; Suzuki, T.; Hase, K.;
- [57] Zhe, W., Hoshink, N., Hon, T., Holo, T., Suzaki, F., Hase, K., Takahashi, D. A novel HDAC1-selective inhibitor attenuates autoimmune arthritis by inhibiting inflammatory cytokine production. *Biol. Pharm. Bull.*, **2022**, *45*(9), 1364-1372. http://dx.doi.org/10.1248/bpb.b22-00321 PMID: 36047206
- [38] Mane, R.R.; Kale, P.P. The roles of HDAC with IMPDH and mTOR with JAK as future targets in the treatment of rheumatoid arthritis with combination therapy. *J. Complement. Integr. Med.*, **2022**, *0*(0)

http://dx.doi.org/10.1515/jcim-2022-0114 PMID: 36409592

[39] Karami, J.; Aslani, S.; Tahmasebi, M.N.; Mousavi, M.J.; Sharafat Vaziri, A.; Jamshidi, A.; Farhadi, E.; Mahmoudi, M. Epigenetics in rheumatoid arthritis; fibroblast-like synoviocytes as an emerging paradigm in the pathogenesis of the disease. *Immunol. Cell Biol.*, **2020**, *98*(3), 171-186.

- http://dx.doi.org/10.1111/imcb.12311 PMID: 31856314
- [40] Angiolilli, C.; Grabiec, A.M.; Ferguson, B.S.; Ospelt, C.; Malvar Fernandez, B.; van Es, I.E.; van Baarsen, L.G.M.; Gay, S.; McKinsey, T.A.; Tak, P.P.; Baeten, D.L.; Reedquist, K.A. Inflammatory cytokines epigenetically regulate rheumatoid arthritis fibroblast-like synoviocyte activation by suppressing HDAC5 expression. Ann. Rheum. Dis., 2016, 75(2), 430-438. http://dx.doi.org/10.1136/annrheumdis-2014-205635 PMID: 25452308
- [41] Björkegren, JLM Atherosclerosis: Recent developments. Cell, 2022, 185(10), 1630-1645.
- [42] Gusev, E.; Sarapultsev, A. Atherosclerosis and inflammation: Insights from the theory of general pathological processes. *Int. J. Mol. Sci.*, **2023**, *24*(9), 7910. http://dx.doi.org/10.3390/ijms24097910 PMID: 37175617
- [43] Shao, B.Z.; Xu, H.Y.; Zhao, Y.C.; Zheng, X.R.; Wang, F.; Zhao, G.R. NLRP3 inflammasome in atherosclerosis: putting out the fire of inflammation. *Inflammation*, 2023, 46(1), 35-46. http://dx.doi.org/10.1007/s10753-022-01725-x PMID: 35953687
- [44] Kong, P.; Cui, Z.Y.; Huang, X.F.; Zhang, D.D.; Guo, R.J.; Han, M. Inflammation and atherosclerosis: signaling pathways and therapeutic intervention. *Signal Transduct. Target. Ther.*, **2022**, 7(1), 131.

http://dx.doi.org/10.1038/s41392-022-00955-7 PMID: 35459215

- [45] Bhattacharya, P.; Kanagasooriyan, R.; Subramanian, M. Tackling inflammation in atherosclerosis: Are we there yet and what lies beyond? *Curr. Opin. Pharmacol.*, **2022**, *66*, 102283. http://dx.doi.org/10.1016/j.coph.2022.102283 PMID: 36037627
- Yang, H.; Sun, Y.; Li, Q.; Jin, F.; Dai, Y. Diverse epigenetic regulations of macrophages in atherosclerosis. *Front. Cardiovasc. Med.*, 2022, 9, 868788. http://dx.doi.org/10.3389/fcvm.2022.868788 PMID: 35425818

[47] Chen, X.; He, Y.; Fu, W.; Sahebkar, A.; Tan, Y.; Xu, S.; Li, H. Histone deacetylases (HDACs) and atherosclerosis: a mechanistic and pharmacological review. *Front. Cell Dev. Biol.*, **2020**, *8*, 581015.

http://dx.doi.org/10.3389/fcell.2020.581015 PMID: 33282862

- [48] Lee, H.T.; Oh, S.; Ro, D.H.; Yoo, H.; Kwon, Y.W. The key role of DNA methylation and histone acetylation in epigenetics of atherosclerosis. *J. Lipid Atheroscler.*, **2020**, *9*(3), 419-434. http://dx.doi.org/10.12997/jla.2020.9.3.419 PMID: 33024734
- [49] Fang, Z.; Wang, X.; Sun, X.; Hu, W.; Miao, Q.R. The role of histone protein acetylation in regulating endothelial function. *Front. Cell Dev. Biol.*, **2021**, *9*, 672447. http://dx.doi.org/10.3389/fcell.2021.672447 PMID: 33996829
- [50] Shen, Z.; Bei, Y.; Lin, H.; Wei, T.; Dai, Y.; Hu, Y.; Zhang, C.; Dai, H. The role of class IIa histone deacetylases in regulating endothelial function. *Front. Physiol.*, 2023, 14, 1091794. http://dx.doi.org/10.3389/fphys.2023.1091794 PMID: 36935751
- [51] Chang, S.; Young, B.D.; Li, S.; Qi, X.; Richardson, J.A.; Olson, E.N. Histone deacetylase 7 maintains vascular integrity by repressing matrix metalloproteinase 10. *Cell*, **2006**, *126*(2), 321-334. http://dx.doi.org/10.1016/j.cell.2006.05.040 PMID: 16873063
- [52] Chen, L.; Shang, C.; Wang, B.; Wang, G.; Jin, Z.; Yao, F.; Yue, Z.; Bai, L.; Wang, R.; Zhao, S.; Liu, E.; Wang, W. HDAC3 inhibitor suppresses endothelial-to-mesenchymal transition via modulating inflammatory response in atherosclerosis. *Biochem. Pharma*col., 2021, 192, 114716.

http://dx.doi.org/10.1016/j.bcp.2021.114716 PMID: 34339713

[53] Nomura, Y.; Nakano, M.; Woo Sung, H.; Han, M.; Pandey, D. Inhibition of HDAC6 Activity Protects Against Endothelial Dysfunction and Atherogenesis *in vivo*: A Role for HDAC6 Neddylation. *Front. Physiol.*, 2021, 12, 675724.

http://dx.doi.org/10.3389/fphys.2021.675724 PMID: 34220539
 [54] Asare, Y.; Campbell-James, T.A.; Bokov, Y.; Yu, L.L.; Prestel,

[54] Asate, F., Campoen-Janes, T.A., Bokov, T., Ju, E.L., Hestel, M.; El Bounkari, O.; Roth, S.; Megens, R.T.A.; Straub, T.; Thomas, K.; Yan, G.; Schneider, M.; Ziesch, N.; Tiedt, S.; Silvestre-Roig, C.; Braster, Q.; Huang, Y.; Schneider, M.; Malik, R.; Haffner, C.; Liesz, A.; Soehnlein, O.; Bernhagen, J.; Dichgans, M. Histone deacetylase 9 activates IKK to regulate atherosclerotic plaque vulnerability. *Circ. Res.*, **2020**, *127*(6), 811-823. http://dx.doi.org/10.1161/CIRCRESAHA.120.316743 PMID: 32546048

- [55] Luan, Y.; Liu, H.; Luan, Y.; Yang, Y.; Yang, J.; Ren, K.D. New insight in HDACs: Potential therapeutic targets for the treatment of atherosclerosis. *Front. Pharmacol.*, **2022**, *13*, 863677. http://dx.doi.org/10.3389/fphar.2022.863677 PMID: 35529430
- [56] Zanza, C.; Romenskaya, T.; Manetti, A.C.; Franceschi, F.; La Russa, R.; Bertozzi, G.; Maiese, A.; Savioli, G.; Volonnino, G.; Longhitano, Y. Cytokine storm in COVID-19: immunopathogenesis and therapy. *Medicina (Kaunas)*, **2022**, *58*(2), 144. http://dx.doi.org/10.3390/medicina58020144 PMID: 35208467
- [57] Gusev, E.; Sarapultsev, A.; Solomatina, L.; Chereshnev, V. SARS-CoV-2-Specific immune response and the pathogenesis of COVID-19. *Int. J. Mol. Sci.*, **2022**, *23*(3), 1716. http://dx.doi.org/10.3390/ijms23031716 PMID: 35163638
- [58] Pires, B.G.; Calado, R.T. Hyper-inflammation and complement in COVID -19. Am. J. Hematol., 2023, 98(S4)(Suppl. 4), S74-S81. http://dx.doi.org/10.1002/ajh.26746 PMID: 36999459
- [59] Zhang, W.; Zhao, Y.; Zhang, F.; Wang, Q.; Li, T.; Liu, Z.; Wang, J.; Qin, Y.; Zhang, X.; Yan, X.; Zeng, X.; Zhang, S. The use of anti-inflammatory drugs in the treatment of people with severe coronavirus disease 2019 (COVID-19): The Perspectives of clinical immunologists from China. *Clin. Immunol.*, **2020**, *214*, 108393. http://dx.doi.org/10.1016/j.clim.2020.108393 PMID: 32222466
- [60] Murakami, N.; Hayden, R.; Hills, T.; Al-Samkari, H.; Casey, J.; Del Sorbo, L.; Lawler, P.R.; Sise, M.E.; Leaf, D.E. Therapeutic advances in COVID-19. *Nat. Rev. Nephrol.*, **2023**, *19*(1), 38-52. http://dx.doi.org/10.1038/s41581-022-00642-4 PMID: 36253508
- [61] Perico, N.; Cortinovis, M.; Suter, F.; Remuzzi, G. Home as the new frontier for the treatment of COVID-19: the case for antiinflammatory agents. *Lancet Infect. Dis.*, **2023**, *23*(1), e22-e33. http://dx.doi.org/10.1016/S1473-3099(22)00433-9 PMID: 36030796
- [62] Li, G.; Hilgenfeld, R.; Whitley, R.; De Clercq, E. Therapeutic strategies for COVID-19: progress and lessons learned. *Nat. Rev. Drug Discov.*, 2023, 22(6), 449-475. http://dx.doi.org/10.1038/s41573-023-00672-y PMID: 37076602
- [63] Shirvaliloo, M. Epigenomics in COVID-19; the link between DNA methylation, histone modifications and SARS-CoV-2 infection. *Epigenomics*, 2021, *13*(10), 745-750. http://dx.doi.org/10.2217/epi-2021-0057 PMID: 33876664
- [64] Behrouj, H.; Vakili, O.; Sadeghdoust, A.; Aligolighasemabadi, N.;
 Khalili, P.; Zamani, M.; Mokarram, P. Epigenetic regulation of autophagy in coronavirus disease 2019 (COVID-19). *Biochem. Biophys. Rep.*, 2022, 30, 101264.
- http://dx.doi.org/10.1016/j.bbrep.2022.101264 PMID: 35469237
 [65] Dey, A.; Vaishak, K.; Deka, D.; Radhakrishnan, A.K.; Paul, S.; Shanmugam, P.; Daniel, A.P.; Pathak, S.; Duttaroy, A.K.; Banerjee, A. Epigenetic perspectives associated with COVID-19 infection and related cytokine storm: an updated review. *Infection*, 2023, 1-16.

http://dx.doi.org/10.1007/s15010-023-02017-8 PMID: 36906872

[66] Rabaan, A.A.; Aljeldah, M.; Shammari, B.R.A.; Alsubki, R.A.; Alotaibi, J.; Alhashem, Y.N.; Alali, N.A.; Sulaiman, T.; Alsalem, Z.; Bajunaid, H.A.; Garout, M.; Alsaffar, H.A.; Almuthree, S.A.; Hudhaiah, D.; Alzaher, A.M.; Alshaikh, F.A.; Alshengeti, A.; Najim, M.A.; Farahat, R.A.; Mohapatra, R.K. Epigenetic targets and pathways linked to SARS-CoV-2 infection and pathology. *Microorganisms*, **2023**, *11*(2), 341. http://dx.doi.org/10.3390/microorganisms11020341 PMID:

http://dx.doi.org/10.3390/microorganisms11020341 PMID 36838306

- [67] Ripamonti, C.; Spadotto, V. HDAC inhibition as potential therapeutic strategy to restore the deregulated immune response in severe COVID-19. *Front Immunol*, 2022, 13, 841716.
- [68] Liu, K.; Zou, R.; Cui, W.; Li, M.; Wang, X.; Dong, J.; Li, H.; Li, H.; Wang, P.; Shao, X.; Su, W.; Chan, H.C.S.; Li, H.; Yuan, S. Clinical HDAC inhibitors are effective drugs to prevent the entry of SARS-CoV2. ACS Pharmacol. Transl. Sci., 2020, 3(6), 1361-1370.

http://dx.doi.org/10.1021/acsptsci.0c00163 PMID: 34778724

[69] Takahashi, Y.; Hayakawa, A.; Sano, R.; Fukuda, H.; Harada, M.; Kubo, R.; Okawa, T.; Kominato, Y. Histone deacetylase inhibitors suppress ACE2 and ABO simultaneously, suggesting a preventive potential against COVID-19. *Sci. Rep.*, **2021**, *11*(1), 3379. http://dx.doi.org/10.1038/s41598-021-82970-2 PMID: 33564039

[70] Saiz, M.L.; DeDiego, M.L.; López-García, D.; Corte-Iglesias, V.; Baragaño Raneros, A.; Astola, I.; Asensi, V.; López-Larrea, C.; Suarez-Alvarez, B. Epigenetic targeting of the ACE2 and NRP1 viral receptors limits SARS-CoV-2 infectivity. *Clin. Epigenetics*, 2021, 13(1), 187.

http://dx.doi.org/10.1186/s13148-021-01168-5

- Sixto-López, Y.; Correa-Basurto, J. HDAC inhibition as neuroprotection in COVID-19 infection. *Curr. Top. Med. Chem.*, 2022, 22(16), 1369-1378. http://dx.doi.org/10.2174/1568026622666220303113445 PMID: 35240959
- [72] Lambrecht, B.N.; Hammad, H. The immunology of asthma. Nat. Immunol., 2015, 16(1), 45-56.

http://dx.doi.org/10.1038/ni.3049 PMID: 25521684

[73] Siroux, V.; Boudier, A.; Bousquet, J.; Bresson, J.L.; Cracowski, J.L.; Ferran, J.; Gormand, F.; Just, J.; Le Moual, N.; Morange, S.; Nadif, R.; Oryszczyn, M.P.; Pison, C.; Scheinmann, P.; Varraso, R.; Vervloet, D.; Pin, I.; Kauffmann, F. Phenotypic determinants of uncontrolled asthma. *J. Allergy Clin. Immunol.*, **2009**, *124*(4), 681-687.e3.

http://dx.doi.org/10.1016/j.jaci.2009.06.010 PMID: 19665764

- [74] Boonpiyathad, T.; Sözener, Z.C.; Satitsuksanoa, P.; Akdis, C.A. Immunologic mechanisms in asthma. *Semin. Immunol.*, 2019, 46, 101333.
- http://dx.doi.org/10.1016/j.smim.2019.101333 PMID: 31703832
 [75] Hammad, H.; Lambrecht, B.N. The basic immunology of asthma. *Cell*, 2021, 184(6), 1469-1485.

http://dx.doi.org/10.1016/j.cell.2021.02.016 PMID: 33711259

- [76] Ora, J.; Calzetta, L.; Matera, M.G.; Cazzola, M.; Rogliani, P. Advances with glucocorticoids in the treatment of asthma: state of the art. *Expert Opin. Pharmacother.*, 2020, 21(18), 2305-2316. http://dx.doi.org/10.1080/14656566.2020.1807514 PMID: 32808828
- [77] He, Y.; Shi, J.; Nguyen, Q.T.; You, E.; Liu, H.; Ren, X.; Wu, Z.; Li, J.; Qiu, W.; Khoo, S.K.; Yang, T.; Yi, W.; Sun, F.; Xi, Z.; Huang, X.; Melcher, K.; Min, B.; Xu, H.E. Development of highly potent glucocorticoids for steroid-resistant severe asthma. *Proc. Natl. Acad. Sci. USA*, **2019**, *116*(14), 6932-6937. http://dx.doi.org/10.1073/pnas.1816734116 PMID: 30894497
- [78] Nadeem, A.; Ahmad, S.F.; Al-Harbi, N.O.; Ibrahim, K.E.; Siddiqui, N.; Al-Harbi, M.M.; Attia, S.M.; Bakheet, S.A. Inhibition of Bruton's tyrosine kinase and IL-2 inducible T-cell kinase suppresses both neutrophilic and eosinophilic airway inflammation in a cockroach allergen extract-induced mixed granulocytic mouse model of asthma using preventative and therapeutic strategy. *Pharmacol. Res.*, **2019**, *148*, 104441.

http://dx.doi.org/10.1016/j.phrs.2019.104441 PMID: 31505252

[79] Nadeem, A.; Ahmad, S.F.; Al-Harbi, N.O.; El-Sherbeeny, A.M.; Alasmari, A.F.; Alanazi, W.A.; Alasmari, F.; Ibrahim, K.E.; Al-Harbi, M.M.; Bakheet, S.A.; Attia, S.M. Bruton's tyrosine kinase inhibitor suppresses imiquimod-induced psoriasis-like inflammation in mice through regulation of IL-23/IL-17A in innate immune cells. *Int. Immunopharmacol.*, **2020**, *80*, 106215.

http://dx.doi.org/10.1016/j.intimp.2020.106215 PMID: 31982823

[80] Nadeem, A.; Alshehri, S.; Al-Harbi, N.O.; Ahmad, S.F.; Albekairi, N.A.; Alqarni, S.A.; Ibrahim, K.E.; Alfardan, A.S.; Alshamrani, A.A.; Bin Salman, S.B.; Attia, S.M. Bruton's tyrosine kinase inhibition suppresses neutrophilic inflammation and restores histone deacetylase 2 expression in myeloid and structural cells in a mixed granulocytic mouse model of asthma. *Int. Immunopharmacol.*, 2023, 117, 109920.

http://dx.doi.org/10.1016/j.intimp.2023.109920 PMID: 36827920

- [81] Weber, A.N.R.; Bittner, Z.; Liu, X.; Dang, T.M.; Radsak, M.P.; Brunner, C. Bruton's tyrosine kinase: an emerging key player in innate immunity. *Front. Immunol.*, 2017, 8, 1454. http://dx.doi.org/10.3389/fimmu.2017.01454 PMID: 29167667
- [82] Islam, R.; Dash, D.; Singh, R. Intranasal curcumin and sodium butyrate modulates airway inflammation and fibrosis via HDAC inhibition in allergic asthma. *Cytokine*, 2022, 149, 155720. http://dx.doi.org/10.1016/j.cyto.2021.155720 PMID: 34634654

- [83] Chiappara, G.; Gagliardo, R.; Siena, A.; Bonsignore, M.R.; Bousquet, J.; Bonsignore, G.; Vignola, A.M. Airway remodelling in the pathogenesis of asthma. Curr. Opin. Allergy Clin. Immunol., 2001, 1(1), 85-93. http://dx.doi.org/10.1097/01.all.0000010990.97765.a1 PMID. 11964675
- [84] Wang, J.; Wen, L.; Wang, Y.; Chen, F. Therapeutic effect of histone deacetylase inhibitor, sodium butyrate, on allergic rhinitis in vivo. DNA Cell Biol., 2016, 35(4), 203-208. http://dx.doi.org/10.1089/dna.2015.3037 PMID: 26859163
- [85] Shabab, T.; Khanabdali, R.; Moghadamtousi, S.Z.; Kadir, H.A.; Mohan, G. Neuroinflammation pathways: a general review. Int. J. Neurosci., 2017, 127(7), 624-633. http://dx.doi.org/10.1080/00207454.2016.1212854 PMID: 27412492
- [86] Rump, K.; Adamzik, M. Epigenetic mechanisms of postoperative cognitive impairment induced by anesthesia and neuroinflammation. Cells, 2022, 11(19), 2954. http://dx.doi.org/10.3390/cells11192954 PMID: 36230916
- Liesz, A.; Zhou, W.; Na, S.Y.; Hämmerling, G.J.; Garbi, N.; [87] Karcher, S.; Mracsko, E.; Backs, J.; Rivest, S.; Veltkamp, R. Boosting regulatory T cells limits neuroinflammation in permanent cortical stroke. J. Neurosci., 2013, 33(44), 17350-17362. http://dx.doi.org/10.1523/JNEUROSCI.4901-12.2013 PMID. 24174668
- [88] Leigh, T.; Scalia, R.G.; Autieri, M.V. Resolution of inflammation in immune and nonimmune cells by interleukin-19. Am. J. Physiol. Cell Physiol., 2020, 319(3), C457-C464. http://dx.doi.org/10.1152/ajpcell.00247.2020 PMID: 32667867
- [89] Dai, Y.; Wei, T.; Shen, Z.; Bei, Y.; Lin, H.; Dai, H. Classical HDACs in the regulation of neuroinflammation. Neurochem. Int., 2021, 150, 105182. http://dx.doi.org/10.1016/j.neuint.2021.105182 PMID: 34509559
- [90] Guo, A.; Li, J.; Luo, L.; Chen, C.; Lu, Q.; Ke, J.; Feng, X. Valproic acid mitigates spinal nerve ligation-induced neuropathic pain in rats by modulating microglial function and inhibiting neuroinflammatory response. Int. Immunopharmacol., 2021, 92, 107332.
- http://dx.doi.org/10.1016/j.intimp.2020.107332 PMID: 33421931 [91] Borgonetti, V.; Governa, P.; Manetti, F.; Galeotti, N. Zingiberene, a non-zinc-binding class I HDAC inhibitor: A novel strategy for the management of neuropathic pain. Phytomedicine, 2023, 111, 154670.

http://dx.doi.org/10.1016/j.phymed.2023.154670 PMID: 36681053

- [92] Liu, Y.F.; Hu, R.; Zhang, L.F.; Fan, Y.; Xiao, J.F.; Liao, X.Z. Effects of dexmedetomidine on cognitive dysfunction and neuroinflammation via the HDAC2/HIF-1α/PFKFB3 axis in a murine model of postoperative cognitive dysfunction. J. Biochem. Mol. Toxicol., 2022, 36(6), e23044. http://dx.doi.org/10.1002/jbt.23044 PMID: 35499365
- [93] Li, Y.; Seto, E. HDACs and HDAC inhibitors in cancer development and therapy. Cold Spring Harb. Perspect. Med., 2016, 6(10), a026831.
- http://dx.doi.org/10.1101/cshperspect.a026831 PMID: 27599530 [94] Singh, T.; Kaur, P.; Singh, P.; Singh, S.; Munshi, A. Differential
- molecular mechanistic behavior of HDACs in cancer progression. Med. Oncol., 2022, 39(11), 171. http://dx.doi.org/10.1007/s12032-022-01770-4 PMID: 35972597
- [95] Hai, R.; He, L.; Shu, G.; Yin, G. Characterization of histone deacetylase mechanisms in cancer development. Front. Oncol., 2021, 11, 700947. http://dx.doi.org/10.3389/fonc.2021.700947 PMID: 34395273
- [96] Patra, S.; Panigrahi, D.P.; Praharaj, P.P.; Bhol, C.S.; Mahapatra, K.K.; Mishra, S.R.; Behera, B.P.; Jena, M.; Bhutia, S.K. Dysregulation of histone deacetylases in carcinogenesis and tumor progression: a possible link to apoptosis and autophagy. Cell. Mol. Life Sci., 2019, 76(17), 3263-3282.
- http://dx.doi.org/10.1007/s00018-019-03098-1 PMID: 30982077 [97] Kim, J.Y.; Cho, H.; Yoo, J.; Kim, G.W.; Jeon, Y.H.; Lee, S.W.;
- Kwon, S.H. Pathological role of HDAC8: Cancer and beyond. Cells, 2022, 11(19), 3161. http://dx.doi.org/10.3390/cells11193161 PMID: 36231123

[98] Hanisch, D.; Krumm, A.; Diehl, T.; Stork, C.M.; Dejung, M.;

Butter, F.; Kim, E.; Brenner, W.; Fritz, G.; Hofmann, T.G.; Roos,

W.P. Class I HDAC overexpression promotes temozolomide resistance in glioma cells by regulating RAD18 expression. Cell Death Dis., 2022, 13(4), 293.

- http://dx.doi.org/10.1038/s41419-022-04751-7 PMID: 35365623 Cai, S.; Chen, W.; Zeng, W.; Cheng, X.; Lin, M.; Wang, J. Roles
- [99] of HDAC2, eIF5, and eIF6 in lung cancer tumorigenesis. Curr. Med. Sci., 2021, 41(4), 764-769.
 - http://dx.doi.org/10.1007/s11596-021-2389-z PMID: 34403101
- [100] Yin, Y.; Zhang, M.; Dorfman, R.G.; Li, Y.; Zhao, Z.; Pan, Y.; Zhou, Q.; Huang, S.; Zhao, S.; Yao, Y.; Zou, X. Histone deacetylase 3 overexpression in human cholangiocarcinoma and promotion of cell growth via apoptosis inhibition. Cell Death Dis., 2017, 8(6), e2856-e2856. http://dx.doi.org/10.1038/cddis.2016.457 PMID: 28569784
- [101] Zhang, S.L.; Zhu, H.Y.; Zhou, B.Y.; Chu, Y.; Huo, J.R.; Tan, Y.Y.; Liu, D.L. Histone deacetylase 6 is overexpressed and promotes tumor growth of colon cancer through regulation of the MAPK/ERK signal pathway. OncoTargets Ther., 2019, 12, 2409-2419

http://dx.doi.org/10.2147/OTT.S194986 PMID: 31118659

[102] Eckschlager, T.; Plch, J.; Stiborova, M.; Hrabeta, J. Histone deacetylase inhibitors as anticancer drugs. Int. J. Mol. Sci., 2017, 18(7), 1414. http://dx.doi.org/10.3390/ijms18071414 PMID: 28671573

- [103] Jenke, R.; Reßing, N.; Hansen, F.K.; Aigner, A.; Büch, T. Anticancer therapy with HDAC inhibitors: mechanism-based combination strategies and future perspectives. Cancers (Basel), 2021, 13(4), 634.
- http://dx.doi.org/10.3390/cancers13040634 PMID: 33562653 [104] Patel, V.K.; Shirbhate, E.; Tiwari, P.; Kore, R.; Veerasamy, R.; Mishra, A.; Rajak, H. Multi-targeted HDAC Inhibitors as Anti-

cancer Agents: Current Status and Future Prospective. Curr. Med. Chem., 2023, 30(24), 2762-2795. http://dx.doi.org/10.2174/0929867329666220922105615 PMID: 36154583

[105] Mehmood, S.A.; Sahu, K.K.; Sengupta, S.; Partap, S.; Karpoormath, R.; Kumar, B.; Kumar, D. Recent advancement of HDAC inhibitors against breast cancer. Med. Oncol., 2023, 40(7), 201

http://dx.doi.org/10.1007/s12032-023-02058-x PMID: 37294406

- [106] Psilopatis, I.; Garmpis, N.; Garmpi, A.; Vrettou, K.; Sarantis, P.; Koustas, E.; Antoniou, E.A.; Dimitroulis, D.; Kouraklis, G.; Karamouzis, M.V.; Marinos, G.; Kontzoglou, K.; Nonni, A.; Nikolettos, K.; Fleckenstein, F.N.; Zoumpouli, C.; Damaskos, C. The Emerging Role of Histone Deacetylase Inhibitors in Cervical Cancer Therapy. Cancers (Basel), 2023, 15(8), 2222. http://dx.doi.org/10.3390/cancers15082222 PMID: 37190151
- [107] Drzewiecka, M.; Gajos-Michniewicz, A.; Hoser, G.; Jaśniak, D.; Barszczewska-Pietraszek, G.; Sitarek, P.; Czarny, P.; Piekarski, J.; Radek, M.; Czyż, M.; Skorski, T.; Śliwiński, T. Histone Deacetylases (HDAC) Inhibitor-Valproic Acid Sensitizes Human Melanoma Cells to Dacarbazine and PARP Inhibitor. Genes (Basel), 2023, 14(6), 1295.

http://dx.doi.org/10.3390/genes14061295 PMID: 37372475

- [108] Roca, M.S.; Moccia, T.; Iannelli, F.; Testa, C.; Vitagliano, C.; Minopoli, M.; Camerlingo, R.; De Riso, G.; De Cecio, R.; Bruzzese, F.; Conte, M.; Altucci, L.; Di Gennaro, E.; Avallone, A.; Leone, A.; Budillon, A. HDAC class I inhibitor domatinostat sensitizes pancreatic cancer to chemotherapy by targeting cancer stem cell compartment via FOXM1 modulation. J. Exp. Clin. Cancer Res., 2022, 41(1), 83. http://dx.doi.org/10.1186/s13046-022-02295-4 PMID: 34980222
- [109] Fan, F.; Liu, P.; Bao, R.; Chen, J.; Zhou, M.; Mo, Z.; Ma, Y.; Liu, H.; Zhou, Y.; Cai, X.; Qian, C.; Liu, X. A dual PI3K/HDAC inhibitor induces immunogenic ferroptosis to potentiate cancer immune checkpoint therapy. Cancer Res., 2021, 81(24), 6233-6245. http://dx.doi.org/10.1158/0008-5472.CAN-21-1547 PMID: 34711611
- [110] Bär, S.I.; Pradhan, R.; Biersack, B.; Nitzsche, B.; Höpfner, M.; Schobert, R. New chimeric HDAC inhibitors for the treatment of colorectal cancer. Arch. Pharm. (Weinheim), 2023, 356(2), 2200422.

http://dx.doi.org/10.1002/ardp.202200422 PMID: 36442846

- [111] He, S.; Dong, G.; Li, Y.; Wu, S.; Wang, W.; Sheng, C. Potent dual BET/HDAC inhibitors for efficient treatment of pancreatic cancer. *Angew. Chem. Int. Ed.*, **2020**, *59*(8), 3028-3032. http://dx.doi.org/10.1002/anie.201915896 PMID: 31943585
- [112] Roy, R.; Ria, T.; RoyMahaPatra, D.; Sk, U.H. Single inhibitors versus dual inhibitors: Role of HDAC in cancer. ACS Omega, 2023, 8(19), 16532-16544. http://dx.doi.org/10.1021/acsomega.3c00222 PMID: 37214715
- [113] Hu, Z.; Wei, F.; Su, Y.; Wang, Y.; Shen, Y.; Fang, Y.; Ding, J.; Chen, Y. Histone deacetylase inhibitors promote breast cancer metastasis by elevating NEDD9 expression. *Signal Transduct. Target. Ther.*, 2023, 8(1), 11. http://dx.doi.org/10.1038/s41392-022-01221-6 PMID: 36604412
- [114] Zhu, J.; Han, S. Histone deacetylase 10 exerts antitumor effects on cervical cancer *via* a novel microRNA-223/TXNIP/Wnt/β-catenin pathway. *IUBMB Life*, **2021**, *73*(4), 690-704. http://dx.doi.org/10.1002/iub.2450 PMID: 33481334