

literature report

Proteolysis in Alzheimer's disease

Can plasmin tip the balance?

Amyloid precursor protein (APP) is a synaptic single-pass transmembrane protein that is best known for its involvement in Alzheimer's disease, a devastating neurological disorder. In Alzheimer's disease patients, amyloid plaques containing aggregated β -amyloid peptide (A β) appear in specific brain regions, triggering an inflammatory response, neuronal cell death and gradual cognitive decline (reviewed in Selkoe, 1999). Aß is derived from APP that is first cleaved at an extracellular position $(\beta$ site), followed by an unusual cleavage within the APP transmembrane segment (γ site), producing ~40–42 amino acid A β peptides, with the longer forms apparently causing the neurotoxicity. This processing pathway has been studied intensively with the goal of devising therapeutic interventions. In recent years, the β -site cleavage enzymes (BACE) have been identified (reviewed in Vassar and Citron, 2000), and the γ cleavage has been shown to be performed by a proteolytic complex containing Presenilin and Nicastrin (Yu et al., 2000). However, APP is also processed by an alternative pathway, in which a different, slightly more membrane-proximal extracellular position (α site) is cleaved, followed by proteolysis at the γ site, as in the A β pathway. The α cleavage is thought to be mediated by extracellular matrix metalloproteases of the tumour necrosis factor α-converting enzyme (TACE)/a disintegrin and metalloprotease (ADAM) family, and it generates a non-toxic peptide termed p3. Unlike the $A\beta$ pathway, which appears to operate throughout the secretory and endosomal compartments (reviewed in Wilson et al., 1999), the p3 pathway is predominantly active at the cell surface (Parvathy et al., 1999).

In this issue of EMBO reports, Ledesma and colleagues (2000) bring attention to another protease that may influence the selection of AB or p3 processing pathways for APP. Starting with previous observations that $A\beta$ is present in rafts, specialized detergent insoluble glycolipid-enriched membrane microdomains (Lee *et al.*, 1998), that raft integrity is important for $A\beta$ production (Simons et al., 1998) and that purified plasmin can degrade AB (Wnendt et al., 1997; Van Nostrand and Porter, 1999; Tucker et al., 2000), Ledesma and co-workers examined neuronal rafts for plasmin interactions with APP and AB. Both plasmin and its inactive precursor, plasminogen, were detected in rafts of cultured hippocampal neurons, and remarkably, active plasmin was largely restricted to the rafts themselves, suggesting a new role for rafts as sites of active proteolysis. Addition of plasmin or tissue plasminogen activator (tPA), which converts plasminogen to plasmin, increased the amount of α cleavage of APP and decreased levels of AB. It is not known whether plasmin may directly cleave the α site of APP or whether these effects occur as a result of the known stimulation of TACE/ADAM metalloproteases by plasmin (Kleiner and Stetler-Stevenson, 1993). Intriguingly, Ledesma *et al.* (2000) reported reduced plasmin levels in Alzheimer's disease brain tissue, indicating that plasmin may be physiologically relevant as a protective factor in this disease.

The results of Ledesma et al. (2000) are bound to re-awaken interest in the idea that upregulation of the α pathway of APP processing might reduce the toxic build-up of A β , either by diverting APP away from the β pathway or by directly degrading existing A β . Ledesma and colleagues propose that A β levels are regulated by a balance of different protease activities acting on the α -, β - and γ -sites of APP in conjunction with an A β clearance system to inhibit $A\beta$ accumulation (Figure 1). The potential involvement of plasmin in these events was previously noted by other groups, who demonstrated that plasmin cleaves AB at certain sites and that exogenously added plasmin blocks AB neurotoxicity, supporting a physiological role for plasmin in APP/AB metabolism (Wnendt et al., 1997; Van Nostrand and Porter, 1999; Tucker et al., 2000). An added twist to this story is that these groups found that $A\beta$ itself induces the plasmin system, leading to increased tPA mRNA levels, an effect caused only by aggregated, not soluble, $A\beta$. Thus, a feedback mechanism may exist whereby toxic A β stimulates the plasmin system, leading to AB clearance in neurons, a mechanism that breaks down due to inflammation and increased expression of tPA inhibitors in Alzheimer's disease. The ideas that elevated AB accumulation may result from reduced plasmin activity (Ledesma et al., 2000) and that $A\beta$ accumulation may initially trigger increased plasmin activity (Wnendt et al., 1997; Van Nostrand and Porter, 1999; Tucker et al., 2000) may not necessarily contradict one another. Perhaps Aβ levels are normally kept within physiological limits by regulated degradation involving a positive plasmin feedback mechanism, but high A β levels might overwhelm this system by causing inflammation-related tPA inhibition, suppressing plasmin activity and promoting plaque formation (Figure 1).

Curiously, upregulation of tPA leads to enhancement of longterm potentiation and learning in mice (Madani et al., 1999), and longitudinal population studies on human subjects have uncovered a positive correlation between linguistic ability and protection from Alzheimer's disease (Snowdon et al., 2000). These observations suggest that there may be a connection between the cognitive and memory deficits seen in Alzheimer's disease and the activity of the plasminogen system. Taken together, these results highlight the need for further analysis of plasmin and associated factors in APP metabolism and Alzheimer's disease. While enhanced deposition of A β due to aberrant γ cleavage has been well documented in relatively rare familial forms of Alzheimer's disease caused by Presenilin gene mutations, it is conceivable that the more prevalent cases of sporadic Alzheimer's disease may involve impaired α pathway activity or A β degradation, perhaps due to plasmin activity in neuronal membrane rafts.

literature report

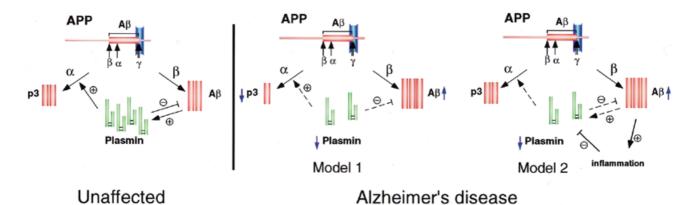


Fig. 1. Possible models for the role of plasmin in the development of Alzheimer's disease. In unaffected individuals (left), plasmin influences the balance of α and β pathway activity by stimulating the α pathway (Ledesma *et al.*, 2000) and degrading A β . A β , in turn, induces the plasmin system in a feedback mechanism (Wnendt *et al.*, 1997; Van Nostrand and Porter, 1999; Tucker *et al.*, 2000). In individuals with Alzheimer's disease (right), reduced plasmin in neuronal rafts shifts the proteolytic balance towards the β pathway, leading to increased A β production and plaque formation (Model 1) (Ledesma *et al.*, 2000). Alternatively, A β deposition may trigger inflammation, which induces inhibitors of the plasmin system, downregulating degradation and clearance of A β (Model 2) (Tucker *et al.*, 2000).

In addition to these effects on APP metabolism, plasminogen has recently been shown to interact with disease-associated forms of the prion protein (Fischer *et al.*, 2000), raising the possibility that the plasminogen system may influence the development or clinical severity of multiple neurodegenerative diseases.

Acknowledgements

G.P. is supported by an American Heart Association Postdoctoral Fellowship Award. M.E.F. is supported by the National Institutes of Health, the Alzheimer's Association and a Merck Research Grant.

References

- Fischer, M.B., Roeckl, C., Parizek, P., Schwarz, H.P. and Aguzzi, A. (2000) Binding of disease-associated prion protein to plasminogen. *Nature*, in press.
- Ikezu, T., Trapp, B.D., Song, K.S., Schlegel, A., Lisanti, M.P. and Okamoto, T. (1998) Caveolae, plasma membrane microdomains for α-secretase-mediated processing of the amyloid precursor protein. *J. Biol. Chem.*, 273, 10485–10495.
- Kleiner, D.E., Jr and Stetler-Stevenson, W.G. (1993) Structural biochemistry and activation of matrix metalloproteases. *Curr. Opin. Cell Biol.*, 5, 891–897.
- Ledesma, M.D., Da Silva, J.S., Crassaerts, K., Delacourte, A., De Strooper, B. and Dotti, C.G. (2000) Brain plasmin influences APP α -cleavage and A β degradation and is reduced in Alzheimer's disease brains. *EMBO rep.* **1**, 530–535.
- Lee, S.-J., Liyanage, U., Bickel, P.E., Xia, W., Lansbury, P.T., Jr and Kosik, K.S. (1998) A detergent-insoluble membrane compartment contains Aβ *in vivo. Nature Med.*, 4, 730–734.
- Madani, R., Hulo, S., Toni, N., Madani, H., Steimer, T., Muller, D. and Vassalli, J.-D. (1999) Enhanced hippocampal long-term potentiation and learning by increased neuronal expression of tissue-type plasminogen activator in transgenic mice. *EMBO J.*, **18**, 3007–3012.
- Parvathy, S., Hussain, I., Karran, E.H., Turner, A.J. and Hooper, N.M. (1999) Cleavage of Alzheimer's amyloid precursor protein by α-secretase occurs at the surface of neuronal cells. *Biochemistry*, **38**, 9728–9734.
- Selkoe, D.J. (1999) Translating cell biology into therapeutic advances in Alzheimer's disease. *Nature*, **399**, A23–A31.
- Simons, M., Keller, P., De Strooper, B., Beyreuther, K., Dotti, C.G. and Simons, K. (1998) Cholesterol depletion inhibits the generation of β-amyloid in hippocampal neurons. *Proc. Natl Acad. Sci. USA*, **95**, 6460–6464.

- Snowdon, D.A., Greiner, L.H. and Markesbery, W.R. (2000) Linguistic ability in early life and the neuropathology of Alzheimer's disease and cerebrovascular disease: findings from the nun study. *Ann. N.Y. Acad. Sci.*, 903, 34–38.
- Tucker, H.M., Kihiko-Ehmann, M., Wright, S., Rydel, R.E. and Estus, S. (2000) The plasmin system is induced by and degrades amyloid-β aggregates. J. Neurosci., 20, 3937–3946.
- Van Nostrand, W.E. and Porter, M. (1999) Plasmin cleavage of the amyloid β-protein: alteration of secondary structure and stimulation of tissue plasminogen activator activity. *Biochemistry*, **38**, 11570–11576.
- Vassar, R. and Citron, M. (2000) A β -generating enzymes: recent advances in β and γ -secretase research. *Neuron*, **27**, 419–422.
- Wilson, C.A., Doms, R.W. and Lee, V.M.-Y. (1999) Intracellular APP processing and $A\beta$ production in Alzheimer disease. *J. Neuropath. Exp. Neurol.*, **58**, 787–794.
- Wnendt, S., Wetzels, I. and Gunzler, W.A. (1997) Amyloid β peptides stimulate tissue-type plasminogen activator but not recombinant prourokinase. *Thromb. Res.*, 85, 217–224.
- Yu, G. et al. (2000) Nicastrin modulates presentiin-mediated notch/glp-1 signal transduction and βAPP processing. Nature, 407, 48–54.



Goran Periz and Mark E. Fortini+

Department of Genetics, University of Pennsylvania School of Medicine, Stellar-Chance Laboratories 709C, 422 Curie Boulevard, Philadelphia, PA 19104, USA

Received October 24, 2000; accepted November 11, 2000

*Corresponding author. Tel: +1 215 573 6446; Fax: +1 215 573 9411; E-mail: fortini@mail.med.upenn.edu

DOI: 10.1093/embo-reports/kvd124