MAMMARY TUMOURS IN THE CAT Size matters, so early intervention saves lives

Joanna Morris

Epidemiology

Mammary neoplasia is the third most common tumour type affecting female cats after lymphoma and skin tumours, accounting for 17% of tumours.¹ Reported incidence is 25.4 per 100,000 female cats per year.¹ Although precise statistics are not available, the incidence of mammary tumours may vary globally depending on cultural accept-

ance of neutering policies. Neutering is less common in Scandinavia and some other parts of Europe than in the UK, for example.

Signalment

Mammary tumours occur in older female cats (mean age 10-12 years),²⁻⁴ and usually those that are entire.^{5,6} Mammary tumours are also reported in male cats (mean age 12.8 years),⁷ but this is rare, accounting for 1-5% of mammary tumours. Siamese cats and other Oriental breeds may be more at risk,⁶ with mammary tumours occurring at a younger age;4,8 however, domestic shorthair cats, being probably the most common cat breed, are also very frequently affected.

Aetiology

As in humans and dogs, hormonal fluctuations associated with repeated oestrous cycles may influence the development of mammary tumours in cats. This is supported by a case control study reporting that cats spayed before 1 year of age had a decreased risk of developing mammary cancer,⁹ and that intact cats were seven times overrepresented in a popula-

tion of cats with mammary tumours compared with a control group.¹⁰ However, cats spayed at less

than a year old do still develop mammary tumours so the effect of early neutering does not eliminate the risk of mammary tumours,⁸ and early studies reporting more tumours in intact cats may possibly have reflected neutering practices at the time rather than a true increased risk.¹¹

Other evidence to support a hormonal aetiology is that oestrogen and progesterone receptors are found in normal mammary tissues and benign tumours but are often lost in malignant tumours and metastases.^{12–17} In addition, exogenous progesterone administration to prevent pregnancy, or for behavioural aggression, causes tumour development (benign and malignant) in both male¹⁸ and female cats.¹⁹ A possible dose-related effect may occur, with increased risk of mammary carcinoma if given regularly rather than intermittently.¹⁰

There is no definitive evidence for a viral aetiology for mammary tumours in cats, although this was proposed in early literature, and obesity has not been implicated, unlike in dogs.

> Joanna S Morris BSc BVSc PhD DipECVIM-CA (Onc) FRCVS School of Veterinary Medicine, College of Medical, Veterinary and Life Sciences, University of Glasgow, Bearsden Road, Bearsden, Glasgow G61 1QH, UK Email: Joanna.Morris@glasgow.ac.uk

DOI: 10.1177/1098612X13483237 © ISFM and AAFP 2013

Practical relevance: Mammary tumours are among the most common neoplasms in both cats and dogs, but the prevalence of malignant histological types is far higher in cats (ratio of malignant:benign is at least 4:1).

Clinical challenges: The more aggressive nature of mammary neoplasia in cats poses challenges for management. Prognosis is affected by tumour size and, therefore, early recognition and treatment of mammary tumours is paramount. Although the primary tumour can be excised surgically, no studies have shown that chemotherapy significantly extends survival time; hence, metastatic spread remains an important clinical problem.

Patient group: Mammary tumours usually affect older female cats, mainly entire females. Siamese and Oriental breeds may be predisposed. Male cats can develop mammary neoplasia, but this is rare. Evidence base: This review summarises the current literature relating to aetiology, pathology, presentation, diagnosis, staging, treatment and prognosis of feline mammary tumours.

Early neutering does not completely eliminate the risk of mammary tumours in cats.





53



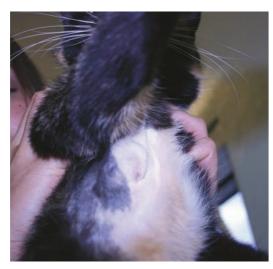


Figure 1 Mammary mass in the thoracic glands of an 11-year-old female entire (FE) domestic shorthair (DSH) cat

Presentation

Cats have four pairs of mammary glands (two thoracic and two abdominal) and, although any gland may be affected, some studies have reported a predisposition for mammary tumours in the caudal glands.^{11,20} Mammary tumours present as a single subcutaneous nodule or mass within the mammary glands (Figures 1 and 2),

which may be discrete and mobile, or attached to underlying tissues and possibly ulcerated in appearance (Figure 3). A few may appear cystic. It is difficult to distinguish benign from malignant nodules in cats, so all should be treated as potentially malignant. Multiple mammary masses within several glands are common (usually ipsilateral but occasionally bilateral) (Figure 4) and were reported in 60%of cats in one study.8 Sometimes, the true extent of the disease cannot be appreciated without clipping the fur. Drainage lymph nodes (inguinal or axillary) may also be visibly or palpably enlarged.

In particularly aggressive inflammatory carcinomas with extensive lymphatic Numerous non-neoplastic hyperplasias involvement, the glands dysplasias can occur in the mammary gland, and may be swollen, hot and although these are much less frequent in cats than in painful.⁴ This presen dogs, all may be mistaken for mammary tumours. These include ductal hyperplasia, duct ectasia (dilation), cysts, tation may be difficult and lobular hyperplasia, all of which may show to differentiate from associated focal fibrosis. Extensive, bilateral gland fibroadenomatous hyperdistension and swelling of the mammary glands may (fibroepithelial plasia also occur in fibroadenomatous hyperplasia, a hypertrophy, feline mammary hypertrophy), although the latter more commonly affects young cats.21



Figure 3 Ulcerated mammary mass in the second abdominal gland of a 21-year-old male neutered DSH cat

and

Differential diagnosis

type of lobular hyperplasia that may follow prolonged metoestrus, pseudopregnancy

or pregnancy, or use of exogenous

progesterone.11,21



Figure 2 Mammary mass in the thoracic glands and axillary lymph node of an 8-year-old FE DSH cat

Diagnosis

If a mammary mass is palpated, confirmation that it is neoplastic requires biopsy of tissue or a fine needle aspirate (FNA) for cytology. Since most feline mammary tumours are malignant, FNAs are more reliable than in dogs and are useful to confirm the diagnosis (Figure 5).





Figure 4 Bilateral mammary masses before (a) and after (b) clipping

The main histological type of mammary tumour in the cat is simple

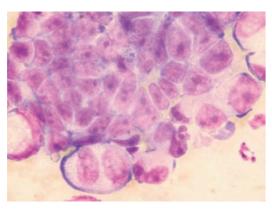


Figure 5 Cytology of feline mammary carcinoma, showing a raft of polygonal neoplastic epithelial cells with anisocytosis and anisokarvosis, some multinucleate cells and prominent nucleoli, often several per nucleus. Courtesy of Elizabeth Villiers

Most feline mammary tumours derive from glandular epithelium and all strictly are adenomas or adenocarcinomas, although the latter are often loosely referred to as carcinomas. Benign tumours are uncommon but, of these, fibroadenoma is most frequently reported, with simple adenoma or duct papilloma rarely seen. The main histological type of mammary tumour in the cat is simple adenocarcinoma derived from the luminal epithelium of the mammary ducts and alveoli (Figure 6). Complex or mixed tumours involving both luminal and myoepithelial cells are extremely infrequent in cats compared with dogs, although they may be associated with a better prognosis.^{22,23} In cats, carcinomas may be tubulopapillary, solid, cribriform or mucinous, although squamous cell carcinoma and mixed carcinosarcoma are also reported.24

Inflammatory mammary carcinoma, which has a particularly poor prognosis owing to an additional inflammatory component blocking the lymphatics and affecting lymph drainage and causing swollen, painful glands, has been reported in three cats with underlying highly malignant, papillary mammary carcinomas.²⁵

Staging

If a mammary tumour is suspected or confirmed, investigations should be carried out to determine the local extent and degree of spread throughout the body prior to surgical excision. As so few mammary masses are benign, and gross appearance alone is an unreliable basis on which to distinguish benign from malignant tumours, complete staging should be routine for all mammary masses. The WHO staging system is usually applied (Table 1);²⁶ however, with many tumours being noticed earlier and in less advanced stages, fewer larger tumours over 5 cm diameter are being reported. Measure-

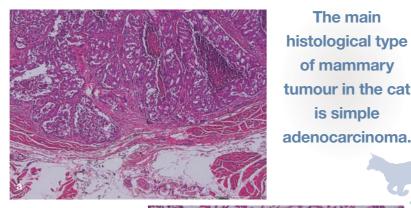
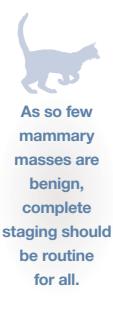
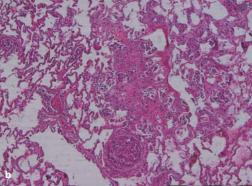


Figure 6 Histology of feline mammary simple adenocarcinoma. Low power views showing primary tumour invading muscle (a), and pulmonary metastasis (b) with tumour cells in blood vessels and lung tissue. Courtesy of Tim Scase





ment of the primary tumour with calipers is important since the size of the tumour influences the prognosis: those <3 cm diameter being associated with better survival rates than those >3 cm (see later).

Staging of confirmed tumours should include palpation and aspiration of local drainage lymph nodes, since more than a quarter of cats have regional metastasis at the time of diagnosis.²⁷ Involvement of multiple axillary lymph nodes is commonly detected by lymphangiography (58–75% cases) but a single inguinal lymph node predominates (84–94% cats).²⁸ Although the axillary and inguinal lymph nodes are those mainly reported as being affected in feline mammary neoplasia (80% cats), the sternal lymph node may also be involved (30% cats).27

Table 1 TNM and cli tumours		nical staging system for feline mammary		
Clinical stage	Tumour diameter (T)	Regional lymph node (N)	Distant metastasis (M)	
I.	<2 cm (T ₁)	Negative (N ₀)	Negative (M ₀)	
Ш	2–3 cm (T ₂)	Negative (N ₀)	Negative (M ₀)	
Ш	>3 cm (T ₃) ≤3 cm (T ₁ −T ₂)	Negative or positive ($N_0 \text{ or } N_1$) Positive (N_1)	Negative (M ₀) Negative (M ₀)	
IV	Any T	Any N	Positive (M ₁)	
Adapted from Owen ²⁶				



Figure 7 Enlarged left medial iliac lymph node chain with surrounding hyperechoic fat detected by abdominal ultrasound at routine staging of an 11-year-old female neutered (FN) Abyssinian with mammary carcinomas of the left caudal abdominal and right cranial thoracic glands

For more distant spread, three-view chest radiography (preferably performed under anaesthesia with inflated lungs) and abdominal ultrasonography should be carried out, since the most common sites of metastasis are the lungs, medial iliac lymph nodes and abdominal organs (Figure 7). Pulmonary metastases usually appear as a miliary pattern on thoracic radiographs, but pleural surfaces can also be affected and in some cases metastatic lung disease can cause pleural effusion (Figure 8). More uncommonly, metastasis to bone can be detected.

Advanced imaging (ie, computed tomography [CT]) of the lungs provides more accurate assessment of metastases (see 'case notes'), and should be used where there is any concern about the radiographic appearance of the lungs (Figure 9).

opacity within the thoracic cavity, with severe retraction of the lung fields from the dorsal wall and effacement of the cardiac silhouette and diaphragm, consistent with pleural effusion More than a quarter of cats

and dorsoventral (b)

of the cat imaged in Figure 7. These views show general increased

radiographs of the thorax

have regional metastasis at the time of diagnosis.



47)

Since most affected cats are elderly, haematology, biochemistry and urinalysis should also be performed to assess for concurrent disease. Paraneoplastic conditions are

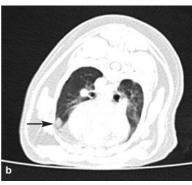


Figure 9 Left lateral radiograph (a) of a 12-year-old FN DSH cat with a mammary carcinoma of the second abdominal gland. A couple of faint, ill-defined soft tissue opacities superimposed on the cardiac silhouette, and suspicious for metastasis, were noted on this view (arrows) but not the right lateral view. CT of the chest confirmed the presence of a 2-3 mm hyperattenuating nodule in the middle right lung lobe and (b) a 4.8 mm hyperattenuating nodule in the caudal part of the left lung lobe (arrow)

rarely reported with mammary tumours, and feline leukaemia and feline virus immunodeficiency virus are not implicated in the aetiology. However, if further treatment, including chemotherapy is being considered, it is important to evaluate viral status since viral immunosuppression may influence whether treatment goes ahead.



Surgical treatment

The mainstay of treatment for mammary tumours is still surgical resection. The extent of surgery is influenced by the lymphatic drainage in the feline mammary gland (see box), as tumour cells spread readily beyond the primary site and complete excision should encompass all known drainage pathways.

Recommended approach

The recommendation based on drainage studies is to perform unilateral or bilateral mammary strips because of possible contact between individual glands and between left and right sides. While radiological imaging studies would suggest that this may not be necessary in every case, additional prognostic analyses do support the use of unilateral or bilateral mammary strips since the extent of surgery appears to make a significant difference to local recurrence/disease-free interval (DFI)³³ and survival time.³⁴ For bilateral strips a 2-week interval is recommended between surgeries although simultaneous bilateral mastectomy can also be performed (Figure 10). Tumour fixation to skin or abdominal fascia necessitates removal of these structures en bloc.³⁵

• Excision of lymph nodes The inguinal lymph node is embedded in the caudal mammary gland and is therefore removed along with the gland as part of a mammary strip. The axillary lymph node should be removed if enlarged or positive for tumour spread on FNA or biopsy, but there is no evidence that prophylactic removal extends survival.

◆ Concurrent ovariohysterectomy There is no evidence that ovariohysterectomy at the time of mastectomy has any benefit on survival or tumour recurrence,⁸ or effect on development of new tumours or carcinoma progression.¹⁰ It might, however, reduce the need for progestin therapy, which may be beneficial.

Figure 10 Bilateral mammary strip in a cat with mammary carcinoma. Courtesy of Kathryn Pratschke



Lymphatic drainage

Lymphatic drainage has been studied by injection of dyes and anatomical dissection of cadavers,^{29,30} and by radiological methods in live healthy cats;²⁸ the latter may be more accurate since dynamic blood pressure influences the direction of natural lymphatic flow. Most studies agree that the first and second (thoracic) glands drain cranially into the axillary lymph nodes; although cadaver dissections have shown that the second gland may drain caudally to the inguinal lymph node,29,30 this has not been seen by radiological methods. The third (abdominal) gland drains both cranially to axillary and caudally to inguinal lymph nodes, and the fourth gland drains just caudally to the inguinal lymph node. Direct drainage from the third and fourth abdominal glands to the medial iliac lymph node was reported in one case each:28 however, direct drainage of the first, second and third glands to the sternal gland has not been confirmed in the cat.28,30

Although connections between mammary glands and between left and right sides have been previously proposed, live imaging does not support this in the healthy cat.²⁸ Drainage may vary between normal glands and glands with mammary tumours, making it difficult to know the precise drainage pathway,³¹ and perhaps making a case for indirect lymphography for each patient to help define the drainage pattern and examine sentinel lymph nodes.³² Potentially this might enable more conservative resections to be performed. As tumour cells spread readily beyond the primary site, complete excision should encompass all known drainage pathways.

Chemotherapy

There is some data to show that chemotherapy may be effective on mammary cell lines in vitro,^{36–38} and that treatment of non-resectable disease in vivo with doxorubicin and cyclophosphamide may shrink tumour size in 50% cases and possibly increase survival (Table 2).^{38–40} The benefit of using chemotherapy as an adjunct to surgical excision of mammary tumours in cats, however, is still not clear (Table 3).

A large, multicentre study of 67 cats receiving adjunctive doxorubicin reported a median survival time of 448 days.41 Although there was no control group in the study, this survival time was deemed longer than historical controls and was similar to that in another study of 23 cats without a control group (460 days) combining adjunctive doxorubicin with the COX-2 inhibitor meloxicam.42 A further study of 73 cats, which included a control group of 36 cats undergoing surgical excision only, reported increased survival time and DFI for the cats receiving postoperative doxorubicin and cyclophosphamide (1406 versus 848 days [survival time] and 676 versus 372 days [DFI]);³⁴ however, the difference was not statistically significant.

Table 2	Effect of chemotherapy (doxorubicin) on mammary carcinomas (gross disease)					
Clinical stage	Number of cats	Treatment	Control group	Median survival (days)	Response	Reference
III or above	14	Doxorubicin (30 mg/m ² IV q2 weeks)	No	215 (30.8 weeks)	>50% response in 9/14	Stolwijk et al ³⁸
III or above	14	Doxorubicin (20–30 mg/m ² IV q3 weeks) + cyclophosphamide (100 mg/m ² PO for 3 days q3 weeks)	No	180	>50% response in 5/14	Jeglum et al ³⁹
III or above	14	Doxorubicin (25 mg/m ² IV) + cyclophosphamide (50 mg/m ² PO for 4 days q3 weeks)	No	90	>50% response in 7/14	Mauldin et al ⁴⁰

 Table 3
 Effect of chemotherapy (doxorubicin) as an adjunct to surgical removal of mammary tumours

Clinical stage	Number of cats	Treatment	Control group	Median survival (days)	Median DFI (days)	Reference
III or below	67	Surgery + doxorubicin*	Historical	448	255	Novosad et al ⁴¹
III or below	37	Surgery alone	Surgery alone	1406	372	McNeill et al34
	36	Surgery + doxorubicin [†] + cyclophosphamide [‡]		848	676	
III or below	23	Surgery + doxorubicin [§] + meloxicam [¶]	Historical	460	269	Borrego et al ⁴²

DFI = disease-free interval

*Doxorubicin 1 mg/kg IV q3 weeks

[†]Doxorubicin dose not given (four cases doxorubicin alone)

[‡]Cyclophosphamide dose not given

[§]Doxorubicin 1 mg/kg IV q3 weeks (one case had vincristine 0.7 mg/m² IV and 13 cases had cyclophosphamide 250 mg/m² IV, 1 week after doxorubicin)

[¶]Meloxicam 0.2 mg/kg PO on day of surgery, then 0.1 mg/kg q24h for 5 days, then 0.025 mg/kg

The benefit of using chemotherapy as an adjunct to surgical excision of mammary tumours in cats is still not clear.

It is possible that with greater numbers and more statistical power, a true benefit of aggressive chemotherapy may become apparent. Alternatively, a different approach with antiangiogenic metronomic (low dose) chemotherapy may prove effective, although low dose chemotherapy using vincristine, cyclophosphamide and methotrexate did not prevent recurrence or metastasis in one report.⁸

Other therapies

Although immunomodulators such as bacillus calmette-Guerin (BCG),⁴³ *Corynebacterium parvum*,⁴⁴ liposome-encapsulated muramyl tripeptide phosphatidylethanolamine (L- MTP-PE)⁴⁵ and oral levamisole⁴⁶ have been tried as intratumoural injections (BCG) or adjuncts to surgical excision of feline mammary tumours, none have proved successful in extending survival time or altering recurrence rate. There are no reports of using antioestrogens such as tamoxifen in cats, probably since most malignant feline mammary tumours lack oestrogen receptors and expected benefits would, therefore, appear to be minimal.

The small molecule inhibitors that target receptor tyrosine kinases (receptor tyrosine kinase inhibitors or RTKIs) are effective in the management of some types of veterinary cancers, particularly those with altered TK activity.⁴⁷ Imatinib and masitinib are well tolerated in cats;^{48–51} however, there is no information about their efficacy against feline mammary tumours. (Further discussion on the use of targeted therapies in cats, with a particular focus on the idiosyncracies of feline patients, is provided in an accompanying article in this special issue.)

Table 4 Prognostic factors for feline mammary tumours				
Factor	Details			
Tumour size*	Diameter <3 cm – median survival 21–24 months Diameter >3 cm – median survival 4–12 months			
Clinical stage [†]	Stage I – median survival 29 months Stage II – median survival 12.5 months Stage III – median survival 9 months Stage IV – median survival 1 month			
Surgical extent	Radical surgery (mammary strip) reduces recurrence rate compared with more conservative mastectomy			
Histopathological grade	Well differentiated – 100% survival at 1 year after surgery Poorly differentiated – 0% survival at 1 year after surgery			
Mitotic index	<2 mitotic figures per high power field give longer survival			
*MacEwen at al ³³ and Viste at al 5^2 ¹ Ito at al ⁵³				

*MacEwen et al³³ and Viste et al.⁵² ¹Ito et al⁵⁴

Prognosis

Prognosis is guarded for most cats with mammary tumours, with deaths mainly attributable to local recurrence or metastasis. The average time between detection and death is reported to be 10–12 months;^{20,35} however, as already mentioned, several factors affect prognosis of feline mammary tumours (Table 4).

Clinical parameters

Prognosis is very much related to the size of the tumour at initial presentation, with tumours of large volume (>27 cm³) or diameter (>3 cm) being associated with shorter survival times (4–12 months).^{8,52,53} The degree of spread at

Table 5 Comparison of mammary tumours in cats and dogs				
	Cats	Dogs		
Aetiology	Hormonal (oestrogen and progesterone)	Hormonal (oestrogen and progesterone) Obesity at a young age		
Signalment	Mean age 10–12 years Siamese/Oriental and DSH cats most affected Usually intact females Rare in males – aggressive disease when seen	Mean age 6–10 years Poodles, dachshunds and spaniels most affected Usually intact females Rare in males – more benign disease when seen		
Staging	Often metastatic spread	Metastatic spread less frequent		
Pathology	80–90% malignant Mainly simple adenocarcinomas	50% benign Complex/mixed benign tumours are common but carcinomas are usually of simple type		
Treatment	Radical mastectomy (strips) advised	Depends on size and site (gland affected) but regional mastectomy or strip advised for most carcinomas		
Chemotherapy	Several studies suggest at least some efficacy of doxorubicin against gross disease or metastasis	No conclusive evidence for efficacy of chemotherapy		
Prognostic factors	Size of tumour Clinical stage Histological grade of tumour Extent of surgery	Size of tumour Clinical stage Histological grade of tumour Histological type of tumour		

initial presentation (eg, regional lymph node metastasis) also dramatically affects prognosis,^{20,33,53} as does the extent of surgery carried out. Radical mastectomy produces a significantly longer DFL,^{33,53} and histological completeness of resection correlates with survival.²⁷ The cat's age may also influence prognosis,^{27,33} although more recent studies dispute this.⁵³

Histological parameters

A wealth of literature exists on histological markers detected by pathologists,⁵⁴ some of which influence prognosis. However, few are routinely used in diagnostic laboratories to offer helpful information to clinicians.

The most useful assessment by pathologists is histological grading, which is significantly related to both overall survival and DFI in univariate and multivariate analyses.55-57 Mitotic index also correlates with survival time,58 but although other proliferation markers such as Ki67,^{57,59} AgNOR⁶⁰ and PCNA⁶¹ may help determine high histological grade, they have not been shown to be independent prognostic markers. Similarly, although expression of hormone receptors (ER and PR) has been examined in feline carcinomas, receptor status does not correlate with overall survival and is not routinely assessed.^{14,15} The epidermal growth factor receptor 2 (Her2/neu) is variably overexpressed in feline carcinomas and, while one study has associated this with survival,⁶² more recent data has not.63 High COX-2 expression in mammary carcinomas has also been linked with poor prognosis,64 but this association is complicated by the fact that it also correlates with expression of the angiogenic factor vascular endothelial growth factor (VEGF), which itself is significantly correlated with overall survival.65 More recently, expression of the cytoplasmic protein kinase, AKT, which is activated by numerous receptor tyrosine kinases such as EGF or by loss of the tumour suppressor gene PTEN (phosphatase and tensin homologue), has been significantly associated with shorter DFI.66

KEY POINTS

- Mammary tumours in cats differ from those in dogs in terms of histological type and behaviour (Table 5).
- Early diagnosis is important for optimal management, as is a more aggressive surgical approach to treatment. In most cases adjuvant chemotherapy also needs to be considered.

Cleo, a 13-year-old FN DSH cat, presented with a 5 x 3 cm mammary mass in her right caudal abdominal gland. She had a short history of being pyrexic and inappetent before presentation, and had clinical signs of concurrent hyperthyroidism (palpable goitre) and an elevated heart rate (250 bpm).

Blood tests Blood tests for haematology, biochemistry and glucose were all within normal limits. Thyroxine (T_4) was also normal (41 nmol/l), although perhaps higher than might be expected for a cat with other illness.

Ultrasound and

echocardiography No significant abnormalities were detected on abdominal ultrasonography. Echocardiography detected mild changes of hypertrophic cardiomyopathy consistent with early primary disease or underlying hyperthyroidism.

Radiography Inflated chest radiographs (see right) revealed a soft tissue opacity in the cranial lung field (right lung on dorsoventral view) at the level of the third rib, and bony changes were noted on the right scapula.

Advanced imaging CT of the chest was performed to further evaluate the nodule in the lung. A high

resolution CT scan with lung field window confirmed the presence of an expansile, cystic lesion in the right lung field (see image a below). The waxing and waning pyrexia was thought to be linked to periodic necrosis within this expansile lung lesion. Metastatic nodules were noted at the dorsal periphery of the





Case notes

Cleo at presentation















right lung lobe (b) and lateral margin of the left lung lobe (c). An expansile lytic lesion with areas of calcification was observed in the dorsal scapular border and scapular spine, consistent with metastatic spread (d and e).

Treatment and outcome Owing to the metastatic spread to lung and scapula, radical surgical excision of the mass was not performed. Cleo was managed with meloxicam and buprenorphine for a few weeks but sadly was then euthanased due to bone pain, weight loss and inappetence.

Funding

The author received no specific grant from any funding agency in the public, commercial or not-for-profit sectors for the preparation of this article.

Conflict of interest

The author does not have any potential conflicts of interest to declare.

References

- 1 Schneider R, Dorn CR and Taylor DO. Factors influencing canine mammary cancer development and postsurgical survival. J Natl Cancer Inst 1969; 43: 1249–1261.
- 2 Misdorp W. Tumors of the mammary gland. In: Meuten DJ (ed). Tumors in domestic animals. Oxford: Blackwell, 2002, pp 575–606.
- Lana SE, Rutteman GR and Withrow S. Feline mammary tumors.
 In: Withrow SJ and Vail DM (eds). Small animal clinical oncology.
 4th ed. St Louis: Elsevier, 2007, pp 628–636.
- 4 Hahn KA and Adams WH. Feline mammary neoplasia: biological behaviour, diagnosis, and treatment alternatives. *Feline Pract* 1997; 25: 5–11.
- 5 Dorn CR, Taylor DO, Schneider R, Hibbard HH and Klauber MR. Survey of animal neoplasms in Alameda and Contra Costa Counties, California. II. Cancer morbidity in dogs and cats from Alameda County. J Natl Cancer Inst 1968; 40: 307–318.
- 6 Hayes HM Jr, Milne KL and Mandell CP. Epidemiological features of feline mammary carcinoma. Vet Rec 1981; 108: 476–479.
- 7 Skorupski KA, Overley B, Shofer FS, Goldschmidt MH, Miller CA and Sorenmo KU. Clinical characteristics of mammary carcinoma in male cats. J Vet Intern Med 2005; 19: 52–55.
- 8 Hayes AA and S Mooney. Feline mammary tumors. Vet Clin North Am Small Anim Pract 1985; 15: 513–520.
- 9 Overley B, Shofer FS, Goldschmidt MH, Sherer D and Sorenmo KU. Association between ovariohysterectomy and feline mammary carcinoma. *J Vet Intern Med* 2005; 19: 560–563.
- 10 Misdorp W, Romijn A and Hart AA. Feline mammary tumors: a case-control study of hormonal factors. *Anticancer Res* 1991; 11: 1793–1797.
- 11 Moore AS and Ogilvie GK. Mammary tumors. In: Ogilvie GK and Moore AS (eds). Feline oncology. A comprehensive guide to compassionate care. Trenton, NJ: Veterinary Learning Systems, 2001, pp 355–367.
- 12 de las Mulas JM, van Niel M, Millan Y, Blankenstein MA, van Mil F and Misdorp W. Immunohistochemical analysis of estrogen receptors in feline mammary gland benign and malignant lesions: comparison with biochemical assay. Domest Anim Endocrinol 2000; 18: 111–125.
- 13 Martin de las Mulas J, van Niel M, Millan Y, Ordas J, Blankenstein MA, van Mil F, et al. Progesterone receptors in normal, dysplastic and tumourous feline mammary glands. Comparison with oestrogen receptors status. *Res Vet Sci* 2002; 72: 153–161.
- 14 Millanta F, Calandrella M, Bari G, Niccolini M, Vannozzi I and Poli A. Comparison of steroid receptor expression in normal, dysplastic, and neoplastic canine and feline mammary tissues. *Res Vet Sci* 2005; 79: 225–232.
- 15 Millanta F, Calandrella M, Vannozzi I and Poli A. Steroid hormone receptors in normal, dysplastic and neoplastic feline

mammary tissues and their prognostic significance. *Vet Rec* 2006; 158: 821–824.

- 16 Morris JS, Nixon C, Bruck A, Nasir L, Morgan IM and Philbey AW. Immunohistochemical expression of TopBP1 in feline mammary neoplasia in relation to histological grade, Ki67, ERalpha and p53. Vet J 2008; 175: 218–226.
- 17 Cardazzo B, Zappulli V, Frassineti F, Patarnello T, Castagnaro M and Bargelloni L. Full-length sequence and expression analysis of estrogen receptor alpha mRNA in feline mammary tumors. *J Steroid Biochem Mol Biol* 2005; 96: 109–118.
- 18 Jacobs TM, Hoppe BR, Poehlmann CE, Ferracone JD and Sorenmo KU. Mammary adenocarcinomas in three male cats exposed to medroxyprogesterone acetate (1990–2006). *J Feline Med Surg* 2010; 12: 169–174.
- 19 Keskin A, Yilmazbas G, Yilmaz R, Ozyigit MO and Gumen A. Pathological abnormalities after long-term administration of medroxyprogesterone acetate in a queen. J Feline Med Surg 2009; 11: 518–521.
- 20 Zappulli V, De Zan G, Cardazzo B, Bargelloni L and Castagnaro M. Feline mammary tumours in comparative oncology. J Dairy Res 2005; 72: 98–106.
- 21 Gimenez F, Hecht S, Craig LE and Legendre AM. Early detection, aggressive therapy: optimizing the management of feline mammary masses. J Feline Med Surg 2010; 12: 214–224.
- 22 Seixas F, Palmeira C, Pires MA and Lopes C. Are complex carcinoma of the feline mammary gland and other invasive mammary carcinoma identical tumours? Comparison of clinicopathologic features, DNA ploidy and follow-up. *Res Vet Sci* 2008; 84: 428–433.
- 23 Seixas F, Pires MA and Lopes CA. Complex carcinomas of the mammary gland in cats: pathological and immunohistochemical features. *Vet J* 2008: 176: 210–215.
- 24 Misdorp W, Else RW, Hellmen E and Lipscomb TP. Histological classification of mammary tumors of the dog and the cat. Second Series. Washington DC: Armed Forces Institute of Pathology, 1999.
- 25 Perez-Alenza MD, Jimenez A, Nieto AI and Pena L. First description of feline inflammatory mammary carcinoma: clinicopathological and immunohistochemical characteristics of three cases. *Breast Cancer Res* 2004; 6: R300–307.
- 26 Owen LN (ed). TNM classification of tumours in domestic animals. Geneva: World Health Organization, 1980, p 53.
- 27 Weijer K and Hart AA. Prognostic factors in feline mammary carcinoma. J Natl Cancer Inst 1983; 70: 709–716.
- 28 Papadopoulou PL, Patsikas MN, Charitanti A, Kazakos GM, Papazoglou LG, Karayannopoulou M, et al. The lymph drainage pattern of the mammary glands in the cat: a lymphographic and computerized tomography lymphographic study. Anat Histol Embryol 2009; 38: 292–299.
- 29 Raharison F and Sautet J. Lymph drainage of the mammary glands in female cats. J Morphol 2006; 267: 292–299.
- 30 Raharison F and Sautet J. **The topography of the lymph vessels** of mammary glands in female cats. *Anat Histol Embryol* 2007; 36: 442–452.
- 31 Patsikas MN, Karayannopoulou M, Kaldrymidoy E, Papazoglou LG, Papadopoulou PL, Tzegas SI, et al. The lymph drainage of the neoplastic mammary glands in the bitch: a lymphographic study. *Anat Histol Embryol* 2006; 35: 228–234.
- 32 Patsikas MN, Papadopoulou PL, Charitanti A, Kazakos GM, Soultani CB, Tziris NE, et al. Computed tomography and radiographic indirect lymphography for visualization of mammary lymphatic vessels and the sentinel lymph node in normal cats.

Vet Radiol Ultrasound 2010; 51: 299-304.

- 33 MacEwen EG, Hayes AA, Harvey HJ, Patnaik AK, Mooney S. and Passe S. **Prognostic factors for feline mammary tumors.** *J Am Vet Med Assoc* 1984; 185: 201–204.
- 34 McNeill CJ, Sorenmo KU, Shofer FS, Gibeon L, Durham AC, Barber LG, et al. **Evaluation of adjuvant doxorubicin-based chemotherapy for the treatment of feline mammary carcinoma.** *J Vet Intern Med* 2009; 23: 123–129.
- 35 Lana SE, Rutteman GR and Withrow SJ. Tumors of the mammary gland. In: Withrow SJ and Vail DM (eds). Small animal clinical oncology. St Louis: Elsevier, 2007, pp 619–636.
- 36 Muleya JS, Nakaichi M, Taura Y, Yamaguchi R and Nakama S. In-vitro anti-proliferative effects of some anti-tumour drugs on feline mammary tumour cell lines. *Res Vet Sci* 1999; 66: 169–174.
- 37 Prop FJ, Weijer K, Spies J, Souw L, Peters K, Erich T, et al. Feline mammary carcinomas as a model for human breast cancer. I. Sensitivity of mammary tumor cells in culture to cytostatic drugs. A preliminary investigation of a predictive test. Anticancer Res 1986; 6: 989–994.
- 38 Stolwijk JA, Minke JM, Rutteman GR, Hoekstra J, Prop FJ and Misdorp W. Feline mammary carcinomas as a model for human breast cancer. II. Comparison of in vivo and in vitro adriamycin sensitivity. *Anticancer Res* 1989; 9: 1045–1048.
- 39 Jeglum KA, deGuzman E and Young KM. Chemotherapy of advanced mammary adenocarcinoma in 14 cats. J Am Vet Med Assoc 1985; 187: 157–160.
- 40 Mauldin GN, Matus RE, Patnaik AK, Bond BR and Mooney SC. Efficacy and toxicity of doxorubicin and cyclophosphamide used in the treatment of selected malignant tumors in 23 cats. J Vet Intern Med 1988: 2: 60–65.
- 41 Novosad CA, Bergman PJ, O'Brien M, McKnight JA, Charney SC, Selting KA, et al. Retrospective evaluation of adjunctive doxorubicin for the treatment of feline mammary gland adeno-carcinoma: 67 cases. J Am Anim Hosp Assoc 2006; 42: 110–120.
- 42 Borrego JF, Cartagena JC and Engel J. Treatment of feline mammary tumours using chemotherapy, surgery and a COX-2 inhibitor drug (meloxicam): a retrospective study of 23 cases (2002–2007)*. Vet Comp Oncol 2009; 7: 213–221.
- 43 Rutten VP, Misdorp W, Gauthier A, Estrada M, Mialot JP, Parodi AL, et al. Immunological aspects of mammary tumors in dogs and cats: a survey including own studies and pertinent literature. Vet Immunol Immunopathol 1990; 26: 211–225.
- 44 Parodi AL, Misdorp W, Mialot JP, Mialot M, Hart AA, Hurtrel M, et al. Intratumoral BCG and *Corynebacterium paroum* therapy of canine mammary tumours before radical mastectomy. *Cancer Immunol Immunother* 1983; 15: 172–177.
- 45 Fox LE, MacEwen EG, Kurzman ID, Dubielzig RR, Helfand SC, Vail DM, et al. Liposome-encapsulated muramyl tripeptide phosphatidylethanolamine for the treatment of feline mammary adenocarcinoma – a multicenter randomized double-blind study. *Cancer Biother* 1995; 10: 125–130.
- 46 MacEwen EG, Hayes AA, Mooney S, Patnaik AK, Harvey HJ, Passe S, et al. Evaluation of effect of levamisole on feline mammary cancer. J Biol Response Mod 1984; 3: 541–546.
- 47 London CA. **Tyrosine kinase inhibitors in veterinary medicine.** *Top Companion Anim Med* 2009; 24: 106–112.
- 48 Bellamy F, Bader T, Moussy A and Hermine O. Pharmacokinetics of masitinib in cats. Vet Res Commun 2009; 33: 831–837.
- 49 Daly M, Sheppard S, Cohen N, Nabity M, Moussy A, Hermine O, et al. Safety of masitinib mesylate in healthy cats. J Vet Intern Med 2011; 25: 297–302.
- 50 Isotani M, Tamura K, Yagihara H, Hikosaka M, Ono K, Washizu

T, et al. Identification of a c-kit exon 8 internal tandem duplication in a feline mast cell tumor case and its favorable response to the tyrosine kinase inhibitor imatinib mesylate. *Vet Immunol Immunopathol* 2006; 114: 168–172.

- 51 Lachowicz JL, Post GS and Brodsky E. A phase I clinical trial evaluating imatinib mesylate (Gleevec) in tumor-bearing cats. *J Vet Intern Med* 2005; 19: 860–864.
- 52 Viste JR, Myers SL, Singh B and Simko E. Feline mammary adenocarcinoma: tumor size as a prognostic indicator. *Can Vet J* 2002; 43: 33–37.
- 53 Ito T, Kadosawa T, Mochizuki M, Matsunaga S, Nishimura R and Sasaki N. Prognosis of malignant mammary tumor in 53 cats. J Vet Med Sci 1996; 58: 723–726.
- 54 Hughes K and Dobson JM. Prognostic histopathological and molecular markers in feline mammary neoplasia. *Vet J* 2012; 194; 19–26.
- 55 Castagnaro M, Casalone C, Bozzetta E, De Maria R, Biolatti B and Caramelli M. Tumour grading and the one-year post-surgical prognosis in feline mammary carcinomas. J Comp Pathol 1998; 119: 263–275.
- 56 Seixas F, Palmeira C, Pires MA, Bento MJ and Lopes C. Grade is an independent prognostic factor for feline mammary carcinomas: a clinicopathological and survival analysis. *Vet J* 2011; 187: 65–71.
- 57 Millanta F, Lazzeri G, Mazzei M, Vannozzi I and Poli A. MIB-1 labeling index in feline dysplastic and neoplastic mammary lesions and its relationship with postsurgical prognosis. *Vet Pathol* 2002; 39: 120–126.
- 58 Preziosi R, Sarli G, Benazzi C, Mandrioli L and Marcato PS. Multiparametric survival analysis of histological stage and proliferative activity in feline mammary carcinomas. *Res Vet Sci* 2002; 73: 53–60.
- 59 Dias Pereira P, Carvalheira J and Gartner F. Cell proliferation in feline normal, hyperplastic and neoplastic mammary tissue – an immunohistochemical study. Vet J 2004; 168: 180–185.
- 60 Castagnaro M, Casalone C, Ru G, Nervi GC, Bozzetta E and Caramelli M. Argyrophilic nucleolar organiser regions (AgNORs) count as indicator of post-surgical prognosis in feline mammary carcinomas. *Res Vet Sci* 1998; 64: 97–100.
- 61 Preziosi R, Sarli G, Benazzi C and Marcato PS. Detection of proliferating cell nuclear antigen (PCNA) in canine and feline mammary tumours. *J Comp Pathol* 1995; 113: 301–313.
- 62 Millanta F, Calandrella M, Citi S, Della Santa D and Poli A. Overexpression of HER-2 in feline invasive mammary carcinomas: an immunohistochemical survey and evaluation of its prognostic potential. *Vet Pathol* 2005; 42: 30–34.
- 63 Rasotto R, Caliari D, Castagnaro M, Zanetti R and Zappulli V. An immunohistochemical study of HER-2 expression in feline mammary tumours. J Comp Pathol 2011; 144: 170–179.
- 64 Millanta F, Citi S, Della Santa D, Porciani M and Poli A. COX-2 expression in canine and feline invasive mammary carcinomas: correlation with clinicopathological features and prognostic molecular markers. *Breast Cancer Res Treat* 2006; 98: 115–120.
- 65 Millanta F, Silvestri G, Vaselli C, Citi S, Pisani G, Lorenzi D, et al. The role of vascular endothelial growth factor and its receptor Flk-1/KDR in promoting tumour angiogenesis in feline and canine mammary carcinomas: a preliminary study of autocrine and paracrine loops. *Res Vet Sci* 2006; 81: 350–357.
- 66 Maniscalco L, Iussich S, de las Mulas JM, Millán Y, Biolatti B, Sasaki N, et al. Activation of AKT in feline mammary carcinoma: a new prognostic factor for feline mammary tumours. *Vet J* 2012; 191: 65–71.