

The genetic variant SLC2A1-rs1105297 is associated with the differential analgesic response to a glucose-based treatment in newborns

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Abstract

Neonatal pain is a critical issue in clinical practice. The oral administration of glucose-based solutions is currently one of the most common and effective nonpharmacologic strategies for neonatal pain relief in daily minor procedures. However, a varying degree of analgesic efficacy has been reported for this treatment. Environmental, maternal, and genetic factors may explain this variability and potentially allow for a personalized analgesic approach, maximizing therapeutic efficacy and preventing side effects. We investigated the exposome (ie, the set of clinical and anthropometric variables potentially affecting the response to the therapy) and the genetic variability of the noradrenaline transporter gene (solute carrier family 6 member 2 [SLC6A2]) and 2 glucose transporter genes (solute carrier family 2 member 1 [SLC2A1] and 2 [SLC2A2]) in relation to the neonatal analgesic efficacy of a 33% glucose solution. The study population consisted in a homogeneous sample of more than 1400 healthy term newborns. No association for the exposome was observed, whereas a statistically significant association between the G allele of SLC2A1-rs1105297 and a fourfold decreased probability of responding to the therapy was identified after multiple-testing correction (odds ratio of 3.98, 95% confidence interval 1.95-9.17; $P = 4.05 \times 10^{-4}$). This allele decreases the expression of SLC2A1-AS1, causing the upregulation of SLC2A1 in the dorsal striatum, which has been suggested to be involved in reward-related processes through the binding of opioids to the striatal mu-opioid receptors. Altogether, these results suggest the involvement of SLC2A1 in the analgesic process and highlight the importance of host genetics for defining personalized analgesic treatments.

Keywords: genetics, genetic variability, neonates, newborn, pharmacogenetics, precision medicine, personalized medicine, glucose, glucose transporters, SLC2A1, SLC2A2, noradrenaline, noradrenaline transporter, SLC6A2

1. Introduction

Pain is a medical issue in neonatal clinical practice, and several studies identified an association between neonatal pain and painrelated stress with short and long-term adverse health outcomes, such as sleep disorders,¹ reduced neuroanatomical development,²⁷ alterations in pain sensitivity,³⁷ and neurological disorders,^{3,16} which may last until infancy and childhood.²

Specific analgesic strategies have been developed to prevent neonatal pain.³³ Pharmacological approaches based on opioids,

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benzodiazepines, or other analgesic drugs are recommended for the most painful and invasive procedures, whereas nonpharmacological strategies are indicated for daily treatments such as heel prick and venipuncture.28,33,39 One of the most common and effective nonpharmacological treatment is the oral administration of a 20% to 33% glucose solution.¹⁰ Despite the high analgesic efficacy of this therapy, a fraction of newborns does not respond to the treatment, suggesting the involvement of host genetic factors.

Pharmacogenetic studies have already demonstrated the relevance of the genetic variability in determining interindividual differences in the response to specific analgesic treatments.^{34,36} For example, polymorphisms in OPRM1, COMT, and CYP2D6 genes have been associated with the efficacy of opioids-based analgesia.^{9,11,47} Some of these associations have been translated into clinical practice accounting for the genetic variability of the patient, when prescribing a pharmacological treatment.^{9,13} For example, the Food and Drug Administration suggests testing for CYP2D6 genotypes for codeine, oliceridine, and tramadol treatments and for CYP2C9 genotypes for meloxicam treatment.^{21,43}

The identification of genetic variants influencing the response to a treatment, in addition to allowing for the potential personalization of the therapy, can provide evidence for the mechanism of action of a drug.^{18,25,49} The mechanism by which glucose induces neonatal analgesia is still unclear, but one of the main hypotheses proposes the involvement of the endogenous

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opioid system.10,19,33 Accordingly, we reported in a previous study the association between a polymorphic variant, the missense mutation rs1799971, which belongs to the mu-opioid receptor (OPRM1) gene, and a reduced analgesia in 1077 healthy term newborns treated with oral glucose.¹⁷

Response to the treatment is a complex trait, and it is likely that many genetic polymorphisms may affect its therapeutic efficacy. With these premises, the goal of this study was to use genetic variability to identify in advance subjects nonresponding to the glucose-based treatment to allow for a different and personalized approach.

Many studies suggested a role for the norepinephrine transporter (NET) in opioids-mediated analgesia, highlighting a relationship between opioids and the noradrenergic system.^{7,42,45} We investigated whether the genetic variability of the solute carrier family 6 member 2 (SLC6A2) gene, coding for NET, could influence the response to the analgesic treatment based on glucose administration.

Moreover, since glucose is the effector of the neonatal analgesia, we hypothesize that genetic variants involved in glucose transport and metabolism may play a role in the response to the therapy. Thus, we also investigated whether single nucleotide polymorphisms (SNPs) within the solute carrier family 2 member 1 and 2 (SLC2A1 and SLC2A2) genes, coding for the ubiquitous glucose transporter GLUT-1 and the low-affinity glucose transporter GLUT-2, respectively, could affect the response to the therapy since SNPs in these 2 genes have been associated with hematic glucose concentration through genomewide association studies.^{12,23}

2. Materials and methods

2.1. Study subjects

The study was performed on 1421 healthy full-term newborns, enrolled between 2015 and 2022 at the Neonatology Unit of the Santa Chiara Hospital of Pisa (Italy). Inclusion criteria were term birth (gestational age of at least 37 weeks) an Apgar score, measured at 5 minutes after birth, of at least 7; and the parental subscription of an informed consent. Specifically, the Apgar score is a prognostic score used to assess the clinical conditions of newborns as soon after delivery. It is composed of 5 items (skin color, pulse, breathing, muscle tone, and reflex irritability) with a score for each one from 0 to 2. A normal Apgar score ranges from 7 to 10.

Exclusion criteria were defined as gestational age being lower than 37 weeks, an Apgar score lower than 7, suspicion of metabolic or genetic syndromes, or denied subscription of the parental informed consent. All subjects included in this study were of self-reported European ethnicity.

The Ethical Committee of Meyer Paediatric Hospital (Florence, Italy), which is the elected IRB for all paediatric studies in the Tuscany region of Italy, approved this study (registration number 84/2015). In addition, all clinical procedures were performed according to the ethical standards of the Declaration of Helsinki (1964).

2.2. Pain assessment

Following the guidelines defined by the Italian Neonatal Society for neonatal pain relief,²² all newborns were subjected to the oral administration of a 33% glucose solution a few minutes before the painful procedure, to provide nonpharmacological analgesia. The painful procedure consisted in a routine heel lancing for neonatal metabolic screenings.

The ABC scale, which has been validated in healthy term newborns,⁶ was used to assess pain after the administration of the therapy. The ABC scale is based on 3 parameters that evaluate crying intensity during the painful procedure: the pitch of the first cry, the constancy in time, and the rhythmicity of crying. 6 A score from 0 to 2 is assigned to each parameter and then each value is added up, and therefore, the ABC score may vary from a minimum of 0 (no pain) to a maximum of 6 (maximum pain intensity).

All clinical procedures including pain assessment were performed by trained personnel of the Neonatology Unit of the Santa Chiara Hospital.

2.3. Variables under study

The environmental exposome was defined as the set of clinical and anthropometric variables potentially influencing the response to the analgesic treatment. More specifically, it comprised gestational age (weeks), maternal age (years), birthweight (kgs, as continuous variable), maternal pregravidic weight (kgs, as continuous variable), neonatal type of feeding (breastfeeding/partial breastfeeding/formula milk), maternal gestational diabetes (yes or no), pregravidic diabetes (yes or no), insulin assumption (yes or no), smoking status (yes or no), sex (male or female), delivery mode (vaginal or caesarean delivery), type of analgesia during delivery (spinal or epidural or total analgesia), and maternal analgesic drugs assumption during the last 3 months of pregnancy (yes or no). Particularly, the variables gestational diabetes and pregravidic diabetes refer to 2 different conditions. The former is a transient glucose metabolism impairment related to pregnancy, whereas the latter indicates that the mother had diabetes (usually type I, insulin dependent) before pregnancy. As some mothers with gestational diabetes and all those with pregravidic diabetes may be treated with insulin, the variable insulin assumption was included.

2.4. Single nucleotide polymorphisms selection

Single nucleotide polymorphisms (SNPs) within the SLC2A1, SLC2A2, and SLC6A2 gene regions were selected based on a tagging approach to cover most of the genetic variability of these genes, using the Haploview Tagger Program software [\(https://](https://www.broadinstitute.org/haploview/haploview) [www.broadinstitute.org/haploview/haploview,](https://www.broadinstitute.org/haploview/haploview) version 4.2).⁵ The criteria for the selection of tagging SNPs were minor allele frequency (MAF) > 0.05 and $r^2 < 0.8$.

A set of 14, 3, and 19 SNPs was selected for SLC2A1, SLC2A2, and SLC6A2 genes, respectively. For each SNP, information related to the chromosome, position, gene, minor allele frequency (MAF) in the 1000 Genomes database for the TSI population, and functional annotation of the SNP (missense, synonymous, intronic, and noncoding) is reported in supplementary files (Table ST1, available at [http://links.lww.com/PAIN/](http://links.lww.com/PAIN/B918) [B918\)](http://links.lww.com/PAIN/B918).

2.5. DNA extraction and genotyping

Genomic DNA was extracted from blood cord with the automated QIAcube Connect extractor using the QIAamp DNA Blood Mini Kit as recommended by the producer. DNA concentration was quantified using a NanoDrop Lite UV–Vis Spectrophotometer (Thermo Fisher Scientific).

Genotyping was performed on 384 well plates using the TaqMan Assay system (Thermo Fisher Scientific, Waltham, MA), including 3.2% of duplicated samples for quality control purposes.

2.6. Statistical analysis

Hardy–Weinberg equilibrium was evaluated using the Pearson chi-square test.

Three different regression models were used to investigate the association between clinical and anthropometric variables and the response to the analgesic therapy. Logistic regression was applied to compare newborns with an ABC score > 0 with those with an ABC score $= 0$; a second logistic regression model was applied on a subset of individuals, to compare newborns not responding to the therapy and showing an high intensity of pain (ABC score \geq 5) with those responding to the therapy (ABC score $= 0$). Finally, an ordered logistic regression model was performed comparing all newborns within each ABC score category "k" (with k assuming values from 0 to 5), with those within the next category " $k + 1$ ", undertaking the proportional odds assumption.

The variables of the environmental exposome associated with the outcome at a P -value $<$ 0.05 were selected and additionally evaluated according to Akaike information criterion (AIC) and Bayesian information criterion (BIC), for covariates selection. In brief, AIC and BIC are mathematical estimators of the goodness of fit of a regression model. They allow the identification of the optimal model among several ones differing only for the number of independent variables included, thus allowing the selection of the ideal set of covariates.

The analyses of association between SNPs and ABC score were performed under dominant and additive allelic inheritance models adjusted for the covariates identified in the previous step. The dominant model compares homozygous carriers for the less common allele and heterozygous subjects (grouped together) with the homozygous carriers for the more common allele. The assumption underlying this model is that one copy of the effect allele is enough to increase the risk for the outcome compared with the group of homozygous subjects for the noneffect allele and that the effect of one copy or 2 copies of the effect allele have the same effect on the phenotype. The additive allelic model, instead, assumes that homozygous individuals for the effect allele and heterozygous individuals have a 2-fold and 1-fold increased risk, respectively, of getting the phenotype of interest compared with homozygous subjects for the other allele.

Bonferroni correction was applied to define a study-wide threshold for statistical significance, by dividing 0.05 by the number of the independent variables tested: $0.05/(36 + 16) =$ 9.62×10^{-4} . All statistical analyses were performed in RStudio, version 4.1.2.

2.7. Functional characterization of the single nucleotide polymorphisms

Several tools were used to investigate the functional effect of the SNPs associated with the response to the therapy. More specifically, the GTEx portal⁴ was used for identifying potential expression quantitative trait loci (eQTLs), which are SNPs that influence the expression of nearby genes; HaploReg v4.1⁴⁴ and RegulomeDB $2.0.3⁸$ were used to investigate the potential influence of the SNPs on DNA regulatory elements. Specifically, HaploReg provides data in relation to the effect of the SNPs on chromatin state and regulatory motifs. RegulomeDB provides a predictive score, intended as a probability score for the SNP to be functionally active, and a rank based on the amount of experimental evidence for the SNP to be functionally active. The rank goes from 1 to 6, where 1 indicates the highest evidence for functional or regulatory potential and 6 the lowest. The Combined Annotation Dependent Depletion (CADD) score was used to evaluate the deleteriousness of the SNPs.³⁵

3. Results

3.1. Genotyping results and quality control

The average genotyping call rate for the 36 SNPs was 98.77%, and the concordance rate between duplicates was higher than 99%. The observed MAF, call rate, and genotypes distribution for each SNP are also reported in Table ST1 [\(http://links.lww.com/](http://links.lww.com/PAIN/B918) [PAIN/B918](http://links.lww.com/PAIN/B918)).

SLC2A1-rs11537641 was not in Hardy–Weinberg equilibrium (Pvalue = 1.96×10^{-15}), but since the distribution of the genotypes was very similar to that reported in 1000 genomes for the TSI population, it was not excluded from the following analysis.

3.2. Characteristics of the study population

The average gestational age was 39.59 ± 1.14 weeks, and the male–female ratio of the newborns was about 1:1 (Table 1). The nonpharmacologic therapy was highly effective, as 141 newborns (of 1421) did not respond to the treatment. Among them, 8 had an ABC score of 1, 59 of 2, 38 of 3, 1 of 4, 21 of 5, and 14 of 6. For a subset of 54 subjects, the ABC score information was not available. Complete information on the variables of the exposome is reported in Table 1.

3.3. Association between the exposome and response to the therapy

None of the tested variables was associated with the response to the analgesic treatment ($P < 0.05$) in the model comparing newborns responding to the therapy with newborns not responding to the therapy (Table 2). However, a suggestive association was observed between maternal insulin assumption and a lower analgesic efficacy, with an OR of 1.86 (95% CI 0.96- 3.37) and a P -value = 0.052.

When comparing newborns responding (ABC score $= 0$) to the therapy with newborns with an ABC score ≥ 5 , partial breastfeeding (ie, feeding modality comprising both maternal breastfeeding and formula feeding) increased 3 times the probability of responding to the therapy compared with complete breastfeeding, with an OR of 0.33 (95% CI 0.10-0.86) and P -value = 0.041.

Two associations were observed in the ordered logistic regression model. Gestational age was associated with a higher analgesic efficacy, with an OR of 0.82 (95% CI 0.70-0.96) and Pvalue $= 0.013$ for each 1 week increase in gestational age. Instead, maternal age was associated with a lower analgesic efficacy with an OR of 1.05 (95% CI 1.01-1.08) and P-value =0.011 for each 1 year increase in maternal age. All results are reported in Table 2.

However, none of the 3 variables (feeding type, gestational age, and maternal age) was included as covariate in the genetic models after evaluation of AIC and BIC criteria.

3.4. Association between genetic variants and response to the analgesic therapy

After multiple-testing correction, one statistically significant association was observed for SLC2A1-rs1105297. The carriers of the G allele had a fourfold decreased chance of responding to

Table 1

The table reports the number of subjects for each variable based on the ABC score group (ABC score = 0 or ABC score > 0). In addition, the average and standard deviation values are reported for numeric variables, whereas the number of subjects for each level is additionally specified for categorical variables.

* It indicates the number of subjects for each variable.

† It stands for standard deviation.

the therapy in the model comparing newborns with ABC score $=$ 0 and newborns with ABC score \geq 5: ORs of 3.98 (95% CI 1.95-9.17, P-value=4.05 \times 10⁻⁴) and 4.18 (95% CI 1.88-10.10, P-value = 7.17×10^{-4}) in the additive allelic and in the dominant models, respectively (Table 3).

All other associations below the threshold of 0.05 in at least 1 of the 3 regression models are reported in Table 3. Notably, 3 SNPs were consistently associated with the ABC score throughout all regression models: SLC2A1-rs11210769, SLC2A1-rs11537641, and SLC6A2-rs12446977. However, none of them reached the adjusted threshold of statistical significance.

For all 3 SNPs, the largest ORs were observed in the model comparing newborns responding to the therapy with newborns with an ABC score \geq 5. More specifically, the C allele of SLC2A1-rs11210769 was associated with a fivefold decreased probability of responding to the analgesic treatment, with ORs of 4.69 (95% CI 1.30-33.33; P-value = 0.050) and 4.85 (95% CI 1.30-34.48; P -value = 0.048), in the additive allelic and in the dominant models, respectively.

The G allele of SLC2A1-rs11537641 was associated with a threefold decreased probability of responding to the therapy, with an OR of 3.14 (95% CI 1.26-9.62) and a P-value = 0.024 in both the additive allelic and the dominant genetic models. For SLC6A2-rs12446977, instead, the carriers of the G allele had a 2 to almost threefold decreased chance of responding to the therapy, depending on the genetic model: ORs of 1.96 (95% CI 1.16-3.33; P -value = 0.012) and 2.76 (95% CI 1.25-6.61; P-value = 0.016), in the additive allelic and dominant models, respectively.

The results for all the 36 SNPs are reported separately for each regression model in supplementary material—Tables ST2-4, available at<http://links.lww.com/PAIN/B918>.

3.5. Functional characterization of the single nucleotide polymorphisms

SLC2A1-rs1105297 is an eQTL for the SLC2A1-AS1 gene in the caudate and putamen of basal ganglia, with homozygous subjects for the G allele showing lower expression levels of SLC2A1-AS1. RegulomeDB assigned a rank of 5 to SLC2A1 rs1105297 (indicating a functional effect of the SNP in a transcription factor binding site or in a DNase site) and a score of 1.0 (corresponding to the maximum predicted value). In addition, both RegulomeDB and HaploReg suggested that the SNP could potentially modify transcription factor binding sites in several brain regions, such as the substantia nigra and the caudate. The CADD score was 0.261. No modification of gene expression was found for SLC2A1-rs11210769. However,

Table 2

Association between clinical and anthropometric variables and response to the therapy.

Variable	Logistic regression 1*		Logistic regression 2+		Ordered logistic regression‡	
	OR (95% CI)	P	OR (95% CI)	P	OR (95% CI)	Ρ
Sex (female vs male)	$0.77(0.54-1.09)$	0.145	$0.71(0.35-1.40)$	0.321	$0.88(0.62 - 1.25)$	0.475
Gestational age, wk	$1.02(0.87 - 1.19)$	0.848	$0.83(0.61 - 1.13)$	0.243	$0.82(0.70-0.96)$	0.013
Maternal age, y	$1.03(1.00-1.07)$	0.081	$1.00(0.94-1.07)$	0.988	$1.05(1.01-1.08)$	0.011
Birthweight, g	$1.00(1.00-1.00)$	0.890	$1.00(1.00-1.00)$	0.821	$1.00(1.00-1.00)$	1.000
Pregravidic weight, kg	$1.00(0.98-1.01)$	0.590	$1.00(0.97 - 1.03)$	0.980	$1.01(0.99-1.02)$	0.393
Feeding (partial breastfeeding vs breastfeeding)	$1.17(0.80 - 1.71)$	0.413	$0.33(0.10-0.86)$	0.041	$1.15(0.78 - 1.67)$	0.483
Feeding (formula milk vs breastfeeding)	$0.82(0.28-1.91)$	0.674	$1.17(0.19-4.07)$	0.830	0.81 (0.28-1.89)	0.656
Maternal gestational diabetic status (negative vs positive)	1.25 (0.79-1.91)	0.325	$0.96(0.32 - 2.32)$	0.928	1.25 (0.80-1.98)	0.330
Maternal pregravidic diabetic status (negative vs positive)	$2.35(0.53 - 7.62)$	0.195			$2.47(0.69 - 8.86)$	0.167
Maternal insulin assumption (no vs yes)	1.86 (0.96-3.37)	0.052	$1.15(0.18-3.90)$	0.853	1.43 (0.72-2.85)	0.312
Maternal smoking habit (negative vs positive)	$0.61(0.18-1.52)$	0.349	$1.44(0.23-4.95)$	0.623	$1.62(0.78-3.37)$	0.195
Delivery mode (vaginal delivery vs caesarean delivery)	$0.99(0.69 - 1.41)$	0.953	1.31 (0.66-2.60)	0.444	$1.11(0.78-1.59)$	0.550
Spinal anaesthesia (no vs yes)	$0.91(0.63 - 1.29)$	0.594	$1.08(0.52 - 2.19)$	0.828	$1.09(0.75-1.56)$	0.659
Epidural anaesthesia (no vs yes)	$1.08(0.69-1.63)$	0.731	$0.69(0.23 - 1.67)$	0.456	$0.68(0.41 - 1.11)$	0.121
Total anaesthesia (no vs yes)	$0.82(0.05-4.29)$	0.852				
Analgesics assumption (no vs yes)	1.24 (0.65-2.21)	0.489	1.90 (0.64-4.62)	0.193	$0.75(0.36-1.58)$	0.449

Table 2 reports the association results for the 3 regression models for each clinical and anthropometric variable of the environmental exposome. Associations with P-value < 0.05 are reported in bold.

* A logistic regression analysis was performed on all individuals to compare individuals not responding to the therapy with individuals responding to the therapy.

† A logistic regression analysis was applied on a subset of individuals, considering nonresponding subjects with a high ABC score (≥5) with responding newborns.

‡ An ordered logistic regression analysis was performed on all individuals, comparing subjects within a ABC score category with those within the higher-level ABC score category.

HaploReg suggested an effect of the SNP on regulatory enhancer sequences in several brain regions, whereas RegulomeDB assigned a rank of 2b (indicating evidence for a functional effect in transcription binding sites) and a score of 0.8. Moreover, SLC2A1-rs11210769 is in perfect LD with 7 other intronic noncoding variants (namely, rs79580038, rs80184186, rs75009191, rs12038788, rs12407435, rs60023956, and rs112981157), all of which are characterized by similar functional profiles according to both HaploReg and RegulomeDB. The CADD score for this variant was 5.537. SLC2A1-rs11537641 was not reported to be an eQTL in brain regions or other tissues of relevance for this study. HaploReg suggested the SNP to modify the effect of an enhancer or a promoter sequence, whereas RegulomeDB assigned a rank of 4 and a score of 0.61. However, the CADD score was quite high, showing a value of 29.50.

For SLC6A2-rs12446977 instead, neither GTEx nor HaploReg supported a potential functional effect of the SNP. RegulomeDB, instead, assigned a rank of 5 and score of 0.59 and suggested an association with a low or quiescent chromatinic state of the DNA region surrounding the SNP in several brain structures, among which the substantia nigra and the caudate nucleus. In addition, SLC6A2-rs12446977 is in perfect LD with 4 other intronic variants (SLC6A2-rs4436775, SLC6A2-rs2397772, SLC6A2-rs12920735, and SLC6A2-rs1861647). According to both HaploReg and RegulomeDB, all 4 genetic variants affect the structure of several DNA regulatory motifs. The CADD score for this variant was 2.341. Table ST5 in supplementary material shows a summary of all the functional annotation of the 4 SNPs, available at [http://links.lww.](http://links.lww.com/PAIN/B918) [com/PAIN/B918](http://links.lww.com/PAIN/B918).

4. Discussion

The oral administration of glucose-based solutions is an effective analgesic treatment to prevent neonatal pain¹⁰; however, around 10% of the newborns do not respond and still perceive pain. The underlying factors—especially genetic ones—contributing to the interindividual variability in response to the treatment have not been completely identified.

We investigated whether the exposome and the genetic variability of 3 transporters coding genes (namely, SLC2A1, SLC2A2, and SLC6A2) were associated with the differential response to the analgesic therapy in a population of more than 1400 healthy term newborns. This is the largest study up to date performed in this setting.

Despite analyzing a wide environmental exposome comprising many nongenetic variables, no statistically significant associations with the response to the therapy were identified. This may suggest that the exposome may not have a strong effect on the response to the therapy, or that we do not have enough statistical power to detect it, because of the relatively limited number of individuals with ABC score > 0 . Considering Bonferroni correction for multiple testing, we identified the statistically significant association between the G allele of SLC2A1-rs1105297 and a lower analgesic efficacy. The G allele of this SNP decreases the expression of the SLC2A1-AS1 gene according to GTEx, HaploReg, and RegulomeDB. SLC2A1-AS1 is a long noncoding RNA that downregulates the expression of SLC2A1.³⁸ The effect on the expression of SLC2A1-AS1 is specific for the putamen and the caudate nucleus, which together constitute the dorsal striatum that is enriched for the expression of mu-opioid receptors.26,48 Several evidence suggest the role of the dorsal striatum in reward-related and motivationTable 3

Association between genetic variants and response to the therapy. SNP Gene EA* NEA† Genetic model Logistic regression 1‡ Logistic regression 2§ Ordered logistic regression‖ OR (95% CI) P OR (95% CI) P OR (95% CI) P rs1105297 SLC2A1 G A Allelic model 1.38 (1.03-1.87) 0.036 3.98 (1.95-9.17) 4.05 x 10⁻⁴ 1.40 (1.04-1.89) 0.026 Dominant model 1.47 (0.98-2.20) 0.063 4.18 (1.88-10.10) 7.17 \times 10⁻⁴ 1.51 (1.01-2.25) 0.045 rs11210769 *SLC2A1* C T Allelic model 1.85 (1.10-3.32) 0.028 4.69 (1.30-33.33) 0.050 1.83 (1.06-3.13) 0.029 Dominant model 1.82 (1.05-3.32) 0.039 4.85 (1.30-34.48) 0.048 1.81 (1.03-3.17) 0.040 rs11537641 *SLC2A1* G A Allelic model 1.77 (1.15-2.78) 0.012 3.15 (1.26-9.62) 0.024 1.78 (1.15-2.78) 0.009 Dominant model 1.76 (1.14-2.78) 0.012 3.15 (1.26-9.62) 0.024 1.78 (1.15-2.75) 0.013 rs12446977 SLC6A2 G A Allelic model 1.35 (1.01-1.80) 0.041 1.96 (1.16-3.33) 0.012 1.37 (1.03-1.82) 0.031 Dominant model 1.58 (1.05-2.38) **0.029** 2.76 (1.25-6.61) **0.016** 1.60 (1.07-2.40) **0.024** rs3820546 *SLC2A1* A G Allelic model 1.33 (1.00-1.77) 0.047 1.27 (0.75-2.17) 0.377 1.32 (1.00-1.75) 0.052 Dominant model 1.67 (1.05-2.74) 0.036 1.93 (0.80-5.52) 0.174 1.69 (1.05-2.73) 0.031 rs3820548 *SLC2A1* G A Allelic model 1.52 (1.10-2.13) **0.013** 1.31 (0.73-2.46) 0.381 1.49 (1.09-2.08) **0.015** Dominant model 1.53 (1.01-2.30) 0.043 1.32 (0.60-2.85) 0.484 1.50 (1.00-2.25) 0.049 rs13330300 *SLC6A2* G A Allelic model 1.41 (0.96-2.15) 0.094 3.28 (1.31-11.11) **0.025** 1.41 (0.93-2.08) 0.099 Dominant model 1.42 (0.90-2.29) 0.143 3.42 (1.28-12.05) 0.027 1.42 (0.89-2.26) 0.143 rs40434 *SLC6A2* G A Allelic model 1.30 (0.97-1.75) 0.075 1.77 (1.02-3.13) **0.045** 1.32 (0.99-1.77) 0.06 Dominant model 1.24 (0.81-1.91) 0.326 2.13 (0.91-5.67) 0.101 1.26 (0.83-1.92) 0.283 rs710216 SLC2A1 A G Allelic model 1.19 (0.84-1.69) 0.326 2.99 (1.23-7.27) 0.016 1.22 (0.87-1.73) 0.248 Dominant model 1.28 (0.84-1.93) 0.249 3.06 (1.21-7.73) 0.018 1.31 (0.87-1.98) 0.192 rs710222 SLC2A1 A G Allelic model 1.02 (0.78-1.35) 0.873 1.66 (0.97-2.84) 0.065 1.01 (0.77-1.33) 0.922 Dominant model 1.09 (0.71-1.68) 0.689 2.54 (1.19-5.42) 0.016 1.15 (0.57-1.34) 0.532 rs1800887 SLC6A2 C T Allelic model 1.20 (0.85-1.72) 0.323 2.18 (0.98-4.88) 0.057 1.19 (0.75-1.76) 0.322 Dominant model 1.16 (0.56-1.31) 0.493 2.66 (1.12-7.41) 0.039 1.17 (0.56-1.30) 0.455

The table reports all associations below the threshold of 0.05 (bold text) in at least 1 of the 3 regression models.

* Effect allele (allele increasing the risk of not responding to the therapy).

† Noneffect allele (allele decreasing the risk of not responding to the therapy).

‡ The logistic regression analysis was performed on all individuals, comparing nonresponding newborns with newborns responding to the analgesic therapy.

§ The logistic regression analysis was performed on a subset of individuals, comparing those with a high ABC score with responding subjects.

‖ The ordered logistic regression analysis was performed on all subjects, comparing those within a specific ABC score category with those within the next ABC score category.

related functions.^{15,30,31} For example, the stimulation of mu-opioid receptors in the dorsal striatum affects eating behavior and generates motivation to gain a reward.¹⁵ In addition, the putamen has been suggested to contribute to sensory aspects of pain.⁴⁰

Therefore, as the glucose-induced analgesia is probably mediated by the release of endogenous opioids.^{10,19,33} the identification of a genetic variant specifically affecting the expression of SLC2A1 in the dorsal striatum, implicated in opioids-mediated reward, supports the involvement of SLC2A1 in this analgesic process.

In addition, we also observed that the C allele of SLC2A1 rs11210769 and the G allele of SLC2A1-rs11537641 were consistently associated with a lower analgesic efficacy throughout all regression models (although never reaching the Bonferroni-adjusted threshold).

Robust evidence for a functional effect in enhancer sequences in the brain, among which the caudate and the substantia nigra regions, were observed for SLC2A1-rs11210769 through HaploReg and RegulomeDB. The alteration of DNA binding sites for transcription factors or other regulatory proteins may directly affect the expression of SLC2A1 or other genes in such brain regions, which are involved in reward-related functions.^{15,30,31}ln line with this hypothesis, HaploReg reported an alteration of a motif (id: M00984) recognized by the transcription factor phosphatidylethanolamine binding protein (PEBP), coded by PEBP1. A study on the expression of PEBP1 in a mouse model highlighted a high expression level in the nucleus accumbens.⁴¹

As the involvement of the nucleus accumbens in reward, motivation and addiction are well established, $14,29$ it is likely that PEBP is involved in reward-related functions too.^{24,41} Thus, the modification of PEBP1 binding site (due to SLC2A1-rs11210769) in the nucleus accumbens may be an additional potential process regulating the response to the therapy.

SLC2A1-rs11537641 is instead a synonymous variant which may regulate the expression of SLC2A1 by influencing the kinetics of mRNA translation, according to codon usage bias principle.³² In brief, codon usage bias refers to the concept for which gene expression is influenced by the differential usage of some synonymous codons over the others. As a consequence of this nonrandom usage, there is a varying bioavailability of different aminoacyl-tRNA carrying the same amino acids, which causes different synonymous codons to be translated at different rates, thus influencing the processing of a transcript.

HaploReg and RegulomeDB both suggest a moderate functional effect for SLC2A1-rs11537641 in enhancer and DNA regulatory sequences in several regions.

An increased expression of SLC2A1 in the dorsal striatum may lead to a higher striatal glucose availability that in turn may affect the glucose-mediated reward and thus partially explain the differential response to the analgesic therapy.

An interesting association was also observed between the G allele of SLC6A2-rs12446977 and a lower analgesic efficacy of the therapy, which was consistent throughout all regression models. The involvement of SLC6A2 in relation to opioidsmediated analgesia has been already investigated.^{7,46} For example, knock-out models for SLC6A2 showed a higher effect of the endogenous opioids-related analgesia compared with wild-type animals because of an increase of extrasynaptic noradrenaline.⁷ However, no data have been published, to the best of our knowledge, on the effect of SLC6A2-rs12446977 (or other SNPs in LD with SLC6A2-rs12446977) on analgesia. Being an intronic variant, it may be possible that SLC6A2 rs12446977 is involved in the splicing process or that it is in LD with other functional SNPs. In line with this possibility, HaploReg reported the alteration of several DNA regulatory elements due to 4 intronic variants in perfect LD with SLC6A2 rs12446977 (SLC6A2-rs4436775, SLC6A2-rs2397772, SLC6A2-rs12920735, and SLC6A2-rs1861647). Such regulatory elements are recognized by different transcription factors and chromatin-remodeling proteins, which may influence the expression of SLC6A2.

A study focusing on the relationship between monoamine transporters and pain response after oral surgery identified a time-dependent association between SLC6A2-rs40434 and analgesia onset in adults. 20 It is possible that a timedependent relationship between the genetic variability of SLC6A2 and analgesia may contribute to the reduced analgesic response observed for some newborns. However, since all newborns were subjected to heel prick puncture 5 minutes after the analgesic treatment, we are not able to explore this possibility. Anyway, as SLC6A2-rs40434 is not in LD with SLC6A2-rs12446977, our result represents an additional independent indication that the genetic variability of the SLC6A2 locus contributes to the interindividual differences in the analgesic responses.

To the best of our knowledge, this is the largest pharmacogenetic study performed in newborns. Another strength is represented by the homogeneity of the neonatal population analyzed and the availability of several maternal and neonatal variables to account for a wide environmental exposome. Possible limitations of this study are the relatively small number of individuals that do not respond to the therapy, and the limited variability in several of the clinical variables analyzed. These 2 factors together limit our power to detect small effects (OR $<$ 1.6) on the outcome for some of the variables tested (analgesics assumption, total and epidural anesthesia, insulin assumption, pregravidic diabetes, and smoking status). Therefore, it is likely that a larger sample size is required to gain sufficient statistical power to identify the genetic contribution of rare allelic variants and small effects of exposome variables.

In conclusions, we observed a study-wide significant association between the G allele of SLC2A1-rs1105297 and a lower analgesic efficacy of the therapy. As the G allele of this SNP also decreases the expression of the antisense RNA SLC2A1-AS1 in the dorsal striatum, which is involved in reward-related functions, we hypothesized that the genetic variability of SLC2A1 may affect the response to the therapy through the regulation of the striatal glucose availability. This result represents a significant step towards the accomplishment of personalized analgesic treatment, which in the future will allow assigning the best regimen according to the genetic variability of the patient, maximizing the chance of analgesic efficacy and minimizing the administration of low-effective treatments or related side effects.

Conflict of interest statement

The authors have no conflicts of interest to declare.

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References

- [1] Abdulkader HM, Freer Y, Garry EM, Fleetwood-Walker SM, McIntosh N. Prematurity and neonatal noxious events exert lasting effects on infant pain behaviour. Early Hum Dev 2008;84:351–355.
- [2] Agakidou E, Tsoni K, Stathopoulou T, Thomaidou A, Farini M, Kontou A, Karagianni P, Sarafidis K. Changes in physicians' perceptions and practices on neonatal pain management over the past 20 years. A survey conducted at two time-points. Front Pediatr 2021;9:10.
- [3] Anand KJS, Palmer FB, Papanicolaou AC. Repetitive neonatal pain and neurocognitive abilities in ex-preterm children. PAIN 2013;154: 1899–1901.
- [4] Ardlie KG, DeLuca DS, Segrè AV, Sullivan TJ, Young TR, Gelfand ET, Trowbridge CA, Maller JB, Tukiainen T, Lek M, Ward LD, Kheradpour P, Iriarte B, Meng Y, Palmer CD, Esko T, Winckler W, Hirschhorn JN, Kellis M, MacArthur DG, Getz G, Shabalin AA, Li G, Zhou YH, Nobel AB, Rusyn I, Wright FA, Lappalainen T, Ferreira PG, Ongen H, Rivas MA, Battle A, Mostafavi S, Monlong J, Sammeth M, Melé M, Reverter F, Goldmann JM, Koller D, Guigó R, McCarthy MI, Dermitzakis ET, Gamazon ER, Im HK, Konkashbaev A, Nicolae DL, Cox NJ, Flutre T, Wen X, Stephens M, Pritchard JK, Tu Z, Zhang B, Huang T, Long Q, Lin L, Yang J, Zhu J, Liu J, Brown A, Mestichelli B, Tidwell D, Lo E, Salvatore M, Shad S, Thomas JA, Lonsdale JT, Moser MT, Gillard BM, Karasik E, Ramsey K, Choi C, Foster BA, Syron J, Fleming J, Magazine H, Hasz R, Walters GD, Bridge JP, Miklos M, Sullivan S, Barker LK, Traino HM, Mosavel M, Siminoff LA, Valley DR, Rohrer DC, Jewell SD, Branton PA, Sobin LH, Barcus M, Qi L, McLean J, Hariharan P, Um KS, Wu S, Tabor D, Shive C, Smith AM, Buia SA, Undale AH, Robinson KL, Roche N, Valentino KM, Britton A, Burges R, Bradbury D, Hambright KW, Seleski J, Korzeniewski GE, Erickson K, Marcus Y, Tejada J, Taherian M, Lu C, Basile M, Mash DC, Volpi S, Struewing JP, Temple GF, Boyer J, Colantuoni D, Little R, Koester S, Carithers LJ, Moore HM, Guan P, Compton C, Sawyer SJ, Demchok JP, Vaught JB, Rabiner CA, Lockhart NC, Ardlie KG, Getz G, Wright FA, Kellis M, Volpi S, Dermitzakis ET. The Genotype-Tissue Expression (GTEx) pilot analysis: multitissue gene regulation in humans. Science 2015;348: 648–660.
- [5] Barrett JC, Fry B, Maller J, Daly MJ. Haploview: analysis and visualization of LD and haplotype maps. Bioinformatics 2005;21:263–265.
- [6] Bellieni Cv, Bagnoli F, Sisto R, Neri L, Cordelli D, Buonocore G. Development and validation of the ABC pain scale for healthy full-term babies. Acta Paediatr Int J Paediatrics 2005;94:1432–1436.
- [7] Bohn LM, Xu F, Gainetdinov RR, Caron MG. Potentiated opioid analgesia in norepinephrine transporter knock-out mice. J Neurosci 2000;20: 9040–9045.
- [8] Boyle AP, Hong EL, Hariharan M, Cheng Y, Schaub MA, Kasowski M, Karczewski KJ, Park J, Hitz BC, Weng S, Cherry JM, Snyder M. Annotation of functional variation in personal genomes using RegulomeDB. Genome Res 2012;22:1790–1797.
- [9] Bright DR, Petry N, Roath E, Gibb T. Engaging pharmacogenomics in pain management and opioid selection. Pharmacogenomics 2021;22: 927–937.
- [10] Bueno M, Yamada J, Harrison D, Khan S, Ohlsson A, Adams-Webber T, Beyene J, Stevens B. A systematic review and meta-analyses of nonsucrose sweet solutions for pain relief in neonates. Pain Res Manag 2013;18:153–161.
- [11] Campa D, Gioia A, Tomei A, Poli P, Barale R. Association of ABCB1/ MDR1 and OPRM1 gene polymorphisms with morphine pain relief. Clin Pharmacol Ther 2008;83:559–566.
- [12] Chen J, Spracklen CN, Marenne G, Varshney A, Corbin LJ, Luan J, Willems SM, Wu Y, Zhang X, Horikoshi M, Boutin TS, Mägi R, Waage J, Li-Gao R, Chan KHK, Yao J, Anasanti MD, Chu AY, Claringbould A, Heikkinen J, Hong J, Hottenga JJ, Huo S, Kaakinen MA, Louie T, März W, Moreno-Macias H, Ndungu A, Nelson SC, Nolte IM, North KE, Raulerson CK, Ray D, Rohde R, Rybin D, Schurmann C, Sim X, Southam L, Stewart ID, Wang CA, Wang Y, Wu P, Zhang W, Ahluwalia TS, Appel EVR, Bielak LF, Brody JA, Burtt NP, Cabrera CP, Cade BE, Chai JF, Chai X, Chang LC, Chen CH, Chen BH, Chitrala KN, Chiu YF, de Haan HG, Delgado GE, Demirkan A, Duan Q, Engmann J, Fatumo SA, Gayán J, Giulianini F, Gong JH, Gustafsson S, Hai Y, Hartwig FP, He J, Heianza Y, Huang T, Huerta-Chagoya A, Hwang MY, Jensen RA, Kawaguchi T, Kentistou KA, Kim YJ, Kleber ME, Kooner IK, Lai S, Lange LA, Langefeld CD, Lauzon M, Li M, Ligthart S, Liu J, Loh M, Long J, Lyssenko V, Mangino M, Marzi C, Montasser ME, Nag A, Nakatochi M, Noce D, Noordam R, Pistis G, Preuss M, Raffield L, Rasmussen-Torvik LJ, Rich SS, Robertson NR, Rueedi R, Ryan K, Sanna S, Saxena R, Schraut KE, Sennblad B, Setoh K, Smith AV, Sparsø T, Strawbridge RJ, Takeuchi F, Tan J, Trompet S, van den Akker E, van der Most PJ, Verweij N, Vogel M, Wang H, Wang C, Wang N, Warren HR, Wen W, Wilsgaard T, Wong A, Wood AR, Xie T, Zafarmand MH, Zhao JH, Zhao W, Amin N, Arzumanyan Z, Astrup A, Bakker SJL, Baldassarre D, Beekman M, Bergman RN, Bertoni A, Blüher M, Bonnycastle LL, Bornstein SR, Bowden DW, Cai Q, Campbell A, Campbell H, Chang YC, de Geus EJC, Dehghan A, Du S, Eiriksdottir G, Farmaki AE, Frånberg M, Fuchsberger C, Gao Y, Gjesing AP, Goel A, Han S, Hartman CA, Herder C, Hicks AA, Hsieh CH, Hsueh WA, Ichihara S, Igase M, Ikram MA, Johnson WC, Jørgensen ME, Joshi PK, Kalyani RR, Kandeel FR, Katsuya T, Khor CC, Kiess W, Kolcic I, Kuulasmaa T, Kuusisto J, Läll K, Lam K, Lawlor DA, Lee NR, Lemaitre RN, Li H, Lin SY, Lindström J, Linneberg A, Liu J, Lorenzo C, Matsubara T, Matsuda F, Mingrone G, Mooijaart S, Moon S, Nabika T, Nadkarni GN, Nadler JL, Nelis M, Neville MJ, Norris JM, Ohyagi Y, Peters A, Peyser PA, Polasek O, Qi Q, Raven D, Reilly DF, Reiner A, Rivideneira F, Roll K, Rudan I, Sabanayagam C, Sandow K, Sattar N, Schürmann A, Shi J, Stringham HM, Taylor KD, Teslovich TM, Thuesen B, Timmers PRHJ, Tremoli E, Tsai MY, Uitterlinden A, van Dam RM, van Heemst D, van Hylckama Vlieg A, van Vliet-Ostaptchouk JV, Vangipurapu J, Vestergaard H, Wang T, Willems van Dijk K, Zemunik T, Abecasis GR, Adair LS, Aguilar-Salinas CA, Alarcón-Riquelme ME, An P, Aviles-Santa L, Becker DM, Beilin LJ, Bergmann S, Bisgaard H, Black C, Boehnke M, Boerwinkle E, Böhm BO, Bønnelykke K, Boomsma DI, Bottinger EP, Buchanan TA, Canouil M, Caulfield MJ, Chambers JC, Chasman DI, Chen YDI, Cheng CY, Collins FS, Correa A, Cucca F, de Silva HJ, Dedoussis G, Elmståhl S, Evans MK, Ferrannini E, Ferrucci L, Florez JC, Franks PW, Frayling TM, Froguel P, Gigante B, Goodarzi MO, Gordon-Larsen P, Grallert H, Grarup N, Grimsgaard S, Groop L, Gudnason V, Guo X, Hamsten A, Hansen T, Hayward C, Heckbert SR, Horta BL, Huang W, Ingelsson E, James PS, Jarvelin MR, Jonas JB, Jukema JW, Kaleebu P, Kaplan R, Kardia SLR, Kato N, Keinanen-Kiukaanniemi SM, Kim BJ, Kivimaki M, Koistinen HA, Kooner JS, Körner A, Kovacs P, Kuh D, Kumari M, Kutalik Z, Laakso M, Lakka TA, Launer LJ, Leander K, Li H, Lin X, Lind L, Lindgren C, Liu S, Loos RJF, Magnusson PKE, Mahajan A, Metspalu A, Mook-Kanamori DO, Mori TA, Munroe PB, Njølstad I, O'Connell JR, Oldehinkel AJ, Ong KK, Padmanabhan S, Palmer CNA, Palmer ND, Pedersen O, Pennell CE, Porteous DJ, Pramstaller PP, Province MA, Psaty BM, Qi L, Raffel LJ, Rauramaa R, Redline S, Ridker PM, Rosendaal FR, Saaristo TE, Sandhu M, Saramies J, Schneiderman N, Schwarz P, Scott LJ, Selvin E, Sever P, ou ShuX, Slagboom PE, Small KS, Smith BH, Snieder H, Sofer T, Sørensen TIA, Spector TD, Stanton A, Steves CJ, Stumvoll M, Sun L, Tabara Y, Tai ES, Timpson NJ, Tönjes A, Tuomilehto J, Tusie T, Uusitupa M, van der Harst P, van Duijn C, Vitart V, Vollenweider P, Vrijkotte TGM, Wagenknecht LE, Walker M, Wang YX, Wareham NJ, Watanabe RM, Watkins H, Wei WB, Wickremasinghe AR, Willemsen G, Wilson JF, Wong TY, Wu JY, Xiang AH, Yanek LR, Yengo L, Yokota M, Zeggini E, Zheng W, Zonderman AB, Rotter JI, Gloyn AL, McCarthy MI, Dupuis J, Meigs JB, Scott RA, Prokopenko I, Leong A, Liu CT, Parker SCJ, Mohlke KL, Langenberg C, Wheeler E, Morris AP, Barroso I, de Haan HG, van den Akker E, van der Most PJ, de Geus EJC, van Dam RM, van Heemst D, van

Hylckama Vlieg A, van Willems van Dijk K, van der Harst P. The transancestral genomic architecture of glycemic traits. Nat Genet 2021;53: 840–860.

- [13] Crews KR, Monte AA, Huddart R, Caudle KE, Kharasch ED, Gaedigk A, Dunnenberger HM, Leeder JS, Callaghan JT, Samer CF, Klein TE, Haidar CE, van Driest SL, Ruano G, Sangkuhl K, Cavallari LH, Müller DJ, Prows CA, Nagy M, Somogyi AA, Skaar TC. Clinical pharmacogenetics implementation consortium guideline for CYP2D6, OPRM1, and COMT genotypes and select opioid therapy. Clin Pharmacol Ther 2021;110: 888–896.
- [14] Deadwyler SA, Hayashizaki S, Cheer J, Hampson RE. Reward, memory and substance abuse: functional neuronal circuits in the nucleus accumbens. Neurosci Biobehav Rev 2004;27:703–711.
- [15] Difeliceantonio AG, Mabrouk OS, Kennedy RT, Berridge KC. Enkephalin surges in dorsal neostriatum as a signal to eat. Curr Biol 2012;22: 1918–1924.
- [16] Doesburg SM, Chau CM, Cheung TPL, Moiseev A, Ribary U, Herdman AT, Miller SP, Cepeda IL, Synnes A, Grunau RE. Neonatal pain-related stress, functional cortical activity and visual-perceptual abilities in schoolage children born at extremely low gestational age. PAIN 2013;154: 1946–1952.
- [17] Erbi I, Ciantelli M, Farinella R, Tuoni C, Gentiluomo M, Moscuzza F, Rizzato C, Bedini A, Faraoni M, Giusfredi S, Tavanti A, Ghirri P, Campa D. Role of OPRM1, clinical and anthropometric variants in neonatal pain reduction. Sci Rep 2020;10:7019.
- [18] Gonçalves E, Segura-Cabrera A, Pacini C, Picco G, Behan FM, Jaaks P, Coker EA, van der Meer D, Barthorpe A, Lightfoot H, Mironenko T, Beck A, Richardson L, Yang W, Lleshi E, Hall J, Tolley C, Hall C, Mali I, Thomas F, Morris J, Leach AR, Lynch JT, Sidders B, Crafter C, Iorio F, Fawell S, Garnett MJ. Drug mechanism-of-action discovery through the integration of pharmacological and CRISPR screens. Mol Syst Biol 2020;16:e9405.
- [19] Harrison D, Beggs S, Stevens B. Sucrose for procedural pain management in infants. Pediatrics 2012;130:918–925.
- [20] Kim H, Lee H, Rowan J, Brahim J, Dionne RA. Genetic polymorphisms in monoamine neurotransmitter systems show only weak association with acute post-surgical pain in humans. Mol Pain 2006;2:24.
- [21] Kumar S, Kundra P, Ramsamy K, Surendiran A. Pharmacogenetics of opioids: a narrative review. Anaesthesia 2019;74:1456–1470.
- [22] Lago P, Garetti E, Merazzi D, Pieragostini L, Ancora G, Pirelli A, Bellieni CV, Neonatology on behalf of the PSG of the IS of. Guidelines for procedural pain in the newborn. Acta Paediatr 2009;98:932–939.
- [23] Lagou V, Mägi R, Hottenga JJ, Grallert H, Perry JRB, Bouatia-Naji N, Marullo L, Rybin D, Jansen R, Min JL, Dimas AS, Ulrich A, Zudina L, Gådin JR, Jiang L, Faggian A, Bonnefond A, Fadista J, Stathopoulou MG, Isaacs A, Willems SM, Navarro P, Tanaka T, Jackson AU, Montasser ME, O'Connell JR, Bielak LF, Webster RJ, Saxena R, Stafford JM, Pourcain BS, Timpson NJ, Salo P, Shin SY, Amin N, Smith AV, Li G, Verweij N, Goel A, Ford I, Johnson PCD, Johnson T, Kapur K, Thorleifsson G, Strawbridge RJ, Rasmussen-Torvik LJ, Esko T, Mihailov E, Fall T, Fraser RM, Mahajan A, Kanoni S, Giedraitis V, Kleber ME, Silbernagel G, Meyer J, Müller-Nurasyid M, Ganna A, Sarin AP, Yengo L, Shungin D, Luan J, Horikoshi M, An P, Sanna S, Boettcher Y, Rayner NW, Nolte IM, Zemunik T, Iperen Evan, Kovacs P, Hastie ND, Wild SH, McLachlan S, Campbell S, Polasek O, Carlson O, Egan J, Kiess W, Willemsen G, Kuusisto J, Laakso M, Dimitriou M, Hicks AA, Rauramaa R, Bandinelli S, Thorand B, Liu Y, Miljkovic I, Lind L, Doney A, Perola M, Hingorani A, Kivimaki M, Kumari M, Bennett AJ, Groves CJ, Herder C, Koistinen HA, Kinnunen L, Faire Ude, Bakker SJL, Uusitupa M, Palmer CNA, Jukema JW, Sattar N, Pouta A, Snieder H, Boerwinkle E, Pankow JS, Magnusson PK, Krus U, Scapoli C, de Geus EJCN, Blüher M, Wolffenbuttel BHR, Province MA, Abecasis GR, Meigs JB, Hovingh GK, Lindström J, Wilson JF, Wright AF, Dedoussis GV, Bornstein SR, Schwarz PEH, Tönjes A, Winkelmann BR, Boehm BO, März W, Metspalu A, Price JF, Deloukas P, Körner A, Lakka TA, Keinanen-Kiukaanniemi SM, Saaristo TE, Bergman RN, Tuomilehto J, Wareham NJ, Langenberg C, Männistö S, Franks PW, Hayward C, Vitart V, Kaprio J, Visvikis-Siest S, Balkau B, Altshuler D, Rudan I, Stumvoll M, Campbell H, van Duijn CM, Gieger C, Illig T, Ferrucci L, Pedersen NL, Pramstaller PP, Boehnke M, Frayling TM, Shuldiner AR, Peyser PA, Kardia SLR, Palmer LJ, Penninx BW, Meneton P, Harris TB, Navis G, Harst Pvander, Smith GD, Forouhi NG, Loos RJF, Salomaa V, Soranzo N, Boomsma DI, Groop L, Tuomi T, Hofman A, Munroe PB, Gudnason V, Siscovick DS, Watkins H, Lecoeur C, Vollenweider P, Franco-Cereceda A, Eriksson P, Jarvelin MR, Stefansson K, Hamsten A, Nicholson G, Karpe F, Dermitzakis ET, Lindgren CM, McCarthy MI, Froguel P, Kaakinen MA, Lyssenko V, Watanabe RM, Ingelsson E, Florez JC, Dupuis J, Barroso I, Morris AP, Prokopenko I. Sex-dimorphic genetic effects and novel loci for fasting glucose and insulin variability. Nat Commun 2021;12:995.
- [24] Ling HH, Mendoza-Viveros L, Mehta N, Cheng HYM. Raf kinase inhibitory protein (RKIP): functional pleiotropy in the mammalian brain. Crit Rev Oncog 2014;19:505–516.
- [25] Lötsch J, Geisslinger G. Pharmacogenetics of new analgesics. Br J Pharmacol 2011;163:447–460.
- [26] Meier IM, Eikemo M, Leknes S. The role of mu-opioids for reward and threat processing in humans: bridging the gap from preclinical to clinical opioid drug studies. Curr Addict Rep 2021;8:306.
- [27] Mooney-Leber SM, Brummelte S. Neonatal pain and reduced maternal care: early-life stressors interacting to impact brain and behavioral development. Neuroscience 2017;342:21–36.
- [28] da Motta GdeCP, da Cunha MLC. Prevention and non-pharmacological management of pain in newborns. Rev Bras Enferm 2015;68:131–135.
- [29] Norgren R, Hajnal A, Mungarndee SS. Gustatory reward and the nucleus accumbens. Physiol Behav 2006;89:531–535.
- [30] Nummenmaa L, Hirvonen J, Hannukainen JC, Immonen H, Lindroos MM, Salminen P, Nuutila P. Dorsal striatum and its limbic connectivity mediate abnormal anticipatory reward processing in obesity. PLoS One 2012;7:e31089.
- [31] Palmiter RD. Dopamine signaling in the dorsal striatum is essential for motivated behaviors. Ann N Y Acad Sci 2008;1129:35–46.
- [32] Parvathy ST, Udayasuriyan V, Bhadana V. Codon usage bias. Mol Biol Rep 2022;49:539.
- [33] Perry M, Tan Z, Chen J, Weidig T, Xu W, Cong XS. Neonatal pain: perceptions and current practice. Crit Care Nurs Clin North Am 2018;30: 549–561.
- [34] Relling Mv, Evans WE. Pharmacogenomics in the clinic. Nature 2015; 526:343–350.
- [35] Rentzsch P, Witten D, Cooper GM, Shendure J, Kircher M. CADD: predicting the deleteriousness of variants throughout the human genome. Nucleic Acids Res 2019;47:D886–D894.
- [36] Roses AD. Pharmacogenetics and future drug development and delivery. Lancet 2000;355:1358–1361.
- [37] Schwaller F, Fitzgerald M. The consequences of pain in early life: injuryinduced plasticity in developing pain pathways. Eur J Neurosci 2014;39: 344–352.
- [38] Shang R, Wang M, Dai B, Du J, Wang J, Liu Z, Qu S, Yang X, Liu J, Xia C, Wang L, Wang D, Li Y. Long noncoding RNA SLC2A1-AS1 regulates aerobic glycolysis and progression in hepatocellular carcinoma via

inhibiting the STAT3/FOXM1/GLUT1 pathway. Mol Oncol 2020;14: 1381–1396.

- [39] Squillaro A, Mahdi EM, Tran N, Lakshmanan A, Kim E, Kelley-Quon LI. Managing procedural pain in the neonate using an opioid-sparing approach. Clin Ther 2019;41:1701–1713.
- [40] Starr CJ, Sawaki L, Wittenberg GF, Burdette JH, Oshiro Y, Quevedo AS, McHaffie JG, Coghill RC. The contribution of the putamen to sensory aspects of pain: insights from structural connectivity and brain lesions. Brain 2011;134:1987–2004.
- [41] Theroux S, Pereira M, Casten KS, Burwell RD, Yeung KC, Sedivy JM, Klysik J. Raf kinase inhibitory protein knockout mice: expression in the brain and olfaction deficit. Brain Res Bull 2007;71:559–567.
- [42] Tura B, Tura SM. The analgesic effect of tricyclic antidepressants. Brain Res 1990;518:19–22.
- [43] USA Food, Drug Administration. Table of Pharmacogenomic Biomarkers in Drug Labeling. 2022. Available at: [https://www.fda.gov/drugs/](https://www.fda.gov/drugs/science-and-research-drugs/table-pharmacogenomic-biomarkers-drug-labeling) [science-and-research-drugs/table-pharmacogenomic-biomarkers](https://www.fda.gov/drugs/science-and-research-drugs/table-pharmacogenomic-biomarkers-drug-labeling)[drug-labeling.](https://www.fda.gov/drugs/science-and-research-drugs/table-pharmacogenomic-biomarkers-drug-labeling) Accessed November 7, 2022.
- [44] Ward LD, Kellis M. HaploReg: a resource for exploring chromatin states, conservation, and regulatory motif alterations within sets of genetically linked variants. Nucleic Acids Res 2012;40:D930–D934.
- [45] Ward NG, Bloom VL, Friedel RO. The effectiveness of tricyclic antidepressants in the treatment of coexisting pain and depression. PAIN 1979;7:331–341.
- [46] Yaksh TL. Pharmacology of spinal adrenergic systems which modulate spinal nociceptive processing. Pharmacol Biochem Behav 1985;22: 845–858.
- [47] Yang Y, Botton MR, Scott ER, Scott SA. Sequencing the CYP2D6 gene: from variant allele discovery to clinical pharmacogenetic testing. Pharmacogenomics 2017;18:673–685.
- [48] Zachariou V, Georgescu D, Sanchez N, Rahman Z, DiLeone R, Berton O, Neve RL, Sim-Selley LJ, Selley DE, Gold SJ, Nestler EJ. Essential role for RGS9 in opiate action. Proc Natl Acad Sci U S A 2003;100:13656–13661.
- [49] Zimmermann M, Murina O, Reijns MAM, Agathanggelou A, Challis R, Tarnauskaite Ž, Muir M, Fluteau A, Aregger M, McEwan A, Yuan W, Clarke M, Lambros MB, Paneesha S, Moss P, Chandrashekhar M, Angers S, Moffat J, Brunton VG, Hart T, de Bono J, Stankovic T, Jackson AP, Durocher D. CRISPR screens identify genomic ribonucleotides as a source of PARP-trapping lesions. Nature 2018;559:285.