SHORT COMMUNICATION

Pathogenic monoallelic variants in *GLIS3* **increase type 2 diabetes risk and identify a subgroup of patients sensitive to sulfonylureas**

Sarah Meulebrouck¹ · Victoria Scherrer¹ · Raphaël Boutry¹ · Bénédicte Toussaint¹ · Emmanuel Vaillant¹ · **Aurélie Dechaume1 · Hélène Loiselle1 · Beverley Balkau² [·](http://orcid.org/0000-0003-2021-413X) Guillaume Charpentier[3](http://orcid.org/0000-0001-6124-5712) · Sylvia Franc3,4 · Michel Marre5,[6](http://orcid.org/0000-0002-3071-1837) · Morgane Baron1 · Martine Vaxillaire1 · Mehdi Derhourhi1 · Mathilde Boissel1 · Philippe Froguel1,7 · Amélie Bonnefond1,[7](http://orcid.org/0000-0001-9976-3005)**

Received: 10 July 2023 / Accepted: 19 September 2023 / Published online: 5 December 2023 © The Author(s) 2023

Abstract

Aims/hypothesis *GLIS3* encodes a transcription factor involved in pancreatic beta cell development and function. Rare pathogenic, bi-allelic mutations in *GLIS3* cause syndromic neonatal diabetes whereas frequent SNPs at this locus associate with common type 2 diabetes risk. Because rare, functional variants located in other susceptibility genes for type 2 diabetes have already been shown to strongly increase individual risk for common type 2 diabetes, we aimed to investigate the contribution of rare pathogenic *GLIS3* variants to type 2 diabetes.

Methods *GLIS3* was sequenced in 5471 individuals from the Rare Variants Involved in Diabetes and Obesity (RaDiO) study. Variant pathogenicity was assessed following the criteria established by the American College of Medical Genetics and Genomics (ACMG). To address the pathogenic strong criterion number 3 (PS3), we conducted functional investigations of these variants using luciferase assays, focusing on capacity of GLIS family zinc fnger 3 (GLIS3) to bind to and activate the *INS* promoter. The association between rare pathogenic or likely pathogenic (P/LP) variants and type 2 diabetes risk (and other metabolic traits) was then evaluated. A meta-analysis combining association results from RaDiO, the 52K study (43,125 individuals) and the TOPMed study (44,083 individuals) was fnally performed.

Results Through targeted resequencing of *GLIS3*, we identifed 105 rare variants that were carried by 395 participants from RaDiO. Among them, 49 variants decreased the activation of the *INS* promoter. Following ACMG criteria, 18 rare variants were classifed as P/LP, showing an enrichment in the last two exons compared with the remaining exons (*p*<5×10−6; OR>3.5). The burden of these P/LP variants was strongly higher in individuals with type 2 diabetes (*p*=3.0×10−3; OR 3.9 [95% CI 1.4, 12]), whereas adiposity, age at type 2 diabetes diagnosis and cholesterol levels were similar between variant carriers and non-carriers with type 2 diabetes. Interestingly, all carriers with type 2 diabetes were sensitive to oral sulfonylureas. A total of 7 P/LP variants were identifed in both 52K and TOPMed studies. The meta-analysis of association studies obtained from RaDiO, 52K and TOPMed showed an enrichment of P/LP *GLIS3* variants in individuals with type 2 diabetes (*p*=5.6×10−5; OR 2.1 [95% CI 1.4, 2.9]).

Conclusions/interpretation Rare P/LP *GLIS3* variants do contribute to type 2 diabetes risk. The variants located in the distal part of the protein could have a direct efect on its functional activity by impacting its transactivation domain, by homology with the mouse GLIS3 protein. Furthermore, rare P/LP *GLIS3* variants seem to have a direct clinical efect on beta cell function, which could be improved by increasing insulin secretion via the use of sulfonylureas.

Keywords ACMG · Functional genetics · GLIS3 · Luciferase assays · Rare variants · Type 2 diabetes

Extended author information available on the last page of the article

Research in context

What is already known about this subject?

- GLIS3 encodes a transcription factor playing a key role in pancreatic beta cell development and function \bullet
- Rare pathogenic bi-allelic GLIS3 variants cause neonatal obesity, whereas frequent SNPs identified through genome-wide association studies modestly increase type 2 diabetes risk
- For several susceptibility genes for type 2 diabetes, it has been demonstrated that rare functional variants can strongly contribute to type 2 diabetes risk

What is the key question?

Do rare pathogenic GLIS3 variants strongly contribute to type 2 diabetes risk and allow the identification of a subgroup of patients?

What are the new findings?

- Rare, pathogenic or likely pathogenic (P/LP) GLIS3 variants strongly increase type 2 diabetes risk, and people with type 2 diabetes carrying a P/LP variant respond well to sulfonylurea treatment
- \bullet BMI, age at diagnosis and cholesterol levels are similar between variant carriers and non-carriers with type 2 diabetes
- P/LP GLIS3 variants are enriched in the C-terminal part of the protein, potentially impacting directly its transactivation domain

How might this impact on clinical practice in the foreseeable future?

People with type 2 diabetes carrying a P/LP GLIS3 variant could be specifically treated with sulfonylureas for longterm restoration of insulin secretion

Introduction

Type 2 diabetes, like many other multifactorial disorders, includes monogenic forms that are rare, more severe, and appear earlier in life than common polygenic forms [[1](#page-4-0)]. Rare, pathogenic, bi-allelic variants in *GLIS3* cause a severe syndrome that includes neonatal diabetes [\[2\]](#page-4-1). GLIS family zinc fnger 3 (GLIS3) is a transcription factor playing a major role in pancreatic beta cell development and function. Notably, it enhances the transcription of *INS* by binding to its promoter and recruiting the transcription factors pancreatic and duodenal homeobox 1 (PDX1), MafA and neuronal differentiation 1 (NEUROD1) [[3](#page-4-2)]. In addition, genomewide association studies (GWAS) identifed frequent SNPs at the *GLIS3* locus associated with common type 2 diabetes risk, and with variation in fasting glucose and beta cell function [[4\]](#page-4-3). We and others previously found strong impact on common type 2 diabetes risk of rare, functional variants located in susceptibility genes identified by GWAS [[1\]](#page-4-0), paving the way for new insights into the underlying pathophysiology and personalised treatment strategies. This approach is particularly fruitful as it is challenging to uncover such information solely through GWAS, where associated SNPs are typically non-coding and exhibit modest efect [[5\]](#page-4-4).

We wondered whether rare, deleterious *GLIS3* variants could be associated with increased risk for common type 2 diabetes, and whether they might impact drug treatment choices. In this respect, a recent study showed an association between rare *GLIS3* variants and increased disease risk [\[6](#page-4-5)], but the lack of pathogenicity assessment using in vitro data may limit its conclusion.

Methods

Study participants We analysed 5471 blood DNA samples accurately sequenced from several population studies included in the Rare Variants Involved in Diabetes and Obesity (RaDiO) study [[7\]](#page-4-6). The RaDiO study and criteria of inclusion are described in detail in the electronic supplementary material (ESM) Methods.

GLIS3 sequencing and variant annotation *GLIS3* DNA sequencing (NM_001042413.2) was previously performed by next-generation sequencing [[7](#page-4-6)]. Only rare variants with a minor allele frequency (MAF) below 1% in any population study in the GnomAD browser (v2.1.1) and in the present study were kept for further analyses. All rare coding variants detected in *GLIS3* had a quality QUAL score higher than 50. No variant presented more than 5% of missing genotype across the participants.

Statistical analyses for association studies The burdens of rare coding variants identifed in RaDiO were analysed as a single cluster using the mixed-efects score test (MiST), as previously described [[7\]](#page-4-6). The association studies between the burdens of variants and clinical traits were adjusted for age, sex, BMI and ancestry (for assessing type 2 diabetes risk, age at type 2 diabetes diagnosis and cholesterol levels), or for age, sex and ancestry (for assessing BMI). Participant ancestry was assessed using the frst fve genotypic principal components as previously described [\[7](#page-4-6)]. The meta-analysis was performed using the generic inverse variance method from the R package meta $[8]$. The common effect model was applied because of low heterogeneity.

Plasmid generation Plasmids encoding wild-type *GLIS3* gene and rare variants were either purchased from Twist Bioscience (San Francisco, CA, USA) or generated from the frst one using the Quick Change site-directed mutagenesis kit from Stratagene (San Diego, CA, USA), and verifed by Sanger sequencing. Please see ESM Methods for more details.

Culture of HEK293 cells Human embryonic kidney 293 **(**HEK293) cells were purchased from American Type Culture Collection (Manassas, VA, USA). These cells were cultured in DMEM containing 10% FBS (vol./vol.) and 50 units/ml penicillin/streptomycin. These cells were regularly tested for mycoplasma contamination.

Luciferase assays HEK293 cells were transfected in suspension using Lipofectamine 2000 Transfection Reagent (Thermo Fisher Scientifc, Waltham, MA, USA), with wildtype or mutated *GLIS3* plasmid, plasmid including the gene encoding frefy luciferase driven by the 5′ fanking region of *INS* containing GLIS binding sites, and plasmid including the gene encoding β-galactosidase, with or without *MAFA* plasmid. Please see ESM Methods for further details.

Results

Through targeted resequencing of *GLIS3* (NM_001042413.2) in 5471 participants from RaDiO [[7\]](#page-4-6), we detected 105 rare coding *GLIS3* variants, including one nonsense variant (p.Y627*), among 395 carriers (ESM Table 1). At this stage of analysis, the burden of rare variants was not associated with type 2 diabetes risk (p_{π} =0.20 [p_{overall} =0.055], where p_{π} is the *p* value for the impact of the burden; with an OR of 0.85 [95% CI, 0.66, 1.1]; Table [1\)](#page-2-0).

To assess the pathogenicity of the 105 variants, we used the criteria from the American College of Medical Genetics and Genomics (ACMG) [[9\]](#page-4-8); notably we developed in vitro assays to address the pathogenic strong criterion number 3 (PS3). Plasmids including each *GLIS3* variant were overexpressed along with their gene reporter assay to assess the ability of each mutant to bind to the 5′ fanking region of *INS* containing GLIS3 binding sites. Furthermore, these variants were evaluated in conjunction with the overexpression of MafA to determine their capacity for recruiting this transcription factor to the *INS* 5' flanking region, consequently enhancing luciferase signalling. When compared with wildtype *GLIS3*, 49 variants decreased luciferase activity with and/or without the addition of *MAFA* in the system, and were considered loss-of-function (ESM Fig. 1).

In comparison with our functional results, the in silico pathogenicity prediction by REVEL had a poor sensitivity as only 4% of loss-of-function variants were predicted to be deleterious (ESM Table 1), but had a high specifcity (ESM Table 1).

Following ACMG criteria including the PS3 criterion, 18 out of 105 variants, carried by 18 unrelated individuals of European ancestry, were found to be pathogenic or likely pathogenic (P/LP; Fig. [1](#page-3-0) and ESM Table 2). Furthermore, P/ LP variants were strongly enriched in the C-terminal part of *GLIS3*, i.e. the last two coding exons after accounting for exon length (p <5×10⁻⁶ with an OR>3.5; Fig. [1](#page-3-0) and ESM Table 3).

We then assessed the effect of the burden of the 18 rare P/ LP *GLIS3* variants on type 2 diabetes risk. This burden was four times higher among participants with type 2 diabetes compared

Table 1 Association analyses between rare coding *GLIS3* variants and type 2 diabetes risk

Association analyses were performed using the MiST method for the burden of the 105 rare *GLIS3* variants, or for the burden of the 18 P/LP variants only

MiST provides a score statistic $S(\pi)$ for the mean effect (π) of the cluster, and a score statistic $S(\tau)$ for the heterogeneous effect (τ) of the cluster. The overall *p* value combined *p* values p_{π} and p_{τ}

 p_{π} , *p* value indicating the impact of the burden; p_{τ} , *p* value indicating the heterogeneity of the burden; T2D, type 2 diabetes

Fig. 1 Location of the 18 P/LP variants in *GLIS3*, shown using arrows. Ser-rich, regions enriched in serine; Pro-rich, region enriched in proline

with controls $(p_{\pi} = 8.0 \times 10^{-3} [p_{\text{overall}} = 3.0 \times 10^{-3}]$; with an OR of 3.9 [95% CI 1.4, 12]; Table [1\)](#page-2-0). We found an equal number of female and male participants carrying a P/LP *GLIS3* variant, either with type 2 diabetes or not (data not shown). Interestingly, all participants with type 2 diabetes carrying a P/LP *GLIS3* variant were treated with sulfonylureas, a class of medication that stimulates the secretion of insulin by activating ATP-dependent potassium channels. In contrast, 50% of non-carriers sufering from type 2 diabetes were treated with sulfonylureas. The sibling of one of the participants with type 2 diabetes (carrying the P/LP variant encoding p.A168G) also presented with type 2 diabetes and was also treated with sulfonylureas. The age at diagnosis, BMI and cholesterol levels were similar among P/LP variant carriers vs non-carriers with type 2 diabetes (ESM Table 4).

We then analysed the association between P/LP variants in *GLIS3* (NM_001042413 [ENST00000381971] transcript) and type 2 diabetes risk in the Type 2 Diabetes Knowledge Portal (using the genetic association interactive tool) [[10](#page-4-9)]. In 43,125 individuals from 52K and in 44,083 individuals from TOPMed, we only found eight P/LP variants per study (i.e. loss-of-function transcript efect estimator [LofTee] variants with a very low MAF in GnomAD; ESM Table 5) that had no efect on disease risk due to a low statistical power (*p*=0.13 with an OR of 1.7 and *p*=0.44 with an OR of 3.3, respectively; variable threshold test). No P/LP missense variants identifed in our study were observed in either TOPMed or the 52K study. However, through a meta-analysis of RaDiO, TOPMed and 52K studies, we found an enrichment of P/LP *GLIS3* variants among individuals with type 2 diabetes (*p*=5.6×10−5 with an OR of 2.1 [1.4, 2.9]).

Discussion

On the basis of functional genetics, we identifed 18 P/LP heterozygous *GLIS3* variants that increase type 2 diabetes risk. This corroborates the continuum between monogenic and polygenic type 2 diabetes [\[5](#page-4-4)], and supports the importance of functional genomics to identify variants associated with metabolic disorders, as previously demonstrated [[1\]](#page-4-0).

Importantly, all the participants carrying a P/LP *GLIS3* variant were treated with sulfonylureas, and their BMI was similar to non-carriers. This suggests that these variants directly alter beta cell function, but that this defect can be compensated for efectively by increasing beta cell insulin secretion. In the context of precision medicine, individuals harbouring a rare P/LP *GLIS3* variant might be deemed suitable candidates for treatment with sulfonylureas, over other medication options.

Furthermore, we observed an enrichment of P/LP variants in the C-terminal domain of GLIS3. In a prior study, two individuals with type 2 diabetes carried two highly rare deleterious variants in the last exons [\[6](#page-4-5)]. We suggest that mutations located at the C-terminal domain of GLIS3 could directly afect its activity by impacting its transactivation domain. Indeed, the transactivation domain of GLIS3 mouse protein, which presents a high homology with human GLIS3, is located in its C-terminal section [\[11](#page-4-10)], and the human c.2338dupC mutation, which generates a truncated protein (missing the last coding exons), strongly decreased GLIS3 transcriptional activity [[2\]](#page-4-1).

Our study has limitations, as our luciferase experiments focused on the ability of GLIS3 to bind to the *INS* promoter only, despite its multiple transactivation role in beta cells. It is thus possible that some variants impacting other aspects of GLIS3 function were not identifed as loss-of-function in this study.

In conclusion, P/LP monoallelic *GLIS3* variants contribute to increased type 2 diabetes risk, in addition to *GLIS3* involvement in monogenic diabetes. Sulfonylureas might be sufficient to manage type 2 diabetes in the carriers.

Supplementary Information The online version contains peer-reviewed but unedited supplementary material available at [https://doi.org/10.](https://doi.org/10.1007/s00125-023-06035-x) [1007/s00125-023-06035-x.](https://doi.org/10.1007/s00125-023-06035-x)

Acknowledgements We are grateful to all individuals included in the different cohort studies. We thank F. Allegaert, and T. Beke (Inserm UMR1283, CNRS UMR8199, European Genomic Institute for Diabetes [EGID], Institut Pasteur de Lille, Lille University Hospital, Lille, France) and M. Vieubled (4P-Pharma, Lille, France) for their technical assistance. Data from this article were obtained as part of the PhD thesis of S. Meulebrouck entitled 'Impact of rare genetic variants on the etiology of type 2 diabetes' (2020). Several data from this article have been previously exposed during an oral presentation entitled 'The impact of rare pathogenic variants of *GLIS3* on type 2 diabetes' at the 58th EASD Annual Meeting (2022).

Data availability Data regarding variants from TOPMed and 52K studies are available on the AMP Type 2 Diabetes Knowledge Portal via this link: [https://t2d.hugeamp.org/gait.html?gene=GLIS3&tests=](https://t2d.hugeamp.org/gait.html?gene=GLIS3&tests=burden%2Cvt&transcript=ENST00000381971) [burden%2Cvt&transcript=ENST00000381971.](https://t2d.hugeamp.org/gait.html?gene=GLIS3&tests=burden%2Cvt&transcript=ENST00000381971)

Data regarding *GLIS3* variants from the GnomAD browser (v2.1.1) via this link: [https://gnomad.broadinstitute.org/gene/ENSG00000107249?](https://gnomad.broadinstitute.org/gene/ENSG00000107249?dataset=gnomad_r2_1) [dataset=gnomad_r2_1](https://gnomad.broadinstitute.org/gene/ENSG00000107249?dataset=gnomad_r2_1)

Other data are available upon request to the corresponding author.

Funding We thank 'France Génomique' consortium (ANR-10- INBS-009). This work was supported by grants from the French National Research Agency (ANR-10-LABX-46 [European Genomics Institute for Diabetes] and ANR-10-EQPX-07-01 [LIGAN-PM]), from the European Research Council (ERC $O\pi O - 101043671$, to AB), and from the National Center for Precision Diabetic Medicine – PreciDIAB, which is jointly supported by the French National Agency for Research (ANR-18-IBHU-0001), by the European Union (FEDER), by the Hauts-de-France Regional Council and by the European Metropolis of Lille (MEL).

Contribution statement SM, PF and AB conceptualised and designed the study. SM, VS, RB, HL and MBaron contributed to functional analysis of each variant. BT, EV and AD performed sequencing experiments. MBoissel and AB performed statistical analyses. MD performed computer analyses. BB, GC, SF, MM, MV and PF contributed to collection of cohort data. SM and AB wrote the frst draft of this manuscript. SM and AB have accessed and verifed all the data from this study. SM, AB and PF are responsible for the integrity of the work as a whole. All the authors revised and approved the fnal version of the manuscript.

Author's relationship and activities AB is a member of the Editorial Board of *Diabetologia*. The authors declare that they have no other relationships or activities that might bias, or be perceived to bias, their work.

Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit<http://creativecommons.org/licenses/by/4.0/>.

References

- 1. Bonnefond A, Semple RK (2022) Achievements, prospects and challenges in precision care for monogenic insulin-defcient and insulin-resistant diabetes. Diabetologia 65(11):1782–1795. [https://](https://doi.org/10.1007/s00125-022-05720-7) doi.org/10.1007/s00125-022-05720-7
- 2. Senée V, Chelala C, Duchatelet S et al (2006) Mutations in GLIS3 are responsible for a rare syndrome with neonatal diabetes mellitus and congenital hypothyroidism. Nat Genet 38(6):682–687. <https://doi.org/10.1038/ng1802>
- 3. ZeRuth GT, Takeda Y, Jetten AM (2013) The Krüppel-like protein Gli-similar 3 (Glis3) functions as a key regulator of insulin transcription. Mol Endocrinol 27(10):1692–1705. [https://doi.org/](https://doi.org/10.1210/me.2013-1117) [10.1210/me.2013-1117](https://doi.org/10.1210/me.2013-1117)
- 4. Wen X, Yang Y (2017) Emerging roles of GLIS3 in neonatal diabetes, type 1 and type 2 diabetes. J Mol Endocrinol 58(2):R73– R85. <https://doi.org/10.1530/JME-16-0232>
- 5. Bonnefond A, Froguel P (2015) Rare and common genetic events in type 2 diabetes: what should biologists know? Cell Metab 21(3):357–368.<https://doi.org/10.1016/j.cmet.2014.12.020>
- 6. Sun J, Have CT, Hollensted M et al (2019) Sequencing reveals protective and pathogenic efects on development of diabetes of rare GLIS3 variants. PLoS One 14(8):e0220805. [https://doi.org/](https://doi.org/10.1371/journal.pone.0220805) [10.1371/journal.pone.0220805](https://doi.org/10.1371/journal.pone.0220805)
- 7. Bonnefond A, Boissel M, Bolze A et al (2020) Pathogenic variants in actionable MODY genes are associated with type 2 diabetes. Nat Metab 2(10):1126–1134.<https://doi.org/10.1038/s42255-020-00294-3>
- 8. Balduzzi S, Rücker G, Schwarzer G (2019) How to perform a meta-analysis with R: a practical tutorial. Evid Based Ment Health 22(4):153–160.<https://doi.org/10.1136/ebmental-2019-300117>
- 9. Richards S, Aziz N, Bale S et al (2015) Standards and guidelines for the interpretation of sequence variants: a joint consensus recommendation of the American College of Medical Genetics and Genomics and the Association for Molecular Pathology. Genet Med 17(5):405–424.<https://doi.org/10.1038/gim.2015.30>
- 10. Costanzo MC, von Grotthuss M, Massung J et al (2023) The Type 2 Diabetes Knowledge Portal: an open access genetic resource dedicated to type 2 diabetes and related traits. Cell Metab 35(4):695-710.e6.<https://doi.org/10.1016/j.cmet.2023.03.001>
- 11. Beak JY, Kang HS, Kim Y-S, Jetten AM (2008) Functional analysis of the zinc fnger and activation domains of Glis3 and mutant Glis3(NDH1). Nucleic Acids Res 36(5):1690–1702. [https://doi.](https://doi.org/10.1093/nar/gkn009) [org/10.1093/nar/gkn009](https://doi.org/10.1093/nar/gkn009)

Publisher's Note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Authors and Afliations

Sarah Meulebrouck¹ · Victoria Scherrer¹ · Raphaël Boutry¹ · Bénédicte Toussaint¹ · Emmanuel Vaillant¹ · **Aurélie Dechaume1 · Hélène Loiselle1 · Beverley Balkau² [·](http://orcid.org/0000-0003-2021-413X) Guillaume Charpentier[3](http://orcid.org/0000-0001-6124-5712) · Sylvia Franc3,4 · Michel Marre5,[6](http://orcid.org/0000-0002-3071-1837) · Morgane Baron1 · Martine Vaxillaire1 · Mehdi Derhourhi1 · Mathilde Boissel1 · Philippe Froguel1,7 · Amélie Bonnefond1,[7](http://orcid.org/0000-0001-9976-3005)**

- \boxtimes Philippe Froguel p.froguel@imperial.ac.uk
- \boxtimes Amélie Bonnefond amelie.bonnefond@inserm.fr
- ¹ Inserm UMR1283, CNRS UMR8199, European Genomic Institute for Diabetes (EGID), Université de Lille, Institut Pasteur de Lille, Lille University Hospital, Lille, France
- ² Inserm U1018 Clinical Epidemiology, Center for Research in Epidemiology and Population Health, Paris-Saclay University, Paris-Sud University, UVSQ, Villejuif, France
- ³ CERITD (Centre d'Étude et de Recherche pour l'Intensifcation du Traitement du Diabète), Evry, France
- ⁴ Department of Diabetes, Sud-Francilien Hospital, Paris-Sud University, Corbeil-Essonnes, France
- ⁵ Institut Necker-Enfants Malades, Inserm, Université de Paris, Paris, France
- ⁶ Clinique Ambroise Paré, Neuilly-sur-Seine, France
- ⁷ Department of Metabolism, Digestion and Reproduction, Imperial College London, London, UK