

NOTES

Pharmacokinetics of Tenofovir in Breast Milk of Lactating Rhesus Macaques

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To study tenofovir transfer into milk, two lactating macaques were given a subcutaneous dose of tenofovir (30 mg/kg of body weight). Peak concentrations and area under the curve values of tenofovir in milk were ~3 and ~20% of those detected in serum, respectively.

For lactating mothers taking medications, the drug concentration in breast milk is usually lower than that in plasma; it is generally of negligible concern for the nursing infant, but some exceptions exist (reviewed in reference 2). A growing number of human immunodeficiency virus (HIV)-infected people in developing countries are gaining access to treatment with anti-HIV drugs, including lactating mothers for whom avoidance of breast-feeding is not always an option. Thus, questions are raised regarding possible biological implications of anti-HIV drugs that may be transferred to the nursing infant (4, 5). It is difficult to predict drug transfer into milk based on physicochemical properties (6). Accordingly, animal models can be useful to gather preliminary information on the transfer of compounds into breast milk prior to obtaining such data from human studies (1).

We performed a pilot study to determine the transfer of tenofovir {9-[2-(phosphonomethoxy)propyl]adenine; PMPA} in breast milk of rhesus macaques. Two healthy lactating adult rhesus macaques (*Macaca mulatta*), which were multiparous and 5 to 11 years of age, were used. The animals were housed in accordance with American Association for Accreditation of Laboratory Animal Care standards, and we strictly adhered to the *Guide for the Care and Use of Laboratory Animals* (9). When necessary, animals were immobilized with ketamine HCl (Parke-Davis, Morris Plains, New Jersey) at a concentration of 10 mg/kg of body weight injected intramuscularly. Both female macaques had been lactating for 10 to 11 weeks, and their infants had been weaned the day prior to the pharmacokinetic study. A single dose of tenofovir (30 mg/kg) was administered subcutaneously. Pre- and postdose blood samples (without anticoagulant) were collected over a 24-h time period (at 0, 0.5, 1, 2, 4, 6, 8, and 24 h, with an additional time point at 10 h for animal 24964) and were spun immediately for the collection of serum; at the same times, all milk that could be expressed manually from both nipples was collected (up to 7.5 ml per time point). Our study has the caveat that this milk collection

schedule (especially the absence of sample collections between 10 and 24 h of dosing) does not completely mimic the more regular drinking activity of a nursing infant macaque. Serum and milk samples were stored at -70°C and subsequently analyzed by MDS Pharma Services (Montreal, Canada) using high-performance liquid chromatography methods with mass spectrometry detection (liquid chromatography-mass spectrometry-mass spectrometry), previously validated for monkey plasma and rat milk (unpublished data). For monkey plasma, the limit of quantitation was 10 ng/ml, standard curve linearity r^2 was 0.999, and within- and between-run quantitative comparison [QC] accuracy and precision were $<2\%$ bias and 95%, respectively; for monkey milk, the limit of quantitation was 10 ng/ml, r^2 was 0.9932, and within-run QC accuracy was $<3\%$ bias. Tenofovir concentrations were measured in whole milk (i.e., without prior separation of the different milk fractions). The values of the pharmacokinetic parameters were derived by noncompartmental analysis with WinNonlin software (version 3.1; Pharsight Corporation, Mountain View, California).

Tenofovir concentrations in serum and milk are shown in

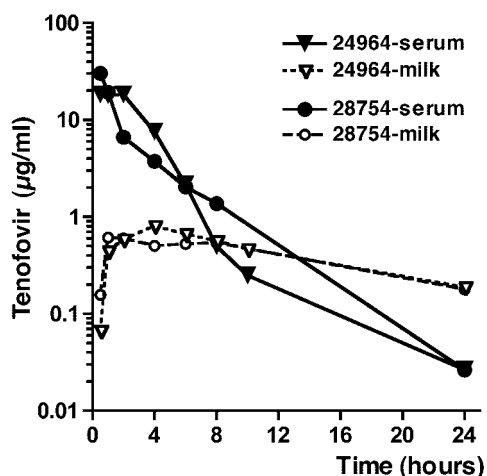


FIG. 1. Concentrations of tenofovir in serum and milk following a single subcutaneous dose of 30 mg of tenofovir/kg of body weight.

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TABLE 1. Tenofovir pharmacokinetics in lactating rhesus macaques

Animal no. ^a	Sample	C _{max} (μg/ml)	T _{max} (h)	C _{24h} (μg/ml)	AUC _{0-∞} (μg · h/ml)	t _{1/2} (h)	CL/F (ml/h/kg)	V _d /F (ml/kg)
24964	Serum	18.3	0.5	0.026	68.9	3.97	435	2,489
	Milk	0.808	4	0.188	12.8	10.3		
28754	Serum	30.2	0.5	0.026	56.2	2.85	534	2,191
	Milk	0.610	1	0.179	12.1	10.9		

^a Animals 24964 and 28754 were 11 and 5 years of age, respectively. A single dose of tenofovir (30 mg/kg) was administered subcutaneously. Abbreviations: C_{max}, maximum concentration; T_{max}, the time to C_{max}; C_{24h}, concentration at 24 h; AUC_{0-∞}, area under the concentration-versus-time curve extrapolated to infinity; t_{1/2}, half-life of terminal elimination phase; CL/F, apparent clearance; V_d/F, apparent volume of distribution based on terminal phase. CL/F and V_d/F were not calculated for milk. Each individual concentration profile was reviewed to assure accuracy of the pharmacokinetic analysis.

Fig. 1, while pharmacokinetic parameters are presented in Table 1. Tenofovir was detected in the milk of both animals, but the peak concentrations (~0.6 to 0.8 μg/ml, corresponding to ~2 to 3 μM) were ~2 to 4% of those detected in serum, with milk area under the curve (AUC) values being ~20% of the serum AUC values.

Other anti-HIV drugs (nevirapine, zidovudine, and lamivudine) are also found in breast milk (7, 8). Treatment of HIV-infected lactating mothers with anti-HIV drugs is expected to benefit the infant indirectly (by improving the mother's health and reducing maternal systemic virus levels, thus potentially lowering the infectivity of the breast milk). However, could such levels of anti-HIV drugs in breast milk have any direct biological effects, either harmful or beneficial for the infant? It was recently reported that concentrations of lamivudine and nevirapine in breast milk were high enough to give detectable serum levels in their nursing infants, which may provide prophylactic effects but may also have toxic effects (R. Shapiro et al., 42nd Ann. Meet. Infect. Dis. Soc. Am., Boston, Late Breaker abstr. LB-1, 2004). Concerns have also been raised that infants who become infected may be exposed for relatively long periods to subtherapeutic levels of drug, which may lead to resistance and limit the future treatment options for the infant (4, 5).

For breast-feeding mothers taking the orally bioavailable prodrug tenofovir disoproxil fumarate, breast milk is expected to contain almost exclusively the parental compound tenofovir, which due to its charged anionic nature exhibits low oral bioavailability in animals (5% in cynomolgus macaques) and is expected to also show low oral bioavailability after ingestion by the nursing infant (3, 10). A previous study in macaques suggests that the concentration of tenofovir in milk is unlikely to give topical prophylaxis against oral HIV infection (13). Instead, infant macaque studies suggest that direct administration of tenofovir disoproxil fumarate to the nursing infant at regimens that give systemic drug levels is needed to prevent infection through breast-feeding (12; K. Van Rompay, J. Lawson, R. Colón, N. Bischofberger, and M. Marthas, XV Int. AIDS Conf., Bangkok, Late Breaker abstr. LbOrB10, 2004). Considering the volume of ingested breast milk, the tenofovir concentrations we observed in breast milk of lactating macaques are unlikely to be toxic for the infant, especially because our previous studies demonstrated a favorable safety profile of prolonged daily treatment of infant macaques with a dose of tenofovir (10 mg/kg subcutaneously) that is much higher than the daily amount of tenofovir likely to be ingested and absorbed from breast milk (11). Because of its low oral bioavailability, small amounts of tenofovir in milk are also very unlikely to select for resistance in an already infected infant, thus preserving future treatment options.

In conclusion, this pilot pharmacokinetic study in lactating

rhesus macaques demonstrates that tenofovir, similar to most other drugs, is found in milk but at lower levels than in maternal blood. The available data suggest that such low tenofovir levels in milk will most likely have no biological effects whatsoever for the nursing infant.

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