

# Persistence of SARS-CoV-2 antibodies over 18 months following infection: UK Biobank COVID-19 Serology Study

JelenaBešević  $\bullet$  ,<sup>1</sup> Ben Lacey  $\bullet$  ,<sup>1</sup> Howard Callen,<sup>1</sup> Wemimo Omiyale,<sup>1</sup> MeganConroy  $\bullet$ ,<sup>1</sup> Qi Feng,<sup>1</sup> Derrick W Crook,<sup>2</sup> Nicola Doherty,<sup>3</sup> Daniel Ebner,<sup>2</sup> David W Eyre,<sup>4</sup> Dan Fry,<sup>3</sup> Edward Horn,<sup>1</sup> E Yvonne Jones,<sup>2</sup> Brian D Marsden,<sup>2</sup> Tim E A Peto,<sup>2</sup> Fenella Starkey,<sup>1</sup> David Stuart,<sup>2</sup> Samantha Welsh,<sup>3</sup> Natasha Wood,<sup>1</sup> Alan Young,<sup>1,3</sup> Allen Young,<sup>1</sup> Mark Effingham,<sup>3</sup> Rory Collins,<sup>1,3</sup> Jo Holliday,<sup>1</sup> Naomi Allen<sup>1,3</sup>

# **ABSTRACT**

**Background** Little is known about the persistence of antibodies after the first year following SARS-CoV-2 infection. We aimed to determine the proportion of individuals that maintain detectable levels of SARS-CoV-2 antibodies over an 18-month period following infection.

**Methods** Population-based prospective study of 20 000 UK Biobank participants and their adult relatives recruited in May 2020. The proportion of SARS-CoV-2 cases testing positive for immunoglobulin G (IgG) antibodies against the spike protein (IgG-S), and the nucleocapsid protein (IgG-N), was calculated at varying intervals following infection.

**Results** Overall, 20 195 participants were recruited. Their median age was 56 years (IQR 39–68), 56% were female and 88% were of white ethnicity. The proportion of SARS-CoV-2 cases with IgG-S antibodies following infection remained high (92%, 95% CI 90%–93%) at 6months after infection. Levels of IgG-N antibodies following infection gradually decreased from 92% (95% CI 88%–95%) at 3months to 72% (95% CI 70%–75%) at 18 months. There was no strong evidence of heterogeneity in antibody persistence by age, sex, ethnicity or socioeconomic deprivation.

**Conclusion** This study adds to the limited evidence on the long-term persistence of antibodies following SARS-CoV-2 infection, with likely implications for waning immunity following infection and the use of IgG-N in population surveys.

# **INTRODUCTION**

Since the emergence of SARS-CoV-2 in December 2019, there have been over 650 million confirmed cases of COVID-19 worldwide.<sup>1</sup> Although vaccination is currently effective at reducing the risk of severe COVID-19, there is ongoing transmission of SARS-CoV-2, driven in part by waning immunity after infection or vaccination.

Following infection, most individuals will develop circulating antibodies to SARS-CoV-[2](#page-3-1).<sup>2</sup> However, the long-term persistence of these antibodies has not been well described. Understanding the persistence of SARS-CoV-2 antibodies would improve our understanding of the protection afforded following infection and inform estimates

# **WHAT IS ALREADY KNOWN ON THIS TOPIC**

⇒ The dynamics of SARS-CoV-2 antibodies following infection has been extensively assessed over the course of a year following infection.

# **WHAT THIS STUDY ADDS**

 $\Rightarrow$  To the best of our knowledge, this study is the first to report the persistence of antibodies over an 18-month period following SARS-CoV-2 infection.

# **HOW THIS STUDY MIGHT AFFECT RESEARCH, PRACTICE OR POLICY**

 $\Rightarrow$  Our findings indicate that over a quarter of individuals are unlikely to have detectable IgG-N antibodies at 18 months after infection. This has implications for immunity following infection, and the use of IgG-N antibodies in population surveys to avoid underestimating prevalence of past infection.

of the likely trajectory of SARS-CoV-2 transmission, especially in populations with limited availability or uptake of vaccines.

We aimed to determine the duration of antibody response following SARS-CoV-2 infection over an 18-month period, and to assess whether this is affected by demographic factors such as age, sex and ethnicity.

# **METHODS**

# **Design and participants**

UK Biobank participants, resident in the mainland UK in May 2020, were invited by email to join the UK Biobank COVID-19 Serology Study.<sup>[3](#page-3-2)</sup> Participants were asked whether they would be willing to take serial blood samples in their own home and answer brief questionnaires about potential COVID-19 symptoms. In order to increase the age range of the study, UK Biobank participants (who are now typically 50–80 years) were asked to forward an email invitation from UK Biobank to their adult (ie,  $\geq 18$  years old) children and/or grandchildren inviting them to join the study. UK Biobank participants and their adult relatives were

► Additional supplemental material is published online only. To view, please visit the journal online [\(http://dx.](http://dx.doi.org/10.1136/jech-2023-220569) [doi.org/10.1136/jech-2023-](http://dx.doi.org/10.1136/jech-2023-220569) [220569](http://dx.doi.org/10.1136/jech-2023-220569)).

1 Nuffield Department of Population Health (NDPH), University of Oxford, Oxford, UK <sup>2</sup>Nuffield Department of Medicine (NDM), University of Oxford, Oxford, UK <sup>3</sup>UK Biobank, Stockport, UK 4 University of Oxford Big Data Institute, Oxford, UK

## **Correspondence to**

Dr Jelena Bešević, Nuffield Department of Population Health (NDPH), University of Oxford, Oxford OX1 2JD, UK; jelena.besevic@ndph.ox.ac.uk

JB and BL contributed equally. JH and NA contributed equally.

Received 17 March 2023 Accepted 8 October 2023 Published Online First 3 November 2023

# Check for updates

© Author(s) (or their employer(s)) 2024. Re-use permitted under CC BY. Published by BMJ.



# **Short report**

then selected by random sampling, stratified by demographic characteristics, with some subsequent oversampling from ethnic minority groups and from urban settings. Efforts were made to select only one participant from a given address.

# **SARS-CoV-2 antibody assessment**

In the first phase of the study, participants were posted a selfsampling blood kit and symptom questionnaire every month for 6months between the end of May and beginning of December 2020. Instructions on how to collect and return the sample (together with a link to an online information video) were included in all sampling kits. Participants were asked to return the sample (containing  $\sim 0.5$  mL capillary blood) and questionnaire by post to the UK Biobank laboratory on the day of sample collection. Samples were tested for immunoglobulin G (IgG) antibodies against the spike protein (IgG-S) of the SARS-CoV-2 virus, determined using an ELISA platform at the Target Discovery Institute laboratory, University of Oxford.<sup>4</sup> In the second phase of the study, participants were sent a final blood sample kit between November 2021 and February 2022. While the first phase of the study measured IgG-S antibodies, the second phase of the study involved testing blood samples

for antibodies against the nucleocapsid protein (IgG-N) of the virus. The change in antibody test used to determine past infection was necessary as the UK vaccination programme began in December 2020 and after this date IgG-S antibodies, which can be present as a result of either infection or vaccination, could not be used to determine infection, whereas IgG-N antibodies, which are generated as a result of infection only, could be used to determine infection. Thriva test kits were used to collect the blood samples, which were subsequently analysed at a nominated Thriva laboratory.

## **Statistical analysis**

Participants with previous SARS-CoV-2 infection were identified by a positive antibody test or positive PCR test (available for existing UK Biobank participants via linkage with the national testing data). The date of infection was estimated as date of PCR or 21 days prior to first positive antibody test (accepting that in some participants the date of infection was likely earlier, but accounting for the lag in time between infection and the generation of detectable levels of antibodies). Individuals with evidence of reinfection, defined as a positive PCR test >90days after the date of first SARS-CoV-2 infection,<sup>5</sup> were excluded.

<span id="page-1-0"></span>

IgG-S antibody analyses restricted to SARS-CoV-2 cases with one or more IgG-S tests following diagnosis. IgG-N antibody analyses restricted to SARS-CoV-2 cases with IgG-N test following diagnosis.

\*Excludes participants of unknown ethnicity.

†Area-level derived measure of socioeconomic deprivation (categories are defined as: <−2 (less deprived), −2 to <2 (average), 2+ (more deprived)).



<span id="page-2-0"></span>**Figure 1** Proportion of SARS-CoV-2 cases seropositive for IgG-S and IgG-N antibodies over an 18-month period following infection. (A) Analyses among 1514 SARS-CoV-2 cases who had one or more subsequent IgG-S antibody tests; 74 individuals are omitted from the plot as they have a duration of follow-up less than 30 days and a further 27 individuals are omitted from the plot as they exceed 180 days duration. Proportions are % (95% CI) of IgG-S seropositive cases among participants that returned a valid test during each period; participants may contribute to more than period. (B) Analyses among 1749 SARS-CoV-2 cases with IgG-N test following diagnosis; 78 individuals are omitted from the plot as they have a duration of follow-up less than 30 days and a further 16 individuals are omitted from the plot as they exceed 630 days duration. Proportions are % (95% CI) of IgG-N seropositive cases among participants that returned a valid test during each period; participants provided one IgG-N test only.

To determine the seroprevalence of IgG-S antibodies following infection in the first phase of the study, the proportion of SARS-CoV-2 cases testing positive for IgG-S antibodies was calculated along with 95% CIs at the following time intervals: 30–60, 60–90, 90–120, 120–150 and 150–180days following the date of infection. The denominators were restricted to cases that returned an analysable sample during the given time interval. The seroprevalence of IgG-N antibodies following infection was similarly calculated for the following time intervals: 30–270, 270–450 and 450–630days. Heterogeneity was assessed using a  $\chi^2$  test. Statistical analysis was performed in R V.4.1.1.

#### **RESULTS**

Overall, 20195 individuals were recruited into the UK Biobank COVID-19 Serology Study. The median age of participants was 56 (IQR 39–68) years, 56% were female and 88% were of white ethnicity; 94% returned one or more blood samples [\(table](#page-1-0) 1; [online supplemental table S1\)](https://dx.doi.org/10.1136/jech-2023-220569).

In the first phase of the study, there were 1514 cases of SARS-CoV-2 infection with at least one subsequent IgG-S antibody test. The proportion of SARS-CoV-2 cases with IgG-S antibodies following infection remained high over the first 6months, with  $\sim$ 92% (95% CI 90%–93%) of those tested between 150 and 180 days after infection having detectable antibodies [\(figure](#page-2-0) 1A). There was no strong evidence of heterogeneity in this proportion by age, sex, ethnicity or socioeconomic deprivation [\(online](https://dx.doi.org/10.1136/jech-2023-220569)  [supplemental table S2](https://dx.doi.org/10.1136/jech-2023-220569)).

There were 1749 cases of SARS-CoV-2 infection with a subsequent IgG-N antibody test in the second phase of the study. There was a progressive reduction in the proportion of individuals with detectable levels of IgG-N antibodies over 18 months following infection [\(figure](#page-2-0) 1B): 92% (95% CI 88%–95%) had IgG-N antibodies at about 3months, 79% (95% CI 75%–83%) at 12 months and 72% (95% CI 70%–75%) at 18 months.

There was no strong evidence of heterogeneity in the persistence of Ig-N antibodies by age, sex, ethnicity or socioeconomic deprivation index; neither was there evidence that seroprevalence varied by method of case ascertainment, as similar estimates were observed at 18 months when limiting the analysis to cases identified via PCR test only [\(online supplemental table](https://dx.doi.org/10.1136/jech-2023-220569) S<sub>3</sub> and figure S<sub>1</sub>).

#### **DISCUSSION**

In this population-based cohort study, the proportion of SARS-CoV-2 cases with IgG-S antibodies following infection remained high at 92% at 6 months. Longer follow-up for IgG-N antibodies found a progressive reduction in the proportion of cases with detectable levels of antibodies over 18 months following infection, falling from 90% at 3months to 79% at 12 months, and 72% at 18 months. There was no evidence of variation in findings for IgG-S or IgG-N by age, sex, ethnicity or socioeconomic deprivation.

Several previous studies on antibody dynamics following SARS-CoV-2 infection report broadly consistent findings to the

# **Short report**

present study at 1year or less of follow-up, but there were no identified studies with 18 months of follow-up.<sup>6-14</sup> One of the largest studies, of nearly 40000 individuals from the USA, found that IgG-S and IgG-N antibodies were detected in  $\sim$ 90% and  $\sim$ 70% of individuals, respectively, at 10 months after infection; however, there was substantial heterogeneity in the IgG-N find-ings by assay platforms.<sup>[11](#page-3-6)</sup>

The long duration of follow-up is a particular strength of this study. The study also used objective case ascertainment and validated antibody tests, and was sufficiently large to enable exploration of study outcomes in important population subgroups. It is a limitation of the study, however, that most cases were ascertained during the first phase of the study, and as such those with the longest follow-up in the second phase were identified mostly by antibody tests (PCR testing was not routinely performed during the early stages of the pandemic). More widespread PCR testing would have identified some cases who did not seroconvert, which may have reduced further the proportion with undetectable antibody levels during follow-up.[15](#page-3-7) Another limitation of the study was that severity of infection, which may affect the persistence of antibodies, was not assessed.<sup>16</sup> Further, individuals with severe disease may have been less likely to return a sample, and as such the findings should be extrapolated with caution to those with severe infections, although this will have accounted for only a small proportion of cases.[17](#page-3-9) This study excluded individuals with evidence of reinfection; however, as the pandemic continues and reinfections increase, the effect on antibody levels in this context (including any difference in effect by major variants, such as Delta and Omicron) needs further study.<sup>18</sup> It would also be valuable to understand further the effect of vaccination on immune response among those with and without natural infection, which was beyond the scope of the study.<sup>[19](#page-3-11)</sup>

In conclusion, although antibody levels are detectable in the vast majority of individuals following infection over a 6-month period, there was an appreciable reduction over 18 months, with over a quarter of individuals having undetectable levels of IgG-N antibodies at this time. This waning of antibody levels over time supports the growing evidence on the limited long-term protection following natural infection, and the importance of vaccine schedules to increase antibody levels over the longer term. It also indicates that the use of IgG-N in population surveys is likely to substantially underestimate past infection at longer durations of follow-up.

**Acknowledgements** We thank all individuals who participated in the UK Biobank Serology Study.

**Contributors** NA and RC initiated the study. JB and BL had full access to all of the data in the study and take responsibility for the accuracy of the data analysis. JB, BL, JH and NA drafted the manuscript and did the statistical analysis. All authors contributed to the design of the study; the acquisition, analysis and interpretation of data; or made critical revision of the manuscript for important intellectual content. JB, BL, JH and NA are the guarantors.

**Funding** The funding for this substudy of UK Biobank was provided by the UK Department of Health and Social Care. The core funding for UK Biobank is provided by the UK Medical Research Council, Wellcome, British Heart Foundation, Cancer Research UK and National Institute for Health Research (grant ref: 223600/Z/21/Z).

**Disclaimer** The funders had no role in the design; in the collection, analysis or interpretation of data; in the writing of the report; or in the decision to submit the paper for publication.

**Competing interests** None declared.

## **Patient consent for publication** Not applicable.

**Ethics approval** This study involves human participants and UK Biobank received approval from the National Information Governance Board for Health and Social

Care and the National Health Service North West Centre for Research Ethics Committee (Ref: 21/NW/0157). Participants gave informed consent to participate in the study before taking part.

**Provenance and peer review** Not commissioned; externally peer reviewed.

**Supplemental material** This content has been supplied by the author(s). It has not been vetted by BMJ Publishing Group Limited (BMJ) and may not have been peer-reviewed. Any opinions or recommendations discussed are solely those of the author(s) and are not endorsed by BMJ. BMJ disclaims all liability and responsibility arising from any reliance placed on the content. Where the content includes any translated material, BMJ does not warrant the accuracy and reliability of the translations (including but not limited to local regulations, clinical guidelines, terminology, drug names and drug dosages), and is not responsible for any error and/or omissions arising from translation and adaptation or otherwise.

**Open access** This is an open access article distributed in accordance with the Creative Commons Attribution 4.0 Unported (CC BY 4.0) license, which permits others to copy, redistribute, remix, transform and build upon this work for any purpose, provided the original work is properly cited, a link to the licence is given, and indication of whether changes were made. See: [https://creativecommons.org/](https://creativecommons.org/licenses/by/4.0/) [licenses/by/4.0/.](https://creativecommons.org/licenses/by/4.0/)

## **ORCID iDs**

Jelena Bešević <http://orcid.org/0000-0003-4927-1840> Ben Lacey <http://orcid.org/0000-0003-0139-2934> Megan Conroy<http://orcid.org/0000-0002-3847-6202>

#### **REFERENCES**

- <span id="page-3-0"></span>1 World Health Organisation. WHO COVID-19 dashboard. 2020. Available: [https://](https://covid19.who.int/) [covid19.who.int/](https://covid19.who.int/) [Accessed 22 Nov 2022].
- <span id="page-3-1"></span>2 Lou B, Li T-D, Zheng S-F, et al. Serology characteristics of SARS-Cov-2 infection after exposure and post-symptom onset. [Eur Respir J](http://dx.doi.org/10.1183/13993003.00763-2020) 2020;56:2000763.
- <span id="page-3-2"></span>3 Biobank UK. COVID-19 serology study waves 1-6 overview document; 2022.
- <span id="page-3-3"></span>4 National SARS-CoV-2 Serology Assay Evaluation Group. Performance characteristics of five immunoassays for SARS-Cov-2: a head-to-head benchmark comparison. [Lancet](http://dx.doi.org/10.1016/S1473-3099(20)30634-4)  [Infect Dis](http://dx.doi.org/10.1016/S1473-3099(20)30634-4) 2020;20:1390–400.
- <span id="page-3-4"></span>5 Centers for Disease Control and Prevention. Coronavirus disease 2019 (COVID-19) 2021 case definition. 2023. Available: [https://ndc.services.cdc.gov/case-definitions/](https://ndc.services.cdc.gov/case-definitions/coronavirus-disease-2019-2021/) [coronavirus-disease-2019-2021/](https://ndc.services.cdc.gov/case-definitions/coronavirus-disease-2019-2021/) [Accessed 21 Feb 2023].
- <span id="page-3-5"></span>6 Ortega N, Ribes M, Vidal M, et al. Seven-month kinetics of SARS-Cov-2 antibodies and role of pre-existing antibodies to human Coronaviruses. [Nat Commun](http://dx.doi.org/10.1038/s41467-021-24979-9) 2021;12:4740.
- Wang Y, Zhang L, Sang L, et al. Kinetics of viral load and antibody response in relation to COVID-19 severity. [J Clin Invest](http://dx.doi.org/10.1172/JCI138759) 2020;130:5235-44.
- 8 Gudbjartsson DF, Norddahl GL, Melsted P, et al. Humoral immune response to SARS-Cov-2 in Iceland. [N Engl J Med](http://dx.doi.org/10.1056/NEJMoa2026116) 2020;383:1724–34.
- 9 Wajnberg A, Amanat F, Firpo A, et al. Robust neutralizing antibodies to SARS-Cov-2 infection persist for months. [Science](http://dx.doi.org/10.1126/science.abd7728) 2020;370:1227-30.
- 10 Isho B, Abe KT, Zuo M, et al. Persistence of serum and saliva antibody responses to SARS-Cov-2 spike antigens in COVID-19 patients. [Sci Immunol](http://dx.doi.org/10.1126/sciimmunol.abe5511) 2020;5:eabe5511.
- <span id="page-3-6"></span>11 Alfego D, Sullivan A, Poirier B, et al. A population-based analysis of the longevity of SARS-Cov-2 antibody seropositivity in the United States. [EClinicalMedicine](http://dx.doi.org/10.1016/j.eclinm.2021.100902) 2021;36:100902.
- 12 Van Elslande J, Oyaert M, Ailliet S, et al. Longitudinal follow-up of IgG antinucleocapsid antibodies in SARS-Cov-2 infected patients up to eight months after infection. [J Clin Virol](http://dx.doi.org/10.1016/j.jcv.2021.104765) 2021;136:104765.
- 13 Krutikov M, Palmer T, Tut G, et al. Prevalence and duration of detectable SARS-Cov-2 nucleocapsid antibodies in staff and residents of long-term care facilities over the first year of the pandemic (VIVALDI study): prospective cohort study in England. Lancet [Healthy Longev](http://dx.doi.org/10.1016/S2666-7568(21)00282-8) 2022;3:e13–21.
- 14 Lumley SF, Wei J, O'Donnell D, et al. The duration, dynamics, and determinants of severe acute respiratory syndrome Coronavirus 2 (SARS-Cov-2) antibody responses in individual healthcare workers. [Clin Infect Dis](http://dx.doi.org/10.1093/cid/ciab004) 2021;73:e699-709.
- <span id="page-3-7"></span>15 Liu W, Russell RM, Bibollet-Ruche F, et al. Predictors of nonseroconversion after SARS-Cov-2 infection. [Emerg Infect Dis](http://dx.doi.org/10.3201/eid2709.211042) 2021;27:2454–8.
- <span id="page-3-8"></span>16 Wang Y, Li J, Li H, et al. Persistence of SARS-Cov-2-specific antibodies in COVID-19 patients. [Int Immunopharmacol](http://dx.doi.org/10.1016/j.intimp.2020.107271) 2021;90:107271.
- <span id="page-3-9"></span>17 Beaney T, Neves AL, Alboksmaty A, et al. Trends and associated factors for COVID-19 hospitalisation and fatality risk in 2.3 million adults in England. [Nat Commun](http://dx.doi.org/10.1038/s41467-022-29880-7) 2022;13:2356.
- <span id="page-3-10"></span>18 Reynolds CJ, Pade C, Gibbons JM, et al. Immune boosting by B.1.1.529 (Omicron) depends on previous SARS-Cov-2 exposure. [Science](http://dx.doi.org/10.1126/science.abq1841) 2022;377:eabq1841.
- <span id="page-3-11"></span>Lin D-Y, Gu Y, Xu Y, et al. Association of primary and booster vaccination and prior infection with SARS-Cov-2 infection and severe COVID-19 outcomes. [JAMA](http://dx.doi.org/10.1001/jama.2022.17876) 2022;328:1415–26.