



# *Review* **Melanoma Antigen Family A (MAGE A) as Promising Biomarkers and Therapeutic Targets in Bladder Cancer**

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**Simple Summary:** The Melanoma Antigen Gene (MAGE) belongs to the larger family of cancer testis antigens. The MAGEA family were the first tumor-associated antigens identified at the molecular level whose expression was consistent in most human cancers and germinal cells. Aberrant expression of MAGEA family is noted in a majority of human malignancies, where they are associated with increased cancer cell proliferation, survival, and resistance to various therapies. This makes them an ideal biomarker and attractive therapeutic target in designing novel therapies. This review mainly focuses on the opportunities for the development of MAGEAs as promising biomarkers and their therapeutic implications in bladder cancer.

**Abstract:** The Melanoma Antigen Gene (MAGE) is a large family of highly conserved proteins that share a common MAGE homology domain. Interestingly, many MAGE family members exhibit restricted expression in reproductive tissues but are abnormally expressed in various human malignancies, including bladder cancer, which is a common urinary malignancy associated with high morbidity and mortality rates. The recent literature suggests a more prominent role for MAGEA family members in driving bladder tumorigenesis. This review highlights the role of MAGEA proteins, the potential for them to serve as diagnostic or prognostic biomarker(s), and as therapeutic targets for bladder cancer.

**Keywords:** bladder cancer; Melanoma Antigen Gene; cancer testis antigen; biomarkers; therapeutic target; signaling pathways; protein–protein interaction; genomic alterations

# **1. Introduction**

Bladder cancer continues to be an important health problem with an estimated 600,000 new cases and 220,000 deaths worldwide [\[1\]](#page-14-0). In the United States, more than 80,000 new cases and 17,000 deaths are predicted this year [\[2\]](#page-14-1). Urothelial carcinoma is the most common type of bladder cancer, which arises from the urothelial cells lining the bladder's inner surface. At presentation, nearly 75% of patients have non-muscleinvasive bladder (NMIBC) cancer and 25% have muscle-invasive (MIBC) disease. About 50% of NMIBC are low-grade, whereas most muscle-invasive or metastatic tumors are highgrade and invade the detrusor muscle [\[2](#page-14-1)[–4\]](#page-14-2). NMIBC is typically managed via cystoscopic



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transurethral resection in combination with adjuvant intravesical chemo- or immunetherapy [\[2](#page-14-1)[,5\]](#page-14-3). Unfortunately, disease recurrence is common (40–75%) despite guidelineconcordant care, and approximately 30% of cases progress to MIBC, which often requires systemic chemotherapy with radical pelvic surgery (cystectomy) or chemo-radiation [\[6,](#page-14-4)[7\]](#page-14-5). Treatment recommendations are based on risk stratification, which presently do not include biological heterogeneity in bladder cancer. One way to address this gap is to identify novel biomarkers and therapeutic targets to manage this life-threatening disease.

In the past decade, the search for diverse biomarkers involved in cancer initiation and/or its progression led to the discovery of cancer–testis antigens (CTAs) as a breakthrough in cancer biology and clinical oncology [\[8–](#page-14-6)[10\]](#page-14-7). CTAs are commonly expressed in various tumor types but have limited expression patterns in normal tissues; therefore, they are proposed as cancer biomarkers for diagnosis [\[11](#page-14-8)[–15\]](#page-15-0). Some of these molecules are progressively increased during carcinogenesis and thus recognized as prognostic markers; both could be effective targets for prevention and/or therapeutic intervention. In the human genome, more than 200 CTA genes have been documented and classified into 44 gene families in the CTA database [\(http://www.cta.lncc.br](http://www.cta.lncc.br) (accessed on 4 September 2023)) and GeneBank [\(https://www.ncbi.nlm.nih.gov/gene](https://www.ncbi.nlm.nih.gov/gene) (accessed on 5 September 2023)) [\[16,](#page-15-1)[17\]](#page-15-2). A few of these families consist of multiple members and CTA gene orthologs and paralogs. CTAs are divided into two groups based on the chromosomal location that include CT-X antigens positioned in the X chromosome or non-X CTAs sited on the autosomes [\[18\]](#page-15-3). The distribution of antigens on the X-chromosome (Xq21-q28 region) is organized in gene clusters harboring groups of direct and inverted repeats, whereas non-X-CTA genes are mostly single-copy genes [\[19\]](#page-15-4). The expression of type I or CT-X is restricted to the X chromosome that comprises three subfamilies, MAGEA, MAGEB, and MAGEC, whereas type II, which is not limited to the X-chromosome, includes MAGED, MAGEE, MAGEF, MAGEG, MAGEH, MAGEL, and Necdin [\[20\]](#page-15-5). Additionally, CTA pseudo-genes were identified in the MAGEA, MAGEB, SSX, CT45, and CT47 families in human and mammalian genomes [\[21\]](#page-15-6). The expression of 174 CTA-encoding genes might be regulated by CTA-non-coding RNAs, which are annotated in the Cancer Genome Atlas [\(https://cancergenome.nih.gov](https://cancergenome.nih.gov) (assessed on 6 October 2023) [\[22\]](#page-15-7). However, both type I and type II groups have a Melanoma Antigen family (MAGE) homology domain (MHD), which is highly conserved within the MAGEA subfamily (>80% identical) consisting of approximately 170 amino acids, except the MAGED proteins, which consist of two MHDs [\[23](#page-15-8)[,24\]](#page-15-9). Structure analysis has documented that MHD is a tandem-winged helix motif that presumably plays a key role in protein–protein interaction [\[25\]](#page-15-10). While other studies have demonstrated the similarity between MAGE proteins having distinct functions [\[26\]](#page-15-11), it was projected that the adaptable MHD undergoes allosteric changes to allow interactions between different protein domains, conferring special properties to MAGE members [\[27](#page-15-12)[,28\]](#page-15-13). The members of MAGED family were considered the ancestral and most conserved with the highest homology of gene and protein sequences between humans and other species based on alignment score data [\(https://www.ncbi.nlm.nih.gov/homologene](https://www.ncbi.nlm.nih.gov/homologene) (accessed on 16 October 2023) (Figure [1\)](#page-2-0).

Members of the MAGEA family were the first tumor-associated antigens identified in humans at the molecular level [\[29–](#page-15-14)[31\]](#page-15-15). To date, 12 family members of MAGEA have been identified, including type I MAGEs, that are characteristically restricted to expression in the testis and are often abnormally expressed in various human malignancies. Preferential intracellular location may be dissimilar for different antigens, such as MAGEA1, MAGEA3, and MAGEA4, which are mostly cytoplasmic, but MAGEA10 is majorly nuclear in localization [\[32](#page-15-16)[,33\]](#page-15-17). The mechanisms that control the unusual re-expression of MAGEAs are still under exploration. MAGEAs are infrequently expressed in somatic tissues; however, epigenetic changes including DNA hypermethylation of CpG dinucleotides in promoters and posttranslational modifications of the histone proteins averts transcription factors from binding, and thus represses the expression of MAGEA genes [\[34\]](#page-15-18). Further studies have shown that DNA methyltransferases (DNMTs) inhibitors, such as 5-aza-2-deoxycytidine,



<span id="page-2-0"></span>result in the increased expression of MAGEA1 in several malignant cells [\[35\]](#page-15-19). This effect can be additionally reinforced using histone deacetylase inhibitors [\[35](#page-15-19)[,36\]](#page-15-20).

**Figure 1.** Human MAGE proteins with the identified common domain, the MAGE homology domain (MHD). Few MAGEs have truncated MHDs and those members that are likely pseudogenes are<br>not listed. not listed.

Exercise cancer promotation and members inverving the VIR organing pairway [by].<br>The overexpression of MAGEA4 facilitates the growth of spontaneously transformed oral keratinocytes through apoptosis inhibition [40]. Other studies have found that MAGEA4<br>increases the survival of cancer cells through its interaction with Gankyrin and Miz1, which are transcriptional partners of c-myc. The recruitment of the MAGEA4-Miz1 transcriptional<br>complex on the Gip1/p21 premater results in the dourneaulation of Gip1/p21, thus enhancing cancer cell survival [41]. Through direct interaction, MAGEA3 and MAGEA6 are reported to be involved in the doleral matter of Tiwith R&T claims the subdition regulating autophagy and adaptation to nutrition stress [42]. This event results in the upregulation of m1OK signaling pathways facilitating early tumor formation [43]. Furthermore, MAGEA11<br>has been shown to interact with S phase kinase-associated protein (Skp2) modulating its specificity and association with cyclin A regulating cell cycle progression [44].<br>Numerous studies detail the syntession nattern of MACEA members in various burner. Several studies demonstrated the role of MAGEA family members in cancer cell proliferation and progression [\[27,](#page-15-12)[37,](#page-15-21)[38\]](#page-15-22). The aberrant expression of MAGEA3 facilitates cervical cancer proliferation and metastasis involving the Wnt signaling pathway [\[39\]](#page-15-23). keratinocytes through apoptosis inhibition [\[40\]](#page-15-24). Other studies have found that MAGEA4 complex on the Cip1/p21 promoter results in the downregulation of Cip1/p21, thus are reported to be involved in the ubiquitination of  $AMPK\alpha$  1 catalytic subunit regulating mTOR signaling pathways facilitating early tumor formation [\[43\]](#page-16-2). Furthermore, MAGEA11

tumors, while only few studies have identified mutations across different tumor types. A somate mutation analysis of the county regions of MAGEA failing showed mutations in<br>one or more members in melanoma patients [45]. An analysis of MAGEA4 mutant proteins in tumor cells exhibited high structural stability but showed changes in thermal stability<br>and folding that affect tumor growth [46]. Based on the mutation analysis, an overall low level of correlation was detected between MAGEAs mutation and antigen expression that Numerous studies detail the expression pattern of MAGEA members in various human somatic mutation analysis of the coding regions of MAGEA family showed mutations in and folding that affect tumor growth [\[46\]](#page-16-5). Based on the mutation analysis, an overall low might contribute to tumor progression.

 $\frac{1}{2}$  Clinical studies have shown a correlation between MAGEA expression and poor progrims in c[a](#page-16-8)ncer patients  $[47]$ . Studies also suggest that MAGEA protein expression is<br>associated with therapeutic resistance [48,49]. There is in[cre](#page-16-7)asing evidence demonstrating the involvement of MAGEA proteins in regulating the processes of cell survival in cancer<br>cells by direct interaction with the p53 tumor suppressor or indirectly by regulating the MAGEA antigens have been investigated for their role in human cancers and as potential  $MAGE$ A antigens have been investigated for their role in human cancers and as potential prognosis in cancer patients [\[47\]](#page-16-6). Studies also suggest that MAGEA protein expression is cells by direct interaction with the p53 tumor suppressor or indirectly by regulating the activity of E3 RING ubiquitin ligases [\[50,](#page-16-9)[51\]](#page-16-10). MAGEA proteins increase the metastatic potential of malignant cells by enhancing cell motility and invasiveness [\[47,](#page-16-6)[52\]](#page-16-11). Overall,

therapeutic targets. This review highlights the role of MAGEA family proteins in bladder cancer as putative diagnostic/prognostic biomarkers and in future directions toward advancements in MAGEA-specific therapies.

#### **2. Expression of MAGEA Family in Bladder Cancer**

There are a few reports on the expression of specific MAGEA family members in bladder cancers. Patard et al. (1995) analyzed the expression of MAGEA1, MAGEA2, MAGEA3, and MAGEA4 and observed the expression of at least one of these genes in 61% of invasive and 28% of superficial tumors, with MAGEA3 and MAGEA4 genes being most frequently expressed [\[53\]](#page-16-12). A study by Picard et al. (2007) analyzed the expression of MAGEA3, MAGEA4, MAGEA8, and MAGEA9 in bladder cancer. The study reported that MAGEA3–9 members were expressed in 30%, 33%, 56%, and 54% of bladder tumors, respectively. Although MAGEA8 was the most frequently expressed, its expression was low overall and mostly confined to the normal urothelium. In comparison, MAGEA9 was expressed at a higher level and was two times more frequent in superficial bladder cancer than in invasive tumors [\[54\]](#page-16-13). Bergeron et al. (2009) showed that MAGEA4 and MAGEA9 were expressed in 38% and 63% of NMIBC, in 48% and 57% of MIBC, 65% and 84% in carcinomas in situ, and 73% and 85% in lymph node metastases, respectively. The expression of MAGEA4 ( $p = 0.007$ ) and MAGEA9 ( $p = 0.012$ ) was associated with higher-grade tumors. In multivariable Cox regression analyses, the expression of MAGEA9 in pTa tumors was associated with recurrence (HR =  $1.829$ ;  $p = 0.010$ ). MAGEA4 expression in these tumors was associated with progression to MIBC ( $HR = 7.417$ ,  $p = 0.013$ ) based on univariate analyses, whereas MAGEA9 expression was further predictive of bladder cancer progression [\[55\]](#page-16-14). A study by Xylinas et al. (2014) showed that MAGEA3 expression was independently associated with an increased risk of bladder cancer recurrence and cancer-specific mortality [\[56\]](#page-16-15). Another study by Dyrskjot et al. (2012) showed that 43% of bladder tumors expressed MAGEA3. A univariate Cox regression analysis of gene expression in NMIBC showed that the expression of MAGEA3 ( $p = 0.026$ ) was significantly associated with a shorter progression-free survival [\[57\]](#page-16-16). A study by Kocher et al. (2002) showed that MAGEA4 protein was significantly expressed at higher levels in transitional cell carcinomas ( $p < 0.001$ ); its positivity was significantly correlated with an invasive phenotype (*p* < 0.001) and high-grade tumors (*p* < 0.0001). A retrospective evaluation of 908 transitional cell carcinomas of the bladder patients demonstrated strong MAGEA4 staining which was associated with decreased tumor-specific survival (*p* < 0.0001) [\[58\]](#page-16-17). Other studies have shown higher expressions of MAGEA2, MAGEA8, and MAGEA10 in high-grade bladder tumors [\[59](#page-16-18)[–63\]](#page-16-19). Overall, these studies suggest that MAGEA members are associated with bladder cancer, grade, stage, and oncologic outcomes.

We performed a meta-analysis combining the results from several independent studies on bladder cancer listed in the TCGA database for detecting differentially expressed MAGEA genes with the potential to increase both the statistical power and generalizability of our analysis. MAGEA family expression was analyzed in  $n = 4536$  patients according to tumor stage and the *p* values were generated by two-sample t-test, comparing pTa and pT2 stage bladder cancer. MAGEA2 (*p* < 0.001), MAGEA3 (*p* = 0.005), MAGEA6 (*p* < 0.001), and MAGEA12 ( $p = 0.01$ ) were found to have stage-dependent expression (highest in pT2 subgroup), suggesting that these MAGEA members might play important roles in the development of urothelial carcinoma (Figure [2\)](#page-4-0). These observations highlight the importance of improving our understanding of the etiology of bladder cancer, as well as the molecular changes underlying aberrant MAGEA expression. However, the clinical and prognostic value of MAGEA family members in the pathobiology of bladder cancer is currently under investigation by our group.

<span id="page-4-0"></span>

bladder cancer stages. The X-axis of the graph representation of the graph represe pT3, pT4, and pTa, and the Y-axis of the graph represents log2 value. pT3, pT4, and pTa, and the Y-axis of the graph represents log2 value. pT3, pT4, and pTa, and the Y-axis of the graph represents log2 value. **Figure 2.** MAGEA expression profile across various stages of bladder cancer. Box and dot-plot of **Figure 2.** MAGEA expression profile across various stages of bladder cancer. Box and dot-plot of bladder cancer stages. The X-axis of the graph represents various stages of bladder cancer,  $pT1$ ,  $pT2$ ,

# **3. Genomic Aberration of MAGEA Genes in Bladder Cancer 3. Genomic Aberration of MAGEA Genes in Bladder Cancer 3. Genomic Aberration of MAGEA Genes in Bladder Cancer**

To gain a better understanding of the molecular alteration of MAGEA proteins in bladder cancer, we used a TCGA bladder cancer patient's  $(n = 4536)$  genomics database to analyze how MAGEA family members could affect bladder tumorigenesis. Genomic<br>how we have al-proliferation and irregularities in cell death in neoplastic cells [\[64](#page-17-0)[,65\]](#page-17-1). It was found that frequencies of genomic alterations among MAGEA family members in bladder cancer were MAGEA1 1.8%, MAGEA2 1.6%, MAGEA3 1.9%, MAGEA4 1.6%, MAGEA6 2.6%, M[AG](#page-5-0)EA8 1.7%, MAGEA10 2%, MAGEA11, 2.1%, and MAGEA12 2.4% (Figure 3A). The most prevalent form of genomic alteration is gene amplification among all MAGEA members  $[66,67]$  $[66,67]$ . The amplification of these genes often has the potential to transform normal cells into neoplastic cells, further hinting at the possibility that the MAGEA family has a significant role in bladder tumorigenesis [\[68\]](#page-17-4). These observations recommend that an  $\frac{1}{100}$  significant role in blad der tumorigenesis to the contribution of  $\frac{1}{100}$  and  $\frac{1}{100}$  and of Enduce anten in during the amplified regions of some might before ingites the as derations, including mutation, homozygous deletion, or amplification, led to uncontrolled a significant fore in bladder tamorigenesis [60]. These observations recommend that an<br>analysis of MAGEAs might be a useful diagnostic tool to determine the invasive potential of bladder cancer. In addition, the amplified regions of some MAGEA genes might serve as potential therapeutic targets.

We also analyzed mutations in the MAGEA family in bladder cancer. MAGEA genes are more frequently mutated in cancer patients, suggesting their critical role in malignancy [69]. In bladder cancer, the somatic mutation frequency is very low and ranges from  $0.1\%$  to  $0.4\%$  in the MAGEA family members identified in 4880 samples from 4158 patients consolidated from 21 studies from the cancer genome atlas (TCGA) (Figure 3B).

<span id="page-5-0"></span>

Figure 3. Genomic alterations in MAGEA family members. Oncoprint of MAGEA1, MAGEA2, MAGEA3, MAGEA4, MAGEA6, MAGEA8, MAGEA10, MAGEA11, and MAGEA12 in bladder cancer patients ( $n = 5436$ ). (A) Color bar represents the individual patient's profile showing gene amplification in red,  $*$  denotes % gene amplification; deletion in blue; and mutation in green. (**B**) Bar represents the summary of MAGEA gene alteration frequency (Y-axis), and the X-axis shows the X-axis sho mutation rate (green), amplification (red), and deep deletion (blue) in bladder cancer patients. represents the summary of MAGEA gene alteration frequency (Y-axis), and the X-axis shows the the mutation rate (green), amplification (red), and deep deletion (blue) in bladder cancer patients. graph represents the summary of MAGEA gene alteration frequency (Y-axis), and the X-axis shows

and MAGEA11. Some of these mutations, including G137W, E232Q, P242L, Y249H, P262R, gene [\[56\]](#page-16-15). Among the noted mutations, the functional impact of P262R was most deleterious, which might contribute to poor outcomes in bladder cancer patients (Figure 4A,B). and MAGEA11. Some of these mutations, including G137W, E232Q, P242L, Y249H, P262K,<br>G296V, R298C, and E314Q, were found in the 229–399 amino acid sequence in the MAGEA6<br>gene [56]. Among the noted mutations, the functional i Moreover, missense mutations were identified in MAGEA6 compared to MAGEA3 G296V, R298C, and E314Q, were found in the 229–399 amino acid sequence in the MAGEA6

<span id="page-6-0"></span>

**Figure 4.** Mutation in the MAGEA family in bladder cancer. (**A**) Mutation plot was generated by the Mutation Mapper tool (cBioportal) showing the structure of MAGEA protein, and the frequency and position of mutations. Green color shows MAGE\_N: Melanoma-associated antigen family N terminal (4–97), and red color shows MAGE: MAGE family (116–286). The green lollipop denotes number and change in the amino acid. (**B**) The table shows the patient's TCGA sample ID, protein change, functional impact, and mutation type. Color dots denotes function impact: red—high, yellow/orange—moderate, and gray—low impact.

#### **4. Gene Network and Signaling Pathways of MAGEA Family in Bladder Cancer** members with other MAGE members, including MAGEA2B, MAGEB10, MAGEB2, and t. Gene network and Signaling Pathways of MAGEA Pamily in bladder Cancer

The altered expression of genes results in changes in gene expression and gene network interaction during cancer progression. A Cytoscape version 3.10.1 (Complex Net-work Analysis) Genemania module [\[70\]](#page-17-6) was used to explore the genetic interaction of MAGEA1, MAGEA2, MAGEA3, MAGEA4, MAGEA6, MAGEA8, MAGEA10, MAGEA11, and MAGEA12 (red color). This software tool provides a critical assessment and integration of protein–protein interactions to assess the associations of potential differentially expressed genes. The size of the circle of each protein represents its degree of connection to other proteins. The analysis demonstrated the interaction of MAGEA family members with other MAGE members, including MAGEA2B, MAGEB10, MAGEB2, and others (blue color). The available scientific literature reports on the interaction between MAGEA and  $MAGEB$  in bladder cancer [\[71\]](#page-17-7). Moreover, both family members share the MAGE domain that influences the tumor microenvironment and promotes cell proliferation (Figure [5A](#page-7-0)).

<span id="page-7-0"></span>

Figure 5. (A) Protein–protein interaction network of MAGEA family members constructed by scape so  $(x)$  is called with  $P(x)$ . Proteins are represented with color nodes, and interactions are represented with color nodes, and interactions are represented with color nodes, and interactions are represented with  $P$ Cytoscape software (version 3.10.1). Proteins are represented with color nodes, and interactions are  $\overline{\phantom{a}}$ represented with edges. The size of the circle of each protein represents its connection degree to other proteins. The red color circle shows the query protein, and the blue color shows the interacting proteins. (**B**) Signaling pathways associated with MAGEA family; the X-axis shows the log(B-H) value and the Y-axis represents the associated pathway in bladder cancer.

We further assessed the alteration in the MAGEA family of proteins affecting various signaling pathways promoting tumorigenesis [\[53](#page-16-12)[–63\]](#page-16-19). We analyzed three independent studies on bladder cancer (GSE154261, GSE57813, and GSE37317) using Ingenuity Pathway Analysis (IPA). Our analysis revealed that the molecular mechanism of cancer, mitochondrial dysfunction, protein ubiquitination, oxidative phosphorylation, and sirtuin signaling pathways are among the top five signaling networks associated with MAGEAs expression in bladder cancer. Furthermore, it was documented that a high expression of MAGEA3 modulates the function of the AMPK pathway and significantly decreases autophagy, leading to the activation of the mTOR signaling pathway [\[72\]](#page-17-8) (Figure [5B](#page-7-0)).

### **5. MAGEA Family as Diagnostic Biomarkers in Bladder Cancer**

Several studies have analyzed the stage-specific expression of the MAGEA family in bladder cancer, indicating their higher levels in invasive disease [\[53–](#page-16-12)[63\]](#page-16-19). Therefore, we explored the TCGA database to determine the expression of different MAGEA members in bladder cancer (BCLA;  $n = 414$ ) compared with normal bladder samples ( $n = 19$ ). The BCLA patient's dataset was not available for MAGEA9, and MAGEA5 and MAGEA7 were excluded from the analysis as they are pseudogenes. In some databases, the expression of MAGEA5 and MAGEA7 are measured, despite being pseudogenes, as they regulate MAGEA5 and MAGEA7 are measured, despite being pseudogenes, as they regulate ononcogenes by serving as miRNA decoys [\[73\]](#page-17-9). Interestingly, MAGEA2, MAGEA3, MAGEA4, cogenes by serving as miRNA decoys [73]. Interestingly, MAGEA2, MAGEA3, MAGEA4, MAGEA6, MAGEA10, MAGEA11, and MAGEA12 exhibited significant differences in their MAGEA6, MAGEA10, MAGEA11, and MAGEA12 exhibited significant differences in expression in bladder cancer compared to normal bladder samples. Other MAGEA family members, including MAGEA1 and MAGEA8, exhibited lower expression in bladder cancer compared to normal bladder specimen[s \(](#page-8-0)Figure 6). A data analysis of bladder cancer patients further uncovers the possibilities of utilizing MAGEA2, MAGEA3, MAGEA4,  $\rm MAGEA6$ ,  $\rm MAGE A10$ ,  $\rm MAGEA11$ , and  $\rm MAGEA12$  members as diagnostic biomarkers for bladder cancer.  $BCLA$  patient's dataset was not available for MAGEA9, and MAGEA5 and MAGEA7 were

<span id="page-8-0"></span>

**Figure 6.** Transcriptomic expression of MAGEA family. Relative expressions of varying MAGEA **Figure 6.** Transcriptomic expression of MAGEA family. Relative expressions of varying MAGEA types in normal samples and bladder cancer samples (BLCA) quantified through a box plot analysis types in normal samples and bladder cancer samples (BLCA) quantified through a box plot analysis of the TCGA dataset. The X-axis shows the expression of 435 patients (BCLA = 414, and Normal = 19), and the Y-axis shows the Fragments Per Kilobase of transcript per Million mapped reads (FPKM) mRNA expression values.

Next, we determined the cancer-specific mortality using a hazard ratio (HR) of MAGEA proteins with overall survival for the assessment of relative risk for bladder cancer aggressiveness, using the GENT2 database, which associates gene expression with the HR. The HR was calculated using the fixed effect model and the random effect model with 95% CI and *p*-value. Based on the data analysis from various studies, the HR value

for the fixed (FE) and random effect (RE) models were as follows: MAGEA1 (FE, 1.06; RE 1.18; *p*-value = 0.07); MAGEA2 (FE, 1.48; RE, 1.48; *p*-value = 0.67); MAGEA3 (FE,1.43; RE, 1.43; *p*-value = 0.33); MAGEA4 (FE, 0.96; RE, 1.21; *p*-value = 0.02); MAGEA6 (FE, 0.99, RE, 1.05; *p*-value = 0.03); MAGEA8 (FE, 1.05, RE, 1.05; *p*-value = 0.86); MAGEA10 (FE, 1.07.; rice, *p*-value = 0.02), MATELLIE (12) 1.00, 12) 1.00, *p*-value = 0.00, *m*-relative (12) 1.00,<br>RE,1.10; *p*-value = 0.11); MAGEA11 (FE, 1.09; RE,0.94; *p*-value = 0.31); and MAGEA12 (FE, 1.00; RE, 1.73; *p*-value < 0.01). The HR ratio of MAGEA6 and MAGEA12 was statistically significant with *p*-value = 0.03; <0.01. Based on these findings, poor survival might be predicted in patients that express elevated levels of MAGEA6 and MAGEA12. To define the variation among different datasets, the heterogeneity was calculated. Heterogeneity was common regardless of the treatment effects by odds ratios or risk differences. Random effects estimates, which incorporate heterogeneity, tended to be less precisely assessed than fixed effects estimates. Therefore, compared with the fixed effect model, the weights assigned under random effects are more balanced. The heterogeneity was 0%; this means that heterogeneity has no importance in the results displayed in the forest plot (Figure [7\)](#page-10-0). and  $= 0.11$ , was denoted the  $(1.05, 1.05, 1.05, 0.94, p$ -value  $= 0.51$ , and was denoted the mates, which incorporate heterogeneity, tended to be less precisely assessed than fixed



**Figure 7.** *Cont*.

<span id="page-10-0"></span>

**Figure 7.** Hazard ratio (HR) of MAGEA proteins in bladder cancer. Forest plots were generated for **Figure 7.** Hazard ratio (HR) of MAGEA proteins in bladder cancer. Forest plots were generated for hazard ratio in bladder cancer patients. HR was calculated using the fixed effect model and the random effect model with 95% CI with the *p*-value of heterogeneity.

# 6. Prognostic Value of MAGEA Gene Family in Bladder Cancer

We analyzed the prognostic value (log rank) of MAGEA family members viz. MAGEA -1, -3, -4, -6, -8, -10, -11, and -12 in bladder cancer patients (n = 408; MAGEA2, n = 406) using KM-Plot [\(http://kmplot.com/analysis](http://kmplot.com/analysis) (accessed on 19 October 2023)). For this, we performed survival analysis in bladder cancer patients using the TCGA database. A hazard ratio (HR) of more than 1.0 was used to predict the potential of genes as prognostic biomarkers. Based on the log rank value of MAGEA family members, we found that MAGEA6 scored the highest log rank (log rank 0.99), followed by MAGEA3 (log rank 0.83), compared to other MAGEA members. These values suggest that MAGEA3 and MAGEA6 may serve as prognostic biomarkers for bladder cancer patients (Figure [8\)](#page-11-0). Additional clinical validation in bladder cancer patients is required to justify the above rationale.

Next, we determined the predicted location and protein–protein interaction of MAGEA3 and MAGEA6 genes in bladder cancer. MAGEA6 protein is intracellular in location and showed direct interaction (physical association) with secreted (intracellular) protein S100A9, CTF1, GFOD1, and APO4. MAGEA6 showed interaction with intracellular proteins such as TULP3, EXOC5, LSM2, and others. Similarly, MAGEA3 showed interaction with secreted intracellular protein S100A9, along with other intracellular proteins. Genomic association revealed a direct correlation of MAGEA3 and MAGEA6 with calcium-binding protein S100A9 (Figure [9A](#page-11-1),B). To further understand the association between MAGEA3 and MAGEA6 with S100A9, we explored the OncoDB cancer database, a large-scale multiomics database, and performed pairwise gene expression correlation analysis between MAGEA3 and MAGEA6 and S100A9 in bladder cancer patients (Figure [10\)](#page-12-0). S100A9 is a secretory protein, and its expression is positively associated with elevated levels of MAGEA3 and MAGEA6 proteins [\[74](#page-17-10)[,75\]](#page-17-11). An overexpression of S100A9 is associated with stage progression, invasion, metastasis, and poor survival in bladder cancer patients [\[76\]](#page-17-12). S100A9 and MAGEA family members may be cooperative oncogenes, given our findings.



ditional clinical validation in bladder cancer patients is required to justify the above ra-

<span id="page-11-0"></span>ditional clinical validation in bladder cancer patients is required to justify the above ra-

Figure 8. Kaplan–Meier plots for overall survival in bladder cancer. These curves were g using the TCGA bladder cancer database of  $n = 408$  patients. The X-axis shows the overall survival in days and the Y-axis shows the overall survival rate. ings.

<span id="page-11-1"></span>

Figure 9. Gene interaction of (A) MAGEA3 and (B) MAGEA6 in bladder cancer. The figure shows the physical association and gene regulatory relationships. The assorted colors of the nodes show the physical association and gene regulatory relationships. The assorted colors of the nodes show the predicted location of the proteins.

<span id="page-12-0"></span>

**Figure 10.** Plot pair-wise gene expression correlation analysis between MAGEA6, with TRM28, and MAGEA28. MAGEA3 showed its correlation with S100A9 in BLCA. The X and Y-axes of the graph show log2 transcript per million (TPM).

# **7. MAGEA Family as Therapeutic Target in Bladder Cancer**

The basic potential strategies for MAGEA-targeted therapy in bladder cancer include immunotherapy against MAGE epitopes, the interruption of MAGEA–partner interactions, and the manipulation of regulatory pathway(s) affecting MAGE function. The role of MAGEA proteins had been established and demonstrated as therapeutic targets in multiple myeloma (MM) [\[77,](#page-17-13)[78\]](#page-17-14) and esophageal cancer [\[79\]](#page-17-15). The first two approaches might have huge benefits because of the limited expression of MAGE proteins in normal tissues [\(https://www.proteinatlas.org](https://www.proteinatlas.org) (accessed on 19 October 2023). The development of small molecule inhibitors interacting with MAGEA protein could have a lesser effect on somatic tissues and therefore minimize side effects [\[50,](#page-16-9)[80](#page-17-16)[–82\]](#page-17-17). The immune-based approaches are preferable due to the strong natural immunogenicity of MAGEA proteins coupled with the fact that germ cells do not express MHC class I antigens [\[30,](#page-15-25)[83–](#page-17-18)[85\]](#page-17-19). Hence, MAGEAtargeted vaccines should not elicit an autoimmune response in the testis. To date, there are approximately 47 oncologic clinical trials (including five involving bladder cancer) with a major focus on MAGE proteins. The clinical trials focusing on MAGEA proteins in bladder cancer are summarized in Table [1.](#page-13-0)



## <span id="page-13-0"></span>**Table 1.** Ongoing clinical trials of MAGEA family in bladder cancer.

Advancements in understanding the biology of MAGEA proteins are still ongoing; in particular, the synthesis of stable peptide inhibitors is a major breakthrough in the use of MAGEA–protein interactions [\[92–](#page-18-2)[94\]](#page-18-3). The hydrocarbon cross-linker stable peptides can be synthesized for targeting surface proteins, protease resistance, and cell permeability [\[95\]](#page-18-4). There is also a growing list of small molecules that have been shown to be effective in inhibiting protein–protein interactions, including MAGEA function. For example, a study by Bhatia et al. (2011) targeted the interaction between the RBCC domain of KAP1 with MAGE proteins [\[96\]](#page-18-5).

MAGEA proteins were the first human tumor-associated antigens identified at the molecular level. These proteins are more frequently expressed in the majority of tumors and are recognized as more potent and promising immunotherapeutic targets. The expression of type I MAGEs is typically restricted to the testis except in various cancers, where abnormal expression defined the term cancer–testis antigen. It is not just genomic dysregulation that induces the aberrant expression of MAGEs; interestingly, MAGEs contribute actively to tumorigenesis. Although several studies have associated MAGEs (including MAGEA2, MAGEA3, MAGEA6, and MAGEA9 expression) with pro-tumorigenic activities, such as p53 dysregulation, enhanced tumor proliferation, or the maintenance of cancer-stem-celllike characteristics [\[97](#page-18-6)[,98\]](#page-18-7), their definitive functions are not still fully understood in the context of bladder cancer.

### **8. Conclusions and Future Directions**

An examination and study of the human bladder cancer database has uncovered the MAGEA family and identified its diverse cellular functions, though this is just the beginning stages of understanding how the MAGEA family contributes to normal physiological processes versus the pathogenesis of bladder cancer. As highlighted in this review, the indepth genomic alteration, molecular structure, genetic interaction, and signaling cascades of the MAGEA family will provide further insight into its molecular role during bladder cancer pathogenesis. MAGEA genes can promote tumor progression through various mechanisms, such as through the activation of androgen receptor (AR), p53 inactivation, and an increase in oxidative phosphorylation (OXPHOS), which is overrepresented in bladder cancer and eventually contributes to highly aggressive and metastatic disease states in bladder cancer patients. Therefore, the MAGEA family has been considered as potential targets for bladder cancer treatment.

The high expression of MAGEA family members in bladder cancer may increase the propensity of proliferation and invasiveness during disease progression. This relationship makes MAGEA family members potential targets for diagnostic biomarkers, especially for muscle-invasive bladder cancer. Due to their highly antigenic properties and other diverse roles during cancer progression, the aberrant expression of MAGEA family, especially MAGEA3 and MAGEA6, may serve as prognostic biomarkers for poor outcomes in bladder cancer patients. Based on the genomic alteration profile of MAGEA proteins, it was speculated that mutation in the protein P262R may change the protein structural configuration which could attract other hormones and proteins facilitating malignant progression. MAGEA proteins may serve as a hormone receptor coregulators and trigger the pathogenic response. More mechanistic studies of the MAGEA family will facilitate the development of targeted therapies.

MAGEA-targeted approaches could be effective in inhibiting or even eliminating MAGEA cancer-promoting activities in various human cancers. Based on the current understanding of MAGEA proteins and their expression in germ cells, targeted therapy may offer a high degree of specificity with minimal side-effects in clinical settings. Thus, MAGEA family seems to be highly attractive area of research based on recent advancements in molecular biology. Additionally, innovative technologies are now available to enhance the immunotherapeutic responses and to target protein–protein interactions, which may be relevant for MAGEA proteins. Therefore, insights into the role of MAGEA proteins at the molecular level, and their association with cancer, should provide exciting new opportunities for exploitation and therapeutic intervention. In summary, this review emphasizes the importance of MAGEA family proteins in bladder cancer.

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#### **References**

- <span id="page-14-0"></span>1. Zhang, Y.; Rumgay, H.; Li, M.; Yu, H.; Pan, H.; Ni, J. The global landscape of bladder cancer incidence and mortality in 2020 and projections to 2040. *J. Glob. Health* **2023**, *13*, 04109. [\[CrossRef\]](https://doi.org/10.7189/jogh.13.04109) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/37712386)
- <span id="page-14-1"></span>2. Schafer, E.J.; Jemal, A.; Wiese, D.; Sung, H.; Kratzer, T.B.; Islami, F.; Dahut, W.L.; Knudsen, K.E. Disparities and Trends in Genitourinary Cancer Incidence and Mortality in the USA. *Eur. Urol.* **2023**, *84*, 117–126. [\[CrossRef\]](https://doi.org/10.1016/j.eururo.2022.11.023) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/36566154)
- 3. Tran, L.; Xiao, J.F.; Agarwal, N.; Duex, J.E.; Theodorescu, D. Advances in bladder cancer biology and therapy. *Nat. Rev. Cancer* **2021**, *21*, 104–121. [\[CrossRef\]](https://doi.org/10.1038/s41568-020-00313-1) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/33268841)
- <span id="page-14-2"></span>4. Lenis, A.T.; Lec, P.M.; Chamie, K.; Mshs, M.D. Bladder Cancer: A Review. *JAMA* **2020**, *324*, 1980–1991. [\[CrossRef\]](https://doi.org/10.1001/jama.2020.17598) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/33201207)
- <span id="page-14-3"></span>5. Chang, S.S.; Bochner, B.H.; Chou, R.; Dreicer, R.; Kamat, A.M.; Lerner, S.P.; Lotan, Y.; Meeks, J.J.; Michalski, J.M.; Morgan, T.M.; et al. Treatment of Non-Metastatic Muscle-Invasive Bladder Cancer: AUA/ASCO/ASTRO/SUO Guideline. *J. Urol.* **2017**, *198*, 552–559. [\[CrossRef\]](https://doi.org/10.1016/j.juro.2017.04.086)
- <span id="page-14-4"></span>6. Witjes, J.A.; Bruins, H.M.; Cathomas, R.; Compérat, E.M.; Cowan, N.C.; Gakis, G.; Hernández, V.; Linares Espinós, E.; Lorch, A.; Neuzillet, Y.; et al. European Association of Urology Guidelines on Muscle-invasive and Metastatic Bladder Cancer: Summary of the 2020 Guidelines. *Eur. Urol.* **2021**, *79*, 82–104. [\[CrossRef\]](https://doi.org/10.1016/j.eururo.2020.03.055)
- <span id="page-14-5"></span>7. DeGeorge, K.C.; Holt, H.R.; Hodges, S.C. Bladder Cancer: Diagnosis and Treatment. *Am. Fam. Physician* **2017**, *96*, 507–514.
- <span id="page-14-6"></span>8. Babatunde, K.A.; Najafi, A.; Salehipour, P.; Modarressi, M.H.; Mobasheri, M.B. Cancer/Testis genes in relation to sperm biology and function. *Iran. J. Basic Med. Sci.* **2017**, *20*, 967–974. [\[CrossRef\]](https://doi.org/10.22038/ijbms.2017.9259)
- 9. Kulkarni, P.; Shiraishi, T.; Rajagopalan, K.; Kim, R.; Mooney, S.M.; Getzenberg, R.H. Cancer/testis antigens and urological malignancies. Nature reviews. *Urology* **2012**, *9*, 386–396. [\[CrossRef\]](https://doi.org/10.1038/nrurol.2012.117)
- <span id="page-14-7"></span>10. Whitehurst, A.W. Cause, and consequence of cancer/testis antigen activation in cancer. *Annu. Rev. Pharmacol. Toxicol.* **2014**, *54*, 251–272. [\[CrossRef\]](https://doi.org/10.1146/annurev-pharmtox-011112-140326)
- <span id="page-14-8"></span>11. Nin, D.S.; Deng, L.W. Biology of Cancer-Testis Antigens and Their Therapeutic Implications in Cancer. *Cells* **2023**, *12*, 926. [\[CrossRef\]](https://doi.org/10.3390/cells12060926) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/36980267)
- 12. Jay, A.; Reitz, D.; Namekawa, S.H.; Heyer, W.D. Cancer testis antigens and genomic instability: More than immunology. *DNA Repair* **2021**, *108*, 103214. [\[CrossRef\]](https://doi.org/10.1016/j.dnarep.2021.103214)
- 13. Hikmet, F.; Rassy, M.; Backman, M.; Méar, L.; Mattsson, J.S.M.; Djureinovic, D.; Botling, J.; Brunnström, H.; Micke, P.; Lindskog, C. Expression of cancer–testis antigens in the immune microenvironment of non-small cell lung cancer. *Mol. Oncol.* **2023**, *17*, 2603–2617. [\[CrossRef\]](https://doi.org/10.1002/1878-0261.13474) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/37341056)
- 14. Kulkarni, P.; Uversky, V.N. Cancer/Testis Antigens: "Smart" Biomarkers for Diagnosis and Prognosis of Prostate and Other Cancers. *Int. J. Mol. Sci.* **2017**, *18*, 740. [\[CrossRef\]](https://doi.org/10.3390/ijms18040740) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/28362316)
- <span id="page-15-0"></span>15. Yao, J.; Caballero, O.L.; Yung, W.K.; Weinstein, J.N.; Riggins, G.J.; Strausberg, R.L.; Zhao, Q. Tumor subtype-specific cancer-testis antigens as potential biomarkers and immunotherapeutic targets for cancers. *Cancer Immunol. Res.* **2014**, *2*, 371–379. [\[CrossRef\]](https://doi.org/10.1158/2326-6066.CIR-13-0088)
- <span id="page-15-1"></span>16. Ren, S.; Zhang, Z.; Li, M.; Wang, D.; Guo, R.; Fang, X.; Chen, F. Cancer testis antigen subfamilies: Attractive targets for therapeutic vaccine (Review). *Int. J. Oncol.* **2023**, *62*, 71. [\[CrossRef\]](https://doi.org/10.3892/ijo.2023.5519)
- <span id="page-15-2"></span>17. Gordeeva, O. Cancer-testis antigens: Unique cancer stem cell biomarkers and targets for cancer therapy. *Semin. Cancer Biol.* **2018**, *53*, 75–89. [\[CrossRef\]](https://doi.org/10.1016/j.semcancer.2018.08.006)
- <span id="page-15-3"></span>18. Hofmann, O.; Caballero, O.L.; Stevenson, B.J.; Chen, Y.T.; Cohen, T.; Chua, R.; Maher, C.A.; Panji, S.; Schaefer, U.; Kruger, A.; et al. Genome-wide analysis of cancer/testis gene expression. *Proc. Natl. Acad. Sci. USA* **2008**, *105*, 20422–20427. [\[CrossRef\]](https://doi.org/10.1073/pnas.0810777105)
- <span id="page-15-4"></span>19. Zendman, A.J.; Ruiter, D.J.; Van Muijen, G.N. Cancer/testis-associated genes: Identification, expression profile, and putative function. *J. Cell. Physiol.* **2003**, *194*, 272–288. [\[CrossRef\]](https://doi.org/10.1002/jcp.10215)
- <span id="page-15-5"></span>20. Sang, M.; Wang, L.; Ding, C.; Zhou, X.; Wang, B.; Wang, L.; Lian, Y.; Shan, B. Melanoma-associated antigen genes—An update. *Cancer Lett.* **2011**, *302*, 85–90. [\[CrossRef\]](https://doi.org/10.1016/j.canlet.2010.10.021)
- <span id="page-15-6"></span>21. Weon, J.L.; Potts, P.R. The MAGE protein family and cancer. *Curr. Opin. Cell Biol.* **2015**, *37*, 1–8. [\[CrossRef\]](https://doi.org/10.1016/j.ceb.2015.08.002) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/26342994)
- <span id="page-15-7"></span>22. Lian, Y.; Meng, L.; Ding, P.; Sang, M. Epigenetic regulation of MAGE family in human cancer progression-DNA methylation, histone modification, and non-coding RNAs. *Clin. Epigenetics* **2018**, *10*, 115. [\[CrossRef\]](https://doi.org/10.1186/s13148-018-0550-8) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/30185218)
- <span id="page-15-8"></span>23. Sang, M.; Lian, Y.; Zhou, X.; Shan, B. MAGE-A family: Attractive targets for cancer immunotherapy. *Vaccine* **2011**, *29*, 8496–8500. [\[CrossRef\]](https://doi.org/10.1016/j.vaccine.2011.09.014) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/21933694)
- <span id="page-15-9"></span>24. Das, B.; Senapati, S. Immunological and functional aspects of MAGEA3 cancer/testis antigen. *Adv. Protein Chem. Struct. Biol.* **2021**, *125*, 121–147. [\[CrossRef\]](https://doi.org/10.1016/bs.apcsb.2020.08.001) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/33931137)
- <span id="page-15-10"></span>25. Newman, J.A.; Cooper, C.D.; Roos, A.K.; Aitkenhead, H.; Oppermann, U.C.; Cho, H.J.; Osman, R.; Gileadi, O. Structures of Two Melanoma-Associated Antigens Suggest Allosteric Regulation of Effector Binding. *PLoS ONE* **2016**, *11*, e0148762. [\[CrossRef\]](https://doi.org/10.1371/journal.pone.0148762) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/26910052)
- <span id="page-15-11"></span>26. Muscatelli, F.; Walker, A.P.; De Plaen, E.; Stafford, A.N.; Monaco, A.P. Isolation, and characterization of a MAGE gene family in the Xp21.3 region. *Proc. Natl. Acad. Sci. USA* **1995**, *92*, 4987–4991. [\[CrossRef\]](https://doi.org/10.1073/pnas.92.11.4987)
- <span id="page-15-12"></span>27. Florke Gee, R.R.; Chen, H.; Lee, A.K.; Daly, C.A.; Wilander, B.A.; Fon Tacer, K.; Potts, P.R. Emerging roles of the MAGE protein family in stress response pathways. *J. Biol. Chem.* **2020**, *295*, 16121–16155. [\[CrossRef\]](https://doi.org/10.1074/jbc.REV120.008029)
- <span id="page-15-13"></span>28. Schäfer, P.; Paraschiakos, T.; Windhorst, S. Oncogenic activity and cellular functionality of melanoma associated antigen A3. *Biochem. Pharmacol.* **2021**, *192*, 114700. [\[CrossRef\]](https://doi.org/10.1016/j.bcp.2021.114700)
- <span id="page-15-14"></span>29. Alsalloum, A.; Shevchenko, J.A.; Sennikov, S. The Melanoma-Associated Antigen Family A (MAGE-A): A Promising Target for Cancer Immunotherapy? *Cancers* **2023**, *15*, 1779. [\[CrossRef\]](https://doi.org/10.3390/cancers15061779)
- <span id="page-15-25"></span>30. Schooten, E.; Di Maggio, A.; van Bergen en Henegouwen, P.M.P.; Kijanka, M.M. MAGE-A antigens as targets for cancer immunotherapy. *Cancer Treat. Rev.* **2018**, *67*, 54–62. [\[CrossRef\]](https://doi.org/10.1016/j.ctrv.2018.04.009)
- <span id="page-15-15"></span>31. Kufer, P.; Zippelius, A.; Lutterbüse, R.; Mecklenburg, I.; Enzmann, T.; Montag, A.; Weckermann, D.; Passlick, B.; Prang, N.; Reichardt, P.; et al. Heterogeneous expression of MAGE-A genes in occult disseminated tumor cells: A novel multimarker reverse transcription-polymerase chain reaction for diagnosis of micrometastatic disease. *Cancer Res.* **2002**, *62*, 251–261. [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/11782385)
- <span id="page-15-16"></span>32. De Plaen, E.; Arden, K.; Traversari, C.; Gaforio, J.J.; Szikora, J.P.; De Smet, C.; Brasseur, F.; van der Bruggen, P.; Lethé, B.; Lurquin, C.; et al. Structure, chromosomal localization, and expression of 12 genes of the MAGE family. *Immunogenetics* **1994**, *40*, 360–369. [\[CrossRef\]](https://doi.org/10.1007/BF01246677) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/7927540)
- <span id="page-15-17"></span>33. Katsura, Y.; Satta, Y. Evolutionary history of the cancer immunity antigen MAGE gene family. *PLoS ONE* **2011**, *6*, e20365. [\[CrossRef\]](https://doi.org/10.1371/journal.pone.0020365)
- <span id="page-15-18"></span>34. Sigalotti, L.; Covre, A.; Zabierowski, S.; Himes, B.; Colizzi, F.; Natali, P.G.; Herlyn, M.; Maio, M. Cancer testis antigens in human melanoma stem cells: Expression, distribution, and methylation status. *J. Cell. Physiol.* **2008**, *215*, 287–291. [\[CrossRef\]](https://doi.org/10.1002/jcp.21380) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/18205182)
- <span id="page-15-19"></span>35. Jakobsen, M.K.; Traynor, S.; Stæhr, M.; Duijf, P.G.; Nielsen, A.Y.; Terp, M.G.; Pedersen, C.B.; Guldberg, P.; Ditzel, H.J.; Gjerstorff, M.F. The Cancer/Testis Antigen Gene VCX2 Is Rarely Expressed in Malignancies but Can Be Epigenetically Activated Using DNA Methyltransferase and Histone Deacetylase Inhibitors. *Front. Oncol.* **2020**, *10*, 584024. [\[CrossRef\]](https://doi.org/10.3389/fonc.2020.584024) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/33634013)
- <span id="page-15-20"></span>36. Monte, M.; Simonatto, M.; Peche, L.Y.; Bublik, D.R.; Gobessi, S.; Pierotti, M.A.; Rodolfo, M.; Schneider, C. MAGE-A tumor antigens target p53 transactivation function through histone deacetylase recruitment and confer resistance to chemotherapeutic agents. *Proc. Natl. Acad. Sci. USA* **2006**, *103*, 11160–11165. [\[CrossRef\]](https://doi.org/10.1073/pnas.0510834103) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/16847267)
- <span id="page-15-21"></span>37. Poojary, M.; Jishnu, P.V.; Kabekkodu, S.P. Prognostic Value of Melanoma-Associated Antigen-A (MAGE-A) Gene Expression in Various Human Cancers: A Systematic Review and Meta-analysis of 7428 Patients and 44 Studies. *Mol. Diagn. Ther.* **2020**, *24*, 537–555. [\[CrossRef\]](https://doi.org/10.1007/s40291-020-00476-5) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/32548799)
- <span id="page-15-22"></span>38. Craig, A.J.; Garcia-Lezana, T.; Ruiz de Galarreta, M.; Villacorta-Martin, C.; Kozlova, E.G.; Martins-Filho, S.N.; von Felden, J.; Ahsen, M.E.; Bresnahan, E.; Hernandez-Meza, G.; et al. Transcriptomic characterization of cancer-testis antigens identifies MAGEA3 as a driver of tumor progression in hepatocellular carcinoma. *PLoS Genet.* **2021**, *17*, e1009589. [\[CrossRef\]](https://doi.org/10.1371/journal.pgen.1009589)
- <span id="page-15-23"></span>39. Gao, X.; Chen, G.; Cai, H.; Wang, X.; Song, K.; Liu, L.; Qiu, T.; He, Y. Aberrantly enhanced melanoma-associated antigen (MAGE)-A3 expression facilitates cervical cancer cell proliferation and metastasis via actuating Wnt signaling pathway. *Biomed. Pharmacother.* **2020**, *122*, 109710. [\[CrossRef\]](https://doi.org/10.1016/j.biopha.2019.109710)
- <span id="page-15-24"></span>40. Müller-Richter, U.D.; Dowejko, A.; Reuther, T.; Kleinheinz, J.; Reichert, T.E.; Driemel, O. Analysis of expression profiles of MAGE-A antigens in oral squamous cell carcinoma cell lines. *Head Face Med.* **2009**, *5*, 10. [\[CrossRef\]](https://doi.org/10.1186/1746-160X-5-10)
- <span id="page-16-0"></span>41. Sakurai, T.; Itoh, K.; Higashitsuji, H.; Nagao, T.; Nonoguchi, K.; Chiba, T.; Fujita, J. A cleaved form of MAGE-A4 binds to Miz-1 and induces apoptosis in human cells. *J. Biol. Chem.* **2004**, *279*, 15505–155014. [\[CrossRef\]](https://doi.org/10.1074/jbc.M310437200) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/14739298)
- <span id="page-16-1"></span>42. Pineda, C.T.; Ramanathan, S.; Fon Tacer, K.; Weon, J.L.; Potts, M.B.; Ou, Y.H.; White, M.A.; Potts, P.R. Degradation of AMPK by a cancer-specific ubiquitin ligase. *Cell* **2015**, *160*, 715–728. [\[CrossRef\]](https://doi.org/10.1016/j.cell.2015.01.034) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/25679763)
- <span id="page-16-2"></span>43. Shaw, R.J. Tumor Metabolism: MAGE-A Proteins Help TRIM Turn Over AMPK. *Curr. Biol.* **2015**, *25*, R418–R420. [\[CrossRef\]](https://doi.org/10.1016/j.cub.2015.03.019) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/25989082)
- <span id="page-16-3"></span>44. Su, S.; Chen, X.; Geng, J.; Minges, J.T.; Grossman, G.; Wilson, E.M. Melanoma antigen-A11 regulates substrate-specificity of Skp2-mediated protein degradation. *Mol. Cell. Endocrinol.* **2017**, *439*, 1–9. [\[CrossRef\]](https://doi.org/10.1016/j.mce.2016.10.006) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/27720894)
- <span id="page-16-4"></span>45. Caballero, O.L.; Zhao, Q.; Rimoldi, D.; Stevenson, B.J.; Svobodová, S.; Devalle, S.; Röhrig, U.F.; Pagotto, A.; Michielin, O.; Speiser, D.; et al. Frequent MAGE mutations in human melanoma. *PLoS ONE* **2010**, *5*, e12773, Erratum in *PLoS ONE* **2010**, *5*, 10.1371/annotation/3ee2788f-0c44-429f-85c3-d626c9fedc21. [\[CrossRef\]](https://doi.org/10.1371/journal.pone.0012773)
- <span id="page-16-5"></span>46. Hagiwara, Y.; Sieverling, L.; Hanif, F.; Anton, J.; Dickinson, E.R.; Bui, T.T.; Andreeva, A.; Barran, P.E.; Cota, E.; Nikolova, P.V. Consequences of point mutations in melanoma-associated antigen 4 (MAGE-A4) protein: Insights from structural and biophysical studies. *Sci. Rep.* **2016**, *6*, 25182. [\[CrossRef\]](https://doi.org/10.1038/srep25182)
- <span id="page-16-6"></span>47. Park, T.S.; Groh, E.M.; Patel, K.; Kerkar, S.P.; Lee, C.C.; Rosenberg, S.A. Expression of MAGE-A and NY-ESO-1 in Primary and Metastatic Cancers. *J. Immunother.* **2016**, *39*, 1–7. [\[CrossRef\]](https://doi.org/10.1097/CJI.0000000000000101)
- <span id="page-16-7"></span>48. Xie, C.; Subhash, V.V.; Datta, A.; Liem, N.; Tan, S.H.; Yeo, M.S.; Tan, W.L.; Koh, V.; Yan, F.L.; Wong, F.Y.; et al. Melanoma associated antigen (MAGE)-A3 promotes cell proliferation and chemotherapeutic drug resistance in gastric cancer. *Cell. Oncol.* **2016**, *39*, 175–186. [\[CrossRef\]](https://doi.org/10.1007/s13402-015-0261-5)
- <span id="page-16-8"></span>49. Duan, Z.; Duan, Y.; Lamendola, D.E.; Yusuf, R.Z.; Naeem, R.; Penson, R.T.; Seiden, M.V. Overexpression of MAGE/GAGE genes in paclitaxel/doxorubicin-resistant human cancer cell lines. *Clin. Cancer Res. Off. J. Am. Assoc. Cancer Res.* **2003**, *9*, 2778–2785.
- <span id="page-16-9"></span>50. Marcar, L.; Ihrig, B.; Hourihan, J.; Bray, S.E.; Quinlan, P.R.; Jordan, L.B.; Thompson, A.M.; Hupp, T.R.; Meek, D.W. MAGE-A Cancer/Testis Antigens Inhibit MDM2 Ubiquitylation Function and Promote Increased Levels of MDM4. *PLoS ONE* **2015**, *10*, e0127713. [\[CrossRef\]](https://doi.org/10.1371/journal.pone.0127713)
- <span id="page-16-10"></span>51. Nardiello, T.; Jungbluth, A.A.; Mei, A.; Diliberto, M.; Huang, X.; Dabrowski, A.; Andrade, V.C.; Wasserstrum, R.; Ely, S.; Niesvizky, R.; et al. MAGE-A inhibits apoptosis in proliferating myeloma cells through repression of Bax and maintenance of survivin. *Clin. Cancer Res. Off. J. Am. Assoc. Cancer Res.* **2011**, *17*, 4309–4319. [\[CrossRef\]](https://doi.org/10.1158/1078-0432.CCR-10-1820) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/21565982)
- <span id="page-16-11"></span>52. Bujas, T.; Marusic, Z.; Peric Balja, M.; Mijic, A.; Kruslin, B.; Tomas, D. MAGE-A3/4 and NY-ESO-1 antigens expression in metastatic esophageal squamous cell carcinoma. *Eur. J. Histochem. EJH* **2011**, *55*, e7. [\[CrossRef\]](https://doi.org/10.4081/ejh.2011.e7) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/21556122)
- <span id="page-16-12"></span>53. Patard, J.J.; Brasseur, F.; Gil-Diez, S.; Radvanyi, F.; Marchand, M.; François, P.; Abi-Aad, A.; Van Cangh, P.; Abbou, C.C.; Chopin, D.; et al. Expression of MAGE genes in transitional-cell carcinomas of the urinary bladder. *Int. J. Cancer* **1995**, *64*, 60–64. [\[CrossRef\]](https://doi.org/10.1002/ijc.2910640112) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/7665250)
- <span id="page-16-13"></span>54. Picard, V.; Bergeron, A.; Larue, H.; Fradet, Y. MAGE-A9 mRNA and protein expression in bladder cancer. *Int. J. Cancer* **2007**, *120*, 2170–2177. [\[CrossRef\]](https://doi.org/10.1002/ijc.22282)
- <span id="page-16-14"></span>55. Bergeron, A.; Picard, V.; LaRue, H.; Harel, F.; Hovington, H.; Lacombe, L.; Fradet, Y. High frequency of MAGE-A4 and MAGE-A9 expression in high-risk bladder cancer. *Int. J. Cancer* **2009**, *125*, 1365–1371. [\[CrossRef\]](https://doi.org/10.1002/ijc.24503)
- <span id="page-16-15"></span>56. Xylinas, E.; Cha, E.K.; Khani, F.; Kluth, L.A.; Rieken, M.; Volkmer, B.G.; Hautmann, R.; Küfer, R.; Chen, Y.T.; Zerbib, M.; et al. Association of oncofetal protein expression with clinical outcomes in patients with urothelial carcinoma of the bladder. *J. Urol.* **2014**, *191*, 830–841. [\[CrossRef\]](https://doi.org/10.1016/j.juro.2013.08.048)
- <span id="page-16-16"></span>57. Dyrskjøt, L.; Zieger, K.; Kissow Lildal, T.; Reinert, T.; Gruselle, O.; Coche, T.; Borre, M.; Ørntoft, T.F. Expression of MAGE-A3, NY-ESO-1, LAGE-1 and PRAME in urothelial carcinoma. *Br. J. Cancer* **2012**, *107*, 116–122. [\[CrossRef\]](https://doi.org/10.1038/bjc.2012.215)
- <span id="page-16-17"></span>58. Kocher, T.; Zheng, M.; Bolli, M.; Simon, R.; Forster, T.; Schultz-Thater, E.; Remmel, E.; Noppen, C.; Schmid, U.; Ackermann, D.; et al. Prognostic relevance of MAGE-A4 tumor antigen expression in transitional cell carcinoma of the urinary bladder: A tissue microarray study. *Int. J. Cancer* **2002**, *100*, 702–705. [\[CrossRef\]](https://doi.org/10.1002/ijc.10540)
- <span id="page-16-18"></span>59. Mengus, C.; Schultz-Thater, E.; Coulot, J.; Kastelan, Z.; Goluza, E.; Coric, M.; Spagnoli, G.C.; Hudolin, T. MAGE-A10 cancer/testis antigen is highly expressed in high-grade non-muscle-invasive bladder carcinomas. *Int. J. Cancer* **2013**, *132*, 2459–2463. [\[CrossRef\]](https://doi.org/10.1002/ijc.27914)
- 60. Mohsenzadegan, M.; Razmi, M.; Vafaei, S.; Abolhasani, M.; Madjd, Z.; Saeednejad Zanjani, L.; Sharifi, L. Co-expression of cancer-testis antigens of MAGE-A6 and MAGE-A11 is associated with tumor aggressiveness in patients with bladder cancer. *Sci. Rep.* **2022**, *12*, 599. [\[CrossRef\]](https://doi.org/10.1038/s41598-021-04510-2)
- <span id="page-16-20"></span>61. Lerut, E.; Van Poppel, H.; Joniau, S.; Gruselle, O.; Coche, T.; Therasse, P. Rates of MAGE-A3 and PRAME expressing tumors in FFPE tissue specimens from bladder cancer patients: Potential targets for antigen-specific cancer immunotherapeutics. *Int. J. Clin. Exp. Pathol.* **2015**, *8*, 9522–9532. [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/26464715)
- 62. Bar-Haim, E.; Paz, A.; Machlenkin, A.; Hazzan, D.; Tirosh, B.; Carmon, L.; Brenner, B.; Vadai, E.; Mor, O.; Stein, A.; et al. MAGE-A8 overexpression in transitional cell carcinoma of the bladder: Identification of two tumour-associated antigen peptides. *Br. J. Cancer* **2004**, *91*, 398–407. [\[CrossRef\]](https://doi.org/10.1038/sj.bjc.6601968) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/15213716)
- <span id="page-16-19"></span>63. Sharma, P.; Shen, Y.; Wen, S.; Bajorin, D.F.; Reuter, V.E.; Old, L.J.; Jungbluth, A.A. Cancer-testis antigens: Expression and correlation with survival in human urothelial carcinoma. *Clin. Cancer Res. Off. J. Am. Assoc. Cancer Res.* **2006**, *12*, 5442–5447. [\[CrossRef\]](https://doi.org/10.1158/1078-0432.CCR-06-0527) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/17000678)
- <span id="page-17-0"></span>64. Vogelstein, B.; Papadopoulos, N.; Velculescu, V.E.; Zhou, S.; Diaz, L.A., Jr.; Kinzler, K.W. Cancer genome landscapes. *Science* **2013**, *339*, 1546–1558. [\[CrossRef\]](https://doi.org/10.1126/science.1235122) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/23539594)
- <span id="page-17-1"></span>65. Chen, J.; Herlong, F.H.; Stroehlein, J.R.; Mishra, L. Mutations of Chromatin Structure Regulating Genes in Human Malignancies. *Curr. Protein Pept. Sci.* **2016**, *17*, 411–437. [\[CrossRef\]](https://doi.org/10.2174/1389203717666160122120008) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/26796307)
- <span id="page-17-2"></span>66. Kang, J.U.; Koo, S.H.; Jeong, T.E.; Kwon, K.C.; Park, J.W.; Jeon, C.H. Multitarget fluorescence in situ hybridization and melanoma antigen genes analysis in primary bladder carcinoma. *Cancer Genet. Cytogenet.* **2006**, *164*, 32–38. [\[CrossRef\]](https://doi.org/10.1016/j.cancergencyto.2005.06.006) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/16364760)
- <span id="page-17-3"></span>67. Karihtala, P.; Kilpivaara, O.; Porvari, K. Mutational signatures and their association with survival and gene expression in urological carcinomas. *Neoplasia* **2023**, *44*, 100933. [\[CrossRef\]](https://doi.org/10.1016/j.neo.2023.100933)
- <span id="page-17-4"></span>68. Jungbluth, A.A.; Busam, K.J.; Kolb, D.; Iversen, K.; Coplan, K.; Chen, Y.T.; Spagnoli, G.C.; Old, L.J. Expression of MAGE-antigens in normal tissues and cancer. *Int. J. Cancer* **2000**, *85*, 460–465. [\[CrossRef\]](https://doi.org/10.1002/(SICI)1097-0215(20000215)85:4%3C460::AID-IJC3%3E3.0.CO;2-N)
- <span id="page-17-5"></span>69. Nasrah, S.; Radi, A.; Daberkow, J.K.; Hummler, H.; Weber, S.; Seaayfan, E.; Kömhoff, M. MAGED2 Depletion Promotes Stress-Induced Autophagy by Impairing the cAMP/PKA Pathway. *Int. J. Mol. Sci.* **2023**, *24*, 13433. [\[CrossRef\]](https://doi.org/10.3390/ijms241713433)
- <span id="page-17-6"></span>70. Shannon, P.; Markiel, A.; Ozier, O.; Baliga, N.S.; Wang, J.T.; Ramage, D.; Amin, N.; Schwikowski, B.; Ideker, T. Cytoscape: A software environment for integrated models of biomolecular interaction networks. *Genome Res.* **2003**, *13*, 2498–2504. [\[CrossRef\]](https://doi.org/10.1101/gr.1239303)
- <span id="page-17-7"></span>71. Ottaviani, S.; Colau, D.; van der Bruggen, P.; van der Bruggen, P. A new MAGE-4 antigenic peptide recognized by cytolytic T lymphocytes on HLA-A24 carcinoma cells. *Cancer Immunol. Immunother. CII* **2006**, *55*, 867–872. [\[CrossRef\]](https://doi.org/10.1007/s00262-005-0053-2) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/16151806)
- <span id="page-17-8"></span>72. Pineda, C.T.; Potts, P.R. Oncogenic MAGEA-TRIM28 ubiquitin ligase downregulates autophagy by ubiquitinating and degrading AMPK in cancer. *Autophagy* **2015**, *11*, 844–846. [\[CrossRef\]](https://doi.org/10.1080/15548627.2015.1034420) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/25945414)
- <span id="page-17-9"></span>73. Tang, Z.; Wei, G.; Zhang, L.; Xu, Z. Signature microRNAs and long noncoding RNAs in laryngeal cancer recurrence identified using a competing endogenous RNA network. *Mol. Med. Rep.* **2019**, *19*, 4806–4818. [\[CrossRef\]](https://doi.org/10.3892/mmr.2019.10143) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/31059106)
- <span id="page-17-10"></span>74. Yasar, O.; Akcay, T.; Obek, C.; Turegun, F.A. Significance of S100A8, S100A9 and calprotectin levels in bladder cancer. *Scand. J. Clin. Lab. Investig.* **2017**, *77*, 437–441. [\[CrossRef\]](https://doi.org/10.1080/00365513.2017.1336567) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/28609200)
- <span id="page-17-11"></span>75. Verma, S.; Shankar, E.; Lin, S.; Singh, V.; Chan, E.R.; Cao, S.; Fu, P.; MacLennan, G.T.; Ponsky, L.E.; Gupta, S. Identification of Key Genes Associated with Progression and Prognosis of Bladder Cancer through Integrated Bioinformatics Analysis. *Cancers* **2021**, *13*, 5931. [\[CrossRef\]](https://doi.org/10.3390/cancers13235931) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/34885040)
- <span id="page-17-12"></span>76. Kim, W.T.; Kim, J.; Yan, C.; Jeong, P.; Choi, S.Y.; Lee, O.J.; Chae, Y.B.; Yun, S.J.; Lee, S.C.; Kim, W.J. S100A9 and EGFR gene signatures predict disease progression in muscle invasive bladder cancer patients after chemotherapy. *Ann. Oncol. Off. J. Eur. Soc. Med. Oncol.* **2014**, *25*, 974–979. [\[CrossRef\]](https://doi.org/10.1093/annonc/mdu037) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/24631944)
- <span id="page-17-13"></span>77. Atanackovic, D.; Hildebrandt, Y.; Jadczak, A.; Cao, Y.; Luetkens, T.; Meyer, S.; Kobold, S.; Bartels, K.; Pabst, C.; Lajmi, N.; et al. Cancer-testis antigens MAGE-C1/CT7 and MAGE-A3 promote the survival of multiple myeloma cells. *Haematologica* **2010**, *95*, 785–793. [\[CrossRef\]](https://doi.org/10.3324/haematol.2009.014464)
- <span id="page-17-14"></span>78. de Carvalho, F.; Vettore, A.L.; Colleoni, G.W. Cancer/Testis Antigen MAGE-C1/CT7: New target for multiple myeloma therapy. *Clin. Dev. Immunol.* **2012**, *2012*, 257695. [\[CrossRef\]](https://doi.org/10.1155/2012/257695)
- <span id="page-17-15"></span>79. Zhang, Y.; Zhang, Y.; Zhang, L. Expression of cancer-testis antigens in esophageal cancer and their progress in immunotherapy. *J. Cancer Res. Clin. Oncol.* **2019**, *145*, 281–291. [\[CrossRef\]](https://doi.org/10.1007/s00432-019-02840-3)
- <span id="page-17-16"></span>80. Srivastava, P.; Paluch, B.E.; Matsuzaki, J.; James, S.R.; Collamat-Lai, G.; Taverna, P.; Karpf, A.R.; Griffiths, E.A. Immunomodulatory action of the DNA methyltransferase inhibitor SGI-110 in epithelial ovarian cancer cells and xenografts. *Epigenetics* **2015**, *10*, 237–246. [\[CrossRef\]](https://doi.org/10.1080/15592294.2015.1017198)
- 81. Hartmann, S.; Brands, R.C.; Küchler, N.; Fuchs, A.; Linz, C.; Kübler, A.C.; Müller-Richter, U.D. Melanoma-associated antigen expression and the efficacy of tyrosine kinase inhibitors in head and neck cancer. *Oncol. Lett.* **2015**, *10*, 1211–1217. [\[CrossRef\]](https://doi.org/10.3892/ol.2015.3345)
- <span id="page-17-17"></span>82. Mitchell, G.; Pollack, S.M.; Wagner, M.J. Targeting cancer testis antigens in synovial sarcoma. *J. Immunother. Cancer* **2021**, *9*, e002072. [\[CrossRef\]](https://doi.org/10.1136/jitc-2020-002072) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/34083416)
- <span id="page-17-18"></span>83. Meng, X.; Sun, X.; Liu, Z.; He, Y. A novel era of cancer/testis antigen in cancer immunotherapy. *Int. Immunopharmacol.* **2021**, *98*, 107889. [\[CrossRef\]](https://doi.org/10.1016/j.intimp.2021.107889) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/34174699)
- 84. Scanlan, M.J.; Gure, A.O.; Jungbluth, A.A.; Old, L.J.; Chen, Y.T. Cancer/testis antigens: An expanding family of targets for cancer immunotherapy. *Immunol. Rev.* **2002**, *188*, 22–32. [\[CrossRef\]](https://doi.org/10.1034/j.1600-065X.2002.18803.x) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/12445278)
- <span id="page-17-19"></span>85. Duperret, E.K.; Liu, S.; Paik, M.; Trautz, A.; Stoltz, R.; Liu, X.; Ze, K.; Perales-Puchalt, A.; Reed, C.; Yan, J.; et al. A Designer Cross-reactive DNA Immunotherapeutic Vaccine that Targets Multiple MAGE-A Family Members Simultaneously for Cancer Therapy. *Clin. Cancer Res. Off. J. Am. Assoc. Cancer Res.* **2018**, *24*, 6015–6027. [\[CrossRef\]](https://doi.org/10.1158/1078-0432.CCR-18-1013) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/30262507)
- <span id="page-17-20"></span>86. Colombel, M.; Heidenreich, A.; Martinez-Pineiro, L.; Babjuk, M.; Korneyev, I.; Surcel, C.; Yakovlev, P.; Colombo, R.; Radziszewski, P.; Witjes, F.; et al. Perioperative chemotherapy in muscle-invasive bladder cancer: Overview and the unmet clinical need for alternative adjuvant therapy as studied in the MAGNOLIA trial. *Eur. Urol.* **2014**, *65*, 509–511. [\[CrossRef\]](https://doi.org/10.1016/j.eururo.2013.10.056) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/24268503)
- <span id="page-17-21"></span>87. Derré, L.; Cesson, V.; Lucca, I.; Cerantola, Y.; Valerio, M.; Fritschi, U.; Vlamopoulos, Y.; Burruni, R.; Legris, A.S.; Dartiguenave, F.; et al. Intravesical Bacillus Calmette Guerin Combined with a Cancer Vaccine Increases Local T-Cell Responses in Non-muscle-Invasive Bladder Cancer Patients. *Clin. Cancer Res.* **2017**, *23*, 717–725. [\[CrossRef\]](https://doi.org/10.1158/1078-0432.CCR-16-1189) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/27521445)
- <span id="page-17-22"></span>88. Morgan, R.A.; Dudley, M.E.; Yu, Y.Y.; Zheng, Z.; Robbins, P.F.; Theoret, M.R.; Wunderlich, J.R.; Hughes, M.S.; Restifo, N.P.; Rosenberg, S.A. High efficiency TCR gene transfer into primary human lymphocytes affords avid recognition of melanoma tumor antigen glycoprotein 100 and does not alter the recognition of autologous melanoma antigens. *J. Immunol.* **2003**, *171*, 3287–3295. [\[CrossRef\]](https://doi.org/10.4049/jimmunol.171.6.3287)
- 89. Johnson, L.A.; Morgan, R.A.; Dudley, M.E.; Cassard, L.; Yang, J.C.; Hughes, M.S.; Kammula, U.S.; Royal, R.E.; Sherry, R.M.; Wunderlich, J.R.; et al. Gene therapy with human and mouse T-cell receptors mediates cancer regression and targets normal tissues expressing cognate antigen. *Blood* **2009**, *114*, 535–546. [\[CrossRef\]](https://doi.org/10.1182/blood-2009-03-211714)
- <span id="page-18-0"></span>90. Robbins, P.F.; Morgan, R.A.; Feldman, S.A.; Yang, J.C.; Sherry, R.M.; Dudley, M.E.; Wunderlich, J.R.; Nahvi, A.V.; Helman, L.J.; Mackall, C.L.; et al. Tumor regression in patients with metastatic synovial cell sarcoma and melanoma using genetically engineered lymphocytes reactive with NY-ESO-1. *J. Clin. Oncol.* **2011**, *29*, 917–924. [\[CrossRef\]](https://doi.org/10.1200/JCO.2010.32.2537)
- <span id="page-18-1"></span>91. Sanderson, J.P.; Crowley, D.J.; Wiedermann, G.E.; Quinn, L.L.; Crossland, K.L.; Tunbridge, H.M.; Cornforth, T.V.; Barnes, C.S.; Ahmed, T.; Howe, K.; et al. Preclinical evaluation of an affinity-enhanced MAGE-A4-specific T-cell receptor for adoptive T-cell therapy. *Oncoimmunology* **2019**, *9*, 1682381. [\[CrossRef\]](https://doi.org/10.1080/2162402X.2019.1682381) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/32002290)
- <span id="page-18-2"></span>92. Marcar, L.; Maclaine, N.J.; Hupp, T.R.; Meek, D.W. Mage-A cancer/testis antigens inhibit p53 function by blocking its interaction with chromatin. *Cancer Res.* **2010**, *70*, 10362–10370. [\[CrossRef\]](https://doi.org/10.1158/0008-5472.CAN-10-1341) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/21056992)
- 93. Jia, Z.C.; Tian, Y.; Huang, Z.M.; Wang, J.X.; Fu, X.L.; Ni, B.; Wu, Y.Z. Identification of a new MAGE-A10 antigenic peptide presented by HLA-A\*0201 on tumor cells. *Cancer Biol. Ther.* **2011**, *11*, 395–400. [\[CrossRef\]](https://doi.org/10.4161/cbt.11.4.14100) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/21124073)
- <span id="page-18-3"></span>94. Roch, N.; Kutup, A.; Vashist, Y.; Yekebas, E.; Kalinin, V.; Izbicki, J.R. Coexpression of MAGE-A peptides, and HLA class I molecules in hepatocellular carcinoma. *Anticancer Res.* **2010**, *30*, 1617–1623. [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/20592351)
- <span id="page-18-4"></span>95. Chowdhury, S.M.; Du, X.; Tolić, N.; Wu, S.; Moore, R.J.; Mayer, M.U.; Smith, R.D.; Adkins, J.N. Identification of cross-linked peptides after click-based enrichment using sequential collision-induced dissociation and electron transfer dissociation tandem mass spectrometry. *Anal. Chem.* **2009**, *81*, 5524–5532. [\[CrossRef\]](https://doi.org/10.1021/ac900853k)
- <span id="page-18-5"></span>96. Bhatia, N.; Yang, B.; Xiao, T.Z.; Peters, N.; Hoffmann, M.F.; Longley, B.J. Identification of novel small molecules that inhibit protein-protein interactions between MAGE and KAP-1. *Arch. Biochem. Biophys.* **2011**, *508*, 217–221. [\[CrossRef\]](https://doi.org/10.1016/j.abb.2011.01.007)
- <span id="page-18-6"></span>97. Gjerstorff, M.F.; Harkness, L.; Kassem, M.; Frandsen, U.; Nielsen, O.; Lutterodt, M.; Møllgård, K.; Ditzel, H.J. Distinct GAGE and MAGE-A expression during early human development indicate specific roles in lineage differentiation. *Hum. Reprod.* **2008**, *23*, 2194–2201. [\[CrossRef\]](https://doi.org/10.1093/humrep/den262)
- <span id="page-18-7"></span>98. Lifantseva, N.; Koltsova, A.; Krylova, T.; Yakovleva, T.; Poljanskaya, G.; Gordeeva, O. Expression patterns of cancer-testis antigens in human embryonic stem cells and their cell derivatives indicate lineage tracks. *Stem Cells Int.* **2011**, *2011*, 795239. [\[CrossRef\]](https://doi.org/10.4061/2011/795239)

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