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DOI <https://doi.org/10.1200/JCO.23.00570>

## ABSTRACT

**PURPOSE** Diffuse large B-cell lymphoma (DLBCL) is cured in more than 60% of patients, but outcomes remain poor for patients experiencing disease progression or relapse (refractory or relapsed DLBCL [rrDLBCL]), particularly if these events occur early. Although previous studies examining cohorts of rrDLBCL have identified features that are enriched at relapse, few have directly compared serial biopsies to uncover biological and evolutionary dynamics driving rrDLBCL. Here, we sought to confirm the relationship between relapse timing and outcomes after second-line (immuno)chemotherapy and determine the evolutionary dynamics that underpin that relationship.

**PATIENTS AND METHODS** Outcomes were examined in a population-based cohort of 221 patients with DLBCL who experienced progression/relapse after frontline treatment and were treated with second-line (immuno)chemotherapy with an intention-to-treat with autologous stem-cell transplantation (ASCT). Serial DLBCL biopsies from a partially overlapping cohort of 129 patients underwent molecular characterization, including whole-genome or whole-exome sequencing in 73 patients.

**RESULTS** Outcomes to second-line therapy and ASCT are superior for late relapse (>2 years postdiagnosis) versus primary refractory (<9 months) or early relapse (9–24 months). Diagnostic and relapse biopsies were mostly concordant for cell-of-origin classification and genetics-based subgroup. Despite this concordance, the number of mutations exclusive to each biopsy increased with time since diagnosis, and late relapses shared few mutations with their diagnostic counterpart, demonstrating a branching evolution pattern. In patients with highly divergent tumors, many of the same genes acquired new mutations independently in each tumor, suggesting that the earliest mutations in a shared precursor cell constrain tumor evolution toward the same genetics-based subgroups at both diagnosis and relapse.

**CONCLUSION** These results suggest that late relapses commonly represent genetically distinct and chemotherapy-naïve disease and have implications for optimal patient management.

## ACCOMPANYING CONTENT

 [Data Supplement](#)

Accepted May 8, 2023

Published June 15, 2023

J Clin Oncol 41:4164-4177

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## INTRODUCTION

Diffuse large B-cell lymphoma (DLBCL) is an aggressive and heterogeneous lymphoma for which standard-of-care rituximab with cyclophosphamide, vincristine, doxorubicin, and prednisone (R-CHOP) immunochemotherapy results in long-term remission in 60%–70% of patients.<sup>1</sup> However, outcomes are poor for the 30%–40% of patients with

primary refractory or relapsed disease (refractory or relapsed DLBCL [rrDLBCL]) even after second-line therapy and autologous stem-cell transplantation (ASCT).<sup>2,3</sup> The landscape of coding and noncoding somatic variants in DLBCL at diagnosis is well established,<sup>4–8</sup> and several studies have examined the mutational landscape of cohorts of rrDLBCL, identifying somatic variants that occur more frequently in rrDLBCL.<sup>9–12</sup> Although several of these mutations are

## CONTEXT

### Key Objective

What are the patterns of tumor evolution that underpin the relationship between relapse timing and outcomes to second-line (immuno)chemotherapy and autologous stem-cell transplantation in patients with diffuse large B-cell lymphoma (DLBCL)?

### Knowledge Generated

Although broad molecular categories (cell-of-origin and genetic subgroups) were consistent between diagnosis and relapse, late relapses arose from serial (and potentially treatment naïve) transformations from persistent common precursor cell populations, whereas earlier relapses were more closely genetically related, implying innate (immuno)chemotherapy resistance. These observations have implications for clinical trial design and patient management, where early and late relapse patient populations should be considered separately and support additional molecular characterization of tumors at relapse to guide precision medicine approaches.

### Relevance (J.W. Friedberg)

These results emphasize the importance of repeat biopsy and molecular analysis of relapsed DLBCL, and further our understanding of malignant B-cell evolutionary dynamics.\*

\*Relevance section written by JCO Editor-in-Chief Jonathan W. Friedberg, MD.

prognostic at diagnosis for the likelihood of relapse, they are insufficient to explain the poor outcomes experienced by patients with rrDLBCL.

Tumor evolution is usually considered to follow one of two models: linear or branching evolution. Linear evolution is defined when the relapse tumor harbors all the variants found at diagnosis along with a set of exclusive variants, implying direct descent of the relapse from the diagnostic tumor. Branching evolution is characterized by exclusive variants in both diagnostic and relapse tumors with a variable number of shared variants (the trunk). In the transformation of follicular lymphoma (FL) to aggressive DLBCL (tFL), this branching evolution is considered evidence of a persistent common precursor cell (CPC) ancestral to both lymphomas.<sup>13-15</sup> Previous studies of DLBCL tumor evolution have leveraged circulating tumor DNA and/or limited targeted capture space to examine the evolutionary dynamics of relapse in small cohorts, providing some evidence that branching evolution predominates.<sup>9,16-18</sup> However, the degree to which persistent CPC populations might contribute to DLBCL relapse is not yet known.

Critically, no studies have yet, to our knowledge, examined the evolution of the mutation landscape together with gene expression profiling (GEP)-based cell-of-origin (COO)<sup>19,20</sup> and dark-zone signature (DZsig)<sup>21,22</sup> classification. More recently, genetics-based classifiers have been developed that leverage co-occurrence of somatic variants to identify shared biology within DLBCL. Intriguingly, the three studies that described genetics-based groups converged on 5-7 overlapping subgroups.<sup>7,8,23-25</sup> The LymphGen

algorithm allows genetics-based subgroup assignment for individual biopsies.<sup>23</sup> These classification systems are becoming the foundation for precision medicine in DLBCL, and while the current assumption is that molecular features that underlie the classification of each tumor would be fixed over time, this requires formal testing.

Here, we examined a large population-based cohort of rrDLBCL showing that response rate and outcomes to second-line (immuno)chemotherapy and ASCT are superior for patients with late relapses relative to primary refractory or early relapse. To examine the genetic and evolutionary relationships between diagnostic and rrDLBCL underlying these clinical differences, we assembled a cohort of 129 patients with multiple DLBCL biopsies and interrogated them with a combination of fluorescence in situ hybridization (FISH) for recurrent rearrangements, GEP for COO and DZsig, and/or whole-genome or whole-exome sequencing of two or more DLBCL tumors per patient.

## PATIENTS AND METHODS

Outcome analyses were performed in the outcomes cohort of 221 patients with de novo DLBCL treated with front-line R-CHOP(-like) therapy who experienced DLBCL progression/relapse (Data Supplement [Tables S1 and S2, online only]). A major inclusion criterion was treatment with second-line (immuno)chemotherapy (89% received GDP [gemcitabine, dexamethasone, and cisplatin] ± rituximab)<sup>26</sup> with intention to treat with consolidative ASCT in patients with (immuno)chemotherapy-responsive disease (Data Supplement [Tables S1 and S2]).

The partially overlapping molecular characterization cohort comprised patients where there was sufficient material for molecular assays from multiple tumors of DLBCL morphology. Additionally, constitutional DNA was required for all patients whose tumors underwent DNA sequencing. The uniform second-line treatment approach required for the outcomes cohort was not an inclusion criterion for this cohort. Thus, patients who were not treated with intention to consolidate with ASCT were included along with patients with indolent lymphoma evident at any time in their disease course, as long as multiple DLBCL biopsies were available. This cohort is enriched for patients who experienced late relapse, reflecting historic patterns of obtaining a biopsy to confirm relapse less frequently in primary refractory disease. In total, 129 patients were identified, of whom 32 had prior indolent lymphoid neoplasms. Among the 97 patients with apparently *de novo* DLBCL at diagnosis, 11 had subsequent indolent lymphoma diagnoses. Pairs of biopsies were interrogated with a combination of break-apart FISH for *MYC*, *BCL2*, and/or *BCL6* rearrangements, digital GEP (NanoString DLBCL90) for COO and DZsig classification,<sup>20,21</sup> and/or whole-genome (WGS) or whole-exome sequencing (WES) (Data Supplement [Supplemental Fig 1 and Tables S3 and S4]). Further details are provided in the Data Supplement. This study was reviewed and approved by the University of British Columbia-BC Cancer Research Ethics Board in accordance with the Declaration of Helsinki. All patients provided written informed consent with the exception of patients where waiver of consent was granted by the Research Ethics Board.

## RESULTS

### Patients With Late Relapse Have Superior Outcomes

Considering previous observations that outcomes to second-line therapies are related to progression/relapse timing,<sup>27,28</sup> we first sought to confirm this observation in a large population-based outcomes cohort. The 221 patients were categorized into three relapse timing categories: Primary refractory disease was defined as progression or relapse within 9 months of diagnosis, approximating 3 months post-end of treatment.<sup>29</sup> Late relapses were defined as more than 24 months after diagnosis, reflecting the definition of EFS24—a validated end point in which patients event free 24 months after immunochemotherapy collectively have superior disease-related outcomes.<sup>30</sup> Early relapses were defined as relapse 9–24 months from diagnosis. We found significant differences in both response rates to second-line (immuno)chemotherapy (Fig 1A) and the proportion of patients who ultimately received consolidative ASCT (Fig 1B), demonstrating superior (immuno)chemosensitivity of tumors of patients experiencing late relapses. Patients experiencing late relapse had significantly superior progression-free survival and overall survival relative to patients with primary refractory or early relapse when considering either time from first progression/relapse (Figs 1C and 1D) or from ASCT (Figs 1E and 1F). Outcome differences persisted after

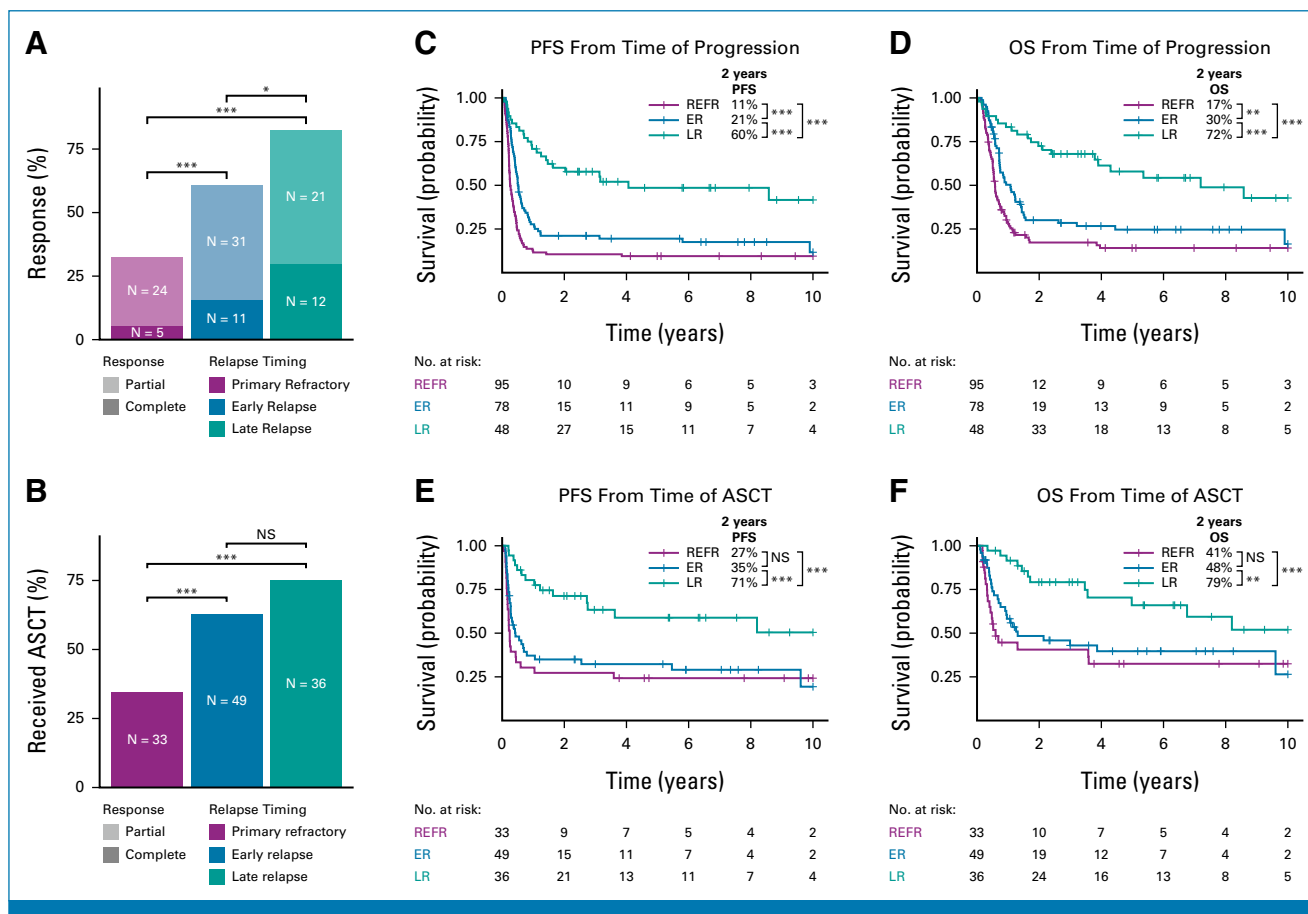
adjusting for age at diagnosis and International Prognostic Index at relapse (Data Supplement [Supplemental Fig 2]).

### Late Relapses Are Highly Genetically Divergent

To examine the underlying tumor biology and patterns of evolution driving the superior outcomes observed in late relapses, we used a molecular characterization cohort of 129 patients that experienced progression/relapse with available serial DLBCL biopsies. DNA sequencing (WGS [n = 68 patients] and WES [n = 5]) was performed on multiple serial DLBCL biopsies along with constitutional DNA (Fig 2), with 21 also included in the outcomes cohort (four primary refractory, seven early relapses, and 10 late relapses).

The use of formalin-fixed paraffin-embedded tissues for most samples resulted in variable sequencing depth (mean, 48.6X across WGS samples and 97X in exomes; Data Supplement [Supplemental Fig 3 and Table S5]). We also performed deep targeted DNA sequencing of genes relevant for LymphGen classification (LySeqST, Data Supplement [Table S6]) on multiple biopsies subjected to WGS from 47 patients and on a single biopsy from another 15 patients. The LySeqST assay achieved a mean depth of 812X across its capture space (Data Supplement [Supplemental Fig 4A and Table S7]). The lower variant allele frequencies of variants detected by LySeqST alone versus genomes demonstrates that it enhanced detection of subclonal variants that fall below the WGS limit of detection (Data Supplement [Supplemental Fig 4B]).

Next, we explored the overall divergence of mutations by comparing the number of shared (common between both biopsies) and exclusive (present in only one biopsy) mutations between the first two DLBCL biopsies in each patient. For this and all subsequent analyses, we pooled the LySeqST and WGS variant calls and only retained variants at positions with a sequencing depth of at least 10 unique molecules in all tumors from the same patient. While primary refractory and early relapse tumors have a rich landscape of variants shared between tumors, many late relapses have few, with most mutations exclusive to either the diagnostic or relapse biopsy (Fig 3A). In both primary refractory and early relapse disease, the number of mutations shared between tumors is strongly correlated with the total number of variants identified at either diagnosis or relapse with slopes nearing unity, demonstrating that most variants are shared between tumors (Fig 3B). In contrast, this correlation was weak in late relapses (Fig 3B). Comparing the percentage of exclusive variants in each tumor to the time between biopsies revealed a clear linear trend, where tumor pairs separated by many years have very few shared variants (Fig 3C and Data Supplement [Table S8]). This trend was consistent when considering time to relapse as a categorical variable (Data Supplement [Supplemental Fig 5]), when the absolute number of exclusive mutations was considered (Data Supplement [Supplemental Fig 6]) and is



**FIG 1.** Relationship between relapse timing and outcomes to second-line therapy. (A and B) The percent of patients in each relapse timing category (A) whose relapse responded to second-line therapy and (B) who received ASCT. Groups were compared with pairwise Fisher's exact tests. (C-F) Kaplan-Meier curves showing PFS and OS from the time of progression or ASCT. *P* values were determined with a log-rank test. \**P* < .05; \*\**P* < .01; \*\*\**P* < .001. ASCT, autologous stem-cell transplantation; ER, early relapse; LR, late relapse; NS, not significant; OS, overall survival; PFS, progression-free survival; REFR, primary refractory.

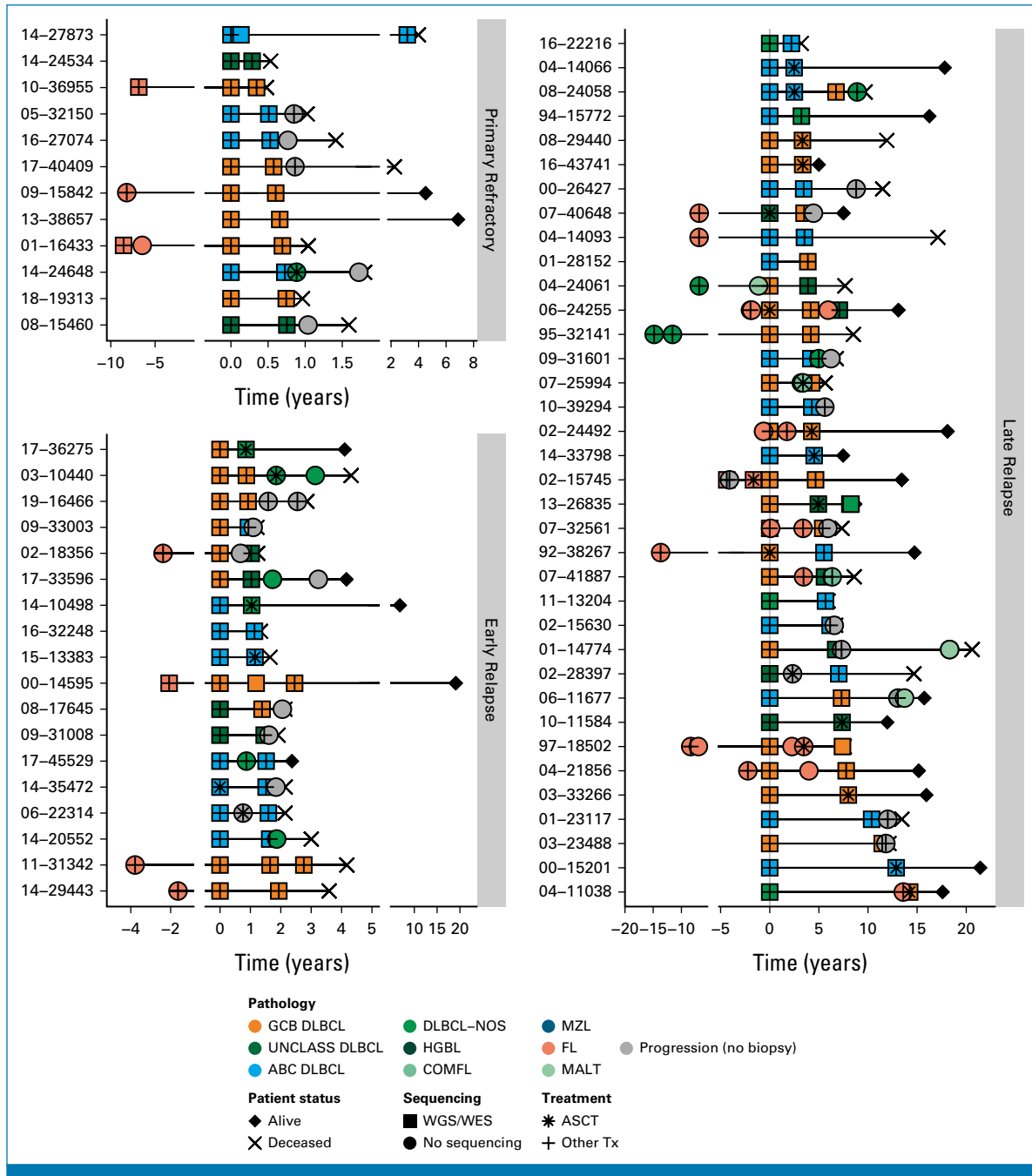
independent of genome coverage (Data Supplement [Table S9]). The linear relationship between exclusive variants and relapse timing was consistent when serial tFL tumors were considered separately from de novo DLBCL (Data Supplement [Supplemental Fig 7]). These results are consistent with a branching evolution model of evolution, where late relapse tumor pairs arise from a CPC harboring few lymphoma-defining mutations.

Given the high degree of divergence observed in some late relapse tumors, we used RNAseq data to identify functional expressed IG receptor rearrangements and confirm clonal relatedness of tumor pairs (Data Supplement [Table S10]). All four primary refractory and nine early relapse patients had concordant IGHV gene usage while 1 of 8 late relapses was discordant (Fig 3D). This lone patient with discordant heavy chain rearrangements also had discordant light chain rearrangements (Fig 3E), suggesting these tumors were not clonally related.

### Temporal Dynamics of Structural Variants

Rearrangements of *MYC*, *BCL2*, and *BCL6* are important drivers of aggressive lymphoma biology and contribute to disease and genetics-based classification.<sup>23,32,33</sup> *BCL2* rearrangement status was concordant in all 100 patients tested (Fig 4A and Data Supplement [Table S3]), consistent with the origin of *BCL2* rearrangements during V(D)J recombination in early B-cell differentiation.<sup>34</sup> In 26 patients where WGS identified *BCL2* breakpoints in two or more tumors, breakpoints were always identical (Data Supplement [Table S11]).

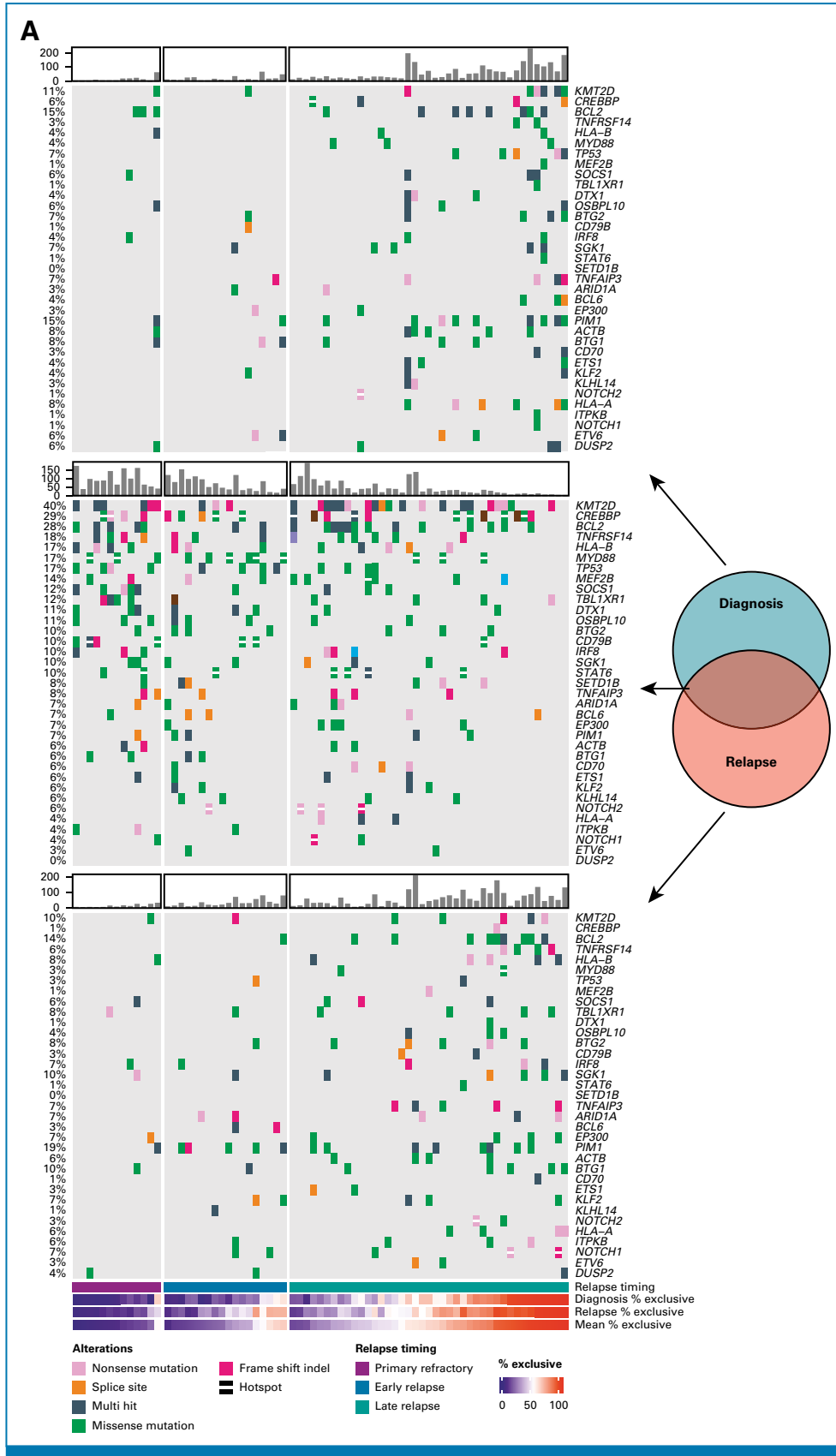
*MYC* and *BCL6* rearrangements were detected in 20 of 114 and 26 of 108 patients, respectively. In contrast to *BCL2* rearrangements, the rate of discordance in rearrangement status between biopsies was substantial at 70% of *MYC*-rearranged patients and 65% of *BCL6*-rearranged patients. Interestingly, in all 10 patients where *BCL6* rearrangements were



**FIG 2.** Sequencing cohort patient histories. Disease and treatment histories for all known biopsies and progression time points for patients for which WGS/WES data were generated, distributed according to relapse timing categories. Five patients were omitted because of incomplete histories. DLBCL tumors are colored according to NanoString cell-of-origin where available or are otherwise labeled DLBCL-NOS. ABC, activated B-cell-like DLBCL; ASCT, autologous stem-cell transplantation; COMFL, composite lymphoma with areas of DLBCL and FL morphology; DLBCL, diffuse large B-cell lymphoma; FL, follicular lymphoma; GCB, germinal center B-cell-like DLBCL; HGBL, high-grade B-cell lymphoma; MALT, extranodal MZL of mucosa-associated lymphoid tissue; MZL, marginal zone lymphoma; NOS, not otherwise specified; PROG, clinical progression without a biopsy; Tx, treatment; UNCLASS, unclassified DLBCL; WES, whole-exome sequencing; WGS, whole-genome sequencing.

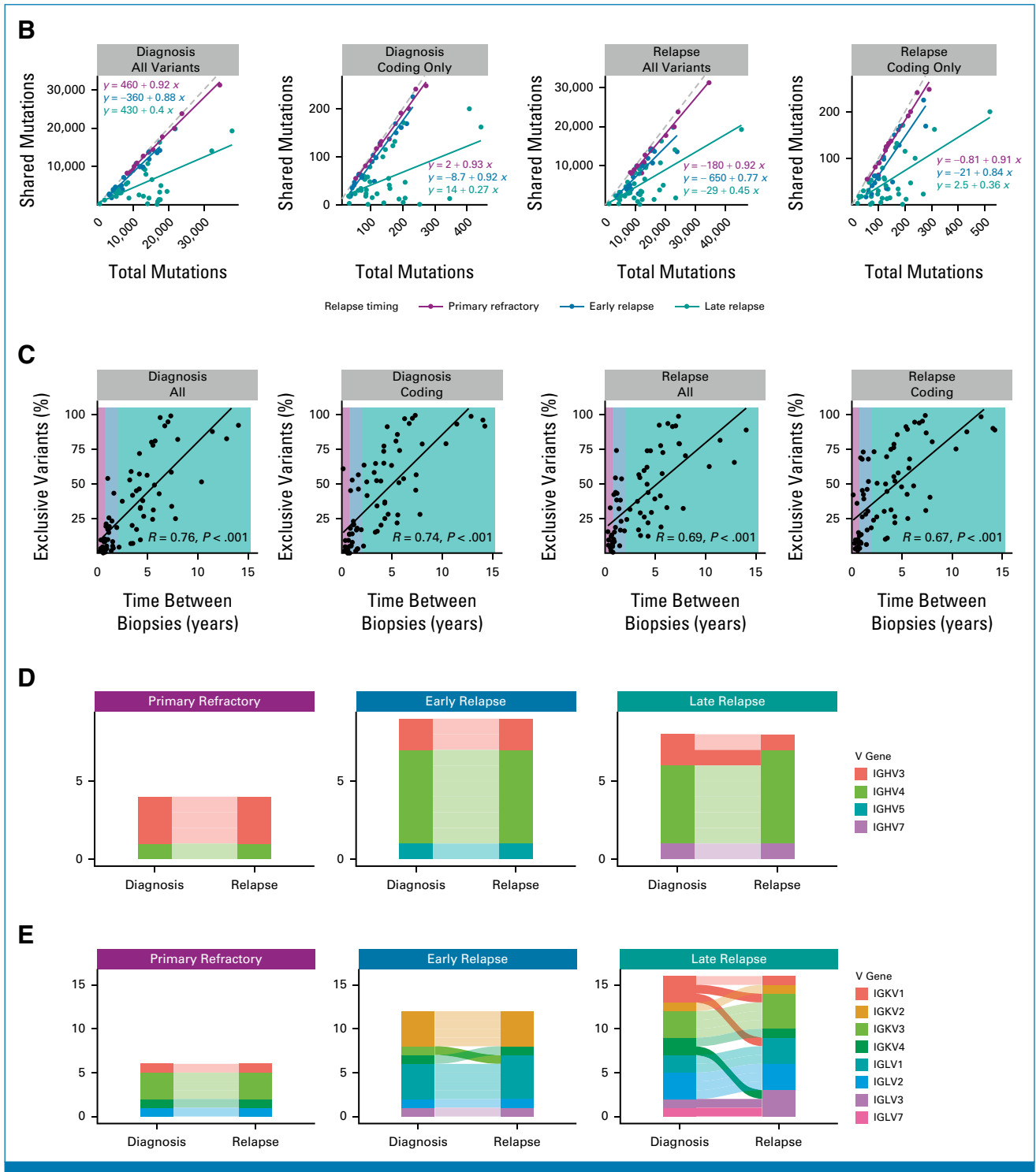
identified by WGS at multiple time points, the breakpoints were identical. *MYC* breakpoints were identified by WGS in multiple tumors from six patients, one of which was cryptic to FISH.<sup>35</sup> One patient with primary refractory disease, two with early relapse, and one late relapse had identical *MYC*

breakpoints in both tumors. However, two patients with late relapses (both high-grade B-cell lymphoma with *MYC* and *BCL2*) had different *MYC* translocation partners at diagnosis and relapse (Fig 3B and Data Supplement [Table S11]). These findings suggest that in some patients experiencing late

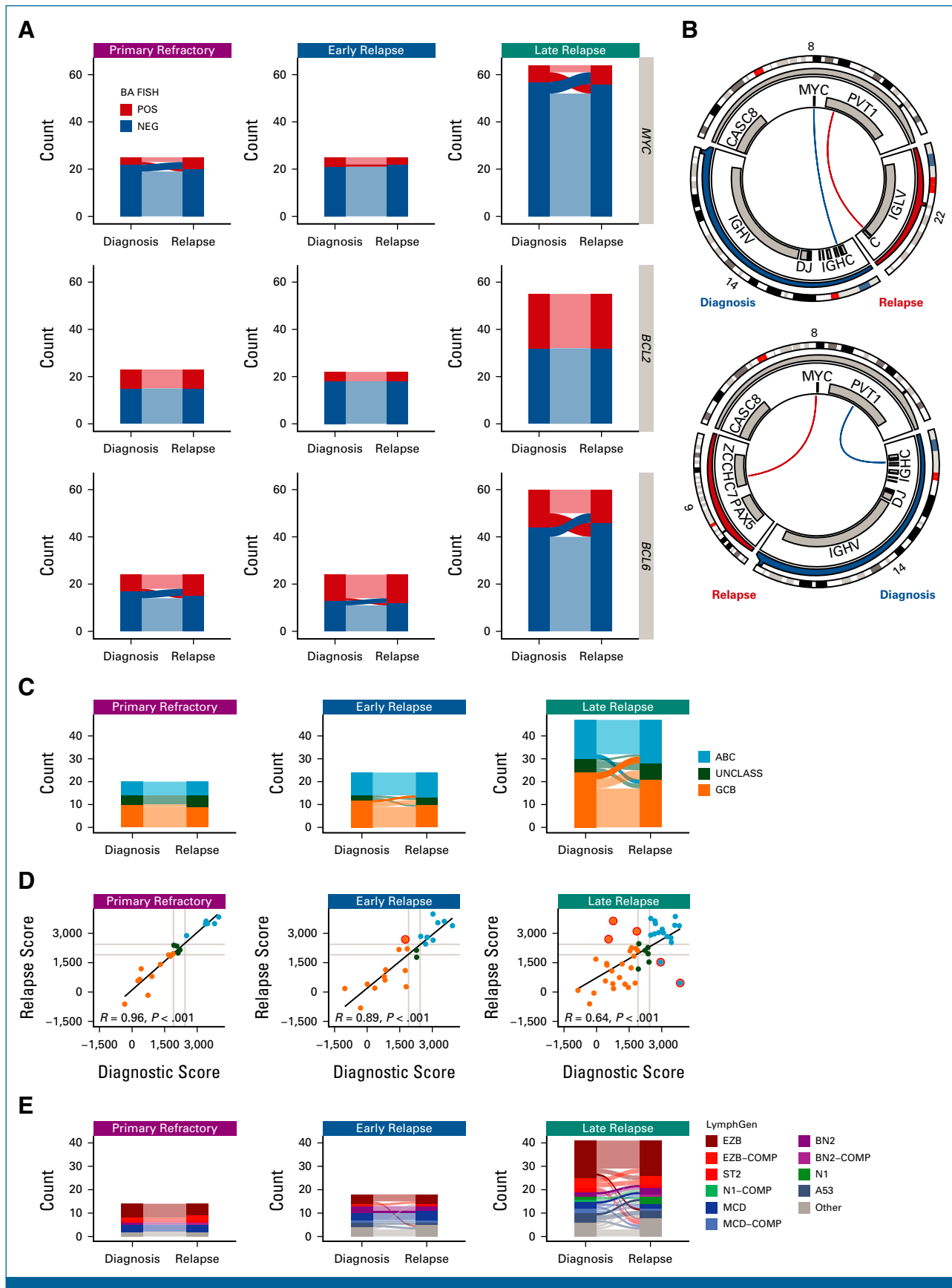


**FIG 3.** Patterns of evolution in diagnostic and relapse tumor pairs. (A) Oncoplots of variants identified exclusively at diagnosis (top), relapse (bottom), or shared between biopsies (middle), highlighting the most frequently mutated genes involved in LymphGen classification. Patients are stratified by relapse timing and ordered by the mean percentage of exclusive variants. (continued on following page)





**FIG 3.** (Continued). Barplots indicate the number of coding mutations present per patient in each mutation subset. (B) The relationship between total variants (all or coding only) at diagnosis or relapse versus the number of mutations shared between tumors. The dashed gray line represents the line of unity. (C) The percent of variants exclusive to either diagnostic or relapse tumors as a function of time between biopsies. R represents the Pearson correlation coefficient. (D) Concordance of heavy chain and (E) light chain V gene usage derived from RNAseq data for tumor pairs colored by V gene subgroup. Light chain rearrangements were more frequently discordant, which may suggest ongoing receptor editing.<sup>31</sup> In all plots, alluvia connecting each tumor pair are opaque for discordant pairs and translucent for concordant pairs. *N.B.* Where V gene usage was discordant but both V genes belong to the same subgroup, the color is consistent across time points.



**FIG 4.** Comparison of structural variants and gene expression profiling and genetic classifications between biopsies. (A) Concordance of BA-FISH results between diagnosis and first relapse for *MYC*, *BCL2*, and *BCL6* translocations. (B) Circos plots showing discordant *MYC* translocations in two patients who experienced late relapse. Top: a tumor pair that was positive for (continued on following page)



**FIG 4.** (Continued). BA-FISH at both timepoints; bottom: a tumor pair that was BA-FISH positive at diagnosis and negative at relapse. (C) Alluvial comparison of COO classifications in diagnostic/relapse pairs stratified by relapse timing. Frank discordance (ABC to GCB or vice versa) is indicated by opaque alluvia. (D) A scatter plot comparing DLBCL90 COO scores across tumor pairs. Red circles highlight frank discordance in COO classification. R values indicate Pearson correlation coefficient. (E) Comparison of LymphGen classifications between tumor pairs. Frank discordance (a switch between two mutually exclusive non-other classifications) is emphasized with opaque alluvia. ABC, activated B-cell-like DLBCL; BA, break-apart; COMP, composite; COO, cell-of-origin; FISH, fluorescence in situ hybridization; GCB, germinal center B-cell-like DLBCL; NEG, negative; POS, positive; UNCLASS, unclassified DLBCL.

relapse, the original *MYC*-translocated aggressive lymphoma is effectively eradicated by treatment, whereas the indolent CPC harboring a *BCL2* translocation or other variants can persist for many years with new *MYC* translocations arising in a subsequent aggressive lymphoma.

### Biological Consistency of Tumor Pairs

We next evaluated the consistency of molecular subgroups using GEP and LymphGen. We observed high levels of concordance of COO subtype between diagnosis and relapse in the 91 patients tested (Data Supplement [Table S3]). None of 20 primary refractory patients, only 1 of 24 early relapse patients (4.2%), and only 5 of 47 late relapses (10.6%) were frankly discordant (ie, activated B-cell-like (ABC) to germinal center B-cell-like (GCB) DLBCL or vice versa; Fig 4C). Comparison of the NanoString linear predictor scores between time points revealed a weaker correlation in late relapse patients (Fig 4D). A similar trend was observed in DZsig scores (Data Supplement [Supplemental Fig 8]).

To evaluate consistency in genetic subgroup assignment, we compared LymphGen classifications across 73 diagnostic/relapse tumor pairs, yielding a genetic classification for 80% of tumors. LymphGen classifications were highly concordant in all relapse timing categories, with discordance mainly occurring in patients with overlapping composite or Other (not assigned to any subgroup with sufficient confidence) tumors (Fig 4E, Data Supplement [Table S12]). A single early relapse patient of 18 (5.6%) patients had frank discordance (BN2 to MCD) and 4 of 41 (9.8%) among late relapses.

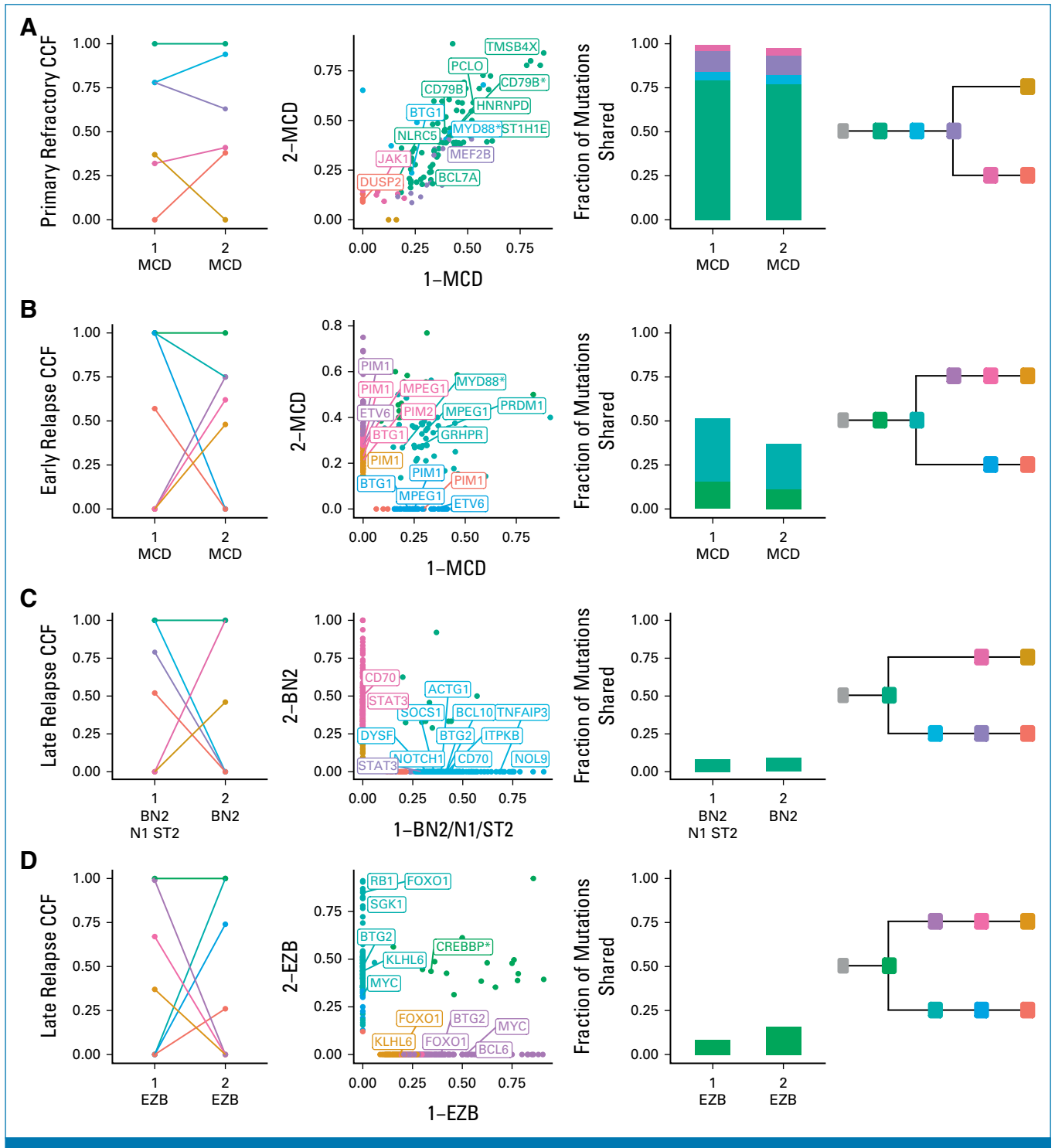
### Convergent Evolution in Divergent Pairs

The relative consistency of molecular subgroups as proxies for tumor biology is at odds with our observation that late relapses share relatively few mutations with the diagnostic tumor. To reconcile these disparities, we performed phylogenetic analyses for each patient, leveraging all coding mutations alongside noncoding mutations in regions known to be affected by aberrant somatic hypermutation (aSHM; Data Supplement [Table S13]).<sup>6</sup> In primary refractory tumors, most somatic variants are found in the shared phylogenetic trunk (clonal in both tumors; Fig 5A and Data Supplement [Supplemental Fig 9]). In early

relapse tumors, the trunk is comparatively shorter with branching evolution giving rise to exclusive mutations in both diagnostic and relapse tumors (Fig 5B). In late relapses, few mutations are in the trunk, with substantial divergence on each branch (Figs 5C and 5D). In the patient with discordant IGHV usage described earlier, the trunk comprised a single shared coding mutation (Fig 5E) providing further evidence that these tumors were not clonally related, arising independently.

Each of the genetic subgroups of DLBCL have distinct hallmark mutations. We examined patterns of evolution involving these hallmark mutations to determine whether they are present in the inferred CPC population. Variant calls from 28 patients with divergent patterns of evolution were used, defined as having at least 25% of mutations exclusive to each tumor. We then identified truncal (shared among all tumors from the same patient) and exclusive mutations. In total, 28 genes had truncal mutations detected in two or more patients (Fig 6A and Data Supplement [Table S14]), including *MYD88*<sup>L265P</sup> (4 of 5 mutated patients) and *CREBBP* KAT domain mutations (5 of 5 mutated patients), which are genetic subgroup hallmark mutations. In contrast, other genetic subgroup-defining mutations were less frequently truncal, including *NOTCH2* PEST domain truncating mutations (1 of 3), *EZH2*<sup>Y646</sup> (0 of 2), and *TET2* mutations (2 of 7). As individual mutations, for example *EZH2*<sup>Y646</sup>, may be considered for treatment selection or prognosis, this finding underscores the importance of recharacterizing late relapses. Loci affected by aSHM, including *BCL2*, *IGLL5*, and *BTG2*, had a high number of both truncal and exclusive mutations, suggesting that aSHM can be an early shared event but continues after divergence.

In addition, we noted that divergent tumor pairs harbored exclusive mutations in the same genes—an example of convergent evolution, where the tumors arrive at the same biology. In the representative MCD-classified early relapse tumor pair, each tumor had independently acquired class-defining mutations in *BTG1*, *PIM1*, and *ETV6* (Fig 5B); similarly the representative late relapse BN2 tumor pair in *CD70* (Fig 5C); and the representative late relapse EZB tumor pair in *FOXO1* and *MYC* (Fig 5D). This pattern of recurrent, independent mutation of the same genes was observed in 16 of 28 patients with divergent tumor evolution (Data Supplement [Tables S15 and S16]). We hypothesized that



**FIG 5.** Representative phylogenetic reconstructions. (A-E) Each row of plots displays data for a single patient. Tumors are labeled according to order of occurrence and LymphGen classification. Subclones are colored consistently across all plots for each patient. From left to right: CCF of subclones estimated by PhyClone; VAF of each variant as a scatter plot with the diagnostic tumor on the x-axis and relapse tumor on the y-axis with selected genes labeled; the fraction of mutations shared between both tumors (ie, all mutations in a cluster with a CCF > 0.1); and the inferred phylogenetic relationship between tumors. Hotspot mutations at *MYD88* L265P and *CD79B* Y179 and missense mutations in the *CREBBP* lysine acetyltransferase (KAT) domain are indicated with an asterisk. The VAF scatter plots without gene labels are also presented in the Data Supplement (Supplemental Fig 9). CCF, cancer cell fraction; VAF, variant allele frequency. (continued on following page)

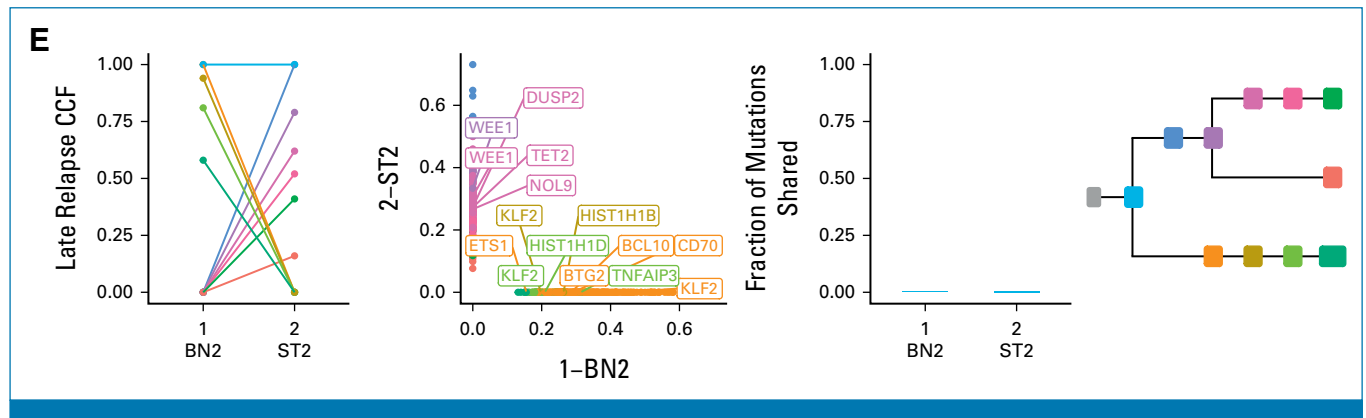


FIG 5. (Continued).

subgroup-defining mutations would be recurrent and concordant with the defined subgroup. Although this was observed in some features, such as MCD-related *PIM1* and *ETV6* in patients with MCD-classified tumors, others, such as *ACTB*, typically associated with the ST2 LymphGen class only recurred in patients without ST2-classified tumors (Fig 6B).

It has been speculated that the shared hallmark mutations between individual LymphGen subgroups and specific indolent lymphomas reflects shared evolutionary history and CPC features.<sup>23,25</sup> As expected on the basis of this model, patients with FL at any time in their disease course had DLBCL tumors predominantly classified as EZB while the few marginal zone lymphoma/extranodal MZL of mucosa-associated lymphoid tissue lymphomas occurred in patients with BN2- and ST2-classified DLBCLs (Fig 6C).

## DISCUSSION

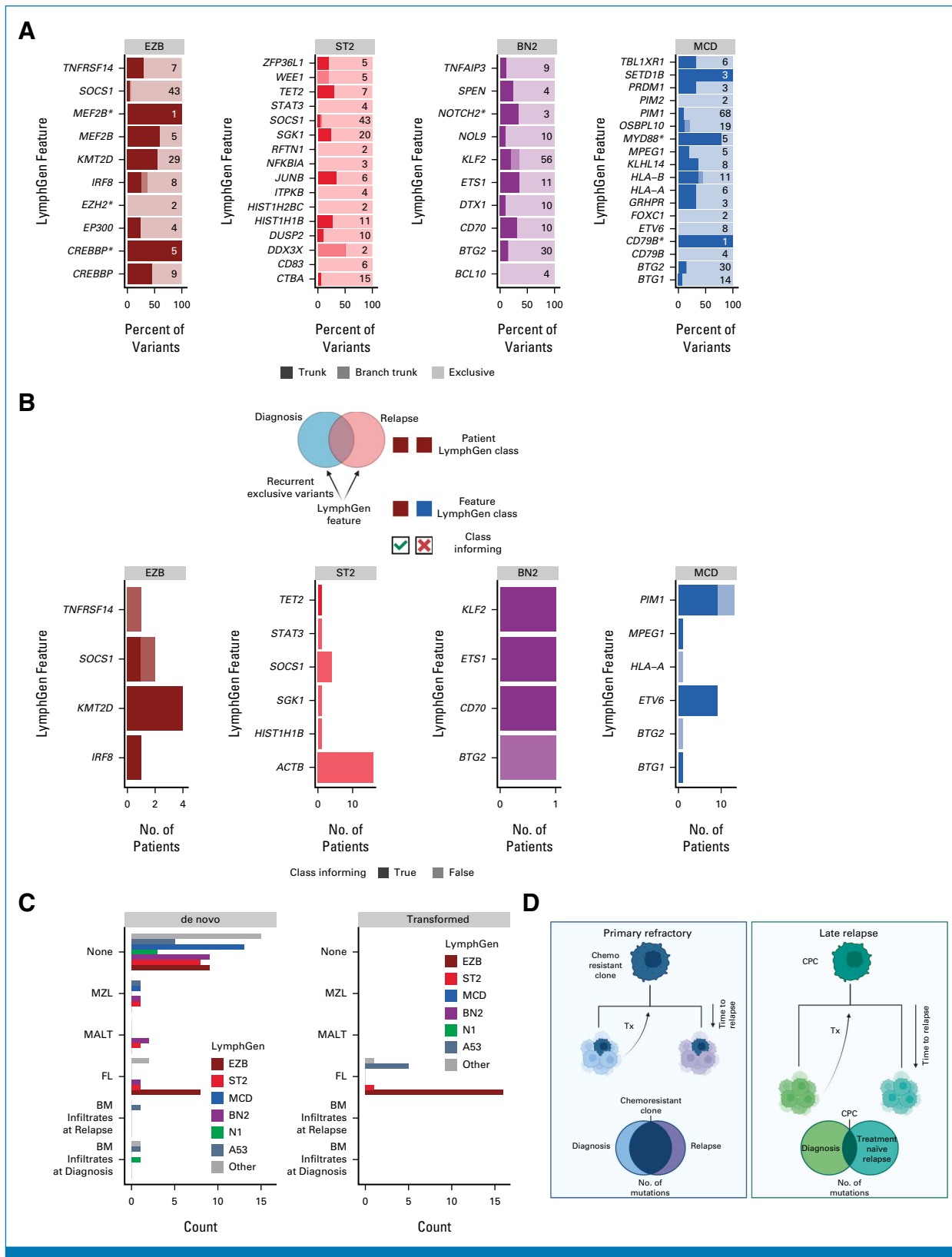
Leveraging multiple metrics of tumor evolution including cytogenetics, GEP, and unbiased genome- and exome-wide sequencing, we have established distinct patterns of tumor evolution that correlate strongly with the timing of DLBCL progression/relapse. The high rate of mutations exclusive to both diagnostic and relapse biopsies shows that branching evolution predominates in late relapses, strongly supporting the existence of persistent CPC populations capable of giving rise to multiple DLBCL manifestations over time. GEP- and genetics-based classifications remain consistent, suggesting that the earliest mutations in a CPC constrain the biology of subsequent DLBCL(s). This constrained evolution may be the basis for the remarkable convergence of the three studies that defined genetic subgroups of DLBCL.<sup>7,8,24,25</sup> Subgroup-defining mutations in the LymphGen classification were sometimes among the inferred CPC mutations identified while others were not consistently clonal, suggesting that additional genomic aberrations or other nongenetic features, such as DNA methylation or tumor microenvironment, are still to be discovered. Furthermore, these CPC mutations appear to constrain the set of loci that acquire mutations

during tumorigenesis. Larger cohorts and model systems will be required to determine the mechanisms of these observed constraints. Patterns of aSHM, known to reflect highly actively transcribed regions, are strongly associated with genetic subgroup<sup>23</sup> and thus may reflect B-cell differentiation states<sup>36,37</sup> that have been occupied during the evolution from CPC to DLBCL.

The patterns of DLBCL tumor evolution observed here help explain the responses to second-line (immuno)chemotherapy observed at disease relapse in DLBCL. In primary refractory disease, the pattern of tumor evolution suggests that innate chemoresistance is present at diagnosis, with little change in the composition of mutations on treatment (Fig 6D). The population of primary refractory patients should, therefore, be the focus in identifying both genetic and nongenetic mechanisms of resistance to frontline immunochemotherapy. In this study and others, these tumors do not typically respond to (immuno)chemotherapy-based second-line regimens, and outcomes are poor<sup>3</sup> while alternatives to chemotherapy result in superior outcomes in this patient population.<sup>38,39</sup>

In contrast, our observations of the biology of late relapse are consistent with elimination of the original DLBCL but persistence of a CPC harboring a very small number of mutations. These CPC populations subsequently give rise to a genetically divergent DLBCL with a large number of newly acquired mutations (Fig 6D). Although they share genetic features, the repertoire of driver mutations in the relapse is not preserved. As these late relapses are effectively chemotherapy naïve, immunochemotherapy-based regimens may remain a rational treatment option.

This divergence in evolutionary patterns and subsequent (immuno)chemotherapy sensitivity associated with relapse timing suggests that clinical trials and routine management should consider patients with early versus late relapse as distinct groups. The trials demonstrating the superiority of second-line chimeric antigen receptor T-cell therapy compared with immunochemotherapy



**FIG 6.** Classification features in divergent tumor pairs. (A) LymphGen classification features that were mutated in two or more tumors from the same patient. Trunk variants (darkest) are identical in all tumors from the same patient; branch trunk are variants not shared across all tumors but are common between at least two; and exclusive are those found in only one tumor. Numbers on each bar represent the total number of variants considered in each feature. (B) LymphGen classification (continued on following page)

**FIG 6.** (Continued). features that acquired exclusive variants in two or more tumors from the same patient. Class informing indicates that the mutations arose in patients in which LymphGen classification matched the class association of the acquired variants. (C) Patient LymphGen classifications stratified according to associated low-grade lymphoma entities. Transformed indicates low-grade disease preceded the first DLBCL diagnosis while de novo indicates that the low-grade diagnosis was made after DLBCL diagnosis. (D) A model of the relationship between relapse timing, evolutionary patterns, and outcomes. BM, bone marrow; CPC, common precursor cell; DLBCL, diffuse large B-cell lymphoma; FL, follicular lymphoma; MALT, extranodal MZL of mucosa-associated lymphoid tissue; MZL, marginal zone lymphoma; Tx, treatment. Created with [BioRender.com](https://BioRender.com).

and ASCT were performed in patients experiencing progression/relapse within 12 months of end of frontline therapy.<sup>38,39</sup> Larger cohorts than those examined in this study will be required to determine whether a specific time-to-relapse interval can be defined that reliably differentiates patients where the disease represents

true rrDLBCL as opposed to new DLBCL arising from a CPC population. Furthermore, particularly when precision medicine approaches are being considered, molecular characterization of the tumor at relapse is recommended as mutations and specific pathway perturbations may differ from those observed at the time of diagnosis.

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## PREPRINT VERSION

Preprint version available on medRxiv.

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## PRIOR PRESENTATION

Presented in part at the International Conference for Malignant Lymphoma, Lugano, Switzerland, June 18-22, 2021 and at the American Society of Hematology annual meeting, Atlanta, GA, December 10-13, 2021. Presented at the American Society of Hematology annual meeting, New Orleans, LA, December 11, 2022.

## SUPPORT

Supported by a Large Scale Applied Research Project funded by Genome Canada (13124), Genome BC (271LYM), Canadian Institutes of Health Research (CIHR; GP1-155873), the British Columbia Cancer Foundation (BCCF) and the Provincial Health Services Authority (PHSA). It was also supported by Terry Fox Research Institute (TFRI) Program Project Grant Nos. (1061, 1108), TFRI Marathon of Hope Cancer Centre Network, the Genome BC Marathon of Hope Cancer Centre program (MOH001), and Grant No. 1P01CA229100 from the National Cancer Institute. R.D.M. and C.S. are supported by Michael Smith Foundation for Health Research, Career Investigator Awards. D.W.S. is supported by a Michael Smith Foundation for Health Research, Health Professional Investigator Award. B.C. is supported by a CIHR Canada Graduate Scholarship Doctoral Award.

## AUTHORS' DISCLOSURES OF POTENTIAL CONFLICTS OF INTEREST

Disclosures provided by the authors are available with this article at DOI <https://doi.org/10.1200/JCO.23.00570>.

## DATA SHARING STATEMENT

Complete sequencing data including WGS/WES, RNAseq, and targeted sequencing will be deposited at the European Genome/Phenome Archive (EGA) under study ID EGAS00001007053. All code for variant calling and phylogenetic analysis are available as part of LCR-modules. An extensive repository of code for statistical analysis is available at <https://github.com/LCR-BCCRC/DLBCL-tumor-evolution>.



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## AUTHORS' DISCLOSURES OF POTENTIAL CONFLICTS OF INTEREST

### Relapse Timing Is Associated With Distinct Evolutionary Dynamics in Diffuse Large B-Cell Lymphoma

The following represents disclosure information provided by authors of this manuscript. All relationships are considered compensated unless otherwise noted. Relationships are self-held unless noted. I = Immediate Family Member, Inst = My Institution. Relationships may not relate to the subject matter of this manuscript. For more information about ASCO's conflict of interest policy, please refer to [www.asco.org/rwc](http://www.asco.org/rwc) or [ascopubs.org/jco/authors/author-center](http://ascopubs.org/jco/authors/author-center).

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No other potential conflicts of interest were reported.