



REVIEW

Bile acids, gut microbiota, and therapeutic insights in hepatocellular carcinoma

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ABSTRACT

Hepatocellular carcinoma (HCC) is a prevalent and aggressive liver malignancy. The interplay between bile acids (BAs) and the gut microbiota has emerged as a critical factor in HCC development and progression. Under normal conditions, BA metabolism is tightly regulated through a bidirectional interplay between gut microorganisms and BAs. The gut microbiota plays a critical role in BA metabolism, and BAs are endogenous signaling molecules that help maintain liver and intestinal homeostasis. Of note, dysbiotic changes in the gut microbiota during pathogenesis and cancer development can disrupt BA homeostasis, thereby leading to liver inflammation and fibrosis, and ultimately contributing to HCC development. Therefore, understanding the intricate interplay between BAs and the gut microbiota is crucial for elucidating the mechanisms underlying hepatocarcinogenesis. In this review, we comprehensively explore the roles and functions of BA metabolism, with a focus on the interactions between BAs and gut microorganisms in HCC. Additionally, therapeutic strategies targeting BA metabolism and the gut microbiota are discussed, including the use of BA agonists/antagonists, probiotic/prebiotic and dietary interventions, fecal microbiota transplantation, and engineered bacteria. In summary, understanding the complex BA-microbiota crosstalk can provide valuable insights into HCC development and facilitate the development of innovative therapeutic approaches for liver malignancy.

KEYWORDS

Bile acid; gut microbiota; hepatocellular carcinoma; therapeutics; microbiota modulation

Introduction

Hepatocellular carcinoma (HCC) is the most common primary liver malignancy worldwide. This disease is typically diagnosed in advanced stages, because of its aggressive nature and rapid progression, thus resulting in high mortality and limited treatment options^{1,2}. HCC arises primarily from chronic liver diseases with various etiologies, including viral hepatitis, alcoholic liver disease, and non-alcoholic fatty liver disease (NAFLD)^{3,4}. The intricate interplay among genetic and environmental factors contributes to the complex tumorigenesis mechanism of HCC. Understanding the risk factors

associated with HCC is essential for the development of diagnostic biomarkers, therapeutic targets, and preventive strategies against this lethal disease.

Recent studies have revealed the emerging role of the gut microbiota in HCC development and progression. The human gastrointestinal tract harbors a diverse and dynamic community of microorganisms forming the gut microbiota, which consists primarily of bacteria but also includes viruses and fungi⁵⁻⁷. These gut microorganisms play critical roles in dietary digestion, such as breaking down carbohydrates, producing vitamins, and metabolizing dietary components that human hosts cannot digest⁸⁻¹⁰. The gut microbiota also interacts with the host, and influences metabolism, immunity, and even brain function^{11,12}. However, the physiological crosstalk between hosts and microorganisms is greatly impaired in disease conditions, because of pathological alterations in gut microbial composition and function, which are commonly known as dysbiosis. Gut dysbiosis has been implicated in promoting chronic liver inflammation, fibrosis, and HCC, through mechanisms such as immunomodulation, toxin release, and altered metabolite regulation, thereby influencing hepatocarcinogenesis.

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Bile acids (BAs) are steroid acids that are synthesized in the liver and promote absorption of dietary lipids. The regulation of BA metabolism involves interactions between host cells and gut microbiota. In the context of HCC patients, there is a notable alteration in BA metabolism and the reciprocal interaction between host cells and gut microbiota. In general, a dysbiotic microbiota can disrupt BA homeostasis, and contribute to hepatocellular damage and carcinogenesis. In contrast, dysregulated BA metabolism reshapes the composition and function of the gut microbiota, and further promotes the occurrence and extent of microbial dysbiosis. Notably, because the intricate interaction between BAs and the gut microbiota is fundamental to HCC development, many studies have investigated the therapeutic potential of approaches targeting BAs and/or the gut microbiota. In this review, we summarize recent research on the interaction between BAs and the gut microbiota in HCC. Therapeutic strategies aimed at modulating BA metabolism and microbial profiles for HCC prevention and treatment are also discussed.

BA metabolism and interaction with the gut microbiota

BA synthesis

The liver plays a crucial role in transforming cholesterol into primary BAs, specifically chenodeoxycholic acid (CDCA) and cholic acid (CA) in humans, and α - and β -muricholic acid (MCA) in rodents. BA synthesis, orchestrated by multiple cytochrome P450 (CYP) enzymes, begins with cholesterol 7 α -hydroxylase (CYP7A1), the rate-limiting enzyme converting cholesterol to 7 α -hydrocholesterol¹³. Subsequently, microsomal sterol 12 α -hydroxylase (CYP8B1) balances CDCA and CA synthesis¹⁴. Concurrently, in various organs, an alternative pathway is initiated by sterol 27 α -hydroxylase (CYP27A1), which converts cholesterol to 27-hydrocholesterol, thereby diversifying the pool of BA precursors¹⁵.

After synthesis, primary BAs undergo conjugation with taurine or glycine, thus increasing their water solubility, and are then transported to the bile duct *via* bile salt export pump (BSEP) and multidrug resistance-associated protein 2 (MRP2). These conjugated BAs are secreted into the duodenum, where they participate in the digestion of dietary fats and fat-soluble vitamins. As much as 95% of conjugated BAs is reabsorbed

by enterocytes through the apical sodium-dependent bile acid transporter (ASBT) in the distal ileum, then released into the portal circulation through basolateral BA transporters, including organic solute transporter α/β (OST α/β) and multidrug resistance protein 3 (MRP3). These reabsorbed BAs are then transferred into hepatocytes through transporters such as Na⁺-taurocholate cotransporting polypeptide (NTCP) and organic anion transporting polypeptide 1 (OATP1)¹⁶, metabolized, and eventually released into bile juice *via* BSEP and MRP2¹⁷. In general, BAs undergo approximately 10 enterohepatic cycles per day, and this cycle is substantially influenced by a variety of BA receptors and the gut microbiota. BA synthesis and transformation by gut microbiota are summarized in **Figure 1**.

BA receptors

BA receptors are necessary for regulating BA metabolism and homeostasis. The main BA receptors include farnesoid X receptor (FXR), Takeda G-protein-coupled receptor 5 (TGR5), pregnane X receptor (PXR), constitutive androstane receptor (CAR), and vitamin D receptor (VDR). These receptors together closely monitor a diverse set of signaling pathways that control BA synthesis, transport, and metabolism.

FXR is a nuclear receptor that acts as a transcription factor and initiates the expression of a diverse set of target genes. In the small intestine, FXR activation leads to the upregulation of fibroblast growth factor (FGF)-15/19, which is subsequently translocated into the liver and interacts with the FGF receptor 4 (FGFR4)/ β -klotho complex on hepatocytes. This complex then inactivates CYP7A1, thus decreasing BA production. Hepatocytic FXR also upregulates BSEP, thereby enhancing BA efflux into the biliary canaliculi while suppressing the BA importer NTCP, and decreasing BA reabsorption from the blood into hepatocytes^{18,19}.

TGR5, also known as G-protein-coupled bile acid receptor (GPBAR1), is expressed in various tissues and serves as a metabolic regulator of BA homeostasis²⁰. TGR5 is the receptor of primary BAs (e.g., CDCA and CA) and microbiota-derived secondary BAs [e.g., lithocholic acid (LCA), deoxycholic acid (DCA), and their conjugated forms]; the later are more potent activators of TGR5²¹. In contrast, TGR5 is critical in BA metabolism. In mice with *Tgr5* deficiency, the total BA amount is significantly lower, by 21%–25%, than that in wild-type mice²² and is accompanied by alterations in BA composition including decreased taurine-conjugated muricholic acid and increased taurocholic acid (TCA) and taurodeoxycholic

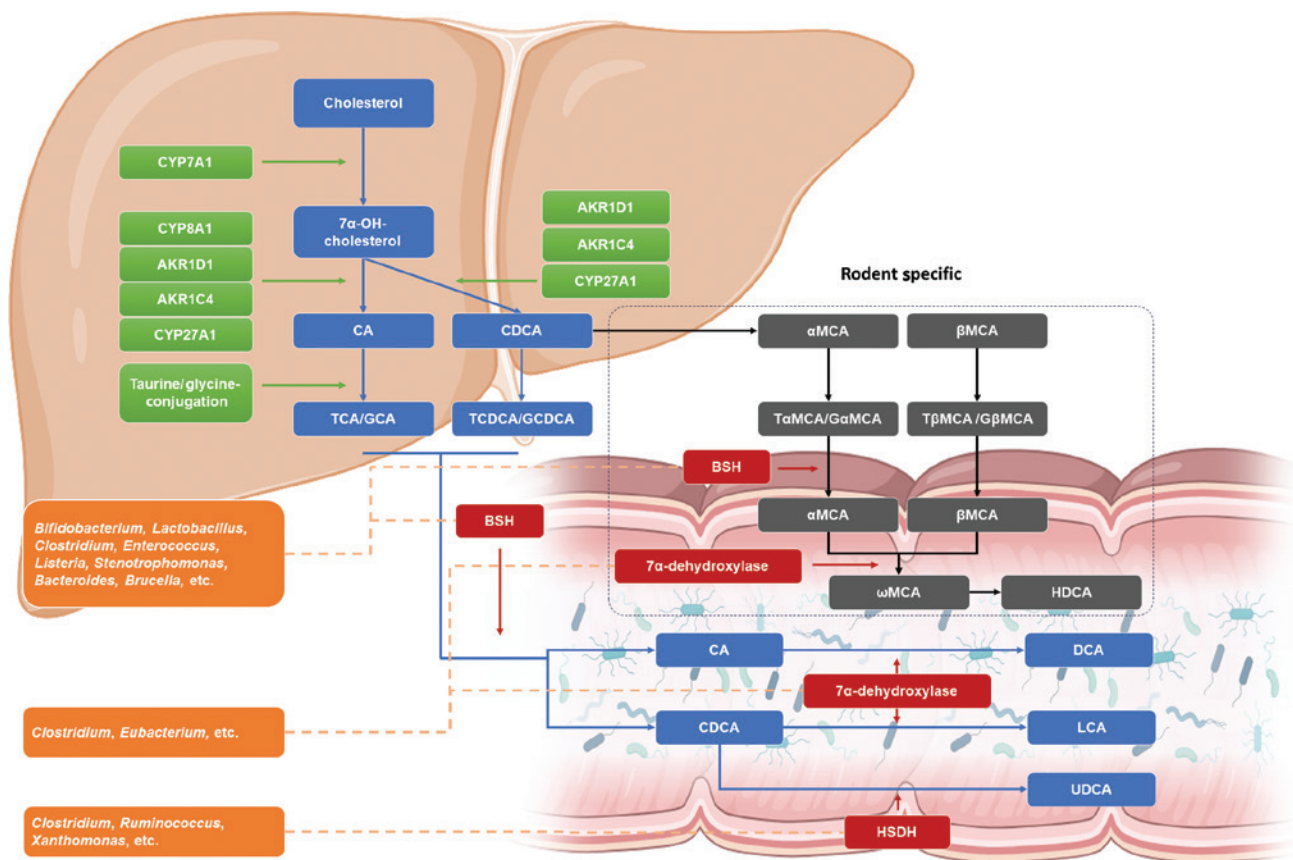


Figure 1 Bile acid synthesis and microbial transformation. In hepatocytes, cholesterol undergoes conversion to 7 α -OH-cholesterol by CYP7A1 or alternatively CYP27A1 (not depicted in this figure), thus catalyzing further synthesis. In the presence or absence of CYP8A1, primary bile acids (BAs), cholic acid (CA), or chenodeoxycholic acid (CDCA) is synthesized. In rodents, CDCA is further transformed into α MCA and β MCA. Primary BAs are conjugated with taurine/glycine, thus enhancing hydrophilicity, before being secreted with bile. In the intestine, secondary BAs are converted from primary BA by gut microorganisms. Bacteria with bile salt hydrolase (BSH) activity, including *Bifidobacterium*, *Lactobacillus*, *Clostridium*, and *Enterococcus*, deconjugate taurine/glycine-conjugated primary BAs. Bacteria including *Clostridium* and *Eubacterium*, expressing 7 α -dehydroxylase, convert CA to DCA and CDCA to LCA. Another enzyme, HSDH, expressed by bacteria including *Clostridium*, *Ruminococcus*, and *Xanthomonas*, transforms CDCA to UDCA. In rodents, α MCA and β MCA are converted to HDCA. BA, bile acids; BSH, bile salt hydrolase; CA, cholic acid; CDCA, chenodeoxycholic acid; CYP7A1, cholesterol 7 α -hydroxylase; CYP8A1, cholesterol 12 α -hydroxylase; CYP27A1, cholesterol 27 α -hydroxylase; DCA, deoxycholic acid; GCA, glycocholic acid; GCDCA, glycochenodeoxycholic acid; G β MCA, glycobetamuricholic acid; G α MCA, glycoalphamuricholic acid; HSDH, hydroxysteroid dehydrogenase; HDCA, hydoxycholic acid; LCA, lithocholic acid; TCA, taurocholic acid; TCDCa, taurochenodeoxycholic acid; T β MCA, taurobetamuricholic acid; T α MCA, tauroalphamuricholic acid; UDCA, ursodeoxycholic acid; α MCA, alphamuricholic acid; β MCA, betamuricholic acid; ω MCA, omegamuricholic acid. Figure generated with BioRender.com.

acid (TDCA)²³. In general, TGR5 activated by BAs stimulates various signaling pathways, such as the NF- κ B, AKT, and ERK pathways, and has potential implications in liver inflammation, hepatocyte proliferation, and other pathological process.

PXR and CAR are crucial nuclear receptors with major roles in regulating drug-metabolizing enzymes and transporters responsible for the metabolism and elimination of xenobiotics²⁴. These receptors are expressed predominantly in the liver but also have high intestinal expression. PXR is activated

by various BAs, whereas CAR is considered an indirect sensor of BAs^{25,26}. Activation of PXR and CAR stimulates BA detoxification, thus potentially counteracting cholestasis²⁷. Indeed, numerous studies have indicated the protective effects of PXR and CAR against BA toxicity. For example, a preclinical study has reported that PXR inhibits the pro-inflammatory response in hepatocytes *in vitro*, as evidenced by inactivation of the NF- κ B signaling pathway²⁸. In mice administered LCA or subjected to bile duct ligation, PXR protects hepatocytes

from injury by promoting the expression of enzymes involved in BA metabolism and transport²⁹. PXR also exhibits anti-inflammatory and anti-fibrosis effects in the intestines³⁰. Meanwhile, CAR, through a different protective mechanism, facilitates BA detoxification in the hepatocytes of CA-treated mice with *Fxr* and *Pxr* deficiencies^{31,32}. CAR also functions locally in intestinal CD4⁺ effector T cells, by contributing to BA detoxification and inflammation resolution by upregulating the anti-inflammatory cytokine interleukin (IL)-10 and BA-detoxifying enzymes and transporters²⁵.

VDR is a nuclear receptor originally known for its binding to calcitriol, the active form of vitamin D. Recent studies have revealed that VDR is also activated by BAs as well as several LCA-associated synthetic compounds, including LCA-acetate and LCA-propionate³³. In the liver, VDR downregulates the small heterodimer partner (SHP), and consequently increases CYP7A1 and BA synthesis³⁴. In a preclinical study, significantly diminished levels of total BAs and CDCA have been observed in mice with *Vdr* deficiency, even after CDCA supplementation, thus indicating that VDR is essential for CDCA metabolism³⁵. VDR also influences the expression of transporters involved in BA uptake and efflux, such as OST α/β , MRP3, MRP2, and BSEP^{36,37}. Notably, VDR interacts with other nuclear receptors including FXR and PXR, and consequently mediates BA metabolism. This crosstalk enables coordinated regulation of BA homeostasis.

BA metabolism is affected by the gut microbiota

In addition to host receptors, the gut microbiota plays a crucial role in BA metabolism, particularly in the synthesis of secondary BAs. These gut microorganisms inhabiting the gut express diverse enzymes that facilitate the conversion of primary BAs into secondary BAs. This transformation encompasses a series of intricate enzymatic reactions, including deconjugation, dehydroxylation, oxidation, epimerization, desulfurization, and re-conjugation³⁸.

Deconjugation is the initial step in BA modification, wherein the enzyme bacterial bile salt hydrolase (BSH) hydrolyzes the taurine and glycine conjugates of primary BAs. BSH activity has been observed in a wide range of bacterial genera, including *Bifidobacterium*, *Lactobacillus*, *Clostridium*, *Enterococcus*, *Listeria*, *Stenotrophomonas*, *Bacteroides*, and *Brucella*³⁹⁻⁴⁵. Once deconjugated, BAs become available for other microorganism-mediated biotransformation. Dehydroxylation, a crucial microbial transformation, involves the conversion of primary

BAs to secondary BAs through the removal of a hydroxyl group. This process is performed primarily by *Lachnospiraceae* and *Peptostreptococcaceae* bacteria, which carry the enzyme 7 α -dehydroxylase⁴⁶⁻⁴⁹. Of note, the product of dehydroxylation depends on the primary BA. For instance, 7 α -dehydroxylation of CDCA results in LCA formation, whereas 7 α -dehydroxylation of CA produces DCA^{48,50}. Moreover, the oxidation and epimerization of BAs are essential for decreasing the toxicity and increasing the hydrophilicity of BAs. Bacterial hydroxysteroid dehydrogenase (HSDH) is an enzyme catalyzing the oxidation of primary BAs to ketonic BAs. These ketonic BAs then undergo epimerization and further generate of β -hydroxy BAs, such as isodeoxycholic acid (IDCA) and isolithocholic acid (ILCA)⁵¹. HSDH is expressed predominantly in the bacterial phyla Actinobacteria, Proteobacteria, and Firmicutes^{52,53}. Several studies have suggested that this microorganism-mediated process helps the gut microbiota maintain resistance against the competitive and hostile intestinal environment⁵⁴. Some bacteria leveraging this characteristic modulate the composition of the BA pool, thus potentially enabling clinical applications. For instance, CDCA can be transformed into ursodeoxycholic acid (UDCA) through isomerization of its C-7 hydroxyl group⁵⁵. This conversion is facilitated by 2 crucial enzymes: 7 α -HSDH and 7 β -HSDH. These enzymes have been identified in various organisms such as *Clostridium*, *Ruminococcus*, and *Xanthomonas*.^{56,57} UDCA has been extensively studied for its health benefits and clinically applied in the treatment of cholestatic liver diseases⁵⁸⁻⁶⁰. Additionally, several bacteria express BA sulfatase enzymes and are involved in BA desulfurization. These enzymes remove sulfate groups from sulfated BAs, thus forming desulfated BAs, which are more easily reabsorbed by the intestine than their sulfated counterparts^{61,62}. Finally, gut microorganisms can re-conjugate unconjugated BAs with phenylalanine, tyrosine, and leucine, in a process that appears to be common in the human gut microbiota⁶³. Microbial modification of BA was listed in **Table 1**.

Recent advances in microbial profiling, such as high-throughput sequencing (e.g., 16S rRNA gene sequencing and metagenomics), and enhanced bioinformatic tools, have spurred an increase in high-quality studies revealing the influence of distinct gut microorganisms on BA metabolism. In a cohort of patients with NAFLD, numerous microorganism-derived BAs, particularly DCA, have been found to increase in tandem with disease activity and fibrosis progression. This increase is concomitant with the enrichment of *Bacteroidetes* and

Table 1 Microbial modification of bile acids

Modification	Process	Related bacteria	Related enzyme	References
Deconjugation	Transfer of glycine/taurine conjugated BAs to unconjugated BAs	Bacteroides, Firmicutes, and Actinobacteria	BSH	39-45
Dehydroxylation	Catalysis of the removal of hydroxyl groups from primary BAs, transforming them into secondary BAs	<i>Lachnospiraceae</i> and <i>Peptostreptococcaceae</i>	<i>Bai</i> operon proteins	46-50
Oxidation and epimerization	Formation of hydroxyl (-OH) or carbonyl (C=O) groups, and change in stereochemistry at a specific carbon atom	Actinobacteria, Proteobacteria, and Firmicutes	HSDH	51-53,56,57
Desulfurization	Removal of sulfate groups from sulfated BA, thus forming desulfated BAs	<i>Clostridium</i> , <i>Ruminococcus</i> , and <i>Xanthomonas</i>	Sulfatase	61,62
Reconjugation	Reconjugation of unconjugated BA with phenylalanine, tyrosine, and leucine	<i>Bacteroides</i> , <i>Bifidobacterium</i> , <i>Enterococcus</i> , and <i>Enterocloster</i>	Not reported	63

BA, bile acid; BSH, bile salt hydrolase; HSDH, hydroxysteroid dehydrogenase.

Lachnospiraceae, which bear genes responsible for DCA generation. In contrast, microbial populations susceptible to DCA are depleted, including *Ruminococcus*, *Prevotella*, *Lactobacillus*, and *Turicibacter*⁶⁴. Another pilot study has shown that disulfiram ameliorates NASH by decreasing *Clostridium*-mediated 7 α -dehydroxylation activity, thus suppressing secondary BA biosynthesis and consequently activating hepatic FXR signaling⁶⁵. Animal studies have provided further detail regarding the effects of gut microorganisms on BA metabolism. For instance, microorganism-derived BAs, such as HDCA, alleviate NAFLD in mice and concurrently enrich probiotic species such as *Parabacteroides distasonis*⁶⁶. In a separate study, *P. distasonis* has been found to produce various BAs, including LCA, DCA, ILCA, and 3-oxolithocholic acid, and to alleviate inflammatory responses by inhibiting Th17 cell differentiation and promoting M2 polarization of macrophages³⁹. Additionally, in mice with liver injury, gut microbial dysbiosis, characterized by the depletion of bacteria with BSH activity, including Bacteroidetes, has been found to inhibit intestinal FXR/FGF-15 signaling, thus ultimately facilitating liver injury development⁶⁷. These findings collectively highlight the intricate interplay between the gut microbial composition and BA metabolism, and have elucidated their major roles in various hepatic conditions.

BA metabolism and its influence on the composition of gut microbiota

Although BAs have been reported to have both positive and negative effects on the gut microbiota, BAs generally decrease

the integrity of bacterial cell membranes, and increase permeability and cell death⁶⁸. The extent of membrane damage depends largely on BA hydrophobicity: higher hydrophobicity is associated with greater detrimental effects on the bacterial membrane⁶⁹. This finding explains why unconjugated BAs often have stronger antibacterial properties than their conjugated counterparts: conjugation markedly decreases lipophilicity⁶⁸. Notably, BAs also exhibit antimicrobial activity through inducing DNA damage, oxidative and pH stress, and chelation of cellular ions such as calcium. BAs also indirectly exert antimicrobial activity by activating the inhibitory FXR pathway, and subsequently inducing inducible nitric oxide synthase (iNOS) and IL-18, thereby facilitating the maintenance of the integrity of the intestinal barrier^{70,71}.

Interestingly, different bacteria exhibit distinct sensitivity to BAs. For example, bacteria expressing BA-metabolizing enzymes tend to have high resistance to the antimicrobial activity of BAs. Gram-positive bacteria appear to be more sensitive to BAs than gram-negative bacteria. However, specific gram-positive bacteria, including *Bifidobacterium*, *Sporolactobacillus*, *Lactobacillus*, and *Bacillus*, are susceptible to BA-induced toxicity^{72,73}. These findings highlight the intricate interactions between BAs and the gut microbiota, and may have promise in clinical applications.

BA-gut microbiota interactions in HCC

In the landscape of HCC, emerging research has examined the complex network of dysregulated gut microbiota and BAs, and

revealed their effects on HCC development. These insights may illuminate potential avenues for innovative diagnostic, therapeutic, and preventive strategies for HCC.

Dysregulated BAs contribute to HCC development

As early as 1940, DCA, a critical secondary BA generated through microorganism-mediated 7α -dehydroxylation, was found to exhibit oncogenic potential⁷⁴, particularly in HCC associated with obesity⁷⁵, thus providing initial evidence linking BAs to HCC. Further support has been provided by another study showing that DCA administration stimulates hepatic stellate cell (HSC) senescence, and causes these cells to exhibit biological behaviors and phenotypes of liver malignancy⁷⁶. On the basis of these insights, DCA is being explored as a potential therapeutic target for HCC. For instance, microbial products such as butyrate might reverse dysregulated BA profiles, including decreased DCA levels in hepatitis and HCC⁷⁷. Another study has further elucidated this association by revealing a positive correlation between elevated DCA levels and HCC, alongside an abundance of gut bacteria with high BSH activity. Subsequent mechanistic investigations have highlighted the anti-tumor potential of a conjugated DCA form (glycodeoxycholic acid, GDCA), whereby gut bacteria rich in BSH activity deconjugate GDCA to DCA, thus promoting HCC development⁷⁸.

Beyond DCA, other BAs including CDCA derivatives, such as LCA and UDCA, have regulatory roles in HCC⁷⁹. LCA contributes to HCC and cholangiocarcinoma development by dysregulating MAFG in hepatocytes, disrupting lipid homeostasis, and subsequently promoting cholestasis injury^{80,81}. In contrast, UDCA has emerged as a promising therapeutic agent for various chronic liver diseases, and its efficacy in the context of HCC is increasingly being recognized. Clinical observations suggest a potential inverse correlation between UDCA use and the incidence of HCC in HCV-associated liver cirrhosis⁸². Pre-clinical experiments have investigated several mechanisms underlying UDCA's anti-HCC effects, including promotion of apoptosis⁸³, facilitation of autophagy⁸², and inhibition of angiogenesis⁸⁴. Glycoursodeoxycholic acid (GUDCA), a microbial derivative of UDCA, has potent anti-tumor properties. Diminished levels of *Bacteroides fragilis* have been found to lead to increased GUDCA levels, thereby alleviating HCC by activating the FXR/RXR pathway⁸⁵.

Aberrant BA metabolism profoundly influences the tumor immune microenvironment (TIME). BAs involved in inflammatory regulation are associated with carcinogenesis through essential signaling pathways, including the NF κ B, COX-2, and STAT3 pathways, and inflammatory factors, such as IL-6, IL-1 β , and TNF- α ⁸⁶. This altered metabolism hampers the function of CD8+ T cells, avoids the recruitment of natural killer T (NKT) cells, and amplifies the polarization of M2-like tumor-associated macrophages, thereby fostering tumor immune escape and contributing to HCC development. Studies have indicated that gut microorganism-mediated conversion of primary-to-secondary BA regulates CXCL16 expression in liver sinusoidal endothelial cells, thus controlling NKT cell accumulation and mediating liver-selective tumor inhibition⁸⁷. BAs also function as signaling mediators by stimulating nuclear receptors and promoting the polarization of M2-like macrophages, thereby creating an immunosuppressive TIME favoring the growth of tumor-initiating cells⁸⁸. Moreover, the overproduction of DCA by specific *Clostridium* species has been found to facilitate the proliferation of regulatory T cells while inhibiting the accumulation of CD103+ dendritic cells. This mechanism may potentially compromise the anti-tumor function of CD8+ T cells and ultimately accelerate the progression of liver cancer⁸⁹.

Regulation of HCC by BA receptors

Key BA receptors such as FXR and TGR5 govern the interplay between BAs and the gut microbiota. Mice lacking *Fxr* expression exhibit elevated serum and hepatic BAs, and therefore are susceptible to spontaneous hepatocarcinogenesis⁹⁰. Liver-specific *Fxr* knockout mice show changes in the expression of the tumor suppressor p53 and cell cycle regulator cyclin D1; however, these effects are reversed by CA supplementation, which disrupts signaling pathways involved in HCC progression⁹¹. Aberrant BA metabolism has been implicated in modulating the TIME, potentially through the elevation of TGR5 methylation, thereby facilitating tumor immune evasion and fostering HCC development⁹².

Beyond FXR and TGR5, other receptors, such as PXR, CAR, and VDR, participate in HCC regulation. High PXR levels in clinical specimens have been associated with poor prognosis in sorafenib-treated patients with HCC. *In vitro* PXR overexpression has been found to facilitate HCC cell persistence through sorafenib treatment⁹³⁻⁹⁵. The role of CAR in HCC is controversial, on the basis of human and rodent studies. In

rodents, phenobarbital, a CAR activator, has been found to support tumor formation in the liver^{96,97}. However, different studies have found inconsistent effects of CAR in human HCC cell lines^{98,99}. VDR polymorphisms and methylation are associated with susceptibility to HCC^{99,100}. VDR has a protective role in HCC development, possibly through the regulation of liver inflammation and fibrosis¹⁰¹. This intricate network of BA receptors regulates BA metabolism and homeostasis, and influences HCC progression. These receptors, known for their roles in normal BA physiology, are increasingly recognized for their effects on pathological processes in HCC, including liver inflammation, hepatocyte proliferation, and other factors contributing to HCC progression. **Table 2** summarizes the roles of the natural ligands and the potential functions of these receptors in HCC.

Gut microbiota dysbiosis and HCC

The connection between gut microbiota and liver functions enables the microbial community to directly influence hepatic processes, thereby profoundly influencing the course of HCC. The underlying mechanisms linking the gut microbiota and HCC involve a complex interplay among a leaky gut barrier, microbial metabolites, host signaling pathways, and immune responses.

Imbalances between commensal and pro-inflammatory bacteria in dysbiotic microbiota may trigger an inflammatory

response and lead to impaired gut barrier integrity¹¹⁴. Increased intestinal permeability can result in influx of increased bacterial ligands and enterotoxins into the portal vein, and subsequently affect the liver. For example, the microbial products lipopolysaccharides activate immune cells *via* the Toll-like receptor 4 (TLR4) and downstream NF- κ B pathway, thus inducing pro-inflammatory cytokines¹¹⁵, and promoting intestinal inflammation and HCC progression¹¹⁶.

Notably, gut microbial dysbiosis alters metabolite profiles. Key gut metabolites associated with HCC include BAs and short-chain fatty acids, the latter of which have been extensively reviewed elsewhere. Although most BAs act locally and are reabsorbed into the liver, a fraction of BAs circulate systemically and function as signaling molecules by activating nuclear receptors, such as FXR and TGR5. The prominent aforementioned roles of FXR and TGR5 in HCC have been extensively documented^{91,92}, thus underscoring the major role of gut microbiota-mediated BA metabolism in the context of HCC.

Gut microbiota dysbiosis, together with altered metabolite profiles and compromised gut barrier integrity, profoundly disrupts immune homeostasis in the liver and consequently fosters carcinogenesis. Clinical studies have revealed augmented numbers of regulatory T cells and diminished numbers of CD3+ and CD8+ T cells in the peripheral blood and tumor specimens of patients with HCC—findings correlated with poorer prognosis¹¹⁷⁻¹¹⁹. Modulation of the gut microbiota

Table 2 Bile acid receptors and their implications in HCC

Bile acid receptor	Endogenous bile acid ligands	Implication in HCC	References
FXR	CDCA > DCA > LCA > CA	FXR is downregulated in HCC; aged FXR knockout mice spontaneously develop HCC; FXR agonists decrease HCC growth and metastasis in mice	102-105
TGR5	LCA > DCA > CDCA > CA	TGR5 knockout mice show pronounced HCC development induced by DEN; TGR5 mediates JAK2 and STAT3 activity, and MMP, ROCK1, and RhoA expression, thereby promoting the development and migration of HCC	106-108
PXR	3-keto-LCA > LCA	PXR mediates sorafenib resistance in HCC cells	94,109,110
CAR	LCA	CAR supports tumor promotion by phenobarbital in rodent liver; CAR does not affect human HCC cell proliferation; CAR activation in humanized CAR mice does not induce tumor formation in the liver	99,111,112
VDR	LCA > 3-keto-LCA > glyco-LCA > 6-keto-LCA	VDR polymorphisms and methylation are associated with HCC; VDR signaling activation in HSC inhibits liver inflammation and fibrosis, thus supporting HCC	100,101,113

CA, cholic acid; CAR, constitutive androstane receptor; CDCA, chenodeoxycholic acid; DCA, deoxycholic acid; FXR, farnesoid X receptor; LCA, lithocholic acid; PXR, pregnane X receptor; TGR5, Takeda G-protein-coupled receptor 5; VDR, vitamin D receptor.

has potential to reinvigorate anti-tumor responses. As previously described, gut microorganism-mediated conversion of primary-to-secondary BAs regulates NKT cell accumulation⁸⁷, and activated NKT cells secrete interferon (IFN)- γ and TNF, both of which are essential for the anti-tumor response¹²⁰. Moreover, individuals with HCC with enrichment in gut probiotics such as *Bifidobacterium longum* and *Enterococcus hirae*, along with higher levels of reactive CD8+ T cells, have been found to experience prolonged disease-free periods¹²¹. These findings thus underscore the substantial effects of the gut microbiota in modulating anti-tumor effects against HCC.

Impaired interaction between BA metabolism and the gut microbiota in HCC

Given that almost all HCC occurs in the milieu of chronic liver diseases or cirrhosis, and that a dysbiotic gut microbiota is a prominent feature, BA homeostasis may be disrupted¹²². The roles of various specific primary and secondary BAs, as well as their derivatives shaped by the gut microbiota, may potentially exhibit substantial variability in the context of HCC. To provide a comprehensive overview, we have compiled evidence from human cohort studies and animal models regarding the profiles of BAs and gut microbiome signatures (Table 3).

A noteworthy aspect of BA dysregulation by gut microbiota in HCC is the imbalance between non-toxic hydrophilic BAs and toxic hydrophobic BAs¹³⁰. Microbial deconjugation/reconjugation activities in the gut influence the hydrophobic or hydrophilic properties of BAs. HCC animal models have shown an intrahepatic increase in hydrophobic BAs and a decrease in hydrophilic BAs^{128,131}. Typically, hydrophobic BAs, primarily unconjugated BAs (some conjugated BAs, such as glycocholic acid are also hydrophobic), are toxic to hepatocytes. These accumulated BAs activate pro-inflammatory signaling pathways, thus inducing liver damage and fibrosis¹³². Enhancing intestinal excretion of hydrophobic BAs through a diet with 2% cholestyramine has been shown to alleviate HCC progression¹²⁸. Chronic inflammation and fibrosis caused by BA accumulation in turn favor the establishment of an immunosuppressive microenvironment in the liver. Additionally, dysregulated BAs promote HCC development by fostering immunosuppressive M2-like tumor-associated macrophage infiltration⁸⁸ and diminishing numbers of NKT cells⁸⁷. These findings underscore the critical roles of gut microorganisms in shaping the hydrophobic or hydrophilic properties of BAs, and controlling HCC development.

HCC is also associated with cholestasis injury, characterized by the abnormal production or excretion of bile, thus leading to changes in intestinal BA composition and further exacerbation of the dysbiotic gut microbiota. People with cholestasis often exhibit gastrointestinal symptoms and intestinal bacterial overgrowth¹³³. Primary biliary cholangitis (PBC), a classic cholestatic disease, is characterized by diminished microbial diversity. Specifically, certain genera involved in BA transformation, such as *Faecalibacterium*, *Bacteroides*, *Clostridium*, *Lactobacillus*, and *Streptococcus*, exhibit alterations in individuals with PBC. However, these alterations are reversed with UDCA therapy¹³⁴. Other cholestatic diseases, including primary sclerosing cholangitis¹³⁵, intrahepatic cholestasis of pregnancy¹³⁶, and parenteral nutrition-associated cholestasis¹³⁷, have been associated with gut microbiota dysbiosis. These results suggest that BA alterations may lead to increased colonization and survival of gut pathobionts, and that therapeutic approaches targeting the gut microbiota may consequently be effective against disorders associated with dysregulated BA metabolism in HCC.

Therapeutic strategies targeting BAs and the gut microbiota against HCC

The intricate relationship between BAs and the gut microbiota has opened new avenues for therapeutic interventions against HCC. Numerous studies have suggested that targeting BAs and microorganisms may feasibly prevent HCC development, inhibit tumor progression, and improve patient outcomes.

Therapeutic strategies targeting BAs

Given their complexity, the therapeutic potential of many agonists and antagonists in BA metabolism has been investigated, and some have shown great promise in treating various liver diseases, including NASH, PBC, and HCC.

The use of UDCA potentially correlates with diminished incidence of HCC in individuals with hepatitis C virus-associated liver cirrhosis. Notably, the 5-year HCC incidence in individuals receiving UDCA is 17.9%, a percentage significantly lower than the incidence of 39.1% in individuals not receiving UDCA¹³⁸. Obeticholic acid (OCA), a synthetic BA analogue, potently activates FXR¹³⁹. Experimental models of HCC have demonstrated the anti-tumor effects of OCA^{140,141}. Currently, clinical data regarding OCA in HCC are lacking;

Table 3 Selected evidence of associations between the gut microbiota and bile acid profile signatures in patients with HCC and HCC mouse models

Population/animal model	Bile acid profile change	Gut microbiome profile change	References
Human studies			
20 HBV-associated HCC and 15 healthy controls	Patients with HCC vs healthy controls: Serum secondary/primary BA ratio↓ Serum secondary BA↓	Patients with HCC vs healthy controls: <i>Bacteroidales</i> , <i>Lactobacillales</i> , <i>Selenomonadales</i> , <i>Verrucomicrobiales</i> , and <i>Enterobacteriales</i> ↑ <i>Clostridiales</i> , <i>Fusobacteriales</i> , <i>Pasteurellales</i> , and <i>Burkholderiales</i> ↓	78
20 healthy controls, 23 patients with NASH, 11 patients with NASH-cirrhosis, 14 patients with NASH-HCC without cirrhosis, and 19 patients with NASH-HCC with cirrhosis	Serum total BAs, primary BAs, GCA, GCDCA, TCA, and TCDCA negatively associated with <i>Lactobacillus</i> abundance	Patients with NASH-HCC vs NASH: <i>Bacteroidetes</i> and <i>Actinobacteria</i> ↓ <i>Lactobacillus</i> ↓	123
Unresectable patients with HCC receiving immune checkpoint inhibitors: 20 responders, 21 non-responders, and 17 healthy controls	Responders vs non-responders: Fecal UDCA, UCA↑ Fecal UDCA, UCA correlated with the abundance of <i>Lachnospirillum</i>	Responders vs non-responders: <i>Lachnospirillum</i> , <i>Lachnospiraceae</i> , and <i>Veillonella</i> ↑ <i>Prevotella</i> 9↓	124
Animal studies			
DEN-induced mouse HCC model	HCC vs control group: Serum secondary/primary BA ratio↓ Serum secondary BA↓	HCC vs control mice: <i>Bifidobacteriales</i> , <i>Lactobacillales</i> , <i>Bacteroidales</i> , and <i>Clostridiales</i> ↓	78
HFD-induced spontaneous mouse NASH-HCC model	NASH-HCC mice vs control group: Total BA from plasma, feces, and liver ↑	NASH-HCC mice vs control mice: <i>Bacteroides</i> and <i>Clostridium cluster XVIII</i> ↑ <i>Streptococcus</i> , <i>Bifidobacterium</i> , and <i>Prevotella</i> ↓	125
Orthotopic transplantation HCC model in nude mice alleviated by XYXD treatment	XYXD treated group vs control group: Primary BAs, including CA, TαMCA, TβMCA, and TCATCA↑ Secondary BAs, including GUDCA, LCA, TLCA, and ILCA↓	XYXD treated group vs control group: <i>Lactobacillus</i> , <i>Bacteroides</i> , and <i>Prevotella</i> ↑ <i>Eubacterium</i> ↓	126
DEN-induced HCC in a T2DM mouse model alleviated by <i>Lactobacillus brevis</i> treatment	<i>Lactobacillus brevis</i> treated group vs control group: Serum total BA↓	<i>Lactobacillus brevis</i> treated group vs control group: <i>Actinomycetes</i> , <i>Alistipes</i> , <i>Bacteroides</i> , <i>Desulfovibrio</i> , <i>Dubosiella</i> , and <i>Firmicus</i> ↑	127
STZ-HFD induced NASH-HCC mouse model	NASH-HCC vs control mice: Hepatic TCA, GCA and TCDCA↑ Plasma TCA, GCA and TCDCA↑	NASH-HCC vs control mice: <i>Clostridium</i> , <i>Bacteroides</i> , and <i>Desulfovibrio</i> ↑ <i>Paasutterella</i> and <i>Akkermansia</i> ↓	128
DEN-induced rat HCC model	HCC vs control group: Fecal CA↑	HCC vs control group: <i>Ruminococcaceae UCG-004</i> ↑ <i>Lachnospiraceae incertae sedis</i> , <i>Prevotella</i> 9, and <i>Prevotellaceae</i> ↓	129
DEN-induced rat HCC model alleviated by celastrol	Celastrol alleviated DEN-induced HCC, and increased hepatic GCDCA, UDCA, TUDCA, and GUDCA	HCC vs control group: <i>Bacteroides fragilis</i> , <i>Bacteroides finegoldii</i> , <i>Bacteroides massiliensis</i> , and <i>Bacteroides uniformis</i> ↑ Celastrol alleviated DEN-induced HCC, and decreased <i>Bacteroides fragilis</i>	85

HBV, hepatitis B virus; HCC, hepatocellular carcinoma; GCA, glycochenodeoxycholic acid; GCDCA, glycochenodeoxycholic acid; TCA, taurocholic acid; TCDCA, taurochenodeoxycholic acid; NASH, non-alcoholic steatohepatitis; UDCA, ursodeoxycholic acid; DEN, diethylnitrosamine; HFD, high-fat diet; XYXD, xiayuxue decoction; CA, cholic acid; TαMCA, tauro-α-muricholic acid; TβMCA, tauro-β-muricholic acid; TCATCA, taurocholic acid; GUDCA, glyoursodeoxycholic acid; LCA, lithocholic acid; TLCA, tauroolithocholic acid; TUDCA, taoursodeoxycholic acid; ILCA, iso-lithocholic acid; STZ, streptozotocin; UCA, ursocholic acid.

however, multiple clinical trials have hinted at its therapeutic effectiveness in treating chronic liver diseases, such as NASH. For instance, in a phase 3 clinical trial, histological fibrosis was ameliorated in 12% of patients in the placebo group compared with 23% in the OCA 25 mg group ($P = 0.0002$)¹⁴². Furthermore, another study investigating the long-term effectiveness of OCA in patients with PBC has demonstrated favorable outcomes observed over a 3-year follow-up period. Notably a significant change was observed in ALP concentrations from baseline [105.2 U/L (SD 87.6)] to 3 years [95.6 U/L (SD121.1)]; $P < 0.0001$ ¹⁴³.

As previously discussed, FXR is a promising therapeutic target for various liver diseases and potentially HCC. Beyond OCA, several other FXR agonists have been developed for management of liver diseases. However, clinical data supporting the effectiveness of various FXR agonists specifically in the treatment of HCC are currently scarce, although these agonists have shown promising potential in managing diverse liver conditions. For instance, in studies focused on NASH, MET409, a synthetic FXR agonist, has shown considerable effects by decreasing liver fat content over a 12-week period in patients with NASH. That study reported mean relative decreases of 55% (80 mg) and 38% (50 mg), as compared with a 6% decrease in the placebo group ($P < 0.001$)¹⁴⁴. Furthermore, other FXR agonists such as nonsteroidal agonists, vonafexor, and tropifexor have shown effects against liver diseases in phase 2 clinical trials¹⁴⁵⁻¹⁴⁷. These findings suggest that FXR agonists may have roles in maintaining liver homeostasis and may potentially serve as alternative therapeutic targets for treating HCC.

Analogues of FGF19, downstream of BA-induced FXR activation, are also being extensively investigated for their therapeutic potential against NASH^{148,149} and cholestatic liver diseases¹⁵⁰. In animal models of HCC, FGF19 analogues have shown anti-fibrosis and anti-tumor effects by suppressing hepatic BA synthesis¹⁵¹. Currently, several ongoing phase 1/2 trials are studying the efficacy of targeting FGF19 in patients with HCC. In particular, FGF401, a potent and selective FGF19-FGFR4 signaling inhibitor, has shown promising results: either FGF401 monotherapy or combined therapy with spartalizumab has shown safety in patients with FGFR4/KLB-positive tumors, including HCC. Preliminary clinical efficacy has also been observed¹⁵².

In contrast, BA pathway antagonists, particularly inhibitors of BA transporters (e.g., ASBT), have also shown therapeutic potential in preclinical studies. ASBT inhibition increases

colonic BA accumulation and decreases the BA pool, thus potentially alleviating liver pathogenesis in cholestatic liver disease and NASH animal models¹⁵³. Similarly, another study has demonstrated the ability of ASBT inhibitors, such as SC-435 and A4250, to modulate BA metabolism and ameliorate liver histology in mice with sclerosing cholangitis^{154,155}. In humans, the ASBT inhibitor odeixibat has shown promising results in alleviating pruritus symptoms and decreasing serum BA in a phase 3 trial of cholestatic liver diseases¹⁵⁶. In contrast, the use of an ASBT inhibitor SHP626 (volixibat) has shown limited therapeutic benefits in patients with NASH in a phase 2 trial¹⁵⁷.

Therapeutic strategies targeting microorganism-mediated BA metabolism

Probiotics, prebiotics, and diet

Probiotics are live microorganisms that offer a wide range of health benefits after being consumed. For instance, a mouse study has demonstrated the prophylactic effect of *Lactobacillus rhamnosus GG* against liver fibrosis by inhibiting hepatic BA synthesis and enhancing BA excretion¹²². *Lactobacillus eosinophil* has potential in alleviating mouse NAFLD through modulating the microorganism-mediated FXR/FGF15 signaling pathway¹⁵⁸. Similarly, *Lactobacillus brevis* has been found to alleviate HCC progression in mice *via* influencing the interplay among the gut microbiota, BAs, and NOTCH1 signaling¹²⁷. Given that probiotics modulate BA metabolism, several clinical trials have investigated the potential of using probiotics, particularly various *Lactobacillus* strains, in the treatment of liver diseases. For instance, the effects of a mixture containing 3 *Lactobacillus plantarum* strains on the BA profile, plasma lipids, and other associated biomarkers is being evaluated through a dose-dependent regimen in a cohort of overweight participants (NCT05378230).

Beyond probiotics, prebiotics—substrates that stimulate the growth and activity of for beneficial gut bacteria—also confer health benefits in humans. In general, prebiotics are more advantageous in managing metabolic conditions than probiotics and specific BA agonists or antagonists, because they stimulate BA production and activate associated receptors in a more natural, controlled manner¹⁵⁹. These characteristics makes prebiotics preferable candidates for alleviating metabolic conditions without inducing serious adverse effects. In addition, emerging research highlights the influence of diet on cancer development¹⁶⁰⁻¹⁶². Personalized dietary interventions

tailored to the gut microbiota in each individual could therefore be feasible for preventing chronic liver diseases and HCC. By optimizing the diet to promote the growth of beneficial commensal organisms and modulate BA metabolism, personalized dietary interventions may restore intestinal homeostasis and support liver health¹⁶³. A compelling example is Pu-erh tea, which has been shown to decrease BSH enriched bacteria and BSH activity. This modulation of BA metabolism, characterized by the accumulation of conjugated BAs in the ileum, attenuates hypercholesterolemia and might have potential as a dietary intervention influencing HCC risk¹⁶⁴.

Fecal microbiota transplantation (FMT)

FMT refers to the transfer of fecal material from a healthy donor to a recipient to restore a balanced gut microbiota¹⁶⁵. FMT is currently clinically approved for treating *Clostridium difficile* infection¹⁶⁶, and has shown promising potential against various intestinal and extra-intestinal diseases^{167,168}. FMT modulates both the BA profile and the gut microbiota. In a recent clinical trial, researchers investigating the effects of oral capsule FMT have observed enhanced microorganism-mediated BA metabolism: specifically, the study reported significant decreases in stool TCA levels in patients receiving FMT with respect to their baselines. Moreover, the BA profiles of FMT recipients began to resemble those of the donors¹⁶⁹. Another clinical trial in obese patients has revealed that, after FMT, *Bacteroides ovatus* and *Phocaeicola dorei* are positively correlated with unconjugated BAs, whereas *Bifidobacterium adolescentis*, *Collinsella aerofaciens*, and *Faecalibacterium prausnitzii* are positively correlated with secondary BAs¹⁷⁰. These promising results have led to further ongoing clinical trials aimed at exploring the application of FMT in HCC. For instance, NCT05750030 is a phase IIa pilot study testing the safety and efficacy of combining FMT with atezolizumab plus bevacizumab in patients who previously did not respond to immunotherapy for advanced HCC. Similarly, NCT05690048 is investigating whether FMT might overcome resistance to atezolizumab/bevacizumab in the context of HCC. Despite limited evidence regarding its specific efficacy in HCC, FMT has shown potential in enhancing liver homeostasis and influencing the hepatic immune microenvironment. However, further comprehensive studies are warranted to ascertain its effectiveness and safety specifically in the treatment of HCC.

Engineered bacteria

Advances in genetic engineering have opened exciting possibilities to create specialized bacteria with therapeutic properties.

Bacteria such as *E. coli* and *Lactococcus lactis* have been used as vehicles for delivering therapeutic recombinant proteins, yet their abilities to persist and survive in the human gut are limited. Engineered bacterial strains are thus required to serve as more effective tools. For example, a study has successfully modified a strain of *Clostridium sporogenes* to heterologously express genes from *Clostridium scindens* responsible for BA conversion; this modification enabled the recombinant bacteria to synthesize DCA and LCA⁴⁸. Hence, developing engineered bacteria with enhanced ability to modulate BA metabolism and the gut microbiota may be feasible to treat liver diseases including HCC.

Immunotherapy

Immunotherapy has pioneered a new treatment paradigm in HCC, setting a novel therapeutic benchmark. In particular, the FDA approval of the combination of atezolizumab (anti-PD1) and bevacizumab (anti-VEGF), the first-line treatment for advanced HCC, marked a major milestone¹⁷¹. Studies increasingly indicate that the interplay between the gut microbiota and the liver influences the tumor microenvironment in HCC, thus resulting in varied responses to immunotherapy^{172,173}. Notably, differences in gut microbiota composition have been observed between patients with HCC who are responders and non-responders to immunotherapy¹⁷⁴, thus suggesting a potential role of microorganisms as prognostic biomarkers for immunotherapy efficacy¹⁷⁵. Furthermore, several bacterial strains stimulate anti-tumor responses to immune checkpoint inhibitors¹⁷⁶⁻¹⁸⁰. Moreover, BA profiles can be used to distinguish responders and non-responders to immunotherapy in HCC. In responders, elevated levels of secondary BAs, including UDCA, TUDCA, UCA, and MDCA, have been found to be accompanied by increases in *Lachnoclostridium*, *Lachnospiraceae*, and *Veillonella*, and a decrease in *Prevotella 9*¹²⁴. Gut microorganisms associated with both TIME and BA metabolism have shown good performance in discriminating 5-year survival (AUC 81%)¹⁸¹. These findings thus underscore the potential of research on liver cancer microbiota in immunotherapy to identify novel therapeutic approaches for managing HCC.

In summary, the diverse therapeutic strategies addressing the intricate interplay between dysregulated gut microbiota and the BA axis have promise in combating HCC. Animal experiments have demonstrated the potential therapeutic efficacy of various microbiota-targeting strategies, such as

probiotics, prebiotics, dietary intervention, and FMT in managing HCC. Preclinical studies on *Lactobacillus* strains have indicated their ability to inhibit liver fibrosis and slow HCC progression. Prebiotics and dietary interventions, which are preferred because of their controlled and natural effects on BA metabolism, have promise in managing liver diseases without causing significant adverse effects. Although combining FMT with immunotherapy is safer and may improve efficacy beyond

that of monotherapy in various cancer types¹⁸², its efficacy and safety in HCC treatment require further validation in human clinical trials. **Figure 2** provides a visual representation of the effects of dysregulated gut microbiota on the BA axis and their contributions to HCC, along with potential therapeutic interventions. Further research and clinical trials are crucial to fully realize the potential of these strategies in the treatment and prevention of HCC.

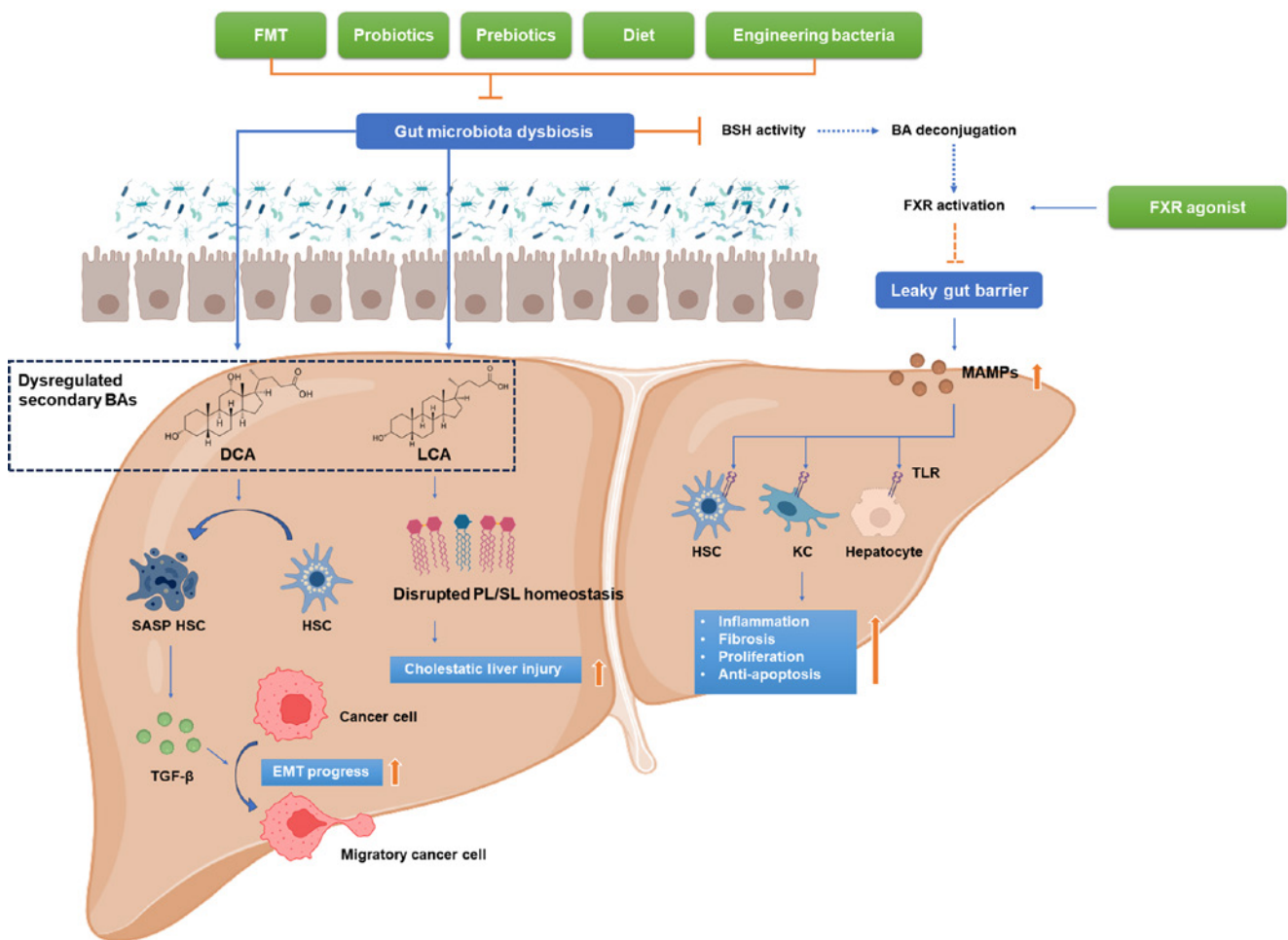


Figure 2 Selected evidence of dysregulation of the gut microbiota-bile acid axis contributes to hepatocellular carcinoma. Dysbiosis of the gut microbiota in HCC contributes to the dysregulation of secondary BA production. DCA induces SASP within quiescent HSCs, thus increasing secretion of cytokines including TGF- β , promoting EMT in cancer cells, and enhancing metastasis. LCA disrupts phospholipid/sphingolipid homeostasis and exacerbates cholestatic liver injury. Dysbiosis also impairs BSH activity, decreases primary BA deconjugation and FXR activation, and results in a compromised gut barrier. With impaired gut barrier function, MAMPs translocate to the liver via the portal vein and activate TLR on HSCs, KCs, and hepatocytes. This cascade intensifies liver inflammation, fibrosis, cell proliferation, and anti-apoptosis, thus facilitating hepatocarcinogenesis. Therapies, including FMT, probiotics, prebiotics, dietary intervention, and engineered bacteria, may alleviate gut microbiota dysbiosis as adjunctive treatments for HCC. Additionally, FXR agonists may improve gut barrier function, thereby decreasing MAMP toxicity and exerting anti-HCC effects. BAs, bile acids; BSH, bile salt hydrolase; DCA, deoxycholic acid; FMT, fecal microbiota transplantation; FXR, farnesoid X receptor; HSC, hepatic stellate cells; KC, Kupffer cells; LCA, lithocholic acid; MAMP, microorganism-associated molecular pattern; PL, phospholipids; SASP, senescence-associated secretory phenotype; SL, sphingolipids. Figure generated with BioRender.com.

Conclusions and future perspectives

The intricate interplay among BAs, gut microbiota, and HCC has illuminated a novel pathway for understanding the complex pathogenesis of this malignancy and exploring innovative therapeutic approaches. The effects of the gut microbiota on BA metabolism, and vice versa, requires deeper exploration of how microbial dysbiosis contributes to HCC initiation and progression.

With the emergence of new analytical tools and strategies, including metagenomics and metabolomics, insights are rapidly being gained into the metabolic interactions between the gut and liver, as well as the signaling pathways that regulate HCC development. The modulation of the gut microbiota and BA profiles is a novel approach to treating HCC. This strategy has immense promise and clearly defines the next frontier in HCC research. Research efforts should focus on: 1) elucidating the crosstalk among the gut microbiota, BAs, and host immune responses in HCC, particularly how microbial dysbiosis and BA dysregulation contribute to chronic inflammation and immune evasion, which are hallmarks of cancer progression; 2) investigating the role of the gut microbiota and BA in modulating the efficacy and adverse effects of existing HCC treatments, such as chemotherapy, immunotherapy, and targeted therapy; 3) exploring the potential of using engineered probiotics to target and modulate BA pathways for HCC prevention and treatment; and 4) understanding the effects of lifestyle factors, such as diet and exercise, on the liver-BA-gut microbiota axis and its relevance to HCC risk and progression.

Several limitations warrant consideration. The translation of preclinical findings into effective clinical therapies presents a substantial challenge. Clinical trials are required to validate the efficacy, safety, and long-term effects of therapeutic strategies targeting BAs and the gut microbiota in HCC prevention and treatment. Furthermore, the microbiome is highly individualized, and is influenced by genetics, diet, environment, and other factors. Therefore, the development of personalized interventions based on an individual's unique microbiome composition and BA metabolism would require careful consideration and validation. Moreover, the liver-BA-gut microbiota axis is a dynamic entity with intricately intertwined individual components. To effectively intervene in HCC and achieve meaningful therapeutic outcomes, comprehensive treatments must address all these interconnected components collectively. Finally, ethical considerations surrounding FMT and engineered bacteria therapies must be carefully addressed.

In conclusion, the relationship between BAs and the gut microbiota in HCC is an exciting and evolving field of research. By targeting these pathways, novel therapeutic approaches may be uncovered to halt or slow HCC progression, and ultimately improve prognosis and quality of life for patients with HCC. Continued exploration of the BA-gut microbiota interaction in HCC is expected to yield valuable insights that will guide the development of innovative therapies in the fight against this deadly disease.

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Conflicts of interest statement

No potential conflicts of interest are disclosed.

Author contributions

Yang Song: conceptualization, literature review, and writing the manuscript.

Harry CH Lau: revision of the manuscript.

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References

1. Kanwal F, Singal AG. Surveillance for hepatocellular carcinoma: current best practice and future direction. *Gastroenterology*. 2019; 157: 54-64.
2. Yao C, Wu S, Kong J, Sun Y, Bai Y, Zhu R, et al. Angiogenesis in hepatocellular carcinoma: mechanisms and anti-angiogenic therapies. *Cancer Biol Med*. 2023; 20: 25-43.
3. McGlynn KA, Petrick JL, El-Serag HB. Epidemiology of hepatocellular carcinoma. *Hepatology*. 2021; 73 Suppl 1: 4-13.
4. Toh MR, Wong EYT, Wong SH, Ng AWT, Loo LH, Chow PK, et al. Global epidemiology and genetics of hepatocellular carcinoma. *Gastroenterology*. 2023; 164: 766-82.
5. Kapitan M, Niemiec MJ, Steimle A, Frick JS, Jacobsen ID. Fungi as part of the microbiota and interactions with intestinal bacteria. *Curr Top Microbiol Immunol*. 2019; 422: 265-301.

6. Zhao L, Shi Y, Lau HC, Liu W, Luo G, Wang G, et al. Uncovering 1058 novel human enteric DNA viruses through deep long-read third-generation sequencing and their clinical impact. *Gastroenterology*. 2022; 163: 699-711.
7. Milani C, Duranti S, Bottacini F, Casey E, Turroni F, Mahony J, et al. The first microbial colonizers of the human gut: composition, activities, and health implications of the infant gut microbiota. *Microbiol Mol Biol Rev*. 2017; 81: e00036-17.
8. Tomioka S, Seki N, Sugiura Y, Akiyama M, Uchiyama J, Yamaguchi G, et al. Cooperative action of gut-microbiota-accessible carbohydrates improves host metabolic function. *Cell Rep*. 2022; 40: 111087.
9. Ellis JL, Karl JP, Oliverio AM, Fu X, Soares JW, Wolfe BE, et al. Dietary vitamin K is remodeled by gut microbiota and influences community composition. *Gut Microbes*. 2021; 13: 1-16.
10. Makki K, Deehan EC, Walter J, Backhed F. The impact of dietary fiber on gut microbiota in host health and disease. *Cell Host Microbe*. 2018; 23: 705-15.
11. Wastyk HC, Fragiadakis GK, Perelman D, Dahan D, Merrill BD, Yu FB, et al. Gut-microbiota-targeted diets modulate human immune status. *Cell*. 2021; 184: 4137-53.e4114.
12. Morais LH, Schreiber HL, Mazmanian SK. The gut microbiota-brain axis in behaviour and brain disorders. *Nat Rev Microbiol*. 2021; 19: 241-55.
13. Chambers KF, Day PE, Aboufarrag HT, Kroon PA. Polyphenol effects on cholesterol metabolism via bile acid biosynthesis, CYP7A1: a review. *Nutrients*. 2019; 11: 2588.
14. Fan L, Joseph JF, Durairaj P, Parr MK, Bureik M. Conversion of chenodeoxycholic acid to cholic acid by human CYP8B1. *Biol Chem*. 2019; 400: 625-8.
15. Jia W, Wei M, Rajani C, Zheng X. Targeting the alternative bile acid synthetic pathway for metabolic diseases. *Protein Cell*. 2021; 12: 411-25.
16. Appelman MD, Wettengel JM, Protzer U, Oude Elferink RPJ, van de Graaf SFJ. Molecular regulation of the hepatic bile acid uptake transporter and HBV entry receptor NTCP. *Biochim Biophys Acta Mol Cell Biol Lipids*. 2021; 1866: 158960.
17. Lu X, Liu L, Shan W, Kong L, Chen N, Lou Y, et al. The role of the sodium-taurocholate co-transporting polypeptide (NTCP) and Bile Salt Export Pump (BSEP) in related liver disease. *Curr Drug Metab*. 2019; 20: 377-89.
18. Park JH, Iwamoto M, Yun JH, Uchikubo-Kamo T, Son D, Jin Z, et al. Structural insights into the HBV receptor and bile acid transporter NTCP. *Nature*. 2022; 606: 1027-31.
19. Al-Aqil FA, Monte MJ, Peleteiro-Vigil A, Briz O, Rosales R, Gonzalez R, et al. Interaction of glucocorticoids with FXR/FGF19/FGF21-mediated ileum-liver crosstalk. *Biochim Biophys Acta Mol Basis Dis*. 2018; 1864: 2927-37.
20. Yang H, Luo F, Wei Y, Jiao Y, Qian J, Chen S, et al. TGR5 protects against cholestatic liver disease via suppressing the NF-kappaB pathway and activating the Nrf2/HO-1 pathway. *Ann Transl Med*. 2021; 9: 1158.
21. Guo C, Chen WD, Wang YD. TGR5, not only a metabolic regulator. *Front Physiol*. 2016; 7: 646.
22. Oteng AB, Liu L. GPCR-mediated effects of fatty acids and bile acids on glucose homeostasis. *Front Endocrinol (Lausanne)*. 2023; 14: 1206063.
23. Donepudi AC, Boehme S, Li F, Chiang JY. G-protein-coupled bile acid receptor plays a key role in bile acid metabolism and fasting-induced hepatic steatosis in mice. *Hepatology*. 2017; 65: 813-27.
24. Cai X, Young GM, Xie W. The xenobiotic receptors PXR and CAR in liver physiology, an update. *Biochim Biophys Acta Mol Basis Dis*. 2021; 1867: 166101.
25. Chen ML, Huang X, Wang H, Hegner C, Liu Y, Shang J, et al. CAR directs T cell adaptation to bile acids in the small intestine. *Nature*. 2021; 593: 147-51.
26. Bhutia YD, Ogura J, Sivaprakasam S, Ganapathy V. Gut microbiome and colon cancer: role of bacterial metabolites and their molecular targets in the host. *Curr Colorectal Cancer Rep*. 2017; 13: 111-8.
27. Thibaut MM, Bindels LB. Crosstalk between bile acid-activated receptors and microbiome in entero-hepatic inflammation. *Trends Mol Med*. 2022; 28: 223-36.
28. Sun M, Cui W, Woody SK, Staudinger JL. Pregnane X receptor modulates the inflammatory response in primary cultures of hepatocytes. *Drug Metab Dispos*. 2015; 43: 335-43.
29. Shin DJ, Wang L. Bile acid-activated receptors: a review on FXR and other nuclear receptors. *Handb Exp Pharmacol*. 2019; 256: 51-72.
30. Flannigan KL, Nieves KM, Szczepanski HE, Serra A, Lee JW, Alston LA, et al. The pregnane X receptor and indole-3-propionic acid shape the intestinal mesenchyme to restrain inflammation and fibrosis. *Cell Mol Gastroenterol Hepatol*. 2023; 15: 765-95.
31. Garcia M, Thirouard L, Sedes L, Monrose M, Holota H, Caira F, et al. Nuclear receptor metabolism of bile acids and xenobiotics: a coordinated detoxification system with impact on health and diseases. *Int J Mol Sci*. 2018; 19: 3630.
32. Poudel S, Huber AD, Chen T. Regulation of nuclear receptors PXR and CAR by small molecules and signal crosstalk: roles in drug metabolism and beyond. *Drug Metab Dispos*. 2023; 51: 228-36.
33. Sasaki H, Masuno H, Kawasaki H, Yoshihara A, Numoto N, Ito N, et al. Lithocholic acid derivatives as potent Vitamin D receptor agonists. *J Med Chem*. 2021; 64: 516-26.
34. Chow EC, Magomedova L, Quach HP, Patel R, Durk MR, Fan J, et al. Vitamin D receptor activation down-regulates the small heterodimer partner and increases CYP7A1 to lower cholesterol. *Gastroenterology*. 2014; 146: 1048-59.
35. Nishida S, Ishizawa M, Kato S, Makishima M. Vitamin D receptor deletion changes bile acid composition in mice orally administered chenodeoxycholic acid. *J Nutr Sci Vitaminol (Tokyo)*. 2020; 66: 370-4.
36. Boyer JL. OSTalpha-OSTbeta guards the ileal enterocyte from the accumulation of toxic levels of bile acids. *Cell Mol Gastroenterol Hepatol*. 2018; 5: 649-50.
37. Liu J, Song Y, Wang Y, Hong H. Vitamin D/vitamin D receptor pathway in non-alcoholic fatty liver disease. *Expert Opin Ther Targets*. 2023: 1-13.
38. Lucas LN, Barrett K, Kerby RL, Zhang Q, Cattaneo LE, Stevenson D, et al. Dominant bacterial phyla from the human gut show

- widespread ability to transform and conjugate bile acids. *mSystems*. 2021; e0080521.
39. Sun H, Guo Y, Wang H, Yin A, Hu J, Yuan T, et al. Gut commensal *Parabacteroides distasonis* alleviates inflammatory arthritis. *Gut*. 2023; 72: 1664-77.
 40. Ruiz L, Sanchez B, Margolles A. Determination of bile salt hydrolase activity in bifidobacteria. *Methods Mol Biol*. 2021; 2278: 149-55.
 41. Foley MH, O'Flaherty S, Allen G, Rivera AJ, Stewart AK, Barrangou R, et al. Lactobacillus bile salt hydrolase substrate specificity governs bacterial fitness and host colonization. *Proc Natl Acad Sci U S A*. 2021; 118.
 42. Chand D, Panigrahi P, Varshney N, Ramasamy S, Suresh CG. Structure and function of a highly active Bile Salt Hydrolase (BSH) from *Enterococcus faecalis* and post-translational processing of BSH enzymes. *Biochim Biophys Acta Proteins Proteom*. 2018; 1866: 507-18.
 43. Yoon S, Yu J, McDowell A, Kim SH, You HJ, Ko G. Bile salt hydrolase-mediated inhibitory effect of *Bacteroides ovatus* on growth of *Clostridium difficile*. *J Microbiol*. 2017; 55: 892-9.
 44. Geng W, Lin J. Bacterial bile salt hydrolase: an intestinal microbiome target for enhanced animal health. *Anim Health Res Rev*. 2016; 17: 148-58.
 45. Marchesini MI, Connolly J, Delpino MV, Baldi PC, Mujer CV, DelVecchio VG, et al. *Brucella abortus* choloylglycine hydrolase affects cell envelope composition and host cell internalization. *PLoS One*. 2011; 6: e28480.
 46. Ridlon JM, Devendran S, Alves JM, Doden H, Wolf PG, Pereira GV, et al. The 'in vivo lifestyle' of bile acid 7 α -dehydroxylating bacteria: comparative genomics, metatranscriptomic, and bile acid metabolomics analysis of a defined microbial community in gnotobiotic mice. *Gut Microbes*. 2020; 11: 381-404.
 47. Chinda D, Takada T, Mikami T, Shimizu K, Oana K, Arai T, et al. Spatial distribution of live gut microbiota and bile acid metabolism in various parts of human large intestine. *Sci Rep*. 2022; 12: 3593.
 48. Funabashi M, Grove TL, Wang M, Varma Y, McFadden ME, Brown LC, et al. A metabolic pathway for bile acid dehydroxylation by the gut microbiome. *Nature*. 2020; 582: 566-70.
 49. Vital M, Rud T, Rath S, Pieper DH, Schluter D. Diversity of bacteria exhibiting bile acid-inducible 7 α -dehydroxylation genes in the human gut. *Comput Struct Biotechnol J*. 2019; 17: 1016-9.
 50. Guzior DV, Quinn RA. Review: microbial transformations of human bile acids. *Microbiome*. 2021; 9: 140.
 51. Tonin F, Otten LG, Arends I. NAD(+)-dependent enzymatic route for the epimerization of hydroxysteroids. *ChemSusChem*. 2019; 12: 3192-203.
 52. Zhang X, Fan D, Hua X, Zhang T. Large-scale production of ursodeoxycholic acid from chenodeoxycholic acid by engineering 7 α - and 7 β -hydroxysteroid dehydrogenase. *Bioprocess Biosyst Eng*. 2019; 42: 1537-45.
 53. Tonin F, Arends I. Latest development in the synthesis of ursodeoxycholic acid (UDCA): a critical review. *Beilstein J Org Chem*. 2018; 14: 470-83.
 54. Ridlon JM, Harris SC, Bhowmik S, Kang DJ, Hylemon PB. Consequences of bile salt biotransformations by intestinal bacteria. *Gut Microbes*. 2016; 7: 22-39.
 55. Song P, Zhang X, Feng W, Xu W, Wu C, Xie S, et al. Biological synthesis of ursodeoxycholic acid. *Front Microbiol*. 2023; 14: 1140662.
 56. Lou D, Tan J, Zhu L, Ji S, Tang S, Yao K, et al. Engineering *Clostridium absonum* 7 α -hydroxysteroid dehydrogenase for enhancing thermostability based on flexible site and $\Delta\Delta G$ prediction. *Protein Pept Lett*. 2018; 25: 230-5.
 57. Ferrandi EE, Bertolesi GM, Polentini F, Negri A, Riva S, Monti D. In search of sustainable chemical processes: cloning, recombinant expression, and functional characterization of the 7 α - and 7 β -hydroxysteroid dehydrogenases from *Clostridium absonum*. *Appl Microbiol Biotechnol*. 2012; 95: 1221-33.
 58. Cifuentes-Silva E, Cabello-Verrugio C. Bile acids as signaling molecules: role of ursodeoxycholic acid in cholestatic liver disease. *Curr Protein Pept Sci*. 2023.
 59. Achufusi TGO, Safadi AO, Mahabadi N. Ursodeoxycholic acid. In: *StatPearls*. Treasure Island, FL: StatPearls Publishing; 2023.
 60. Laschtowitz A, de Veer RC, Van der Meer AJ, Schramm C. Diagnosis and treatment of primary biliary cholangitis. *United European Gastroenterol J*. 2020; 8: 667-74.
 61. Camilleri M. Bile acid detergency: permeability, inflammation, and effects of sulfation. *Am J Physiol Gastrointest Liver Physiol*. 2022; 322: G480-8.
 62. Huang S, Pang D, Li X, You L, Zhao Z, Cheung PC, et al. A sulfated polysaccharide from *Gracilaria Lemaneiformis* regulates cholesterol and bile acid metabolism in high-fat diet mice. *Food Funct*. 2019; 10: 3224-36.
 63. Quinn RA, Melnik AV, Vrbancac A, Fu T, Patras KA, Christy MP, et al. Global chemical effects of the microbiome include new bile-acid conjugations. *Nature*. 2020; 579: 123-9.
 64. Smirnova E, Muthiah MD, Narayan N, Siddiqui MS, Puri P, Luketic VA, et al. Metabolic reprogramming of the intestinal microbiome with functional bile acid changes underlie the development of NAFLD. *Hepatology*. 2022; 76: 1811-24.
 65. Lei Y, Tang L, Chen Q, Wu L, He W, Tu D, et al. Disulfiram ameliorates nonalcoholic steatohepatitis by modulating the gut microbiota and bile acid metabolism. *Nat Commun*. 2022; 13: 6862.
 66. Kuang J, Wang J, Li Y, Li M, Zhao M, Ge K, et al. Hyodeoxycholic acid alleviates non-alcoholic fatty liver disease through modulating the gut-liver axis. *Cell Metab*. 2023; 35: 1752-66.e1758.
 67. Liu Y, Kang W, Liu S, Li J, Liu J, Chen X, et al. Gut microbiota-bile acid-intestinal Farnesoid X receptor signaling axis orchestrates cadmium-induced liver injury. *Sci Total Environ*. 2022; 849: 157861.
 68. Sannasiddappa TH, Lund PA, Clarke SR. In vitro antibacterial activity of unconjugated and conjugated bile salts on *Staphylococcus aureus*. *Front Microbiol*. 2017; 8: 1581.
 69. Gupta S, Arora A, Saini V, Mehta D, Khan MZ, Mishra DK, et al. Hydrophobicity of Cholic acid-derived amphiphiles dictates the antimicrobial specificity. *ACS Biomater Sci Eng*. 2022; 8: 4996-5007.

70. Tie Y, Huang Y, Chen R, Li L, Chen M, Zhang S. Current insights on the roles of gut microbiota in inflammatory bowel disease-associated extra-intestinal manifestations: pathophysiology and therapeutic targets. *Gut Microbes*. 2023; 15: 2265028.
71. Kisthardt SC, Thanissery R, Pike CM, Foley MH, Theriot CM. The microbial-derived bile acid lithocholate and its epimers inhibit *Clostridioides difficile* growth and pathogenicity while sparing members of the gut microbiota. *J Bacteriol*. 2023; 205: e0018023.
72. Schopping M, Zeidan AA, Franzen CJ. Stress response in bifidobacteria. *Microbiol Mol Biol Rev*. 2022; 86: e0017021.
73. Ruiz L, Margolles A, Sanchez B. Bile resistance mechanisms in *Lactobacillus* and *Bifidobacterium*. *Front Microbiol*. 2013; 4: 396.
74. Cook JW, Kennaway EL, Kennaway NM. Production of tumours in mice by deoxycholic acid. *Nature*. 1940; 145: 627.
75. Yoshimoto S, Loo TM, Atarashi K, Kanda H, Sato S, Oyadomari S, et al. Obesity-induced gut microbial metabolite promotes liver cancer through senescence secretome. *Nature*. 2013; 499: 97-101.
76. Nguyen PT, Kanno K, Pham QT, Kikuchi Y, Kakimoto M, Kobayashi T, et al. Senescent hepatic stellate cells caused by deoxycholic acid modulates malignant behavior of hepatocellular carcinoma. *J Cancer Res Clin Oncol*. 2020; 146: 3255-68.
77. Sheng L, Jena PK, Hu Y, Liu HX, Nagar N, Kalanetra KM, et al. Hepatic inflammation caused by dysregulated bile acid synthesis is reversible by butyrate supplementation. *J Pathol*. 2017; 243: 431-41.
78. Shen R, Ke L, Li Q, Dang X, Shen S, Shen J, et al. Abnormal bile acid-microbiota crosstalk promotes the development of hepatocellular carcinoma. *Hepatol Int*. 2022; 16: 396-411.
79. Liu T, Yang H, Fan W, Tu J, Li TWH, Wang J, et al. Mechanisms of MAFG Dysregulation in cholestatic liver injury and development of liver cancer. *Gastroenterology*. 2018; 155: 557-71.e514.
80. Han JC, Yu J, Gao YJ. Lipidomics investigation of reversal effect of glycyrrhizin (GL) towards lithocholic acid (LCA)-induced alteration of phospholipid profiles. *Pharm Biol*. 2014; 52: 1624-8.
81. Matsubara T, Tanaka N, Patterson AD, Cho JY, Krausz KW, Gonzalez FJ. Lithocholic acid disrupts phospholipid and sphingolipid homeostasis leading to cholestasis in mice. *Hepatology*. 2011; 53: 1282-93.
82. Wang F, Qin C, Li Y, Qu W, Liu H, Li B, et al. Ursodeoxycholic acid induces autophagy via LC3B to suppress hepatocellular carcinoma in vivo and in vitro. *Int J Clin Exp Pathol*. 2017; 10: 11805-13.
83. Zhu L, Shan LJ, Liu YJ, Chen D, Xiao XG, Li Y. Ursodeoxycholic acid induces apoptosis of hepatocellular carcinoma cells in vitro. *J Dig Dis*. 2014; 15: 684-93.
84. Lin W, Li S, Meng Y, Huang G, Liang S, Du J, et al. UDCA inhibits hypoxic hepatocellular carcinoma cell-induced angiogenesis through suppressing HIF-1 α /VEGF/IL-8 intercellular signaling. *Front Pharmacol*. 2021; 12: 755394.
85. Zeng D, Zhang L, Luo Q. Celastrol-regulated gut microbiota and bile acid metabolism alleviate hepatocellular carcinoma proliferation by regulating the interaction between FXR and RXR α in vivo and in vitro. *Front Pharmacol*. 2023; 14: 1124240.
86. Fu J, Yu M, Xu W, Yu S. Research progress of bile acids in cancer. *Front Oncol*. 2021; 11: 778258.
87. Ma C, Han M, Heinrich B, Fu Q, Zhang Q, Sandhu M, et al. Gut microbiome-mediated bile acid metabolism regulates liver cancer via NKT cells. *Science*. 2018; 360: eaan5931.
88. Sun R, Zhang Z, Bao R, Guo X, Gu Y, Yang W, et al. Loss of SIRT5 promotes bile acid-induced immunosuppressive microenvironment and hepatocarcinogenesis. *J Hepatol*. 2022; 77: 453-66.
89. Loo TM, Kamachi F, Watanabe Y, Yoshimoto S, Kanda H, Arai Y, et al. Gut microbiota promotes obesity-associated liver cancer through PGE(2)-mediated suppression of antitumor immunity. *Cancer Discov*. 2017; 7: 522-38.
90. Jia W, Xie G, Jia W. Bile acid-microbiota crosstalk in gastrointestinal inflammation and carcinogenesis. *Nat Rev Gastroenterol Hepatol*. 2018; 15: 111-28.
91. Kong B, Zhu Y, Li G, Williams JA, Buckley K, Tawfik O, et al. Mice with hepatocyte-specific FXR deficiency are resistant to spontaneous but susceptible to cholic acid-induced hepatocarcinogenesis. *Am J Physiol Gastrointest Liver Physiol*. 2016; 310: G295-302.
92. Xia JK, Tang N, Wu XY, Ren HZ. Deregulated bile acids may drive hepatocellular carcinoma metastasis by inducing an immunosuppressive microenvironment. *Front Oncol*. 2022; 12: 1033145.
93. Wang H, Chu F, Zhang XF, Zhang P, Li LX, Zhuang YL, et al. TPX2 enhances the transcription factor activation of PXR and enhances the resistance of hepatocellular carcinoma cells to antitumor drugs. *Cell Death Dis*. 2023; 14: 64.
94. Jiang Q, Ma Y, Han J, Chu J, Ma X, Shen L, et al. MDM2 binding protein induces the resistance of hepatocellular carcinoma cells to molecular targeting agents via enhancing the transcription factor activity of the Pregnane X receptor. *Front Oncol*. 2021; 11: 715193.
95. Chen Y, Zeng Q, Liu X, Fu J, Zeng Z, Zhao Z, et al. LINE-1 ORF-1p enhances the transcription factor activity of pregnenolone X receptor and promotes sorafenib resistance in hepatocellular carcinoma cells. *Cancer Manag Res*. 2018; 10: 4421-38.
96. Hori T, Yokobori K, Moore R, Negishi M, Sueyoshi T. CAR requires Gadd45 β to promote phenobarbital-induced mouse liver tumors in early stage. *Front Oncol*. 2023; 13: 1217847.
97. Hori T, Saito K, Moore R, Flake GP, Negishi M. Nuclear receptor CAR suppresses GADD45B-p38 MAPK signaling to promote phenobarbital-induced proliferation in mouse liver. *Mol Cancer Res*. 2018; 16: 1309-18.
98. Li Z, Kwon SM, Li D, Li L, Peng X, Zhang J, et al. Human constitutive androstane receptor represses liver cancer development and hepatoma cell proliferation by inhibiting erythropoietin signaling. *J Biol Chem*. 2022; 298: 101885.
99. Braeuning A, Gavrillov A, Brown S, Wolf CR, Henderson CJ, Schwarz M. Phenobarbital-mediated tumor promotion in transgenic mice with humanized CAR and PXR. *Toxicol Sci*. 2014; 140: 259-70.
100. Abdalla M, Khairy E, Louka ML, Ali-Labib R, Ibrahim EA. Vitamin D receptor gene methylation in hepatocellular carcinoma. *Gene*. 2018; 653: 65-71.

101. Duran A, Hernandez ED, Reina-Campos M, Castilla EA, Subramaniam S, Raghunandan S, et al. p62/SQSTM1 by binding to vitamin D receptor inhibits hepatic stellate cell activity, fibrosis, and liver cancer. *Cancer Cell*. 2016; 30: 595-609.
102. Wang H, Chen J, Hollister K, Sowers LC, Forman BM. Endogenous bile acids are ligands for the nuclear receptor FXR/BAR. *Mol Cell*. 1999; 3: 543-53.
103. Su H, Ma C, Liu J, Li N, Gao M, Huang A, et al. Downregulation of nuclear receptor FXR is associated with multiple malignant clinicopathological characteristics in human hepatocellular carcinoma. *Am J Physiol Gastrointest Liver Physiol*. 2012; 303: G1245-53.
104. Yang F, Huang X, Yi T, Yen Y, Moore DD, Huang W. Spontaneous development of liver tumors in the absence of the bile acid receptor farnesoid X receptor. *Cancer Res*. 2007; 67: 863-7.
105. Deuschle U, Schuler J, Schulz A, Schluter T, Kinzel O, Abel U, et al. FXR controls the tumor suppressor NDRG2 and FXR agonists reduce liver tumor growth and metastasis in an orthotopic mouse xenograft model. *PLoS One*. 2012; 7: e43044.
106. Chen WD, Yu D, Forman BM, Huang W, Wang YD. Deficiency of G-protein-coupled bile acid receptor Gpbar1 (TGR5) enhances chemically induced liver carcinogenesis. *Hepatology*. 2013; 57: 656-66.
107. Li CL, Lin YK, Chen HA, Huang CY, Huang MT, Chang YJ. Smoking as an independent risk factor for hepatocellular carcinoma due to the α 7-Nachr modulating the JAK2/STAT3 signaling axis. *J Clin Med*. 2019; 8: 1391.
108. Cai Y, Zeng M, Chen YZ. The pharmacological mechanism of Huashi Baidu Formula for the treatment of COVID-19 by combined network pharmacology and molecular docking. *Ann Palliat Med*. 2021; 10: 3864-95.
109. Staudinger JL, Goodwin B, Jones SA, Hawkins-Brown D, MacKenzie KI, LaTour A, et al. The nuclear receptor PXR is a lithocholic acid sensor that protects against liver toxicity. *Proc Natl Acad Sci U S A*. 2001; 98: 3369-74.
110. Feng F, Jiang Q, Cao S, Cao Y, Li R, Shen L, et al. Pregnane X receptor mediates sorafenib resistance in advanced hepatocellular carcinoma. *Biochim Biophys Acta Gen Subj*. 2018; 1862: 1017-30.
111. Haines C, Elcombe BM, Chatham LR, Vardy A, Higgins LG, Elcombe CR, et al. Comparison of the effects of sodium phenobarbital in wild type and humanized constitutive androstane receptor (CAR)/pregnane X receptor (PXR) mice and in cultured mouse, rat and human hepatocytes. *Toxicology*. 2018; 396-397: 23-32.
112. Yamada T, Okuda Y, Kushida M, Sumida K, Takeuchi H, Nagahori H, et al. Human hepatocytes support the hypertrophic but not the hyperplastic response to the murine nongenotoxic hepatocarcinogen sodium phenobarbital in an in vivo study using a chimeric mouse with humanized liver. *Toxicol Sci*. 2014; 142: 137-57.
113. Quan Y, Yang J, Qin T, Hu Y. Associations between twelve common gene polymorphisms and susceptibility to hepatocellular carcinoma: evidence from a meta-analysis. *World J Surg Oncol*. 2019; 17: 216.
114. Shen S, Khatiwada S, Behary J, Kim R, Zekry A. Modulation of the gut microbiome to improve clinical outcomes in hepatocellular carcinoma. *Cancers (Basel)* 2022; 14: 2099.
115. Dapito DH, Mencin A, Gwak GY, Pradere JP, Jang MK, Mederacke I, et al. Promotion of hepatocellular carcinoma by the intestinal microbiota and TLR4. *Cancer Cell*. 2012; 21: 504-16.
116. Yu LX, Yan HX, Liu Q, Yang W, Wu HP, Dong W, et al. Endotoxin accumulation prevents carcinogen-induced apoptosis and promotes liver tumorigenesis in rodents. *Hepatology*. 2010; 52: 1322-33.
117. Diedrich T, Kummer S, Galante A, Drolz A, Schlicker V, Lohse AW, et al. Characterization of the immune cell landscape of patients with NAFLD. *PLoS One*. 2020; 15: e0230307.
118. Gabrielson A, Wu Y, Wang H, Jiang J, Kallakury B, Gatalica Z, et al. Intratumoral CD3 and CD8 T-cell densities associated with relapse-free survival in HCC. *Cancer Immunol Res*. 2016; 4: 419-30.
119. Panneerselvam S, Wilson C, Kumar P, Abirami D, Pamarthi J, Reddy MS, et al. Overview of hepatocellular carcinoma: from molecular aspects to future therapeutic options. *Cell Adh Migr*. 2023; 17: 1-21.
120. Lynch L, Nowak M, Varghese B, Clark J, Hogan AE, Toxavidis V, et al. Adipose tissue invariant NKT cells protect against diet-induced obesity and metabolic disorder through regulatory cytokine production. *Immunity*. 2012; 37: 574-87.
121. Rong Y, Dong Z, Hong Z, Jin Y, Zhang W, Zhang B, et al. Reactivity toward *Bifidobacterium longum* and *Enterococcus hirae* demonstrate robust CD8(+) T cell response and better prognosis in HBV-related hepatocellular carcinoma. *Exp Cell Res*. 2017; 358: 352-9.
122. Liu Y, Chen K, Li F, Gu Z, Liu Q, He L, et al. Probiotic *Lactobacillus rhamnosus* GG prevents liver fibrosis through inhibiting hepatic bile acid synthesis and enhancing bile acid excretion in mice. *Hepatology*. 2020; 71: 2050-66.
123. Sydor S, Best J, Messerschmidt I, Manka P, Vilchez-Vargas R, Brodesser S, et al. Altered microbiota diversity and bile acid signaling in cirrhotic and noncirrhotic NASH-HCC. *Clin Transl Gastroenterol*. 2020; 11: e00131.
124. Lee PC, Wu CJ, Hung YW, Lee CJ, Chi CT, Lee IC, et al. Gut microbiota and metabolites associate with outcomes of immune checkpoint inhibitor-treated unresectable hepatocellular carcinoma. *J Immunother Cancer*. 2022; 10: e004779.
125. Yamada S, Takashina Y, Watanabe M, Nagamine R, Saito Y, Kamada N, et al. Bile acid metabolism regulated by the gut microbiota promotes non-alcoholic steatohepatitis-associated hepatocellular carcinoma in mice. *Oncotarget*. 2018; 9: 9925-39.
126. Deng Z, Ouyang Z, Mei S, Zhang X, Li Q, Meng F, et al. Enhancing NKT cell-mediated immunity against hepatocellular carcinoma: role of XYXD in promoting primary bile acid synthesis and improving gut microbiota. *J Ethnopharmacol*. 2023; 318(Pt B): 116945.
127. Chen S, Han P, Zhang Q, Liu P, Liu J, Zhao L, et al. *Lactobacillus brevis* alleviates the progress of hepatocellular carcinoma and type 2 diabetes in mice model via interplay of gut microflora, bile acid and NOTCH 1 signaling. *Front Immunol*. 2023; 14: 1179014.

128. Xie G, Wang X, Huang F, Zhao A, Chen W, Yan J, et al. Dysregulated hepatic bile acids collaboratively promote liver carcinogenesis. *Int J Cancer*. 2016; 139: 1764-75.
129. Zhang Z, Wang D, Qiao S, Wu X, Cao S, Wang L, et al. Metabolic and microbial signatures in rat hepatocellular carcinoma treated with caffeic acid and chlorogenic acid. *Sci Rep*. 2017; 7: 4508.
130. Wang C, Yang M, Zhao J, Li X, Xiao X, Zhang Y, et al. Bile salt (glycochenodeoxycholate acid) induces cell survival and chemoresistance in hepatocellular carcinoma. *J Cell Physiol*. 2019; 234: 10899-906.
131. Xing L, Zhang Y, Li S, Tong M, Bi K, Zhang Q, et al. A dual coverage monitoring of the bile acids profile in the liver-gut axis throughout the whole inflammation-cancer transformation progressive: reveal hepatocellular carcinoma pathogenesis. *Int J Mol Sci*. 2023; 24: 4258.
132. Allen K, Jaeschke H, Copple BL. Bile acids induce inflammatory genes in hepatocytes: a novel mechanism of inflammation during obstructive cholestasis. *Am J Pathol*. 2011; 178: 175-86.
133. Liu Chen Kiow J, Vincent C, Sidani S, Bouin M. High occurrence of small intestinal bacterial overgrowth in primary biliary cholangitis. *Neurogastroenterol Motil*. 2019; 31: e13691.
134. Tang R, Wei Y, Li Y, Chen W, Chen H, Wang Q, et al. Gut microbial profile is altered in primary biliary cholangitis and partially restored after UDCA therapy. *Gut*. 2018; 67: 534-41.
135. Sabino J, Vieira-Silva S, Machiels K, Joossens M, Falony G, Ballet V, et al. Primary sclerosing cholangitis is characterised by intestinal dysbiosis independent from IBD. *Gut*. 2016; 65: 1681-9.
136. Zhan Q, Qi X, Weng R, Xi F, Chen Y, Wang Y, et al. Alterations of the human gut microbiota in intrahepatic cholestasis of pregnancy. *Front Cell Infect Microbiol*. 2021; 11: 635680.
137. Hourigan SK, Moutinho TJ Jr., Berenz A, Papin J, Guha P, Bangiolino L, et al. Gram-negative microbiota blooms in premature twins discordant for parenteral nutrition-associated cholestasis. *J Pediatr Gastroenterol Nutr*. 2020; 70: 640-4.
138. Tarao K, Fujiyama S, Ohkawa S, Miyakawa K, Tamai S, Hirokawa S, et al. Ursodiol use is possibly associated with lower incidence of hepatocellular carcinoma in hepatitis C virus-associated liver cirrhosis. *Cancer Epidemiol Biomarkers Prev*. 2005; 14: 164-9.
139. Kulkarni AV, Tevethia HV, Arab JP, Candia R, Premkumar M, Kumar P, et al. Efficacy and safety of obeticholic acid in liver disease-A systematic review and meta-analysis. *Clin Res Hepatol Gastroenterol*. 2021; 45: 101675.
140. Gou H, Liu S, Liu L, Luo M, Qin S, He K, et al. Obeticholic acid and 5 β -cholanolic acid 3 exhibit anti-tumor effects on liver cancer through CXCL16/CXCR6 pathway. *Front Immunol*. 2022; 13: 1095915.
141. Attia YM, Tawfiq RA, Ali AA, Elmazar MM. The FXR agonist, obeticholic acid, suppresses HCC proliferation & metastasis: role of IL-6/STAT3 signalling pathway. *Sci Rep*. 2017; 7: 12502.
142. Younossi ZM, Ratziu V, Loomba R, Rinella M, Anstee QM, Goodman Z, et al. Obeticholic acid for the treatment of non-alcoholic steatohepatitis: interim analysis from a multicentre, randomised, placebo-controlled phase 3 trial. *Lancet*. 2019; 394: 2184-96.
143. Trauner M, Nevens F, Shiffman ML, Drenth JPH, Bowlus CL, Vargas V, et al. Long-term efficacy and safety of obeticholic acid for patients with primary biliary cholangitis: 3-year results of an international open-label extension study. *Lancet Gastroenterol Hepatol*. 2019; 4: 445-53.
144. Harrison SA, Bashir MR, Lee KJ, Shim-Lopez J, Lee J, Wagner B, et al. A structurally optimized FXR agonist, MET409, reduced liver fat content over 12 weeks in patients with non-alcoholic steatohepatitis. *J Hepatol*. 2021; 75: 25-33.
145. Patel K, Harrison SA, Elkhatab M, Trotter JF, Herring R, Rojter SE, et al. Cilofexor, a nonsteroidal FXR agonist, in patients with noncirrhotic NASH: a phase 2 randomized controlled trial. *Hepatology*. 2020; 72: 58-71.
146. Ratziu V, Harrison SA, Loustaud-Ratti V, Bureau C, Lawitz E, Abdelmalek M, et al. Hepatic and renal improvements with FXR agonist vonafexor in individuals with suspected fibrotic NASH. *J Hepatol*. 2023; 78: 479-92.
147. Sanyal AJ, Lopez P, Lawitz EJ, Lucas KJ, Loeffler J, Kim W, et al. Tropifexor for nonalcoholic steatohepatitis: an adaptive, randomized, placebo-controlled phase 2a/b trial. *Nat Med*. 2023; 29: 392-400.
148. Harrison SA, Abdelmalek MF, Neff G, Gunn N, Guy CD, Alkhoury N, et al. Aldafermin in patients with non-alcoholic steatohepatitis (ALPINE 2/3): a randomised, double-blind, placebo-controlled, phase 2b trial. *Lancet Gastroenterol Hepatol*. 2022; 7: 603-16.
149. Rinella ME, Trotter JF, Abdelmalek MF, Paredes AH, Connelly MA, Jaros MJ, et al. Rosuvastatin improves the FGF19 analogue NGM282-associated lipid changes in patients with non-alcoholic steatohepatitis. *J Hepatol*. 2019; 70: 735-44.
150. Luo J, Ko B, Elliott M, Zhou M, Lindhout DA, Phung V, et al. A nontumorigenic variant of FGF19 treats cholestatic liver diseases. *Sci Transl Med*. 2014; 6: 247ra100.
151. Gadaleta RM, Scialpi N, Peres C, Cariello M, Ko B, Luo J, et al. Suppression of hepatic bile acid synthesis by a non-tumorigenic FGF19 analogue protects mice from fibrosis and hepatocarcinogenesis. *Sci Rep*. 2018; 8: 17210.
152. Chan SL, Schuler M, Kang YK, Yen CJ, Edeline J, Choo SP, et al. A first-in-human phase 1/2 study of FGF401 and combination of FGF401 with spartalizumab in patients with hepatocellular carcinoma or biomarker-selected solid tumors. *J Exp Clin Cancer Res*. 2022; 41: 189.
153. Huang S, Wu Y, Zhao Z, Wu B, Sun K, Wang H, et al. A new mechanism of obeticholic acid on NASH treatment by inhibiting NLRP3 inflammasome activation in macrophage. *Metabolism*. 2021; 120: 154797.
154. Rao A, Kusters A, Mells JE, Zhang W, Setchell KD, Amanso AM, et al. Inhibition of ileal bile acid uptake protects against nonalcoholic fatty liver disease in high-fat diet-fed mice. *Sci Transl Med*. 2016; 8: 357ra122.
155. Baghdasaryan A, Fuchs CD, Osterreicher CH, Lemberger UJ, Halilbasic E, Pahlman I, et al. Inhibition of intestinal bile acid absorption improves cholestatic liver and bile duct injury in a mouse model of sclerosing cholangitis. *J Hepatol*. 2016; 64: 674-81.

156. Thompson RJ, Arnell H, Artan R, Baumann U, Calvo PL, Czubkowski P, et al. Odevixibat treatment in progressive familial intrahepatic cholestasis: a randomised, placebo-controlled, phase 3 trial. *Lancet Gastroenterol Hepatol.* 2022; 7: 830-42.
157. Newsome PN, Palmer M, Freilich B, Sheikh MY, Sheikh A, Sarles H, et al. Volixibat in adults with non-alcoholic steatohepatitis: 24-week interim analysis from a randomized, phase II study. *J Hepatol.* 2020; 73: 231-40.
158. Luo M, Yan J, Wu L, Wu J, Chen Z, Jiang J, et al. Probiotics alleviated nonalcoholic fatty liver disease in high-fat diet-fed rats via gut microbiota/FXR/FGF15 signaling pathway. *J Immunol Res.* 2021; 2021: 2264737.
159. Wang J, Zhao H, Zheng L, Zhou Y, Wu L, Xu Y, et al. FGF19/SOCE/NFATc2 signaling circuit facilitates the self-renewal of liver cancer stem cells. *Theranostics.* 2021; 11: 5045-60.
160. Ocvirk S, O'Keefe SJD. Dietary fat, bile acid metabolism and colorectal cancer. *Semin Cancer Biol.* 2021; 73: 347-55.
161. Li M, Wang S, Li Y, Zhao M, Kuang J, Liang D, et al. Gut microbiota-bile acid crosstalk contributes to the rebound weight gain after calorie restriction in mice. *Nat Commun.* 2022; 13: 2060.
162. Yang J, Yu J. The association of diet, gut microbiota and colorectal cancer: what we eat may imply what we get. *Protein Cell.* 2018; 9: 474-87.
163. Ji X, Wang J, Li Z, Shen Q, Tuo J, Bi J, et al. Dietary fat intake and liver cancer risk: a prospective cohort study in Chinese women. *Cancer Biol Med.* 2021; 19: 370-83.
164. Huang F, Zheng X, Ma X, Jiang R, Zhou W, Zhou S, et al. Theabrownin from Pu-erh tea attenuates hypercholesterolemia via modulation of gut microbiota and bile acid metabolism. *Nat Commun.* 2019; 10: 4971.
165. Wang JW, Kuo CH, Kuo FC, Wang YK, Hsu WH, Yu FJ, et al. Fecal microbiota transplantation: review and update. *J Formos Med Assoc.* 2019; 118 Suppl 1: S23-31.
166. Gupta S, Mullish BH, Allegretti JR. Fecal microbiota transplantation: the evolving risk landscape. *Am J Gastroenterol.* 2021; 116: 647-56.
167. Chen Q, Fan Y, Zhang B, Yan C, Zhang Q, Ke Y, et al. Capsulized fecal microbiota transplantation induces remission in patients with ulcerative colitis by gut microbial colonization and metabolite regulation. *Microbiol Spectr.* 2023; 11: e0415222.
168. Ren YD, Ye ZS, Yang LZ, Jin LX, Wei WJ, Deng YY, et al. Fecal microbiota transplantation induces hepatitis B virus e-antigen (HBeAg) clearance in patients with positive HBeAg after long-term antiviral therapy. *Hepatology.* 2017; 65: 1765-8.
169. Allegretti JR, Kassam Z, Mullish BH, Chiang A, Carrellas M, Hurtado J, et al. Effects of fecal microbiota transplantation with oral capsules in obese patients. *Clin Gastroenterol Hepatol.* 2020; 18: 855-63.e852.
170. Bustamante JM, Dawson T, Loeffler C, Marfori Z, Marchesi JR, Mullish BH, et al. Impact of fecal microbiota transplantation on gut bacterial bile acid metabolism in humans. *Nutrients.* 2022; 14: 5200.
171. Finn RS, Qin S, Ikeda M, Galle PR, Ducreux M, Kim TY, et al. Atezolizumab plus bevacizumab in unresectable hepatocellular carcinoma. *N Engl J Med.* 2020; 382: 1894-905.
172. Zhou A, Tang L, Zeng S, Lei Y, Yang S, Tang B. Gut microbiota: a new piece in understanding hepatocarcinogenesis. *Cancer Lett.* 2020; 474: 15-22.
173. Iida N, Dzutsev A, Stewart CA, Smith L, Bouladoux N, Weingarten RA, et al. Commensal bacteria control cancer response to therapy by modulating the tumor microenvironment. *Science.* 2013; 342: 967-70.
174. Zheng Y, Wang T, Tu X, Huang Y, Zhang H, Tan D, et al. Gut microbiome affects the response to anti-PD-1 immunotherapy in patients with hepatocellular carcinoma. *J Immunother Cancer.* 2019; 7: 193.
175. Oh B, Boyle F, Pavlakis N, Clarke S, Eade T, Hruby G, et al. The gut microbiome and cancer immunotherapy: can we use the gut microbiome as a predictive biomarker for clinical response in cancer immunotherapy? *Cancers (Basel)* 2021; 13: 4824.
176. Park JS, Gazzaniga FS, Wu M, Luthens AK, Gillis J, Zheng W, et al. Targeting PD-L2-RGMB overcomes microbiome-related immunotherapy resistance. *Nature.* 2023; 617: 377-85.
177. Griffin ME, Espinosa J, Becker JL, Luo JD, Carroll TS, Jha JK, et al. Enterococcus peptidoglycan remodeling promotes checkpoint inhibitor cancer immunotherapy. *Science.* 2021; 373: 1040-6.
178. Mager LF, Burkhard R, Pett N, Cooke NCA, Brown K, Ramay H, et al. Microbiome-derived inosine modulates response to checkpoint inhibitor immunotherapy. *Science.* 2020; 369: 1481-9.
179. Tanoue T, Morita S, Plichta DR, Skelly AN, Suda W, Sugiura Y, et al. A defined commensal consortium elicits CD8 T cells and anti-cancer immunity. *Nature.* 2019; 565: 600-5.
180. Routy B, Le Chatelier E, Derosa L, Duong CPM, Alou MT, Daillere R, et al. Gut microbiome influences efficacy of PD-1-based immunotherapy against epithelial tumors. *Science.* 2018; 359: 91-7.
181. Huang H, Ren Z, Gao X, Hu X, Zhou Y, Jiang J, et al. Integrated analysis of microbiome and host transcriptome reveals correlations between gut microbiota and clinical outcomes in HBV-related hepatocellular carcinoma. *Genome Med.* 2020; 12(1): 102.
182. Baruch EN, Youngster I, Ben-Betzalel G, Ortenberg R, Lahat A, Katz L, et al. Fecal microbiota transplant promotes response in immunotherapy-refractory melanoma patients. *Science.* 2021; 371: 602-9.

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