REVIEW

Intestinal microbiota regulates the gut‑thyroid axis: the new dawn of improving Hashimoto thyroiditis

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Abstract

Intestinal microbiota plays an indispensable role in the host's innate immune system, which may be related to the occurrence of many autoimmune diseases. Hashimoto thyroiditis (HT) is one of the most common autoimmune diseases, and there is plenty of evidence indicating that HT may be related to genetics and environmental triggers, but the specifc mechanism has not been proven clearly. Signifcantly, the composition and abundance of intestinal microbiota in patients with HT have an obvious diference. This phenomenon led us to think about whether intestinal microbiota can afect the progress of HT through some mechanisms. By summarizing the potential mechanism of intestinal microfora in regulating Hashimoto thyroiditis, this article explores the possibility of improving HT by regulating intestinal microbiota and summarizes relevant biomarkers as therapeutic targets, which provide new ideas for the clinical diagnosis and treatment of Hashimoto thyroiditis.

Keywords Intestinal microbiota · Hashimoto thyroiditis · Gut-thyroid axis · Biomarker · Autoimmune disease

Introduction

Hashimoto thyroiditis (HT) is a kind of chronic autoimmune thyroiditis. According to statistics, the global prevalence of HT has reached 10–12%, with a high incidence of 30–50 years old and female to male predominance of $7-10:1$ [\[1](#page-13-0)-3]. In the past decades, the incidence rate of HT has gradually increased, which has become a global public health problem [[4,](#page-13-2) [5](#page-13-3)]. Furthermore, in areas of iodine sufficiency, HT is the most common cause of hypothyroidism in adults [\[6](#page-13-4)]. In a study on the incidence rate of thyroid diseases among people in regions with low, sufficient or excessive iodine intake in China, the cumulative incidence rate

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of HT was 0.2%, 1%, and 1.3%, respectively [[7\]](#page-13-5). Pathologically, Hashimoto thyroiditis is characterized by lymphoplasmacytic infltration, tissue fbrosis, lymphatic follicular formation, parenchymal atrophy and acidophilic change of follicular cells [[1\]](#page-13-0). The most common clinical manifestation is goiter with or without hypothyroidism. The pressure of the enlarged thyroid on the neck can lead to dysphonia, dyspnea and dysphagia. Due to the loss of thyroid function and primary hypothyroidism, HT patients often have other symptoms involving multiple systems of the body. Gastrointestinal symptoms are always caused by changes in serum thyroid hormone, among which constipation is the most common clinical symptom. The intestinal peristalsis of HT patients is often signifcantly weakened because of decreased thyroid hormone, which even lead to pseudo-obstruction or intestinal obstruction. In addition, the hypotension of gallbladder and changes in bile composition caused by low thyroid hormone will lead to an increase in the formation of bile duct stones [[8\]](#page-13-6). The autoimmune performance of HT is based on the interaction between environmental factors and genetic susceptibility [[9\]](#page-13-7), but its specific pathogenesis has not been clarifed. Because HT usually accompanies patients for life and is difficult to recover, which often seriously affects the normal work and quality of life of patients. At present, there is still a lack of efective treatment methods, so it is urgent to fnd an efective intervention for curing disease.

There are trillions of microorganisms in the human gastrointestinal tract, forming a reciprocal relationship with the host. Intestinal microbiota plays a key role in maintaining the homeostasis of the digestive tract, limiting the colonization of pathogens, and regulating metabolism [[10–](#page-13-8)[12](#page-13-9)]. At the same time, the microbiota is the basis for the correct development of gut-associated lymphoid tissue (GALT) and initiates the immune response of GALT through pattern recognition receptor/pathogen-associated molecular patterns (PRR–PAMP) recognition and epigenetic regulator (such as short-chain fatty acid) [[13](#page-13-10)]. In return, the host provides the living environment and nutrients for the gut microbiota and afects the composition of the microbiota through the immune response induced by the symbiotic flora [\[14,](#page-13-11) [15](#page-13-12)]. Therefore, the imbalance between the host immune system and intestinal microbiota may lead to disease. The research on intestinal microbiota mostly focuses on gastrointestinal diseases. In clinical practice, the treatment based on regulating intestinal microbiota has been applied to infammatory bowel disease [[16\]](#page-13-13), irritable bowel syndrome [\[17](#page-13-14)], and liver cirrhosis [[18\]](#page-13-15), which has become a hot research topic in the world. However, changes in intestinal microbiota are also described in some extraintestinal diseases, such as type 2 diabetes [[19\]](#page-13-16), rheumatoid arthritis [\[20](#page-13-17)], systemic lupus erythematosus [\[21](#page-13-18)], and other autoimmune diseases.

In recent years, with the in-depth study of intestinal microbiota, the correlation between intestinal fora and HT began to receive attention. Both animal models and patients have confrmed that HT can afect the composition and abundance of intestinal microorganisms, leading to dysbiosis of intestinal microbiota [\[22–](#page-13-19)[24](#page-13-20)]. On the one hand, the dysbiosis of intestinal microbiota leads to the damage of the gut barrier, and then bacterial translocation, which destroys the immune tolerance in thyroid autoimmunity through a series of mechanisms, including molecular simulation, bystander activation, and epitope spreading [[12\]](#page-13-9). On the other hand, increasing evidence shows that microbiota regulates the immune response, changes the balance of T cell subsets, afects the absorption of trace elements and thyroid hormone metabolism through the gut-thyroid axis, and finally affects the thyroid function $[25, 26]$ $[25, 26]$ $[25, 26]$ $[25, 26]$. In addition, the bacterial antigen may enhance the activation of infammasomes (like NLRP1, NLRP3, NLRC4, and AIM2) and increase the expression of related cytokines by binding with antibodies in blood [\[27](#page-13-23), [28\]](#page-13-24). In addition to the microbiota, its metabolites, such as short-chain fatty acids and secondary bile acids, also play an important role in the gut-thyroid axis [\[29\]](#page-13-25). In this article, we will discuss the above mechanisms, explore the possibility of improving HT patients by interfering with intestinal microbiota, and summarize the level changes of relevant biomarkers in HT patients to discuss the potential as a new diagnosis and treatment marker, such as zonulin protein, infammatory body, short-chain fatty acid,

secondary bile acid, IL-18, and IL-1. It is helpful to provide theoretical support for the further development and application of microecological therapy in HT.

Interaction between gut microbiota and thyroid gland: gut‑thyroid axis

Dysbiosis in HT

The intestinal microbiota is mainly composed of bacteria, viruses, fungi, prokaryotic communities and other microorganisms, in which aerobic bacteria and anaerobic bacteria are the main bacteria. In the human healthy intestine, the most representative bacterial phyla are Firmicutes and Bacteroides, followed by Proteobacteria, Actinomyces, Fusobacteria and Verruciformes [\[12](#page-13-9), [30,](#page-13-26) [31](#page-14-0)]. Under normal circumstances, the interaction between intestinal microbiota and host maintains a dynamic balance, however, when the human micro-ecosystem exceeds the self-regulation ability, intestinal dysbiosis can lead to the occurrence of disease [\[32](#page-14-1), [33](#page-14-2)]. Studies in diferent regions have compared the diferences in microbiota between HT patients and healthy people, and various data support the occurrence of intestinal dysbiosis in HT (Table [1\)](#page-2-0). In a meta-analysis demonstrating the relationship between intestinal microbiota and autoimmune thyroid disease (AITD), researchers found that benefcial bacteria (such as *Lactobacillus* and *Bifdobacterium*) in the intestines of patients with AITD decreased and harmful microbiota (such as *Bacteroides fragilis*) increased significantly [\[34\]](#page-14-3). Zhao et al. observed that the composition of intestinal microbiota in HT patients were signifcantly diferent from the composition of intestinal microbiota in the healthy people: the relative abundance of *Blautia, Roseburia, Ruminococcus_torques_*group, *Romboutsia*, *Dorea*, *Fusicatenibacter* and *Eubacterium_ hallii_* group genera increased, while the relative abundance of *Fecalibacterium*, *Bacteroides*, *Prevotella_9* and *Lachnoclostridiuum* genera decreased [\[35](#page-14-4)]. Several other studies also confrmed the reduction of *Faecalibacterium*, *Bacteroides*, *Prevotellaceae* and *Lachnoclostridium* [[36](#page-14-5)[–38](#page-14-6)]. In addition, research that the proportion of *Firmicutes/Bacteroidetes* (F/B) in the intestinal microbiota of HT patients increased. Because Firmicutes and Bacteroidetes are the main dominant microbiota at the level of the phyla, their proportion is related to the disease susceptibility, which may refect the ecosystem of the gastrointestinal tract to infer the disease status of HT [\[35](#page-14-4), [39](#page-14-7)]. Ishaq et al. used 16S rRNA gene sequencing to analyze the microbiota in HT, the fndings showed that at the gate level, the abundance of Proteobacteria and Cyanobacteria was higher, the level of Actinobacteria was improved, while the abundance of Firmicutes and Bacteroides was lower; at the level of families and genera, *Enterobacteriaceae*, *Alcaligenaceae* and *Parasutterella* genera increased, while *Prevotellaceae*, *Ruminococcaceae,*

Veillonellaceae and *Dialister* decreased. At the same time, the data from real-time PCR showed that *Bifdobacterium* and *Lactobacillus* were signifcantly decreased in HT patients [\[36](#page-14-5)]. Interestingly, in another research, Liu et al. showed that the intestinal abundance of *Bifdobacterium* in HT patients increased with the development of the disease [[37\]](#page-14-8). Moreover, some study results showed that the species richness index Chao1 of HT patients had a signifcant increase, which may indicate the excessive growth of intestinal bacteria [\[34](#page-14-3), [36](#page-14-5)]. Although high microbial diversity is usually associated with better health outcomes, it also possibly causes destructive efects, such as increased protein breakdown and decreased polyphenol conversion, mucus secretion, and epithelial turnover [\[40\]](#page-14-9). Therefore, although the intestinal microbiota of HT patients has changed, whether it is to help the host fght against or promote disease needs more animal experiment to prove.

Efect of microbiota on the immune system

Microbiota and innate immunity

Commensal bacteria are recognized as the basis for the correct development of GALT. The innate immune cells in GALT, including innate lymphocytes (ILC), macrophages, dendritic cells, and intestinal epithelial cells (IECs), trigger the immune response by non-specifc recognition of pathogens [[42](#page-14-10)[–44](#page-14-11)]. In healthy conditions, the intestinal microbiota forms a symbiotic relationship with the host and interacts with epithelial cells and immune mucosal cells to maintain immune homeostasis through PRR-PAMP recognition [[15,](#page-13-12) [45](#page-14-12), [46\]](#page-14-13). Intestinal microbiota plays a key role in regulating the development of antigen presenting cells (APCs) [[47](#page-14-14)]. Atarashi et al. demonstrated that microbial-derived ATP can stimulate the dendritic cell subpopulation expressing CD70 and CX3CR1 on its surface, which induces the differentiation of Th17 cells [[48](#page-14-15)]. Conventional natural killer (NK) cells, a kind of ILC, can produce IFN- γ , TNF- α and granulocyte–macrophage colony stimulating factor (GM-CSF) to participate in the immune response [[49\]](#page-14-16). Sanos et al. found that GF mice lack NKp46 cells that produce IL-22, indicating that intestinal microflora may play a key role in promoting the diferentiation of IL-22NKp46 cells [\[50](#page-14-17)]. In addition, intestinal microbiota was found to be dependent on the TLR-Myd88 pathway to promote the migration of mast cells to the intestine by inducing CXCR2 ligands from IECs [\[51](#page-14-18), [52](#page-14-19)]. Professional antigen presenting cells (macrophages

and dendritic cells) and ILC can block self-reactive cells and play a crucial role in autoimmune diseases [[53](#page-14-21), [54](#page-14-22)].

Microbiota and adaptive immunity

CD4+T cells are key components of the adaptive immune system. Previous studies have shown that the composition of microbiota affects the balance between the two main effector T cell populations (IL17+Th17 Treg and CD25+Foxp3+Treg) and induces the transformation of Th1 to Th2 [\[55](#page-14-23)[–58](#page-14-24)]. A considerable number of microbial species have been confrmed to participate in intestinal immunity, for example, the segmented flamentous bacterium(SFB) in the intestine can induce Th17 cells in the intestinal lamina propria [\[59](#page-14-25), [60](#page-14-26)]; Polysaccharide A (PSA) from Bacteroides fragilis can induce the expansion of CD4+T cells systemically and reverses the Th1 /Th2 imbalance, and exerts systemic anti-infammatory activities by enhancing the production of IL-10 and by promoting the level and function of IL-10+Foxp3+Treg cells [[61–](#page-14-27)[63\]](#page-14-28). Based on the above fndings, we have reason to assume that changes in the relative abundance of specifc symbiotic microbiota in HT may afect the diferentiation of efector T cells, thus afecting the process of disease. In addition, the components of microbiota may induce B cell activation factors and participate in the induction of IgA-producing B cells. Recently, it has been demonstrated that the immunogenicity of diferent bacterial strains plays a pivotal role in the induction of regulatory B cells, which are crucial in the suppression of infammatory immune response. In diferent germ-free (GF) animal models (mouse, pig) [\[64](#page-14-29), [65\]](#page-14-30), it was observed that the number of macrophages and neutrophils decreased, CD4+ and CD8+ cells decreased, Th1 /Th2 regulation imbalance, Th17 and TREG diferentiation decreased, IgA level decreased and plasma cell number decreased [\[47](#page-14-14)]. The above results show that intestinal microbiota plays an important role in regulating innate and adaptive immunity, and the absence of microbiota will cause serious immune system disorders.

Microbiota and thyroid autoimmunity

Microbiota can participate in the body's autoimmunity through a variety of mechanisms and play an important role in the development of autoantigen tolerance, including molecular simulation, bystander activation, and epitope spreading. Molecular simulation has been proven to be an extremely common mechanism. Through this mechanism, the microbiota can escape immune response and regulate the biosynthesis and metabolic pathway of the host [\[66](#page-14-31)[–68](#page-14-32)]. The study found that some microbiota, such as *Lactobacillus*, *Bifdobacterium* and *Helicobacter pylori*, can induce thyroid autoimmunity by through molecular simulation, because some bacterial proteins have structural homology with human thyroid globulin (hTg)and thyroid peroxidase (TPO), and can selectively bind human TPO and Tg antibodies, and compete with natural antigens to bind autoantibodies [\[69](#page-14-33), [70](#page-14-34)]. Bevenga et al. proved that 16 Borrelia proteins not only have signifcant amino acid sequence homology with thyroid stimulating hormone receptor (TSHR) but also with Tg, TPO, and sodium-iodide symporter (NIS) [[71\]](#page-14-35).

Secondly, sometimes the microbiota may not have antigens similar to the host structure, but it can induce co-stimulation and cytokine production through antigen presenting cells, leading to the activation of bystander T cells [\[72](#page-14-36), [73](#page-14-37)]. Arata et al. showed that hTg-specifc transferred cells had initiated bystander activation of naive host lymphocytes, which was confrmed to exist in HT [[74](#page-14-38)]. They proposed that bystander activation might be achieved through the following ways: by provoking cell death and thus the release of cellular antigens, increasing the visibility or abundance of antigens; by attracting and potentiating antigen presenting cells; or by disturbing the cytokine balance through the infammation that is associated with infection.

Finally, intramolecular epitope spreading is considered to be the mechanism of the expansion of autoimmune response to many sites of protein autoantigen; after the initial reaction to some dominant epitopes, antibodies against multiple secondary or recessive epitopes are produced by intramolecular epitope spreading [\[75](#page-14-39), [76\]](#page-14-40). Thrasyvoulides et al. proved that peptide TgP41 not only includes the epitopes of Grave's Disease (GD)-related autoantibodies, but also the main immunogenic epitopes of experimentally induced Tg specifc antibodies, which can drive the spreading of B cell epitopes [[76\]](#page-14-40). In addition, dysbiosis may produce new epitopes with auto-immunogenicity through post-translational modifcation of proteins, including citrullination and acetylation of active lysine, etc., and then induce autoimmune reactions [[77\]](#page-14-41). In fact, in several autoimmune diseases such as rheumatoid arthritis (RA) and systemic lupus erythematosus (SLE), anti-citrulline protein antibodies have been used as good diagnostic markers in the clinic [[78](#page-15-0)].

The intestinal barrier in HT

Under normal circumstances, the intestinal mucosal barrier can effectively prevent the pathogen from entering the circulatory system. However, when the integrity of the intestinal barrier is damaged, the submucosal immune cells are exposed to bacteria, dietary antigens and autoantigens, which can result in adverse immune activation or tolerance response failure, leading to autoimmune diseases [\[79](#page-15-1)]. Nowadays, there is a growing body of evidence proving that intestinal microbiota plays a crucial role in maintaining the integrity of the intestinal barrier [[80](#page-15-2)]. In extreme cases, studies on germ-free animals confrmed that the lack of intestinal microbiota would lead to the degradation of intestinal mucosal barrier function, the specifc manifestations are the reduction of the overall intestinal surface area, shorter intestinal villi, the reduction of intestinal crypt, the increase in intestinal permeability as well as the thinning and instability of mucous layer [[81,](#page-15-3) [82\]](#page-15-4). A large number of studies show that intestinal dysbiosis, bacterial overgrowth and increased intestinal permeability (intestinal leakage) contribute to the development of HT [\[25,](#page-13-21) [40,](#page-14-9) [83\]](#page-15-5). Demir et al. found that the increased intestinal permeability caused by the change of intestinal microbiota was related to the higher level of zonulin, a protein responsible for regulating the intercellular connection [[84\]](#page-15-6). Zonulin is a paracrine signal protein secreted by small intestinal epithelial cells, a physiological regulator of the tight connection between cells, and is involved in macromolecular transport, integrity of epithelial and endothelial barrier, and immune tolerance in intestinal mucosa $[85]$ $[85]$. Similar to the changes in patients with IBD, patients with HT also have abnormal intestinal barrier permeability. Therefore, it can be considered as one of the major causes of HT that the intestinal barrier function is damaged in the presence of infammation, and the increased intestinal permeability leads to the antigen that could not pass through the intestinal barrier entering the systemic circulation and activating the immune system, and the generated antibody attacking the thyroid tissue [[86](#page-15-8)].

Microbiota and autophagy defect

There is also interaction between intestinal microbiota and autophagy, which is closely related to the pathogenesis of thyroid autoimmune diseases. Autophagy is a highly conservative physiological process. The components in cells undergo lysosome-mediated self-digestion and circulation, and damaged or aged biological macromolecules and organelles are removed from the cytoplasm [[87\]](#page-15-9). On the one hand, intestinal microbiota and its metabolites can control autophagy through mTOR pathway [[88\]](#page-15-10). On the other hand, autophagy affects the composition of intestinal microorganisms. It can be observed that the abundance of proinfammatory bacteria (such as *Candidatus arthromitus*) and pathogenic bacteria (such as *Pasteurellaceae* family) is increased, while the number of anti-infammatory bacteria (such as *Akkermansia muciniphila* and members of the *Lachnospiraceae* family) is decreased in mice with intestinal specific autophagy-related gene 5 (ATG5)deficiency [\[89](#page-15-11)]. Meanwhile, autophagy defect will change the expression level of the tight junction protein claudin-2 in the intestinal mucosa. Diferent from the function of most tight junctions (such as occludin, claudin-1, ZO-1), the expression of claudin-2 protein increases the cell bypass permeability, leading to the destruction of the intestinal epithelial barrier and the increase in bacterial translocation and transmission, and then resulting intestinal dysbiosis [\[90,](#page-15-12) [91](#page-15-13)]. In addition to mediating infammatory reaction and immunity, autophagy is also an important mechanism for clearing excess reactive oxygen species (ROS) to prevent cell damage and death. Autophagy defects lead to the accumulation of depolarized mitochondria and proteins and induce the release of infammatory body activator (ROS or mitochondrial DNA) [\[92,](#page-15-14) [93](#page-15-15)]. As an important pathogenesis, infammatory factors and ROS also participate in the infammatory process of HT. Normally, ROS is essential for thyroid hormone synthesis, but excessive levels of oxidative stress can induce damage to thyroid follicular cells (TFCs) and promote the occurrence of HT by causing thyroid infammation [[94](#page-15-16)]. The elevated serum concentration of IL-23 can be detected in HT patients, which is because IL-23 promotes the development of HT by promoting Th17 cell diferentiation and IL-17 secretion [[95,](#page-15-17) [96\]](#page-15-18). Recent evidence has revealed a new mechanism that IL-23, as a strong inducer of AKT/mTOR signal pathway, can inhibit the autophagy activity ofTFCs, resulting in excessive ROS accumulation [\[97](#page-15-19)]. In addition, Lu et al. also found that Caveolin-1 deficiency inhibited the autophagic activity of TFCs and induced AKT /mTOR activation, which may be involved in the pathogenesis of HT [\[98\]](#page-15-20). In turn, excessive ROS promotes the production of infammatory reactions and proinfammatory cytokines and IL-23. This forms a positive feedback cycle and aggravates the severity of the disease [[99\]](#page-15-21).

Microbiota and metabolites regulate infammation

The intestinal microbiota has been proven to be able to participate in the regulation of body infammation. Microbiota can activate pro-infammatory or anti-infammatory processes, and the imbalance between the two will trigger the thyroid autoimmune process [\[32](#page-14-1)]. The increased intestinal permeability caused by dysbiosis makes the antigen enters the circulation and activates the immune system. At the same time, the antibodies in the blood may react with bacterial antigens and enhance the activation of infammatory bodies in the thyroid [[34](#page-14-3)]. Intestinal microbiota can increase the activation of infammasome through fagellin, type III secretory system (T3SS), lipopolysaccharide (LPS), toxin, etc. Among them, the expression of NLRP3 mainly locates in the TFCs near the lymphoid infltration area, which can recognize bacteria, viruses or other pathogens, and promote the activation of caspase-1 precursor to caspase-1, thereby activating IL-1 β and IL-18 [[100](#page-15-22)]. IL-1β can promote the recruitment and activation of neutrophils and dendritic cells, and diferentiation of Th17 and Tregs. At the same time, IL-1β induces the expression of Fas/FasL and intercellular adhesion molecule-1 on TFCs and interferes with the integrity of thyroid epithelium [[101\]](#page-15-23). IL-18 can activate intestinal epithelial cells to produce antimicrobial peptides (AMPs) and stimulate the production of Th1, NK and NKT to secrete IFN-γ, or combine with IL-12 to induce more IL-17-secreting γδ T cells [\[102](#page-15-24), [103\]](#page-15-25). The abnormal expression or dysfunction of the infammasome is related to autoimmune and organ damage as well as various diseases and has been confrmed in autoimmune diseases, such as multiple sclerosis and lupus nephropathy [\[104](#page-15-26), [105\]](#page-15-27). Guo et al. confrmed the relationship between HT and the expression level of infammatory corpuscles. They found that the signifcantly enhanced expression of NLRP1, NLRP3, NLRC4, AIM2, ASC, caspase-1, pre-IL-1β, pre-IL-18, mRNA and protein in thyroid tissue of HT patients [\[28](#page-13-24)]. Therefore, they proposed the mechanism that infammasome contributes to the development of HT, that is, the activation of infammasome in TFCs induces immune response by mediating cell death and release of bioactive cytokines. It can be seen that intestinal microbiota induces the expression of some infammatory cytokines and chemokines by enhancing the activation of infammasome and then promotes the infammatory reaction to participate in the pathogenesis of HT.

Lipopolysaccharides (LPS) are the main components of the cell wall of Gram negative bacteria, which can refect the level. The binding of LPS to Toll-like receptor 4 (TLR-4) activates a wide range of cellular signaling pathways, such as NF-aκB and MAPK pathways, thereby inducing chronic subclinical infammatory processes and TNF, IL-1 β Expression and secretion of pro-infammatory cytokines such as IL-6. [[106](#page-15-28)]. The latest research proposes the key mechanism of thyroid homeostasis disorder caused by bacterial LPS, including increased expression of Tg and NIS gene, TLR4-κB (NF-kB) pathway and regulating the activities of diferent deiodinase (type I and type II) [\[107](#page-15-29)].

In addition to the microbiota itself, various metabolites produced by the interaction between the host and the microbiota also participate in the regulation of infammatory responses. Short-chain fatty acids (SCFAs) are metabolites secreted by intestinal microbiota during the fermentation of dietary fber, mainly including butyrate, acetate and propionate (the ratio is 60:20:20), which can afect immune regulation and have anti-inflammatory effects [[29\]](#page-13-25). It has shown that SCFAs can inhibit the infammation that induced by LPS through G protein-coupled receptors (GPCRs) and histone deacetylases (HDACs). Free fatty acid receptor 3 (FFAR3) and FFAR2 are the most important GPCR receptors of SCFAs. SCFAs can inhibit IL-6, IL-1β and TNF-α by activating FFAR receptors to exert anti-infammatory effects $[108]$ $[108]$. Butyrate is regard as the most effective inhibitor of HDACs, and it can reduce IFN-γ by inhibiting HDAC, while resulting in increase of naive CD4+T cells and Treg cells [\[109](#page-15-31)], and inhibiting TNF and the activity of NF-κB in monocytes and neutrophils activated by LPS [[110,](#page-15-32) [111](#page-15-33)], and increasing the IL-10 produced by macrophages to regulate T cell activity in other tissues [\[112\]](#page-15-34), and is related to the inhibition of NLRP3 infammasome activation through GPR109A [[113–](#page-15-35)[115](#page-15-36)]. In the other hand, SCFAs can induce intestinal cell diferentiation together with thyroid hormones, which enhances the integrity of the epithelial barrier, and participate in the regulation of the thyroid-gut axis [[116\]](#page-15-37).

Another important substance, bile acid (BA) metabolites, are produced by microbial biotransformation of BA that pass through the enterohepatic circulation, which plays an important role in regulating intestinal infammation by regulating immune cell maturation and cytokine release through farnesoid X receptor (FXR) and G protein-coupled receptor 5 (TGR5) receptors [[117,](#page-15-38) [118\]](#page-15-39). There are studies confirming that BA exerts anti-inflammatory effects by affecting the development of $ROR\gamma + Treg$ cells in the colon of mice [[119](#page-15-40)]. In addition, trimethylamine N-oxide (TMAO), an intestinal microbiota-dependent product, was found to enhance the activation and formation of NLRP3 infammasome, ASC, IL-1b and caspase-1 [[120](#page-15-41)]. Interestingly, microbial metabolism also has a strong impact on cytokine production, which is mainly mediated by the tryptophan metabolite tryptophol that has strong inhibitory efects on the TNF- α response [[121\]](#page-15-42).

Intestinal microbiota afects thyroid function

Trace elements have a prominent impact on the interaction between microbiota and thyroid and participate in the conversion and metabolism of thyroid hormones, thus afecting the function of the thyroid. It is well known that iodine is an essential element for the synthesis of thyroid hormone, and intestinal microbiota has an obvious regulatory efect on iodine, mainly including two aspects: One is to afect the concentration of thyroid hormone by controlling the intake, degradation and hepato-enteral circulation of iodine [[24](#page-13-20)]; and the second is to regulate the expression and activity of NIS through LPS and metabolites to afect the iodine metabolism of the thyroid [[122](#page-16-0)]. If long-term exposure to a high iodine environment may increase the immunogenicity of thyroglobulin and then activate the autoimmune reaction, leading to the destruction of thyroid tissue [[123\]](#page-16-1). For microbiota, the adverse efects of high-dose iodine may be caused by the oxidation of cytoplasmic and membrane components [\[124](#page-16-2)]. Liu et al. found that excessive iodine promotes pyroptosis of thyroid follicular epithelial cells in HT through the ROS-NF-κB-NLRP3 pathway [\[125](#page-16-3)].

The thyroid is the largest selenium (Se) reservoir in the body. Selenium can promote the activity of CD4+CD25+FOXP3+regulatory T cells (Treg), inhibit the secretion of cytokines, prevent the apoptosis of follicular cells, and avoid the production of cytokines in thyroiditis [\[126](#page-16-4)]. Selenium deficiency will also lead to the reduction of hormone and enzyme activity and the decrease of peripheral T3 synthesis [[127](#page-16-5)]. Calomme et al. found that *Lactobacillus* converts sodium selenite in cells into selenocysteine and

selenomethionine, thus promoting the absorption of selenium as organic selenium by the human body, which proves that the metabolism of selenium is afected by microbiota [\[128](#page-16-6)]. In addition, intestinal microbiota can isolate selenium and limit the availability of the host [\[129](#page-16-7)]. The resident microbiota in the colon metabolize selenium, so it will not be absorbed by the upper digestive tract. In turn, selenium afects the composition and colonization of intestinal microbiota. Kasaikina found that selenium changed the diversity of microbiota in mouse models, which indicated that, *Porphyromonadaceae* phylotypes 1 and 3 and *Tannerella* phylotype 2 had increased reaction to selenium, while *Alistipes phylotype* 1 and *Parabacteroides* phylotype 3 declined [[129](#page-16-7)].

Iron (Fe) and zinc (Zn) are also important elements supporting thyroid function. Iron is necessary for the efective use of iodine and the synthesis of thyroid hormone. Iron deficiency will lead to thyroid hormone impaired synthesis, storage, and secretion [[130](#page-16-8)]. On the one hand, iron affects the composition of microorganisms. In the mice model, an iron-supplemented diet changed microbial diversity and increased the abundance of *Clostridiales* and *Lactobacillales* [[131\]](#page-16-9). Because siderophores of bacteria (mainly *Enterobactin*) have a high affinity for iron, it provides a heme-rich condition for the growth of pathogenic bacteria [[132\]](#page-16-10). On the other hand, by producing SCFAs to reduce the pH, the microbiota can improve iron bioavailability in the colon [\[133\]](#page-16-11). Iron supplementation increased *Enterobacteriaceae* and *Bacteroides* and decreased *Lactobacillus* and *Bifdobacterium*, with the decrease in butyrate and propionate [\[134,](#page-16-12) [135](#page-16-13)]. This change is explained by the role of infammation in promoting the microbiota [\[40](#page-14-9)].

In addition, the microbiota can afect the level of zinc in the body, and the availability of zinc in turn determines the status of intestinal fora. In broiler chickens with chronic zinc deficiency, it was observed that the intestinal population of *Proteobacteria* was signifcantly increased, and the number of Firmicutes correspondingly decreased [\[136](#page-16-14)]. Zinc deficiency can affect the synthesis of thyrotropin releasing hormone (TRH), and the levels of thyrotropin (TSH), T3 and T4 will decrease [[137\]](#page-16-15). In return, hypothyroidism can also lead to zinc deficiency.

At present, the role of microbiota in iodothyronine metabolism has been revealed. The microbiota can uncouple the sulfated glucuronide derivatives of iodothyronine via bacterial sulfate esterase or β-glucuronidase to improve the reab-sorption of thyroid hormone in enterohepatic circulation [[26,](#page-13-22) [138](#page-16-16)]. On the other hand, inhibition of 5-deiodinase activity by microbiota reduces the transformation of T4 to T3 and rT3 [\[139](#page-16-17)]. A study based on the rats model found that intestinal bacteria can absorb unbound iodothyronine and even compete with albumin to bind thyroid hormone [[25,](#page-13-21) [139\]](#page-16-17). It is worth mentioning that some microbiota also plays a role in stabilizing thyroid hormone fuctuations and improving the utilization of levothyroxine. For example, Escherichia coli can act as a reservoir of T3 by combining T3 with bacterial thyroid-binding protein [[40](#page-14-9)]; *Lactobacilli* and *Bifdobacteriaceae* have a signifcantly lower adjustment requirement of T4, indicating that the supplement of microbiota has increased levothyroxine availability and stabilizes thyroid function [[140\]](#page-16-18).

Efect of thyroid on the gastrointestinal tract

Previous studies have confrmed the role of thyroid function in the gastrointestinal system. T3 is considered to be the most important regulator of the development and differentiation of intestinal epithelial cells [[12](#page-13-9)], and the lack of T3 is often an important reason for hypothyroidism. The variation in blood concentration of thyroid hormone causes a change in gastrointestinal neuromotor function, leading to the occurrence of gastrointestinal dysfunction, which can be clinically manifested as constipation and intestinal obstruction [[141\]](#page-16-19). Diarrhea in hypothyroidism is mainly due to the increase in bacterial growth caused by intestinal hypomotility, and excessive bacteria contribute to damage to gastrointestinal neuromuscular function [[142\]](#page-16-20). At the same time, severe hypothyroidism may lead to esophageal peristalsis. When the proximal end is involved, mucous edema causes difficulty in swallowing, while the distal end may have esophagitis and hiatal hernia [\[141\]](#page-16-19). A gastric myoelectric study by Gunsar et al. showed a positive correlation between dyspepsia and hypothyroidism score [[143](#page-16-21)]. Due to muscle edema and decreased myoelectric rhythm, patients with hypothyroidism often have gastric motility disorders, resulting in delayed gastric emptying and gastric acid defciency. In particular cases, hypothyroidism may also be responsible for refractory gastrointestinal bleeding in routine treatment [[144\]](#page-16-22). On the other hand, thyroid function will also affect intestinal microbiota. Studies by Lauritano et al. showed that patients with signifcantly decreased thyroid function are more likely to have intestinal bacterial overgrowth [[142\]](#page-16-20) (Fig. [1\)](#page-7-0).

The relationship between HT and related digestive diseases

The relationship between gastrointestinal tract and thyroid was proposed as early as the 1950s and was called "thyrogastric syndrome." This consistency can be explained by the origin of the common germ layer and the specifc composition of the microbiota. In recent years, the common mechanism between Hashimoto's thyroiditis and digestive tract diseases has attracted attention [[40\]](#page-14-9).

Fig. 1 In steady-state conditions, microorganisms and their metabolites or food source antigen are the basis of intestinal-related lymphoid tissue (GALT) correct development, through pattern recognition receptor/pathogen-related molecular pattern (PRR–PAMP) recognition start intestinal immunity, promote the dendritic cells, T cells, B cells, innate lymphocyte diferentiation or maturation. However, when the intestinal barrier integrity is destroyed and the intes-

HT and celiac disease (CD)

Celiac disease is a small intestinal infammatory disease with autoimmune feature, which is triggered and maintained by the ingestion of the storage proteins (gluten) of wheat, barley, and rye. Because the molecular structure of glutenin is similar to that of thyroid tissue, it can lead to multiple autoimmune thyroid diseases. A singlecenter retrospective case–control study by Bibb ò et al. showed that HT is the most common autoimmune disease in patients with celiac disease $[145]$ $[145]$ $[145]$. In addition, their symptoms often overlap, so we have reason to believe that they may be connected through the gut-thyroid axis. Non-celiac wheat sensitivity (NCWS) is a non-allergic and non-autoimmune disease. It can also increase the expression and activation of TNF- α through toll-like receptors (such as TLR2 and TLR4) $[146]$ $[146]$ $[146]$. According to the gutthyroid axis in HT, the pathogenesis of CD and NCWS can be explained from the following aspects: shared cytokine, molecular simulation, abnormal post-translational modifcation of proteins, malabsorption of essential

tinal mucosal permeability is increased, the intestinal fora and its metabolism can promote caspase-1 precursor activation to caspase-1 through the NLRP 3 inflammasome, and then activate IL-1 β and IL-18, and can also control autophagy through the mTOR pathway. When the intestinal infammatory cascade expands and the infammatory products enter the circulatory system, it may cause the occurrence and development of Hashimoto's thyroiditis

micronutrients for the thyroid and damaged intestinal barrier [[25](#page-13-21)].

HT and Hp infection gastritis

In recent years, massive studies have already demonstrated that *Helicobacter pylori* (Hp) infection is closely related to the incidence of peptic ulcer and gastric cancer. During the past 2 decades, the correlation between Hp and other nongastrointestinal diseases has been revealed. The relationship between thyroid autoimmunity and Hp can be explained by molecular simulation. Cellini M et al. described three mechanisms that ultimately activate Th1 autoreactive lymphocytes to link Hp and HT. Firstly, CD4+T cells recognize Hp epitopes with a similar structure to H/K/ATPase on the thyroid and activate Th1 to induce apoptosis. Secondly, dendritic cells present Hp epitopes to naive T cells, with a lack of peripheral immune tolerance, the Th1 will be activated. Finally, INF-γ can stimulate MHCII expression in follicular thyroid cells [[147](#page-16-25), [148\]](#page-16-26). In addition, Hp can produce a cytotoxin-associated gene A (CagA). It has been found that

the cag-A positive Hp strains show a nucleotide sequence similar to the thyroid peroxidase (TPO) sequence, illustrating that serum CagA positive increases the risk of autoim-mune thyroid disease [\[149\]](#page-16-27).

HT and autoimmune atrophic gastritis(AAG)

HT is found in nearly 40% of patients with autoimmune atrophic gastritis (AAG), who have a large number of antiparietal cell antibodies in the serum [\[150](#page-16-28)]. Because gastric acid secretion is greatly reduced in AAG, iron absorption is too poor to promote the synthesis of T3 and T4. In addition, it is found that the levels of serum gastrin, chromogranin A and the proliferation of enterochromafn-like cells (ECL) were signifcantly related to the coexistence of autoimmune diseases [[151](#page-16-29)].

The above fndings provide theoretical support from multiple perspectives. Therefore, we can further understand the role and mechanism of gut-thyroid by exploring the relationship between HT and other gastrointestinal diseases, and then broaden the clinical thoughts of HT and its complications, develop new therapies and drugs, and fnally further improve the diagnosis and treatment level of HT.

Potential therapeutic strategies by regulating the gut‑thyroid axis

Although L-thyroxine (LT4) is currently a common treatment option for HT, it can only alleviate the lack of thyroxine, but cannot completely cure HT. Therefore, it is necessary to fnd an efective treatment by exploring the mechanism. To sum up, intestinal microbiota can efectively regulate the process of HT by increasing intestinal permeability, molecular simulation, bystander activation, epitope spreading, activation of infammatory bodies, autophagy defects, trace elements absorption, thyroid hormone metabolism, and so on. At the same time, the classifcation and metabolic pathway level of intestinal microbiota refect the interaction between intestinal microbiota and thyroid hormone, which allows us to give more consideration to traditional drug treatment for HT patients. Therefore, based on the relevant mechanism of the gut-thyroid axis, we summarized the following treatment strategies as auxiliary interventions.

Intestinal non‑absorbable antibiotics

For small intestinal bacterial overgrowth (SIBO) in HT patients, the treatment goal is to alleviate symptoms by eradicating bacterial overgrowth [[152\]](#page-16-30). Antibiotic treatment is the basis for treating SIBO. Appropriate antibiotic treatment can improve the symptoms of gastrointestinal neuromuscular function damage by reducing the gastrointestinal bacterial burden, and then changing the composition of microbiota to regulate intestinal permeability. A meta-analysis of 10 prospective clinical studies on the treatment of SIBO patients with non-systemic antibiotics showed that the normalization rate of breath test in the antibiotic group was higher than that in the placebo group, indicating the efectiveness of antibiotics in regulating the fora [\[153](#page-16-31)]. Rifaximin is the frst non-aminoglycoside intestinal antibiotic, which acts locally in the intestine and is not absorbable after oral administration, and is characterized by a broad antibacterial spectrum and strong antibacterial activity. The research results of Lauritano et al. showed that the overall incidence of adverse events of rifaximin was signifcantly lower than that of metronidazole group, showing that the side efects of non-absorbable antibiotics were less [\[154\]](#page-16-32). The results of a drug resistance study showed that the concentration of stool Staphylococcus spp and stool Coliforms would be temporarily reduced during the treatment of rifaximin [[155](#page-16-33)]. Xu et al. showed that rifaximin changed the bacterial population in the ileum of rats, increasing the relative abundance of Lactobacillus, and preventing mucosal infammation and impaired intestinal barrier [\[156](#page-16-34)]. Maccaferri et al. found that rifaximin did not afect the overall composition of intestinal microflora, but caused an increased concentration of Bifidobacterium, Atopobium and Faecalibacterium prausnitzii, accompanied by increased short-chain fatty acids [[157](#page-16-35)]. There is evidence to support that a higher dose of rifaximin $(1200-1600 \text{ mg/d})$ is an effective treatment method, which can achieve SIBO purifcation without increasing the inci-dence of side effects [[158](#page-16-36), [159](#page-16-37)].

Rebuild intestinal ecology

Because antibiotics can only remove bacteria but cannot restore normal fora, antibiotic treatment alone cannot completely solve the microbial ecological imbalance related to SIBO. Therefore, it is necessary to think about how to rebuild intestinal fora, and the application of probiotics has become an attractive choice. Generally, we defne some microbiota that is benefcial to the stability of microbiota as probiotics, which can enhance the intestinal barrier function, reduce infammatory reaction and stabilize the intestinal microbiota. Common probiotics include *Lactobacillus*, *Bifdobacterium*, *Lactococcus*, *Streptococcus* and *Enterococcus*. As mentioned above, *Lactobacillus* and *Bifdobacterium* can participate in the occurrence of HT through molecular simulation. However, *Lactobacillus* and *Bifdobacterium*, as probiotics, regulate the immune response, showing antiinflammatory effects and protecting the body from pathogens [\[34](#page-14-3)]. Studies have shown that *Lactobacillus* protects TH17 cells and supports barrier integrity by secreting IL-22 and IL-17. A study on *Lactobacillus* reuteri found that the beneficial effect on the mouse thyroid was achieved by

promoting the production of IL-10 and the enhancement of Treg cells [[12](#page-13-9)]. In addition, in the context of intestinal barrier dysfunction and infammation, the yeast *S. boulardii* has also been widely studied. Its beneficial effects are mainly through antibacterial and antitoxic activities and nutritional effects on intestinal mucosa $[160]$ $[160]$. In recent years, the role of *Akkermansia muciniphila* on the intestinal barrier has also been revealed, which improves the function of intestinal barrier by restoring the thickness of mucous layer and tight junction protein as well as producing specifc antibacterial and bioactive lipids with anti-infammatory properties [[161](#page-16-39)]. A study on the treatment of Grave's disease with probiotics supplemented with methimazole showed that *Bifdobacterium longum* supplemented with methimazole could improve thyroid function and reduce the concentration of TRAb in patients with GD [\[162\]](#page-16-40). In another double-blind randomized placebo-controlled trial, the probiotic compound LAB4 showed an obvious immunomodulatory effect [\[163\]](#page-16-41). It can be seen from this that it is feasible to improve HT through probiotics controlling microbiota in future. In addition to its efect on the intestinal barrier and immunity, probiotics can also reduce the fuctuation of thyroid hormone. Spaggiari et al. found that compared with the control group, the regulatory demand of T4 in the experimental group was signifcantly reduced, manifesting that the mixture of *Lactobacillus* and *Bifdobacterium* (VSL#3) increased the utilization of levothyroxine and stabilized thyroid function. However, the results of this study also show that the intake of probiotics of this genus will not change the susceptibility of HT patients or improve hypothyroidism [[140\]](#page-16-18).

Prebiotics are a kind of non-digestible food ingredient that can improve host health by selectively stimulating the growth and/or activity of intestinal benefcial bacteria [\[164](#page-16-42)]. The carbohydrate hydrolase of the microbiota promotes the fermentation of prebiotics to produce hydrogen, methane, carbon dioxide, SCFA, and other products. Currently available prebiotics include human milk oligosaccharides (HMOs), lactulose and inulin derivatives. Talebi et al. showed that supplementation of synbiotics (a combination of probiotics and prebiotics) can relieve constipation in patients with hypothyroidism and is benefcial to thyroid function, but no efect on thyroid peroxidase has been observed [[165\]](#page-16-43). Based on the fact that the benefcial efects of microbiota are mediated by the secretion of various metabolites, the concept of postbiotics is proposed. According to Tsilingiri et al., postbiotics include any substances released by or produced by the metabolic activities of microorganisms, which directly or indirectly have beneficial effects on the host [\[166\]](#page-16-44). At present, the available postbiotic drugs include cell-free supernatant, exopolysaccharides, enzymes, cell wall fragments, short-chain fatty acids, bacterial lysates, vitamins, phenol-derived metabolites and aromatic amino acids. In the existing studies, epigenetic elements show the characteristics of immune regulation, anti-infammation, anti-oxidation and anti-cancer [\[167](#page-17-0)]. These potential mechanisms suggest possible strategies for epigenetic elements to regulate the fora of HT patients. Although probiotics, prebiotics and postbiotics can play a certain role in the prevention and treatment of autoimmune diseases, considering that most probiotics research depends on animal models, the exact relationship between probiotics and HT still needs to be further explored to provide a new way for clinical regulation of intestinal dysbiosis or thyroid function in HT patients.

Fecal microflora transplantation (FMT) is direct evidence of the interaction between intestinal microfora and various diseases. At present, the specifc mechanism of FMT is not clear, but it can be speculated that the potential mechanism is the improvement of microbial diversity, donor phage regulation of the recipient fora and altered microbial metabolite production [[168\]](#page-17-1). In patients with recurrent Clostridium dif-ficile infection, FMT has achieved amazing results [[169](#page-17-2)]. In 2020, China released the Chinese experts consensus on standardized methodology and clinical application of fecal microbiota transplantation, and FMT was included in the guidelines as treatment of recurrent or refractory Clostridium difficile infection $[169]$. Furthermore, this consensus pointed out that FMT would be gradually applied to the treatment of autoimmune diseases. In future, the research on the prevention and treatment of HT by FMT may have important clinical signifcance.

Signifcantly, due to individual diferences, the diferent sensitivity of intestinal microbiota to biological intervention may lead to inconsistency in results. Therefore, the key to the individualized treatment of ecological therapy in future is to explore the premise of efective probiotics implantation and understand the degree of inter-individual variation of the sensitivity of microbiota to probiotics intervention.

Dietary therapy and trace element intake

The regulation of diet on microbiota directly afects the characteristics of infammation. Because of the rapid and repeatable response of microbiota to dietary intervention, the rational design of personalized diet has become an important microbiota regulation method [\[32\]](#page-14-1). A study of mice showed that after the transformation from a low-fat and high-fber diet to a "Western-style diet" (WDs) with high sugar, high fat and low fber, they had serious ecological disorders, resulting in loss of secretory IgA function, inhibition of Treg cells producing IL-10, damage of intestinal barrier, and immune imbalance [\[170](#page-17-3)]. In addition, mice on a Western diet showed increased Firmicutes phylum and decreased Bacteroidetes phylum. Similarly, the regulation of diet on microbial ecology has also been verifed in human model research [\[171](#page-17-4)]. Diferent food types afect the production of microbial metabolism. Fiber, polyphenols, tryptophan and glucosinolates (vegetarian) increase SCFA, which is benefcial to Bifdobacterium population, while carnitine, choline and fat (a diet rich in animal protein) increase the production of secondary bile acids [[40\]](#page-14-9).

In addition, metabonomics research and germ-free mouse experiments showed that intestinal microbiota played a key role in the production of TMAO [[100\]](#page-15-22). Symbiotic bacteria metabolize dietary lipid phosphatidylcholine and red meat component L-carnitine, which leads to the accumulation of TMAO, suggesting that our targeted inhibition of this reaction can reverse the accumulation of TMAO and improve the development of disease [\[172,](#page-17-5) [173\]](#page-17-6). The above results show that dietary fber plays a crucial role in improving intestinal ecology. During the fermentation of dietary fber, intestinal microbiota can produce SCFAs or metabolites with anti-infammatory properties and maintain intestinal homeostasis. Therefore, it is suggested that the diet structure of HT patients should be adjusted accordingly: (1) Autoimmune Protocol (AIP) diet. Its basic principle is to avoid foods, additives, or medications (e.g., nonsteroidal anti-infammatory drugs) that can trigger intestinal infammation, dysbiosis, and/or symptomatic food intolerance. And it emphasizes eating more fresh and nutritious foods, bone soup and fermented foods, including a 6-week elimination stage and a 5-week maintenance stage [[174\]](#page-17-7). A pilot study showed that AIP diet and lifestyle plan can reduce infammation and regulate the immune system, signifcantly improving the health-related quality of life and symptom in middle-aged female HT patients [[175\]](#page-17-8). (2) Anti-infammatory diet. Focus on foods with high nutrition, such as whole grains with high fber, vegetables rich in polyphenols and foods rich in omega-3 fatty acids. According to the current research results, the antioxidant properties of the Mediterranean diet may be the most benefcial for HT patients [[176\]](#page-17-9). (3) A gluten-free diet. The study found that a gluten-free diet can improve the symptoms of malabsorption and hypothyroidism, reduce the demand for levothyroxine, reduce intestinal infammation, and have a positive impact on the development and performance of autoimmunity [[177,](#page-17-10) [178](#page-17-11)]. However, this diet is very strict and difficult to follow, which leads to the risk of nutritional deficiency. The current research has not confrmed that patients with HT should adopt a gluten-free diet, so it is not recommended [[179\]](#page-17-12).

Because iodine, selenium, zinc, vitamin D, etc., play a part in thyroid function through diferent ways of the gutthyroid axis, excessive iodine intake, selenium defciency and the use of specific drugs (such as amiodarone) are closely related to the onset of HT.

When probiotics are combined with diferent trace elements, they may have a synergistic efect on the host. Lactobacillus and Bifdobacterium seem to have a negative correlation with dietary iron, while a positive correlation with selenium and zinc [[25](#page-13-21)]. In HT, the above two kinds of bacteria decrease, suggesting that intestinal fora may afect the intake of necessary trace elements for the synthesis of thyroid hormone. Therefore, the intake of trace elements can be reasonably adjusted according to the change of microbiota. For example, selenium supplementation can stimulate the immune system, inhibit the expression of human leucocyte antigen-DR (HLA-DR) in thyroid cells and reduce the thyroid autoimmune function [[123\]](#page-16-1). In addition, vitamin D defciency is one of the reasons for HT, and the more vitamin D defciency, the greater the possibility of HT. Thomas et al. found that the diversity of microbiota is closely related to active vitamin D [\[180\]](#page-17-13). Studies have shown that 1α -25(OH)2D3, the active form of vitamin D, can balance the redox system and regulate immune tolerance, so vitamin D supplementation may alleviate the disease activity of HT. A double-blind randomized controlled trial found that the levels of TGAb and TSH in the vitamin D group were signifcantly lower than those in the placebo group, but the levels of TPOAb were not signifcantly lower [\[181](#page-17-14)]. Considering the existing research and the low cost and minimal side efects of vitamin D, it may be recommended to monitor and supplement patients with HT.

Biomarkers provide new targets

Clinically, serum T3, T4, TSH and anti-thyroid antibody are common biological indicators for diagnosing HT. Recently, with the in-depth study of the mechanism of the gut-thyroid axis, more and more new biomarkers have been found. We summarized some important molecules mentioned above, and hope to provide new therapeutic targets for improving Hashimoto's thyroiditis by detecting these molecules (Table [2\)](#page-11-0).

The increase in plasma zonulin levels in HT patients suggests that zonulin may play a role in the pathogenesis of HT as a marker of intestinal permeability damage. Zonulin is the only physiological medium known to reversibly regulate intestinal permeability (IP) [[182\]](#page-17-15). Under normal conditions, zonula occluden 1 (ZO-1) forms tight junction (TJs) complexes with the help of actin flament binding to form selective permeability of intestinal barrier [\[84](#page-15-6)]. In the presence of zonulin, activated epidermal growth factor receptor (EGFR) and protease-activated receptor 2 (PAR2) trigger the activation of phospholipase C, and then, protease-activated receptor α (PKC- α) is activated. Then the activated PKC- α triggers the release of ZO-1 and actin flament, which results in the loose conformation of TJs, increased barrier permeability and excessive abnormal antigenic transportation [[84,](#page-15-6) [183](#page-17-16)]. In addition, intestinal fatty acid binding protein (I-FABP) and diamine oxidase (DAO) were also used to evaluate the integrity of the intestinal barrier and were verifed. They are cytoplasmic proteins in intestinal epithelial

cells, which are released into the circulation system immediately when the intestinal epithelium is destroyed. A study on Grave's disease pointed out that the increased levels of I-FABP and zonulin can damage intestinal epithelial cells through direct contact, toxin release and innate immune activation, which is probably due to the imbalance of microbiota. Although this study did not fnd the level diference of DAO, it inspired us to the biomarkers related to intestinal leakage [[184\]](#page-17-17).

Tschopp et al. frst proposed the concept and composition of infammasomes: (1) pattern-recognition receptors (PRR) or NOD-like receptors (NLR) or AIM2-like receptors (ALR); (2) apoptosis-associated speck-like proteins containing caspase recruitment domains (ASCs); and (3) caspase proteases [\[185](#page-17-18)]. As mentioned above, TFCs express toll-like receptors, which induce the activation of the innate immune system by responding to the PAMPs and damageassociated molecular patterns (DAMPs) on the surface of microorganisms, making lymphocytes product multiple proinflammatory cytokines, such as IL-1β, IL-18, IFN- $γ$, and TNF- α [\[186](#page-17-19)]. The pro-inflammatory cytokines secreted by infltrating lymphocytes lead to an increase in the expression of infammasomes in thyroid cells and enhance the activation of infammasomes induced by DAMP [\[187\]](#page-17-20). The two

mechanisms form a feedback loop to prolong the immune response of thyroid tissue. As one of the most important members of the infammasome family, NLRP3 widely exists in epithelial cells and immune cells. Many microorganisms and their metabolites can regulate the expression of NLRP3. For example, Bacillus fragilis can activate the expression of NLRP3, and Hp can promote the activation of NLRP3 and caspase-1 and the secretion of IL-1b through TLR4, MyD88 and NF-kB [[188\]](#page-17-21). It is worth noting that the subsequent effector molecule IL-1b downstream of the inflammasomes can regulate the microbiota by regulating the production of AMPs [[189\]](#page-17-22). Guo et al. found that the mRNA levels of NLRP1 and ASC in thyroid tissue of AITD group were correlated with the levels of TPOAb and TgAb in serum, indicating that NLRP1/ASC may be a potential biomarker of AITD [[28\]](#page-13-24).

SCFAs are metabolites of dietary fber fermented by symbiotic bacteria, and play a key role in the development, function and regulation of the immune system. SCFA, especially butyrate, can inhibit the production of pro-infammatory cytokines induced by NF-кB in myeloid cells and promotes the production of regulatory T cells by inhibiting pro-infammatory HDAC [\[189](#page-17-22), [190\]](#page-17-23). Butyrate and propionate are HDAC inhibitors that epigenetically regulate

gene expression. In addition to immunomodulatory and anti-infammatory efects, butyrate is also the main energy source of intestinal epithelial cells, which can induce mucus production and strengthen the tight connection between cells together with thyroid hormone, playing an important role in maintaining the intestinal barrier [[40\]](#page-14-9). Small-scale studies comparing HT and healthy human individuals found that the number of bacteria producing SCFA in HT decreased [\[35,](#page-14-4) [36](#page-14-5)], while the fecal and plasma levels of SCFA remained to be studied. Anaerobic Bacteroides, Bifdobacteria, Eubacteria, Streptococcus and lactobacillus are the main bacteria that produce SCFAs. Although the change in microbiota is diverse in species, it is more consistent in function. Therefore, as a product related to bacterial fermentation pathways, SCFA may be better used for disease monitoring and biomarker discovery.

Lipopolysaccharide (LPS) and D-lactic acid can be used as sensitive indicators of the translocation of intestinal bacteria and their metabolites. High levels of LPS and D-lactic acid indicate increased translocation. The efect of LSP on thyroid homeostasis helps to explain the pathogenesis of HT: LPS can directly affect thyroid cells by increasing the expression of thyroglobulin (Tg) and NIS genes [\[122](#page-16-0)]; secondly, LPS regulates thyroid cell function through TLR4- NF-kB pathway on thyroid cells [[191](#page-17-26)]. In addition, as an endotoxin, LPS can inhibit the activity of hepatic type I iodothyronine deiodinase (D1) and conversely activates type II iodothyronine deiodinase (D2) in the hypothalamus and anterior pituitary, thus promoting the conversion of thyroxine (T4) to triiodothyronine (T3) [[192](#page-17-30), [193\]](#page-17-27). In addition, there is evidence that deoxycholic acid (DCA) may also become a marker of the gut-thyroid axis in HT. Primary bile acids (BAs) derived by Cholesterol are regulated by the nuclear FXR and the TGR5 and convert into secondary bile acids through 7α-dehydroxylation reaction, which includes deoxycholic acid and lithocholic acid [[194,](#page-17-24) [195\]](#page-17-25). Many dominant genera in human intestines can produce secondary bile acids, of which Clostridium is the most active. Deoxycholic acid is believed to decrease bacterial overgrowth by inducing membrane damage [[29](#page-13-25)]. According to the metabolic spectrum, the main bile acid of HT patients is DCA, which prompts the possible relationship between DCA and the overgrowth of intestinal bacteria in HT patients [[196\]](#page-17-31).

Microbial post-translation modification of protein (PTMP) is of great signifcance in inducing autoimmune diseases. Dysbiosis may contribute to autoimmune diseases through inappropriate PTMP. The modifcations in the intestine include de/phosphorylation, de/acetylation, ubiquitination, methylation, citrullination, carbamylation, etc. Their enzymatic apparatus is capable to transform naïve/self or non-self-peptides to autoimmunogenic ones [[77\]](#page-14-41). Based on an epigenetics study, Lu et al. showed that the increased protein levels of the trimethylation of histone H3 lysine 4 (H3K4me3) and Mixed lineage leukemia 1 (MLL1) in HT patients, which can be explained by the abnormal H3K4 trimethylation in lymphocytes and monocytes of thyroid tissue [[197](#page-17-28)]. In addition, another study found that LPS stimulation could enhance H3K4me3 in macrophages [\[198\]](#page-17-29). CagA is a marker for the cag pathogenicity island, including the genes necessary for infammation enhancement caused by pathogenic strains. A meta-analysis showed that the overall positive rate of cagA in patients with AITD was signifcantly higher than that in the control group, with significant differences [[149](#page-16-27)]. Because the cag-A positive Hp strain showed a nucleotide sequence similar to the TPO sequence, there may be a cross-reaction between the antibody produced by Hp infection and the thyroid antigen, showing the signifcant correlation between cagA positive and AITD [\[147,](#page-16-25) [199\]](#page-17-32).

Discussion

Intestinal microbiota and thyroid interact with each other through the gut-thyroid axis and play a key role in the pathogenesis of HT. The excessive growth of small intestinal bacteria leads to dysbiosis, leading to intestinal barrier damage, bacterial antigen and its metabolites migrating to the outside of the intestine, which can increase the activation of inflammatory bodies and regulate inflammatory response. Cross-reaction, autophagy defect and functional imbalance of T lymphocyte subsets caused by intestinal dysbiosis directly lead to thyroid tissue damage. Trace elements afect the metabolism of thyroid hormone to participate in the development of HT through the role of intestinal microbiota. Although HT is closely related to intestinal dysbiosis, because of the numerous types and individual diferences of intestinal microfora, it is necessary to deeply understand the structure and mechanism of intestinal microfora to provide more possibilities for accurate medical treatment of HT. In future, the treatment strategy of HT will be around the gutthyroid axis gradually, aiming to slow down the progress of HT and improve the symptoms of patients by rebuilding the intestinal microbial ecology. Biomarkers can not only provide the basis for diagnosis but also guide the selection of intervention strategies. The occurrence of intestinal leakage suggests that we can intervene in the early stage of the disease, to restore the intestinal tight junction, repair the intestinal barrier, and prevent the intestinal immune cells and infammatory factors from migrating out of the intestine. The restoration of intestinal homeostasis can select diferent antibiotics according to the changes in the microbial composition of HT patients and combine probiotics and other biological agents to eliminate pathogenic bacteria while restoring normal fora, which maintains an ingenious balance. In addition, the restoration of intestinal microbial ecology can be achieved through dietary changes, such as the intake of high-fber and low-fat foods, which will promote the transformation of microbial metabolism to anti-infammatory. However, since most studies on microbiota are based on animal models, it is necessary to make full use of clinical big data and more in vivo studies to verify the efectiveness of the above treatment schemes in future.

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References

- 1. Ralli M, et al. Hashimoto's thyroiditis: an update on pathogenic mechanisms, diagnostic protocols, therapeutic strategies, and potential malignant transformation. Autoimmun Rev. 2020;19(10):102649.
- 2. Klubo-Gwiezdzinska J, Wartofsky L. Hashimoto thyroiditis: an evidence-based guide to etiology, diagnosis and treatment. Pol Arch Intern Med. 2022;132(3):16222.
- 3. McLeod DSA, Cooper DS. The incidence and prevalence of thyroid autoimmunity. Endocrine. 2012;42(2):252–65.
- 4. Ott J, et al. The incidence of lymphocytic thyroid infltration and Hashimoto's thyroiditis increased in patients operated for benign goiter over a 31-year period. Virchows Archiv Int J Pathol. 2011;459(3):277–81.
- 5. Ragusa F, et al. Hashimotos' thyroiditis: epidemiology, pathogenesis, clinic and therapy. Best Pract Res Clin Endocrinol Metab. 2019;33(6):101367.
- 6. Chaker L, et al. Hypothyroidism. Lancet (London, England). 2017;390(10101):1550–62.
- 7. Teng W, et al. Efect of iodine intake on thyroid diseases in China. N Engl J Med. 2006;354(26):2783–93.
- 8. Caturegli P, De Remigis A, Rose NR. Hashimoto thyroiditis: clinical and diagnostic criteria. Autoimmun Rev. 2014;13(4–5):391–7.
- 9. Tomer Y. Mechanisms of autoimmune thyroid diseases: from genetics to epigenetics. Annu Rev Pathol. 2014;9:147–56.
- 10. Pickard JM, et al. Gut microbiota: Role in pathogen colonization, immune responses, and infammatory disease. Immunol Rev. 2017;279(1):70–89.
- 11. de Vos WM, et al. Gut microbiome and health: mechanistic insights. Gut. 2022;71(5):1020–32.
- 12. Virili C, et al. Gut microbiota and Hashimoto's thyroiditis. Rev Endocr Metab Disord. 2018;19(4):293–300.
- 13. Suzuki K, et al. GALT: organization and dynamics leading to IgA synthesis. Adv Immunol. 2010;107:153–85.
- 14. Hooper LV, Macpherson AJ. Immune adaptations that maintain homeostasis with the intestinal microbiota. Nat Rev Immunol. 2010;10(3):159–69.
- 15. Kamada N, et al. Role of the gut microbiota in immunity and infammatory disease. Nat Rev Immunol. 2013;13(5):321–35.
- 16. Mallon P, et al. Probiotics for induction of remission in ulcerative colitis. In: The Cochrane database of systematic reviews, no. 4; 2007. p. CD005573.
- 17. Simon E, et al. Probiotics, prebiotics, and synbiotics: implications and benefcial efects against irritable bowel syndrome. Nutrients. 2021;13(6):2112.
- 18. Liu Y, et al. Probiotic Lactobacillus rhamnosus GG prevents liver fbrosis through inhibiting hepatic bile acid synthesis and enhancing bile acid excretion in mice. Hepatology (Baltimore, MD). 2020;71(6):2050–66.
- 19. Gurung M, et al. Role of gut microbiota in type 2 diabetes pathophysiology. EBioMedicine. 2020;51:102590.
- 20. Zaiss MM, et al. The gut-joint axis in rheumatoid arthritis. Nat Rev Rheumatol. 2021;17(4):224–37.
- 21. Choi S-C, et al. Gut microbiota dysbiosis and altered tryptophan catabolism contribute to autoimmunity in lupus-susceptible mice. Sci Transl Med. 2020;12(551):eaax2220.
- 22. Shin NR, et al. Chemically or surgically induced thyroid dysfunction altered gut microbiota in rat models. FASEB J. 2020;34(6):8686–701.
- 23. Su X, et al. Gut dysbiosis is associated with primary hypothyroidism with interaction on gut-thyroid axis. Clin Sci (Lond, Engl: 1979). 2020;134(12):1521–35.
- 24. Cayres LCdF, et al. Detection of alterations in the gut microbiota and intestinal permeability in patients with Hashimoto thyroiditis. Front Immunol. 2021;12:579140.
- 25. Knezevic J, et al. Thyroid-gut-axis: how does the microbiota infuence thyroid function? Nutrients. 2020;12(6):1769.
- 26. Virili C, Centanni M. "With a little help from my friends"—the role of microbiota in thyroid hormone metabolism and enterohepatic recycling. Mol Cell Endocrinol. 2017;458:39–43.
- 27. Tomasello G, et al. Dysmicrobism, infammatory bowel disease and thyroiditis: analysis of the literature. J Biol Regul Homeost Agents. 2015;29(2):265–72.
- 28. Guo Q, et al. Cytokine secretion and pyroptosis of thyroid follicular cells mediated by enhanced NLRP3, NLRP1, NLRC4, and AIM2 infammasomes are associated with autoimmune thyroiditis. Front Immunol. 2018;9:1197.
- 29. Fenneman AC, et al. Gut microbiota and metabolites in the pathogenesis of endocrine disease. Biochem Soc Trans. 2020;48(3):915–31.
- 30. Schroeder BO, Bäckhed F. Signals from the gut microbiota to distant organs in physiology and disease. Nat Med. 2016;22(10):1079–89.
- 31. Donaldson GP, Lee SM, Mazmanian SK. Gut biogeography of the bacterial microbiota. Nat Rev Microbiol. 2016;14(1):20–32.
- 32. Levy M, et al. Dysbiosis and the immune system. Nat Rev Immunol. 2017;17(4):219–32.
- 33. Fujisaka S, Watanabe Y, Tobe K. The gut microbiome: a core regulator of metabolism. J Endocrinol. 2022;256(3):e220111.
- 34. Gong B, et al. Association between gut microbiota and autoimmune thyroid disease: a systematic review and meta-analysis. Front Endocrinol. 2021;12:774362.
- 35. Zhao F, et al. Alterations of the gut microbiota in Hashimoto's thyroiditis patients. Thyroid. 2018;28(2):175–86.
- 36. Ishaq HM, et al. Molecular estimation of alteration in intestinal microbial composition in Hashimoto's thyroiditis patients. Biomed Pharmacother. 2017;95:865–74.
- 37. Liu J, et al. Analysis of gut microbiota diversity in Hashimoto's thyroiditis patients. BMC Microbiol. 2022;22(1):318.
- 38. Cornejo-Pareja I, et al. Diferential microbial pattern description in subjects with autoimmune-based thyroid diseases: a pilot study. J Person Med. 2020;10(4):192.
- 39. Stojanov S, Berlec A, Štrukelj B. The infuence of probiotics on the frmicutes/bacteroidetes ratio in the treatment of obesity and infammatory bowel disease. Microorganisms. 2020;8(11):1715.
- 40. Fröhlich E, Wahl R. Microbiota and thyroid interaction in health and disease. Trends Endocrinol Metab. 2019;30(8):479–90.
- 41. Liu S, et al. The composition of gut microbiota in patients bearing Hashimoto's thyroiditis with euthyroidism and hypothyroidism. Int J Endocrinol. 2020;2020:5036959.
- 42. Jiao Y, et al. Crosstalk between gut microbiota and innate immunity and its implication in autoimmune diseases. Front Immunol. 2020;11:282.
- 43. Medzhitov R, Janeway C. The Toll receptor family and microbial recognition. Trends Microbiol. 2000;8(10):452–6.
- 44. Woo V, Alenghat T. Host–microbiota interactions: epigenomic regulation. Curr Opin Immunol. 2017;44:52–60.
- 45. Sekirov I, et al. Gut microbiota in health and disease. Physiol Rev. 2010;90(3):859–904.
- 46. Thoo L, Noti M, Krebs P. Keep calm: the intestinal barrier at the interface of peace and war. Cell Death Dis. 2019;10(11):849.
- 47. Wu H-J, Wu E. The role of gut microbiota in immune homeostasis and autoimmunity. Gut Microbes. 2012;3(1):4–14.
- 48. Atarashi K, et al. ATP drives lamina propria T(H)17 cell differentiation. Nature. 2008;455(7214):808–12.
- 49. Satoh-Takayama N, et al. Microbial flora drives interleukin 22 production in intestinal NKp46+ cells that provide innate mucosal immune defense. Immunity. 2008;29(6):958–70.
- 50. Sanos SL, et al. RORgammat and commensal microfora are required for the diferentiation of mucosal interleukin 22-producing NKp46+ cells. Nat Immunol. 2009;10(1):83–91.
- 51. Kunii J, et al. Commensal bacteria promote migration of mast cells into the intestine. Immunobiology. 2011;216(6):692–7.
- 52. Bischof SC, Krämer S. Human mast cells, bacteria, and intestinal immunity. Immunol Rev. 2007;217:329–37.
- 53. Lavin Y, et al. Regulation of macrophage development and function in peripheral tissues. Nat Rev Immunol. 2015;15(12):731–44.
- 54. Virili C, Stramazzo I, Centanni M. Gut microbiome and thyroid autoimmunity. Best Pract Res Clin Endocrinol Metab. 2021;35(3):101506.
- 55. Round JL, Mazmanian SK. Inducible Foxp3+ regulatory T-cell development by a commensal bacterium of the intestinal microbiota. Proc Natl Acad Sci USA. 2010;107(27):12204–9.
- 56. Yasuda K, Takeuchi Y, Hirota K. The pathogenicity of Th17 cells in autoimmune diseases. Semin Immunopathol. 2019;41(3):283–97.
- 57. Cekanaviciute E, et al. Gut bacteria from multiple sclerosis patients modulate human T cells and exacerbate symptoms in mouse models. Proc Natl Acad Sci USA. 2017;114(40):10713–8.
- 58. Ostman S, et al. Impaired regulatory T cell function in germ-free mice. Eur J Immunol. 2006;36(9):2336–46.
- 59. Ivanov II, et al. Induction of intestinal Th17 cells by segmented flamentous bacteria. Cell. 2009;139(3):485–98.
- 60. Wang Y, et al. Induction of intestinal Th17 cells by flagellins from segmented filamentous bacteria. Front Immunol. 2019;10:2750.
- 61. Telesford KM, et al. A commensal symbiotic factor derived from Bacteroides fragilis promotes human CD39(+)Foxp3(+) T cells and Treg function. Gut Microbes. 2015;6(4):234–42.
- 62. Mazmanian SK, et al. An immunomodulatory molecule of symbiotic bacteria directs maturation of the host immune system. Cell. 2005;122(1):107–18.
- 63. Wang Y, et al. A commensal bacterial product elicits and modulates migratory capacity of CD39(+) CD4 T regulatory subsets in the suppression of neuroinfammation. Gut Microbes. 2014;5(4):552–61.
- 64. Mørland B, Midtvedt T. Phagocytosis, peritoneal influx, and enzyme activities in peritoneal macrophages from germfree, conventional, and ex-germfree mice. Infect Immun. 1984;44(3):750–2.
- 65. Zhang W, et al. Lactic acid bacterial colonization and human rotavirus infection infuence distribution and frequencies of monocytes/macrophages and dendritic cells in neonatal gnotobiotic pigs. Vet Immunol Immunopathol. 2008;121(3–4):222–31.
- 66. Benvenga S, Guarneri F. Molecular mimicry and autoimmune thyroid disease. Rev Endocr Metab Disord. 2016;17(4):485–98.
- 67. Sousa MdAC, Paraná R, Andrade LJdO. Sequence similarity between thyroid self-protein and hepatitis C virus polyprotein: possible triggering mechanism of autoimmune thyroiditis. Arq Gastroenterol. 2016;53(3):185–91.
- 68. Cusick MF, Libbey JE, Fujinami RS. Molecular mimicry as a mechanism of autoimmune disease. Clin Rev Allergy Immunol. 2012;42(1):102–11.
- 69. Kiseleva EP, et al. The role of components of Bifidobacterium and Lactobacillus in pathogenesis and serologic diagnosis of autoimmune thyroid diseases. Benef Microbes. 2011;2(2):139–54.
- 70. Masala S, et al. Lack of humoral response against Helicobacter pylori peptides homologous to human ZnT8 in Hashimoto's thyroiditis patients. J Infect Dev Ctries. 2015;9(6):631–4.
- 71. Benvenga S, et al. Human thyroid autoantigens and proteins of Yersinia and Borrelia share amino acid sequence homology that includes binding motifs to HLA-DR molecules and T-cell receptor. Thyroid. 2006;16(3):225–36.
- 72. Ruf WE, Kriegel MA. Autoimmune host-microbiota interactions at barrier sites and beyond. Trends Mol Med. 2015;21(4):233–44.
- 73. Chervonsky AV. Microbiota and autoimmunity. Cold Spring Harb Perspect Biol. 2013;5(3):a007294.
- 74. Arata N, et al. By-stander activation in autoimmune thyroiditis: studies on experimental autoimmune thyroiditis in the GFP+ fuorescent mouse. Clin Immunol (Orlando, Fla). 2006;121(1):108–17.
- 75. McLachlan SM, Rapoport B. Thyroid autoantibodies display both "original antigenic sin" and epitope spreading. Front Immunol. 2017;8:1845.
- 76. Thrasyvoulides A, Lymberi P. Evidence for intramolecular B-cell epitope spreading during experimental immunization with an immunogenic thyroglobulin peptide. Clin Exp Immunol. 2003;132(3):401–7.
- 77. Lerner A, Aminov R, Matthias T. Dysbiosis may trigger autoimmune diseases via inappropriate post-translational modifcation of host proteins. Front Microbiol. 2016;7:84.
- 78. Muller S, Radic M. Citrullinated autoantigens: from diagnostic markers to pathogenetic mechanisms. Clin Rev Allergy Immunol. 2015;49(2):232–9.
- 79. Macdonald TT, Monteleone G. Immunity, inflammation, and allergy in the gut. Science (New York, NY). 2005;307(5717):1920–5.
- 80. Natividad JMM, Verdu EF. Modulation of intestinal barrier by intestinal microbiota: pathological and therapeutic implications. Pharmacol Res. 2013;69(1):42–51.
- 81. Meslin JC, Fontaine N, Andrieux C. Variation of mucin distribution in the rat intestine, caecum and colon: efect of the bacterial fora. Comp Biochem Physiol Part A Mol Integr Physiol. 1999;123(3):235–9.
- 82. Čaja F, et al. Immune activation by microbiome shapes the colon mucosa: Comparison between healthy rat mucosa under conventional and germ-free conditions. J Immunotoxicol. 2021;18(1):37–49.
- 83. Paray BA, et al. Leaky gut and autoimmunity: an intricate balance in individuals health and the diseased state. Int J Mol Sci. 2020;21(24):9770.
- 84. Demir E, et al. The relationship between elevated plasma zonulin levels and Hashimoto's thyroiditis. Turk J Med Sci. 2022;52(3):605–12.
- 85. Fasano A. Zonulin and its regulation of intestinal barrier function: the biological door to infammation, autoimmunity, and cancer. Physiol Rev. 2011;91(1):151–75.
- 86. Manfredo Vieira S, et al. Translocation of a gut pathobiont drives autoimmunity in mice and humans. Science (New York, NY). 2018;359(6380):1156–61.
- 87. Yang Z, Klionsky DJ. Eaten alive: a history of macroautophagy. Nat Cell Biol. 2010;12(9):814–22.
- 88. Haq S, et al. Autophagy: roles in intestinal mucosal homeostasis and infammation. J Biomed Sci. 2019;26(1):19.
- 89. Yang L, et al. Impaired autophagy in intestinal epithelial cells alters gut microbiota and host immune responses. Appl Environ Microbiol. 2018;84(18):e00880–18.
- 90. Zhang C, et al. Inhibition of autophagic degradation process contributes to claudin-2 expression increase and epithelial tight junction dysfunction in TNF-α treated cell monolayers. Int J Mol Sci. 2017;18(1):157.
- 91. Hu C-AA, et al. Autophagy and tight junction proteins in the intestine and intestinal diseases. Anim Nutr (Zhongguo Xu Mu Shou Yi Xue Hui). 2015;1(3):123–7.
- 92. Zhou R, et al. A role for mitochondria in NLRP3 infammasome activation. Nature. 2011;469(7329):221–5.
- 93. Foerster EG, et al. How autophagy controls the intestinal epithelial barrier. Autophagy. 2022;18(1):86–103.
- 94. Burek CL, Rose NR. Autoimmune thyroiditis and ROS. Autoimmun Rev. 2008;7(7):530–7.
- 95. Konca Degertekin C, et al. Circulating Th17 cytokine levels are altered in Hashimoto's thyroiditis. Cytokine. 2016;80:13–7.
- 96. Lee Y, et al. Induction and molecular signature of pathogenic TH17 cells. Nat Immunol. 2012;13(10):991–9.
- 97. Zheng T, et al. Increased Interleukin-23 in Hashimoto's thyroiditis disease induces autophagy suppression and reactive oxygen species accumulation. Front Immunol. 2018;9:96.
- 98. Lu Q, et al. Caveolin-1 regulates autophagy activity in thyroid follicular cells and is involved in Hashimoto's thyroiditis disease. Endocr J. 2018;65(9):893–901.
- 99. Tao T, Xu H. Autophagy and thyroid disease. Adv Exp Med Biol. 2020;1207:435–43.
- 100. Pan H, et al. NLRP3 and gut microbiota homeostasis: progress in research. Cells. 2022;11(23):3758.
- 101. Zhao R, Zhou H, Su SB. A critical role for interleukin-1β in the progression of autoimmune diseases. Int Immunopharmacol. 2013;17(3):658–69.
- 102. Papadakis KA, et al. Dominant role for TL1A/DR3 pathway in IL-12 plus IL-18-induced IFN-gamma production by peripheral blood and mucosal CCR9+ T lymphocytes. J Immunol (Baltimore, Md: 1950). 2005;174(8):4985–90.
- 103. Lalor SJ, et al. Caspase-1-processed cytokines IL-1beta and IL-18 promote IL-17 production by gammadelta and CD4 T cells that mediate autoimmunity. J Immunol (Baltimore, Md: 1950). 2011;186(10):5738–48.
- 104. Shao S, et al. Therapeutic potential of the target on NLRP3 inflammasome in multiple sclerosis. Pharmacol Ther. 2021;227:107880.
- 105. Anton-Pampols P, et al. The role of infammasomes in glomerulonephritis. Int J Mol Sci. 2022;23(8):4208.
- 106. Kayagaki N, et al. Noncanonical infammasome activation by intracellular LPS independent of TLR4. Science (New York, NY). 2013;341(6151):1246–9.
- 107. Jiang W, et al. The relationships between the gut microbiota and its metabolites with thyroid diseases. Front Endocrinol. 2022;13:943408.
- 108. Brown AJ, et al. The Orphan G protein-coupled receptors GPR41 and GPR43 are activated by propionate and other short chain carboxylic acids. J Biol Chem. 2003;278(13):11312–9.
- 109. Aoyama M, Kotani J, Usami M. Butyrate and propionate induced activated or non-activated neutrophil apoptosis via HDAC inhibitor activity but without activating GPR-41/GPR-43 pathways. Nutrition (Burbank, Los Angeles County, Calif). 2010;26(6):653–61.
- 110. Segain JP, et al. Butyrate inhibits infammatory responses through NFkappaB inhibition: implications for Crohn's disease. Gut. 2000;47(3):397–403.
- 111. He J, et al. Short-chain fatty acids and their association with signalling pathways in infammation, glucose and lipid metabolism. Int J Mol Sci. 2020;21(17):6356.
- 112. Chang PV, et al. The microbial metabolite butyrate regulates intestinal macrophage function via histone deacetylase inhibition. Proc Natl Acad Sci USA. 2014;111(6):2247–52.
- 113. Macia L, et al. Metabolite-sensing receptors GPR43 and GPR109A facilitate dietary fibre-induced gut homeostasis through regulation of the infammasome. Nat Commun. 2015;6:6734.
- 114. Yang W, et al. Intestinal microbiota-derived short-chain fatty acids regulation of immune cell IL-22 production and gut immunity. Nat Commun. 2020;11(1):4457.
- 115. Pan X, et al. Butyrate ameliorates caerulein-induced acute pancreatitis and associated intestinal injury by tissue-specifc mechanisms. Br J Pharmacol. 2019;176(23):4446–61.
- 116. Meng S, et al. Short-chain fatty acids and thyroid hormone interact in regulating enterocyte gene transcription. Surgery. 1999;126(2):293–8.
- 117. Devlin AS, Fischbach MA. A biosynthetic pathway for a prominent class of microbiota-derived bile acids. Nat Chem Biol. 2015;11(9):685–90.
- 118. Lavelle A, Sokol H. Gut microbiota-derived metabolites as key actors in infammatory bowel disease. Nat Rev Gastroenterol Hepatol. 2020;17(4):223–37.
- 119. Song X, et al. Microbial bile acid metabolites modulate gut RORγ+ regulatory T cell homeostasis. Nature. 2020;577(7790):410–5.
- 120. Zhang X, et al. Trimethylamine-N-oxide promotes vascular calcifcation through activation of NLRP3 (nucleotide-binding domain, leucine-rich-containing family, pyrin domain-containing-3) infammasome and NF-κB (nuclear factor κB) signals. Arterioscler Thromb Vasc Biol. 2020;40(3):751–65.
- 121. Schirmer M, et al. Linking the human gut microbiome to inflammatory cytokine production capacity. Cell. 2016;167(4):1125.
- 122. Nicola JP, et al. NF-kappaB p65 subunit mediates lipopolysaccharide-induced $Na(+)/I(-)$ symporter gene expression by involving functional interaction with the paired domain transcription factor Pax8. Mol Endocrinol (Baltimore, Md). 2010;24(9):1846–62.
- 123. Jin B, Wang S, Fan Z. Pathogenesis markers of Hashimoto's disease-a mini review. Front Biosci (Landmark Edition). 2022;27(10):297.
- 124. Leung AM, Braverman LE. Consequences of excess iodine. Nat Rev Endocrinol. 2014;10(3):136–42.
- 125. Liu J, et al. Excessive iodine promotes pyroptosis of thyroid follicular epithelial cells in Hashimoto's thyroiditis through the ROS-NF-κB-NLRP3 pathway. Front Endocrinol. 2019;10:778.
- 126. Duntas LH. The role of iodine and selenium in autoimmune thyroiditis. Horm Metab Res. 2015;47(10):721–6.
- 127. Drutel A, Archambeaud F, Caron P. Selenium and the thyroid gland: more good news for clinicians. Clin Endocrinol. 2013;78(2):155–64.
- 128. Calomme M, et al. Seleno-lactobacillus. An organic selenium source. Biol Trace Elem Res. 1995;47(1–3):379–83.
- 129. Kasaikina MV, et al. Dietary selenium afects host selenoproteome expression by infuencing the gut microbiota. FASEB J. 2011;25(7):2492–9.
- 130. Zimmermann MB. The infuence of iron status on iodine utilization and thyroid function. Annu Rev Nutr. 2006;26:367–89.
- 131. Constante M, et al. Iron supplements modulate colon microbiota composition and potentiate the protective efects of probiotics in dextran sodium sulfate-induced colitis. Infamm Bowel Dis. 2017;23(5):753–66.
- 132. Chieppa M, Giannelli G. Immune cells and microbiota response to iron starvation. Front Med. 2018;5:109.
- 133. Bouglé D, et al. Influence of short-chain fatty acids on iron absorption by proximal colon. Scand J Gastroenterol. 2002;37(9):1008–11.
- 134. Dostal A, et al. Low iron availability in continuous in vitro colonic fermentations induces strong dysbiosis of the child gut microbial consortium and a decrease in main metabolites. FEMS Microbiol Ecol. 2013;83(1):161–75.
- 135. Yilmaz B, Li H. Gut microbiota and iron: the crucial actors in health and disease. Pharmaceuticals (Basel, Switzerland). 2018;11(4):98.
- 136. Reed S, et al. Chronic zinc defciency alters chick gut microbiota composition and function. Nutrients. 2015;7(12):9768–84.
- 137. Severo JS, et al. The role of zinc in thyroid hormones metabolism. Int J Vitam Nutr Res. 2019;89(1–2):80–8.
- 138. Rutgers M, et al. Enterohepatic circulation of triiodothyronine (T3) in rats: importance of the microfora for the liberation and reabsorption of T3 from biliary T3 conjugates. Endocrinology. 1989;125(6):2822–30.
- 139. Nguyen TT, et al. 5'- and 5-deiodinase activities in adult rat cecum and large bowel contents inhibited by intestinal microfora. Am J Physiol. 1993;265(3 Pt 1):E521–4.
- 140. Spaggiari G, et al. Probiotics ingestion does not directly afect thyroid hormonal parameters in hypothyroid patients on levothyroxine treatment. Front Endocrinol. 2017;8:316.
- 141. Daher R, et al. Consequences of dysthyroidism on the digestive tract and viscera. World J Gastroenterol. 2009;15(23):2834–8.
- 142. Lauritano EC, et al. Association between hypothyroidism and small intestinal bacterial overgrowth. J Clin Endocrinol Metab. 2007;92(11):4180–4.
- 143. Gunsar F, et al. Efect of hypo- and hyperthyroidism on gastric myoelectrical activity. Dig Dis Sci. 2003;48(4):706–12.
- 144. Fukunaga K. Refractory gastrointestinal bleeding treated with thyroid hormone replacement. J Clin Gastroenterol. 2001;33(2):145–7.
- 145. Bibbò S, et al. Chronic autoimmune disorders are increased in coeliac disease: a case-control study. Medicine. 2017;96(47):e8562.
- 146. Rotondi Aufero V, Fasano A, Mazzarella G. Non-celiac gluten sensitivity: how its gut immune activation and potential dietary management difer from celiac disease. Mol Nutr Food Res. 2018;62(9):e1700854.
- 147. Cuan-Baltazar Y, Soto-Vega E. Microorganisms associated to thyroid autoimmunity. Autoimmun Rev. 2020;19(9):102614.
- 148. Cellini M, et al. Hashimoto's thyroiditis and autoimmune gastritis. Front Endocrinol. 2017;8:92.
- 149. Shi W-J, et al. Associations of helicobacter pylori infection and cytotoxin-associated gene A status with autoimmune thyroid diseases: a meta-analysis. Thyroid. 2013;23(10):1294–300.
- 150. Checchi S, et al. Prevalence of parietal cell antibodies in a large cohort of patients with autoimmune thyroiditis. Thyroid. 2010;20(12):1385–9.
- 151. Nicolaou A, et al. Predictive value of gastrin levels for the diagnosis of gastric enterochromafn-like cell hyperplasia in patients with Hashimoto's thyroiditis. Neuroendocrinology. 2014;99(2):118–22.
- 152. Rao SSC, Bhagatwala J. Small intestinal bacterial overgrowth: clinical features and therapeutic management. Clin Transl Gastroenterol. 2019;10(10):e00078.
- 153. Shah SC, et al. Meta-analysis: antibiotic therapy for small intestinal bacterial overgrowth. Aliment Pharmacol Ther. 2013;38(8):925–34.
- 154. Lauritano EC, et al. Antibiotic therapy in small intestinal bacterial overgrowth: rifaximin versus metronidazole. Eur Rev Med Pharmacol Sci. 2009;13(2):111–6.
- 155. Kim M-S, et al. The efect of rifaximin on gut fora and Staphylococcus resistance. Dig Dis Sci. 2013;58(6):1676–82.
- 156. Xu D, et al. Rifaximin alters intestinal bacteria and prevents stress-induced gut infammation and visceral hyperalgesia in rats. Gastroenterology. 2014;146(2):484.
- 157. Maccaferri S, et al. Rifaximin modulates the colonic microbiota of patients with Crohn's disease: an in vitro approach using a continuous culture colonic model system. J Antimicrob Chemother. 2010;65(12):2556–65.
- 158. Lauritano EC, et al. Rifaximin dose-fnding study for the treatment of small intestinal bacterial overgrowth. Aliment Pharmacol Ther. 2005;22(1):31–5.
- 159. Scarpellini E, et al. High dosage rifaximin for the treatment of small intestinal bacterial overgrowth. Aliment Pharmacol Ther. 2007;25(7):781–6.
- 160. McFarland LV. Systematic review and meta-analysis of Saccharomyces boulardii in adult patients. World J Gastroenterol. 2010;16(18):2202–22.
- 161. Wieërs G, et al. How probiotics afect the microbiota. Front Cell Infect Microbiol. 2019;9:454.
- 162. Huo D, et al. Probiotic Bifidobacterium longum supplied with methimazole improved the thyroid function of Graves' disease patients through the gut-thyroid axis. Commun Biol. 2021;4(1):1046.
- 163. Williams EA, et al. Clinical trial: a multistrain probiotic preparation signifcantly reduces symptoms of irritable bowel syndrome in a double-blind placebo-controlled study. Aliment Pharmacol Ther. 2009;29(1):97–103.
- 164. Roberfroid M. Prebiotics: the concept revisited. J Nutr. 2007;137(3 Suppl 2):830S-837S.
- 165. Talebi S, et al. The effects of synbiotic supplementation on thyroid function and infammation in hypothyroid patients: a randomized, double-blind, placebo-controlled trial. Complement Ther Med. 2020;48:102234.
- 166. Tsilingiri K, Rescigno M. Postbiotics: what else? Benef Microbes. 2013;4(1):101–7.
- 167. Żółkiewicz J, et al. Postbiotics-a step beyond pre- and probiotics. Nutrients. 2020;12(8):2189.
- 168. Zheng L, et al. Fecal microbiota transplantation in the metabolic diseases: current status and perspectives. World J Gastroenterol. 2022;28(23):2546–60.
- 169. van Nood E, et al. Duodenal infusion of donor feces for recurrent Clostridium difficile. N Engl J Med. 2013;368(5):407-15.
- 170. Maslowski KM, Mackay CR. Diet, gut microbiota and immune responses. Nat Immunol. 2011;12(1):5–9.
- 171. David LA, et al. Diet rapidly and reproducibly alters the human gut microbiome. Nature. 2014;505(7484):559–63.
- 172. Zhu W, et al. Gut microbial metabolite TMAO enhances platelet hyperreactivity and thrombosis risk. Cell. 2016;165(1):111–24.
- 173. Wang Z, et al. Non-lethal inhibition of gut microbial trimethylamine production for the treatment of atherosclerosis. Cell. 2015;163(7):1585–95.
- 174. Konijeti GG, et al. Efficacy of the autoimmune protocol diet for inflammatory bowel disease. Inflamm Bowel Dis. 2017;23(11):2054–60.
- 175. Abbott RD, Sadowski A, Alt AG. Efficacy of the autoimmune protocol diet as part of a multi-disciplinary, supported lifestyle intervention for Hashimoto's thyroiditis. Cureus. 2019;11(4):e4556.
- 176. Ruggeri RM, et al. Infuence of dietary habits on oxidative stress markers in Hashimoto's thyroiditis. Thyroid. 2021;31(1):706–7.
- 177. Virili C, et al. Atypical celiac disease as cause of increased need for thyroxine: a systematic study. J Clin Endocrinol Metab. 2012;97(3):E419–22.
- 178. Krysiak R, Szkróbka W, Okopień B. The efect of gluten-free diet on thyroid autoimmunity in drug-naïve women with Hashimoto's thyroiditis: a pilot study. Exp Clin Endocrinol Diabetes. 2019;127(7):417–22.
- 179. Mikulska AA, et al. Metabolic characteristics of Hashimoto's thyroiditis patients and the role of microelements and diet in the disease management-an overview. Int J Mol Sci. 2022;23(12):6580.
- 180. Thomas RL, et al. Vitamin D metabolites and the gut microbiome in older men. Nat Commun. 2020;11(1):5997.
- 181. Chahardoli R, et al. Can supplementation with vitamin D modify thyroid autoantibodies (anti-TPO Ab, Anti-Tg Ab) and thyroid profle (T3, T4, TSH) in Hashimoto's thyroiditis? A double blind, randomized clinical trial. Horm Metab Res. 2019;51(5):296–301.
- 182. Fasano A, et al. Zonulin, a newly discovered modulator of intestinal permeability, and its expression in coeliac disease. Lancet (London, England). 2000;355(9214):1518–9.
- 183. Tripathi A, et al. Identifcation of human zonulin, a physiological modulator of tight junctions, as prehaptoglobin-2. Proc Natl Acad Sci USA. 2009;106(39):16799–804.
- 184. Zheng D, et al. Elevated levels of circulating biomarkers related to leaky gut syndrome and bacterial translocation are associated with Graves' disease. Front Endocrinol. 2021;12:796212.
- 185. Martinon F, Burns K, Tschopp J. The infammasome: a molecular platform triggering activation of infammatory caspases and processing of proIL-beta. Mol Cell. 2002;10(2):417–26.
- 186. Kawashima A, et al. Demonstration of innate immune responses in the thyroid gland: potential to sense danger and a possible trigger for autoimmune reactions. Thyroid. 2013;23(4):477–87.
- 187. Liu Z, et al. Thyrocyte interleukin-18 expression is up-regulated by interferon-γ and may contribute to thyroid destruction in Hashimoto's thyroiditis. Int J Exp Pathol. 2010;91(5):420–5.
- 188. Seo S-U, et al. Distinct commensals induce interleukin-1β via NLRP3 inflammasome in inflammatory monocytes to promote intestinal infammation in response to injury. Immunity. 2015;42(4):744–55.
- 189. Yao X, et al. Remodelling of the gut microbiota by hyperactive NLRP3 induces regulatory T cells to maintain homeostasis. Nat Commun. 2017;8(1):1896.
- 190. Furusawa Y, et al. Commensal microbe-derived butyrate induces the diferentiation of colonic regulatory T cells. Nature. 2013;504(7480):446–50.
- 191. Nicola JP, et al. Functional toll-like receptor 4 conferring lipopolysaccharide responsiveness is expressed in thyroid cells. Endocrinology. 2009;150(1):500–8.
- 192. Sánchez E, et al. Contribution of TNF-alpha and nuclear factorkappaB signaling to type 2 iodothyronine deiodinase activation in the mediobasal hypothalamus after lipopolysaccharide administration. Endocrinology. 2010;151(8):3827–35.
- 193. Baur A, et al. Efects of proinfammatory cytokines on anterior pituitary 5'-deiodinase type I and type II. J Endocrinol. 2000;167(3):505–15.
- 194. Doden H, et al. Metabolism of oxo-bile acids and characterization of recombinant 12α-hydroxysteroid dehydrogenases from bile acid 7α-dehydroxylating human gut bacteria. Appl Environ Microbiol. 2018;84(10):e00235–18.
- 195. Hylemon PB, et al. Bile acids as regulatory molecules. J Lipid Res. 2009;50(8):1509–20.
- 196. Liu J, et al. Serum metabolomic patterns in patients with autoimmune thyroid disease. Endocr Pract. 2020;26(1):82–96.
- 197. Lu X, et al. Changes in histone H3 lysine 4 trimethylation in Hashimoto's thyroiditis. Arch Med Sci AMS. 2022;18(1):153–63.
- 198. Zhao S, et al. H3K4 methylation regulates LPS-induced proinfammatory cytokine expression and release in macrophages. Shock (Augusta, Ga). 2019;51(3):401–6.
- 199. Bassi V, et al. Identification of a correlation between Helicobacter pylori infection and Graves' disease. Helicobacter. 2010;15(6):558–62.

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