Genome-wide SNP-sex interaction analysis of susceptibility to idiopathic pulmonary fibrosis

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Abstract

Background

Idiopathic pulmonary fibrosis (IPF) is a chronic lung condition that is more prevalent in males than females. The reasons for this are not fully understood, with differing environmental exposures due to historically sex-biased occupations, or diagnostic bias, being possible explanations. To date, over 20 independent genetic variants have been identified to be associated with IPF susceptibility, but these have been discovered when combining males and females. Our aim was to test for the presence of sex-specific associations with IPF susceptibility and assess whether there is a need to consider sex-specific effects when evaluating genetic risk in clinical prediction models for IPF.

Methods

We performed genome-wide single nucleotide polymorphism (SNP)-by-sex interaction studies of IPF risk in six independent IPF case-control studies and combined them using inverse-variance weighted fixed effect meta-analysis. In total, 4,561 cases (1,280 females and 2,281 males) and 23,500 controls (8,360 females and 14,528 males) of European genetic ancestry were analysed. We used polygenic risk scores (PRS) to assess differences in genetic risk prediction between males and females.

Findings

Three independent genetic association signals were identified. All showed a consistent direction of effect across all individual IPF studies and an opposite direction of effect in IPF susceptibility between females and males. None had been previously identified in IPF susceptibility genome-wide association studies (GWAS). The predictive accuracy of the PRSs were similar between males and females, regardless of whether using combined or sex-specific GWAS results.

Interpretation

We prioritised three genetic variants whose effect on IPF risk may be modified by sex, however these require further study. We found no evidence that the predictive accuracy of common SNP-based PRSs varies significantly between males and females.

Research in context

Evidence before this study

The prevalence of IPF is higher in males than females. IPF risk has a genetic component, but analyses have only been performed in studies where males and females have been combined. One previous study reported sex-specific differences in association for the *MUC5B* promoter variant, rs35705950, however the finding was not replicated in an independent study. No genome-wide association studies assessing for different genetic risk factors between males and females have been conducted for IPF. It is not known whether approaches to predict individuals at risk of IPF should take sex-specific genetic risk into consideration.

Added value of this study

This was the largest study to test whether there are genetic variants whose effects on IPF susceptibility are different in males and females. The *MUC5B* promotor variant rs35705950 did not show a different magnitude of effect in males vs females. We identified three genetic variants with opposite directions of effect on IPF risk in males vs females. Our polygenic risk score analyses suggested that genetic prediction based on data from males and females separately did not perform better than when males and females were combined.

Implications of all available evidence

Although we found some preliminary evidence of genetic variants with sex-specific effects on IPF risk, our analyses suggest that genome-wide genetic risk from common single nucleotide polymorphisms is similar in males and females. This is important when considering integration of polygenic risk scores into clinical prediction models for IPF. There may be other forms of genetic variation, such as complex structural variation or rare variants, not captured in this analysis, that may improve risk prediction for males and females separately.

Introduction

Idiopathic pulmonary fibrosis (IPF) is a progressively fibrotic lung disease with a median survival time after diagnosis of 3-5 years¹. In the USA and Europe, IPF is estimated to have a disease prevalence of 0.63 to 7.6 per 100,000 people². The number of people diagnosed with IPF is increasing and males are more likely to be diagnosed than women^{3,4}. However, the reason why the disease is more prevalent in males is not understood. Different environmental exposures between males and females, notably occupations such as carpentry which have traditionally been more common amongst men⁵, could explain some of the observed difference. Diagnostic bias may also play a role with men being over diagnosed and women being undiagnosed with IPF⁶. As well as prevalence differences, there are survival differences with men having worse survival after IPF diagnosis than females⁷. Differences in genetic predisposition between males and females may be an additional factor in prevalence differences, however, this has not yet been extensively studied.

IPF is a complex polygenic disease with multiple genes implicated in susceptibility. The genetic variant rs35705950 in the *MUC5B* gene promoter has been shown to increase a person's risk of IPF 5-fold for each copy of the risk allele⁸⁻¹¹ and has been estimated to explain more than three times more disease liability than the other known common IPF risk variants combined¹². In recent years, genome-wide association studies (GWAS), examining genetic variants across the genome, have identified over 20 genetic loci associated with IPF risk¹³⁻¹⁶. In addition to providing new insight into disease biology, polygenic risk scores (PRS) derived from GWAS data have shown potential utility in identifying individuals at highest risk of pulmonary fibrosis¹⁷.

We hypothesised that there might be different biological mechanisms that promote IPF susceptibility in males and females, and that genetic associations that differ between males and females, may pinpoint the genes and pathways involved. To test this, we performed a genome-wide single nucleotide polymorphism (SNP)-by-sex interaction meta-analysis of IPF risk in six independent clinically-defined IPF case-control studies. Given the increasing interest in the use of PRS as a clinical tool for diagnosis in complex diseases¹⁸, we additionally tested whether PRS derived from sex-combined association data performed differently in males and females or whether derivation of PRS from sex-specific data might improve predictive accuracy.

Methods

Studies

We performed a SNP-by-sex interaction meta-analysis for IPF risk using six independent IPF case-control studies, all of which have been previously described; US¹⁹ (formerly referred to as Chicago), Colorado¹³, UK²⁰, UUS¹¹, Genentech²¹ and CleanUP-UCD^{22,23} (**Table 1**). In short, unrelated participants from the six studies were included in this analysis if they were of genetically-determined European ancestry and had sex-at-birth recorded. We only included participants who passed genotyping quality control and cases were defined using the relevant American Thoracic Society/European Respiratory Society guidelines^{24,25} (**Supplementary Methods**).

Genome-wide SNP-by-sex Interaction Analyses

Genome-wide SNP-by-sex interaction analyses of IPF risk were performed separately in each of the six studies and meta-analysed using PLINK 1.9 (www.cog-genomics.org/plink/1.9/) 26 (**Supplementary Methods**). Analyses were performed using autosomal SNPs. We used $P < 5 \times 10^{-8}$ as the threshold for genome-wide significance and $P < 1 \times 10^{-6}$ for suggestive significance in the meta-analysis. Independent sentinel variants were defined using distance-based and conditional analysis methods (**Supplementary Methods**).

As all available datasets with both male and female participants were included within the genomewide discovery analysis to maximise statistical power, we applied Meta-Analysis Model-based Assessment of replicability (MAMBA²⁷) to assess the posterior probability of replication for all SNPs with a meta-analysis $P<1\times10^{-6}$. SNPs with a MAMBA posterior probability of replication (PPR) >90% were considered to be robust across the contributing studies and likely to replicate in future studies.

Male-specific and female-specific effect estimates were calculated for all sentinel variants passing the above criteria. We sought validation of male-specific effect sizes and direction in a male-only IPF case control study, IPFJES (IPF Job Exposure Study)²⁸, comprising 416 male IPF cases and 2,465 male controls (**Figure 1**, **Supplementary Methods**). No independent female-only datasets were available at the time of conducting this study.

Bioinformatic investigation of significant signals

Annotation of variants was performed using Variant Effect Predictor (VEP)²⁹. We used GTEx to assess whether the sentinel variants were eQTLs for gene expression in up to 49 tissues (including lung and non-lung tissues) and the coloc package³⁰ in R version 4.2.1 to test if sentinel variants were eQTLs for gene expression in lung or cultured fibroblasts (**Figure 1, Supplementary Methods**). We conducted phenome-wide association studies (PheWAS) using PhenoScanner^{31,32} and Open Targets³³ to examine whether the signals were also associated with other phenotypes.

Polygenic risk score analyses

As well as looking at the effect individual genetic variants have on disease risk, the effect multiple SNPs have can be explored using PRS. The scores are constructed by taking the weighted sum (usually weighted by the SNP effects from a GWAS data) across many SNPs. It can then be tested whether these scores are predictors of disease risk. We wanted to test whether the predictive accuracy of PRS in predicting IPF risk differed between males and females. The predictive accuracy of PRS was evaluated in two ways: 1) the predictive performance of the PRS derived from sexcombined IPF susceptibility GWAS¹⁴ was evaluated in males and females separately ('standard PRS') and 2) the predictive performance of the PRS derived from sex-specific GWAS was evaluated in males and females separately ('sex-specific PRS') (Figure 1). For 1) we first evaluated a 19-variant PRS representing previously reported common genome-wide significant (P<5x10⁻⁸) signals of association with IPF14. For both 1) and 2) we further incorporated genome-wide data using an approach that varied the threshold used for inclusion of variants in the PRS (PRSice v2.3.5³⁴). 'Base data' were derived from sex-combined and sex-specific meta-analyses of the US, Colorado, UK, UUS and Genentech datasets. The 'target dataset' was the independent CleanUP-UCD study comprising 465 cases (93 females and 372 males) and 2,455 controls (530 females and 1,925 males). Area Under the Curve (AUC) differences were tested using DeLong's test (Supplementary Methods).

Results

The genome-wide sex interaction analysis of IPF risk was performed in up to 4,561 cases (comprising 1,280 females and 2,281 males) and 23,500 controls (8,360 females and 14,528 males). A total of 8,485,642 genetic variants were included in the meta-analysis and there was no evidence of inflated test statistics (Figure 2 & Figure S1).

Three independent sentinel variants with interaction $P<1\times10^{-6}$ and MAMBA PPR>90% were identified (Table S1). All three variants had consistent direction of effect across all contributing studies (Figure 3a, b, c).

The sentinel variant rs62040020, which resided within an intron of JPT2 (Jupiter microtubule associated homolog 2) on chromosome 16 (effect allele frequency (EAF) = 10.6%), was measured in all six studies with a high imputation quality (R² > 0.88 across all six studies) and was nominally significant (P<0.05) in four of the six studies (Table S1 & Figure 4a). When tested for association with IPF risk in females and males separately, the minor allele (allele = C) of rs62040020 was associated with increased risk of IPF in females (odds ratio (OR) 1.34, 95% confidence intervals (CI) 1.15-1.55, $P=1x10^{-4}$) and decreased risk in males (OR 0.82, 95% CI 0.74-0.92, $P=1x10^{-3}$) (Figure 5). Accordingly, if we instead took the major allele to be the effect allele (allele = G), the direction of effect would be in the opposite direction (i.e., increased risk of IPF in males and decreased risk in females). In the maleonly IPFJES study, the association effect was close to the null and non-significant (OR 0.99, 95% CI 0.77-1.27, P=0.939) (Figure S2a). In lung and/or cultured fibroblasts, the C allele of rs62040020 (associated with increased IPF risk in females, decreased risk in males) was associated with increased expression of FAHD1 (Fumarylacetoacetate Hydrolase Domain Containing 1), MEIOB (meiosis specific with OB-fold) and NUBP2 (NUBP Iron-Sulfur Cluster Assembly Factor 2, Cytosolic) and decreased expression of MRPS34 (mitochondrial ribosomal protein S34) (Table S2). This allele was also related to changes in splicing of HAGH (hydroxyacylglutathione hydrolase) in lung, cultured fibroblasts and a range of other tissues. However, rs62040020 was not the most significant variant associated with expression of these genes at this locus, with colocalisation analyses for male sex-specific GWAS results suggesting the GWAS and eQTL association signals were driven by different variants (Figure S3 & Table S3, Supplementary Methods). The C allele of this variant was previously associated with reduced monocyte percentage in UK Biobank (NEALE round 2 results: http://www.nealelab.is/ukbiobank/, $P=1.7x10^{-4}$) (Table S2).

For the other two SNPs, rs1756167317 and rs1663078846, the sentinel variants were of low frequency (EAF: 1-5%) and were nominally significant in 3 out of 5 contributing studies and 2 out of 5 contributing studies, respectively (Figures 3b, 3c & Table S1). The A allele of rs1756167317, located in an intron of PRR7 (Proline rich 7, synaptic) at chromosome 5 (Figure 4b), was associated with increased risk of IPF in females (OR 2.18, 95% CI 1.56-3.04, P=5x10-6) and decreased risk in males (OR 0.63, 95% CI 0.47-0.86, $P=3\times10^{-3}$) (Figure 5). This SNP showed a consistent direction and size of effect in the male only IPFJES study (OR 0.68, 95% CI 0.36-1.30, P=0.247) (Figure S2b). For the sentinel variant of the signal on chromosome 2, rs1663078846, the C allele was associated with increased risk of IPF in females (OR 1.69, 95% CI 1.33-2.14, $P=2x10^{-5}$) and decreased risk in males (OR 0.76, CI 0.63-0.92, $P=6\times10^{-3}$) (Figure 5). This intergenic variant (Figure 4c) did not show a consistent effect in males in the IPFJES study (OR 1.07, 95% CI 0.74-1.55, P=0.702)(Figure S2c). In PhenoScanner and Open Targets no associations with other traits for rs1756167317 or rs1663078846 were found.

No variants met genome-wide significance ($P < 5 \times 10^{-8}$), one was borderline genome-wide significant (rs1756167317, $P = 5.76 \times 10^{-8}$). None of the common previously reported IPF susceptibility variants, including the MUC5B promoter variant rs35705950, were observed to have a significant sex interaction effect when accounting for multiple testing (Table S4) although the DSP (desmoplakin) variant, rs2076295, had a nominally significant interaction effect (P=0.03) (Figure S4).

Polygenic risk scores

For the 'standard PRS' analysis, there was no difference in the AUC between males and females for the 19-variant PRS (AUC males: 80.3% vs AUC females: 80.8%, DeLong P = 0.85) (Table S5). When constructing multiple PRS (i.e., not limiting to published IPF susceptibility variants) the most predictive p-value threshold (P_T) was $P_T < 4.5 \times 10^{-4}$ for males and $P_T < 5 \times 10^{-4}$ for females (Figure S5a & Figure S5b), and whilst the AUC estimated was slightly lower for males than for females, the difference was not statistically significant (male AUC: 80.2% vs female AUC: 81.8%, DeLong P = 0.54).

For the 'sex-specific PRS' analysis, the male specific PRS predictive accuracy was slightly higher than the female specific PRS predictive accuracy, but the difference was not statistically significant (male-specific PRS AUC: 78.2% vs female-specific PRS AUC: 76%, DeLong P = 0.47) (Table S5) (Figure S5c & Figure S5d). The AUCs observed in this analysis were smaller than those generated in the 'standard PRS' analysis, which might be explained by the decrease in sample size of the training set (i.e., less accurate effect sizes).

The PRS results suggest that the predictive accuracy of IPF PRSs is not statistically different between males and females, regardless whether using combined or sex-specific GWAS results.

Discussion

We performed the first genome-wide SNP-by-sex interaction analysis of IPF risk in clinically-defined cases and identified three independent signals that were suggestively significant at $P<1\times10^{-6}$. To test whether a combination of variant effects (including known IPF susceptibility variants, as well as those not reaching statistical significance) predict IPF susceptibility in males and females differently, we performed PRS analyses. We found that the predictive accuracies of PRSs were not sexdependent suggesting that PRS developed from sex-combined association statistics are largely generalisable across sexes.

Although none of the genetic variants analysed reached genome-wide significance in our interaction analysis, three sentinel variants met a less stringent threshold for significance and were consistent across all studies included in the meta-analysis. None of the variants replicated in the male-only IPFJES study, but this could have been because the study was under-powered to validate the male-specific effects. Further data are needed to provide confidence in these signals and to confirm the likely causal genes at these loci. These new signals however may offer further insight into sex-specific mechanisms in IPF. *FAHD1*, *HAGH* and *MRPS34* have all been implicated in mitochondrial function; mitochondrial dysfunction has been widely implicated in age-related disease such as IPF³⁵. *HAGH* was also amongst 2,940 genes differentially expressed between IPF cases and controls³⁶. However, the sex-specific effect of these genes has not been investigated.

None of the 19 previously reported common IPF genetic variants 14 demonstrated a suggestively significant sex-interaction ($P < 1 \times 10^{-6}$). Previously, a sex-stratified meta-analysis conducted across six biobank studies 15 observed a larger effect size in males compared with females for the MUC5B variant rs35705950. The effect was not replicated in a clinically-defined IPF sub-study of FinnGen or in the four clinically-defined case-control studies (four of the six studies used were included in our present study). We also did not observe this difference in our analysis (which included additional datasets). We have previously highlighted differences in genetic association effect sizes when defining IPF from routine electronic healthcare records compared to clinically defined cohorts 37 suggesting that case definition heterogeneity might account for the biobank finding.

Studies of interaction effects require larger sample sizes than GWAS of main effects on disease risk as we are testing for a difference in effect size between two subgroups of participants; we cannot exclude the possibility that there are additional sex-specific genetic association signals yet to be discovered with larger sample sizes. However, our PRS analysis also included variants not meeting stringent statistical significance and did not suggest that there were any large effect sex-specific signals yet to be detected. There were more males than females in the analysis, which would affect the 95% CI of the AUC, with females having a wider AUC 95% CI than males.

It could be that there are sex differences, but we might not see these differences on the autosomal chromosomes as sex chromosomes were not analysed. Furthermore, given the known role of

mitochondrial function it may be that sex differences might be observed in mitochondrial DNA. Our PRS analysis was not intended to define the optimum PRS for IPF in either or both sexes, but rather to indicate whether future efforts should focus on sex-specific PRS development. Our study was limited to variants with MAF>1% and as such we cannot exclude the potential for rare variants with sex-specific effects. All data included in this study was derived from individuals of European ancestry; our findings may not be generalisable to other ancestries and larger genetic studies of ILD in non-European ancestry populations, with appropriate representation of both sexes, are urgently needed.

In summary, our genome-wide SNP-by-sex interaction analysis identified three potential sexinteraction signals which require further validation and functional investigation. Our polygenic risk score analysis suggests that PRS derived from sex-combined IPF SNP association studies perform similarly in males and females with no significant benefit in deriving sex-specific PRS.

Data Availability

Full summary statistics for genome-wide SNP-sex interaction meta-analyse can be accessed from https://github.com/genomicsITER/PFgenetics.

Ethics

This research was conducted with appropriate ethics approval.

The PROFILE study (which provided samples for the UK and UUS studies) had institutional ethics approval at the University of Nottingham (NCT01134822 – ethics reference 10/H0402/2) and Royal Brompton and Harefield NHS Foundation Trust (NCT01110694 – ethics reference 10/H0720/12). UK samples were recruited across multiple sites with individual ethics approval (University of Edinburgh Research Ethics Committee [The Edinburgh Lung Fibrosis Molecular Endotyping (ELFMEN) Study NCT04016181] 17/ES/0075, NRES Committee South West – Southmead, Yorkshire and Humber Research Ethics Committee 08/H1304/54 and Nottingham Research Ethics Committee 09/H0403/59). Spanish samples were recruited under ethics approval by ethics committee from the Hospital Universitario N.S. de Candelaria (reference of the approval: PI-19/12). The UUS study also included individuals from clinical trials with ethics approval (ACE [NCT00957242] and PANTHER [NCT00650091]). For the UCSF cohort, sample and data collection were approved by the University of California San Francisco Committee on Human Research and all patients provided written informed consent. For the Vanderbilt cohort, the Institutional Review Boards from Vanderbilt University approved the study and all participants provided written informed consent before enrolment. For individuals recruited at the University of Chicago, consenting patients with IPF who were prospectively enrolled in the institutional review board-approved ILD registry (IRB#14163A) were included. Individuals recruited at the University of Pittsburgh Medical Centre had ethics approval from the University of Pittsburgh Human Research Protection Office (referenceSTUDY20030223: Genetic Polymorphisms in IPF). Individuals from the COMET (NCT01071707) and Lung Tissue Research Consortium (NCT02988388) studies were also included in the Chicago study. All subjects in the Colorado study gave written informed consent as part of IRBapproved protocols for their recruitment at each site and the GWAS study was approved by the National Jewish Health IRB and Colorado Combined Institutional Review Boards (COMIRB). Subjects in the Genentech study provided written informed consent for whole-genome sequencing of their DNA. Ethical approval was provided as per the original clinical trials (INSPIRE [NCT00075998], RIFF [NCT01872689], CAPACITY [NCT00287729 and NCT00287716] and ASCEND [NCT01366209]). Individuals in the CleanUP-UCD study included individuals from clinical trials with ethics approval

(NCT02759120). These samples were genotyped under University of Virginia ethics approval (IRB 20845). IPFJES involved human participants and was approved by East Midlands - Nottingham 1 Research Ethics Committee REC reference: 17/EM/0021IRAS project ID: 203355. Participants gave informed consent to participate in the study before taking part.

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The UK, UUS CleanUP-UCD and IPFJES studies selected controls from UK Biobank under application 648. This research used the SPECTRE High Performance Computing Facility at the University of Leicester. For the purpose of open access, a CC BY or equivalent licence will be applied to any author accepted.

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Global Biobank Engine:

Global Biobank Engine, Stanford, CA (URL: http://gbe.stanford.edu) [19th May 2022 accessed]. G. McInnes, Y. Tanigawa, C. DeBoever, A. Lavertu, J. E. Olivieri, M. Aguirre, M. A. Rivas, Global Biobank Engine: enabling genotype-phenotype browsing for biobank summary statistics. Bioinformatics (2019). doi:10.1093/bioinformatics/bty999

Conflicts of Interest

AA declares funding from NIH (K23HL146942); consulting fees from Genentech, Inogen, Medscape, Abbvie, PatientMpower and Boehringer Ingelheim; payment or honoraria for lectures, presentations, speakers bureaus, manuscript writing or educational events from Boehringer Ingelheim. ADS is a full-time employee of Genentech/Roche with stock and stock options in Roche. AFG was a full-time employee of PPD, Part of Thermo Fisher Scientific until June 2023. BGG declares fellowship funding from Wellcome Trust (221680/Z/20/Z). BLY is a full-time employee of Genentech/Roche with stock and stock options in Roche. CF declares funding Ministerio de Ciencia e Innovación, Instituto de Salud Carlos III and Instituto Tecnológico y de Energías Renovables; honoraria in educational events from Fundación Instituto Roche. DAS declares being the founder and chief scientific officer of Eleven P15, Inc., a company dedicated to the early diagnosis and treatment of pulmonary fibrosis. HP declares grant payment to institution from Boehringer Ingelheim Ltd;

consulting fees from Boehringer Ingelheim Ltd, Roche Limited, Trevi Therapeutics, Pilant Therapeutics; speaker fees from Boehringer Ingelheim Ltd; member of TIPAL trial management group, trustee for Action for Pulmonary Fibrosis, member of scientific advisory board for European Pulmonary Fibrosis Federation. IN declares funding from National Institutes of Health (UG3HL145266) to institution; grant funding to institution from Veracyte; consulting fees from Boerhinger Ingelheim and Sanofi. IPH declares funding from Wellcome Trust and NIHR; vice chair Trustees for Asthma + Lung UK. JMO declares funding from National Institutes of Health (R01HL169166 & K23HL138190); consulting fees from Boehringer Ingelheim, Lupin pharmaceuticals, AmMax Bio, Roche and Veracyte; patent for TOLLIP TT genotype for NAC use in IPF; participation on a Data Safety Monitoring Board or Advisory Board for Endeavor Biomedicines, Novartis and Genentech; Associate editor for CHEST, on Program Committee for American Thoracic Society and Editorial board for AJRCCM. LVW declares funding from UK Research and Innovation (MR/V00235X/1) and GSK/Asthma + Lung UK (Professorship (C17-1)) to complete this work; funding from Orion Pharma, GSK, Genentech, AstraZeneca, Nordic Bioscience, Sysmex (OGT); Consulting fees Galapagos, Boehringer Ingelheim, GSK; support for attending meetings and/or travel Genentech; participation on Advisory Board for Galapagos; leadership or fiduciary roles as Associate Editor for European Respiratory Journal and Medical Research Council Board member and Deputy Chair. MKBW declares funding from National Institutes of Health (K23HL146942); consulting fees from Genentech, Inogen, Medscape, Abbvie, PatientMpower and Boehringer Ingelheim; payment or honoraria for lectures, presentations, speakers bureaus, manuscript writing or educational events from Boehringer Ingelheim. MN is a full-time employee of Genentech/Roche with stock and stock options in Roche. NK declares grant funding from National Institutes of Health; grant funding to institution from BMS, Boehringer Ingelheim and Three Lakes Foundation; consultancy fees from Biogen Idec, Boehringer Ingelheim, Third Rock, Pliant, Samumed, NuMedii, Theravance, Three Lake Partners, Astra Zeneca, RohBar, Veracyte, Augmanity, CSL Behring, Thyron, Gilead, Galapagos, Chiesi, Arrowhead, Sofinnova, GSK and Merk; patent for new therapies for IPF (Biotech), new therapies for ARDS (Biotech) and new Biomarkers in IPF (Biotech); equity in Pilant and Thyron; reports serving as the scientific founder of Thyron. PLM declares grant funding to institution from AstraZeneca; consultancy fees from Hoffman-La Roche, Boehringer Ingelheim, AstraZeneca, Trevi and Qureight; speaker fees from Boehringer Ingelheim and Hoffman-La Roche. RGJ declares funding from UK Research and Innovation (MR/V00235X/1); that their institute received funding from Astra Zeneca, Biogen, Galecto, GlaxoSmithKline, Nordic Biosciences, RedX and Pliant; consulting fees from AstraZeneca, Brainomix, Bristol Myers Squibb, Chiesi, Cohbar, Daewoong, GlaxoSmithKline, Veracyte, Resolution Therapeutics and Pliant; payment for lectures and presentations received from Boehringer Ingelheim, Chiesi, Roche, PatientMPower, AstraZeneca; payment for expert testimony from Pinsent Masons LLP; participation on a Data Safety Monitoring Board or Advisory Board for Boehringer Ingelheim, Galapagos, Vicore; leadership or fiduciary role for NuMedii and president for Action for Pulmonary Fibrosis. SPH declares grant funding to institution from Boehringer Ingelheim; consulting fees from Trevi therapeutics; payment or honoraria for lectures, presentations, speakers bureaus, manuscript writing or educational events from Chiesi and Trevi therapeutics; support for attending meetings and/or travel from Chiesi; Participation on a Data Safety Monitoring Board or Advisory Board for Trevi therapeutics; Chair for BTS Standards of Care Committee (till November 2022) and Trustee for Action for Pulmonary Fibrosis. TMM declares consulting fees from Boehringer Ingelheim, Roche/Genentech, Astra Zeneca, Bayer, Blade Therapeutics, Bristol-Myers Squibb, CSL Behring, Galapagos, Galecto, GlaxoSmithKline, IQVIA, Pfizer, Pliant, Respivant, Sanofi, Theravance, Trevi, Veracyte and Vicore; participation on a Data Safety Monitoring Board or Advisory Board for Fibrogen, Blade Therapeutics and Nerre. XRS is a full-time employee of Genentech/Roche with stock and stock options in Roche.

Manuscript Tables

Table 1: IPF case-control cohorts

	Discovery cohorts																Validation cohort	
	US		Colorado		UK		UUS		CleanUP-UCD		Genentech		Total			IPFJES		
	Cases	Controls	Cases	Controls	Cases	Controls	Cases	Controls	Cases	Controls	Cases	Controls	Cases	Controls	Total	Cases	Controls	
Males	374	241	1,017	2,289	433	2,356	597	7,210	372	1,925	488	507	3,281	14,528	17,809	416	2,465	
Females	138	269	498	2,394	179	1,010	196	2,790	93	530	176	1,367	1,280	8,360	9,640	NA	NA	
Total	512	510	1,515	4,683	612	3,366	793	10,000	465	2,455	664	1,874	4,561	22,888	27,449	416	2,465	

Manuscript Figures

Figure 1: Overview of the SNP-by-sex interaction analysis and polygenic risk score analysis

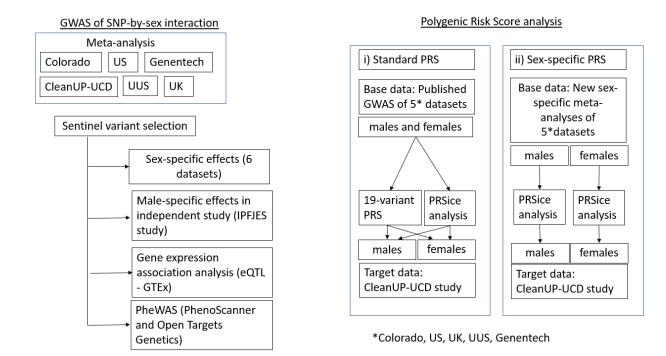


Figure 2: Manhattan plot of meta-analysed sex-interaction results. The chromosomal position is on the x-axis and the $-\log(p\text{-value})$ for each genetic variant in the sex-interaction meta-analysis is on the y-axis. Variants present in at least 3 studies are presented. The blue horizontal line represents the 1×10^{-6} p-value threshold and the red horizontal line represents 5×10^{-8} p-value threshold (genome-wide significance threshold).

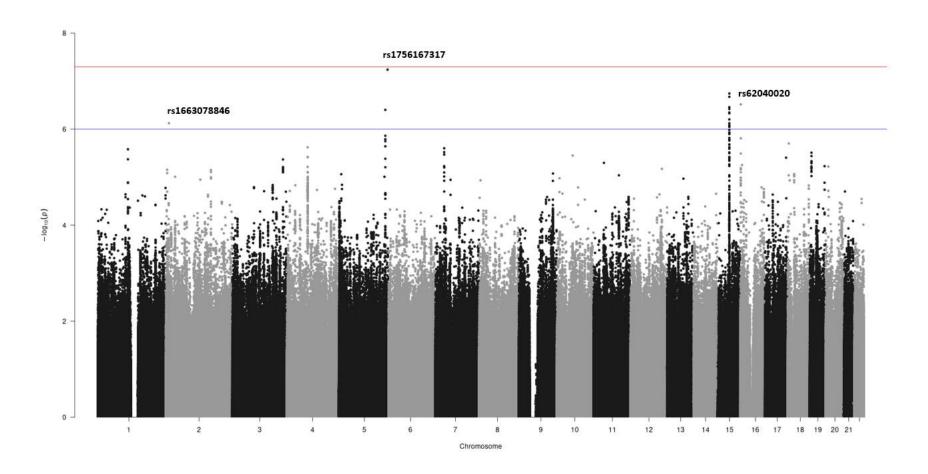
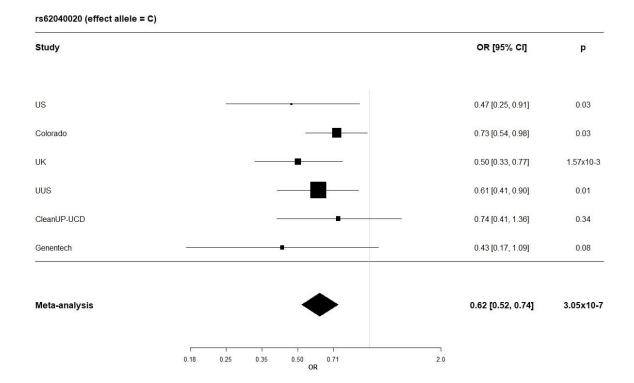


Figure 3: Forest plots showing SNP-sex interaction odds ratio by study and the meta-analysed results for a) rs62040020, b) rs1756167317 and c) rs1663078846.

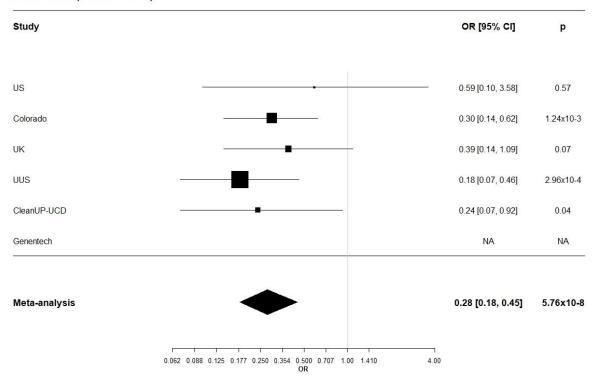
OR = odds ratio and CI = confidence interval

a)



b)

rs1756167317 (effect allele = A)



c)

rs1663078846 (effect allele = C)

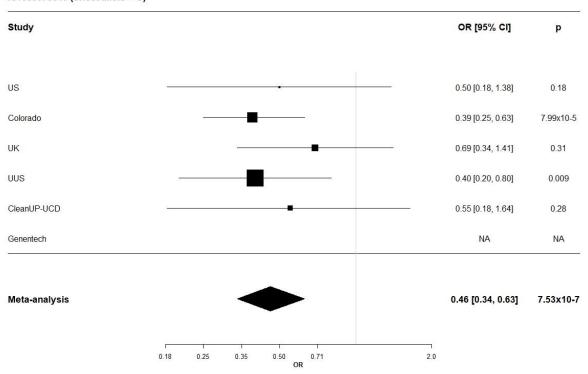
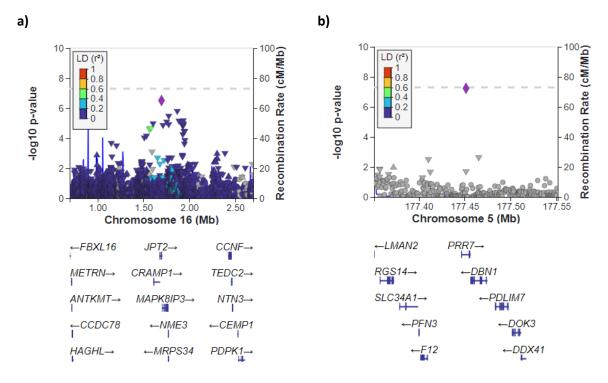


Figure 4: Region plots for a) rs62040020, b) rs1756167317 and c) rs1663078846. The chromosomal position is on the x-axis and the $-\log(p\text{-value})$ for each genetic variant in the sexinteraction meta-analysis is on the y-axis.



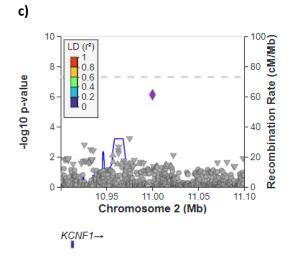
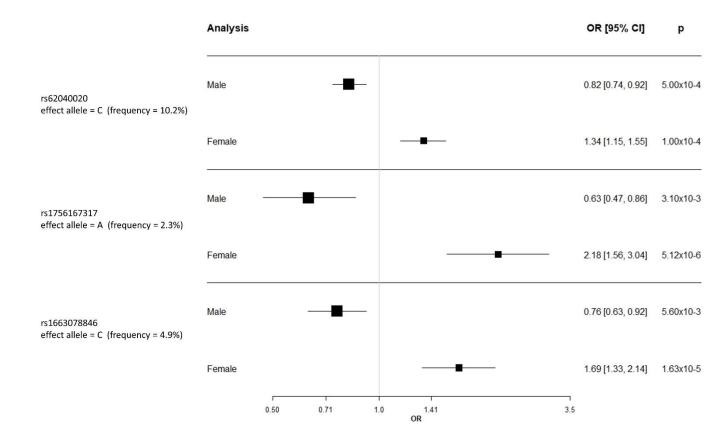


Figure 5: Forest plot for sex stratified results meta-analysed across US, Colorado, UK, UUS, CleanUP-UCD and Genentech.

OR = odds ratio and CI = confidence interval



References

- 1. Fernández Fabrellas E, Peris Sánchez R, Sabater Abad C, Juan Samper G. Prognosis and follow-up of idiopathic pulmonary fibrosis. *Medical Sciences*. 2018;6(2):51.
- 2. Gupta RS, Koteci A, Morgan A, George PM, Quint JK. Incidence and prevalence of interstitial lung diseases worldwide: A systematic literature review. *BMJ Open Respiratory Research*. 2023;10(1):e001291.
- 3. Maher TM, Bendstrup E, Dron L, et al. Global incidence and prevalence of idiopathic pulmonary fibrosis. *Respiratory research*. 2021;22(1):1-10.
- 4. Pergolizzi Jr JV, LeQuang JA, Varrassi M, Breve F, Magnusson P, Varrassi G. What do we need to know about rising rates of idiopathic pulmonary fibrosis? A narrative review and update. *Adv Ther*. 2023;40(4):1334-1346.
- 5. Trethewey SP, Walters GI. The role of occupational and environmental exposures in the pathogenesis of idiopathic pulmonary fibrosis: A narrative literature review. *Medicina*. 2018;54(6):108.
- 6. Assayag D, Morisset J, Johannson KA, Wells AU, Walsh SL. Patient gender bias on the diagnosis of idiopathic pulmonary fibrosis. *Thorax*. 2020;75(5):407-412.
- 7. Zaman T, Moua T, Vittinghoff E, Ryu JH, Collard HR, Lee JS. Differences in clinical characteristics and outcomes between men and women with idiopathic pulmonary fibrosis: A multicenter retrospective cohort study. *Chest.* 2020;158(1):245-251.
- 8. Seibold MA, Wise AL, Speer MC, et al. A common MUC5B promoter polymorphism and pulmonary fibrosis. *N Engl J Med*. 2011;364(16):1503-1512.

- 9. Moore C, Blumhagen RZ, Yang IV, et al. Resequencing study confirms that host defense and cell senescence gene variants contribute to the risk of idiopathic pulmonary fibrosis. *American journal of respiratory and critical care medicine*. 2019;200(2):199-208.
- 10. Zhu Q, Zhang X, Zhang S, et al. Association between the MUC5B promoter polymorphism rs35705950 and idiopathic pulmonary fibrosis: A meta-analysis and trial sequential analysis in caucasian and asian populations. *Medicine*. 2015;94(43).
- 11. Allen RJ, Guillen-Guio B, Oldham JM, et al. Genome-wide association study of susceptibility to idiopathic pulmonary fibrosis. *American journal of respiratory and critical care medicine*. 2020;201(5):564-574.
- 12. Leavy OC, Ma S, Molyneaux PL, et al. Proportion of idiopathic pulmonary fibrosis risk explained by known common genetic loci in european populations. *American journal of respiratory and critical care medicine*. 2021;203(6):775-778.
- 13. Fingerlin TE, Murphy E, Zhang W, et al. Genome-wide association study identifies multiple susceptibility loci for pulmonary fibrosis. *Nat Genet*. 2013;45(6):613-620.
- 14. Allen RJ, Stockwell A, Oldham JM, et al. Genome-wide association study across five cohorts identifies five novel loci associated with idiopathic pulmonary fibrosis. *Thorax*. 2022;77(8):829-833.
- 15. Partanen JJ, Häppölä P, Zhou W, et al. Leveraging global multi-ancestry meta-analysis in the study of idiopathic pulmonary fibrosis genetics. *Cell genomics*. 2022;2(10).
- 16. Dhindsa RS, Mattsson J, Nag A, et al. Identification of a missense variant in SPDL1 associated with idiopathic pulmonary fibrosis. *Communications biology*. 2021;4(1):392.

- 17. Moll M, Peljto AL, Kim JS, et al. A polygenic risk score for idiopathic pulmonary fibrosis and interstitial lung abnormalities. *American Journal of Respiratory and Critical Care Medicine*. 2023;208(7):791-801.
- 18. Lewis CM, Vassos E. Polygenic risk scores: From research tools to clinical instruments. *Genome medicine*. 2020;12(1):1-11.
- 19. Noth I, Zhang Y, Ma S, et al. Genetic variants associated with idiopathic pulmonary fibrosis susceptibility and mortality: A genome-wide association study. *The Lancet respiratory medicine*. 2013;1(4):309-317.
- 20. Allen RJ, Porte J, Braybrooke R, et al. Genetic variants associated with susceptibility to idiopathic pulmonary fibrosis in people of european ancestry: A genome-wide association study. *The Lancet respiratory medicine*. 2017;5(11):869-880.
- 21. Dressen A, Abbas AR, Cabanski C, et al. Analysis of protein-altering variants in telomerase genes and their association with MUC5B common variant status in patients with idiopathic pulmonary fibrosis: A candidate gene sequencing study. *The Lancet Respiratory Medicine*. 2018;6(8):603-614.
- 22. Martinez FJ, Yow E, Flaherty KR, et al. Effect of antimicrobial therapy on respiratory hospitalization or death in adults with idiopathic pulmonary fibrosis: The CleanUP-IPF randomized clinical trial. *JAMA*. 2021;325(18):1841-1851.
- 23. Allen RJ, Oldham JM, Jenkins DA, et al. Longitudinal lung function and gas transfer in individuals with idiopathic pulmonary fibrosis: A genome-wide association study. *The Lancet Respiratory Medicine*. 2023;11(1):65-73.

- 24. Raghu G, Collard HR, Egan JJ, et al. An official ATS/ERS/JRS/ALAT statement: Idiopathic pulmonary fibrosis: Evidence-based guidelines for diagnosis and management. *American journal of respiratory and critical care medicine*. 2011;183(6):788-824.
- 25. Raghu G, Remy-Jardin M, Myers JL, et al. Diagnosis of idiopathic pulmonary fibrosis. an official ATS/ERS/JRS/ALAT clinical practice guideline. *American journal of respiratory and critical care medicine*. 2018;198(5):e44-e68.
- 26. Chang CC, Chow CC, Tellier LC, Vattikuti S, Purcell SM, Lee JJ. Second-generation PLINK: Rising to the challenge of larger and richer datasets. *Gigascience*. 2015;4(1):s13742-8.
- 27. McGuire D, Jiang Y, Liu M, et al. Model-based assessment of replicability for genome-wide association meta-analysis. *Nature communications*. 2021;12(1):1964.
- 28. Reynolds CJ, Sisodia R, Barber C, et al. What role for asbestos in idiopathic pulmonary fibrosis? findings from the IPF job exposures case—control study. *Occup Environ Med*. 2023;80(2):97-103.
- 29. McLaren W, Gil L, Hunt SE, et al. The ensembl variant effect predictor. *Genome Biol*. 2016;17(1):1-14.
- 30. Giambartolomei C, Vukcevic D, Schadt EE, et al. Bayesian test for colocalisation between pairs of genetic association studies using summary statistics. *PLoS genetics*. 2014;10(5):e1004383.
- 31. Staley JR, Blackshaw J, Kamat MA, et al. PhenoScanner: A database of human genotype—phenotype associations. *Bioinformatics*. 2016;32(20):3207-3209.
- 32. Kamat MA, Blackshaw JA, Young R, et al. PhenoScanner V2: An expanded tool for searching human genotype–phenotype associations. *Bioinformatics*. 2019;35(22):4851-4853.

- 33. Ochoa D, Hercules A, Carmona M, et al. The next-generation open targets platform: Reimagined, redesigned, rebuilt. *Nucleic Acids Res.* 2023;51(D1):D1353-D1359.
- 34. Choi SW, O'Reilly PF. PRSice-2: Polygenic risk score software for biobank-scale data. *Gigascience*. 2019;8(7):giz082.
- 35. Zank DC, Bueno M, Mora AL, Rojas M. Idiopathic pulmonary fibrosis: Aging, mitochondrial dysfunction, and cellular bioenergetics. *Frontiers in medicine*. 2018;5:10.
- 36. DePianto DJ, Chandriani S, Abbas AR, et al. Heterogeneous gene expression signatures correspond to distinct lung pathologies and biomarkers of disease severity in idiopathic pulmonary fibrosis. *Thorax*. 2014:thoraxjnl-204596.
- 37. Leavy OC, Allen RJ, Kraven LM, et al. The use of genetic information to define idiopathic pulmonary fibrosis in UK biobank. *Chest.* 2023;163(2):362-365.