

# Genomic epidemiology and antifungal-resistant characterization of *Candida auris*, Colombia, 2016–2021

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**ABSTRACT** Since 2016, in Colombia, ongoing transmission of *Candida auris* has been reported in multiple cities. Here, we provide an updated description of *C. auris* genomic epidemiology and the dynamics of antifungal resistance in Colombia. We sequenced 99 isolates from *C. auris* cases with collection dates ranging from June 2016 to January 2021; the resulting sequences coupled with 103 previously generated sequences from *C. auris* cases were described in a phylogenetic analysis. All *C. auris* cases were clade IV. Of the 182 isolates with antifungal susceptibility data, 67 (37%) were resistant to fluconazole, and 39 (21%) were resistant to amphotericin B. Isolates predominately clustered by country except for 16 isolates from Bogotá, Colombia, which grouped with isolates from Venezuela. The largest cluster ( $N = 166$  isolates) contained two subgroups. The first subgroup contained 26 isolates, mainly from César; of these, 85% ( $N = 22$ ) were resistant to fluconazole. The second subgroup consisted of 47 isolates from the north coast; of these, 81% ( $N = 38$ ) were resistant to amphotericin B. Mutations in the *ERG11* and *TAC1B* genes were identified in fluconazole-resistant isolates. This work describes molecular mechanisms associated with *C. auris* antifungal resistance in Colombia. Overall, *C. auris* cases from different geographic locations in Colombia exhibited high genetic relatedness, suggesting continued transmission between cities since 2016. These findings also suggest a lack of or minimal introductions of different clades of *C. auris* into Colombia.

**IMPORTANCE** *Candida auris* is an emerging fungus that presents a serious global health threat and has caused multiple outbreaks in Colombia. This work discusses the likelihood of introductions and local transmission of *C. auris* and provides an updated description of the molecular mechanisms associated with antifungal resistance in Colombia. Efforts like this provide information about the evolving *C. auris* burden that could help guide public health strategies to control *C. auris* spread.

**KEYWORDS** *Candida auris*, antifungal, resistance, genomics, WGS, epidemiology

*Candida auris* is a multidrug-resistant yeast capable of causing outbreaks of infections associated with high mortality rates (1). Genomic analysis of *C. auris* infections from global regions showed six major clades (I–VI) (2). In Colombia, genomic sequencing revealed that all *C. auris* cases were clade IV and suggested ongoing transmission of *C. auris* in multiple cities (3).

Regarding antifungal susceptibility, about 35% of *C. auris* isolates were resistant to fluconazole and 33% were resistant to amphotericin B (4), mainly from the north-coast region of Colombia (3). Molecular mechanisms associated with azole resistance often involve mutations in the *ERG11* and *TAC1B* genes. For polyenes, molecular mechanisms conferring resistance are largely unknown; a previous report identified two non-synonymous substitutions related with high MIC values of amphotericin B (3).

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In this study, we investigated genomic epidemiology of *C. auris* cases from 2016 to 2021 in Colombia to obtain an updated description of *C. auris* transmission dynamics and molecular characterization of antifungal resistance.

## MATERIALS AND METHODS

In 2016, the Colombian Instituto Nacional de Salud (INS) published the National Alert (5), which requested that public health laboratories to send all suspected or confirmed *C. auris* isolates to the INS.

Additional Materials and Methods are described in the Supplemental Material. Briefly, the U.S. Centers for Disease Control and Prevention received 99 *C. auris* isolates and performed species identification, antifungal susceptibility testing (AFST), and whole-genome sequencing (WGS; BioProject [PRJNA1003896](#); Tables S1 and S2). WGS data were used to generate a phylogenetic tree and pairwise distance comparisons and to identify mutations related to antifungal drug resistance.

## RESULTS

Phylogenetic analysis of 99 newly and 83 previously generated sequences revealed that all *C. auris* in Colombia were clade IV (maximum single nucleotide polymorphisms [SNPs] difference to the reference sequence: 205 SNPs). Isolates predominately clustered by country. Colombian isolates were dispersed in two clusters (C1 and C2). Most Colombian isolates ( $n = 166$ , 91%) clustered with the previously described Colombian isolates in cluster C1 with collection dates from 2015 to 2016 (Fig. 1A).

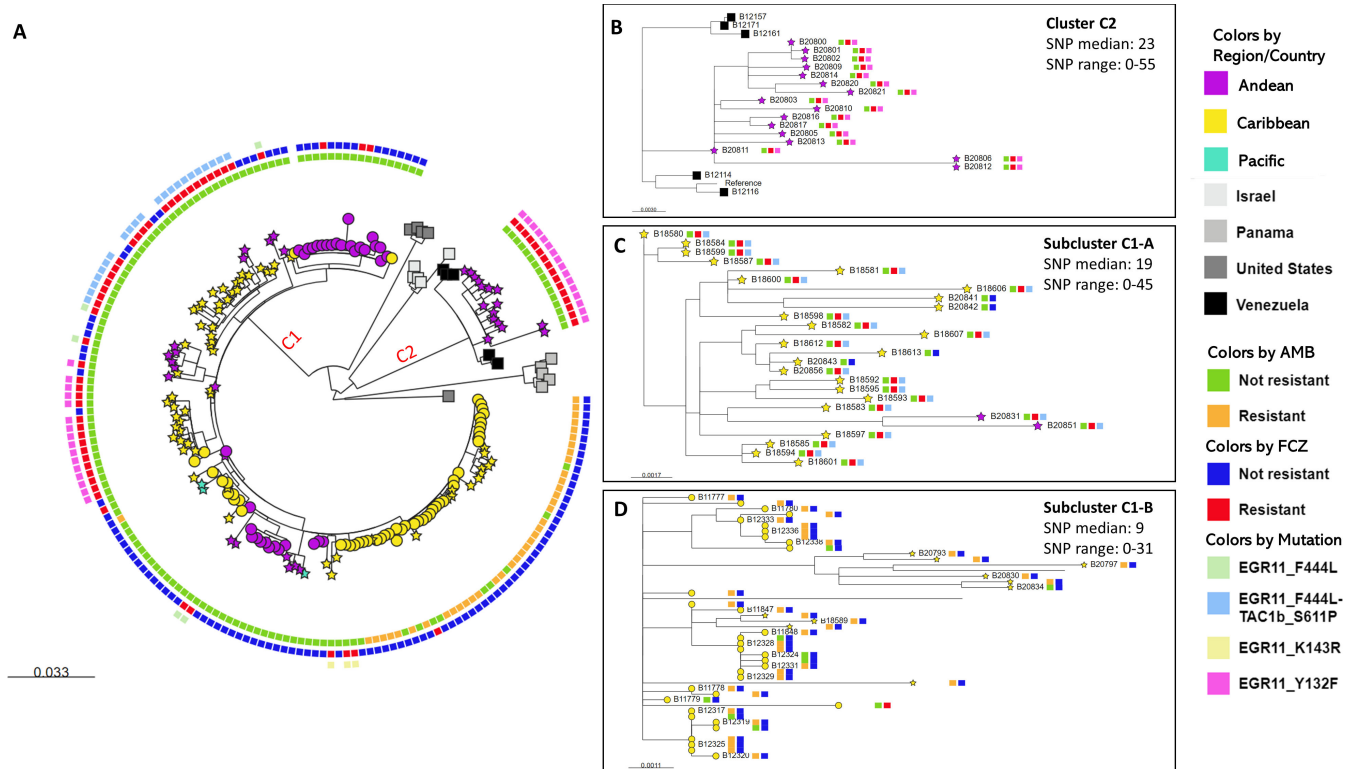
Cluster C2 was separated by 152 SNPs from cluster C1. Cluster C2 comprised 16 isolates from Bogotá collected between May and November 2020 and five isolates from Venezuela collected between 2012 and 2015 (Fig. 1A). All Colombian isolates in this cluster and two Venezuelan isolates were fluconazole-resistant (Fig. 1B; Table S1). Within cluster C1, we highlighted two subclusters (C1-A and C1-B) of isolates with bootstrap values >98% and with high (>80%) proportion of antifungal-resistant isolates (Fig. 1C through D; Fig. S2).

Among all 182 Colombian isolates, 67 (37%) were resistant to fluconazole, 39 (21%) were resistant to amphotericin B, and only one was resistant to anidulafungin (Fig. S1). We found that 62 (93%) of the 67 fluconazole-resistant isolates had known mutations in *ERG11* and *TAC1B* genes. The Y132F mutation in the *ERG11* gene was the most prevalent mutation, observed in 32 (48%) azole-resistant isolates (Table S3). All fluconazole-resistant isolates in subcluster C1-A ( $n = 22$ ), collected in 2016, carried the concomitant mutations F444L in the *ERG11* gene and S611P in the *TAC1B* gene. Significant differences were observed between MIC values, supporting the cumulative effect of the concomitant mutation in *ERG11* and *TAC1B* genes (Fig. 2B). All ( $n = 16$ ) fluconazole-resistant isolates in cluster C2 carried the mutation Y132F. Of the 39 amphotericin B-resistant isolates, we found that 38 (97%) carried the substitutions S108N in *FLO8* (PSK76257) gene and I139T in PSK74852 gene. Both substitutions were also found in eight susceptible isolates with amphotericin B MIC values  $\geq 0.75$  and in only one isolate with amphotericin B MIC value of 0.25 mg/ $\mu$ L.

## DISCUSSION

Here, we provided an update on the genomic epidemiology of *C. auris* in Colombia. All *C. auris* cases in Colombia continue to be of clade IV. This contrasts with several countries experiencing *C. auris* outbreaks that have identified multiple *C. auris* clades (6). The uniformity of clade IV isolates could be due to (1) lack of introductions of other clades or (2) lack of subsequent transmission after introductions of other clades. Alternatively, isolates from other *C. auris* clades could have been missed by our sampling.

Previously, a global description of *C. auris* reported genetic clustering by country within clade IV (6). Here, we observed a weaker phylogeographic structure as isolates

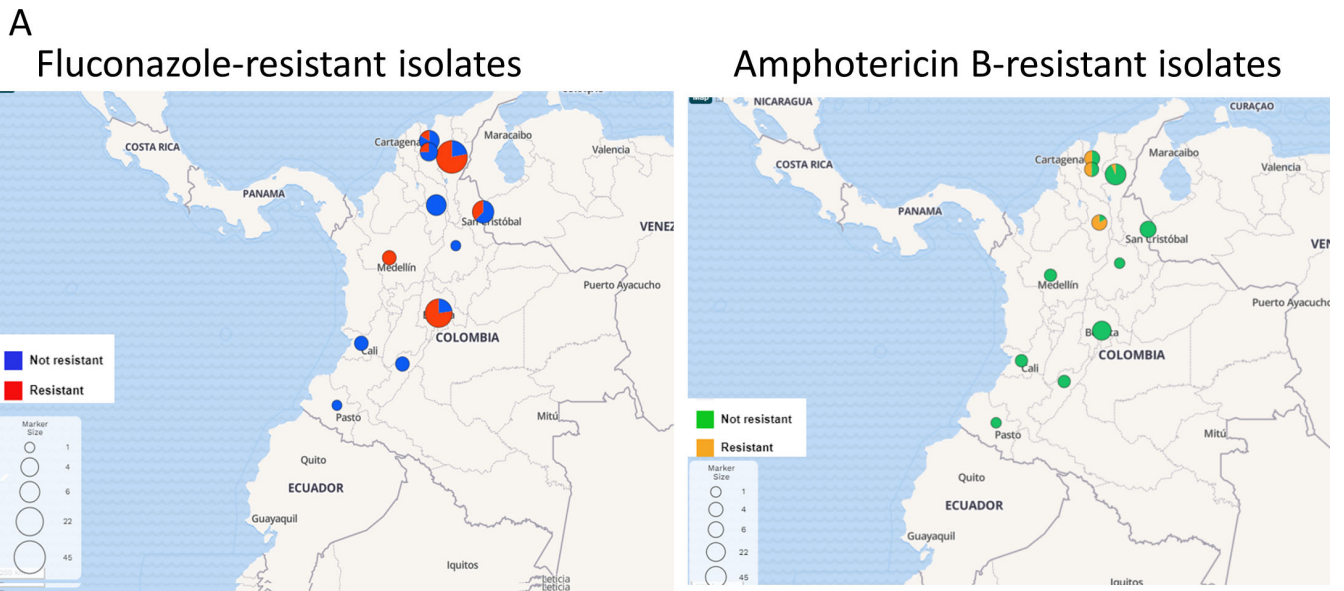


**FIG 1** (A) Phylogenetic tree using maximum likelihood method; the majority of isolates clustered by country. Cluster C1 consists of Colombian isolates, and cluster C2 consists of Venezuelan isolates and fluconazole-resistant Colombian isolates. Taxa color represents country or Colombian region: Andean (Antioquia, Bogotá-Cundinamarca, Norte de Santander, Huila), Caribbean (Atlántico, Bolívar, Cesar, Magdalena), and Pacific (Nariño, Valle); isolates from countries other than Colombia are shown as squares. Colombian isolates sequenced in previous studies are shown as circles, and Colombian isolates sequenced in this study are shown as stars. External circle colors represent resistant and no resistant phenotypes for fluconazole and amphotericin B; presence of mutation in *EGR11* and *TACb1* genes. (B) Zoom-in view of cluster C2. (C) Zoom-in view of subcluster C1-A, which contains mainly fluconazole-resistant isolates. (D) Zoom-in view of subcluster C1-B, which contains mainly amphotericin B-resistant isolates. For panels B, C, and D, the squares to the right of the label represent resistant and non-resistant phenotypes and mutation in *EGR11* and *TACb1* genes.

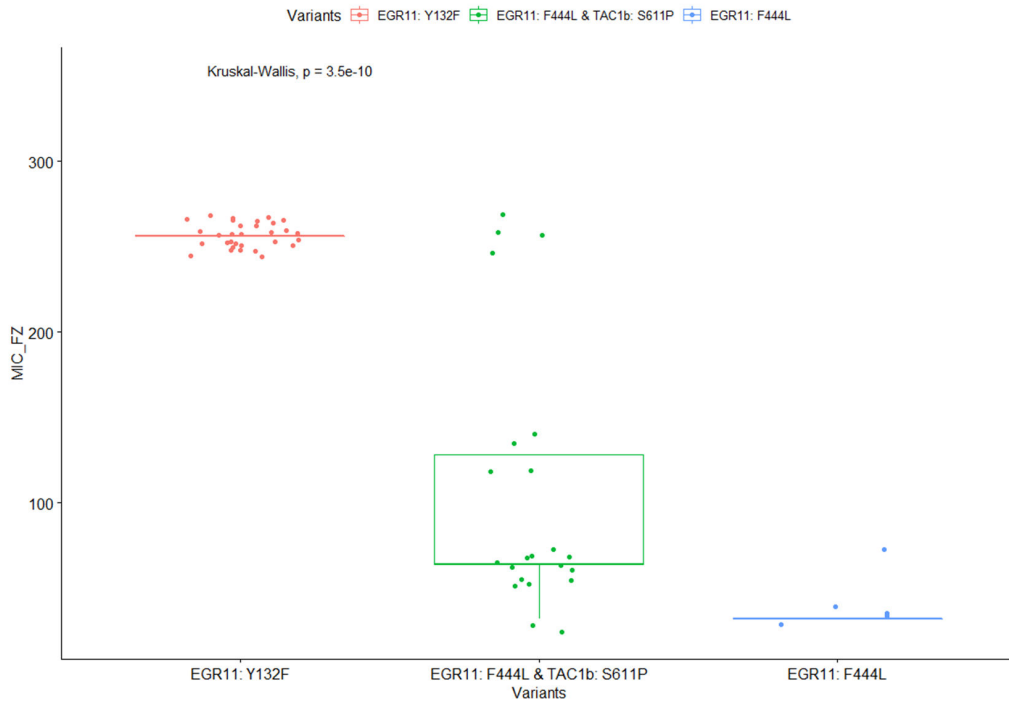
from Colombia were dispersed into two clusters. Cluster C2 included fluconazole-resistant isolates collected in 2020 from cases in Bogotá and Venezuela. The remaining Colombian isolates clustered with cases from Colombia (cluster C1). Interestingly, all clinical isolates from Bogotá collected before 2020 were susceptible; this could suggest a later introduction of fluconazole-resistant cases from Venezuela. We hypothesize that an introduction of clade IV cases occurred in Colombia, possibly from Venezuela where *C. auris* is circulating since 2012 (4). Such a recent introduction could have given rise to the C2 cluster. Additional phylogeographic and molecular clock analyses are needed to estimate when this described introduction could have occurred.

Low median SNP differences were observed among isolates in both subclusters C1-A and C1-B (Fig. 1) spanning a 6-year period, suggesting that most cases in Colombia (cluster C1) are the result of ongoing transmission since 2016.

We concluded that amphotericin B-resistant isolates clustered (subcluster C1-B) and were predominately collected from the northern region of the country, which was previously reported by Escandon et al. (3). Additionally, the fluconazole-resistant isolates from Colombia were predominately collected from the northeast region and Bogotá, in the central region. We observed that the most common mutation, *ERG11* Y132F, was associated with the highest MICs for fluconazole compared to other mutations (Fig. 2B). Y132F was present in all isolates in cluster C2 from 2020. The second most common genotype associated with fluconazole resistance had both mutations in *ERG11* (F444L) and *TAC1B* (S611P; Fig. 2B). Interestingly, the genotypes *ERG11* F444L and *TAC1B* S611P



**B**



**FIG 2** (A) Geographic distribution of antifungal-resistant isolates. Maps were generated from metadata containing AFST data for each sample using Microreact (<http://microreact.org>). (B) R boxplot on common mutation in *ERG11* and *TAC1B* genes and MIC values (mg/μL) of fluconazole-resistant isolates. Isolates with mutation Y132F in the *ERG11* gene had significantly higher fluconazole MIC values (MIC value median = 256) than isolates with concomitant mutations F444L in the *ERG11* gene and S611P in the *TAC1B* gene (MIC value median = 64). The isolates with concomitant mutations also had significantly higher fluconazole MIC values than isolates with the single mutation F444L in the *ERG11* gene (MIC value median = 32). Significant differences between MIC values support the cumulative effect of the concomitant mutation in *ERG11* and *TAC1B* genes. Isolates B11790 and B11087 with mutations *ERG11* K143R were excluded from the graphic. jitter R function was used to add minimal and random noise to the MIC values to visualize the number of isolates at each value.

have been reported only in Colombian clade IV isolates to date (7) and were restricted to C1-A, which were collected in 2016 or later. The emergence of a new mechanism of fluconazole resistance in 2016 and the introduction of resistant *C. auris* from another

country in 2020 could explain the increase in MIC values of fluconazole-resistant isolates observed by Escandon et al. (4).

In contrast with the observed increase in MIC values of fluconazole-resistant isolates, resistance to amphotericin B has not changed significantly over time (Table S4). In amphotericin B-resistant isolates, we observed the substitution S108N in *FLO8* (PSK76257) and the substitution I139T in PSK74852, which were also previously reported in Colombian isolates (3). If and how the described substitutions lead to a reduction in amphotericin B susceptibility are unknown.

One limitation is that the patient history of antifungal treatment was unknown. Some cases with resistant infections could have acquired resistance in response to therapy rather than transmission of drug-resistant *C. auris*. However, given the low genetic differences between some cases and the common phylogeny, transmission seems more likely. A second limitation is that specimens were collected by convenience sampling and therefore are not representative of *C. auris* cases in Colombia.

In conclusion, the findings provided evidence of ongoing spread and a better understanding of molecular mechanisms associated with *C. auris* antifungal resistance in Colombia. This work contributed to the understanding of the *C. auris* epidemiology and factors associated with transmission in Colombia and could be useful to guide prevention and control strategies.

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## DATA AVAILABILITY

The data underlying this study are available in the GenBank Nucleotide Database at <https://www.ncbi.nlm.nih.gov/bioproject/>, and can be accessed with the accession number PRJNA1003896.

## ADDITIONAL FILES

The following material is available [online](#).

## Supplemental Material

Supplemental material (mSphere00577-23-S0001.docx). Supplemental text, Fig. S1 and S2, and Tables S1 to S4.

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