

ARTICLE OPEN



Brain tissue- and cell type-specific eQTL Mendelian randomization reveals efficacy of *FADS1* and *FADS2* on cognitive function

Xueyan Wu^{1,2,6}, Lei Jiang^{1,2,6}, Hongyan Qi^{1,2,6}, Chunyan Hu^{1,2}, Xiaojing Jia^{1,2}, Hong Lin^{1,2}, Shuangyuan Wang^{1,2}, Lin Lin^{1,2}, Yifang Zhang³, Ruizhi Zheng^{1,2}, Mian Li^{1,2}, Tiange Wang^{1,2}, Zhiyun Zhao^{1,2}, Min Xu^{1,2}, Yu Xu^{1,2}, Yuhong Chen^{1,2}, Jie Zheng^{1,2,4,5}✉, Yufang Bi^{1,2}✉ and Jieli Lu^{1,2}✉

© The Author(s) 2024

Epidemiological studies suggested an association between omega-3 fatty acids and cognitive function. However, the causal role of the fatty acid desaturase (*FADS*) gene, which play a key role in regulating omega-3 fatty acids biosynthesis, on cognitive function is unclear. Hence, we used two-sample Mendelian randomization (MR) to estimate the gene-specific causal effect of omega-3 fatty acids ($N = 114,999$) on cognitive function ($N = 300,486$). Tissue- and cell type-specific effects of *FADS1*/*FADS2* expression on cognitive function were estimated using brain tissue cis-expression quantitative trait loci (cis-eQTL) datasets (GTEx, $N \leq 209$; MetaBrain, $N \leq 8,613$) and single cell cis-eQTL data ($N = 373$), respectively. These causal effects were further evaluated in whole blood cis-eQTL data ($N \leq 31,684$). A series of sensitivity analyses were conducted to validate MR assumptions. Leave-one-out MR showed a *FADS* gene-specific effect of omega-3 fatty acids on cognitive function [$\beta = -1.3 \times 10^{-2}$, 95% confidence interval (CI) $(-2.2 \times 10^{-2}, -5 \times 10^{-3})$, $P = 2 \times 10^{-3}$]. Tissue-specific MR showed an effect of increased *FADS1* expression in cerebellar hemisphere and *FADS2* expression in nucleus accumbens basal ganglia on maintaining cognitive function, while decreased *FADS1* expression in nine brain tissues on maintaining cognitive function [colocalization probability (PP.H4) ranged from 71.7% to 100.0%]. Cell type-specific MR showed decreased *FADS1*/*FADS2* expression in oligodendrocyte was associated with maintaining cognitive function (PP.H4 = 82.3%, respectively). Increased *FADS1*/*FADS2* expression in whole blood showed an effect on cognitive function maintenance (PP.H4 = 86.6% and 88.4%, respectively). This study revealed putative causal effect of *FADS1*/*FADS2* expression in brain tissues and blood on cognitive function. These findings provided evidence to prioritize *FADS* gene as potential target gene for maintenance of cognitive function.

Translational Psychiatry (2024)14:77; <https://doi.org/10.1038/s41398-024-02784-4>

INTRODUCTION

Cognitive dysfunction is an important issue in the aging population [1]. However, changes in brain function start to occur several years before the diagnose of cognitive impairment [2]. Hence, identifying factors associated with the development of cognitive impairment is of great societal interest.

Evidence suggested that fatty acids play an important role in cognition [3]. Previous studies reported that omega-3 fatty acids were associated with cognitive and mental health [4, 5]. However, recent observational studies and randomized controlled trials (RCTs) have shown inconsistent evidence [6–9]. Therefore, other line of evidence is needed to clarify whether there is a causal effect of omega-3 fatty acids on cognitive function. Omega-3 fatty acids were influenced by genetic factors [10–12]. Delta-5 desaturase and delta-6 desaturase are key rate-limiting enzymes

that crucial in a series of elongation and desaturation reactions of omega-3 fatty acids, which are encoded by two genes: fatty acid desaturase 1 (*FADS1*) and fatty acid desaturase 2 (*FADS2*) [13, 14]. Several studies reported the associations between single nucleotide polymorphisms (SNPs) in the *FADS* loci and omega-3 fatty acids concentrations [15–17], implying that variants in the *FADS* gene region modify the activity of polyunsaturated fatty acids desaturation. However, evidence between *FADS1*/*FADS2* gene expression and their own cognitive impairment is limited [18]. In addition, *FADS1* and *FADS2* gene are expressed in multiple human tissues and cells. The role of expression levels of *FADS1* and *FADS2* in different tissues and cell types on cognitive function needs further investigation.

Mendelian randomization (MR) analysis is an emerging method that using genetic variants as instrumental variables (IVs) to infer

¹Department of Endocrine and Metabolic Diseases, Shanghai Institute of Endocrine and Metabolic Diseases, Ruijin Hospital, Shanghai Jiao Tong University School of Medicine, Shanghai, China. ²Shanghai National Clinical Research Center for Endocrine and Metabolic Diseases, Key Laboratory for Endocrine and Metabolic Diseases of the National Health Commission of the PR China, Shanghai National Center for Translational Medicine, Ruijin Hospital, Shanghai Jiao Tong University School of Medicine, Shanghai, China. ³Network and Information Center, Shanghai Jiao Tong University, Shanghai, China. ⁴Shanghai Digital Medicine Innovation Center, Ruijin Hospital, Shanghai Jiao Tong University School of Medicine, Shanghai, China. ⁵MRC Integrative Epidemiology Unit (IEU), Bristol Medical School, University of Bristol, Oakfield House, Oakfield Grove, Bristol BS8 2BN, UK. ⁶These authors contributed equally: Xueyan Wu, Lei Jiang, Hongyan Qi. ✉email: Jie.Zheng@bristol.ac.uk; byf10784@rjh.com.cn; jielilu@hotmail.com

Received: 26 July 2023 Revised: 8 January 2024 Accepted: 16 January 2024

Published online: 05 February 2024

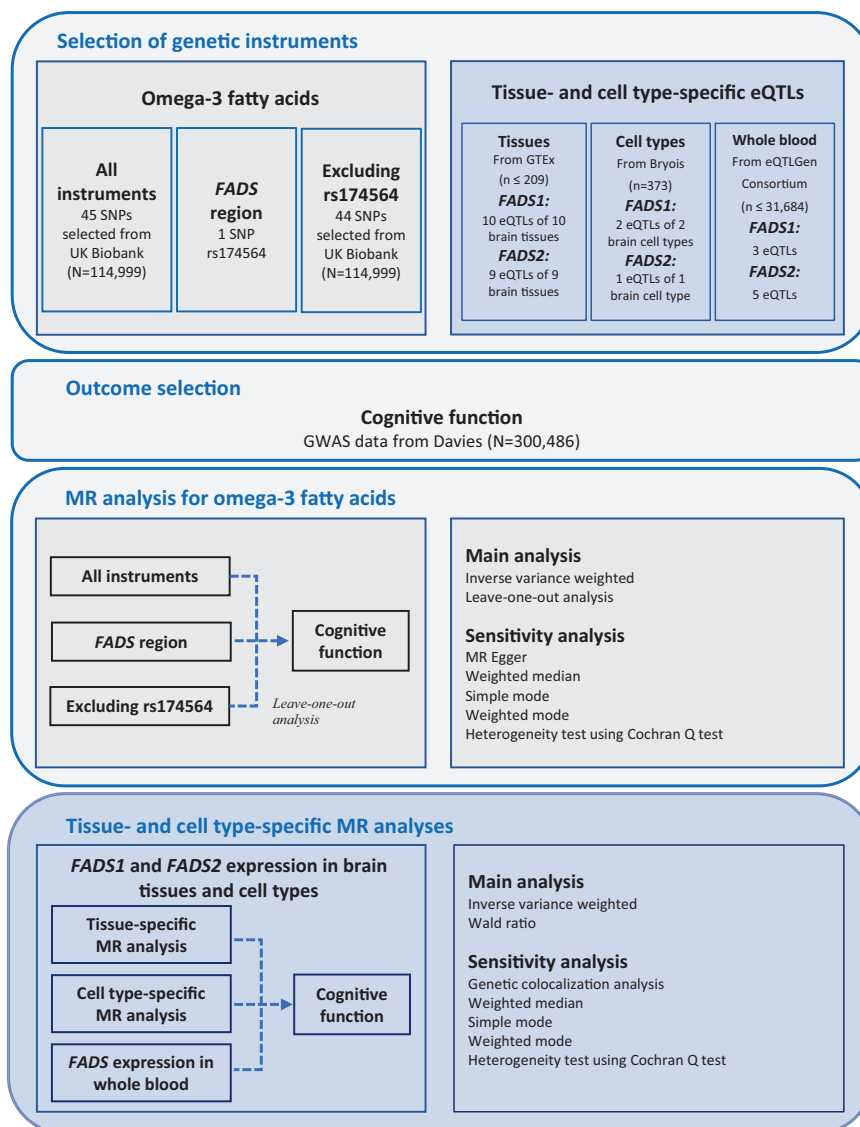


Fig. 1 Flow chat of the whole study design. SNP single nucleotide polymorphism, eQTL expression quantitative trait loci, GWAS Genome-wide association study, MR Mendelian randomization.

the causal effect of an exposure on an outcome [5, 19–21]. Due to specificity of IVs, the MR estimates are not commonly subject to confounding bias and reverse causation [22]. MR has also been applied to detect putative causal effect of tissue-specific gene expression and a wide range of diseases using expression quantitative trait loci (eQTLs) as instruments [23–25]. However, the eQTL relationship was highly dependent on cell type and eQTLs that from bulk tissue samples may mask the cell specificity of genetic regulatory effects [26]. With development of novel omics tools, especially single-cell sequencing technology [27–29] and genetic colocalization methodologies [30], estimating the effect of gene on disease in single-cell level will provide novel insight of disease etiology and molecular mechanism soon. In addition to tissue specificity, recent studies have demonstrated that many eQTL effects are cell type-specific [31], as well as genes showing cell type-specific effects including *FADS1* and *FADS2* [27]. By using eQTLs of diverse cell types will help us to supplement the potential molecular mechanisms that underlie cognitive function.

Therefore, the aim of this study was to investigate the causal effect of omega-3 fatty acids on cognitive function within and outside the *FADS* region by using MR method. To identify potential target gene, the tissue- and cell type-specific causal

effects of *FADS1* and *FADS2* gene expression on cognitive function were evaluated using cis-eQTL-based MR and colocalization.

METHODS

Overall study design

Figure 1 presented the overall design of the study. In this study, i) we applied a two-sample MR analysis to determine whether omega-3 fatty acids have causal effect on cognitive function within and outside the *FADS* region; ii) conducting tissue- and cell type-specific MR analyses to assess tissue- and cell type-dependent effects of *FADS1* and *FADS2* expression in brain and blood on cognitive function. It is important to note that we applied the MR Steiger filtering approach to exclude cis-eQTLs with potential reverse causality [32]. Ethical approval of all data was obtained in the original studies.

Data sources

Genetic instruments of omega-3 fatty acids. Genome-wide association study (GWAS) results in individuals of mostly European ancestry were obtained from the UK Biobank (up to 114,999 individuals) for plasma concentration of omega-3 fatty acids [33]. This is one of the largest available GWASs of circulating polyunsaturated fatty acids. SNPs were excluded if it had a minor allele frequency no more than 0.01 or did not

reach the significant genome-wide association level ($P \leq 5 \times 10^{-8}$) (Supplementary Table 1).

Genetic instruments of FADS1 and FADS2 expression in brain and blood. Brain tissue-specific cis-eQTL data of FADS1 and FADS2 expression was obtained from the GTEx project (v8; <https://gtexportal.org/home/>). For each tissue, the independent cis-eQTL that passed the false discovery rate (FDR) threshold (with $FDR < 0.05$) was selected as instrument for the tissue-specific analysis, which resulted in 10 cis-eQTLs of the 10 tissues for FADS1 gene and nine cis-eQTLs of the nine tissues for FADS2 gene respectively (Supplementary Table 2A). Besides, the results were also validated using the brain cis-eQTL data from the MetaBrain consortium (<https://www.metabrain.nl>), which is a large scale eQTL meta-analysis of previously published human brain eQTL datasets ($N \leq 8,613$) [34]. For consistency, we selected the significant cis-eQTLs ($q\text{-value} < 0.05$) for FADS1 and FADS2 genes with $FDR < 0.05$. After selection, three cis-eQTLs of FADS1 gene derived from three brain tissues were selected (Supplementary Table 2A).

Single-cell cis-eQTL data of FADS1 and FADS2 expression was obtained from a brain cell type cis-eQTL study, which including eight brain cell types from 373 brain samples that published by Bryois et al. [27]. The cis-eQTLs (with $FDR < 0.05$) were identified in two cell types for FADS1 and one cell type for FADS2 expression respectively. Same as tissue-specific instruments, only cis-eQTL with the strongest association for each cell type was selected as instrument for the cell-type specific analysis (Supplementary Table 2B).

The cis-eQTL associations of FADS1 and FADS2 expression derived from whole blood in 31,470 individuals made available by the eQTLGen Consortium [35], and the study included rigorous quality control (Supplementary Table 1).

Outcome data. The GWAS summary statistics of cognitive function was extracted from Davies et al. [3], which included 300,486 individuals of European ancestry from 57 population-based cohorts brought together by the Cohorts for Heart and Aging Research in Genomic Epidemiology (CHARGE), the Cognitive Genomics Consortium (COGENT) consortia, and the UK Biobank. Cognitive function in the three cohorts was estimated by applying a consistent method of extracting a general cognitive function component from cognitive test, which has been reported in more details in the original study [3] (Supplementary Table 3).

Statistical analyses

MR analysis of omega-3 fatty acids on cognitive function. For each omega-3 fatty acids instrument set, we harmonized the SNP-omega-3 fatty acids and SNP-cognitive function data and did the univariable MR analysis by using the TwosampleMR R package (version 0.5.6). In total, 45 SNPs were selected from the UK Biobank as IVs for omega-3 fatty acids, and the primary analysis used the inverse variance weighted (IVW) method to estimate the causal effect.

Leave-one-out analysis: We further conducted leave-one-out analysis and assessed the causal effect of single SNP rs174564 within the FADS region by using the Wald ratio method [36]. Considering the potential effect of rs174564 on cognitive function, we further excluded it from 45 instruments for omega-3 fatty acids to estimate the causal effect of the other variants outside the FADS region on cognitive function.

LD Score regression analysis of omega-3 fatty acids on cognitive function. Considering the GWAS data of omega-3 fatty acids and cognitive function have minor sample overlap, which may induce spurious correlation. We employed linkage disequilibrium score regression (LDSC, v.1.0.1) analysis to evaluate the genetic correlation between omega-3 fatty acids and cognitive function and to test the existence of sample overlap [37, 38]. The LD scores from the European 1000 Genomes Project dataset were referenced [39].

Tissue- and cell type-specific MR analyses

MR analysis of FADS1 and FADS2 expression in brain tissues on cognitive function: For tissue-specific MR analysis, we estimated the putative causal effects of FADS1 expression in 10 brain tissues and FADS2 expression in nine brain tissues using data from the GTEx. MR analysis of FADS1 expression in three brain tissues on cognitive function were also conducted using the MetaBrain data. The Wald ratio [36] method was used since one instrument were available for each tissue. FDR correction was applied using the Benjamini-Hochberg method [40].

MR analysis of FADS1 and FADS2 expression in brain single cell on cognitive function: In cell type-specific MR analysis, the putative causal effects of FADS1 expression in two brain cell types and FADS2 expression in one brain cell type on cognitive function were estimated by using the Wald ratio method [36]. FDR was computed using the Benjamini-Hochberg method [40].

MR analysis of FADS1 and FADS2 expression in whole blood on cognitive function: We further used three cis-eQTLs of FADS1 expression and five cis-eQTLs of FADS2 expression derived from whole blood respectively to estimate the causal effects of expression of these two genes on cognitive function by using IVW method. Moreover, a novel MR method with automated instrument determination (MRAID) was applied [41].

MR sensitivity analysis. We conducted a set of sensitivity analyses to estimate the effects using methods that were robust to other forms of pleiotropy using MR-Egger, weighted median, simple mode, and weighted mode, as each method can obtain consistent estimate of the causal effect if the pleiotropic effect is independent of the effect on the exposure. Cochran's Q test for inverse variance weighted analysis was conducted to assess the presence of heterogeneity between individual SNP [42].

Genetic colocalization analysis. To examine the posterior probability for a shared causal variant between FADS1/FADS2 expression and cognitive function for the candidate MR signal [43], we used a Bayesian colocalization method that is noted as COLOC [30]. A colocalization probability (PP.H4) $> 70\%$ would suggest that the two genetic association signals are likely to share the same causal variant. Besides, we used an approximate colocalization analysis which is called LD check [44]. We estimated the linkage disequilibrium (LD) r^2 between each cis-eQTL against all variants with GWAS $P < 1 \times 10^{-3}$ in the region associated with cognitive function. In this analysis, $r^2 > 0.7$ between each cis-eQTL and cognitive function variants was considered as approximate colocalization.

RESULTS

We selected 45 omega-3 fatty acids variants as instruments, which were selected from Borges CM ($N = 114,999$). Besides, we selected 10 cis-eQTLs and nine cis-eQTLs respectively which is the strongest cis-eQTL for each brain tissue from the GTEx v8 database, and three cis-eQTLs from the MetaBrain data for tissue-specific MR analysis. For brain cell type-specific MR analysis, we used two cis-eQTLs from two cell types and one cis-eQTL from one cell type respectively that published from Bryois ($N = 373$). For instruments of FADS1 and FADS2 expression in whole blood, we selected three cis-eQTLs and five cis-eQTLs respectively that from eQTLGen Consortium ($N \leq 31,684$). All the above cis-eQTLs were tested for Steiger filtering method so that there is no potential reverse causality. For FADS1 and FADS2 expression in different tissues, mean F statistics ranged from 8.5 to 581.5, indicating that most instruments were unlikely to be subject to weak instrument bias. F statistics for hippocampus and substantia nigra is less than 10 (Supplementary Table 2A). For FADS1 and FADS2 expression in different cell types, the F statistics only for inhibitory neurons is less than 10 (Supplementary Table 2B). We kept all instruments but with caution that three of these cis-eQTL dataset could suffer from weak instrument bias.

Effect of omega-3 fatty acids on cognitive function

We investigated the causal effect of omega-3 fatty acids on cognitive function using genetic variants within and outside the FADS region. Little evidence was observed to support a causal effect using the IVW method [$\beta = -6 \times 10^{-3}$, 95% confidence interval (CI) $(-1.8 \times 10^{-2}, 6 \times 10^{-3})$, $P = 3.3 \times 10^{-1}$], although weighted median and weighted mode estimates suggested potential causal effects [$\beta = -1.2 \times 10^{-2}$, 95% CI $(-2 \times 10^{-2}, -4 \times 10^{-3})$, $P = 3 \times 10^{-3}$; $\beta = -1.3 \times 10^{-2}$, 95% CI $(-2.1 \times 10^{-2}, -5 \times 10^{-3})$, $P = 3 \times 10^{-3}$, respectively] (Fig. 2A). Besides, strong evidence of heterogeneity was observed for the overall effect of omega-3 fatty acids on cognitive function (P -value of the Q

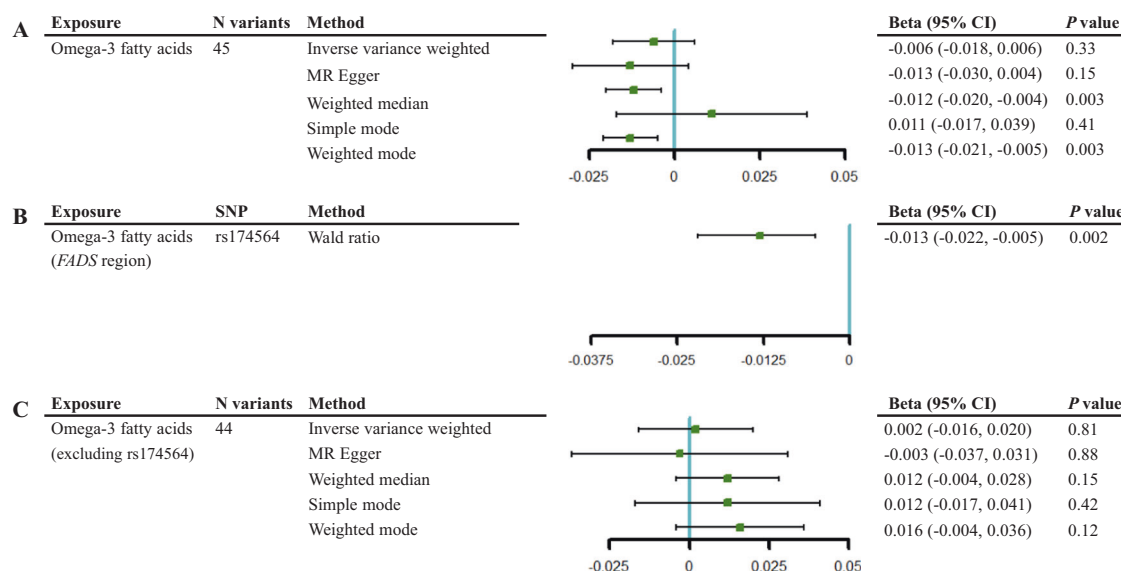


Fig. 2 Mendelian randomization analysis of the causal effect of omega-3 fatty acids on cognitive function within and outside the *FADS* region. **A** All instruments of omega-3 fatty acids on cognitive function. **B** Single SNP within the *FADS* region of omega-3 fatty acids on cognitive function. **C** SNPs outside the *FADS* region of omega-3 fatty acids on cognitive function. The vertical line in this plot indicates the null of beta = 0 and the error bars correspond to 95% confidence intervals. CI confidence interval.

test = 5.5×10^{-17}) (Supplementary Table 4). Specially, leave-one-out analysis indicated that the potential effect on cognitive function was driven by a single variant, rs174564, within the *FADS* region [$\beta = -1.3 \times 10^{-2}$, 95% CI (-2.2×10^{-2} , -5×10^{-3}), $P = 2 \times 10^{-3}$] (Fig. 2B, Supplementary Fig. 1). The estimated effect using instruments outside the *FADS* region showed little evidence by using IVW and the other sensitivity MR methods ($P > 0.05$) (Fig. 2C). In addition, the LDSC results showed that there was no genome-wide genetic correlation between omega-3 fatty acids and cognitive function after controlling for sample overlap (intercept = -1.3×10^{-2} , $P = 0.36$, Supplementary Table 5).

Tissue- and cell type-specific effect of *FADS1* and *FADS2* expression on cognitive function

Due to the key role of *FADS* gene on cognitive function, we investigated the tissue- and cell type-specific causal effect of *FADS1* and *FADS2* expression on cognitive function (Figs. 3 and 4). As brain is closely related to cognitive function, we focused on explored the causal effect of *FADS1* and *FADS2* gene expression on cognitive function using cis-eQTL data from 10 and nine brain tissues respectively (e.g., amygdala, cortex, etc). The MR and colocalization analyses suggested putative causal effects of *FADS1* expression in 10 brain tissues and *FADS2* expression in one brain tissue on cognitive function, and these associations passed FDR threshold of 0.05: increased expression levels of *FADS1* gene in cerebellar hemisphere showed a cognitive function maintenance effect. While, decreased expression levels of *FADS1* in nine additional brain tissues showed effects on maintaining cognitive function, including cerebellum, spinal cord cervical c-1, hypothalamus, cortex, hippocampus, putamen basal ganglia, anterior cingulate cortex BA24, caudate basal ganglia, and frontal Cortex BA9. In addition, the significant results with colocalization evidence for cerebellum, cortex and hippocampus were validated in the MetaBrain data and were all directionally consistent with the MR effects in GTEx (Fig. 3A). For *FADS2*, increased expression levels in nucleus accumbens basal ganglia showed a possible maintenance effect of cognitive function. The MR and colocalization results suggested little evidence to support causality for *FADS2* in other eight brain tissues (Fig. 3B, Supplementary Table 6).

Secondly, we estimated the cell type-specific causal effect of gene expression of *FADS1* and *FADS2* on cognitive function using

brain single-cell cis-eQTL data. *FADS1* and *FADS2* expression in one cell type showed MR and colocalization evidence: decreased levels of *FADS1* and *FADS2* expression in oligodendrocytes showed cognitive function maintenance effect. Causal effect of *FADS1* expression on cognitive function was not observed in inhibitory neurons (Fig. 4, Supplementary Table 7).

In order to further verified the role of *FADS1* and *FADS2* gene expression on cognitive function in whole blood, we estimated the causal effect using three cis-eQTLs for *FADS1* and five cis-eQTLs for *FADS2* respectively. MR analysis indicated that increased expression levels of *FADS1* and *FADS2* in whole blood showed effects on cognitive function maintenance [IVW $\beta = 9 \times 10^{-3}$, 95% CI (3×10^{-3} , 1.5×10^{-2}), $P = 5 \times 10^{-3}$; IVW $\beta = 5 \times 10^{-3}$, 95% CI (1×10^{-4} , 1×10^{-2}), $P = 4.6 \times 10^{-2}$; respectively]. In sensitivity analysis, weighted median suggested that increased expression levels of *FADS1* was associated with maintenance of cognitive function [$\beta = 9 \times 10^{-3}$, 95% CI (3×10^{-3} , 1.5×10^{-2}), $P = 4 \times 10^{-3}$], while the estimates showed little causal evidence using other sensitivity MR methods (Fig. 5). Little evidence of heterogeneity was observed (P -value of all the Q test > 0.05) (Supplementary Table 4). The MRSAID method showed that *FADS1* expression in whole blood had robust causal effect on cognitive function ($P = 0.02$) and directionally consistent with the MR effects from the IVW method, while the causal effect of *FADS2* were not observed (Supplementary Table 8). After performing colocalization analysis with the candidate MR signal, we observed compelling evidence of gene colocalization between expression of *FADS1* and *FADS2* and cognitive function (PP.H4 = 86.6% and 88.4%, respectively) (Fig. 6, Supplementary Table 9).

Furthermore, to understand the link between omega-3 fatty acids variants and *FADS* gene variants, we estimated the LD between them. The omega-3 fatty acids variant rs174564 is located in the intron of *FADS2* gene and it is in strong LD ($r^2 > 0.7$) with several of the *FADS1* and *FADS2* cis-eQTLs/instruments we used. This suggested that omega-3 fatty acids and *FADS1*/*FADS2* cis-eQTLs are likely to represent the same genetic signal in the *FADS* region. Therefore, the effect of *FADS1*/*FADS2* expression on cognitive function could be related to the omega-3 fatty acids variant rs174564. The pairwise LD r^2 between each cis-eQTL and rs174564 was presented in Supplementary Table 10.

Finally, we attempted to identify potential mechanistic pathways between omega-3 fatty acids, *FADS1*/*FADS2* genes, and

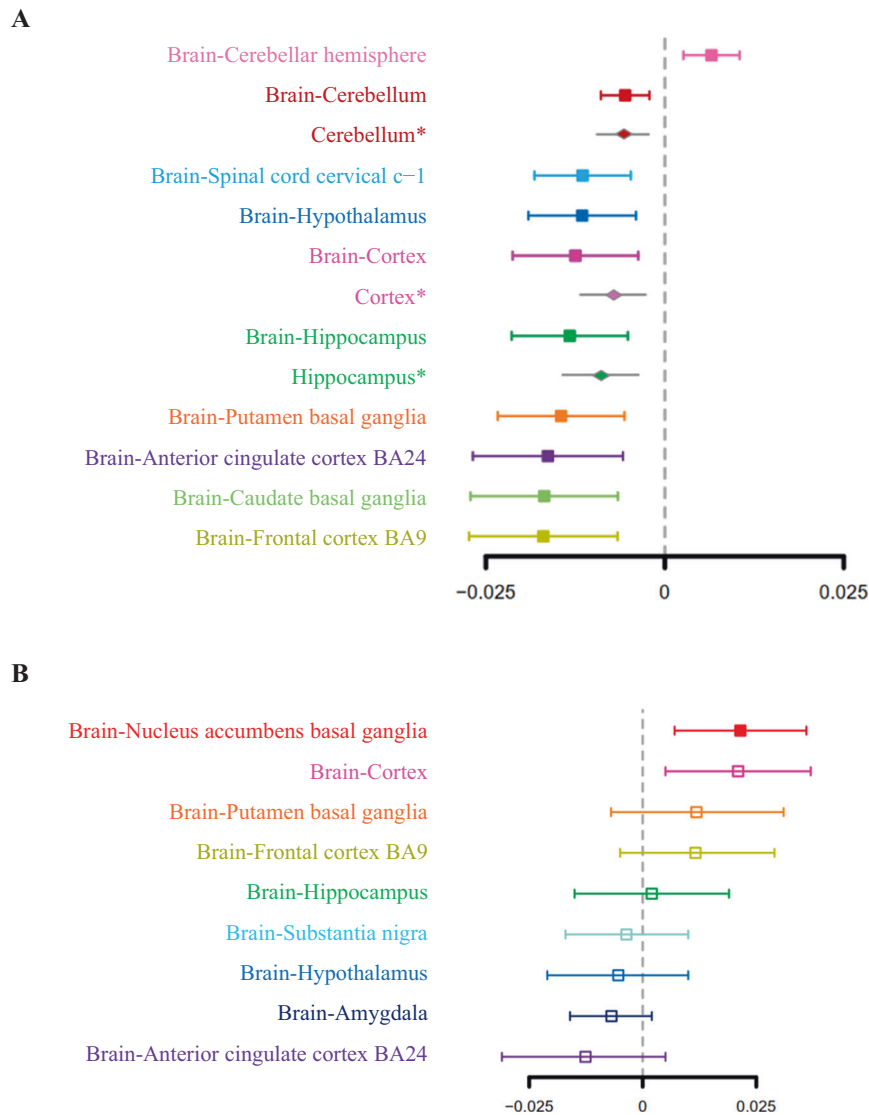


Fig. 3 Forest plot illustrating the brain tissue-dependent association for *FADS1* and *FADS2* expression on cognitive function. **A** *FADS1*. **B** *FADS2*. The vertical line in this plot indicates the null of $\beta = 0$ and the error bars correspond to 95% confidence intervals. Solid squares represented results that passed the LD check, while hollow squares represented results that failed the LD check. Asterisks represented results using the MetaBrain database.

cognitive function through MELODI Presto [45]. The results showed that potential intermediates between omega-3 fatty acids or *FADS1/FADS2* genes and cognitive function were mostly associated with metabolic or neurological diseases, such as non-insulin dependent diabetes, metabolic syndrome, obesity and Alzheimer's disease (Supplementary Table 11).

DISCUSSION

In this study, we found that *FADS1* and *FADS2* expression in different brain tissues and cell types showed causal effect on cognitive function using genetic tools. Data on the expression of *FADS1* and *FADS2* in whole blood further confirmed this finding. Our results revealed that *FADS1* and *FADS2* are likely to be two causal genes influencing cognitive function, while the *FADS* gene as potential target gene, may be functional especially in specific cell type.

Previous epidemiology studies reported a protective effect of omega-3 fatty acids on cognitive function, and this effect is particularly pronounced in individuals with early and mild cognitive impairment [46]. However, no benefit was observed

when subjects with diagnosed Alzheimer's disease were supplemented with omega-3 fatty acids as well as in many other population-based studies [47–49]. These inconsistencies may be attributed to interventions in RCTs that have been carried out too late to against the progression of cognitive impairment and are vulnerable to confounding factors. In our MR analysis, we found a weak negative association between omega-3 fatty acids and cognitive function using some MR methods, while the sensitivity analysis suggested strong evidence of heterogeneity, which suggested that a few genes may drive the causal effect between the two. Leave-one-out analysis further suggested that the effect of omega-3 fatty acids on cognitive function is more likely to be driven by SNP within the *FADS* gene region rather than a general effect of omega-3 fatty acids.

As a natural extension, we investigated the impact of *FADS1* and *FADS2* gene expression on cognitive function. The *FADS* variants have been reported to be associated with cognitive function in previous studies, but the association has only been studied in the context of the effect of *FADS* gene variation on children or offspring [50–53]. Genetic variants in the *FADS1/FADS2* region are associated with

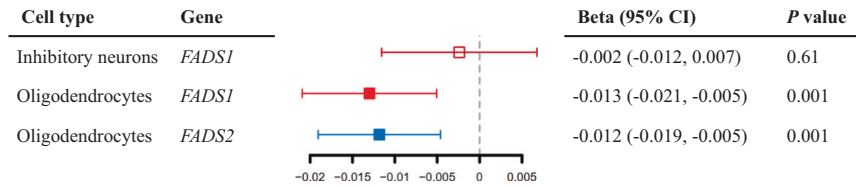


Fig. 4 Forest plot illustrating the causal effect association for single cell gene expression of *FADS1* and *FADS2* on cognitive function. The vertical line in this plot indicates the null of beta = 0 and the error bars correspond to 95% confidence intervals. Solid squares represented results that passed the LD check, while hollow squares represented results that failed the LD check. CI confidence interval.

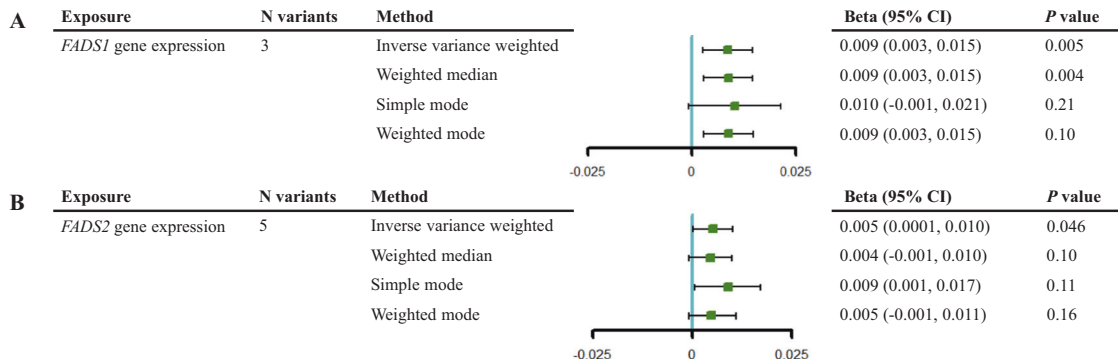


Fig. 5 Mendelian randomization analysis of the causal effect of *FADS1* and *FADS2* gene expression in whole blood on cognitive function. **A** *FADS1*. **B** *FADS2*. The vertical line in this plot indicates the null of beta = 0 and the error bars correspond to 95% confidence intervals. CI confidence interval.

maternal long-chain polyunsaturated fatty acid status and could modified cognitive development of infants [50]. In addition, *FADS1*/*FADS2* genetic variants have been reported to be associated with behavioral outcomes in children [52, 53]. However, one issue to be resolved is whether there is an association between *FADS* gene expression and cognitive function, and whether this association is influenced by tissue type, especially brain tissue. Our tissue-specific MR analysis showed that increased levels of *FADS1* expression in cerebellar hemisphere and *FADS2* expression in nucleus accumbens basal ganglia may maintain cognitive function, while decreased levels of *FADS1* expression in other nine brain tissues, including cerebellum, spinal cord cervical c-1, hypothalamus, cortex, hippocampus, putamen basal ganglia, anterior cingulate cortex BA24, caudate basal ganglia, and frontal Cortex BA9, may benefit cognitive function maintenance. It is accepted that the cerebellum played a possible role in the mediation of cognitive processes [54]. Previous studies have showed that individuals with Parkinson's disease had significant atrophy of left cerebellar hemisphere [55, 56]. A broad variety of cognitive and linguistic deficits can occur after cerebellar damage [57–59]. The most popular mechanism of cerebellar involvement in cognitive functions is Schmahmann's dysmetria of thought theory, which assuming that the way the cerebellum regulates movement may also influence mental processes [60]. Besides, basal ganglia are critical for several cognitive, motor and emotional functions and are part of a complex functional circuit [61–63]. Early animal experiments confirmed the relationship between basal ganglia and cognitive and memory function, which pointed out that this relationship may be related to the cholinergic neuronal impulse transmission in the basal ganglia and the role of dopamine neurons for reward learning [64, 65]. Human studies have also reported that basal ganglia may play an integrative role in cognitive information processing and that the electrical activity of multifunctional clusters of neuronal populations may underlie this nonspecific integrative effect [66]. In this study, we revealed a putative causal mechanism that increased expression levels of *FADS1* gene in cerebellar hemisphere and *FADS2* gene in nucleus accumbens basal ganglia are associated with maintenance of cognitive function.

It is important to notice that eQTL effect of the same gene could be different dependent on the tissues or cell types of the human

brain. In tissue-specific MR analysis, we observed that both *FADS1* and *FADS2* expression in cortex showed MR evidence, which decreased *FADS1* expression levels and increased *FADS2* expression levels showed maintenance effect on cognitive function. However, the causal effect of *FADS2* were not confirmed by colocalization evidence. Similar with cortex, decreased expression levels of *FADS1* in anterior cingulate cortex BA24 and frontal cortex BA9 was associated with maintaining cognitive function. This directional inconsistency may be due to the limitation of tissue sample size or there may be different pathways involved in *FADS1*/*FADS2* expression in cortex on cognition. More datasets of larger independent tissue-specific eQTL data and additional genetic methods, such as transcriptome-wide association study (TWAS), should be considered in future studies to further improve the statistical power and identify true causal genes with functions [67–70]. Previous studies have affirmed the role of the anterior cingulate cortex and frontal cortex in social cognition and cognitive control [55, 56, 71, 72], and we supplied new evidence for this association at genetic level. Furthermore, our cell type-specific MR analysis used single-cell brain cis-eQTL data highlighted the important role of *FADS1*/*FADS2* gene in oligodendrocytes. Recently, Kenigsbuch et al. [73] confirmed that oligodendrocyte state was associated with brain pathologies among multiple central nervous system diseases. Our findings provided new evidence that decreased expression levels of *FADS1*/*FADS2* in oligodendrocytes could influence cognitive function. The potential mechanism causing the differences between tissues and cells need further investigation.

To further verify our findings, we also used cis-eQTL data in whole blood and found a protective effect of *FADS1* and *FADS2* expression in blood on cognition. Additionally, the causal effect of *FADS1* gene was also confirmed by MR-IVW method, which provided additional evidence to prove the robustness of this finding. As one of the main MR approaches, the IVW method relies on pre-selected independent SNPs as instruments for MR analysis and could not account for horizontal pleiotropy [74]. MR-IVW uses multiple correlated genetic variants and account for correlated and uncorrelated pleiotropy [41]. While, the causal effect of *FADS2* gene was only observed using the IVW method. Despite the good

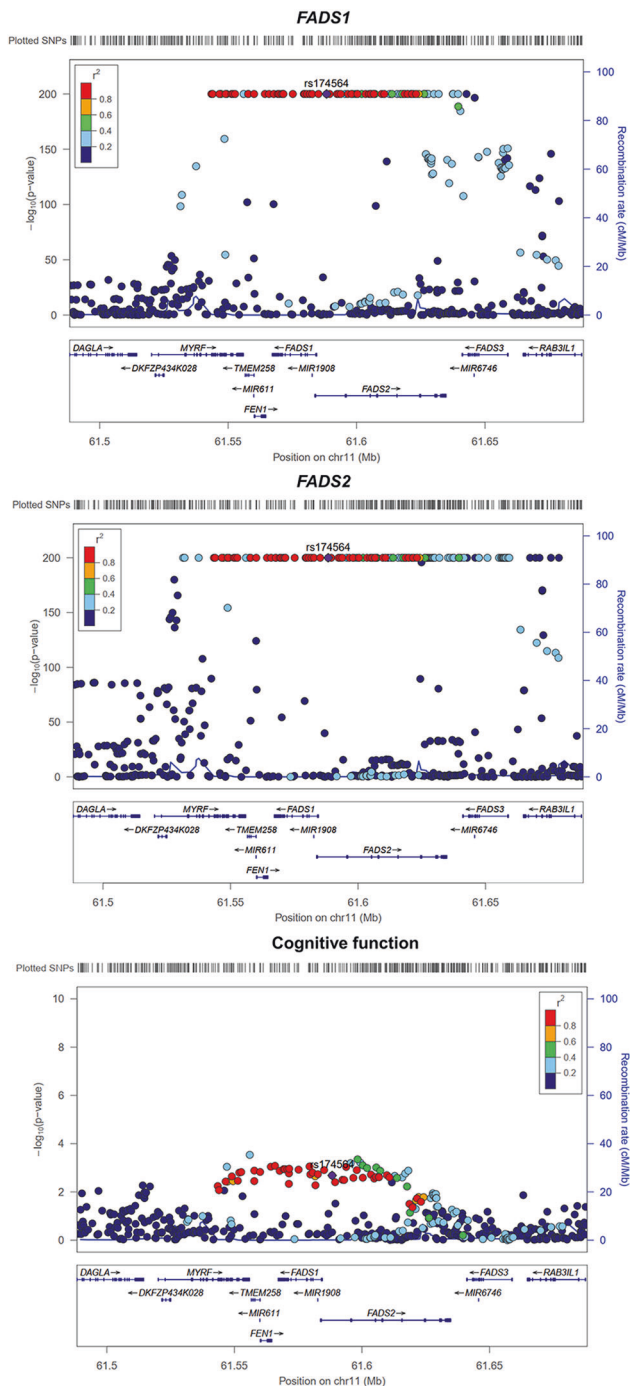


Fig. 6 Regional association plots of *FADS1* and *FADS2* expression in whole blood on cognitive function in the *FADS* region. The candidate signal within *FADS* region is rs174564.

statistical power from both IVW or MR-IVW analysis [41], the effect of *FADS2* on cognitive function needs to be investigated in future studies. Importantly, our colocalization evidence confirmed the causal effect of *FADS1/FADS2* expression levels on cognitive function. It is well known that *FADS1* and *FADS2* polymorphisms could modulate fatty acid metabolism [75]. The results from MELODI Presto also identified and prioritized metabolic and neurological diseases as potential intermediates between omega-3 fatty acids or *FADS1/FADS2* genes and cognitive function, which provided direction for future mechanistic studies.

There are some strengths in our study. First, we have used large-scale GWAS data of omega-3 fatty acids, tissue and single-cell sequencing cis-eQTL data of gene expression and GWAS data of cognitive function, which brought good instrument strength and statistical power to our study. Second, traditional studies tend to focus on the association between omega-3 fatty acids and cognition, while we proposed for the first time that *FADS1* and *FADS2* expression in multiple brain tissues and cell types had different effect on cognitive function. Third, we have supplemented the mechanism of cognition at the genetic level by providing evidence to prioritize *FADS1* and *FADS2* as two potential target genes on cognition, which could be functional in brain.

Our study has several limitations. Firstly, the instruments of omega-3 fatty acids and a small proportion of the outcome samples were obtained from the UK Biobank, which have minor sample overlap issue. However, there was no sample overlap between cis-eQTL data and the outcome GWAS, which means the vast majority of the MR results will not be influenced by the sample overlap issue. Secondly, there were limited number of instruments for the cis-eQTL data, which means most of the MR sensitivity methods such as MR-Egger were not applicable. However, we systematically conducted colocalization analysis to enhance the causal evidence of our findings. Thirdly, the *F* statistics of hippocampus, substantia nigra and inhibitory neurons were lower than the common threshold of 10, the weak instrument bias need to be carefully considered when interpreting the findings. However, our top findings were observed in oligodendrocytes, which showed good instrument strength. Large-scale single-cell eQTL studies are needed in the future to provide better statistical power.

In conclusion, our MR analysis showed novel insight between *FADS1/FADS2* gene expression and cognitive function by using tissue and single cell cis-eQTL data and state-of-the-art methods such as genetic colocalization. Integrating these novel data and methods suggested that *FADS1* and *FADS2* expression levels could influence cognitive function in different brain tissues and cell types. Our results provided clues for the understanding of the genetic mechanism of cognitive function and improved the current knowledge of *FADS* gene and cognition. Future studies are needed to prioritize *FADS1/FADS2* as potential target genes for maintenance of cognitive function.

DATA AVAILABILITY

GWAS data of omega-3 fatty acids are available from the UK Biobank (<https://www.ukbiobank.ac.uk>). Brain tissue cis-eQTL data of *FADS* gene can be obtained from the GTEx project (v8; <https://gtexportal.org/home>) and the MetaBrain consortium (<https://www.metabrain.nl>). Single-cell sequencing cis-eQTL data of *FADS* gene can be accessed from the respective publication. The cis-eQTL data of *FADS* gene expression in whole blood are available on the eQTLGen Consortium (<https://eqtlgen.org>). The GWAS summary statistics for cognitive function are available from the respective publication.

CODE AVAILABILITY

The main statistical analyses were conducted using TwoSampleMR R package (v.0.5.6). Colocalization analysis was conducted using coloc R package (v.5.2.0).

REFERENCES

- Lövdén M, Fratiglioni L, Glymour MM, Lindenberg U, Tucker-Drob EM. Education and Cognitive Functioning Across the Life Span. *Psychol Sci. Public Interest.* 2020;21:6–41.
- Beason-Held LL, Goh JO, An Y, Kraut MA, O'Brien RJ, Ferrucci L, et al. Changes in brain function occur years before the onset of cognitive impairment. *J Neurosci.* 2013;33:18008–14.
- Davies G, Lam M, Harris SE, Trampush JW, Luciano M, Hill WD, et al. Study of 300,486 individuals identifies 148 independent genetic loci influencing general cognitive function. *Nat Commun.* 2018;9:2098.

4. Daiello LA, Gongvatana A, Dunsiger S, Cohen RA, Ott BR. Association of fish oil supplement use with preservation of brain volume and cognitive function. *Alzheimers Dement*. 2015;11:226–35.
5. Jones HJ, Borges MC, Carnegie R, Mongan D, Rogers PJ, Lewis SJ, et al. Associations between plasma fatty acid concentrations and schizophrenia: a two-sample Mendelian randomisation study. *Lancet Psychiatry*. 2021;8:1062–70.
6. Alex A, Abbott KA, McEvoy M, Schofield PW, Garg ML. Long-chain omega-3 polyunsaturated fatty acids and cognitive decline in non-demented adults: a systematic review and meta-analysis. *Nutr Rev*. 2020;78:563–78.
7. Øyen J, Kvestad I, Midtbø LK, Graff IE, Hysing M, Stormark KM, et al. Fatty fish intake and cognitive function: FINS-KIDS, a randomized controlled trial in pre-school children. *BMC Med*. 2018;16:41.
8. Andrieu S, Guyonnet S, Coley N, Cantet C, Bonnefoy M, Bordes S, et al. Effect of long-term omega 3 polyunsaturated fatty acid supplementation with or without multidomain intervention on cognitive function in elderly adults with memory complaints (MAPT): a randomised, placebo-controlled trial. *Lancet Neurol*. 2017;16:377–89.
9. Wood AHR, Chappell HF, Zullyniak MA. Dietary and supplemental long-chain omega-3 fatty acids as moderators of cognitive impairment and Alzheimer's disease. *Eur J Nutr*. 2022;61:589–604.
10. Lemaitre RN, Tanaka T, Tang W, Manichaikul A, Foy M, Kabagambe EK, et al. Genetic loci associated with plasma phospholipid n-3 fatty acids: a meta-analysis of genome-wide association studies from the CHARGE Consortium. *PLoS Genet*. 2011;7:e1002193.
11. Shin SY, Fauman EB, Petersen AK, Krumsiek J, Santos R, Huang J, et al. An atlas of genetic influences on human blood metabolites. *Nat Genet*. 2014;46:543–50.
12. Wu JH, Lemaitre RN, Manichaikul A, Guan W, Tanaka T, Foy M, et al. Genome-wide association study identifies novel loci associated with concentrations of four plasma phospholipid fatty acids in the de novo lipogenesis pathway: results from the Cohorts for Heart and Aging Research in Genomic Epidemiology (CHARGE) consortium. *Circ Cardiovasc Genet*. 2013;6:171–83.
13. Nakamura MT, Nara TY. Structure, function, and dietary regulation of delta6, delta5, and delta9 desaturases. *Annu Rev Nutr*. 2004;24:345–76.
14. Tosi F, Sartori F, Guarini P, Olivieri O, Martinelli N. Delta-5 and delta-6 desaturases: crucial enzymes in polyunsaturated fatty acid-related pathways with pleiotropic influences in health and disease. *Adv Exp Med Biol*. 2014;824:61–81.
15. Malerba G, Schaeffer L, Xumerle L, Klopp N, Trabetti E, Biscola M, et al. SNPs of the FADS gene cluster are associated with polyunsaturated fatty acids in a cohort of patients with cardiovascular disease. *Lipids*. 2008;43:289–99.
16. Rzehak P, Heinrich J, Klopp N, Schaeffer L, Hoff S, Wolfram G, et al. Evidence for an association between genetic variants of the fatty acid desaturase 1 fatty acid desaturase 2 (FADS1 FADS2) gene cluster and the fatty acid composition of erythrocyte membranes. *Br J Nutr*. 2009;101:20–6.
17. Xie L, Innis SM. Genetic variants of the FADS1 FADS2 gene cluster are associated with altered (n-6) and (n-3) essential fatty acids in plasma and erythrocyte phospholipids in women during pregnancy and in breast milk during lactation. *J Nutr*. 2008;138:2222–8.
18. Schuchardt JP, Köbe T, Witte V, Willers J, Gingrich A, Tesky V, et al. Genetic Variants of the FADS Gene Cluster Are Associated with Erythrocyte Membrane LC PUFA Levels in Patients with Mild Cognitive Impairment. *J Nutr Health Aging*. 2016;20:611–20.
19. Zhou S, Zhu G, Xu Y, Gao R, Li H, Han G, et al. Mendelian Randomization Study on the Putative Causal Effects of Omega-3 Fatty Acids on Low Back Pain. *Front Nutr*. 2022;9:819635.
20. Luo J, le Cessie S, Blauw GJ, Franceschi C, Noordam R, van Heemst D. Systemic inflammatory markers in relation to cognitive function and measures of brain atrophy: a Mendelian randomization study. *Geroscience*. 2022;44:2259–70.
21. Smith GD, Ebrahim S. 'Mendelian randomization': can genetic epidemiology contribute to understanding environmental determinants of disease? *Int J Epidemiol*. 2003;32:1–22.
22. Emdin CA, Khera AV, Kathiresan S. Mendelian Randomization. *JAMA*. 2017;318:1925–6.
23. Taylor K, Davey Smith G, Relton CL, Gaunt TR, Richardson TG. Prioritizing putative influential genes in cardiovascular disease susceptibility by applying tissue-specific Mendelian randomization. *Genome Med*. 2019;11:6.
24. Richardson TG, Hemani G, Gaunt TR, Relton CL & Davey Smith G. A transcriptome-wide Mendelian randomization study to uncover tissue-dependent regulatory mechanisms across the human phenome. *Nat Commun*. 2020;11:185.
25. Khankari NK, Keaton JM, Walker VM, Lee KM, Shuey MM, Clarke SL, et al. Using Mendelian randomisation to identify opportunities for type 2 diabetes prevention by repurposing medications used for lipid management. *EBioMedicine*. 2022;80:104038.
26. Neavin D, Nguyen Q, Daniszewski MS, Liang HH, Chiu HS, Wee YK, et al. Single cell eQTL analysis identifies cell type-specific genetic control of gene expression in fibroblasts and reprogrammed induced pluripotent stem cells. *Genome Biol*. 2021;22:76.
27. Bryois J, Calini D, Macnair W, Foo L, Ulrich E, Ortmann W, et al. Cell-type-specific cis-eQTLs in eight human brain cell types identify novel risk genes for psychiatric and neurological disorders. *Nat Neurosci*. 2022;25:1104–12.
28. Soskic B, Cano-Gamez E, Smyth DJ, Ambridge K, Ke Z, Matte JC, et al. Immune disease risk variants regulate gene expression dynamics during CD4(+) T cell activation. *Nat Genet*. 2022;54:817–26.
29. Yazari S, Alquicira-Hernandez J, Wing K, Senabouth A, Gordon MG, Andersen S, et al. Single-cell eQTL mapping identifies cell type-specific genetic control of autoimmune disease. *Science*. 2022;376:eabf3041.
30. Giambartolomei C, Vukcevic D, Schadt EE, Franke L, Hingorani AD, Wallace C, et al. Bayesian test for colocalisation between pairs of genetic association studies using summary statistics. *PLoS Genet*. 2014;10:e1004383.
31. Ishigaki K, Kochi Y, Suzuki A, Tsuchida Y, Tsuchiya H, Sumitomo S, et al. Polygenic burdens on cell-specific pathways underlie the risk of rheumatoid arthritis. *Nat Genet*. 2017;49:1120–5.
32. Hemani G, Tilling K, Davey Smith G. Orienting the causal relationship between imprecisely measured traits using GWAS summary data. *PLoS Genet*. 2017;13:e1007081.
33. Borges MC, Haycock PC, Zheng J, Hemani G, Holmes MV, Davey Smith G, et al. Role of circulating polyunsaturated fatty acids on cardiovascular diseases risk: analysis using Mendelian randomization and fatty acid genetic association data from over 114,000 UK Biobank participants. *BMC Med*. 2022;20:210.
34. de Klein N, Tsai EA, Vochteloo M, Baird D, Huang Y, Chen CY, et al. Brain expression quantitative trait locus and network analyses reveal downstream effects and putative drivers for brain-related diseases. *Nat Genet*. 2023;55:377–88.
35. Vösa U, Claringbould A, Westra HJ, Bonder MJ, Deelen P, Zeng B, et al. Large-scale cis- and trans-eQTL analyses identify thousands of genetic loci and polygenic scores that regulate blood gene expression. *Nat Genet*. 2021;53:1300–10.
36. Lawlor DA, Harbord RM, Sterne JA, Timpson N. & Davey Smith G. Mendelian randomization: using genes as instruments for making causal inferences in epidemiology. *Stat Med*. 2008;27:1133–63.
37. Bulik-Sullivan BK, Loh PR, Finucane HK, Ripke S, Yang J, Patterson N, et al. LD Score regression distinguishes confounding from polygenicity in genome-wide association studies. *Nat Genet*. 2015;47:291–5.
38. Bulik-Sullivan B, Finucane HK, Anttila V, Gusev A, Day FR, Loh PR, et al. An atlas of genetic correlations across human diseases and traits. *Nat Genet*. 2015;47:1236–41.
39. Hemani G, Zheng J, Elsworth B, Wade KH, Haberland V, Baird D, et al. The MR-Base platform supports systematic causal inference across the human phenome. *Elife*. 2018;7:e34408.
40. Benjamini Y, Hochberg Y. Controlling the False Discovery Rate: A Practical and Powerful Approach to Multiple Testing. *J R Stat Soc Ser B*. 1995;57:289–300.
41. Yuan Z, Liu L, Guo P, Yan R, Xue F, Zhou X. Likelihood-based Mendelian randomization analysis with automated instrument selection and horizontal pleiotropic modeling. *Sci Adv*. 2022;8:eabl5744.
42. Bowden J, Del Greco MF, Minelli C, Davey Smith G, Sheehan N, Thompson J. A framework for the investigation of pleiotropy in two-sample summary data Mendelian randomization. *Stat Med*. 2017;36:1783–802.
43. Hormozdizari F, van de Bunt M, Segrè AV, Li X, Joo JWJ, Bilow M, et al. Colocalization of GWAS and eQTL Signals Detects Target Genes. *Am J Hum Genet*. 2016;99:1245–60.
44. Zheng J, Haberland V, Baird D, Walker V, Haycock PC, Hurler MR, et al. Phenome-wide Mendelian randomization mapping the influence of the plasma proteome on complex diseases. *Nat Genet*. 2020;52:1122–31.
45. Elsworth B, Gaunt TR. MELODI Presto: a fast and agile tool to explore semantic triples derived from biomedical literature. *Bioinformatics* 2021;37:583–5.
46. Zhu RZ, Chen MQ, Zhang ZW, Wu TY, Zhao WH. Dietary fatty acids and risk for Alzheimer's disease, dementia, and mild cognitive impairment: A prospective cohort meta-analysis. *Nutrition* 2021;90:111355.
47. Bischoff-Ferrari HA, Vellas B, Rizzoli R, Kressig RW, da Silva JAP, Blauth M, et al. Effect of Vitamin D Supplementation, Omega-3 Fatty Acid Supplementation, or a Strength-Training Exercise Program on Clinical Outcomes in Older Adults: The DO-HEALTH Randomized Clinical Trial. *JAMA*. 2020;324:1855–68.
48. Brainard JS, Jimoh OF, Deane KHO, Biswas P, Donaldson D, Maas K, et al. Omega-3, Omega-6, and Polyunsaturated Fat for Cognition: Systematic Review and Meta-analysis of Randomized Trials. *J Am Med Dir Assoc*. 2020;21:1439–e21.
49. Lin PY, Cheng C, Satyanarayanan SK, Chiu LT, Chien YC, Chuu CP, et al. Omega-3 fatty acids and blood-based biomarkers in Alzheimer's disease and mild cognitive impairment: A randomized placebo-controlled trial. *Brain Behav Immun*. 2022;99:289–98.
50. Gonzalez Casanova I, Schoen M, Tandon S, Stein AD, Barraza Villarreal A, DiGirolamo AM, et al. Maternal FADS2 single nucleotide polymorphism modified the impact of prenatal docosahexaenoic acid (DHA) supplementation on child neurodevelopment at 5 years: Follow-up of a randomized clinical trial. *Clin Nutr*. 2021;40:5339–45.

51. Yeates AJ, Love TM, Engström K, Mulhern MS, McSorley EM, Grzesik K, et al. Genetic variation in FADS genes is associated with maternal long-chain PUFA status but not with cognitive development of infants in a high fish-eating observational study. *Prostaglandins Leukot Ess Fat Acids*. 2015;102-3:13–20.
52. Brookes KJ, Chen W, Xu X, Taylor E, Asherson P. Association of fatty acid desaturase genes with attention-deficit/hyperactivity disorder. *Biol Psychiatry*. 2006;60:1053–61.
53. Jensen HA, Harsløf LB, Nielsen MS, Christensen LB, Ritz C, Michaelsen KF, et al. FADS single-nucleotide polymorphisms are associated with behavioral outcomes in children, and the effect varies between sexes and is dependent on PPAR genotype. *Am J Clin Nutr*. 2014;100:826–32.
54. Murdoch BE. The cerebellum and language: historical perspective and review. *Cortex*. 2010;46:858–68.
55. Ridderinkhof KR, Ullsperger M, Crone EA, Nieuwenhuis S. The role of the medial frontal cortex in cognitive control. *Science*. 2004;306:443–7.
56. Amodio DM, Frith CD. Meeting of minds: the medial frontal cortex and social cognition. *Nat Rev Neurosci*. 2006;7:268–77.
57. Baillieux H, De Smet HJ, Dobbeleir A, Paquier PF, De Deyn PP, Mariën P. Cognitive and affective disturbances following focal cerebellar damage in adults: a neuropsychological and SPECT study. *Cortex*. 2010;46:869–79.
58. Richter S, Gerwig M, Aslan B, Wilhelm H, Schoch B, Dimitrova A, et al. Cognitive functions in patients with MR-defined chronic focal cerebellar lesions. *J Neurol*. 2007;254:1193–203.
59. Starowicz-Filip A, Prochwicz K, Kłosowska J, Chrobak AA, Myszk A, Bętkowska-Korpała B, et al. Cerebellar Functional Lateralization From the Perspective of Clinical Neuropsychology. *Front Psychol*. 2021;12:775308.
60. Schmahmann JD. From movement to thought: anatomic substrates of the cerebellar contribution to cognitive processing. *Hum Brain Mapp*. 1996;4:174–98.
61. Hampton AN, Bossaerts P, O'Doherty JP. The role of the ventromedial prefrontal cortex in abstract state-based inference during decision making in humans. *J Neurosci*. 2006;26:8360–7.
62. Middleton FA, Strick PL. Anatomical evidence for cerebellar and basal ganglia involvement in higher cognitive function. *Science*. 1994;266:458–61.
63. Gunaydin LA, Kreitzer AC. Cortico-Basal Ganglia Circuit Function in Psychiatric Disease. *Annu Rev Physiol*. 2016;78:327–50.
64. White NM. Mnemonic functions of the basal ganglia. *Curr Opin Neurobiol*. 1997;7:164–9.
65. Packard MG, Knowlton BJ. Learning and memory functions of the Basal Ganglia. *Annu Rev Neurosci*. 2002;25:563–93.
66. Rektor I, Bares M, Brázdil M, Kanovský P, Rektorová I, Sochorová D, et al. Cognitive- and movement-related potentials recorded in the human basal ganglia. *Mov Disord*. 2005;20:562–8.
67. Mai J, Lu M, Gao Q, Zeng J, Xiao J. Transcriptome-wide association studies: recent advances in methods, applications and available databases. *Commun Biol*. 2023;6:899.
68. Gamazon ER, Wheeler HE, Shah KP, Mozaffari SV, Aquino-Michaels K, Carroll RJ, et al. A gene-based association method for mapping traits using reference transcriptome data. *Nat Genet*. 2015;47:1091–8.
69. Yuan Z, Zhu H, Zeng P, Yang S, Sun S, Yang C, et al. Testing and controlling for horizontal pleiotropy with probabilistic Mendelian randomization in transcriptome-wide association studies. *Nat Commun*. 2020;11:3861.
70. Liu L, Zeng P, Xue F, Yuan Z, Zhou X. Multi-trait transcriptome-wide association studies with probabilistic Mendelian randomization. *Am J Hum Genet*. 2021;108:240–56.
71. Bush G, Luu P, Posner MI. Cognitive and emotional influences in anterior cingulate cortex. *Trends Cogn Sci*. 2000;4:215–22.
72. Fellows LK, Farah MJ. Is anterior cingulate cortex necessary for cognitive control? *Brain*. 2005;128:788–96.
73. Kenigsbuch M, Bost P, Halevi S, Chang Y, Chen S, Ma Q, et al. A shared disease-associated oligodendrocyte signature among multiple CNS pathologies. *Nat Neurosci*. 2022;25:876–86.
74. Verbanck M, Chen CY, Neale B, Do R. Detection of widespread horizontal pleiotropy in causal relationships inferred from Mendelian randomization between complex traits and diseases. *Nat Genet*. 2018;50:693–8.
75. Koletzko B, Reischl E, Tanjung C, Gonzalez-Casanova I, Ramakrishnan U, Meldrum S, et al. FADS1 and FADS2 Polymorphisms Modulate Fatty Acid Metabolism and Dietary Impact on Health. *Annu Rev Nutr*. 2019;39:21–44.

ACKNOWLEDGEMENTS

This study was supported by the National Natural Science Foundation of China (82370810, 82372347, 82088102 and 82170819), National Key Research and Development Program of China (2022YFC2505202, 2022YFC2505201, 2022YFC2505203 and 2021YFA1301103), Science and Technology Commission of Shanghai Municipality (23JS1400900, 23Y11908400 and 23XD1422400), Innovative research team of high-level local universities in Shanghai. JZ was supported by the Academy of Medical Sciences (AMS) Springboard Award, the Wellcome Trust, the Government Department of Business, Energy and Industrial Strategy (BEIS), the British Heart Foundation and Diabetes UK (SBF006\1117).

AUTHOR CONTRIBUTIONS

Conceptualization, XW, JZ and JL; Data curation, XW, HQ, CH, XJ and LL; Formal analysis, XW, HQ and CH; Funding acquisition, JZ, YB and JL; Methodology, JZ; Project administration, JZ, YB and JL; Resources, YB and JL; Supervision, JL; Validation, HL and SW; Writing - original draft, XW, HQ and CH; Writing - review & editing, LJ, YZ, RZ, ML, TW, ZZ, MX, YX and YC. All authors have read and agreed to the final version of the manuscript.

COMPETING INTERESTS

The authors declare no competing interests. Some icons in Graphical Abstract were created with BioRender.com.

ADDITIONAL INFORMATION

Supplementary information The online version contains supplementary material available at <https://doi.org/10.1038/s41398-024-02784-4>.

Correspondence and requests for materials should be addressed to Jie Zheng, Yufang Bi or Jieli Lu.

Reprints and permission information is available at <http://www.nature.com/reprints>

Publisher's note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons license, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons license and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this license, visit <http://creativecommons.org/licenses/by/4.0/>.

© The Author(s) 2024