**ENVIRONMENTAL MICROBIOLOGY - REVIEW** 





# A critical review on bioaerosols—dispersal of crop pathogenic microorganisms and their impact on crop yield

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#### Abstract

Bioaerosols are potential sources of pathogenic microorganisms that can cause devastating outbreaks of global crop diseases. Various microorganisms, insects and viroids are known to cause severe crop diseases impeding global agro-economy. Such losses threaten global food security, as it is estimated that almost 821 million people are underfed due to global crisis in food production. It is estimated that global population would reach 10 billion by 2050. Hence, it is imperative to substantially increase global food production to about 60% more than the existing levels. To meet the increasing demand, it is essential to control crop diseases and increase yield. Better understanding of the dispersive nature of bioaerosols, seasonal variations, regional diversity and load would enable in formulating improved strategies to control disease severity, onset and spread. Further, insights on regional and global bioaerosol composition and dissemination would help in predicting and preventing endemic and epidemic outbreaks of crop diseases. Advanced knowledge of the factors influencing disease onset and progress, mechanism of pathogen attachment and penetration, dispersal of pathogens, life cycle and the mode of infection, aid the development and implementation of species-specific and region-specific preventive strategies to control crop diseases. Forthcoming studies on the development of an appropriately stacked *R* gene with a wide range of resistance to crop diseases would enable proper management and yield. The article reviews various aspects of pathogenic bioaerosols, pathogen invasion and infestation, crop diseases and yield.

Keywords Bioaerosols · Crop diseases · Yield loss · Source · Global impact · Biotic factors

# Introduction and relevance of bioaerosols in crop diseases

Bioaerosols are the subset of atmospheric aerosol particles that are of biological origin and vary from a few nm to 100  $\mu$ m in size [182]. They are ubiquitous in the lower parts of the atmosphere i.e., the planetary boundary layer (PBL) as they are majorly released from the Earth's surface due to various natural and anthropogenic activities [304]. They use PBL as a medium to enable and enhance their transportation and dissemination, spread and distribution,

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Pooja Shivanand pooja.shivanand@ubd.edu.bn; poojashivanand@outlook.com evolution, mutation, emergence of new variant, adaptation, improved pathogenicity, development of wide host specificity, and drug resistance [112, 180, 181, 400, 479, 610]. Their unique nature to carry the particles to longer distances with the air currents makes them a key factor in the dispersal of reproductive and other units of plants, animals, and pathogenic microbes across the geographical barrier [124, 182]. Bioaerosols are well known for their beneficial role in the climate system [180] and are equally hazardous to ecosystem health including crop health when they harbor pathogenic microbes in them [124, 610]. They act as agents of spread and dispersal of the human and crop pathogenic microbes that could have deleterious effect on the public and agricultural health of a country [4, 65, 66, 124, 137, 167, 183, 206, 286, 624]. Notably, the pristine atmospheric air present in the vegetated regions play a vital role in the dissemination of the beneficial as well as the crop pathogenic microbes to various ecological niches enhancing their colonization on various substrates [120, 251, 462, 466].

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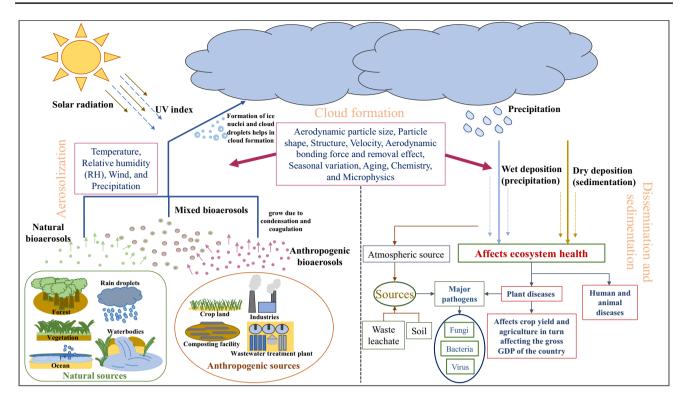
Till date, a significant proportion of the particulate matters in the atmospheric air are said to be composed of bioaerosols which could harbor various pathogenic microbes that could hamper the agricultural sector and economy. Accordingly, bioaerosols were found to contribute to about 24% of the total particulate matters and about 5-10% of the suspended particulate matters in the atmospheric air [4, 373]. Further, various researchers worldwide like [26, 66, 82, 345], Burt [73, 194], and [388] have discussed various crop and plant pathogenic diseases namely rust, smut, powdery mildew, downy mildew, etc. that are generally transmitted by the bioaerosols. Hence, it is highly essential and relevant to study the crop pathogenic bioaerosols and their diversity as they could impact the agricultural sector in the local and the global scale [124]. Furthermore, the uncertainties prevailing in the current understanding of the climate change, the role of bioaerosols in the terrestrial interactions, and anthropogenic influence posed by human activities hampering the agricultural production emphasizes the need to have better insight on the crop pathogenic bioaerosols [529, 533, 541]. Manmade activities like land use pattern and thus liberated bioaerosols have always had a negative implication on the climate and precipitation [402]. Such altered precipitation cycle and climate change would affect the vegetation and agricultural sector thus negatively influencing the economy of a country where agriculture plays a vital role in the gross domestic product (GDP). In this perspective, this article will attempt to summarize and highlight the role of bioaerosols in the spread of crop diseases, various microbial pathogens, mechanism of infection, and the possible preventive measures that need to be followed. Moreover, the manuscript will emphasize on the following objectives: (i) in general the role of bioaerosols in crop diseases and yield loss is understudied, therefore the manuscript elaborates their role and importance in crop disease leading to heavy yield losses; (ii) it also ventures the vital role of meteorological and atmospheric factors in the spread and aerosolization of bioaerosols which would help us in understanding the basic information and knowledge on the influence of changing climatic factors over the increased crop diseases worldwide; (iii) it gives the readers an insight on the diverse pathogenic microbes, source and mode of spread of pathogenic bioaerosols and their route of entry in crops which could help farmers and officials in preparing the crop protection strategies. In order to achieve all these objectives, the manuscript carefully attempts to cover most of the details related to the disease caused by biotic and meso-biotic factors using stringent and specific methodologies in data collection especially the literatures based on the generalized and specific search strings like "bioaerosols and crop diseases, sources of bioaerosols, factors influencing aerosolization and dissemination of bioaerosols, fungal crop diseases, bacterial crop diseases, meso-biotic factors, viral crop diseases, invasion of crop pathogens, etc." excluding the details and data related to the crop diseases caused by abiotic factors like spoilage due to extreme temperature or rainfall and other implications of bioaerosols in ecosystem.

# Source of bioaerosols- factors influencing their release and dissemination

As already discussed, bioaerosols are the aerosol particles that contain a biological unit in them which could either be a living cell or a dead material [22, 83, 103, 124, 182, 209, 564, 610, 630]. First study on bioaerosols was reported in the early nineteenth century and many other studies addressing various aspects of the bioaerosols were carried out since then [413]. As addressed by various researchers worldwide, they play an imperative role in the dissemination of various biological units, dissemination of disease and infection, helping in the genetic exchange and development of new variants, and in the development and evolution dynamics of the ecosystem [66, 120, 148, 216, 217, 224, 354, 378, 460, 463, 469, 516, 527, 598]. Though bioaerosols transport the microbes to longer distances, their vertical spread across the troposphere or PBL has not yet been addressed properly [124].

#### Natural activities as source of bioaerosols

Natural activities observed on the land surfaces and the water surfaces act as an important source of bioaerosols emission and spread [227, 337, 401, 551, 552, 623] (Fig. 1). Further, wind movement over the land surface, water surface, and vegetated regions including the cryptogamic covers contribute to large numbers of bioaerosols from the biosphere [29, 62, 65, 149] as leaf surface area contributes to four times the area as compared to the terrestrial ground surface area [604]. Also, [140] has reported that the dry air and strong wind would favor the dissemination of bioaerosols to longer distances. Movement and activities observed in trees, plants, and crops including the leaf movement were also found to contribute tremendous quantities of bioaerosols to the atmospheric air especially the ice nucleating (IN) microbes which are pathogenic to crops and plants [124, 393, 458, 470, 596]. Furthermore, cryptogamic covers that comprise the region blanketed with the growth of fungi, bacteria, cyanobacteria, lichens, algae, and bryophytes play an imperative role in the release and spread of bioaerosols [37, 61, 67, 120, 122, 124, 164, 183, 366, 511, 526]. Bioaerosols liberation is further enhanced when these cryptogamic covers (which approximately covers almost 1/3rd of the ground surface



**Fig. 1** Schematic representation of the natural and anthropogenic bioaerosols sources, their release from the biosphere along with their role in the ice nuclei formation influencing the climate change, and

their role in affecting crop health and ecosystem health through wet and dry deposition

area) are disturbed by the wind, rainfall, and natural activities of human and animals [149, 182]. Such bioaerosols emitted from the vegetated region and cryptogamic covers would mainly comprise of pollen grains, bacterial cells, small seeds, cyanobacteria, fungal spores and segments, plant debris and segments, dead animal units and cells, algae, insects and their segments, etc. [22, 67, 81, 83, 103, 124, 209, 351, 373, 482, 564, 610].

Likewise, virus particles, viroids, cyanobacteria, and bacteria dominate the bioaerosols liberated from the waterbodies and the ocean surfaces along with very sparse presence of archaea, fungi, protozoa, and algae [92, 302, 361, 425, 553]. Splashing, bubble bursting, rain droplets, wave breaking, collapsing of bubble cavity, spume droplets, river water movement and flow are the various activities that are responsible for the release of bioaerosols from the waterbodies (Fig. 1) [12, 16, 92, 120, 124, 160, 254, 280, 323, 463, 493, 593, 608]. Also, Mayol et al. [376] has stated that ocean surfaces can exchange millions of microbes with atmospheric air i.e., millions of microorganisms/square meters of air every day and about 10% of which could thrive in the atmospheric air for longer time of even four days. This shows that meteorological conditions prevailing over ocean surfaces favor the survival of bioaerosols for longer periods compared to terrestrial surfaces.

# Role of anthropogenic activities on bioaerosols emission and dissemination

On the other hand, anthropogenic activities, or manmade activities (Fig. 1) like agriculture, solid waste management, wastewater treatment, waste transportation, biomass burning, reuse of solid waste, landfills, road dust, industrial activities, and waste discharge [14, 143, 171, 228, 283, 353, 355, 439] contribute immensely to the release of anthropogenic bioaerosols into the atmospheric air. Human, birds, and animals are said to expel and excrete pathogenic microbes and lots of dead cell debris in the bioaerosols that are released from them [47, 48, 83, 625]. Chen et al. [89] has stated that bacterial bioaerosols dominated the human populated region compared to fungal bioaerosols. Similarly, waste treatment plays an imperative role in the dispersal and dissemination of pathogenic microbes in the bioaerosols, which includes municipal wastewater/sewage treatment, composting, waste dumping yards, landfills, etc. [99, 142, 304]. Further, [99] has stated that actinomycetes and fungal bioaerosols are the majorly released bioaerosols from the composting site as most of bacterial species are killed due to the high temperatures prevailing in the compost. Also, [219] reported the predominance of mesophilic bacteria, psychrophilic bacteria, and microfungi in the bioaerosols collected from

a facility containing both wastewater treatment plant and a composting site. This is mainly due to the organic dust released from the composting facility which enables the specific growth of mesophilic and thermophilic species [274, 499]. Hence, wastewater treatment plants, composting facilities, and landfill sites are considered as potential anthropogenic source of pathogenic and non-pathogenic bioaerosols [70, 90, 176, 219, 407, 434]. However, [143] have identified that the animal-feeding operations and animal husbandry release high quantities of pathogenic bioaerosols that could have lethal effects on animals and human. Further, bioaerosols released in the indoor environment are also considered as anthropogenic bioaerosols as they are majorly released from human activities like day-to-day activities and also due to coughing, sneezing, talking, and breathing which could expel human respiratory and oral microbiota[221, 275, 417, 616], the skin microbes as a result of skin aberrations and shedding activities [175, 348, 421], and floor dusts rich in pathogenic microbes [243, 562]. On the other note, [99, 284, 430, 535] and [255] have reported the abundance of bacterial species like Staphylococcus aureus, Pseudomonas aeruginosa, and Escherichia coli in the bioaerosols collected from the industrial sites. Whereas E. coli was the predominant species reported in the bioaerosols released from the wastewater and waste treatment site [255]. Study reported by [455] stated that bacterial bioaerosols were the major bioaerosols released during the activity of land spreading of the semi-solid sewage sludge. Also, a study conducted by [138] and [80] explained the presence of bacterial and viral bioaerosols downwind the sewage sludge disposal site. Furthermore, agricultural practices were said to release considerable quantities of bioaerosols to the atmospheric air [333]. Though the bioaerosols released due to anthropogenic activities affect agricultural sector and various aspects of ecosystem, many studies till date elucidate and characterize the diversity and pathogenic properties of human and animal pathogenic bioaerosols. Crop pathogenic bioaerosols and their implication on the crop/agricultural health are less investigated till date.

### Influence of meteorological factors and aerodynamic particle size on bioaerosols aerosolization

Globally, many researchers have stated the significant role of meteorological factors and aerodynamic diameter in the prolonged persistence of bioaerosols in the atmospheric air [191, 287, 490, 558, 611]. Concurrently, the concentration of bioaerosols in the atmospheric air decides the transportation capacity of bioaerosols across continents and plays a pivotal role in microbial migration and colonization [124, 182]. Further, [276] have reported that meteorological factors greatly influence the aerosolization properties of bioaerosols. In specific, meteorological factors like relative humidity (RH), temperature, wind direction and speed, and precipitation are said to have major impact on the bacterial bioaerosols release and concentration in the atmospheric air and are reported to vary with seasons [124, 134, 140, 192, 193, 222, 304]. Accordingly, Miquel [391] has stated that fungal spore concentration and emission in the atmospheric air was also guided by seasonal variations and wind direction. Furthermore, it was observed that the movement of bioaerosols and their dispersal is majorly governed by the particle size (aerodynamic diameter) and the flux density [611]. These factors decide the dry and wet deposition properties of bioaerosol particles and influence their spread and transport over longer distances. It is also reported that the particles of high aerodynamic diameter settle down rapidly compared to a smaller particle [124]. Researchers worldwide have estimated that the aerodynamic particle size diameter of pollen grains could range between 17 and 58 µm [312, 539], fungal spores could range from 1 to 30 µm in diameter [213], bacteria could range from 1.25 to 8 µm [512, 567], viruses ranges  $\leq 0.3 \,\mu m$  [563] and all other biological particles like fungal segments, bacterial clusters, plant and animal segments could be present in varying size ranges [276].

The other two major factors influencing aerosolization of particles are bonding force and removal effect [276]. The balance maintained between these two forces acts as the driving force for the efficient aerosolization of any particle from the surface. Bonding force can be further defined as the electrostatic force created especially when the surface and the particles are differently charged which are mainly dependent on or influenced by the factors like temperature, moisture content, and the radiation balance [544]. On the other hand, it is the removal effect which is mainly guided by the aerodynamic force or drag developed due to the air movement, electric charge, particle impaction, inertia, and movement of surface away from the particles which are influenced by wind force, raindrop impaction, and physical agitations [542]. Further, the shape and structure of organisms present in the bioaerosols also decide the sedimentation, settlement, and dispersion rate of the bioaerosols [124, 125]. All these uncertainties like the seasonality, type of biological particles in the bioaerosols, life cycle of the biological particle, seasonal variations, aging, chemistry, and microphysics of the particles govern the dispersal and dissemination of bioaerosols in the atmospheric air [72].

# Microbe specific aerosolization and dissemination properties

Microbiota comprises of a variety of diverse microbial communities categorized into different taxonomic groups namely the algae, protozoa, slime molds, fungi, bacteria, archaea, and viruses with the major taxonomic levels classified being Kingdom, Division, Class, Order, Family, Genus, and Species [457]. Each group of which has their own characteristic physical, chemical, and genomic properties that differentiate them from each other [32, 457]. Aerosolization efficiency of these organisms are further dependent on and decided by their physical properties like shape, size, velocity in air etc. [124, 126, 130, 152, 285, 410, 414]. Bioaerosols containing these microorganisms and the number of microbes present in the bioaerosols are dependent on the surrounding environment and the meteorological factors and could even contribute to huge concentration per unit of surface area measured [337, 401, 622]. Archaea, being primitive organisms known till date are reported to be less observed in the bioaerosols as they are majorly present in extreme environments [501]. Nevertheless, the fact that they play a pivotal role in the Earth's biogeochemical cycles, their aerosolization properties are understudied due to the limitations existing in the efficient characterization of archaea. The only report available till date is their presence in the bioaerosols collected from composting sites as stated by [29, 395], and [568]. Similarly, algae and cyanobacteria were reported to get aerosolized from both the terrestrial and marine environments [124, 374, 375], but studies on their airborne quantitative measurements show that they are present in very low concentrations of ~ 300-500 cells/m<sup>3</sup> of the sampled air [478]. Globally, various researchers like [320, 468], Dubovik [141], Reisser [478, 514], and [416] have stated that Chlorophycean and Xanthophycean are the major species of algae that were observed in the bioaerosols and have also stated that their aerodynamic diameter ranged  $\leq 10 \,\mu\text{m}$  which favor their rapid aerosolization. Cyanobacteria being one of the successful groups of the microorganisms in the terrestrial habitat, are abundantly available in the marine surface contributing to the global carbon and nitrogen availability and exchange [124, 540]. Chroococcus limenticus, Lyngbya lagerheimii, and Schizothrix purpurascens are the well know species of cyanobacteria that were reported to be airborne [147]. Similarly, Phormidium fragile and Nostoc muscorum were reported in the Cairo, Egypt [147, 197].

Bacteria being one of the most abundant microorganisms present in the bioaerosols were reported to be emitted at the rate of 0.7–2.58 Tg yr<sup>-1</sup> [71, 242, 264]. The characteristic size range of the bacterial cells enable their easy aerosolization and they are often found as single cell as well as in agglomerates and clusters in association with other particles like soil, plant segments, dust, and other debris [59, 334]. Bacterial bioaerosols contain many bacterial pathogens that are lethal to human, plants, and animals and in addition to that they can act as biomarkers indicating the atmospheric, climatic, and environmental changes and anthropogenic influences [218, 356, 403, 404]. Further, [304, 309, 448, 610], and [356] have stated that meteorological factors, type of anthropogenic activities observed, seasonality, and the source materials present could highly influence the aerosolization properties, concentration, and diversity of bacterial species observed in the bioaerosols of the atmospheric air. [127], reported that bacterial concentration in the atmospheric air were found to increase with the increased temperature and wind speed. Release and aerosolization of bacterial particles can occur due to various activities like wind aerosolizing the bacterial particles from the substrates, sedimentation and settlement of particles from different layers of atmospheric air with higher concentration, aerosolization from plant surfaces, and sunlight shifting the electrostatic charge of the particles [276, 334, 570]. Firmicutes, proteobacteria (including alpha, beta, gamma, delta, and epsilonproteobacteria), Verrucomicrobia, Cyanobacteria, Acidobacteria, Planctomycetes and Chloroflexi are the major bacterial phyla reported till date in the atmospheric bioaerosols [163, 230, 304, 512]. Also, [125, 163], and [159] observed that the concentration and diversity of bacterial bioaerosols varied among the urban and rural regions with high concentrations of bacterial bioaerosols in the urban region. Further, [63], have reported that different land-use activities like agricultural fields, suburban areas, and forests release very high concentrations of bacterial bioaerosols of up to  $10^5$  to  $10^6$  cells/m<sup>3</sup> of air. Among the waterbodies, bubble bursting and sea sprays are the activities that liberate the bacterial bioaerosols from the marine environment to the atmospheric air [12, 324, 364] and it is also reported that bacterial bioaerosols liberated from the ocean surface has shorter life span compared to the one liberated from the land surface [71].

Likewise, the most predominant microorganism of all in the bioaerosols are the fungi, fungal spores, and the fungal segments which contribute to a load of about 8-186 Tg yr<sup>-1</sup> [148, 235, 242, 264, 511]. Fungal concentration in the atmospheric air and their aerosolization are highly dependent on meteorological conditions and maximum aerosolization of fungal particles are favored by temperature of 25-30 °C, relative humidity of 60–70%, and wind speed  $\leq 1$  m/second. Further, it was observed that fungal concentration in the atmospheric air reduces with increased wind speed of  $\geq 5$  m/ second [335]. [75] has reported that the maximum release of fungal spores in atmospheric air is high during the dry conditions i.e., during afternoon time when the temperature and wind speed are favorable for the spore release from the conidial chains. Similarly, some wet air spora like Nigrospora sp. are reported to release spores favorably during high humid conditions like mid-morning [557] and it was also reported that their spore concentration reduces with increased altitude. Further, [260, 313], and [467] have reported that the spore releasing mechanism of fungi is highly dependent on the surface tension and osmotic pressure. [351], stated that the plant pathogens like Cladosporium, Alternaria, Penicillium, Aspergillus, and Epicoccum sp. that could cause smut and rust disease of plants are the major species found in the atmospheric bioaerosols. Similarly, Ascomycota and Basidiomycota were found to be the dominant phyla observed in the bioaerosols and Basidiomycota to dominate marine environments [180, 183, 610]. Consecutively, viruses are the tiny airborne microbes in the bioaerosols that are as small as ~20 nm in size and are reported to be always attached to other suspended particles in the atmospheric air [135, 627]. Only limited studies are available on the virus bioaerosols persistence and existence in the atmospheric air. It was observed that the environmental factors like altered temperature, humidity, solar radiation, and UV index inactivate the virus particles present in the bioaerosols [124, 461]. Further, Hugh-Jones and Wright in 1970 [252], have stated that wind speed and humidity (over 60%) play a pivotal role in the dispersion and spread of foot and mouth viral disease. Similarly, spread of Aujeszky's disease (a rabies like disease of pigs) was found to be high during winter especially when the temperatures were 2-3 °C higher than the normally recorded temperature [96, 97]. Further, it has been reported that sprays from waterbodies acted as source of virus containing bioaerosols, their liberation, and dissemination [38-40]. Moreover, virus bioaerosols were also liberated from infected animals, humans, and birds [276, 634]. Likewise, high concentrations of virus bioaerosols were found to aerosolize from sewage treatment plant, sludge dumping site or treatment plants, and marine environment [79, 124, 182, 319, 517, 543]. However, the virus bioaerosols mass concentration in the particulate matter (PM) was always found to be in negligible quantities, though the concentration of virus bioaerosols were found to be high in certain conditions and activities as mentioned above.

# Major crop pathogenic bioaerosols and their implications in crop health and yield

Bioaerosols transmits several plant and crop pathogens causing various crop diseases hampering the agricultural production and yield creating significant economic losses worldwide [66, 73, 194]. Pathogenic bioaerosols gets sedimented and deposited as wet and dry deposition on the plants and crop surfaces during various stages of crop growth causing severe diseases and infections. Further, pathogenic bioaerosols settled and colonized on the soil acts as a vital source of seedling and crop infections. Fungi being the predominant crop pathogens by nature were reported to cause severe infections to a variety of crops worldwide. Bacteria being the second predominant microbial community causing various crop infection are followed by viruses, nematodes, etc. Phyllosphere of plants acts as an important niche for microbial colonization with a diverse lifestyle as epiphytes, saprophytes, and pathogens [41, 337]. Epiphytes are generally found to be present in the leaf surface and the pathogens like the foliar pathogens tend to ingress into the leaf tissues i.e., the intercellular space causing infections.

# Fungal crop pathogens- their mechanism of invasion and disease onset

About 19,000 fungal species have been identified till date to cause various crop diseases like leaf spot, rust, anthracnose, blight, wilt, scab, galls, coils, damping-off, mildew, cankers, rots, die-back, smut, warts, etc. [182, 265]. Spore dispersal in the environment is dependent on two major dispersal mechanisms like active [574] and passive dispersal [259] where active dispersal is enabled by forcible dispersion of the spores by fungi itself as a part of their life cycle and passive dispersal is through the bioaerosols, animals, insects, etc. As illustrated in Fig. 2a, the fungal spores settle on the leaf or plant surface by the dry and wet deposition of the fungal bioaerosols and starts invading the plants and crops causing various diseases. Fungal spores are generally categorized into two types i.e., non-motile and motile spores. Where in non-motile spores like the ascospores (sexual spores), urediniospores of rust fungi, sclerotia, conidiospores, oospores, and sporangiospores proliferate at the settlement site whereas the motile spores like the zoospores of chytrid fungi with flagella tend to move to a favourable location after sedimentation on to the leaf surface [215].

Survival of a plant pathogenic microbe is majorly dependent upon the proliferation, growth, and reproductive strategies adopted to exploit nutrient and environmental condition available in the host [20]. Fungal spore invasion into the plant tissue is facilitated by a sequence of proliferation stages that guides the process of germination and formation of appressorium which enables the development of the penetration hyphae that penetrates across the cuticle into the epidermis layer (Fig. 2a) [385]. After sedimentation on the leaf surface, the spores adhere to the plant surface with the help of the adhesion molecules present on the spore surface [7]. Apparently, a germination tube emerges from the spore utilizing the polyols like glycogens, trehalose, and sugar alcohols that are present in the fungal spores as energy source [328, 371]. Further the formation of the melanized appressoria enables the penetration of the appressorium peg into the cuticle with the turgor pressure developed in the appressorium [94, 214, 225, 229, 556, 585]. Penetrated hyphae grow between the epithelial cells (intracellular hyphae) in the intercellular space causing the infection utilizing the host nutrients [453] with the formation of various infective structures like hyphopodia [139, 432, 545], haustorium in the epithelial cells [384, 605] and infection cushions [123, 385] severing the infection.

Similarly, the notorious necrotrophic fungi which follows various phytopathogenic strategies for infecting and killing the crops by absorbing the nutrients available in the plant cells with the production of various cell wall toxins and degradation enzymes leading to various diseases like root rots, hypersensitive reactions, and alteration of the plant metabolism [226, 380]. They also suppress the host plant defence

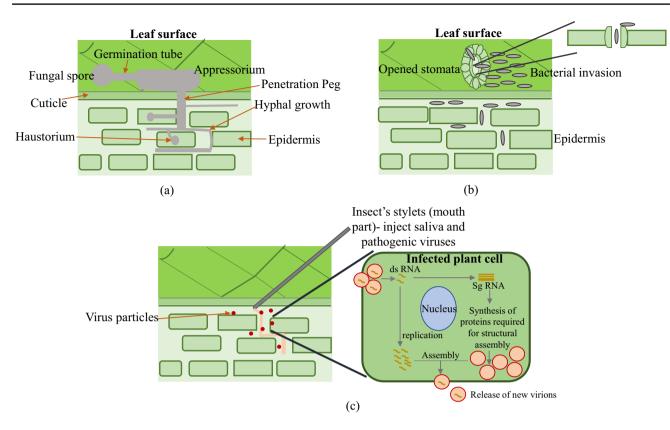


Fig. 2 Illustration of the various modes of invasion by the disease-causing crop pathogens: **a** fungal invasion of the leaf tissues; **b** bacterial invasion of the leaf tissues

mechanisms by manipulating the machinery, altering the plant gene regulating the membrane permeability, and by utilizing the amino acid present in the plant cell [549] aiding the fungal infection and disease progression.

#### Mechanism of bacterial invasion into plant tissues

Numerous bacterial pathogens causing various plant diseases have been discussed by various researchers worldwide [360]. They are mostly passive invaders and enter the plant tissues through all the possible opening on the plant surface like the stomata, rhizoplane (expanding root points), base of trichomes, grooves along the veins, hydathodes, nectarthodes, lenticles in stem and roots, and through cut wounds and scars [34, 247, 322, 336, 337]. Among all the discussed passive openings in the plants, bacterial phytopathogens use stomata as their common entry point as stomata provides access to substomatal chamber and into the mesophyll [371] (Fig. 2b).

Stomatal movement (opening and closing) in the plants are majorly controlled by the plant's immune system by developing a turgor pressure in the guard cells [441]. Further, environmental factors like draught, stress, light intensity, UV intensity, relative humidity, and the concentration of carbon dioxide in the atmospheric air influence the activity of guard cells affecting the stomatal movement [161, 405, 502]. In order to overcome all these barriers, bacterial pathogens alter the plant immune system with the secretion of hormones like jasmonic acid, coronatine (secreted by *Pseudomonas syringae*) promoting the stomatal opening favouring bacterial entry and colonization in the intercellular region [172, 202, 383]. Apparently, they develop various mechanisms to adopt to the relatively unfavourable conditions prevailing in the leaf surface and intercellular spaces with the development of a microenvironment favouring their survival in the area of colonization [69, 257]. Additionally, bacterial pathogens exist in aggregates and biofilms to resist and overcome the unfavourable conditions prevailing during their colonization in the leaf surface and intercellular space as well [110, 396, 397]. They also secrete various enzymes and proteins to adsorb the nutrients available in the plant tissues causing infections [121].

#### Invasion of viral phytopathogens

Unlike other phytopathogens, viral phytopathogens spread through insect vectors, nematodes, and also through bioaerosols especially the pollen grains play a vital role in carrying the viruses [195, 372]. Insect bite, bioaerosols transmitting the infected plant sap and debris, carry over from generation to generation, and the invasion of nematodes carrying the virions are the major entry routes of virus into plant tissues [68, 78, 253, 497]. Similarly, contaminated seeds and soil are also among the potential sources of phytopathogenic viral transmission and infection [371]. Generally, viruses alter the host mechanisms by upregulating or downregulating the host proteins in order to select the host cells in the plant tissues which favours their infection, replication, and pathogenicity [509]. Viruses use the plants cells as their replication factories by further altering the inner membrane and mechanisms of the host cell. Figure 2c elaborates the viral invasion and replication inside the plant cells. The infected cells generate multiple copies of viruses by promoting the replication of sub-genomic viral RNAs, and the replication of the whole genomic viral RNAs which are then packed into the capsid synthesized by the expression of the subgenomic RNAs [272, 301]. Generally, viral infection spread form cell to cell transmission of the viruses, and also by the transmission of the infected sap and debris by the insects and bioaerosols. Further, infected soil and the invasion of nematodes carrying the viruses from the infected soil also play a vital role in spreading the viral infection.

Further, the evolutionary factors like the genetic drift, gene flow, natural selection, mutation, and recombination enabling the evolution of resistant and virulent variants makes it a vital pathogen hampering the crop yield and growth [187, 190, 300]. Henceforth, the disease management strategies are majorly focused on the prevention of phytopathogenic viral infections by restricting the entry of viruses into plants and development of resistant varieties rather than its post-infection cure [488].

### Pests and vectors using air as a medium to infect crops

#### Nematodes as pest of crops

Nematodes are the microscopic organisms that can spread through air and water, especially their eggs have the tendency to get aerosolized in air. They are the most devastating pests of crops causing a global loss in agricultural sectors to about 80 billion USD [278, 419]. Researchers have identified about 4100 different species of plant pathogenic nematodes worldwide [118]. These nematodes also act as carriers of various crop pathogenic microorganisms especially the viruses and help in spreading the infections to crops [371].

Nematodes are categorized into three types namely ectoparasites, semi-endoparasites, and endoparasites. Generally, ectoparasitic nematodes dwell outside the host and benefit from the host with the long feeding style inserted into the plants which enables their nutrient absorption. Whereas the semi-endoparasitic nematodes partially enter the roots of the crops for feeding and the posterior end of their body is kept intact in the soil. Endoparasitic nematodes completely dwell inside the roots and other parts of the crops feeding on the internal tissues following a migratory or sedentary lifestyle. Migratory endoparasites feed on the root cells immediately after entering the roots and migrate to other parts of the crops causing a severe damage to the plant tissues and cell death [154, 531, 566]. Similarly, sedentary endoparasites enter the vascular cylinder of the plants and causes redifferentiation of the host cells into multinucleate cells for reproduction and the hypertrophic feeding cells [119, 278, 440, 525]. Researchers like [113, 199, 207, 236, 525], and [381] have reported that endoparasitic infections are initiated with the entry of juveniles near the root tips. Further, they migrate intercellularly to the vascular cylinder to enable the formation of feeding cells by the degradation of the cell walls of the plant cells, inhibiting the anti-nematode enzymes secreted by the plants, suppressing the immune system, and enhancing the secretion of the proteins required for the formation of the feeding cells. They also induce the formation of the multinucleate giant cells by the repeated nuclear division at the favourable locations where the cytoplasmic division is almost absent [1, 157]. However, many parasitic nematodes are reported to have complex interaction with their host plants to receive a rich and continuous food source [584]. Root nematodes feed exclusively on the root tissues whereas, the aerial nematodes migrate aerially to the stem and leaves and feed on the bulbs, foliage and the stem [314, 579]. Sedentary root nematodes are known to cause devastating infection compared to the migratory nematodes and this is mainly enabled by the specific exploitation of the host immune responses altering the host defence mechanism [53, 278, 314].

#### Insect as pests and vectors

Insects are the six-legged small invertebrates of the *Arthrop-oda* phyla that mostly feed on plants and crops for their nutrition. For example, bees feed on nectar and pollen from plants, larvae of many insects like beetles, moths, and flies live on plants and crops feeding on leaves and plant parts, and bugs thrive on plants saps as their major source of nutrients. Almost all the four stages of metamorphosis of insects are known to involve plants as their habitat. Among which, caterpillars are the most important stage of the insect's life cycle which causes significant damage to the crops and plants. Similarly, they act as vectors transmitting various fungal, bacterial, and majorly virus pathogens that could cause severe infections to crop impeding the crop yield.

General plant injuries caused by the plant feeding insects include (i) consumption of the infested parts (leaves, stems, roots, and flowers) of the plants by chewing, (ii) pit feeding of leaves by caterpillars, beetles, and flea beetles, (iii) edge notch of leaves caused by weevils, grasshoppers, large caterpillars, and katydids, (iv) semi-circular cut causes by cutter bees, (v) leaf mining by beetles, flies, sawflies, and moths, (vi) stem boring by long-horned beetles, and (vii) metallic wood boring beetles, engraver beetles, clearwing moth, American plum borer, and moths. Similarly, root chewing insects like weevils and root maggots, and sap feeding insects like aphids, leafhoppers, thrips and scales also cause major damage to plants. Oviposition damage is also a serious problem caused by insects when the insects lay their eggs in deep tissues of plants especially in the stems [105, 449].

Transmission of vector-borne pathogens like bacteria and viruses are enabled by the colonization of the pathogens through insects in the plants. Plants vascular systems help in the transportation and colonization of the pathogens to various parts of the plants. Further, phloem favours the transportation and nutrient supply required for the pathogens with the help of the nutrient rich sap and the sieve elements available in them [60, 343, 346, 606]. Despite the low nutrient contents available in the xylem some pathogens have also been observed to colonize the xylem [28, 454, 473]. Along with the pathogens many insect pests like whiteflies, aphids, psyllids, and leafhoppers were also reported to acquire their nutrients from the xylem and phloem of the plants [454]. Purcell [472] has stated that the specialized mouth parts of these insects enable their penetration to the epidermis to reach the preferred locations like the mesophyll and vascular system involving phloem and xylem. This activity favours the transportation of the pathogens present in the phloem and xylem of the infected plants through their body parts making them a potential vector for transmitting diseases to healthy plants [411, 433].

# Major crop diseases and their global implications

Generally, crop/plant diseases are majorly caused by biotic factors supported by the abiotic factors. The factors influencing the crop diseases are classified into three major categories as the abiotic, biotic, and the meso-biotic factors [588].

#### Disease caused by abiotic factors

Abiotic factors are the reason for the general deficiencies of the crops and the associated diseases. They are caused by malnutrition including minerals and ions, impaired soil conditions like type and fertility, relative humidity, temperature, reduced or excess light source, reduced and excess water availability, wind or aeration conditions, concentration of the  $CO_2$  available, effect of impurities carried by the aerosols, and by the presence of toxic compounds in the soil and air [136, 246, 305, 588]. The major diseases caused by abiotic factors include chlorosis, stunted growth, interveinal chlorosis, purplish-red colouring, necrosis, black tip of mango due to  $SO_2$  toxicity, whiptail of cauliflower crops caused due to molybdenum deficiency, khaira disease of rice caused by zinc deficiency, hollow and black heart of potato caused by excessive usage of  $CO_2$  during postharvest storage, and apple bitter pit caused due to calcium deficiency [459].

#### Disease caused by biotic factors

Crop diseases caused by biotic factors are the diseases caused by various crop pathogenic species belonging to the kingdom Fungi, Chromista, Monera, Animalia, etc. Bioaerosols are considered as the key factor in the spread and dissemination of the disease-causing pathogens to the crops worldwide [124, 183]. Along with bioaerosols, various environmental factors are also involved in influencing the sedimentation, survival, and penetration or ingress of the pathogens into the crop tissues [484, 550, 588]. Moreover, these factors influence the onset of disease symptoms and progression of disease in the crops. Table 1 describes the details of the specific/optimum temperature, relative humidity, water activity, light intensity, and nutritional requirement of various pathogens causing major crop diseases. Temperature being an important meteorological parameter which plays a pivotal role in deciding the weather conditions especially the four different seasons, directly influences the susceptibility of the crops to infections, influences the pathogen survival, transport, germination on the host as well as source, and also alters the visual disease symptoms in the crops [201, 248, 459]. Similarly, relative humidity observed and the water activity of the soil like the dry and wet conditions alters and triggers the disease onset, progression, severity, and dissemination [101]. It is reported that dry soil and a high relative humidity (almost > 85%) substantially enhances the disease onset, severity, and spread compared to the wet soil and low relative humidity levels [108]. Also, pathogen invasion, survival, and onset of disease symptoms were reported to be dependent on the CO<sub>2</sub> concentration in the atmospheric air and the light availability. It is also estimated by the researchers worldwide that the elevated  $CO_2$ concentration and increased light availability including the long day-length period helps in promoting as well as controlling the infections [145, 296, 341, 524, 587]. Further, soil conditions like the pH of the soil and the nutrients available (nitrogen, calcium, potassium, phosphorus, zinc, manganese, molybdenum, and microelements) influences the pathogen entry into the plants through the roots [7, 465]. Table 2 elaborates the symptoms, details of the susceptible crops, and the pathogen responsible for the various crop diseases reported worldwide.

It is well evident from numerous studies worldwide that leaves are the main organs of interest for the crop pathogens as it provides the required space for its settlement and entry into the plant's inner tissues [371]. Powdery mildew,

Table 1	Details of the s	pecific and optimum	n range of abiotic fac	ctors required for the	e crop pathogenic infections
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Disease caused	Pathogen	Optimum tem- perature, °C	Optimum rela- tive humidity, %	Water activity	Light intensity	CO <sub>2</sub> concentration	Reference
Onion smut disease	Urocystis cepule	10 -12	-	Dry soil	-	-	[17]
Yellow dwarf virus disease of barley	Yellow dwarf virus	16	-	-	-	Low CO <sub>2</sub> con- centration	[238]
Cyst nematode of potato	Globodera pal- lida	15	-	-	-	-	[377]
Bunchy top of bananas disease	Banana bunchy top virus (BBTV)	25	-	-	-	-	[111, 327]
Seedling death of cereals disease	Fusarium nivale	-	-	Dry soil	-	-	[101]
Downey mildew of rose plants	Peronospora sparsa	15-20	85-100	Low moisture in soil	-	-	[491]
Early blight of potato	Alternaria solani	28-30	-	-	-	-	[597]
Late blight of potato	Phytophthora infestans	12-20 and above	-	Low moisture in soil	High light intensity	-	[7]
Bacterial leaf blight of rice	Xanthomonas oryzae	27	-	-	-	-	[204]
False smut of rice	Ustilaginoidea virens	20	-	-	-	-	[162]
Papaya ring spot	Papaya Ring Spot Virus (PRSV)	26-31	-	-	-	-	[575]
Black rust of wheat	Puccinia graminis	25-30	>85	High leaf wet- ness	Darkness	Elevated CO <sub>2</sub> concentration	[248]
Stripped rust of wheat	Puccinia stri- formis	18-30	>85	High leaf wet- ness	Low light inten- sity	Low CO <sub>2</sub> con- centration	[248]
Loose smut of wheat	Ustilago nuda var, tritici	16-22	-	-	-	-	[613]
Brown spot of rice	Hel- minthosporium oryzae	28-30	-	-	-	-	[31]
Sheath blight of rice	Rhizoctonia solani	28-32	-	-	-	-	[519]
Damping off	Pythium sp., Phytophthora sp., Rhizoc- tonia sp., Fusarium sp., Sclerotia sp.	< 24	-	Low moisture in soil	-	-	[7]
Club root of cab- bage	Plasmodiophora brassicae	18-25	-	Wet acidic soil with 70-80% moisture	-	-	[50]
Leaf spot of cole crops	Alternaria bras- sicicola	28	-	High leaf wet- ness	-	-	[248]
Downy mildew of cole crops	Peronospora parasitica	12-27	-	Low moisture in soil	-	-	[7]
Downy mildew/ crazy top of maize	Peronoscleros- pora sorghiis	21-33	-	Low moisture in soil	-	-	[7]
Anthracnose of citrus	Colletotrichum acutatum	High tempera- tures	-	-	-	-	[290]

#### Table 1 (continued)

potato

Disease caused	Pathogen	Optimum tem- perature, °C	Optimum rela- tive humidity, %	Water activity	Light intensity	CO <sub>2</sub> concentration	Reference
Bunt of wheat	Tilletia contro- versa	High tempera- tures	-	-	-	_	[55]
Root knot of coffee	Meloidogyne incognita	High tempera- tures	-	-	-	-	[200]
Wilt of tomato	Ralstonia solan- acearum	-	>75	High moisture in soil	-	-	[262, 306]
Rice blight	Pyricularia oryzae	-	>85	-	-	-	[24]
Wheat blotch	Septoria tritici	-	-	-	High light intensity of 8000 lux	-	[572]
Bean rust	Uromyces pha- seoli	17-22	>95	-	Both high and low intensity	-	[392]
Downy mildew of grapes	Plasmopora viticola	12-30	>85	Low moisture in soil	Long day light	-	[321]
Turnip mosaic infection	Turnip mosaic virus	High tempera- tures	Less humidity	Less moisture	-	-	[465]
Bunch rot of grapes	Botrytis cineria	20-25	100	-	-	-	[98]
Crown rot of wheat	Fusarium pseu- dogramine- arum	-	-	-	-	Elevated CO <sub>2</sub> concentration	[382]
Smut of barley	Ustilago hordei	-	-	-	-	Elevated CO <sub>2</sub> concentration	[359]
Downey mildew of barley	Blumeria graminis	-	-	-	-	Low CO <sub>2</sub> con- centration	[237]
Leaf spots caused by foliar nematodes	Aphelenchoides ritzemabosi	22	High humidity	High moisture	-	-	[84]
Early blight of tomatoes and potatoes	Alternaria solani	28-30	Humid envi- ronment is required	Alternate dry and moist condi- tions	Day light is essential for development	-	[86]
Smut disease of sugarcane	Sporisorium scitamineum	25-30	65-70	-	-	-	[475]
Wart disease of	Synchytrium	10-27	-	Moist soil with	-	-	[426]

rainfall exceeding 70 cm

downy mildew, and leaf spots are the prominent leaf diseases of the crops, and the causative agents include fungi, bacteria, and nematodes causing characteristic symptomatic disease which on progression leads to altered crop growth and reduced yield. Powdery mildew is the fungal pathogenic infection of crops caused by various fungal species affecting a wide range of crops like fruits, vegetables, cereals, and other common and ornamental plants (Table 2). These fungi invade the epidermis layer of the leaves immediately after the sedimentation of the spores on the leaf surface by supressing the host immune response [211]. This causes the green island effect on the infected leaves i.e., the leaf area of the fungal colonization remains green whilst the surrounding area has chlorosis leading to the systematic cell death and formation of nutrient sinks

endobioticum

promoting monosaccharide transport at the site of infection [174]. Which enables the utilization of the monosaccharide by the fungi favouring the fungal colonization with the formation of the characteristic symptomatic white powdery growth on the leaves surface that leads to about 30% loss in the crop yield [249, 548]. Downy mildew is caused by *Oomycetes* [104] that hampers the crop quality and yield worldwide and have caused severe losses during 1970s of about 7.5 billion USD in USA as stated by USDA [580, 582]. They have a broader host range compared to the powdery mildew, almost infecting all the crops with the characteristic yellow lesions on the leaves (Table 2) [491]. Xu and Pettitt [620] has reported that the symptomatic lesions formed on the leaves, stem, peduncles, calyxes, and petals by downy mildew are very difficult

Crop disease	Symptoms	Crops affected	Pathogen involved	Reference
Blight disease	Blight is caused by bacterial or fungal infestation with characteristic symp- toms like sudden and severe yellowing, browning, spotting, withering, and drying of leaves, flowers, fruits, twigs, sheath, stem, and the whole plant	Potato, com, chestnut, citrus, fruit, rice, wheat, tomato, grass, oak, grains, pulses, cereals, vegetables, etc.	Fungi: Phytophthora infestans, Cochliobo- lus heterostrophus, Bipolaris maydis, Cryphonectria parasitica, Alternaria sp., Alternaria triticina, Tubakia iowen- sis, Colletotrichum capsici, Phytophthora parasitica, Cochliobolus heterostrophus Bacteria: Erwinia amylovora, Xanthomonas oryzae, Burkholderia plantarii, Xan- thomonas axonopodis	[133, 234, 358, 422, 451, 452, 480, 500]
Rot disease	Rot disease is caused by bacteria, fungi, and Oomycetes. This affects the roots, basal or bulb rots, heart rot, grey rot, seed rot, soft rot, red rot, tuber rot, and wood rot. It is characterized by plant decomposi- tion and putrefaction with a hard, dry, spongy, watery, mushy, or slimy tissue decay	Howers, fruits, vegetables, wood, celery, barley, bean, carrot, faba bean, lettuce, oat, oilseed, pea, rye, soybean, sugar beet, tomato, triti- cale, wheat, alfalfa etc.	Fungi: Fusarium avenaceum, Fusarium cul- morum, Fusarium graminearum, Fusarium pseudograminearum, Fusarium solani, Fusarium verticillioides, Phoma betae, Phoma terrestris, Phoma sclerotioides, Thielaviopsis basicola Oomycetes: Aphanomyces cochlioides, Aph- anomyces euteiches, Phytophthora citroph- thora, Phytophthora citroph- thora, Phytophthora citroph- thora cactorum, Phytophthora cactorum, Phytophth	[36, 43, 91, 115, 155, 156, 223, 241, 330, 573, 607, 621]
Wilt disease	Wilt is caused by bacterial, fungal and nematode pathogen. Affects the vascu- lar system, alters the xylem activities by hindering the transport of water to different parts of plants. Leads to rapid death of plants and trees	Cucurbits, elm trees, mimosa, oak, persimmon, pine, Stewart's wilt of corn, chillies, potatoes, tomatoes, aubergine, banana, geranium, ginger, tobacco, sweet peppers, olives, cotton	Fungi: Ophiostoma ulmi, Fusarium oxyspo- rum, Bretziella fagacearum, Acromonium diospyri, Verticillium sp. Bacteria: Erwinia tracheiphila, Ralstonia solanacearum, Pantoea stewartia Nematode: Bursaphelenchus xylophilus	[114, 131, 153, 177, 369, 513, 518]
Rust disease	Fungal disease that affects the eco- nomically important crops. Appears as yellow, orange, red, brown, and black powdery pustules on the leaves, stems, young shoots, and fruits causing leaf rust, stem rust, stripe rust, crown rust, blister rust, apple rust, coffee rust, bean rust, leek rust, etc.	Wheat, rye, coffee, gooseberry, cere- als, cedar- apple, grasses, crabap- ple, white pine, asparagus, pine, bean, chrysanthemum, hollyhock, snapdragon, sugarcane, ornamental plants	Fungi: Puccinia graminis, Puccinia strii- formis, Puccinia recondita, Puccinia sorghi, Phakopsora pachyrhizi, Melampsora epitea, Melampsora lini, Cronartium ribicola, Gymnosporangium juniperi-virginianae, Hemileia vastartix, Phakopsora meibomiae, Puccinia coronata, Puccinia henerocallidis, Puccinia triticina, Uromyce appendicu- latus, Puccinia melanocephala, Puccinia Puccinia telanocephala, Puccinia Puccinia telanocephala, Puccinia	[151, 256]

Table 2 (continued)				
Crop disease	Symptoms	Crops affected	Pathogen involved	Reference
Smut disease	A fungal disease identified with their dirt like sori which is dark coloured, thick-walled structures containing dust like teliospores. Sori generally appears in the leaves, flowers, stems, bulbs, and seeds. Commonly known as covered smut, loose smut, false smut	Rice, sugarcane, maize, barley wheat oats, forage grass, potato	Fungi: Ustilago esculenta, Sporisorium scitamineum, Ustilago maydis, Ustilago nuda, Ustilago tritici, Thecaphora solani, Ustilago virens	[18, 51, 269, 530]
Canker disease	Cankers are caused by both fungal and bacterial pathogen. Generally, occurs on woody trunks with the characteristic lesions with irregu- larly sunken, swollen, flattened, cracked, discoloured, and dead areas on the stem, twigs, and trunk	Sweet and sour cherries, tomatoes, strawberries, kiwi fruit, hazelnuts, olives, apple, stone fruit, almond, blueberry, chestnut, apricot, citrus fruits, butternut, rapeseed, rose, oak, mulberry	Fungi: Valsa mali, Neonectria galligena, Sirococcus clavigignenti-juglandacearum, Seiridium cardinale, Geosmithia puterilli, Discula destructiva, Eutypa lata, Thy- romectria austro-americana, Lachnellula willkommii, Gibberella baccata, Diplodia quercina, Fusarium circinatum, Apiogno- monia veneta, Leptosphaeria maculans, Leptosphaeria coniothyrium, Crypto- sporella umbrina, Gremmeniella abietina Bacteria: Pseudomonas syringae, Clavibac- ter michiganensis, Xanthomonas camp- estris, Xanthomonas citri, Xanthomonas axonopodis, Pseudomonas savastanoi, Xanthomonas populi	[64, 74, 165, 168, 178, 179, 212, 316, 481, 505, 506, 599, 609]
Blast disease	Fungal and bacterial disease that affects grains, leaves, leaf collars, nodes, panicles, and seedlings. It forms, elliptical leaf spots with grey, white centre and red to brown margins	Rice, wheat, millets, citrus, rye, bar- ley, cereals, apple	Fungi: Magnaporthe oryzae, Hel- minthosporium sativum, Magnaporthe grisea Bacteria: Pseudomonas syringae	[196, 232, 263, 626]
Leaf spot disease	Caused by bacteria, fungi, viruses, nematodes, and insects which can lead to cankers and blights. It is a light-coloured circular spot with brown coloured edge	Olives, legumes, vegetables, cereals, pulses, fruits, almost all crops are affected	Fungi: Phoma sp., Sclerotinia sp., Alternaria brassicae, Cercosporidium personatum, Cercospora dolichi, Pseudocercospora griseola, Phyllosticta sp., Colletotrichum gloeosporioides Bacteria: Xanthomonas campestris, Pseu- domonas savastanoi, Curtobacterium flac- cumfaciens, Xanthomonas begoniae Virus: Trichovirus, Cocadviroid Nematodes: Aphelenchoides ritzemabosi Insects: Aphids, leafhoppers, lace bugs, plant bugs, mites,	[23, 84, 233, 297, 318, 332, 435]

Crop disease	Symptoms	Crops affected	Pathogen involved	Reference
Downy mildew disease	Majorly caused by Oomycetes. Symptoms include the appearance of yellow to white patches on the leaf surface especially due to the growth of white to greyish cotton like fungi. With time the infected leaves turn crisp, brown, brittle, and fall off	Cucumber, cantaloupe, pumpkin, squash, watermelon, basil, grapes, hops, ornamental plants, soyabeans, spinach, sunflower, rose plants, blackberries, caneberries, and almost all crops are susceptible	Fungi: Peronospora belbahrii, Pseudoper- onospora cubensis, Plasmopara viticola, Pseudoperonospora humuli, Peronospora manshurica, Peronospora effuse, Plas- mopara halstedii, Peronospora sparsa	[321, 471, 491, 631]
Powdery mildew disease	A fungal disease with characteris- tic spots or patches with white to greyish powdery appearance on the leaves	Affects all plants including fruits, vegetables, cereals, grasses, weeds, shrubs, roses, creepers. Mainly infects wheat, barley, grapes, leg- umes, onions, strawberries, apple, pears, melons, gourds, pumpkins	Fungi: Blumeria graminis, Microsphaera dif- fusa. Erysiphe necator, Leveillula taurica, Podosphaera leucotricha, Podosphaera xanthii, Erysiphe cichoracearum, Leveil- lula taurica, Microsphaera syringae, Podosphaera aphanis, Sawadaea tulasnei, Erysiphe berberidis, Golovinomyces orontii	[2, 387, 474, 494]
Anthracnose disease	A common fungal disease of leaves with the characteristic symptomatic spots, cupping, and curling of leaves leading to blight and cankers. Also affects stems, fruits, and flowers	Tomatoes, melons, chillies, oak, maple, ash, avocado, cashew, mango, banana, vegetables, cucum- ber, bean, pumpkin, spinach, almost all plants grown in shade	Fungi: Colletotrichum sp., Gloeosporium sp.	[116, 565]
Gall disease	Formation of round and wart-like growth on the lower stems and branches mostly near the soil line	Woody plants, shrubs, grapes, rasp- berries, blackberries, roses	Bacteria: Agrobacterium tumefaciens, Agro- bacterium rubi, Agrobacterium vitis	[33, 205]
Wart disease	Otherwise known as black wart, black scab, potato turnour, potato cancer or canker, cauliflower disease, warty disease with symptoms like forma- tion of green parenchymatous tissues on stems base, stolon buds, and tuber eyes which may turn black on maturation	Potatoes	Fungi: Synchytrium endobioticum	[426]
Scab disease	Twisted and puckered leaves, black circular scabby spot on the under- side of the leaves, spot looks velvety, sooty, olive-green appearance on the upper side, finally the leaves turn yellow and drop down	Apple, potato	Fungi: Venturia inaequalis Bacteria: Streptomyces scabies, Streptomy- ces bottropensis, Streptomyces stellisca- biei, Streptomyces aureofaciens	[203, 495]
Damping off disease	It is soilborne disease. Rotting of stem and root tissues, leaf spotting, decay of seeds and seedlings	Affects all agricultural and forestry crops	Fungi: Rhizoctonia solani, Aphanomyces cochlioides, Pythium sp., Phytophthora sp., Botrytis sp., Fusarium sp., Cylindrocladium sp., Diplodia sp., Phoma sp., Alternaria sp.	[315]

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Crop disease	Symptoms	Crops affected	Pathogen involved	Reference
Mosaic disease	Discoloration of leaves, mottled leaves, yellow spots and stripes on the leaves, twisting of young leaves, stunted growth, poor leaf and fruits formation, reduced photosynthesis, and reduced grain production	Tobacco, pepper, potato, tomato, egg- plant, cucumber, petunia, melons, squash, spinach, celery, beet, cere- als, wheat, sugarcane, lettuce, maize, and other plants	Virus: Tobacco mosaic virus, cauliflower mosaic virus, sugarcane mosaic virus, let- tuce mosaic virus, maize mosaic virus Transmitting insect vectors: insect, aphids, leafhoppers	[87, 338, 344, 446, 601]
Leaf roll disease	Severely affected crops appear stunted and erect with leaves rolling upwards and appear leathery, chlo- rotic, and die eventually. Necrotic lesions are observed in the tuber tissues of potatoes	Grapes, potato	Virus: Grapevine leafroll associated virus, potato leafroll virus Transmitting insect vectors: Aphids	[307, 363]
Leaf curl disease	Upward and downward curling of leaves with thickening of leaves	Cotton, Papaya, Okra, Chilly, Capsi- cum, Tomato, Tobacco	Virus: Leaf curl virus Transmitting insect vectors: White flies	[362]
Stunt disease	Discoloration and distortion of leaves with stunted growth. Vulnerable to secondary infections	Peanuts	Virus: Peanut stunt virus Transmitting insect vectors: Aphids and sap insects	[408, 443]
Root knot disease	Formation of root-knot galls that results in stunted growth, poor qual- ity and yield of crops, discoloration and nutrient deficiency, and reduces resistance to draught conditions and secondary infections by other pathogens	All horticultural and field crops	Nematode: Meloidogyne javanica, Meloi- dogyne arenaria, Meloidogyne incognita, Meloidogyne hapla	[268]
Cyst disease	Stunted growth, yellow discoloration, and death	Soyabean, peas, sugar beet, potato	Nematode: Heterodera schachtii, Heterodera goettingiana, Heterodera glycines, Glo- bodera pallida	[245, 377]
Lesion disease	Plant root stress with necrotic lesions. Lesions turn from reddish-brown to black. Initially appear as spotty lesions and coalesce to form larger lesions with time	Over 400 crops species including potato, peanut, monocots, fruits, cereals	Nematodes: Pratylenchus pratensis, Pratylenchus brachyurus, Pratylenchus coffeae, Pratylenchus penetrans, Pratylen- chus scribneri, Pratylenchus vulnus, Pratylenchus zeae, Pratylenchus crenatus, Pratylenchus thornei	[394]
Sting disease	Ceases the growth of the root tips leading to reduced strength, slow growth, stunted growth, and even- tual death of crops. Foliage appears nutrient deficient with yellow to red discoloration which leads to chronic wilting	All agricultural crops, vegetables, grains, and trees	Nematode: Belonolaimus longicaudatus	[310]

Table 2 (continued)

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Table 2 (continued)

Crop disease	Symptoms	Crops affected	Pathogen involved	Reference
Chloric dwarf disease	Depending on crop variety and environmental conditions it causes stunted growth, reduced internode length, foliar irregularities, chloro- sis, necrosis, purpling of leaflets, downward curling of leaves	Tomato	Viroid: Tomato chloric dwarf viroid - <i>Pospi</i> - [591, 592] <i>viroid</i>	[591, 592]
Fruit crinkle disease	Russet ring symptom, leaf pucker symptom, and appearance of green crinkles	Apple	Viroid: Apple fruit crinkle viroid - Apscavi- roid	[329]
Citrus exocortis disease	Affects trees and plants with the characteristic symptoms like scaling of bark, severe chlorosis, and stunted growth	Citrus, tomato	Viroid: Citrus exocortis viroid - Pospiviroid	[600]
Columnea latent disease	Stunted growth, distorted and vein necrosis. Necrosis observed in stem, branch, and leaves	Tomato, cucumber, potato, eggplant, capsicum, petunia, ornamental plants	Viroid: Columnea latent viroid - Pospiviroid	[560]
Tomato apical stunt disease	Stunting, apical proliferation, apical leaf narrowing and yellowing, leaf crinkling, tissue brittleness, necrosis	Tomato	Viroid: Tomato apical stunt viroid - Pospi- viroid	[21]
Tomato planta macho disease	Stunted growth, exhibit strong epi- nasty in leaves, and produce fewer fruits	Tomato	Viroid: Tomato planta macho viroid - Pospi- viroid	[372]
Potato spindle tuber disease	Stunted growth, chlorosis, reddening or purpling of leaves, brittle leaves, waviness or distorted leaf margins, significant reduction in the quality of fruits and tubers	Potato, tomato, ornamental plants	Viroid: Potato spindle tuber viroid - Pospi- viroid	[520]

to differentiate from the lesions formed due to nutritional deficiencies, spray injury, and black spots [85]. Further, researchers have stated that the yellowing and abscise formation on the leaves are the severe symptoms observed on the leaves surface followed by enlargement of abscise, appearance of black spots, lesions on the petals, leaf fall in about one month period of infection, arrest of root formation, and collapsing of buds [6, 85, 186, 620]. Similarly, leaf spots affecting almost all the crops and plants are caused by various species of fungi, bacteria, and nematodes as describe in Table 2. Fungal leaf spot disease occurs when the fungal spore settles on a wet leaf surface on a warm weather which enables the immediate sporulation and formation of characteristic irregularly edged spots measuring about 0.5–1.5 cm demarcated with black edges, brown to reddish spot, and yellow halo. Brown leaf spot disease caused by the fungal pathogens such as Cercospora henningsii, Cercospora manihotis, and Cercospora vicosae were reported to cause a yield loss of about 20-30% in a wide range of crops [555]. While the white spot causing fungal species like Cercospora caribaea, that are found mostly in lower temperatures are known to cause considerable defoliation in casava plants [614]. Consequently, bacterial leaf spots appear as water-soaked lesions of 0.6 cm in diameter with characteristic black colour. They develop only during cool and wet conditions especially during rainy seasons or as sprinkler effect in the fields using sprinkler irrigation methods. Xanthomonas sp. and *Pseudomonas* sp. are the predominant pathogens found to cause bacterial leaf spots in many economically important crops hampering their quality and yield [518]. Unlike the common root nematodes, the foliar nematodes like Aphelenchoides ritzemabosi, Aphelenchoides besseyi, and Aphelenchoides fragariae have distinguishing properties of feeding on the leaves. Infection generally occurs when the nematode crawls up to the leaves from the soil or carried by wind and water film to the leaf surface, where they enter the leaf surfaces through stomata and cause progressive diseases as they feed on the parenchymal cells. The major symptoms include the formation of 0.25-1 cm necrotic lesions on the leaf surface near the vein especially occurrence of stripe necrotic lesions on the leaves with parallel veins and angular necrotic lesions on the leaves with netted veins. With time the stripe lesions spread to most of the leaves causing leaf blotches and the angular lesions progress to fan shaped pattern with the characteristic yellow-brown-grey colour. Severe diseases cause chlorosis of the crops with stunted growth [84, 350].

However, blight, anthracnose, wilt, blast, rust, and smut are also some of the major crop/plant diseases caused by the pathogens that target the leaves as their mode of colonization and entry into the plant's inner tissues. Blight and anthracnose disease of crops are caused by both bacterial and fungal pathogens (Table 2) which leads to the sudden spotting, drying, and withering of leaves, flowers, twigs, sheath, stem, and the whole plant affecting the yield with progressive crop loss [628]. There are many pathogens that belong to a widespread genera like Phytophthora, Alternaria, Cochliobolus, Bipolaris, Cryphonectria, Tubakia, Colletotrichum, Gloeosporium etc. (Table 2). Of which, Phytophthora genus was reported to have 120 crop/plant pathogenic species and the most prominent species being the Phytophthora infestans causing devastating disease i.e., late blight of potato and tomato [281]. The other prominently known fungal blight disease is the sheath blight caused by Rhizoctonia solani on paddy crops causing around 50% loss in the crop yield worldwide during favourable conditions [46, 220]. It is a soilborne Basidiomycete containing 100 different crop pathogenic species affecting various crops with sheath blight, banded leaf, aerial blight, and brown patch [9, 635]. Similarly, the bacterial pathogen Xanthomonas oryzae are the causative agent of bacterial blight and leaf streak disease in paddy crops which leads to about 16-50% yield loss worldwide depending on the paddy variety grown [267, 427]. Ou in 1985 [436] have stated that bacterial blight especially the kresek syndrome under favourable conditions could cause even 75% loss in the crop yield worldwide. Analogously, [340] have observed that bacterial leaf streak under favourable conditions could cause severe to very severe damage in the paddy crops leading to about 8-32% loss in yield. The major symptoms of the bacterial blight include the appearance of water-soaked lesions on the tips and the margins of leaves which may progress to chlorosis and necrosis [325] and the symptoms of bacterial leaf streaks are the formation of water-soaked lesions anywhere on the leaf surface resulting in translucent and yellowish streaks which on time turns greyish-white, die, and wither [267, 420].

The causative agents of the wilt disease in crops are the fungi, bacteria, and nematodes and are generally caused due to the moisture stress created by the pathogens while attacking the vascular system of the crops [308, 554]. The common symptoms include the water loss in the leaves and stem leading to dryness and death of the plant. Fungal pathogens like Ophiostoma ulmi, Fusarium oxysporum, Bretziella fagacearum, Acromonium diospyri, Verticillium sp. etc. are well known for their ability to cause wilt disease in various economically important crops like vegetables, flowers, and fruits (Table 2) [369]. Affected leaves turn to dull green with water-soaked appearance which curl upward and cling to the stem, and the margins are often observed to turn yellow to bronze in colour [270, 271, 342]. Verticillium sp. are the prominent species among the wilt causing fungal pathogens that affects a variety of crops and plants like trees, shrubs, vines, flowers, ornamental plants, vegetables, fruits, and economically important crops [49, 177, 450, 528]. There are about ten reported species of Verticillium namely Verticillium dahliae, Verticillium albo-atrum, Verticillium alfalfae, Verticillium longisporum, Verticillium nonalfalfae, Verticillium tricorpus, Verticillium zaregamsianum, Verticillium isaacii, Verticillium nubilum, and Verticillium klebahnii, among which Verticillium dahliae is the potential pathogen with economically importance [146, 258, 288, 295, 450, 528]. The specific symptoms include the early drop of green leaves from the twigs and branches leading to complete defoliation, necrosis, and death [288]. Bacterial wilt caused by the genera Corynebacterium, Erwinia, Pseudomonas, Ralstonia, Pantoea, and Xanthomonas species (Table 2) induces the formation of the discoloured water-soaked lesions resulting in stunting, wilting, and withering of leaves which on progression to stems lead to the formation of the lesions in the stem with characteristic bacterial ooze from the infected stems [153]. Nematode, Bursaphelenchus xylophilus a causative agent of pinewood wilt disease leads to a severe economic loss in various countries causing dramatic irreparable changes to the pine wood available in their native forest [8, 518].

Apparently, blast is a crop disease caused by the fungal and bacterial pathogens that majorly infects the cereals including the economically important varieties affecting the grains, leaves, leaf collars, nodes, panicles, and seedlings with the characteristic symptoms like elliptical leaf spots with grey, white centre and red to brown margins (Table 2) [292]. Fungal pathogens like *Magnaporthe oryzae*, *Hel*minthosporium sativum, Cochliobolus miyabeanus, and Magnaporthe grisea are known for their pathogenic effects on crops among which Magnaporthe oryzae and Magna*porthe grisea* are the potential pathogen affecting the paddy crops at various growth stages leading to severe losses to about 10–30% worldwide [523]. The necrotrophic fungi Cochliobolus miyabeanus causing rice brown spots and the hemibiotrophic bacteria Xanthomonas oryzae causing rice blight are generally found in association with the blast fungal species in the paddy crops leading to the improved crop and yield loss [279]. The bacterial blast also known as the citrus blast is caused by Pseudomonas syringae especially during the winter seasons with the formation of black lesions which leads to the curling and withering of leaves [263]. Also, it is well known for its blossom blast disease on apple with the characteristic disease symptoms including the necrosis of the lateral flower buds which on disease progression gets wilted and drooped [196]. Furthermore, rust and smut diseases are the significant fungal diseases that affect a wide range of crops and plants hampering their growth, yield, and quality substantially. About 7000 species of Basidiomycetes are known to cause yellow, black powdery, brown, orange, and red rust to various crops and plant worldwide [42, 66, 117]. Among these the most important genera includes the Puccinia, Phakopsora, Melampsora, Cronartium, Gymnosporangium, Hemileia, and Uromyces sp. (Table 2). These fungi are the biotrophic organisms that depend on their host for the nutrients, growth, and reproduction [11, 106, 144]. Wheat being the mostly affected crops by the rust fungi are susceptible to the infection caused by three potential species namely Puccinia striiformis causing yellow stripe rust [339], *Puccinia triticina* causing leaf brown rust [56, 208], and Puccinia graminis f. sp. tritici causing brown rust of stem [45, 326, 521] which leads to about 15-20% annual yield loss [166]. Yellow stripe rust is a devastating disease that causes an annual loss of about 5.5 million tons of wheat worldwide [42]. More than 88% of the wheat varieties worldwide are known to be vulnerable to the yellow stripe rust resulting in almost 100% yield loss with the characteristic stunted growth of the crops and immature kernels [42, 88, 166, 602]. Prank et al. [464] have estimated that the Puccinia graminis f. sp. tritici causing brown stem rust of wheat crops could cause as close to 100% loss in the yield of the crops with the formation of characteristic symptomatic brick red to brown lesions containing urediniospores on the leaves, sheath, stem, awns, and glumes. Around 90% of the wheat crop varieties worldwide are susceptible to the brown rust associated with reduced grain size and lodging of the crops [326, 521]. Wheat leaf rust is a common and widely distributed disease [56, 250], and the yield losses are attributed by the reduction in the kernel size and the number of grains in the head [250, 299]. Similarly, crown rust of cereals and the coffee rust are the other two imperative rust disease leading to severe economic loss caused by the pathogens Puccinia coronata and Hemileia vastatrix respectively. Former affects the crops with the formation of light orangish pustules on the leaf sheath, peduncles, and awn leading to stunted growth and reduced yield [166, 412]. Whereas the later forms yellow-orange powdery lesions with pale yellow chlorotic spots leading to leaf fall with a reduction in the quality and quantity of the flowers and fruits produced [571]. Smut is the other disease caused by *Basidiomycete* fungal pathogens which leads to great economic losses and are characterized by the formation of a dirt like structures or soot-like spores called as "sori" on the leaves, grains, and the ears of the affected crops. It mostly affects cereals, spices, vegetables like potato, and some cash crops like sugarcane by the fungal species belonging to the genera Ustilago, Sporisorium, and Thecaphora (Table 2). Sporisorium scitamineum is the causative agent of whip smut disease of sugarcane crops in about 120 countries worldwide [475]. It is generally characterized by the emergence of the smut whip in sugarcane crops which usually causes stunted growth, thin and slender canes with broad spaced nodes, appearance of whip like sorus at the top of the stalk or at either side of the canes with narrow leaves, profuse tillers, and spindlier shoot with an average yield loss of up to 12-75% [100, 424, 475, 489]. The common smut (boil or blister smut) of cereals especially maize is caused Ustilago maydis with the formation of thick and fleshy galls filled with spores impeding the yield of the crops [477]. Similarly, *Ustilago nuda, Ustilago nigra, Ustilago hordei, Ustilago avenae, Ustilago kolleri,* and *Ustilago tritici* are the other potential species that causes smut of other cereals like wheat, oats, barley, etc. [386]. *Thecaphora solani* is responsible for the destructive disease caused to potato crops with high yield loss of about > 90% worldwide [18].

Canker is the disease affecting various parts of the crops like leaves, stem, woody trunk, fruits etc. with characteristic symptoms like lesions with irregular sunken appearance, swollen, flattened, cracked, discoloured, and dead areas caused by both bacteria and fungi. Valsa mali, Neonectria galligena, Sirococcus clavigignenti-juglandacearum, Seiridium cardinale, Geosmithia putterillii, Discula destructiva, Eutypa lata, Thyronectria austro-americana, Lachnellula willkommii, Gibberella baccata, Diplodia quercina, Fusarium circinatum, Apiognomonia veneta, Leptosphaeria maculans, Leptosphaeria coniothyrium, Cryptosporella umbrina, and Gremmeniella abietina are some of the fungal species and the species like Pseudomonas syringae, Clavibacter michiganensis, Xanthomonas campestris, Xanthomonas citri, Xanthomonas axonopodis, Pseudomonas savastanoi, and Xanthomonas populi are some of the bacterial species causing cankers to various crops [64, 74, 165, 168, 178, 179, 212, 316, 331, 481, 505, 506, 599, 609]. Symptoms of fungal stem or branch cankers are associated with dieback syndrome and are caused by almost 150 different species of fungi infecting about 130 species of woody hosts [3, 10, 162, 294, 537]. Besides, bacterial cankers of citrus fruits are characterized by the lesions on the twigs, leaves, and fruits which leads to defoliation and fruit drop causing severe economic loss [267, 503].

Neoplastic plant disease like crown galls (tumours) of the herbaceous plants are caused by bacterial species such as Agrobacterium tumefaciens, Agrobacterium rubi, and Agrobacterium vitis on the stems, branches, leaves, roots, canes, and veins with a characteristic wound appearing like cauliflower head leading to severe damage with weakened appearance, stunted growth, and death of the plants [33, 205]. However, potato warts caused by the fungi Synchytrium endobioticum are said to result in substantial economic losses worldwide as it is a social disease observed in almost all the countries with severe damage to the tubers [426]. Nevertheless, scab disease of potatoes caused by the bacterial pathogens Streptomyces scabies, Streptomyces bottropensis, Streptomyces stelliscabiei, and Streptomyces aureofaciens are known to cause severe loss of the tubers with the formation of crustaceous lesions on the leaves, stem, fruits, and tubers [495]. Furthermore, apple scabs caused by the Ascomycetous fungi Venturia inaequalis leads to considerable yield loss worldwide affecting the quality and quantity of the fruits produced [203]. Likewise, damping-off is a soilborne infection caused by

fungal pathogens like *Rhizoctonia solani*, *Aphanomyces cochlioides*, *Pythium sp.*, *Phytophthora sp.*, *Botrytis sp.*, *Fusarium sp.*, *Cylindrocladium sp.*, *Diplodia sp.*, *Phoma sp.*, and *Alternaria sp.* affecting almost all agricultural and forestry crops causing severe damage to the yield and leads to death of crops worldwide [315].

Parasites like nematodes are also known to cause various diseases to crop globally, namely the root-knot disease, cyst disease, leaf spots, sting, wilt, and lesion disease. Root-knot disease is caused by the nematodes of the genus Meloidogyne with the formation of the root-knot galls containing the nematodes in roots of the crops [576, 615]. The major species of the root-knot nematodes include Meloidogyne javanica, Meloidogyne arenaria, Meloidogyne incognita, and *Meloidogyne hapla* [173]. It is estimated by researchers like [268, 496, 632], and [429] that these nematodes cause severe damage to economically important crops especially the varieties like tomato, watermelon, pepper, eggplant, potato, carrot, and cucumber leading to a yield loss of about 15-25% and sometimes as high as 75% with an estimated loss of about 100 billion USD per year worldwide. Nematode cyst disease is caused by the nematodes like Heterodera schachtii, Heterodera goettingiana, Heterodera glycines, and Globodera pallida affecting various crops like soya beans, peas, sugar beet, and potatoes causing a yield loss of about 10–30% and estimated loss of about 1 billion USD [245, 377]. Lesion nematode disease is also a devastating disease of root tissues that are majorly soilborne and are caused by Pratylenchus pratensis, Pratylenchus brachyurus, Pratylenchus coffeae, Pratylenchus penetrans, Pratylenchus scribneri, Pratylenchus vulnus, Pratylenchus zeae, Pratylenchus crenatus, and Pratylenchus thornei affecting various crops including cereals, peanuts, potatoes, fruits, and all monocots leading to a yield loss of about 50-85% and estimated loss of about 216 billion USD worldwide [394, 418, 423]. Similarly, Pratylenchus penetrans causes about 30-50% yield loss of potatoes with the characteristic early dying disease associated with premature vein senescence [317, 486, 487]. Sting nematode Belonolaimus longicaudatus is a soilborne ectoparasitic nematode that generally affects all agricultural crops especially the peanuts with the characteristic chronic wilting leading to great economic losses [310].

#### Disease caused by meso-biotic factors

Disease caused by the meso-biotic factors are the disease caused by parasites like viruses and viroids that are capable of existing in dormant stage for a relatively longer periods, gets activated once after entering a favourable host cell, and requires a host for their survival [302, 379]. Virus pathogens are transmitted to healthy plants through sap contamination, vectors, nematodes, soilborne, seedborne, and pollen borne transmissions [508]. Of which, insects (vectors) play a vital

role with the existence of 200 known species of aphids that transmit the mosaic viruses, > 100 species of leafhoppers known to transmit viruses causing yellow discoloration in crops, and other insects like whiteflies, thrips, mealybug, planthoppers, grasshoppers, scales, and some beetle which help in transmitting a wide range of viruses affecting agricultural crops [189]. Nematodes feeding on roots are also responsible for the virus transmission in the host plants from the soil for e.g., grape fanleaf virus, tobacco and tomato ringspot virus, and many strawberry viruses are known to be transmitted by nematodes [19, 185]. Similarly, many studies worldwide report that viruses like big vein of lettuce, soilborne wheat mosaic, and tobacco necrosis viruses are transmitted by the swimming spores of the soil inhabiting fungal pathogens [129, 589].

Viruses cause various damages to the host crops like the mosaic damage, yellowish discoloration, chlorosis, stunted growth, and necrosis which significantly hampers the growth and yield of the crops to about 40-100% [357, 508]. Mosaic viruses affects the foliage of the crops with the formation of yellowish to dark green patches, mottled appearance, curled leaves, and appearance of light-coloured veins [87]. Many mosaic viruses are known till date to affect a vast range of crops. Wherefrom, the most potential viruses are the tobacco mosaic virus, rice stripe mosaic virus, cauliflower mosaic virus, sugarcane mosaic virus, lettuce mosaic virus, and maize mosaic virus infecting tobacco, paddy, pepper, potato, tomato, eggplant, cucumber, petunia, melons, squash, spinach, celery, beet, cereals, wheat, sugarcane, lettuce, maize, and other economically important agricultural crops [633]. Among these, the rice stripe mosaic viruses affecting paddy crops, transmitted by flies were reported to have a field incidence of about 70% with severe loss in the grain production [87, 601]. Tobacco mosaic virus is of profound economic significance as it causes mottled browning of tobacco leaves and stunted growth hampering the quality and market value of the leaves [338]. The virus has also been reported to infect various solanaceous crops affecting their yield and quality [446]. Sugarcane mosaic virus being the next potential pathogen belonging to the genera Potyvirus and family Potyviridae are transmitted by aphids [344] and causes severe damage to the economically significant sugar and energy producing crops sugarcane by reducing their yield to about 10-50% and sometime as high as about 60-80% worldwide [595, 612].

Similarly, the leaf roll viruses affecting the fruits (grapes) and vegetable (potato) crops are transmitted by aphids with the characteristic symptoms like stunted growth with erected appearance and leaves roll upwards with leathery and chlorotic texture and appearance [363]. Grapevine leafroll disease caused by grapevine leafroll virus have been known since 1936 for their symptomatic downward leaf rolling, chlorosis, and delayed ripening of fruits reducing the fruit quality and yield to about 40% [13, 367]. It is also estimated by [25] that the virus results in a loss of about 25,000-40,000 USD per hectare of vineyard. Potato leafroll virus being the most devastating virus of the leafroll viruses which belongs to the genus Polerovirus and family Luteoviridae leads to a global yield loss of 20 million tons i.e., 90% loss worldwide [303]. [307] has observed the latent infection caused by the virus in the planting tubers directly affects the rate of tuber germination, crop strength, and the yield of potatoes. Likewise, the leaf curl viruses belonging to the Geminiviridae family are generally transmitted through whiteflies and are known to affect a variety of crops with stunted growth, chlorosis, curling of leaves, and considerable reduction in the fruit yield globally [107, 198, 399, 442, 586]. Among the know leaf curl viruses, tomato yellow leaf curl virus is a potential pathogen causing devastating disease of many crops and tomatoes with severe effects on the fruits especially the depletion in the fruit taste, reduced size, un-uniform ripening of fruits, and severe yield losses worldwide [362, 438, 538]. Further, peanut stunt virus of the family Bromoviridae are known for their stunt disease of peanut crops and soyabean with severe dwarfing or stunting of peanut crops with an annual yield loss of about 10-50% [408, 443].

Likewise, the viroid, which are also called as mini-viruses lacking capsid are the low molecular weight single stranded RNA (usually the size ranges between 246–399 nucleotides), covalently closed, circular, highly structured non-coding RNAs that replicate autonomously affecting a vast range of economically important crops [302]. Further, they can cause non-symptomatic latent infections in their host which on transmission to susceptible hosts causes devastating infections. Table 3 lists the available information of various viroids known to affect the crops and their lineage. Viroid belongs to the two major families namely Pospiviroidae and Avsunviroidae with the common genera Pospiviroid, Hostuviroid, Cocadviroid, Apscaviroid, and Coleviroid of Pospiviroidae family and Avsunviroid, Pelamoviroid, and Elaviroid of Avsunviroidae family (Table 3) [128, 437]. Pospiviroids are majorly known to affect tomato crops with the characteristic diseases like chloric dwarf disease, citrus exocortis disease, columnea latent disease, tomato apical stunt disease, and tomato planta macho disease (Table 2) leading to chlorosis, bronzing, leaf distortion, and growth reduction with an estimated yield loss of about 39-82% worldwide [591, 592]. Similarly, potato spindle tuber disease caused by Pospiviroid are known to affect tomatoes and potatoes with a field infection rate of about 25-50% affecting the tuber and fruit quality resulting in a yield loss of about 80% [520]. Nonetheless, the economically important hop stunt disease caused by hop stunt viroid of Hostuviroid has shown a field infection rate of about 10-20% [365] to multiple crops like mulberry, almond, apple, apricot, cherry, peach, pear, plum, pistachio, citrus fruits, and the flower Hibiscus rosa-sinensis with severe loss in quality and yield [15, 347, 349, 619]. Further, fruit crinkle disease caused by the fruit crinkle viroid of the genus *Apscaviroid* is known to have a narrow host range affecting apple fruits with a significant loss in yield and quality of the fruits [329]. Similarly, *Avsunviroid, Pelamoviroid*, and *Elaviroid* of *Avsunviroidae* family are known to cause severe yield loss of fruits and vegetable especially the crops like avocado, chrysanthemum, peach, and eggplant hampering the fruit quality and yield worldwide [77, 93, 158, 510].

# Devastating crop diseases of Southeast Asian nations

The ten member states association forming the Southeast Asian nations include the countries like the Brunei Darussalam, Cambodia, Indonesia, Laos, Malaysia, Myanmar, Philippines, Singapore, Thailand, and Vietnam [244]. A wide variety of crops are cultivated in these countries namely cotton, millets, peanuts, paddy, sunflower, maize, palm, soybean, and sugarcane that are affected by major crop diseases impeding the crop yield and causing severe to heavy economic losses. Paddy crops with profound significance in the Southeast Asian nations are majorly affected by the devastating and destructive biotrophic fungal pathogen Rhizoctonai solani with a yield loss of about 6 million tons of rice grains annually [635]. Similarly, Naqvi [409] has reported the prevalence of bacterial blight of paddy caused by Xanthomonas oryzae in the countries like Philippines and Indonesia leading to severe loss in the grain yield. Oña et al. [431] has reported the epidemic developed by the bacterial leaf streak disease of paddy causing severe loss in the yield of the crops especially in the variety IR1695 compared to IR24. Another study on the devastating pathogen, Fusarium oxysporum (soilborne fungi) causing the Fusarium wilt of banana was reported by [398] in Philippines outlines the severity of infection and loss caused by the TR4 strain. Further, [57] has documented the major pathogenic disease affecting the vegetables in Philippines and has reported that the bacterial wilt caused by Ralstonia solanacearum has led to severe damage and yield loss of the crops like tomato, pepper, eggplant, bitter gourd, and lettuce. Further, the study has also stated the prevalence of the devastating diseases like downy mildew affecting bitter gourd and Chinese cabbage, Cercospora disease affecting pepper, eggplant, bitter gourd, and tomato, Phytophthora disease of tomato, eggplant, pepper, and Chinese cabbage, Fusarium wilt of tomato and pepper, disease of tomato crops caused by *Phytoplasma* sp. and Corynespora cassiicola, and various other diseases caused to tomato which includes bacterial canker, bacterial speck, target spot, Septoria leaf spot, and pith necrosis. Similarly, it is reported that the popular fruit trees Durian is

highly vulnerable to the stem canker or patch canker disease caused by Phytophthora palmivora impeding the fruit yield and fruit quality in Sungai Liang, Brunei Darussalam [522]. Furthermore, Pearce et al. [447] and Bigirimana et al. [52] has reviewed the destructive rice sheath disease caused by Sarocladium oryzae causing considerable reduction in the crop yield in the countries like Brunei Darussalam, Indonesia, Philippines, Vietnam, and Thailand. Pyricularia oryzae causing destructive rice blast, leaf blast, and neck blast of paddy crops in Cambodia [95] is reported to cause elliptical lesions on the leaves during the vegetative and the reproductive phases of the crops, grain sterility, reduced grain size, loss in yield, and reduced grain quality [35, 289]. Also, [311] has reported an annual yield loss of about 55 million UDS in Southeast Asian countries due to the devastating infection of paddy cause by Pyricularia oryzae. According to the study reported by [102], bacterial sheath brown rot of rice caused by Pseudomonas fuscovaginae was found to be prevalent in Cambodia leading to sever loss in the grain yield with the appearance of the characteristic brown lesions on the sheath. It was also reported that the coinfection caused by several bacterial pathogens like Acidovorax avenae, Burkholderia gladioli, Burkholderia cepacian, and Pantoea ananatis resulted in the severing of the disease along with the formation of sheath lesions and grain discoloration [27, 534].

Casava being the most importantly cultivated food crops of the Southeast Asian countries has been majorly affected by a variety of disease-causing pathogens [569]. The most prevalent disease of casava is the casava mosaic disease caused by the cassava mosaic begomoviruses of the genus Begomovirus and Geminiviridae family resulting in the poor leaf development with reduced photosynthesis leading to the poor tuber quality with improved yield loss [581]. Shallots are one among the crops with high economic value worldwide and are largely cultivated in the South Kalimantan of Indonesia are reported to be infected by the Moler disease by Fusarium oxysporum and anthracnose by Colletotrichum sp. causing a severe damage to crop with a resulting yield loss ranging from 24 to 100%. Likewise, Adiyoga et al. [5], Bambang and Khusnul [30], and [188] have reported that Fusarium wilt disease, anthracnose, and porch blotch of onions could lead to a sever yield loss of about 27-27%, 21-100%, and 30% respectively in Southeast Asian countries and Indonesia. Further, bacterial stalk rot of corn a devastating disease caused by the bacterial species Dickeya zeae is reported to cause severe yield loss of corn in Indonesia [547]. Similarly, [546] has reported the vulnerability of the corn crops to various insects like grasshopper, katydid, sweet potato bug, derbid plant hopper, cotton stainer, largid bug, corn leaf hopper, corn rootworms, green chafer beetle, tussock moth, Asian lady beetles, transverse lady beetle, and assassin bug along with two fungal pathogenic

#### Table 3 Details of various viroid known to affect crops and their yield (classification as given in [170])

Disease	Viroid	Size of the viroid RNA, nucleotides	Family	Genera	Host plant	Variants known	New host infected	Reference
Chloric dwarf disease	Chloric dwarf viroid	360	Pospiviroidae	Pospiviroid	Tomato	2	_	[591, 592]
Citrus exocortis disease	Citrus exocortis viroid	366-475			Tomato, citrus	86	-	[600]
Columnea latent disease	Columnea latent viroid	359-456			Columnea con- sanguinea, Nematanthus sp., Brunfel- sia pauciflora	17	Tomato	[560]
Tomato apical stunt disease	Tomato apical stunt viroid	360-363			Tomato	5	-	[21]
Tomato planta macho disease	Tomato planta macho viroid	360			Tomato	2	-	[372]
Potato spindle tuber disease	Potato spindle tuber viroid	341-364			Potato	109	Tomato, avocado	[520]
Chrysanthemum stunt disease	Chrysanthemum stunt viroid	348-356			Chrysanthemum	19	-	[406]
Iresine disease	Iresine viroid	370			Iresine herbstii	3	-	[590]
Mexican papita disease	Mexican papita viroid	359-360			Solanum cardio- phyllum	6	-	[370]
Hop stunt disease	Hop stunt viroid	294-303		Hostuviroid	Citrus, grape- vine, <i>Prunus</i> sp.	144	Hop, cucumber	[282]
Coconut cadang- cadang disease	Coconut cadang- cadang viroid	246-301		Cocadviroid	Coconut palm	8	Monocots and African oil palm	[231, 476]
Coconut tinangaja disease	Coconut tinangaja viroid	254			Coconut palm	2	-	[240]
Citrus bark crack- ing disease	Citrus bark crack- ing viroid	284-286			Citrus	6	-	[507]
Hop latent disease	Hop latent viroid	255-256			Нор	10	-	[445]
Apple scar skin disease	Apple scar skin viroid	329-333		Apscaviroid	Apple, pear	8	-	[515]
Apple dimple fruit disease	Apple dimple fruit viroid	306			Apple	2	-	[485]
Apple fruit crin- kle disease	Apple fruit crin- kle viroid	368-372			Apple	29	-	[329]
Australian grape- vine disease	Australian grape- vine viroid	369			Grapes	1	-	[150]
Citrus bent leaf disease	Citrus bent leaf viroid	315-329			Citrus	24	-	[76]
Citrus dwarfing disease	Citrus dwarfing viroid	291-297			Citrus	53	-	[109]
Grapevine yel- low speckle 1 disease	Grapevine yellow speckle 1 viroid	365-368			Grapes	49	-	[617]
Grapevine yel- low speckle 2 disease	Grapevine yellow speckle 2 viroid	363			Grapes	1	-	[561]
Pear blister canker disease	Pear blister canker viroid	314-316			Pear, quince	18	-	[169]
Coleus blumei -1 disease	Coleus blumei -1 viroid	248-251		Coleviroid	Coleus blumei, Mentha sp.	9	-	[578]
Coleus blumei -2 disease	Coleus blumei -2 viroid	295-301			Coleus blumei	1	-	[184]
Coleus blumei -3 disease	Coleus blumei -3 viroid	361-364			Ocimum basili- cum, Melissa officinalis	3	-	[536]
Coleus blumei -5 disease	Coleus blumei -5 disease	NA			Plectranthus scutellarioides	NA	Aurora black cherry	[577]

Peach latent

viroid

mosaic viroid

Eggplant latent

Table 3 (contin	ued)							
Disease	Viroid	Size of the viroid RNA, nucleotides	Family	Genera	Host plant	Variants known	New host infected	Reference
Avocado sun blotch disease	Avocado sun blotch viroid	239-251	Avsunviroidae	Avsunviroid	Avocado	83	_	[77]
Chrysanthemum chlorotic mottle disease	Chrysanthemum chlorotic mottle viroid	397-401		Pelamoviroid	Chrysanthemum	21	-	[93]

Elaviroid

NA, Not available

mosaic disease

Eggplant latent

Peach latent

disease

infections caused by Fusarium sp. and Puccinia sp. affecting the crop yield in Malaysia. Myanmar being the second largest producer of sesame seeds globally experiences an annual yield loss of about 5-50% by phyllody, 10-75% by charcoal rot caused by *Macrophomina phaseolina*, 5–50% by Cercospora leaf spot, and 5% by bacterial leaf spot caused by Xanthomonas campestris [389].

335-351

332-335

Furthermore, [54] have reported the prevalence of various pathogens infecting a variety of crops like paddy disease of sheath and bakanae caused by Gibberella fujikurol, paddy sheath blight by Rhizoctonia sp., sheath rot by Sarocladium attenuatum, rice blast by Pyricularia oryzae, paddy false smut by Ustilaginoidea virens, downy mildew of soya bean by Peronospora manshurica, anthracnose of soya bean by Colletotrichum lindemuthianum, late blight of vegetables by Phytophthora infestans, fruit leaf anthracnose by Glomerella cingulate, and coffee leaf rust by Hemileia vastatrix impeding the crop yield. Similarly, Phytoplasma sp., a destructive pathogen infecting sugar cane in Vietnam are reported to reduces the sucrose content of the crops to about 60-80% [239].

# Measures to control crop diseases- overview of the resistant varieties

Globally, around 16-30% of the crops grown are estimated to be lost to the devastating infections caused by pathogens [427, 428, 498]. To control the crop and yield losses, several practices have been followed by the farmers worldwide like the usage of healthy seeds for sowing, seed and soil treatment, crop rotation, usage of biological and chemical pesticides, etc., but development of host resistance have attracted researchers worldwide as it is considered as a comparatively reliable method ever [415, 583]. Generally, plant's innate immune system provides protection against the ingression of pathogens, but the species specificity expressed by the immune system makes it difficult to manage the infections caused by multiple pathogens [510]

[158]

[261]. Hence, deployment of the gene conferred resistant varieties of crops are considered as a potential measure to control crop diseases in an effective, eco-friendly, robust, and economic means [267]. Ji et al. [266] have reported the identification of about 40 different resistant R genes of paddy crops which could confer resistance against the major paddy crop infections especially the bacterial blight caused by Xanthomonas oryzae. Among which, the most frequently used R genes as rice breeders are xa5, Xa7, xa13, Xa21, and Xa23 due to their wider resistance spectrum compared to the other known genes. Xu et al. [618] have reported the multiple pathogenic resistance conferred by the transformation of the specific R genes namely Xa7and Xa21 to Yihui 1577 rice variety. Similarly, Pathi et al. [444] have stated that the generated loss-of-function mutations in the Lox3 gene of maize crops using the Cas endonuclease technology improved the durability in the resistant properties of the crops to various pathogens. The Pto gene of the tomato crops were found to confer resistance against the pathogen Pseudomonas syringae which are well known for their devastating vegetable and fruit crop diseases [293, 368, 456, 504, 559]. Further, identification of the NLR gene [352, 493] in various Arabidopsis crops led to a significant progress in the genetic resistance to crop diseases as these genes conferred resistance to wide range of pathogens which include fungi [132, 277], bacteria [44, 210, 390], oomycetes [58], and viruses [298, 603]. Furthermore, discovery of R gene stacking or pyramiding helped in the deployment of multiple NLR gene into a single potato cultivar improving the resistance of the crops to multiple diseases [273, 291]. Identification of the quantitative trait locus (QTLs) of various pathogens (cucumber mosaic virus, zucchini yellow mosaic virus, whitefly-transmitted begomoviruses, melon chlorotic mosaic virus, tomato leaf curl virus, watermelon chlorotic stunt virus, Fusarium wilt, downy mildew, and powdery mildew) and insects (aphids and whitefly) that are co-localized in the Vat gene were found to confer resistance in fruit trees and crops improving the fruit yield and quality [483, 492, 532, 594, 629].

168

9

Peach nectarine

Eggplant

Likewise, several researchers worldwide are actively working in identifying and improving the resistance gene that could confer resistance to a vast range of pathogens that could be used to protect a wide range of crops from the pathogenic infections.

# Conclusion

Bioaerosols act as potential source of crop pathogens and enable their dissemination across geographical barriers. Their dispersal in the atmospheric air is largely dependent upon several intrinsic and extrinsic factors like the availability of source, meteorological factors, aerosolization behaviour, properties of the microorganisms, etc. Once dispersed, they can cause severe to very severe diseases in crops. Crop pathogenic bioaerosols are generated due to various natural and anthropogenic activities such as rain, splashing of water, usage of sprayers in crop field, mechanical activities observed in the canopy, especially the movement of leaves, ooze of infected crops, infected plant sap etc. Leaves and external tissues of crops play a pivotal role in the ingress of pathogens from the surface to the inner parenchymal tissues of the crops. Also, it is reported that about 30% of the global crop production is lost to the devastating diseases caused by the crop pathogens known till date. Hence, it is imperative to have a better understanding of the crop pathogen dissemination through bioaerosols, sedimentation, invasion, and the onset of symptomatic disease of the region-specific crop cultivated. To achieve this, a detailed study on the season-specific regional diversity and community composition of bioaerosols and their implications on the crops must be conducted globally. Global modelling of the data obtained would give us a better insight on the dispersion and spread of crop pathogens on a global scale. Based on the knowledge and information obtained on the season-specific regional diversity, bioaerosol composition, and global modelling, proper measures required to control various crop diseases like crop rotation practises, usage of resistant varieties, fumigation of soil, seed selection, etc. must be developed and implemented. Moreover, advanced research on the resistant varieties have shown promising results in controlling multiple crop pathogens. Though many studies have explained the identification and existence of various R genes responsible for gene conferred resistance in crops, good understanding on the mechanism of the gene conferred plant response and resistance could give a better insight in the development of an appropriate and potential gene for the crop resistance. Further, forthcoming studies on the identification and development of a crop specific gene mediated resistance to multiple pathogens would enable the development of potential resistant varieties of crops in the future era.

### Future need and measures to be adopted

Crop diseases and pests are critical problems in farming since earlier days. Although there have been many advancements in the understanding and eradication of various crop diseases worldwide, further measures need to be taken to improve the crop yield and crop resistance to diseases to overcome the yield loss caused by pathogens. Following are the future needs that need to be adopted and implemented by the farmers and officials to control the pathogenic crop diseases.

- Proper insight on the regional climatic conditions and the bioaerosols seasonal community composition
- Implementation of various farm practises like seed selection, season-specific crop growth, crop rotation, soil fumigation, soil treatment, seed treatment, etc.
- Breeding and adaptation of the resistant crop varieties
- Elimination of the source of bioaerosols
- Early prediction of spread of crop pathogenic bioaerosols and onset of disease
- Thorough knowledge of the disease cycle followed by each pathogen would help in improving the disease management strategies
- Proper understanding of the regional bioaerosols composition and implementation of appropriate methods e.g., growing appropriate resistant crop varieties
- Dissemination of knowledge to officials and farmers regarding the region-specific disease management strategies

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# References

- Abad P, Castagnone-Sereno P, Rosso MN, et al (2009) Invasion, feeding and development. Root-knot Nematodes 163–181. https://doi.org/10.1079/9781845934927.0163
- Abasova LV, Aghayeva DN, Takamatsu E (2018) Erysiphe azerbaijanica and E. linderae: Two new powdery mildew species (Erysiphales) belonging to the Microsphaera lineage of Erysiphe. Mycoscience 59:181–187
- Adams GC, Wingfield MJ, Common R, Roux J (2005) Phylogenetic relationships and morphology of Cytospora species and related teleomorphs (Ascomycota, Diaporthales, Valsaceae) from Eucalyptus. Stud Mycol 52
- 4. Adhikari A, Reponen T, Grinshpun SA et al (2006) Correlation of ambient inhalable bioaerosols with particulate matter and

ozone: A two-year study. Environ Pollut 140:16–28. https://doi. org/10.1016/j.envpol.2005.07.004

- Adiyoga W, Moekasan TK, Uhan TS, Suenaryo E, Hendarsih (2000) Present status of pest and disease management on food and vegetable crops and its future development. PEI (Perhimpunan Entomologi Indonesia), Surakarta and PT. PCI, Jakarta
- Aegerter BJ, Nuñez JJ, Davis RM (2003) Environmental factors affecting rose downy mildew and development of a forecasting model for a nursery production system. Plant Dis 87:732–738. https://doi.org/10.1094/PDIS.2003.87.6.732
- Agrios GN (2005) Plant pathology, 5th edn. Elsevier Academic Press, London, UK
- Akbulut S, Yüksel B, Serin M, Erdem M (2015) Comparison of pathogenic potential of Bursaphelenchus species on conifer seedlings between greenhouse and outdoor conditions. Phytoparasitica 43:209–214. https://doi.org/10.1007/s12600-014-0433-2
- Akhtar J, Kumar Jha V, Kumar A, Lal HC (2009) Occurrence of banded leaf and sheath blight of maize in Jharkhand with reference to diversity in Rhizoctonia solani. Asian J Agric Sci 1:32–35
- Al Raish SM, Saeed EE, Sham A et al (2020) Molecular characterization and disease control of stem canker on royal poinciana (Delonix regia) caused by neoscytalidium dimidiatum in the United Arab Emirates. Int J Mol Sci 21:1033. https://doi.org/10. 3390/ijms21031033
- Ali S, Rodriguez-Algaba J, Thach T et al (2017) Yellow rust epidemics worldwide were caused by pathogen races from divergent genetic lineages. Front Plant Sci 8:1057. https://doi.org/10.3389/ fpls.2017.01057
- Aller JY, Kuznetsova MR, Jahns CJ, Kemp PF (2005) The sea surface microlayer as a source of viral and bacterial enrichment in marine aerosols. J Aerosol Sci 36:801–812. https://doi.org/10. 1016/j.jaerosci.2004.10.012
- 13. Almeida RPP, Daane KM, Bell VA et al (2013) Ecology and management of grapevine leafroll disease. Front Microbiol 4:94. https://doi.org/10.3389/fmicb.2013.00094
- Alvarado MJ, Lonsdale CR, Yokelson RJ et al (2015) Investigating the links between ozone and organic aerosol chemistry in a biomass burning plume from a prescribed fire in California chaparral. Atmos Chem Phys 15:6667–6688. https://doi.org/10. 5194/acp-15-6667-2015
- Amari K, Ruiz D, Gómez G et al (2007) An important new apricot disease in Spain is associated with Hop stunt viroid infection. Eur J Plant Pathol 118:173–181. https://doi.org/10.1007/s10658-007-9127-7
- Amato P, Parazols M, Sancelme M et al (2007) An important oceanic source of micro-organisms for cloud water at the Puy de Dôme (France). Atmos Environ 41:8253–8263
- Anderson PJ (1926) Comparative susceptibility of onion varieties and of species of Allium to *Urocystis cepulae*. J Agric Res 31:275–285
- Andrade O, Muñoz G, Galdames R et al (2004) Characterization, in vitro culture, and molecular analysis of Thecaphora solani, the causal agent of potato smut. Phytopathology 94:875–882. https:// doi.org/10.1094/PHYTO.2004.94.8.875
- Andret-Link P, Laporte C, Valat L et al (2004) Grapevine fanleaf virus: still a major threat to the grapevine industry. J Plant Pathol 86:183–195. https://doi.org/10.1080/01904167.2015.1112950
- Andrews JH, Harris RF (2000) The ecology and biogeography of microorganisms on plant surfaces. Phytopathology 38:145–180
- Antignus Y, Lachman O, Pearlsman M (2007) The spread of Tomato apical stunt viroid (TASVd) in greenhouse tomato crops is associated with seed transmission and bumble bee activity. Plant Dis 91:47–50
- Ariya PA, Amyot M (2004) New directions: The role of bioaerosols in atmospheric chemistry and physics. Atmos Environ 38:1231–1232. https://doi.org/10.1016/j.atmosenv.2003.12.006
- Arshad M, Suhail A (2010) Studying the sucking insect pest community in transgenic Bt cotton. Int J Agric Biol 12:764–768

- Asibi AE, Chai Q, Coulter JA (2019) Rice blast: A disease with implications for global food security. Agronomy 9:451. https:// doi.org/10.3390/agronomy9080451
- Atallah SS, Gómez MI, Fuchs MF, Martinson TE (2012) Economic impact of grapevine leafroll disease on Vitis vinifera cv. Cabernet franc in Finger Lakes vineyards of New York. Am J Enol Vitic 63:73–79. https://doi.org/10.5344/ajev.2011.11055
- Aylor DE, Taylor GS (1982) Aerial dispersal and drying of Peronospora tabacina conidia in tobacco shade tents. Proc Natl Acad Sci 79:697–700.https://doi.org/10.1073/pnas.79.2.697
- 27. Azegami K (2001) Burkholderia spp. associated with rice. In: Mew TW, Cottyn BG (eds) Seed health and seed-associated microorganisms for rice disease management. IRRI, Limited Proceedings No. 6., Los Banos, Philippines
- Bae C, Han SW, Song YR et al (2015) Infection processes of xylem-colonizing pathogenic bacteria: possible explanations for the scarcity of qualitative disease resistance genes against them in crops. Theor Appl Genet 128:1219–1229. https://doi.org/10. 1007/s00122-015-2521-1
- Baertsch C, Paez-Rubio T, Viau E, Peccia J (2007) Source tracking aerosols released from land-applied class B biosolids during high-wind events. Appl Environ Microbiol 73:4522–4531. https://doi.org/10.1128/AEM.02387-06
- Bambang HI, Khusnul M (2014) Effectiveness of resistance and biopesticide induction on Cercospora and Anthracnose leaves in chili (Capsicum annuum L.). Planta Tropika J Agro Sci 2:106–114
- Barnwal MK, Kotasthane A, Magculia N et al (2013) A review on crop losses, epidemiology and disease management of rice brown spot to identify research priorities and knowledge gaps. Eur J Plant Pathol 136:443–457
- Barrow GH and Feltham RKA (1993) Cowan and steel's manual for identification of medical bacteria, 3rd edn. Cambridge University Press, Cambridge, pp 331. https://doi.org/10.1017/ CBO9780511527104
- Barton IS, Fuqua C, Platt TG (2018) Ecological and evolutionary dynamics of a model facultative pathogen: Agrobacterium and crown gall disease of plants. Physiol Behav 20:16–29. https://doi. org/10.1111/1462-2920.13976.Ecological
- Bashan Y, Sharon E, Oleon Y, Henis Y (1981) Scanning electron and ligbl microscopy of infection and symptom development in tomato leaves infected with Pseudomonas tomalo. Physiol Plant Palhol 19:3944
- Bastiaans L (1991) Ratio between virtual and visual lesion size as measure to describe reduction in leaf photosynthesis of rice due to leaf blast. Phytopathology 81:611–615
- Bates GD, Rothrock CS, Rupe JC (2008) Resistance of the soybean cultivar Archer to Pythium damping-off and root rot caused by several Pythium spp. Plant Dis 92:763–766
- Bauer H, Kasper-Giebl A, Löflund M, Giebl H, Hitzenberger R, Zibuschka F, Puxbaum H (2002) The contribution of bacteria and fungal spores to the organic carbon content of cloud water, precipitation and aerosols. Atmos Res 64:109–119
- Baylor ER, Baylor MB (1980) Surf-to-wind transfer of viruses. Ann NY Acad Sci 353:201–208
- Baylor ER, Baylor MB, Blanchard DC et al (1977) Water-to-air transfer of virus. Science 198:575–580
- Baylor ER, Peters V, Baylor MB (1977) Water-to-air transfer of virus. Science 197:763–764
- Beattie GA, Lindow SE (1995) The Secret Life of Foliar Bacterial Pathogens on Leaves. Annu Rev Phytopathol 33:145–172. https://doi.org/10.1146/annurev.phyto.33.1.145
- Beddow JM, Pardey PG, Chai Y et al (2015) Research investment implications of shifts in the global geography of wheat stripe rust. Nat Plants 1:15132. https://doi.org/10.1038/nplants.2015.132

- Benfradj N, Metoui N, Boughalleb N (2016) Screening for tolerance of different citrus rootstocks against zoospores of Phytophthora nicotianae in infested soil. J Phytopathol Pest Management 3:63–75
- Bent AF, Kunkel BN, Dahlbeck D et al (1994) RPS2 of Arabidopsis thaliana: a leucine-rich repeat class of plant disease resistance genes. Science (80-) 265:1856–1860
- 45. Berlin A, Samils B, Djurle A et al (2013) Disease development and genotypic diversity of Puccinia graminis f. sp. avenae in Swedish oat fields. Plant Pathol 62:32–40. https://doi.org/10. 1111/j.1365-3059.2012.02609.x
- Bernardes-de-Assis J, Storari M, Zala M et al (2009) Genetic structure of populations of the rice-Infecting pathogen rhizoctonia solani AG-1 IA from China. Phytopathology 99:1090–1099. https://doi.org/10.1094/PHYTO-99-9-1090
- 47. Bhangar S, Huffman JA, Nazaroff WW (2014) Size-resolved fluorescent biological aerosol particle concentrations and occupant emissions in a university classroom. Indoor Air 24:604–617. https://doi.org/10.1111/ina.12111
- Bhangar S, Adams RI, Pasut W, Huffman JA, Arens EA, Taylor JW, Bruns TD, Nazaroff WW (2016) Chamber bioaerosol study: human emissions of size-resolved fluorescent biological aerosol particles. Indoor Air 26:193–206
- Bhat RG, Subbarao KV (1999) Host range specificity in Verticillium dahliae. Phytopathology 89:1218–1225. https://doi.org/10. 1094/PHYTO.1999.89.12.1218
- Bhattacharya I, Dutta S, Mondal S, Mondal B (2014) Special issue: Clubroot disease on Brassica crops in India. Canadian J Plant Pathol 36:154–160. https://doi.org/10.1080/07060661. 2013.875064
- Bhuiyan SA, Magarey RC, McNeil MD, Aitken KS (2021). Sugarcane Smut, Caused by Sporisorium scitamineum, a major disease of sugarcane: A contemporary review. Phytopathol 111. https://doi.org/10.1094/PHYTO-05-21-0221-RVW
- Bigirimana V de P, Hua GKH, Nyamangyoku OI, Hôfte M (2015) Rice sheath rot: An emerging ubiquitous destructive disease complex. Front Plant Sci 6. https://doi.org/10.3389/fpls. 2015.01066
- Bird DMK, Kaloshian I (2003) Are roots special? Nematodes have their say. Physiol Mol Plant Pathol 62:115–123. https://doi. org/10.1016/S0885-5765(03)00045-6
- Black R, Jonglaekha N (1989) Plant diseases and other aspects of plant protection in northern thailand with special reference to highland development programmes. Trop Pest Manag 35:289– 296. https://doi.org/10.1080/09670878909371383
- Boland GJ, Melzer MS, Hopkin A, Higgins V, Nassuth A (2004) Climate change and plant diseases in Ontario. Canadian J Plant Pathol 26:335–350
- Bolton MD, Kolmer JA, Garvin DF (2008) Wheat leaf rust caused by Puccinia triticina. Mol Plant Pathol 9:563–575. https:// doi.org/10.1111/j.1364-3703.2008.00487.x
- 57. Borines L, Sagarino R, Calamba R et al (2015) Potential of Chitosan for the Control of Tomato Bacterial Wilt Caused by Ralstonia solanacearum (Smith) Yabuuchi et al. Ann Trop Res 37:57–69. https://doi.org/10.32945/atr3725.2015
- Botella MA, Parker JE, Frost LN et al (1998) Three genes of the arabidopsis RPP1 complex resistance locus recognize distinct Peronospora parasitica avirulence determinants. Plant Cell 10:1847–1860. https://doi.org/10.1105/tpc.10.11.1847
- Bovallius Å, Bucht B, Roffey R, Anas P (1978) Long-Range Transmission of Bacteria. Appl Environ Microbiol 35:1231– 1232. https://doi.org/10.1111/j.1749-6632.1980.tb18922.x
- Bove JM, Garnier M (2003) Phloem-and xylem-restricted plant pathogenic bacteria. Plant Sci 164:423–438
- 61. Bowers RM, Clements N, Emerson JB et al (2013) Seasonal variability in bacterial and fungal diversity of the near-surface

atmosphere. Environ Sci Technol 47:12097–12106. https://doi. org/10.1021/es402970s

- Bowers RM, McLetchie S, Knight R, Fierer N (2011) Spatial variability in airborne bacterial communities across land-use types and their relationship to the bacterial communities of potential source environments. ISME J 5:601–612. https://doi. org/10.1038/ismej.2010.167
- Bowers RM, Sullivan AP, Costello EK et al (2011) Sources of bacteria in outdoor air across cities in the midwestern United States. Appl Environ Microbiol 77:6350–6356. https://doi.org/ 10.1128/AEM.05498-11
- Broders KD, Boland GJ (2011) Reclassification of the butternut canker fungus, Sirococcus clavigignenti-juglandacearum, into the genus Ophiognomonia. Fungal Biol 115:70–79. https://doi. org/10.1016/j.funbio.2010.10.007
- Brodie EL, DeSantis TZ, Moberg Parker JP et al (2007) Urban aerosols harbor diverse and dynamic bacterial populations. Proc Natl Acad Sci U S A 104:299–304.https://doi.org/10.1073/pnas. 0608255104
- Brown JKM, Hovmøll MS (2002) Aerial dispersal of pathogens on the global and continental scales and its impact on plant disease. Science (80-) 297:537–541. https://doi.org/10.1126/scien ce.1072678
- Brown RM, Larson DA, Bold HC (1964) Airborne algae: their abundance and heterogeneity. Science 143:583–585. https://doi. org/10.1126/science.143.3606.583
- Brunt AA, CAB International (1996) Viruses of plants: descriptions and lists from the VIDE database / edited by Alan Brunt ... [et al.]. CAB International, Wallingford, Oxon, UK. http://www.loc.gov/catdir/enhancements/fy0637/96185674-t.html
- Bunster L, Fokkema NJ, Schippers B (1989) Effect of Surface-Active Pseudomonas spp. on Leaf Wettability. Appl Environ Microbiol 55:1340–1345. https://doi.org/10.1128/aem.55.6. 1340-1345.1989
- Burdsall AC, Xing Y, Cooper CW, Harper WF (2021) Bioaerosol emissions from activated sludge basins: Characterization, release, and attenuation. Sci Total Environ 753:141852
- Burrows SM, Butler T, Jöckel P et al (2009) Bacteria in the global atmosphere - Part 2: Modeling of emissions and transport between different ecosystems. Atmos Chem Phys 9:9281–9297. https://doi.org/10.5194/acp-9-9281-2009
- Burrows SM, Rayner PJ, Butler T, Lawrence MG (2013) Estimating bacteria emissions from inversion of atmospheric transport: Sensitivity to modelled particle characteristics. Atmos Chem Phys 13:5473–5488. https://doi.org/10.5194/acp-13-5473-2013
- Burt PJA (1995) The potato and the pathogen: the Irish potato famine of 1845. Weather 50:342–346. https://doi.org/10.1002/j. 1477-8696.1995.tb05502.x
- Butler MI, Stockwell PA, Black MA et al (2013) Pseudomonas syringae pv. actinidiae from recent outbreaks of kiwifruit bacterial canker belong to different clones that originated in China. PLoS One 8:e57464. https://doi.org/10.1371/journal.pone.00574 64
- Calderon C, Lacey J, McCartney HA, Rosas I (1995) Seasonal and diurnal variation of airborne basidiomycete spore concentrations in Mexico City. Grana 34:260–268. https://doi.org/10.1080/ 00173139509429055
- Cao MJ, Atta S, Liu YQ et al (2009) First report of Citrus bent leaf viroid and Citrus dwarfing viroid from Citrus in Punjab, Pakistan. Plant Dis 93:840–840
- Carabez JRS, Ortiz DT, Pérez MRV, Peña HB (2019) The avocado sunblotch viroid: An invisible foe of avocado. Viruses 11:491. https://doi.org/10.3390/v11060491
- 78. Card SD, Tapper BA, Lloyd-West C, Wright KM (2013) Assessment of fluorescein-based fluorescent dyes for tracing

Neotyphodium endophytes in planta. Mycologia 105:221–229. https://doi.org/10.3852/12-062

- 79. Carducci A, Arrighi S, Ruschi A (1995) Detection of coliphages and enteroviruses in sewage and aerosol from an activated sludge wastewater treatment plant. Lett Appl Microbiol 21:207–209. https://doi.org/10.1111/j.1472-765X.1995.tb01042.x
- Carducci A, Tozzi E, Rubulotta E et al (2000) Assessing airborne biological hazard from urban wastewater treatment. Water Res 34:1173–1178. https://doi.org/10.1016/S0043-1354(99)00264-X
- Carotenuto F, Georgiadis T, Gioli B et al (2017) Measurements and modeling of surface-atmosphere exchange of microorganisms in Mediterranean grassland. Atmos Chem Phys 17:14919– 14936. https://doi.org/10.5194/acp-17-14919-2017
- Carvalho CR, Fernandes RC, Carvalho GMA et al (2011) Cryptosexuality and the genetic diversity paradox in coffee rust, Hemileia vastatrix. PLoS One 6:e26387. https://doi.org/10.1371/ journal.pone.0026387
- Castillo JA, Staton SJR, Taylor TJ et al (2012) Exploring the feasibility of bioaerosol analysis as a novel fingerprinting technique. Anal Bioanal Chem 403:15–26. https://doi.org/10.1007/ s00216-012-5725-0
- Chalanska A, Bogumił A, Łabanowski G (2017) Management of foliar nematode Aphelenchoides ritzemabosi on Anemone hupehensis using plant extracts and pesticides. J Plant Dis Prot 124:437–443
- Chase AR, Daughtrey ML (2013) Rose downy mildew review. Greenh Prod News Mag. 2013:32–34
- Chaudhary A, Yadav J, Gupta A, Gupta K (2021) Integrated disease management of early blight (Alternaria Solani) of Potato. 77–81. https://doi.org/10.26480/trab.02.2021.77.81
- Chen K, Wang Y, Zhang R et al (2019) CRISPR/Cas genome editing and precision plant breeding in agriculture. Annu Rev Plant Biol 70:667–697. https://doi.org/10.1146/annurev-arpla nt-050718-100049
- Chen W, Wellings C, Chen X et al (2014) Wheat stripe (yellow) rust caused by Puccinia striiformis f. sp. tritici. Mol Plant Pathol 15:433–446. https://doi.org/10.1111/mpp.12116
- Chen X, Ran P, Ho K et al (2012) Concentrations and size distributions of airborne microorganisms in guangzhou during summer. Aerosol Air Qual Res 12:1336–1344. https://doi.org/10. 4209/aaqr.2012.03.0066
- Chen Y-H, Yan C, Yang Y-F, Ma J-X (2021) Quantitative microbial risk assessment and sensitivity analysis for workers exposed to pathogenic bacterial bioaerosols under various aeration modes in two wastewater treatment plants. J Neurol Sci 755:142615. https://doi.org/10.1016/j.scitotenv.2020.142615
- Chittem K, Porter L, McPhee K, Khan M, Goswami RS (2010) Fusarium avenaceum as causal agent of root rot in field peas and its control. In Phytopathology. AMER PHYTOPATHOLOGI-CAL SOC, 3340 PILOT KNOB ROAD, ST PAUL, MN 55121 USA, pp S25–S25
- 92. Cho BC, Hwang CY (2011) Prokaryotic abundance and 16S rRNA gene sequences detected in marine aerosols on the East Sea (Korea). FEMS Microbiol Ecol 76:327–341. https://doi.org/ 10.1111/j.1574-6941.2011.01053.x
- Cho WK, Jo Y, Jo KM, Kim KH (2013) A current overview of two viroids that infect chrysanthemums: Chrysanthemum stunt viroid and Chrysanthemum chlorotic mottle viroid. Viruses 5:1099–1113. https://doi.org/10.3390/v5041099
- Choi W, Dean RA (1997) The adenylate cyclase gene MAC1 of Magnaporthe grisea controls appressorium formation and other aspects of growth and development. Plant Cell 9:1973–1983. https://doi.org/10.1105/tpc.9.11.1973
- 95. Chou C, Castilla N, Hadi B et al (2020) Rice blast management in Cambodian rice fields using Trichoderma harzianum and a

resistant variety. Crop Prot 135:104864. https://doi.org/10.1016/j. cropro.2019.104864

- Christensen LS, Mousing J, Mortensen S, Soerensen KJ, Strandbygaard SB, Henriksen CA, Andersen JB (1990) Evidence of long distance airborne transmission of Aujeszky's disease (pseudorabies) virus. Vet Rec 127:471–474
- Christensen LS, Mortensen S, Bøtner A, Strandbygaard BS, Rønsholt L, Henriksen CA, Andersen JB (1993) Further evidence of long distance airborne transmission of Aujeszky's disease (pseudorabies) virus. Vet Rec 132:317–321. https://doi.org/10.1136/ vr.132.13.317
- Ciliberti N, Fermaud M, Roudet J, Rossi V (2015) Environmental conditions affect Botrytis cinerea infection of mature grape berries more than the strain or transposon genotype. Phytopathol 105:1090–1096
- Coccia AM, Gucci PMB, Lacchetti I et al (2010) Airborne microorganisms associated with waste management and recovery: biomonitoring methodologies. Ann Ist Super Sanità 46:288–292. https://doi.org/10.4415/ANN
- Comstock JC (2000) Smut. In: Rott P, Bailey RA, Comstock JC, Croft BJ, Sauntally AS (eds) A Guide to Sugarcane Diseases. CIRAD/ISCCT Publishers; Montpellier, France, pp 181–185
- Cook RJ, Flentj¢ NT (1967) Chlamydospore germination and germling survival of Fusarium solani f. pisi in soil as affected by soil water and pea seed exudation. Phytopathol 57:178–82
- 102. Cother EJ, Noble DH, van de Ven RJ et al (2010) Bacterial pathogens of rice in the Kingdom of Cambodia and description of a new pathogen causing a serious sheath rot disease. Plant Pathol 59:944–953. https://doi.org/10.1111/j.1365-3059.2010.02310.x
- Cox CS, Wathes CM (1995) Bioaerosols Handbook. Lewis Publishers, Boca Raton
- 104. Crandall SG, Rahman A, Quesada-Ocampo LM et al (2018) Advances in diagnostics of downy mildews: Lessons learned from other oomycetes and future challenges. Plant Dis 102:265– 275. https://doi.org/10.1094/PDIS-09-17-1455-FE
- 105. Cranshaw WS (2004) Onion thrips in onions XXV. Agricultural Experiment Station, Colorado State University, Fort Collins, CO. http://wiki.bugwood.org/uploads/OnionThrips-Onions.pdf. Accessed Apr 2015
- Cummins GB, Hiratsuka Y (2003) Illustrated Genera of Rust Fungi. American Phytopathol. Society Press, St Paul, MN
- 107. Czosnek H (2008) Acquisition, circulation and transmission of begomoviruses by their whitefly vectors. In: Palombo EA, Kirkwood CD (eds) Viruses in the Environment. Research Signpost, Trivandrum, Kerala, India, pp 29–44
- Damiri N (2011) Climate change, environment and plant diseases development. In: Proceedings of the International Seminar. Sriwijaya University cooperation with CRISU and CUPT, pp 200–205
- 109. Dang T, Lavagi-Craddock I, Bodaghi S, Vidalakis G (2021) Next-generation sequencing identification and characterization of MicroRNAs in dwarfed citrus trees infected with citrus dwarfing viroid in high-density plantings. Front Microbiol 2. https:// doi.org/10.3389/fmicb.2021.646273
- Danhorn T, Fuqua C (2007) Biofilm formation by plant-associated bacteria. Annu Rev Microbiol 61:401–422. https://doi.org/ 10.1146/annurev.micro.61.080706.093316
- 111. Dato KMG, Dégbègni MR, Atchadé MN et al (2021) Spatial parameters associated with the risk of banana bunchy top disease in smallholder systems. PLoS One 16:e0260976. https://doi.org/ 10.1371/journal.pone.0260976
- Davies CR, Wohlgemuth F, Young T et al (2021) Evolving challenges and strategies for fungal control in the food supply chain. Fungal Biol Rev 36:15–26

- Davis EL, Hussey RS, Mitchum MG, Baum TJ (2008) Parasitism proteins in nematode-plant interactions. Curr Opin Plant Biol 11:360–366. https://doi.org/10.1016/j.pbi.2008.04.003
- 114. De Beer ZW, Marincowitz S, Duong T, Wingfield M (2017) Bretziella, a new genus to accommodate the oak wilt fungus, Ceratocystis fagacearum (Microascales, Ascomycota). MycoKeys 27:1–19. https://doi.org/10.3897/mycokeys.27.20657
- 115. de Gruyter J, Aveskamp MM, Woudenberg JHC et al (2009) Molecular phylogeny of Phoma and allied anamorph genera: towards a reclassification of the Phoma complex. Mycol Res 113:508–519
- 116. de Silva DD, Groenewald JZ, Crous PW et al (2019) Identification, prevalence and pathogenicity of Colletotrichum species causing anthracnose of Capsicum annuum in Asia. IMA Fungus 10:8. https://doi.org/10.1186/s43008-019-0001-y
- 117. Dean R, Van Kan JAL, Pretorius ZA et al (2012) The Top 10 fungal pathogens in molecular plant pathology. Mol Plant Pathol 13:414–430. https://doi.org/10.1111/j.1364-3703.2011.00783.x
- Decraemer W, Hunt DJ (2006) Structure and classification. In: Nematology P (ed) Perry RN, Moens M. CAB International, Wallingford, UK, pp 3–32
- Decraemer W, Hunt DJ (2013) Structure and classification. In: Perry RN, Moens M (Eds) Plant Nematology, 2 edn. Wallingford, UK: CABI Publishing, pp 3–39. https://doi.org/10.1079/97817 80641515.0003
- 120. DeLeon-Rodriguez N, Lathem TL, Rodriguez-R LM et al (2013) Microbiome of the upper troposphere: Species composition and prevalence, effects of tropical storms, and atmospheric implications. Proc Natl Acad Sci U S A 110:2575–2580.https://doi.org/ 10.1073/pnas.1212089110
- Delmotte N, Knief C, Chaffron S et al (2009) Community proteogenomics reveals insights into the physiology of phyllosphere bacteria. Proc Natl Acad Sci U S A 106:16428–16433.https:// doi.org/10.1073/pnas.0905240106
- 122. Delort AM, Vaïtilingom M, Amato P et al (2010) A short overview of the microbial population in clouds: Potential roles in atmospheric chemistry and nucleation processes. Atmos Res 98:249–260. https://doi.org/10.1016/j.atmosres.2010.07.004
- Demirci E, Timur Döken M (1998) Host penetration and infection by the anastomosis groups of Rhizoctonia solani kühn isolated from potatoes. Turkish J Agric For 22:609–613. https://doi. org/10.3906/tar-97020
- 124. Després VR, Alex Huffman J, Burrows SM et al (2012) Primary biological aerosol particles in the atmosphere: A review. Tellus, Ser B Chem Phys Meteorol 64. https://doi.org/10.3402/tellusb.v64i0.15598
- 125. Després VR, Nowoisky JF, Klose M et al (2007) Characterization of primary biogenic aerosol particles in urban, rural, and highalpine air by DNA sequence and restriction fragment analysis of ribosomal RNA genes. Biogeosciences 4:1127–1141. https://doi. org/10.5194/bg-4-1127-2007
- 126. Di-Giovanni F, Kevan PG, Nasr ME (1995) The variability in settling velocities of some pollen and spores. Grana 34:39–44. https://doi.org/10.1080/00173139509429031
- 127. Di Giorgio C, Krempff A, Guiraud H et al (1996) Atmospheric pollution by airborne microorganisms in the city of Marseilles. Atmos Environ 30:155–160. https://doi.org/10.1016/1352-2310(95)00143-M
- Di Serio F, Flores R, Verhoeven JTJ et al (2014) Current status of viroid taxonomy. Arch Virol 159:3467–3478. https://doi.org/ 10.1007/s00705-014-2200-6
- 129. Diaz-Cruz GA, Smith CM, Wiebe KF, Cassone BJ (2017) First Complete Genome Sequence of Tobacco necrosis virus D Isolated from Soybean and from North America. Genome Announc 5:e00781-e817. https://doi.org/10.1128/genomeA.00781-17

- Diehl K, Quick C, Matthias-Maser S et al (2001) The ice nucleating ability of pollen Part I: Laboratory studies in deposition and condensation freezing modes. Atmos Res 58:75–87
- 131. Dita M, Barquero M, Heck D, Mizubuti ESG et al (2018) Fusarium Wilt of banana: Current knowledge on epidemiology and research needs toward sustainable disease management. Front Plant Sci 9:1468. https://doi.org/10.3389/fpls.2018.01468
- 132. Dixon MS, Jones DA, Keddie JS et al (1996) The tomato Cf-2 disease resistance locus comprises two functional genes encoding leucine-rich repeat proteins. Cell 84:451–459. https://doi.org/10. 1016/S0092-8674(00)81290-8
- 133. Doddaraju P, Kumar P, Gunnaiah R, Gowda AA et al (2019) Reliable and early diagnosis of bacterial blight in pomegranate caused by Xanthomonas axonopodis pv. punicae using sensitive PCR techniques. Sci Rep 9:10097. https://doi.org/10.1038/ s41598-019-46588-9
- 134. Dong L, Qi J, Shao C et al (2016) Concentration and size distribution of total airborne microbes in hazy and foggy weather. Sci Total Environ 541:1011–1018. https://doi.org/10.1016/j.scito tenv.2015.10.001
- Dongsheng D (2006) From the smallest virus to the biggest gene: Marching towards gene therapy for Duchenne muscular dystrophy. Discov Med 6:103–108
- Dordas C (2008) Role of nutrients in controlling plant diseases in sustainable agriculture. A review. Agron Sustain Dev 28:33–46
- Douwes J, Thorne P, Pearce N, Heederik D (2003) Bioaerosol health effects and exposure assessment: Progress and prospects. Ann Occup Hyg 47:187–200. https://doi.org/10.1093/annhyg/ meg032
- Dowd SE, Gerba CP, Pepper IL, Pillai SD (2000) Bioaerosol transport modeling and risk assessment in relation to biosolid placement. Encycl Ecol 29:343–348. https://doi.org/10.1016/ B978-0-12-409548-9.11137-6
- Du M, Schardl CL, Nuckles EM, Vaillancourt LJ (2005) Using mating-type gene sequences for improved phylogenetic resolution of Collectotrichum species complexes. Mycologia 97:641– 658. https://doi.org/10.1080/15572536.2006.11832795
- 140. Du P, Du R, Ren W et al (2018) Variations of bacteria and fungi in PM2.5 in Beijing, China. Atmos Environ 172:55–64
- Dubovik IE (2002) Migrations of aerophytic algae and their colonization on different substrata. Int J Algae 4:48–55
- 142. Duchaine C, Roy CJ (2020) Bioaerosols and airborne transmission: Integrating biological complexity into our perspective. Sci Total Environ 825:154117. https://doi.org/10.1016/j.scitotenv. 2022.154117
- 143. Dungan RS, Leytem AB (2009) Qualitative and quantitative methodologies for determination of airborne microorganisms at concentrated animal-feeding operations. World J Microbiol Biotechnol 25:1505–1518. https://doi.org/10.1007/ s11274-009-0043-1
- Duplessis S, Joly DJ, Dodds PN (2012) Rust effectors. In: Martin F, Kamoun S (eds) Effectors in Plant - Microbes Interactions. Wiley Blackwell, Oxford, pp 155–193
- 145. Eastburn DM, Degennaro MM, Delucia EH et al (2010) Elevated atmospheric carbon dioxide and ozone alter soybean diseases at SoyFACE. Glob Chang Biol 16:320–330. https://doi.org/10. 1111/j.1365-2486.2009.01978.x
- 146. EFSA Plh Panel (2014) Scientific opinion on the pest categorisation of Verticillium dahliae Kleb. EFSA J 12:3928. https://doi. org/10.2903/j.efsa.2014.3928
- 147. El-Gamal AD (2008) Aerophytic cyanophyceae (Cyanobacteria) from some Cairo Districts. Egypt Pakistan J Biol Sci 11:1293–1302
- 148. Elbert W, Taylor PE, Andreae MO, Pöschl U (2007) Contribution of fungi to primary biogenic aerosols in the atmosphere: Wet and dry discharged spores, carbohydrates, and inorganic

ions. Atmos Chem Phys 7:4569–4588. https://doi.org/10.5194/ acp-7-4569-2007

- Elbert W, Weber B, Burrows S et al (2012) Contribution of cryptogamic covers to the global cycles of carbon and nitrogen. Nat Geosci 5:459–462. https://doi.org/10.1038/ngeo1486
- Elleuch A, Marrakchi M, Perreault JP, Fakhfakh H (2003) First report of Australian grapevine viroid from the Mediterranean region. J Plant Pathol 53–57
- 151. Ellis Jeffrey G, Lagudah Evans S, Spielmeyer Wolfgang, Dodds Peter N (2014) The past, present and future of breeding rust resistant wheat. Front. Plant Sci 5. https://doi.org/10.3389/fpls. 2014.00641
- Ellstrand NC (1992) Gene Flow by Pollen: Implications for plant conservation genetics. Oikos 63:77–86. https://doi.org/10.2307/ 3545517
- 153. Elsayed TR, Jacquiod S, Nour EH et al (2020) Biocontrol of bacterial Wilt disease through complex interaction between tomato plant, antagonists, the indigenous rhizosphere microbiota, and Ralstonia solanacearum. Front Microbiol 10:1–15. https://doi. org/10.3389/fmicb.2019.02835
- Endo BY, Veech JA (1970) Morphology and histochemistry of soybean roots infected with heterodera glycines. Phytopathology 60:1493–1498
- 155. Eranthodi A, Schneiderman D, Harris LJ et al (2020) Enniatin production influences Fusarium avenaceum virulence on potato tubers, but not on durum wheat or peas. Pathogens. https://doi. org/10.3390/pathogens9020075
- 156. Erginbas-Orakci G, Morgounov A, Dababat A (2018) Determination of resistance in winter wheat genotypes to the dryland root rots caused by Fusarium culmorum in Turkey. Uluslararası Tarım ve Yaban Hayatı Bilimleri Dergisi. 4:193–202. https://doi.org/10. 24180/ijaws.414501
- 157. Escobar C, Barcala M, Cabrera J, Fenoll C (2015) Overview of root-knot nematodes and giant cells. In: Escobar C, Fenoll C (eds) Plant nematode interactions: a view on compatible interrelationships, vol 73, Advances in Botanical Research. (Oxford, UK: Elsevier), pp 1–32. https://doi.org/10.1016/bs.abr.2015.01.001
- 158. Fadda Z, Daròs JA, Fagoaga C et al (2003) Eggplant latent viroid, the candidate type species for a new genus within the family avsunviroidae (Hammerhead Viroids). J Virol 77:6528– 6532. https://doi.org/10.1128/jvi.77.11.6528-6532.2003
- 159. Fahlgren C, Bratbak G, Sandaa RA et al (2011) Diversity of airborne bacteria in samples collected using different devices for aerosol collection. Aerobiologia (Bologna) 27:107–120. https://doi.org/10.1007/s10453-010-9181-z
- 160. Fahlgren C, Gómez-Consarnau L, Zábori J et al (2015) Seawater mesocosm experiments in the Arctic uncover differential transfer of marine bacteria to aerosols. Environ Microbiol Rep 7:460–470. https://doi.org/10.1111/1758-2229.12273
- 161. Fan LM, Zhao Z, Assmann SM (2004) Guard cells: A dynamic signaling model. Curr Opin Plant Biol 7:537–546. https://doi. org/10.1016/j.pbi.2004.07.009
- 162. Fan XL, Bezerra JDP, Tian CM, Crous PW (2020) Cytospora (Diaporthales) in China. Persoonia 45:1–45. https://doi.org/ 10.3767/persoonia.2020.45.01
- Fang Z, Ouyang Z, Zheng H et al (2007) Culturable airborne bacteria in outdoor environments in Beijing, China. Microb Ecol 54:487–496
- 164. Favero-Longo SE, Sandrone S, Matteucci E et al (2014) Spores of lichen-forming fungi in the mycoaerosol and their relationships with climate factors. Sci Total Environ 466–467:26–33. https://doi.org/10.1016/j.scitotenv.2013.06.057
- 165. Ference CM, Gochez AM, Behlau F et al (2018) Recent advances in the understanding of Xanthomonas citri ssp. citri pathogenesis and citrus canker disease management. Mol Plant Pathol 19:1302–1318. https://doi.org/10.1111/mpp.12638

- 166. Figueroa M, Hammond-Kosack KE, Solomon PS (2018) A review of wheat diseases—a field perspective. Mol Plant Pathol 19:1523–1536. https://doi.org/10.1111/mpp.12618
- 167. Fisher MC, Henk DA, Briggs CJ et al (2012) Emerging fungal threats to animal, plant and ecosystem health. Nature 484:186– 194. https://doi.org/10.1038/nature10947
- 168. Fitt BDL, Brun H, Barbetti MJ, Rimmer SR (2006) World-wide importance of phoma stem canker (Leptosphaeria maculans and L. biglobosa) on oilseed Rape (Brassica napus). Eur J Plant Pathol 114:3–15. https://doi.org/10.1007/s10658-005-2233-5
- 169. Flores R, Hernández C, Llácer G, Desvignes JC (1991) Identification of a new viroid as the putative causal agent of pear blister canker disease. J Gen Virol 72:1199–1204. https://doi. org/10.1099/0022-1317-72-6-1199
- 170. Flores R, Randles JW, Bar-Joseph M, Diener TO (1998) A proposed scheme for viroid classification and nomenclature. Adv Virol 143:623–629
- 171. Flores-Tena FJ, Pardavé LM, Valenzuela I (2007) Estudio aerobiológico de la zona San Nicolás, municipio de Aguascalientes, México. Invest Cienc 15:13–18
- 172. Fonseca S, Chico JM, Solano R (2009) The jasmonate pathway: the ligand, the receptor and the core signalling module. Curr Opin Plant Biol 12:539–547. https://doi.org/10.1016/j.pbi.2009.07.013
- 173. Forghani F, Hajihassani A (2020) Recent advances in the development of environmentally benign treatments to control rootknot nematodes. Front Plant Sci 11:1125. https://doi.org/10. 3389/fpls.2020.01125
- 174. Fotopoulos V, Gilbert MJ, Pittman JK et al (2003) The monosaccharide transporter gene, AtSTP4, and the cell-wall invertase, Atβfruct1, are induced in arabidopsis during infection with the fungal biotroph Erysiphe cichoracearum. Plant Physiol 132:821–829. https://doi.org/10.1104/pp.103.021428
- 175. Fox K, Fox A, Elßner T et al (2010) MALDI-TOF mass spectrometry speciation of staphylococci and their discrimination from micrococci isolated from indoor air of schoolrooms. J Environ Monit 12:917–923. https://doi.org/10.1039/b925250a
- 176. Fracchia L, Pietronave S, Rinaldi M, Martinotti MG (2006) The assessment of airborne bacterial contamination in three composting plants revealed site-related biological hazard and seasonal variations. J Appl Microbiol 100:973–984. https://doi.org/10. 1111/j.1365-2672.2006.02846.x
- 177. Fradin EF, Thomma BPHJ (2006) Physiology and molecular aspects of Verticillium wilt diseases caused by V. dahliae and V. albo-atrum. Mol Plant Pathol 7:71–86. https://doi.org/10.1111/j. 1364-3703.2006.00323.x
- 178. Franceschini A, Szklarczyk D, Frankild S et al (2013) STRING v9.1: Protein-protein interaction networks, with increased coverage and integration. Nucleic Acids Res 41:808–815. https://doi. org/10.1093/nar/gks1094
- 179. Francis DM, Kabelka E, Bell J et al (2001) Resistance to bacterial canker in tomato (Lycopersicon hirsutum LA407) and its progeny derived from crosses to L. esculentum. Plant Dis 85:1171–1176. https://doi.org/10.1094/PDIS.2001.85.11.1171
- Frohlich-Nowoisky J, Burrows SM, Xie Z et al (2012) Biogeography in the air: Fungal diversity over land and oceans. Biogeosciences 9:1125–1136. https://doi.org/10.5194/bg-9-1125-2012
- Frohlich-Nowoisky J, Hill TCJ, Pummer BG et al (2015) Ice nucleation activity in the widespread soil fungus Mortierella alpina. Biogeosciences 12:1057–1071. https://doi.org/10.5194/ bg-12-1057-2015
- Frohlich-Nowoisky J, Kampf CJ, Weber B et al (2016) Bioaerosols in the Earth system: Climate, health, and ecosystem interactions. Atmos Res 182:346–376. https://doi.org/10.1016/j.atmos res.2016.07.018
- Frohlich-Nowoisky J, Pickersgill DA, Després VR, Pöschl U (2009) High diversity of fungi in air particulate matter. Proc Natl

Acad Sci U S A 106:12814–12819.https://doi.org/10.1073/pnas. 0811003106

- 184. Fu FH, Li SF, Jiang DM, Wang HQ, Liu AQ, Sang LW (2011) First report of Coleus blumei viroid 2 from commercial coleus in China. Plant Dis 95:494–494
- 185. Fuchs M, Abawi GS, Marsella-Herrick P et al (2010) Occurrence of tomato ringspot virus and Tobacco ringspot virus in highbush blueberry in New York State. J Plant Pathol 92:451–459
- Gomez SY, Arbel aez G, (2005) Effect of temperature on the latency periodand production of sporangia of Peronospora sparsa Berkeley on three varieties of rose. Agron Colomb 23:246–255
- 187. Gago S, Elena SF, Flores R, Sanjuán R (2009) Extremely high mutation rate of a hammerhead viroid. Science (80-) 323:1308. https://doi.org/10.1126/science.1169202
- Galande DR, Simon S (2019) Effect of Intercropping on Purple Blotch (Alternaria porri) of Onion (Allium cepa L.). Int J Curr Microbiol Appl Sci 8:1105–1111. https://doi.org/10.20546/ijcmas.2019.802.129
- Gallet R, Michalakis Y, Blanc S (2018) Vector-transmission of plant viruses and constraints imposed by virus-vector interactions. Curr Opin Virol 33:144–150. https://doi.org/10.1016/j. coviro.2018.08.005
- 190. Gandía M, Rubio L, Palacio A, Duran-Vila N (2005) Genetic variation and population structure of an isolate of Citrus exocortis viroid (CEVd) and of the progenies of two infectious sequence variants. Arch Virol 150:1945–1957. https://doi.org/10.1007/ s00705-005-0570-5
- 191. Ganthaler A, Mayr S (2015) Temporal variation in airborne spore concentration of Chrysomyxa rhododendri: Correlation with weather conditions and consequences for Norway spruce infection. For Pathol 45:443–449. https://doi.org/10.1111/efp. 12190
- 192. Gao M, Jia R, Qiu T et al (2015) Seasonal size distribution of airborne culturable bacteria and fungi and preliminary estimation of their deposition in human lungs during non-haze and haze days. Atmos Environ 118:203–210. https://doi.org/10. 1016/j.atmosenv.2015.08.004
- 193. Gao M, Yan X, Qiu T et al (2016) Variation of correlations between factors and culturable airborne bacteria and fungi. Atmos Environ 128:10–19. https://doi.org/10.1016/j.atmosenv. 2015.12.008
- García-Blázquez G, Göker M, Voglmayr H et al (2008) Phylogeny of Peronospora, parasitic on Fabaceae, based on ITS sequences. Mycol Res 112:502–512. https://doi.org/10.1016/j. mycres.2007.10.007
- 195. Garcia-Ruiz H (2019) Host factors against plant viruses. Mol Plant Pathol 20:1588–1601. https://doi.org/10.1111/mpp. 12851
- 196. Gašić K, Pavlović, Santander RD et al (2018) First report of pseudomonas syringae pv. syringae associated with bacterial blossom blast on apple (Malus pumila) in the United States. Plant Dis 102:1848. https://doi.org/10.1094/ PDIS-01-18-0184-PDN
- 197. Genitsaris S, Kormas KA, Moustaka-Gouni M (2011) Airborne algae and cyanobacteria: occurrence and related health effects. Front Biosci 3:772–787. https://doi.org/10.1109/leoswt.2008. 4444364
- 198. Ghanim M, Medina V (2007) Localization of tomato yellow leaf curl virus in its whitefly vector Bemisia tabaci. Tomato Yellow Leaf Curl Virus Dis Manag Mol Biol Breed Resist 171–183. https://doi.org/10.1007/978-1-4020-4769-5\_10
- 199. Gheysen G, Mitchum MG (2011) How nematodes manipulate plant development pathways for infection. Curr Opin Plant Biol 14:415–421. https://doi.org/10.1016/j.pbi.2011.03.012
- 200. Ghini R, Hamada E, Pedro JM, Marengo J, Gonçalves R (2008) Risk analysis of climate change on coffee nematodes and leaf

miner in Brazil. Pesqui Agrop Bras – PAB 43. https://doi.org/ 10.1590/S0100-204X2008000200005

- Gill CC, Westdal PH (1966) Virus Diseases of cereals and vector populations in the Canadian prairies during 1965. Canadian Plant Dis Surv 46:18
- 202. Gimenez-Ibanez S, Boter M, Ortigosa A et al (2017) JAZ2 controls stomata dynamics during bacterial invasion. New Phytol 213:1378–1392. https://doi.org/10.1111/nph.14354
- 203. Gladieux P, Zhang XG, Afoufa-Bastien D et al (2008) On the origin and spread of the scab disease of apple: Out of central Asia. PLoS One 3:e1455. https://doi.org/10.1371/journal.pone. 0001455
- Gnanamanickam SS, Priyadarisini VB, Narayanan NN, Vasudevan P, Kavitha S (1999) An overview of Bacterial blight disease of rice and strategies for its management. Curr Sci 77:1435–1443
- Gohlke J, Deeken R (2014) Plant responses to Agrobacterium tumefaciens and crown gall development. Front Plant Sci 5:155. https://doi.org/10.3389/fpls.2014.00155
- 206. Górny RL, Reponen T, Willeke K et al (2002) Fungal fragments as indoor air biocontaminants. Appl Environ Microbiol 68:3522– 3531. https://doi.org/10.1128/AEM.68.7.3522-3531.2002
- 207. Goverse A, Smant G (2014) The activation and suppression of plant innate immunity by parasitic nematodes. Annu Rev Phytopathol 52:243–265. https://doi.org/10.1146/annur ev-phyto-102313-050118
- Goyeau H, Park R, Schaeffer B, Lannou C (2006) Distribution of pathotypes with regard to host cultivars in French wheat leaf rust populations. Phytopathology 96:264–273. https://doi.org/10. 1094/PHYTO-96-0264
- Graham B, Guyon P, Maenhaut W et al (2003) Composition and diurnal variability of the natural Amazonian aerosol. J Geophys Res Atmos 108. https://doi.org/10.1029/2003jd004049
- 210. Grant MR, Godiard L, Straube E, Ashfield T, Lewald J, Sattler A, Innes RW, Dangl JL (1995) Structure of the Arabidopsis RPM1 gene enabling dual specificity disease resistance. Science 269:843–846
- 211. Green JR, Carver TLW, Gurr SJ (2002) The formation and function of infection and feeding structures. In Bélanger RR, Bushnell WR, Dik AJ, Carver TLW (eds) The Powdery Mildews: A Comprehensive Treatise, American Phytopathological Society Press, pp 66–82. http://hdl.handle.net/2160/4098
- 212. Greer G, Saunders C (2012) The costs of Psa-V to the New Zealand kiwifruit industry and the wider community. Agribusiness and Economics Researh Unit, Lincoln University, New Zealand
- 213. Gregory PH (1973) The Microbiology of the Atmosphere, 2nd edn. Leonard Hill Books, Plant Series Monographs, Plymouth
- Grenville-Briggs LJ, Anderson VL, Fugelstad J et al (2008) Cellulose synthesis in Phytophthora infestans is required for normal appressorium formation and successful infection of potato. Plant Cell 20:720–738. https://doi.org/10.1105/tpc.107.052043
- 215. Griffin DH (1996) Fungal physiology. John Wiley & Sons
- Griffin DW (2004) Terrestrial microorganisms at an altitude of 20,000 m in Earth's atmosphere. Aerobiologia (Bologna) 20:135– 140. https://doi.org/10.1023/B:AERO.0000032948.84077.12
- 217. Griffin DW, Garrison VH, Herman JR, Shinn EA (2001) African desert dust in the caribbean atmosphere: Microbiology and public health. Entomol Exp Appl 17:203–213
- Grinshpun SA, Clark JM (2005) Measurement and characterization of bioaerosols. J Aerosol Sci 36:553–555
- Grisoli P, Rodolfi M, Villani S et al (2009) Assessment of airborne microorganism contamination in an industrial area characterized by an open composting facility and a wastewater treatment plant. Environ Res 109:135–142. https://doi.org/10.1016/j.envres.2008.11.001

- Groth DE (2008) Effects of cultivar resistance and single fungicide application on rice sheath blight, yield, and quality. Crop Prot 27:1125–1130. https://doi.org/10.1016/j.cropro.2008.01.010
- 221. Guo J, Xiong Y, Shi C et al (2020) Characteristics of airborne bacterial communities in indoor and outdoor environments during continuous haze events in Beijing: Implications for health care. Environ Int 139:105721. https://doi.org/10.1016/j.envint. 2020.105721
- 222. Haas D, Galler H, Luxner J et al (2013) The concentrations of culturable microorganisms in relation to particulate matter in urban air. Atmos Environ 65:215–222. https://doi.org/10.1016/j. atmosenv.2012.10.031
- 223. Hagerty CH, Cuesta-Marcos A, Cregan PB et al (2015) Mapping Fusarium solani and Aphanomyces euteiches root rot resistance and root architecture quantitative trait loci in common bean. Crop Sci 55:1969–1977
- 224. Hallar AG, Chirokova G, McCubbin I, Painter TH, Wiedinmyer C, Dodson C (2011) Atmospheric bioaerosols transported via dust storms in the western United States. Geophys Res Lett 38:L17801. https://doi.org/10.1029/2011GL048166
- Hamer JE, Howard RJ, Chumley FG, Valent B (1988) A mechanism for surface attachment in spores of a plant phytopathogenic fungus. Science 239:288–290
- 226. Hammond-Kosack KE, Parker JE (2003) Deciphering plant-pathogen communication: fresh perspectives for molecular resistance breeding. Curr Opin Biotechnol 14:177–193. https://doi.org/10. 1016/s0958-1669(03)00035-1
- 227. Hantsch L, Braun U, Scherer-Lorenzen M, Bruelheide H (2013) Species richness and species identity effects on occurrence of foliar fungal pathogens in a tree diversity experiment. Ecosphere 4:81. https://doi.org/10.1890/ES13-00103.1
- Hara K, Zhang D (2012) Bacterial abundance and viability in long-range transported dust. Atmos Environ 47:20–25. https:// doi.org/10.1016/j.atmosenv.2011.11.050
- 229. Hardham AR (2001) The cell biology behind Phytophthora pathogenicity. Aust Plant Pathol 30:91–98
- Harrison RM, Jones AM, Biggins PDE et al (2005) Climate factors influencing bacterial count in background air samples. Int J Biometeorol 49:167–178. https://doi.org/10.1007/ s00484-004-0225-3
- Haseloff J, Mohamed N, Symons R (1982) Viroid RNAs of cadang-cadang disease of coconuts. Nature 299:316–321. https:// doi.org/10.1038/299316a0
- 232. Hayashi K, Yoshida T, Hayano-Saito Y (2019) Detection of white head symptoms of panicle blast caused by Pyricularia oryzae using cut-flower dye. Plant Methods 15:159. https://doi.org/10. 1186/s13007-019-0548-z
- 233. Hayes RJ, Trent MA, Truco MJ et al (2014) The inheritance of resistance to bacterial leaf spot of lettuce caused by Xanthomonas campestris pv. vitians in three lettuce cultivars. Hortic Res 1:14066. https://doi.org/10.1038/hortres.2014.66
- 234. He YW, Wu J, Cha J-S, Zhang L-H (2010) Rice bacterial blight pathogen Xanthomonas oryzae pv. oryzae produces multiple DSF-family signals in regulation of virulence factor production. BMC Microbiol 10:187. https://doi.org/10.1186/ 1471-2180-10-187
- Heald CL, Spracklen DV (2009) Atmospheric budget of primary biological aerosol particles from fungal spores. Geophys Res Lett 38:L09806. https://doi.org/10.1029/2009GL037493
- Hewezi T, Baum TJ (2013) Manipulation of plant cells by cyst and root-knot nematode effectors. Mol Plant-Microbe Interact 26:9–16. https://doi.org/10.1094/MPMI-05-12-0106-FI
- 237. Hibberd J, Whitbread R, Farrar J (1996) Effect of 700 μmol mol-1 CO2 and infection by powdery mildew on the growth and carbon partitioning of barley. New Phytol 134:309–315

- 238. Hibberd JM, Whitbread R, Farrar JF (1996) Effect of elevated concentrations of CO2 in infection of barley by Erysiphe graminis. Physiol Mol Plant Pathol 48:37–53
- Hoat TX, Quan MV, Anh DTL et al (2015) Phytoplasma diseases on major crops in Vietnam. Phytopathogenic Mollicutes 5:S69. https://doi.org/10.5958/2249-4677.2015.00029.8
- Hodgson R, Wall G, Randles J (1998) Specific identification of Coconut Tinangaja Viroid for differential Field diagnosis of viroids in Coconut Palm. Phytopathol 88:774–781. https://doi. org/10.1094/PHYTO.1998.88.8.774
- Hollingsworth CR, Gray FA, Koch DW, Groose R, Heald TE (2010) Distribution of Phoma sclerotioides and incidence of brown root rot of alfalfa in Wyoming, U.S.A. Canadian J Plant Pathol 215–217. https://doi.org/10.1080/07060660309507071
- Hoose C, Kristjánsson JE, Burrows SM (2010) How important is biological ice nucleation in clouds on a global scale? Environ Res Lett 5:024009. https://doi.org/10.1088/1748-9326/5/2/024009
- 243. Hospodsky D, Qian J, Nazaroff WW et al (2012) Human occupancy as a source of indoor airborne bacteria. PLoS One 7:e34867. https://doi.org/10.1371/journal.pone.0034867
- 244. Hotez PJ, Bottazzi ME, Strych U et al (2015) Neglected tropical diseases among the association of Southeast Asian Nations (ASEAN): Overview and update. PLoS Negl Trop Dis 9:e0003575. https://doi.org/10.1371/journal.pntd.0003575
- Hu W, Strom N, Haarith D et al (2018) Mycobiome of cysts of the soybean cyst nematode under long term crop rotation. Front Microbiol 9. https://doi.org/10.3389/fmicb.2018.00386
- 246. Hua J (2013) Modulation of plant immunity by light, circadian rhythm, and temperature. Curr Opin Plant Biol 16:406–413. https://doi.org/10.1016/j.pbi.2013.06.017
- Huang J-S (1986) Ultrastructure of bacterial penetration in plants. Annu Rev Phytopathol 24:141–157
- Huber L, Gillespie TJ (1992) Modelling leaf wetness in relation to plant disease epidemiology. Annu Rev Phytopathol 30:553–577
- Hückelhoven R (2005) Powdery mildew susceptibility and biotrophic infection strategies. FEMS Microbiol Lett 245:9–17. https://doi.org/10.1016/j.femsle.2005.03.001
- Huerta-Espino J, Singh RP, Germán S et al (2011) Global status of wheat leaf rust caused by Puccinia triticina. Euphytica 179:143–160. https://doi.org/10.1007/s10681-011-0361-x
- 251. Huffman JA, Prenni AJ, Demott PJ et al (2013) High concentrations of biological aerosol particles and ice nuclei during and after rain. Atmos Chem Phys 13:6151–6164. https://doi.org/10. 5194/acp-13-6151-2013
- Hugh-Jones ME, Wright PB (1970) Studies on the 1967–8 footand-mouth disease epidemic. The relation of weather to the spread of disease. J Hyg (Lond) 68:253–271. https://doi.org/10. 1017/s0022172400028722
- Hull R (2014) Plant Virology, 5th edn. Academic Press, Cambridge, MA. https://doi.org/10.1016/C2010-0-64974-1
- 254. Hultin KAH, Krejci R, Pinhassi J et al (2011) Aerosol and bacterial emissions from Baltic Seawater. Atmos Res 99:1–14. https:// doi.org/10.1016/j.atmosres.2010.08.018
- 255. Hurtado L, Rodríguez G, López J et al (2014) Characterization of atmospheric bioaerosols at 9 sites in Tijuana, Mexico. Atmos Environ 96:430–436. https://doi.org/10.1016/j.atmosenv.2014. 07.018
- 256. Hurtado-Gonzales OP, Valentini G, Gilio TA et al (2017) Fine mapping of Ur-3, a historically important rust resistance locus in common bean. G3 (Bethesda) 7(2):557–569. https://doi.org/ 10.1534/g3.116.036061
- 257. Hutchison ML, Johnstone K (1993) Evidence for the involvement of the surface active properties of the extracellular toxin tolaasin in the manifestation of brown blotch disease symptoms by

Pseudomonas tolaasii on Agaricus bisporus. Physiol Mol Plant Pathol 42:373–384. https://doi.org/10.1016/S0885-5765(05) 80013-X

- 258. Inderbitzin P, Bostock RM, Davis RM et al (2011) Phylogenetics and taxonomy of the fungal vascular wilt pathogen Verticillium, with the descriptions of five new species. PLoS One 6:e28341. https://doi.org/10.1371/journal.pone.0028341
- 259. Ingold CT (1953) Dispersal in Fungi. Oxford University Press, Oxford
- Ingold CT (1999) Active liberation of reproductive units in terrestrial fungi. Mycologist 13:113–116. https://doi.org/10.1016/ S0269-915X(99)80040-8
- 261. Islam MT, Croll D, Gladieux P et al (2016) Emergence of wheat blast in Bangladesh was caused by a South American lineage of Magnaporthe oryzae. BMC Biol 14:1–12. https://doi.org/10. 1186/s12915-016-0309-7
- 262. Islam TMD, Toyota K (2004) Suppression of bacterial wilt of tomato by Ralstonia solanacearum by incorporation of composts in soil and possible mechanism. Microbes Environ 19:53–60. https://doi.org/10.1264/jsme2.2004.53
- 263. Ivanović Ž, Perović T, Popović T et al (2017) Characterization of pseudomonas syringae pv. syringae, causal agent of citrus blast of mandarin in Montenegro. Plant Pathol J 33:21–33. https://doi. org/10.5423/PPJ.OA.08.2016.0161
- 264. Jacobson MZ, Streets DG (2009) Influence of future anthropogenic emissions on climate, natural emissions, and air quality. J Geophys Res Atmos 114:D08118. https://doi.org/10.1029/2008J D011476
- 265. Jain A, Sarsaiya S, Wu Q et al (2019) A review of plant leaf fungal diseases and its environment speciation. Bioengineered 10:409–424. https://doi.org/10.1080/21655979.2019.1649520
- 266. Ji Z, Wang C, Zhao K (2018) Rice routes of countering xanthomonas oryzae. Int J Mol Sci 19:3008. https://doi.org/10.3390/ ijms19103008
- 267. Jiang N, Yan J, Liang Y et al (2020) Resistance genes and their interactions with bacterial blight/leaf streak pathogens (Xanthomonas oryzae) in Rice (Oryza sativa L.)—an updated review. Rice 13:1–12. https://doi.org/10.1186/s12284-019-0358-y
- Jiang Y, Zhou H, Chen L et al (2018) Nematodes and microorganisms interactively stimulate soil organic carbon turnover in the macroaggregates. Front Microbiol 9:2803. https://doi.org/10. 3389/fmicb.2018.02803
- Jiehua Q, Shuai M, Yizhen D, Shiwen H, Yanjun K (2019) Ustilaginoidea virens: A fungus infects rice flower and threats world rice production. Rice Science 26:199–206. https://doi.org/10. 1016/j.rsci.2018.10.007
- 270. Jimenez-Diaz RM, Jimenez-Gasco M del M (2011) Integrated management of fusarium wilt diseases. In: Alves-Santos FM, Diez JJ (eds) Research Signpost, Control of. Research Signpost, Kerala, India, pp 177–215
- 271. Jiménez-Díaz RM, Trapero-Casas A, de la Colina JC (1989) Races of *fusarium oxysporum* F. Sp. Ciceri infecting chickpeas in Southern Spain. In: Tjamos EC, Beckman CH (eds) Vascular wilt diseases of plants. NATO ASI Series, vol 28. Springer, Berlin, Heidelberg. https://doi.org/10.1007/978-3-642-73166-2\_39
- 272. Jin X, Cao X, Wang X et al (2018) Three-dimensional architecture and biogenesis of membrane structures associated with plant virus replication. Front Plant Sci 9. https://doi.org/10.3389/fpls. 2018.00057
- 273. Jo KR, Kim CJ, Kim SJ et al (2014) Development of late blight resistant potatoes by cisgene stacking. BMC Biotechnol 14:1–10. https://doi.org/10.1186/1472-6750-14-50
- 274. Jo KR, Kim CJ, Kim SJ, Kim TY, Bergervoet M, Jongsma MA, Visser RG, Jacobsen E, Vossen JH (2014) Development of late blight resistant potatoes by cisgene stacking. BMC Biochem 14:1–10. https://doi.org/10.1186/1472-6750-14-50

- 276. Jones AM, Harrison RM (2004) The effects of meteorological factors on atmospheric bioaerosol concentrations - A review. Sci Total Environ 326:151–180. https://doi.org/10.1016/j.scitotenv. 2003.11.021
- 277. Jones DA, Thomas CM, Hammond-kosack KE et al (1994) Isolation of the Tomato Cf-9 Gene for Resistance to Cladosporium fulvum by Transposon Tagging. Science (80-) 266:789–793. https://doi.org/10.1126/science.7973631
- 278. Jones JT, Haegeman A, Danchin EGJ et al (2013) Top 10 plantparasitic nematodes in molecular plant pathology. Mol Plant Pathol 14:946–961. https://doi.org/10.1111/mpp.12057
- 279. Jones K, Kim DW, Park JS, Khang CH (2016) Live-cell fluorescence imaging to investigate the dynamics of plant cell death during infection by the rice blast fungus Magnaporthe oryzae. BMC Plant Biol 16:1–8. https://doi.org/10.1186/s12870-016-0756-x
- Joung YS, Ge Z, Buie CR (2017) Bioaerosol generation by raindrops on soil. Nat Commun 8. https://doi.org/10.1038/ncomm s14668
- Kamoun S, Furzer O, Jones JDG et al (2015) The Top 10 oomycete pathogens in molecular plant pathology. Mol Plant Pathol 16:413–434. https://doi.org/10.1111/mpp.12190
- 282. Kappagantu M, Bullock JM, Nelson ME, Eastwell KC (2017) Hop stunt viroid: Effect on host (Humulus lupulus) transcriptome and its interactions with hop powdery mildew (Podospheara macularis). Mol Plant Microbe Interact 30:842–851
- Karra S, Katsivela E (2007) Microorganisms in bioaerosol emissions from wastewater treatment plants during summer at a Mediterranean site. Water Res 41:1355–1365
- 284. Kaushik R, Balasubramanian R (2013) Discrimination of viable from non-viable Gram-negative bacterial pathogens in airborne particles using propidium monoazide-assisted qPCR. Sci Total Environ 449:237–243. https://doi.org/10.1016/j.scitotenv.2013. 01.065
- 285. Kawecki TJ, Ebert D (2004) Conceptual issues in local adaptation. Ecol Lett 7:1225–1241. https://doi.org/10.1111/j.1461-0248.2004.00684.x
- Kellogg CA, Griffin DW (2006) Aerobiology and the global transport of desert dust. Trends Ecol Evol 21:638–644. https:// doi.org/10.1016/j.tree.2006.07.004
- 287. Kembel SW, Jones E, Kline J et al (2012) Architectural design influences the diversity and structure of the built environment microbiome. ISME J 6:1469–1479. https://doi.org/10.1038/ ismej.2011.211
- 288. Keykhasaber M, Thomma BPHJ, Hiemstra JA (2018) Verticillium wilt caused by Verticillium dahliae in woody plants with emphasis on olive and shade trees. Eur J Plant Pathol 150:21–37. https://doi.org/10.1007/s10658-017-1273-y
- Khan MAI, Bhuiyna MR, Hossain MS et al (2014) Neck blast disease influences grain yield and quality traits of aromatic rice. Comptes Rendus - Biol 337:635–641. https://doi.org/10. 1016/j.crvi.2014.08.007
- 290. Khan M, Atiq M (2020) Technical Report: Prediction of citrus canker disease and its management. https://doi.org/10.13140/ RG.2.2.33830.96329/1
- 291. Kim HJ, Lee HR, Jo KR et al (2012) Broad spectrum late blight resistance in potato differential set plants MaR8 and MaR9 is conferred by multiple stacked R genes. Theor Appl Genet 124:923–935. https://doi.org/10.1007/s00122-011-1757-7
- 292. Kim S, Kim CY, Park SY et al (2020) Two nuclear effectors of the rice blast fungus modulate host immunity via transcriptional reprogramming. Nat Commun 11:5845. https://doi.org/ 10.1038/s41467-020-19624-w

- 293. Kim YJ, Lin NC, Martin GB (2002) Two distinct Pseudomonas effector proteins interact with the Pto kinase and activate plant immunity. Cell 109:589–598. https://doi.org/10.1016/S0092-8674(02)00743-2
- 294. Kirk PM, Canoon PF, Minter DW, Stalpers JA (2008) Ainsworth and Bisby's Dictionary of the Fungi, 10th edn. CABI, Wallingford. https://doi.org/10.1079/9780851998268.0000
- 295. Klosterman SJ, Atallah ZK, Vallad GE, Subbarao KV (2009) Diversity, pathogenicity, and management of verticillium species. Annu Rev Phytopathol 47:39–62. https://doi.org/10.1146/ annurev-phyto-080508-081748
- 296. Kobayashi T, Ishiguro K, Nakajima T et al (2006) Effects of elevated atmospheric CO2 concentration on the infection of rice blast and sheath blight. Phytopathology 96:425–431. https://doi.org/10.1094/PHYTO-96-0425
- 297. Köhl J, Van Tongeren CAM, Groenenboom-de Haas BH, Van Hoof RA, Driessen R, Van Der Heijden L (2010) Epidemiology of dark leaf spot caused by Alternaria brassicicola and A. brassicae in organic seed production of cauliflower. Plant Pathol 59:358–367
- Köhm BA, Goulden MG, Gilbert JE et al (1993) A potato virus X resistance gene mediates an induced, nonspecific resistance in protoplasts. Plant Cell 5:913–920. https://doi.org/10.2307/3869659
- 299. Kolmer JA, Hughes ME (2005) Physiologic specialization of Puccinia triticina on wheat in the United States in 2003. Plant Dis 89:1201–1206. https://doi.org/10.1094/ PDIS-11-17-1701-SR
- 300. Kong P, Rubio L, Polek ML, Falk BW (2000) Population structure and genetic diversity within California Citrus tristeza virus (CTV) isolates. Virus Genes 21:139–145. https://doi.org/10. 1023/A:1008198311398
- 301. Kovalev N, de Castro Martín IF, Pogany J et al (2016) Role of Viral RNA and Co-opted Cellular ESCRT-I and ESCRT-III Factors in Formation of Tombusvirus Spherules Harboring the Tombusvirus Replicase. J Virol 90:3611–3626. https://doi.org/ 10.1128/jvi.02775-15
- Kovalskaya N, Hammond RW (2014) Molecular biology of viroid-host interactions and disease control strategies. Eff Grain Boundaries Paraconductivity YBCO 228:48–60. https://doi.org/ 10.1016/j.plantsci.2014.05.006
- 303. Kreuze JF, Souza-Dias JAC, Jeevalatha A, Figueira AR, Valkonen JPT, Jones RAC (2020) Viral diseases in potato. In: Campos H, Ortiz O (eds) The potato crop. Springer, Cham
- 304. Krishnamoorthy S, Muthalagu A, Priyamvada H et al (2020) On distinguishing the natural and human-induced sources of airborne pathogenic viable bioaerosols: characteristic assessment using advanced molecular analysis. SN Appl Sci 2:1162. https:// doi.org/10.1007/s42452-020-2965-z
- Krupa S, McGrath MT, Andersen CP et al (2001) Ambient ozone and plant health. Plant Dis 85:4–12
- 306. Kumar D, Bhatt J, Sharma RL (2017) Efficacy of different bio control agents against Meloidogyne incognita and Fusarium oxysporum on Black gram (Vigna mungo L). Int J Current Microb Applied Sci 6:2287–2291
- 307. Kumar RR, Ansar M, Rajani K et al (2020) First report on molecular basis of potato leaf roll virus (PLRV) aggravation by combined effect of tuber and prevailing aphid. BMC Res Notes 13:523. https://doi.org/10.1186/s13104-020-05370-1
- 308. Kuroda K (2001) Responses of Quercus sapwood to infection with the pathogenic fungus of a new wilt disease vectored by the ambrosia beetle Platypus quercivorus. J Wood Sci 47:425–429. https://doi.org/10.1007/BF00767893
- 309. Kuske CR (2006) Current and emerging technologies for the study of bacteria in the outdoor air. Curr Opin Biotechnol 17:291–296. https://doi.org/10.1016/j.copbio.2006.04.001

- Kutsuwa K, Dickson DW, Brito JA et al (2015) Belonolaimus longicaudatus: An emerging pathogen of Peanut in Florida. J Nematol 47:87–96
- 311. Kuyek D, Biothai G et al (2000) Blast, biotech and big business: Implications of corporate strategies on Rice research in Asia. In: GRAIN. http://scholar.google.com/scholar?hl=en&btnG=Searc h&q=intitle:Blast,+biotech+and+big+business#1. Accessed 28 Sep 2022
- Lacey J (1991) Aggregation of spores and its effect on aerodynamic behaviour. Grana 30:437–445. https://doi.org/10.1080/ 00173139109432005
- Lacey J (1996) Spore dispersal Its role in ecology and disease: The British contribution to fungal aerobiology. Mycol Res 100:641–660
- Lambert KN, Bekal S (2002) Introduction to plant parasitic nematodes. Plant Health Instr 10:1094–1218
- Lamichhane JR, Dürr C, Schwanck AA et al (2017) Integrated management of damping-off diseases. A review. Agron Sustain Dev 37:1–25. https://doi.org/10.1007/s13593-017-0417-y
- 316. Lamichhane JR, Varvaro L, Parisi L et al (2014) Disease and frost damage of woody plants caused by pseudomonas syringae: Seeing the forest for the trees. Adv Agron 126:235–295. https:// doi.org/10.1016/B978-0-12-800132-5.00004-3
- 317. LaMondia JA (2006) Management of lesion nematodes and potato early dying with rotation crops. J Nematol 38:442–448
- Landeras E, Estefanía TV, Máximo B, Ana G (2017) Short communication: Occurrence of angular leaf spot caused by Pseudocercospora griseola in Phaseolus vulgaris in Asturias, Spain. Spanish J Agricul Res 15:e10SC03. https://doi.org/10.5424/sjar/ 2017153-10798
- Langley RL, Morrow WEM (2010) Livestock handling-minimizing worker injuries. J Agromedicine 15:226–235. https://doi.org/ 10.1080/1059924X.2010.486327
- 320. Laundon JR (1985) *Desmococcus Olivaceus*—the name of the common subaerial green alga. Taxon 34:671–672
- 321. Lazazzara V, Bueschl C, Parich A, Pertot I, Schuhmacher R, Perazzolli M (2018) Downy mildew symptoms on grapevines can be reduced by volatile organic compounds of resistant genotypes. Sci Rep 8:1618. https://doi.org/10.1038/s41598-018-19776-2
- Leben C (1969) Colonization of soybean buds by bacteria: observations with the scanning electron microscope. Can J Microbiol 15:319–321
- 323. Leck C, Bigg EK (2005) Biogenic particles in the surface microlayer and overlaying atmosphere in the central Arctic Ocean during summer. Tellus B Chem Phys Meteorol 57:305–316. https:// doi.org/10.3402/tellusb.v57i4.16546
- 324. Lee C, Sultana CM, Collins DB et al (2015) Advancing model systems for fundamental laboratory studies of sea spray aerosol using the microbial loop. J Phys Chem A 119:8860–8870. https:// doi.org/10.1021/acs.jpca.5b03488
- 325. Lee SW, Han M, Park CJ, Seo YS, Jeon JS (2011) The molecular mechanisms of rice resistance to the bacterial blight pathogen, Xanthomonas oryzae pathovar oryzae. In: Kader DM (ed) Advances in botanical research, vol 60. Academic Press, San Diego, pp 51–87
- 326. Leonard KJ, Szabo LJ (2005) Stem rust of small grains and grasses caused by Puccinia graminis. Mol Plant Pathol 6:99–111. https://doi.org/10.1111/j.1364-3703.2005.00273.x
- 327. Lestari S, Hidayat S (2020) IOP Conference Series: Earth and Environmental Science Survey and detection of Banana bunchy top virus in Java Survey and detection of Banana bunchy top virus in Java. IOP Conf Ser Earth Environ Sci 583. https://doi. org/10.1088/1755-1315/583/1/012022
- Lewis DH, Smith DC (1967) Sugar alcohols (Polyols) in fungi and green plants. I. Distribution physiology and metabolism. New Phytologist 66:143–184

- 329. Li C, Yaegashi H, Kishigami R et al (2020) Apple russet ring and apple green crinkle diseases: Fulfillment of Koch's postulates by virome analysis, amplification of full-length cDNA of viral genomes, in vitro transcription of infectious Viral RNAs, and reproduction of symptoms on fruits of apple T. Front Microbiol 11:1627. https://doi.org/10.3389/fmicb.2020.01627
- 330. Li HL, Yuan HX, Fu B et al (2012) First report of Fusarium pseudograminearum causing Crown Rot of Wheat in Henan. China Plant Dis 96:1065. https://doi.org/10.1094/ PDIS-01-12-0007-PDN
- 331. Li J, Pang Z, Duan S et al (2019) The in planta effective concentration of oxytetracycline against Candidatus Liberibacter asiaticus for suppression of citrus Huanglongbing. Phytopathol 109:2046–2054
- 332. Liberti D, Marais A, Svanella-Dumas L et al (2005) Characterization of Apricot pseudo-chlorotic leaf spot virus, a novel Trichovirus isolated from stone fruit trees. Phytopathol 95:420–426
- 333. Lighthart B (1984) Microbial aerosols: Estimated contribution of combine harvesting to an airshed. Appl Environ Microbiol 47:430–432. https://doi.org/10.1128/aem.47.2.430-432.1984
- Lighthart B (1997) The ecology of bacteria in the alfresco atmosphere. FEMS Microbiol Ecol 23:263–274
- Lin WH, Li CS (2000) Associations of fungal aerosols, air pollutants, and meteorological factors. Aerosol Sci Technol 32:359– 368. https://doi.org/10.1080/027868200303678
- 336. Lindow SE (2002) Differential survival of solitary and aggregated cells of Pseudomonas syringae on leaves. Phytopathol 92:S97
- 337. Lindow SE, Brandl MT (2003) Microbiology of the phyllosphere. Appl Environ Microbiol 69:1875–1883. https://doi.org/10.1128/ AEM.69.4.1875-1883.2003
- 338. Liu C, Nelson RS (2013) The cell biology of tobacco mosaic virus replication and movement. Front Plant Sci 4:1–11. https:// doi.org/10.3389/fpls.2013.00012
- Liu M, Hambleton S (2010) Taxonomic study of stripe rust, Puccinia striiformis sensu lato, based on molecular and morphological evidence. Fungal Biol 114:881–899. https://doi.org/10.1016/j. funbio.2010.08.005
- 340. Liu W, Liu J, Triplett L et al (2014) Novel insights into rice innate immunity against bacterial and fungal pathogens. Annu Rev Phytopathol 52:213–241. https://doi.org/10.1146/annur ev-phyto-102313-045926
- Long SP, Zhu XG, Naidu SL, Ort DR (2006) Can improvement in photosynthesis increase crop yields? Plant, Cell Environ 29:315– 330. https://doi.org/10.1111/j.1365-3040.2005.01493.x
- Lopez-Escudero FJ, Blanco-Lopez MA (2001) Effect of a single or double soil solarization to control Verticillium wilt in established olive orchards in Spain. Plant Dis 85:489–496. https://doi. org/10.1094/PDIS.2001.85.5.489
- Lough TJ, Lucas WJ (2006) Integrative plant biology: role of phloem long-distance macromolecular trafficking. Annu Rev Plant Biol 57:203–232. https://doi.org/10.1146/annurev.arplant. 56.032604.144145
- 344. Lu G, Wang Z, Xu F et al (2021) Sugarcane mosaic disease: Characteristics, identification and control. Microorganisms 9:1984. https://doi.org/10.3390/microorganisms9091984
- 345. Lucas GB, Campbell CL, Lucas LT (1992) Diseases caused by airborne fungi. In: Introduction to Plant Diseases. Springer, US, Boston, MA, pp 586–594. https://doi.org/10.1007/ 978-1-4615-7294-7
- 346. Lucas WJ (2006) Plant viral movement proteins: Agents for cell-to-cell trafficking of viral genomes. Virology 344:169–184. https://doi.org/10.1016/j.virol.2005.09.026
- 347. Luigi M, Manglli A, Tomassoli L, Faggioli F (2013) First report of Hop stunt viroid in Hibiscus rosa-sinensis in Italy. New Dis Reports 27:14–14. https://doi.org/10.5197/j.2044-0588.2013. 027.014

- 348. Mackintosh CA, Lidwell OM, Towers AG, Marples RR (1978) The dimensions of skin fragments dispersed into the air during activity. J Hyg (Lond) 81:471–480. https://doi.org/10.1017/ S0022172400025341
- Maddahian M, Massumi H, Heydarnejad J et al (2019) Biological and molecular characterization of hop stunt viroid variants from pistachio trees in Iran. J Phytopathol 167:163–173. https://doi. org/10.1111/jph.12783
- 350. Madej T, Janowicz K, Błaszkowski J (2000) The diseases of ornamental plants caused by Aphelenchoides ritzemabosi in association with fungi. Arch Phytopathol Plant Prot 33:141–148. https://doi.org/10.1080/03235400009383338
- Madelin TM (1994) Fungal aerosols: A review. J Aerosol Sci 25:1405–1412. https://doi.org/10.1016/0021-8502(94)90216-X
- 352. Maekawa T, Kracher B, Vernaldi S et al (2012) Conservation of NLR-triggered immunity across plant lineages. Proc Natl Acad Sci U S A 109:20119–20123. https://doi.org/10.1073/pnas.12180 59109
- 353. Maki T, Hara K, Iwata A et al (2017) Variations in airborne bacterial communities at high altitudes over the Noto Peninsula (Japan) in response to Asian dust events. Atmos Chem Phys 17:11877–11897. https://doi.org/10.5194/acp-17-11877-2017
- 354. Maki T, Kakikawa M, Kobayashi F et al (2013) Assessment of composition and origin of airborne bacteria in the free troposphere over Japan. Atmos Environ 74:73–82. https://doi.org/10. 1016/j.atmosenv.2013.03.029
- 355. Maki T, Lee KC, Kawai K et al (2019) Aeolian dispersal of bacteria associated with desert dust and anthropogenic particles over continental and oceanic surfaces. J Geophys Res Atmos 124:5579–5588. https://doi.org/10.1029/2018JD029597
- 356. Maki T, Susuki S, Kobayashi F et al (2010) Phylogenetic analysis of atmospheric halotolerant bacterial communities at high altitude in an Asian dust (KOSA) arrival region, Suzu City. Sci Total Environ 408:4556–4562. https://doi.org/10.1016/j.scito tenv.2010.04.002
- 357. Maksimov IV, Sorokan AV, Burkhanova SV et al (2019) Mechanisms of plant tolerance to RNA viruses induced by plantgrowth-promoting microorganisms. Plants 8:575. https://doi. org/10.3390/plants8120575
- 358. Manching Heather C, Balint-Kurti Peter J, Stapleton Ann E (2014) Southern leaf blight disease severity is correlated with decreased maize leaf epiphytic bacterial species richness and the phyllosphere bacterial diversity decline is enhanced by nitrogen fertilization. Front Plant Sci 5. https://doi.org/10. 3389/fpls.2014.00403
- 359. Manning W, Tiedemann A (1995) Climate change: potential effects of increased atmospheric carbon dioxide (CO2), ozone (O3), and ultraviolet-B (UV-B) radiation on plant diseases. Environ Poll 88:219–245
- 360. Mansfield J, Genin S, Magori S et al (2012) Top 10 plant pathogenic bacteria in molecular plant pathology. Mol Plant Pathol 13:614–629. https://doi.org/10.1111/j.1364-3703.2012. 00804.x
- 361. Maranger R, Bird DF (1995) Viral abundance in aquatic systems: A comparison between marine and fresh waters. Mar Ecol Prog Ser 121:217–226. https://doi.org/10.3354/meps121217
- 362. Marchant WG, Gautam S, Hutton SF, Srinivasan R (2020) Tomato yellow leaf curl virus-resistant and -susceptible tomato genotypes similarly impact the virus population genetics. Front Plant Sci 11:599697. https://doi.org/10.3389/fpls.2020.599697
- Maree HJ, Almeida RPP, Bester R et al (2013) Grapevine leafroll-associated virus 3. Front Microbiol 4:82. https://doi.org/10. 3389/fmicb.2013.00082
- Marks R, Kruczalak K, Jankowska K, Michalska M (2001) Bacteria and fungi in air over the Gulf of Gdansk and Baltic sea. J Aerosol Sci 32:237–250

- 365. Marquez-molins J, Gomez G, Pallas V (2020) Hop stunt viroid : A polyphagous pathogenic RNA that has shed light on viroid – host interactions. Mol Plant Patholo 22:153–162. https://doi.org/ 10.1111/mpp.13022
- Marshall WA, Chalmers MO (1997) Airborne dispersal of antarctic terrestrial algae and cyanobacteria. Ecography (Cop) 20:585–594. https://doi.org/10.1111/j.1600-0587.1997.tb00427.x
- 367. Martelli GP, Abou Ghanem-Sabanadzovic N, Agranovsky AA et al (2012) Taxonomic revision of the family closteroviridae with special reference to the grapevine leafroll-associated members of the genus ampelovirus and the putative species unassigned to the family. J Plant Pathol 94:7–19
- 368. Martin GB, Brommonschenkel SH, Chunwongse J et al (1993) Map-based cloning of a protein kinase gene conferring disease resistance in tomato. Science (80-) 262:1432–1436
- Martínez-arias C, Sobrino-plata J, Gil L et al (2021) Priming of plant defenses against ophiostoma novo-ulmi by elm (Ulmus minor mill.) fungal endophytes. J Fungi 7:687. https://doi.org/ 10.3390/jof7090687
- 370. Martinez-Soriano JP, Galindo-Alonso J, Maroon CJM et al (1996) Mexican papita viroid: Putative ancestor of crop viroids. Proc Natl Acad Sci 93:9397-9401
- 371. Masurkar P, Bajpai R, Sahu V et al (2018) Invasion and nutrient acquisition strategies of phytopathogens: Fungi, bacteria and viruses. Int J Curr Microbiol Appl Sci 7:3132–3146. https://doi. org/10.20546/ijcmas.2018.708.335
- 372. Matsushita Y, Yanagisawa H (2018) Distribution of Tomato planta macho viroid in germinating pollen and transmitting tract. Virus Genes 54:124–129. https://doi.org/10.1007/ s11262-017-1510-7
- 373. Matthias-Maser S, Peters K, Jaenicke R (1995) Seasonal variation of primary biological aerosol particles. J Aerosol Sci 26:S545–S546. https://doi.org/10.1016/0021-8502(95)97180-M
- 374. May NW, Gunsch MJ, Olson NE et al (2018) Unexpected contributions of sea spray and lake spray aerosol to inland particulate matter. Environ Sci Technol Lett 5:405–412. https://doi. org/10.1021/acs.estlett.8b00254
- 375. May NW, Olson NE, Panas M et al (2018) Aerosol emissions from great lakes harmful algal blooms. Environ Sci Technol 52:397–405. https://doi.org/10.1021/acs.est.7b03609
- 376. Mayol E, Jiménez MA, Herndl GJ et al (2014) Resolving the abundance and air- sea fluxes of airborne microorganisms in the North Atlantic Ocean. Front Microbiol 5:557. https://doi. org/10.3389/fmicb.2014.00557
- 377. Mburu H, Cortada L, Haukeland S et al (2020) Potato cyst nematodes: A new threat to potato production in East Africa. Front Plant Sci 11:670. https://doi.org/10.3389/fpls.2020. 00670
- McCarthy M (2001) Dust clouds implicated in spread of infection. Lancet 358:478. https://doi.org/10.1016/S0140-6736(01) 05677-X
- 379. Mehetre GT, Leo VV, Singh G et al (2021) Current developments and challenges in plant viral diagnostics: A systematic review. Viruses 13:412. https://doi.org/10.3390/v13030412
- 380. Meinhardt LW, Costa GG, Thomazella DP et al (2014) Genome and secretome analysis of the hemibiotrophic fungal pathogen, Moniliophthora roreri, which causes frosty pod rot disease of cacao: mechanisms of the biotrophic and necrotrophic phases. BMC Genomics 27:164. https://doi.org/10.1186/ 1471-2164-15-164
- Mejias J, Truong NM, Abad P et al (2019) Plant proteins and processes targeted by parasitic nematode effectors. Front Plant Sci 10:970. https://doi.org/10.3389/fpls.2019.00970
- 382. Melloy P, Hollaway G, Luck J et al (2010) Production and fitness of Fusarium pseudograminearum inoculum at elevated carbon dioxide in FACE. Global Change Biology – Glob

Change Biol 16. https://doi.org/10.1111/j.1365-2486.2010. 02178.x

- Melotto M, Zhang L, Oblessuc PR, He SY (2017) Stomatal defense a decade later. Plant Physiol 174:561–571. https://doi. org/10.1104/pp.16.01853
- Mendgen K, Dressler E (1983) Culturing Puccinia coronata on a Cell Monolayer of the Avena sativa Coleoptile. J Phytopathol 108:226–234. https://doi.org/10.1111/j.1439-0434.1983.tb00583.x
- 385. Meng S, Torto-Alalibo T, Chibucos MC et al (2009) Common processes in pathogenesis by fungal and oomycete plant pathogens, described with Gene Ontology terms. BMC Microbiol 9:S7. https://doi.org/10.1186/1471-2180-9-S1-S7
- Menzies JG, Turkington TK, Knox RE (2009) Testing for resistance to smut diseases of barley, oats and wheat in western Canada. Can J Plant Pathol 31:265–279. https://doi.org/10.1080/ 07060660909507601
- Mieslerova B, Kitner M, Křístková E, Majeský Ľ, Lebeda A (2020) Powdery Mildews on Lactuca species – A complex view of host-pathogen interactions. Crit Rev Plant Sci 39:1–28. https:// doi.org/10.1080/07352689.2020.1752439
- Milgroom MG, Wang K, Lipari SE, Kaneko S (1996) Intercontinental population structure of the chestnut blight fungus, Cryphonectria parasitica. Mycol Soc Am 88:179–190. https://doi.org/ 10.1080/00275514.1942.12020904
- Min YY, Toyota K (2019) Occurrence of different kinds of diseases in sesame cultivation in Myanmar and their impact to sesame yield. J Exp Agric Int 38:1–9. https://doi.org/10.9734/ jeai/2019/v38i430309
- 390. Mindrinos M, Katagiri F, Yu GL, Ausubel FM (1994) The A. thaliana disease resistance gene RPS2 encodes a protein containing a nucleotide-binding site and leucine-rich repeats. Cell 78:1089–1099. https://doi.org/10.1016/0092-8674(94)90282-8
- 391. Miquel P (1883) Les Organismes Vivants de l'atmosphere. Gauthier-Villars, Paris
- 392. Mmbaga MT, Steadman JR, Stavely JR (1996) The use of host resistance in disease management of rust in common bean. Integr Pest Manag Rev 1:191–200. https://doi.org/10.1007/Bf00139763
- Moffett BF, Getti G, Henderson-Begg SK, Hill TCJ (2015) Ubiquity of ice nucleation in lichen' possible atmospheric implications. Lindbergia 38:39–43. https://doi.org/10.25227/linbg.01070
- 394. Mokrini F, Viaene N, Waeyenberge L et al (2018) Investigation of resistance to Pratylenchus penetrans and P. thornei in international wheat lines and its durability when inoculated together with the cereal cyst nematode Heterodera avenae, using qPCR for nematode quantification. Eur J Plant Pathol 151:875–889. https:// doi.org/10.1007/s10658-018-1420-0
- 395. Moletta M, Delgenes JP, Godon JJ (2007) Differences in the aerosolization behavior of microorganisms as revealed through their transport by biogas. Sci Total Environ 379:75–88
- 396. Monier JM, Lindow SE (2003) Differential survival of solitary and aggregated bacterial cells promotes aggregate formation on leaf surfaces. Proc Natl Acad Sci U S A 100:15977–15982. https://doi.org/10.1073/pnas.2436560100
- 397. Monier JM, Lindow SE (2004) Frequency, size, and localization of bacterial aggregates on bean leaf surfaces. Appl Environ Microbiol 70:346–355. https://doi.org/10.1128/AEM.70.1.346-355.2004
- 398. Montiflor MO, Vellema S, Digal LN (2019) Coordination as management response to the spread of a global plant disease: a case study in a major Philippine banana production area. Front Plant Sci 10:1048. https://doi.org/10.3389/fpls.2019.01048
- 399. Moriones E, Navas-Castillo J (2000) Tomato yellow leaf curl virus, an emerging virus complex causing epidemics worldwide. Virus Res 71:123–134. https://doi.org/10.1016/S0168-1702(00) 00193-3

- 400. Morris CE, Conen F, Alex Huffman J et al (2014) Bioprecipitation: A feedback cycle linking Earth history, ecosystem dynamics and land use through biological ice nucleators in the atmosphere. Glob Chang Biol 20:341–351. https://doi.org/10.1111/gcb.12447
- 401. Morris CE, Kinkel LL (2002) Fifty years of phyllosphere microbiology: significant contributions to research in related fields. In: Lindow SE, Hecht-Poinar EI, Elliott VJ (eds) Phyllosphere Microbiology. APS Press, Saint Paul, USA, pp 365–375
- 402. Morris CE, Leyronas C, Nicot PC (2014) Movement of Bioaerosols in the Atmosphere and the Consequences for Climate and Microbial Evolution. In: Colbeck I, Mihalis L (eds) Aerosol Science: Technology and Applications. John Wiley & Sons, Hoboken, NJ, pp 393–416
- 403. Morris CE, Sands DC, Bardin M et al (2011) Microbiology and atmospheric processes: Research challenges concerning the impact of airborne micro-organisms on the atmosphere and climate. Biogeosciences 8:17–25. https://doi.org/10.5194/ bg-8-17-2011
- 404. Morris CE, Sands DC, Vinatzer BA et al (2008) The life history of the plant pathogen Pseudomonas syringae is linked to the water cycle. ISME J 2:321–334. https://doi.org/10.1038/ismej. 2007.113
- 405. Mustilli A-C, Merlot S, Vavasseur A et al (2002) Arabidopsis OST1 protein kinase mediates the regulation of stomatal aperture by abscisic acid and acts upstream of reactive oxygen species production. Am Soc Plant Biol 14:3089–3099. https://doi.org/ 10.1105/tpc.007906
- 406. Nabeshima T, Doi M, Hosokawa M (2017) Comparative analysis of Chrysanthemum stunt viroid accumulation and movement in two Chrysanthemum (Chrysanthemum morifolium) cultivars with differential susceptibility to the viroid infection. Front Plant Sci 8. https://doi.org/10.3389/fpls.2017.01940
- 407. Nag R, Monahan C, Whyte P et al (2021) Risk assessment of Escherichia coli in bioaerosols generated following land application of farmyard slurry. Sci Total Environ 791:148189. https:// doi.org/10.1016/j.scitotenv.2021.148189
- 408. Nam M, Park SJ, Kim YJ et al (2012) First report of peanut stunt virus on glycine max in Korea. Plant Pathol J 28:330. https://doi. org/10.5423/PPJ.DR.07.2011.0138
- 409. Naqvi SAH (2019) Bacterial leaf blight of rice: An overview of epidemiology and management with special reference to-indiansub-continent. Pakistan J Agric Res 32:359–380. https://doi.org/ 10.17582/journal.pjar/2019/32.2.359.380
- 410. Nathan R, Schurr FM, Spiegel O et al (2008) Mechanisms of long-distance seed dispersal. Trends Ecol Evol 23:638–647. https://doi.org/10.1016/j.tree.2008.08.003
- 411. Nault LR, Ammar ED (1989) Leafhopper and planthopper transmission of plant viruses. Annu Rev Entomol 34:503–529. https:// doi.org/10.1146/annurev.en.34.010189.002443
- 412. Nazareno ES, Li F, Smith M et al (2018) Puccinia coronata f. sp. avenae: a threat to global oat production. Mol Plant Pathol 19:1047–1060. https://doi.org/10.1111/mpp.12608
- 413. Neff JC, Ballantyne AP, Farmer GL et al (2008) Increasing eolian dust deposition in the western United States linked to human activity. Nat Geosci 1:189–195. https://doi.org/10.1038/ngeo133
- 414. Neilson RP, Pitelka LF, Solomon AM et al (2005) Forecasting regional to global plant migration in response to climate change. Bioscience 55:749–759. https://doi.org/10.1641/0006-3568(2005)055[0749:FRTGPM]2.0.CO;2
- 415. Nelson A (2017) Crop-health survey aims to fill data gaps. Nature 541:464. https://doi.org/10.1038/541464a
- 416. Neustupa J, Škaloud P (2010) Diversity of subaerial algae and cyanobacteria growing on bark and wood in the lowland tropical forests of singapore. Plant Ecol Evol 143:51–62. https://doi.org/ 10.5091/plecevo.2010.417

- 417. Nicas M, Nazaroff WW, Hubbard A (2005) Toward understanding the risk of secondary airborne infection: Emission of respirable pathogens. J Occup Environ Hyg 2:143–154. https://doi.org/ 10.1080/15459620590918466
- 418. Nicol JM, Rivoal R (2007) Global knowledge and its application for the integrated control and management of nematodes on wheat. In: Ciancio A, Mukerji KG (eds) Integrated management and biocontrol of vegetable and grain crop nematodes. Springer, Dordrecht, The Netherlands, pp 251–294. https://doi.org/10. 1007/978-1-4020-6063-2
- 419. Nicol JM, Turner SJ, Coyne DL et al (2011) Current nematode threats to world agriculture. In Genomics and Molecular Genetics of Plant-Nematode Interactions; Springer: Dordrecht, The Netherlands, 21–44
- Niño-Liu DO, Ronald PC, Bogdanove AJ (2006) Xanthomonas oryzae pathovars: Model pathogens of a model crop. Mol Plant Pathol 7:303–324. https://doi.org/10.1111/j.1364-3703.2006.00344.x
- 421. Noble WC, Habbema JDF, Van Furth R, Smith I, De Raay C (1976) Quantitative studies on the dispersal of skin bacteria into the air. J Med Microbiol 9:53–61. https://doi.org/10.1099/00222 615-9-1-53
- 422. Nowicki M, Kozik EU, Foolad MR (2013) Late blight of tomato. In: Varshney RK, Tuberosa R (eds).Translational Genomics for Crop Breeding. John Wiley & Sons Ltd, pp 241–65
- Nyaku ST, Affokpon A, Danquah A, Brentu FC (2017) Harnessing useful rhizosphere microorganisms for nematode control. Nematol - Concepts, Diagnosis Control 153. https://doi.org/10. 5772/intechopen.69164
- 424. Nzioki H, Jamoza J, Olweny C, Rono J (2010) Characterization of physiologic races of sugarcane smut (Ustilago scitaminea) in Kenya. African J Microbiol Res 4:1694–1697
- 425. O'dowd C, Ceburnis D, Ovadnevaite J et al (2015) Connecting marine productivity to sea-spray via nanoscale biological processes: Phytoplankton dance or death disco? Sci Rep 5:14883. https://doi.org/10.1038/srep14883
- 426. Obidiegwu JE, Flath K, Gebhardt C (2014) Managing potato wart: A review of present research status and future perspective. Theor Appl Genet 127:763–780. https://doi.org/10.1007/ s00122-014-2268-0
- 427. Oerke EC (2006) Crop losses to pests. J Agr Sci 144:31-43
- Oerke EC, Dehne HW (2004) Safeguarding production Losses in major crops and the role of crop protection. Crop Prot 23:275– 285. https://doi.org/10.1016/j.cropro.2003.10.001
- Oka Y, Chet I, Spiegel Y (1993) Control of the rootknot nematode Meloidogyne javanica by Bacillus cereus. Biocontrol Sci Technol 3:115–126. https://doi.org/10.1080/095831593093552 67
- 430. Olaya Escobar DR, Perez Rojas FA (2006) Caracterización cualitativa - cuantitativa de Bioaerosoles relacionados con factores meteorológicos y material particulado en Puente Aranda. Universidada de La Salle, Bogotá D.C
- 431. Ona I, Cruz CV, Nelson RJ, Leach JE, Mew TW (1998) Epidemic development of bacterial blight on rice carrying resistance genes Xa-4, Xa-7, and Xa-10. Plant Dis 82:1337–1340. https://doi.org/ 10.1094/PDIS.1998.82.12.1337
- 432. Onyile AB, Edwards HH, Gessner R V (1982) Adhesive material of the hyphopodia of Buergenerula spartinae. 74:777–784
- 433. Orlovskis Z, Canale MC, Thole V et al (2015) Insect-borne plant pathogenic bacteria: Getting a ride goes beyond physical contact. Curr Opin Insect Sci 9:16–23. https://doi.org/10.1016/j.cois. 2015.04.007
- 434. Orsini M, Laurenti P, Boninti F et al (2002) A molecular typing approach for evaluating bioaerosol exposure in wastewater treatment plant workers. Water Res 36:1375–1378. https://doi.org/10. 1016/S0043-1354(01)00336-0

- 435. Osdaghi E, Jones JB, Sharma A, Goss EM, Abrahamian P, Newberry EA, Potnis N, Carvalho R, Choudhary M, Paret ML, Timilsina S, Vallad GE (2021) A centenary for bacterial spot of tomato and pepper. Mol Plant Pathol 22:1500. https://doi.org/10. 1111/mpp.13125
- 436. Ou SH (1985) Rice disease, 2nd edn. Commonwealth Mycology Institute, Kew, Surrey, UK
- 437. Owens RA (2007) Potato spindle tuber viroid: The simplicity paradox resolved? Mol Plant Pathol 8:549–560. https://doi.org/ 10.1111/j.1364-3703.2007.00418.x
- 438. Ozores-hampton M, Mcavoy E, Sargent S, Roberts P (2010) Evaluation of tomato yellow leaf curl virus (tylcv) resistant and Fusarium crown rot (fcr) resistant tomato variety under commercial conditions in southwest Florida. Tomato Inst Proc 53:11–15
- 439. Pahari AK, Dasgupta D, Patil RS, Mukherji S (2016) Emission of bacterial bioaerosols from a composting facility in Maharashtra, India. Waste Manag 53:22–31. https://doi.org/10.1016/j.wasman. 2016.04.027
- 440. Palomares-Rius JE, Escobar C, Cabrera J et al (2017) Anatomical alterations in plant tissues induced by plant-parasitic nematodes. Front Plant Sci 8:1987. https://doi.org/10.3389/fpls.2017.01987
- 441. Panchal S, Melotto M (2017) Stomate-based defense and environmental cues. Plant Signal Behav 12:e1362517. https://doi.org/ 10.1080/15592324.2017.1362517
- 442. Pappu SS, Pappu HR, Langston DB et al (2000) Outbreak of Tomato yellow leaf curl virus (Family Geminiviridae) in Georgia. Plant Heal Prog 1:34. https://doi.org/10.1094/ php-2000-0601-02-hn
- 443. Pasev G, Radeva-Ivanova V, Manoussopoulos Y et al (2018) First report of Peanut stunt virus on beans in Bulgaria. New Dis Reports 38:9–9. https://doi.org/10.5197/j.2044-0588.2018.038.009
- 444. Pathi KM, Rink P, Budhagatapalli N et al (2020) Engineering Smut resistance in maize by site-directed mutagenesis of LIPOXYGENASE 3. Front Plant Sci 11:543895. https://doi.org/ 10.3389/fpls.2020.543895
- 445. Patzak J, Henychová A, Krofta K, Svoboda P, Malí rová I (2021) The influence of Hop Latent Viroid (HLVd) infection on gene expression and secondary metabolite contents in Hop (Humulus lupulus L.) Glandular Trichomes. Plants 10:2297. https://doi.org/ 10.3390/plants10112297
- 446. Pazarlar S, Gümüs M, Öztekin GB (2013) The effects of tobacco mosaic virus infection on growth and physiological parameters in some pepper varieties (Capsicum annuum L.). Not Bot Horti Agrobot Cluj-Napoca 41:427–433. https://doi.org/10.15835/nbha4129008
- 447. Pearce D, Bridge P, Hawksworth D (2001) Species concept in Sarocladium, the causal agent of sheath rot in Rice and Bamboo blight. https://doi.org/10.1007/978-94-017-2157-8\_20
- 448. Peccia J, Hernandez M (2006) Incorporating polymerase chain reaction-based identification, population characterization, and quantification of microorganisms into aerosol science: a review. Atmos Environ 40:3941–3961. https://doi.org/10.1016/j.atmos env.2006.02.029
- 449. Pedigo LP, Rice ME (2006) Entomology and pest management. 5th ed. Pearson Prentice Hall. Columbus, OH
- Pegg GF, Brady BL (2002) Verticillium Wilts. CABI Publishing, Wallingford, UK
- 451. Peil A, Bus V, Geider K et al (2009) Improvement of fire blight resistance in apple and pear. Int J Plant Breeding 3:1–27
- 452. Perelló AE, Sisterna MN (2006) Leaf blight of wheat caused by Alternaria triticina in Argentina. Plant Pathol 55:303–303
- 453. Perfect SE, Green JR (2001) Infection structures of biotrophic and hemibiotrophic fungal plant pathogens. Mol Plant Pathol 2:101–108. https://doi.org/10.1046/j.1364-3703.2001.00055.x

- 454. Perilla-Henao LM, Casteel CL (2016) Vector-borne bacterial plant pathogens: Interactions with hemipteran insects and plants. Front Plant Sci 7:1163. https://doi.org/10.3389/fpls.2016.01163
- 455. Pillai SD, Widmer KW, Dowd SE, Ricke SC (1996) Occurrence of airborne bacteria and pathogen indicators during land application of sewage sludge. Appl Environ Microbiol 62:296–299. https://doi.org/10.1128/aem.62.1.296-299.1996
- 456. Piquerez SJM, Harvey SE, Beynon JL, Ntoukakis V (2014) Improving crop disease resistance: Lessons from research on Arabidopsis and tomato. Front Plant Sci 5:671. https://doi.org/ 10.3389/fpls.2014.00671
- 457. Pitt TL, Barer MR (2012) Classification, identification and typing of micro-organisms. Med Microbiol Eighteenth Ed 24–38. https://doi.org/10.1016/B978-0-7020-4089-4.00018-4
- 458. Pöhlker C, Wiedemann KT, Sinha B et al (2012) Biogenic potassium salt particles as seeds for secondary organic aerosol in the Amazon. Science (80-) 337:1075–1078. https://doi.org/10.1126/ science.1223264
- Pokhrel B (2020) Review on post-harvest handling to reduce loss of fruits and vegetables. Int J Hortic Food Sci Table 2:48–52
- 460. Polymenakou PN, Mandalakis M, Stephanou EG, Tselepides A (2008) Particle size distribution of airborne microorganisms and pathogens during an intense African dust event in the eastern Mediterranean. Environ Health Perspect 116:292–296. https:// doi.org/10.1289/ehp.10684
- 461. Posada JA, Redrow J, Celik I (2010) A mathematical model for predicting the viability of airborne viruses. J Virol Methods 164:88–95. https://doi.org/10.1016/j.jviromet.2009.12.004
- 462. Pöschl U, Martin ST, Sinha B et al (2010) Rainforest aerosols as biogenic nuclei of clouds and precipitation in the Amazon. Science 329:1513–1516. https://doi.org/10.1126/science.1191056
- 463. Pósfai M, Li J, Anderson JR, Buseck PR (2003) Aerosol bacteria over the Southern Ocean during ACE-1. Atmos Res 66:231–240. https://doi.org/10.1016/S0169-8095(03)00039-5
- 464. Prank M, Kenaley SC, Bergstrom GC et al (2019) Climate change impacts the spread potential of wheat stem rust, a significant crop disease. Environ Res Lett 14:124053. https://doi. org/10.1088/1748-9326/ab57de
- 465. Prasch CM, Sonnewald U (2013) Simultaneous application of heat, drought, and virus to Arabidopsis plants reveals significant shifts in signaling networks. Plant Physiol 162:1849– 1866. https://doi.org/10.1104/pp.113.221044
- 466. Prenni AJ, Tobo Y, Garcia E et al (2013) The impact of rain on ice nuclei populations at a forested site in Colorado. Geophys Res Lett 40:227–231. https://doi.org/10.1029/2012GL053953
- 467. Pringle A, Patek SN, Fischer M, Stolze J, Money NP (2005) The captured launch of a ballistospore. Mycologia 97:866–871. https://doi.org/10.3852/mycologia.97.4.866
- 468. Printz H (1921) Subaerial algae from south africa. Det Kongelige Norske Videnskabers Selskabs Skrifter
- 469. Prospero JM, Blades E, Mathison G, Naidu R (2005) Interhemispheric transport of viable fungi and bacteria from Africa to the Caribbean with soil dust. Aerobiologia (Bologna) 21:1–19. https://doi.org/10.1007/s10453-004-5872-7
- 470. Pummer BG, Budke C, Augustin-Bauditz S et al (2015) Ice nucleation by water-soluble macromolecules. Atmos Chem Phys 15:4077–4091. https://doi.org/10.5194/acp-15-4077-2015
- 471. Purayannur S, Cano LM, Bowman MJ et al (2021) The effector repertoire of the hop downy mildew pathogen Pseudoperonospora humuli. Front Genet 11:910. https://doi.org/10.3389/ fgene.2020.00910
- 472. Purcell AH (1982) Insect vector relationships with prokaryotic plant pathogens. Annu Rev Phytopathol 20:397–417. https:// doi.org/10.1146/annurev.py.20.090182.002145

- 473. Purcell AH, Hopkins DL (1996) Fastidious xylem-limited bacterial plant pathogens. Annu Rev Phytopathol 34:131–151. https://doi.org/10.1146/annurev.phyto.34.1.131
- 474. Qiu W, Feechan A, Dry I (2015) Current understanding of grapevine defense mechanisms against the biotrophic fungus (Erysiphe necator), the causal agent of powdery mildew disease. Hortic Res 2:15020. https://doi.org/10.1038/hortres.2015.20
- 475. Rajput MA, Rajput NA, Syed RN et al (2021) Sugarcane smut: Current knowledge and the way forward for management. J Fungi 7:1095. https://doi.org/10.3390/jof7121095
- 476. Randles JW, Rodriguez MJ, Imperial JS (1988) Cadang-cadang disease of coconut palm. Microbiol Sci 5:18–22
- 477. Redkar A, Matei A, Doehlemann G (2017) Insights into host cell modulation and induction of new cells by the corn smut Ustilago maydis. Front Plant Sci 8:899. https://doi.org/10.3389/fpls.2017. 00899
- 478. Reisser W (2002) Algae living on trees. In: Seckbach J (ed) Symbiosis: Mechanisms and Model Systems. Kluwer Academic Publishers, The Netherlands, pp 387–395
- Rhodes J (2019) Rapid Worldwide Emergence of Pathogenic Fungi. Cell Host Microbe 26:12–14. https://doi.org/10.1016/j. chom.2019.06.009
- 480. Rigling D, Prospero S (2017) Cryphonectria parasitica, the causal agent of chestnut blight: invasion history, population biology and disease control. Mol Plant Pathol 19:7–20
- 481. Della RG, Danti R, Williams N et al (2019) Molecular analyses indicate that both native and exotic pathogen populations serve as sources of novel outbreaks of Cypress Canker Disease. Biol Invasions 21:2919–2932. https://doi.org/10.1007/ s10530-019-02022-9
- Rogerson A, Detwiler A (1999) Abundance of airborne heterotrophic protists in ground level air of South Dakota. Atmos Res 51:35–44. https://doi.org/10.1016/S0169-8095(98)00109-4
- 483. Romay G, Pitrat M, Lecoq H et al (2019) Resistance against melon chlorotic mosaic virus and tomato leaf curl New Delhi virus in melon. Plant Dis 103:2913–2919. https://doi.org/10. 1094/PDIS-02-19-0298-RE
- Romon M, Soustre-Gacougnolle I, Schmitt C et al (2013) RNA silencing is resistant to low-temperature in grapevine. PLoS One 8:e82652. https://doi.org/10.1371/journal.pone.0082652
- 485. Roumi V, Gazel M, Caglayan K (2017) First report of Apple dimple fruit viroid in apple trees in Iran. New Dis Rep 35:3. https://doi.org/10.5197/j.2044-0588.2017.035.003
- 486. Rowe RC, Riedel RM, Martin MJ (1985) Synergistic interactions between Verticillium dahliae and Pratylenchus penetrans in potato early dying disease. Phytoapthology 75:412–418
- 487. Rowe RC, Davis JR, Powelson ML, Rouse DI (1987) Potato early dying: Causal agents and management strategies. Plant Dis 71:482–489
- 488. Rubio L, Galipienso L, Ferriol I (2020) Detection of plant viruses and disease management: Relevance of genetic diversity and evolution. Front Plant Sci 11:1092. https://doi.org/10.3389/fpls. 2020.01092
- 489. Rutherford RS, McFarlane SA, Van Antwerpen T, McFarlane K (2003) Use of varieties to minimise losses from sugarcane diseases in South Africa. Proc S Afr Sug Technol Ass 180–188
- Sache I (2000) Short-distance dispersal of wheat rust spores by wind and rain. Agronomie 20:757–767
- 491. Salgado-Salazar C, Shiskoff N, Daughtrey M et al (2018) Downy mildew: A serious disease threat to rose health worldwide. Plant Dis 102:1873–1882. https://doi.org/10.1094/ PDIS-12-17-1968-FE
- 492. Salinier J, Lefebvre V, Besombes D et al (2022) The INRAE centre for vegetable germplasm: geographically and phenotypically diverse collections and their use in genetics and plant breeding. Plants 11:347. https://doi.org/10.3390/plants11030347

- 493. Salmeron JM, Oldroyd GED, Rommens CMT et al (1996) Tomato Prf is a member of the leucine-rich repeat class of plant disease resistance genes and lies embedded within the Pto kinase gene cluster. Cell 86:123–133. https://doi.org/10.1016/S0092-8674(00)80083-5
- 494. Sanchez-Martín J, Rubiales D, Prats E (2011) Resistance to powdery mildew (Blumeria graminis f.sp. avenae) in oat seedlings and adult plants. Plant Pathol 60:846–856
- 495. Sarwar A, Latif Z, Zhang S et al (2018) Biological control of potato common scab with rare Isatropolone C compound produced by plant growth promoting streptomyces A1RT. Front Microbiol 9:1126. https://doi.org/10.3389/fmicb.2018.01126
- 496. Sasser JN, Eisenback JD, Carter CC, Triantaphyllou AC (1983) The international Meloidogyne project-its goals and accomplishments. Annu Rev Phytopathol 21:271–288. https://doi.org/10. 1146/annurev.py.21.090183.001415
- 497. Sastry KS (2013) Seed-borne plant virus diseases. Springer Science & Business Media 67–73
- 498. Savary S, Willocquet L, Pethybridge SJ et al (2019) The global burden of pathogens and pests on major food crops. Nat Ecol Evol 3:430–439. https://doi.org/10.1038/s41559-018-0793-y
- 499. Sawyer B, Elenbogen G, Rao KC et al (1993) Bacterial aerosol emission rates from municipal wastewater aeration tanks. Appl Environ Microbiol 59:3183–3186. https://doi.org/10.1128/aem. 59.10.3183-3186.1993
- Saxena A, Raghuwanshi R, Gupta VK, Singh HB (2016) Chilli anthracnose: the epidemiology and management. Front Microbiol 7:1527. https://doi.org/10.3389/fmicb.2016.01527
- Schleper C, Jurgens G, Jonuscheit M (2005) Genomic studies of uncultivated archaea. Nat Rev Microbiol 3:479–488. https://doi. org/10.1038/nrmicro1159
- 502. Schroeder JI, Allen GJ, Hugouvieux V et al (2001) Guard cell signal transduction. Annu Rev Plant Biol 52:627–658. https:// doi.org/10.1146/annurev.arplant.52.1.627
- 503. Schubert TS, Rizvi SA, Sun X, Gottwald TR, Graham JH, Dixon WN (2001) Meeting the challenge of eradicating citrus canker in Florida—again. Plant Dis 85:340–356. https://doi.org/10.1094/ PDIS.2001.85.4.340
- Scofield SR, Tobias CM, Rathjen JP et al (1996) Molecular basis of gene-for-gene specificity in bacterial speck disease of tomato. 274:2063–2065
- Scortichini M (2002) Bacterial canker and decline of European hazelnut. Plant Dis 86:704–709. https://doi.org/10.1094/PDIS. 2002.86.7.704
- 506. Scortichini M, Marcelletti S, Ferrante P et al (2012) Pseudomonas syringae pv. actinidiae: A re-emerging, multi-faceted, pandemic pathogen. Mol Plant Pathol 13:631–640. https://doi.org/10.1111/j.1364-3703.2012.00788.x
- 507. Seigner L, Liebrecht M, Keckel L et al (2020) Real-time RT-PCR detection of Citrus bark cracking viroid (CBCVd) in hops including an mRNA-based internal positive control. J Plant Dis Prot 127:763–767. https://doi.org/10.1007/s41348-020-00317-x
- 508. Seo JK, Kim MK, Kwak HR et al (2018) Molecular dissection of distinct symptoms induced by tomato chlorosis virus and tomato yellow leaf curl virus based on comparative transcriptome analysis. Virology 516:1–20. https://doi.org/10.1016/j.virol.2018.01.001
- 509. Serra-Soriano M, Navarro JA, Genoves A, Pallás V (2015) Comparative proteomic analysis of melon phloem exudates in response to viral infection. J Proteomics 124:11–24. https://doi. org/10.1016/j.jprot.2015.04.008
- 510. Serra P, Bertolini E, Martínez MC et al (2017) Interference between variants of peach latent mosaic viroid reveals novel features of its fitness landscape: Implications for detection. Sci Rep 7:4285. https://doi.org/10.1038/srep42825

- 511. Sesartic A, Dallafior TN (2011) Global fungal spore emissions, review and synthesis of literature data. Biogeosciences 8:1181– 1192. https://doi.org/10.5194/bg-8-1181-2011
- 512. Shaffer BT, Lighthart B (1997) Survey of the culturable airborne bacteria at four diverse locations in oregon: urban, rural, forest and coastal. Pap Knowl Towar a Media Hist Doc 34:167–177
- 513. Shapiro LR, Paulson JN, Arnold BJ et al (2018) An introduced crop plant is driving diversification of the virulent bacterial pathogen Erwinia tracheiphila. mBio 9:e01307-18. https://doi.org/10.1128/mBio.01307-18
- 514. Sharma NK, Rai AK, Singh S, Brown RM (2007) Airborne algae: Their present status and relevance. J Phycol 43:615–627. https:// doi.org/10.1111/j.1529-8817.2007.00373.x
- 515. Sharma U, Watpade S, Gupta B et al (2020) Economic losses due to infection by apple scar skin viroid in Himachal Pradesh, India. Virusdisease 31:490–496. https://doi.org/10.1007/ s13337-020-00625-8
- 516. Shivaji S, Chaturvedi P, Suresh K et al (2006) Bacillus aerius sp. nov., Bacillus aerophilus sp. nov., Bacillus stratosphericus sp. nov. and Bacillus altitudinis sp. nov., isolated from cryogenic tubes used for collecting air samples from high altitudes. Int J Syst Evol Microbiol 56:1465–1473. https://doi.org/10.1099/ijs.0.64029-0
- 517. Sigari G, Panatto D, Lai P et al (2006) Virological investigation on aerosol from waste depuration plants. J Prev Med Hyg 47:4–7
- 518. Silva H, Anjo SI, Manadas B et al (2021) Comparative Analysis of Bursaphelenchus xylophilus Secretome Under Pinus pinaster and P. pinea Stimuli. Front Plant Sci 12:668064. https://doi.org/ 10.3389/fpls.2021.668064
- 519. Singh P, Mazumdar P, Harikrishna J, Babu S (2019) Sheath blight of rice: a review and identification of priorities for future research. Planta 250. https://doi.org/10.1007/ s00425-019-03246-8
- 520. Singh RP (2014) The discovery and eradication of potato spindle tuber viroid in Canada. VirusDisease 25:415–424. https://doi.org/10.1007/s13337-014-0225-9
- 521. Singh RP, Huerta-Espino J, Bhavani S et al (2011) Race non-specific resistance to rust diseases in CIMMYT spring wheats. Euphytica 179:175–186. https://doi.org/10.1007/ s10681-010-0322-9
- 522. Sivapalan A, Hj Hamdan F, Junaidy MAHM (2007) Patch canker of Durio zibethinus caused by Phytophthora palmivora in Brunei Darussalam. Plant Dis 81. https://doi.org/10.1094/PDIS.1997. 81.1.113C
- 523. Skamnioti P, Gurr SJ (2009) Against the grain: safeguarding rice from rice blast disease. Trends Biotechnol 27:141–150. https:// doi.org/10.1016/j.tibtech.2008.12.002
- 524. Skelsey P, Cooke DEL, Lynott JS, Lees AK (2016) Crop connectivity under climate change: future environmental and geographic risks of potato late blight in Scotland. Glob Chang Biol 22:3724–3738. https://doi.org/10.1111/gcb.13368
- 525. Smant G, Helder J, Goverse A (2018) Parallel adaptations and common host cell responses enabling feeding of obligate and facultative plant parasitic nematodes. Plant J 93:686–702. https:// doi.org/10.1111/tpj.13811
- 526. Smets W, Moretti S, Denys S, Lebeer S (2016) Airborne bacteria in the atmosphere: Presence, purpose, and potential. Atmos Environ 139:214–221. https://doi.org/10.1016/j.atmosenv.2016. 05.038
- 527. Smith DJ, Timonen HJ, Jaffe DA, Griffin DW, Birmele MN, Perry KD, Ward PD, Roberts MS (2013) Intercontinental dispersal of bacteria and archaea by transpacific winds. Appl Environ Microbiol 79:1134–1139. https://doi.org/10.1128/AEM.03029-12
- 528. Smith IM, Dunez J, Lelliot RA et al (1988) European handbook of plant diseases. Blackewell Scientific publications, Oxford etc., 583 S

- 529. Smith M, Matavulj P, Mimić G et al (2022) Why should we care about high temporal resolution monitoring of bioaerosols in ambient air? Sci Total Environ 826:154231. https://doi.org/10. 1016/j.scitotenv.2022.154231
- Snetselaar K, McCann M (2017) Ustilago maydis, the corn smut fungus, has an unusual diploid mitotic stage. Mycologia 109:140–152. https://doi.org/10.1080/00275514.2016.1274597
- 531. Sobczak M, Avrova A, Jupowicz J et al (2005) Characterization of susceptibility and resistance responses to potato cyst nematode (Globodera spp.) infection of tomato lines in the absence and presence of the broad-spectrum nematode resistance Hero gene. Mol Plant-Microbe Interact 18:158–168. https://doi.org/10.1094/ MPMI-18-0158
- 532. Solmaz I, Sari N, Dogimont C, Pitrat M (2016) Evaluation of Turkish melon accessions for resistance to Fusarium wilt, downy mildew, powdery mildew, Cucumber mosaic virus and Zucchini yellow mosaic virus. XXIX International Horticultural Congress on Horticulture: Sustaining Lives, Livelihoods and Landscapes (IHC2014) 1127:133–140. https://doi.org/10.17660/ActaHortic. 2016.1127.22
- 533. Solomon SD, Qin D, Manning M et al (2007) Climate Change 2007: The Physical Science Basis: Working Group I Contribution to the Fourth Assessment Report of the IPCC. Cambridge Univ Press 4
- 534. Song WY, Kang MH, Kim HM (1999) Current status of bacterial brown stripe of rice caused by Acidovorax avenae subsp. avenae. Plant Dis Agric 5:69–76
- 535. Soto T, Lozano M, Vicente-Soler J et al (2009) Microbiological survey of the aerial contamination in urban areas of the city of Murcia, Spain. An Biol 31(2009):7–13
- 536. Spieker RL (1996) In vitro-generated 'inverse' chimeric Coleus blumei viroids evolve in vivo into infectious RNA replicons. J Gen Virol 77:2839–2846
- 537. Spielman LJ (1985) A monograph of Valsa on hardwoods in North America. Can J Bot 63:1355–1378. https://doi.org/10. 1139/b85-190
- 538. Srinivasan R, Riley D, Diffie S et al (2012) Whitefly population dynamics and evaluation of whitefly-transmitted tomato yellow leaf curl virus (TYLCV)-resistant tomato genotypes as whitefly and TYLCV reservoirs. J Econ Entomol 105:1447–1456. https:// doi.org/10.1603/EC11402
- 539. Stanley R, Linskins H (1974) Pollen: Biology, Chemistry and Management. Springer-Verlag, Berlin
- 540. Stewart I, Falconer IR (2008) Cyanobacteria and cyanobacterial toxins. In: Walsh PJ, Smith SL, Fleming LE, Solo-gabriele HM, Gerwick WH (eds) Oceans and human health: risks and remedies from the seas. Academic Press, Burlington, MA, pp 271–296
- 541. Stocker TF, Qin D, Plattner G-K et al (2013) IPCC, 2013: Climate Change 2013: The Physical Science Basis. Contribution of Working Group I to the Fifth Assessment Report of the Intergovernmental Panel on Climate Change. Cambridge University Press, Cambridge, University and New York, NY, USA
- Stout JE (2001) Dust and environment in the southern high plains of North America. J Arid Environ 47:425–441. https://doi.org/ 10.1006/jare.2000.0732
- Strauch D, Ballarini G (1994) Hygienic aspects of the production and agricultural use of animal wastes. J Vet Med Ser B 41:176– 228. https://doi.org/10.1111/j.1439-0450.1994.tb00222.x
- 544. Su H, Van Bruggen AHC, Subbarao KV (2000) Spore release of Bremia lactucae on lettuce is affected by timing of light initiation and decrease in relative humidity. Phytopathology 90:67–71. https://doi.org/10.1094/PHYTO.2000.90.1.67
- 545. Sukno SA, García VM, Shaw BD, Thon MR (2008) Root infection and systemic colonization of maize by Collectorichum graminicola. Appl Environ Microbiol 74:823–832. https://doi. org/10.1128/AEM.01165-07

- 546. Sulong Y, Zakaria AJ, Mohamed S et al (2019) Survey on pest and disease of corn (Zea Mays Linn) grown at BRIS soil area. J Agrobiotechnol 10:2180–1983
- 547. Suriani PB, Junaid M, Muis A (2021) The presence of bacterial stalk rot disease on corn in Indonesia: A review. IOP Conf Ser Earth Environ Sci 911:012058. https://doi.org/10.1088/1755-1315/911/1/012058
- 548. Sutton PN, Henry MJ, Hall JL (1999) Glucose, and not sucrose, is transported from wheat to wheat powdery mildew. Planta 208:426–430. https://doi.org/10.1007/s004250050578
- 549. Sweigard JA, Carroll AM, Farrall L et al (1998) Magnaporthe grisea pathogenicity genes obtained through insertional mutagenesis. Mol Plant-Microbe Interact 11:404–412. https://doi.org/10. 1094/MPMI.1998.11.5.404
- 550. Szittya G, Silhavy D, Molnár A et al (2003) Low temperature inhibits RNA silencing-mediated defence by the control of siRNA generation. EMBO J 22:633–640. https://doi.org/10.1093/ emboj/cdg74
- 551. Taha H, Shivanand P, Khoo DH et al (2020) Identification of culturable petroleum-degrading bacteria and fungi from petroleum-contaminated sites in Brunei Darussalam. J Environ Sci Heal - Part A Toxic/Hazard Subst Environ Eng 55:1542–1547. https://doi.org/10.1080/10934529.2020.1826238
- 552. Taha H, Shivanand P, Shahminan NIN et al (2020b) Isolation and identification of culturable bacteria and fungi from mixed dipterocarp and mangrove forests of Brunei Darussalam. Proc Natl Acad Sci India Sect B - Biol Sci 90:523–530.https://doi. org/10.1007/s40011-019-01119-4
- 553. Taha H, Shivanand P, Zainudin MAA, Hadanan NA (2021) Identification of culturable marine fungi and bacteria from coastal region in Brunei Darussalam: Identification of culturable marine microbes. Biodiversitas Journal of Biological Diversity 22:3. https://doi.org/10.13057/biodiv/d220332
- 554. Takahata Y (2001) Changes of xylem pressure potential in Quercus serrata saplings inoculated with Raffaelea sp. (in Japanese). Abstracts of the 112th Annual Meeting of the Japanese Forestry Society 112:284.
- 555. Takatsu A, Fukuda S, Hahn SK, Caveness FE (1990) Integrated pest management for tropical root and tuber crops. In: Hahn, S.K. and Caveness, F.E. (eds) Proceedings of the Workshop on the Global Status and of Prospects for IPM of Root and Tuber Crops, Ibadan, Nigeria, 25–30 October 1987. IITA, Ibadan, Nigeria, pp 127–131
- 556. Talbot NJ (2007) Fungal genomics goes industrial. Nat Biotechnol 25:542–543. https://doi.org/10.1038/nbt0507-542
- 557. Tan TK, Teo TS, Lee BW et al (1992) Variations in tropical airspora in Singapore. Mycol Res 96:221–224. https://doi.org/ 10.1016/S0953-7562(09)80969-6
- 558. Tang JW (2009) The effect of environmental parameters on the survival of airborne infectious agents. J R Soc Interface 6:S737– S746. https://doi.org/10.1098/rsif.2009.0227.focus
- 559. Tang X, Frederick RD, Zhou J et al (1996) Initiation of plant disease resistance by physical interaction of AvrPto and Pto kinase. Science (80-) 274:2060–2063
- 560. Tangkanchanapas P, Haegeman A, Höfte M, De Jonghe K (2021) Reassessment of the Columnea latent viroid (CLVd) taxonomic classification. Microorganisms 9:1117. https://doi.org/10.3390/ microorganisms9061117
- 561. Tangkanchanapas P, Reanwarakorn K, Juenak H, De Jonghe K (2017) First report of Grapevine yellow speckle viroid-2 infecting grapevine (Vitis vinifera) in Thailand. New Dis Rep 36:6. https://doi.org/10.5197/j.2044-0588.2017.036.006
- 562. Täubel M, Rintala H, Pitkäranta M et al (2009) The occupant as a source of house dust bacteria. J Allergy Clin Immunol 124:834– 840. https://doi.org/10.1016/j.jaci.2009.07.045

- 563. Taylor SE, Brown JD (1988) Illusion and well-being: A social psychological perspective on mental health. Psychol Bull 103:193–210. https://doi.org/10.1037/0033-2909.103.2.193
- 564. Tesson SVM, Skjøth A, Šantl-temkiv T (2016) Airborne microalgae: Insights, opportunities, and challenges. Appl Environ Microbiol 82:1978–1991. https://doi.org/10.1128/AEM.03333-15.Editor
- 565. Than PP, Prihastuti H, Phoulivong S, Taylor PW, Hyde KD (2008) Chilli anthracnose disease caused by Colletotrichum species. J of Zhejiang University Science B 9:764–778. https://doi. org/10.1631/jzus.B0860007
- 566. Thomason IJ, Rich JR, O'Melia FC (1976) Pathology and histopathology of Pratylenchus scribneri infecting snap bean and lima bean. J Nematol 8:347–352
- 567. Thompson WAR (1981) editor. Black's medical dictionary, 33 ed. Adam and Charles Black
- 568. Thummes K, Schäfer J, Kämpfer P, Jäckel U (2007) Thermophilic methanogenic Archaea in compost material: Occurrence, persistence and possible mechanisms for their distribution to other environments. Syst Appl Microbiol 30:634–643. https:// doi.org/10.1016/j.syapm.2007.08.001
- 569. Tokunaga H, Baba T, Ishitani M et al (2018) Correction to: Sustainable Management of Invasive Cassava Pests in Vietnam, Cambodia, and Thailand: Application of Cutting-edge Science and Technology in Developing Countries. Crop Production under stressful conditions. Springer, Singapore, pp E1–E1
- Tong Y, Lighthart B (1999) Diurnal distribution of total and culturable atmospheric bacteria at a rural site. Aerosol Sci Technol 30:246–254. https://doi.org/10.1080/027868299304822
- 571. Toniutti L, Breitler JC, Etienne H et al (2017) Influence of environmental conditions and genetic background of Arabica coffee (C. Arabica L) on leaf rust (Hemileia vastatrix) pathogenesis. Front Plant Sci 8:2025. https://doi.org/10.3389/fpls.2017.02025
- 572. Toropova EY, Kazakova OA, Piskarev VV (2020) Septoria blotch epidemic process on spring wheat varieties. Vavilovskii Zh Genetiki Selektsii 24:139–148. https://doi.org/10.18699/VJ20.609
- Toth IK, Kenneth SB, Maria CH, Paul RJB (2003) Soft rot erwiniae: from genes to genomes. Mol Plant Pathol 4:17–30
- 574. Trail F (2007) Fungal cannons: Explosive spore discharge in the Ascomycota. FEMS Microbiol Lett 276:12–18. https://doi.org/ 10.1111/j.1574-6968.2007.00900.x
- 575. Tripathi S, Suzuki JY, Ferreira SA, Gonsalves D (2008) Papaya ringspot virus-P: characteristics, pathogenicity, sequence variability and control. Mol Plant Pathol 9:269–280. https://doi.org/ 10.1111/j.1364-3703.2008.00467.x
- 576. Trudgill DL, Blok VC (2001) Apomictic, polyphagous root-knot nematodes: Exceptionally successful and damaging biotrophic root pathogens. Annu Rev phytopathol 39:53–77. https://doi.org/ 10.1146/annurev.phyto.39.1.53
- 577. Tsushima T, Sano T (2015) First report of Coleus blumei viroid 5 infection in vegetatively propagated clonal coleus cv.'Aurora black cherry'in Japan. New Dis Rep 32(7). https://doi.org/10. 5197/j.2044-0588.2015.032.007
- 578. Tsushima T, Sano T (2018) A point-mutation of Coleus blumei viroid 1 switches the potential to transmit through seed. J Gen Virol 99:393–401. https://doi.org/10.1099/jgv.0.001013
- 579. Tytgat T, De Meutter J, Gheysen G, Coomans A (2000) Sedentary endoparasitic nematodes as a model for other plant parasitic nematodes. Nematology 2:113–121. https://doi.org/10.1163/ 156854100508827
- USDA (2013) USDA National Nutrient Database for Standard Reference, Release 26. Nutrient Data Laboratory Home Page, http://www.ars.usda.gov/ba/bhnrc/ndl. Accessed 7 October 2022
- 581. Uke A, Tokunaga H, Utsumi Y et al (2021) Cassava mosaic disease and its management in Southeast Asia. Plant Mol Biol 109:301–311. https://doi.org/10.1007/s11103-021-01168-2

- USDA (2015) The plantss database (http://plants.usda.gov, May 2011). National Plant Data Team, Greensboro. Accessed 7 October 2022
- 583. van Esse HP, Reuber TL, van der Does D (2020) Genetic modification to improve disease resistance in crops. New Phytol 225:70–86. https://doi.org/10.1111/nph.15967
- 584. Van Megen H, Van Den Elsen S, Holterman M et al (2009) A phylogenetic tree of nematodes based on about 1200 full-length small subunit ribosomal DNA sequences. Nematology 11:927– 950. https://doi.org/10.1163/156854109X456862
- Vaneault-Fourrey C, Barooah M, Egan M et al (2006) Autophagic fungal cell death is necessary for infection by the rice blast fungus. Science (80-) 312:580–583. https://doi.org/10.1126/science. 1124550
- 586. Varma A, Malathi VG (2003) Emerging geminivirus problems: A serious threat to crop production. Ann Appl Biol 142:145–164. https://doi.org/10.1111/j.1744-7348.2003.tb00240.x
- 587. Váry Z, Mullins E, Mcelwain JC, Doohan FM (2015) The severity of wheat diseases increases when plants and pathogens are acclimatized to elevated carbon dioxide. Glob Chang Biol 21:2661–2669. https://doi.org/10.1111/gcb.12899
- Velásquez AC, Castroverde CDM, He SY (2018) Plant-pathogen warfare under changing climate conditions. Curr Biol 28:R619– R634. https://doi.org/10.1016/j.cub.2018.03.054
- 589. Verbeek M, Dullemans AM, van Bekkum PJ, van der Vlugt RAA (2013) Evidence for Lettuce big-vein associated virus as the causal agent of a syndrome of necrotic rings and spots in lettuce. Plant Pathol 62:444–451. https://doi.org/10.1111/j.1365-3059.2012.02645.x
- 590. Verhoeven JT, Roenhorst JW, Hooftman M et al (2015) A pospiviroid from symptomless portulaca plants closely related to iresine viroid 1. Virus Res 205:22–26. https://doi.org/10.1016/j.virus res.2015.05.005
- 591. Verhoeven JTJ, Botermans M, Meekes ETM, Roenhorst JW (2012) Tomato apical stunt viroid in the Netherlands: Most prevalent pospiviroid in ornamentals and first outbreak in tomatoes. Eur J Plant Pathol 133:803–810. https://doi.org/10.1007/s10658-012-0005-6
- 592. Verhoeven JTJ, Jansen CCC, Willemen TM et al (2004) Natural infections of tomato by Citrus exocortis viroid, Columnea latent viroid, Potato spindle tuber viroid and Tomato chlorotic dwarf viroid. Eur J Plant Pathol 110:823–831. https://doi.org/10.1007/s10658-004-2493-5
- Veron F (2015) Ocean spray. Annu Rev Fluid Mech 47:507– 538. https://doi.org/10.1146/annurevfluid-010814-014651
- 594. Villeneuve F, Latour F, Théry T et al (2014) The control of soil-borne vascular diseases: Limits of genetic resistance of cultivars and rootstocks for controlling Fusarium oxysporum F. Sp. Melonis (Melon) and Verticillium Sp. (Eggplant). Acta Hortic. 57–65
- 595. Viswanathan R, Balamuralikrishnan M (2005) Impact of mosaic infection on growth and yield of sugarcane. Sugar Tech 7:61–65. https://doi.org/10.1007/BF02942419
- 596. von Blohn N, Mitra SK, Diehl K, Borrmann S (2005) The ice nucleating ability of pollen: Part III: New laboratory studies in immersion and contact freezing modes including more pollen types. Atmos Res 78:182–189
- 597. Waals JE, Korsten L, Aveling T (2001) A review of early blight of potato. Afr Plant Prot 7:91–102
- 598. Wainwright M, Wickramasinghe NC, Narlikar JV, Rajaratnam P (2003) Microorganisms cultured from stratospheric air samples obtained at 41 km. FEMS Microbiol Lett 218:161–165. https:// doi.org/10.1016/S0378-1097(02)01138-2
- 599. Wang T, Jia ZH, Zhang JY et al (2020) Identification and analysis of nbs-lrr genes in actinidia chinensis genome. Plants 9:1350. https://doi.org/10.3390/plants9101350

- 600. Wang Y, Wu J, Qiu Y et al (2019) Global transcriptomic analysis reveals insights into the response of 'etrog' citron (Citrus medica L.) to Citrus Exocortis viroid infection. Viruses 11. https://doi. org/10.3390/v11050453
- 601. Wang Z, Chen B, Zhang T et al (2021) Rice stripe mosaic disease: Characteristics and control strategies. Front Microbiol 12:715223. https://doi.org/10.3389/fmicb.2021.715223
- 602. Wellings CR (2011) Global status of stripe rust: A review of historical and current threats. Euphytica 179:129–141. https:// doi.org/10.1007/s10681-011-0360-y
- 603. Whitham S, Dinesh-kumar SP, Choi D et al (1994) The product of the tobacco mosaic virus resistance gene N: similarity to toll and the interleukin-1 receptor. Cell 78:1101–1115. https://doi. org/10.1016/0092-8674(95)90399-2
- Whittaker RH, Likens GE (1973) Primary production: The biosphere and man. Hum Ecol 1:357–369. https://doi.org/10.1007/BF01536732
- 605. Wietholter N, Graessner B, Mierau M, Mort AJ, Moerschbacher BM (2003) Differences in the methyl ester distribution of homogalacturonans from near-isogenic wheat lines resistant and susceptible to the wheat stem rust fungus. Mol Plant Microbe Interact 16:945–952
- 606. Will T, Furch ACU, Zimmermann MR (2013) How phloemfeeding insects face the challenge of phloem-located defenses. Front Plant Sci 4:336. https://doi.org/10.3389/fpls.2013.00336
- 607. Williamson-Benavides BA, Dhingra A (2021) Understanding root rot disease in agricultural crops. Horticulturae 7:33. https:// doi.org/10.3390/horticulturae7020033
- 608. Wilson TW, Ladino LA, Alpert PA et al (2015) A marine biogenic source of atmospheric ice-nucleating particles. Nature 525:234–238. https://doi.org/10.1038/nature14986
- 609. Wingfield MJ, Hammerbacher A, Ganley RJ et al (2008) Pitch canker caused by Fusarium circinatum - A growing threat to pine plantations and forests worldwide. Australas Plant Pathol 37:319–334. https://doi.org/10.1071/AP08036
- 610. Womack AM, Bohannan BJM, Green JL (2010) Biodiversity and biogeography of the atmosphere. Philos Trans R Soc B Biol Sci 365:3645–3653. https://doi.org/10.1098/rstb.2010.0283
- 611. Woo C, An C, Xu S, Yamamoto N (2018) Taxonomic diversity of fungi deposited from the atmosphere. ISME J 12:2051–2060. https://doi.org/10.1038/s41396-018-0160-7
- 612. Wu L, Zu X, Wang S, Chen Y (2012) Sugarcane mosaic virus e Long history but still a threat to industry. Crop Prot 42:74–78. https://doi.org/10.1016/j.cropro.2012.07.005
- 613. Wunderle J, Leclerque A, Schaffrath U, Slusarenko A, Koch E (2012) Assessment of the loose smut fungi (Ustilago nuda and U. tritici) in tissues of barley and wheat by fluorescence microscopy and real-time PCR. Eur J Plant Pathol 133:865–875. https://doi.org/10.1007/s10658-012-0010-9
- 614. Wydra K, Msikita W (1998) An overview of the present situation of cassava diseases in West Africa. In: Akoroda MO, Ekanayake IJ (eds) Root Crops for Poverty Alleviation. Proceedings of the Sixth Triennial Symposium of the International Society for Tropical Root Crops (ISTRC), Lilongwe, Malawi, 22–28 October 1995. ISTRC (International Society for Tropical Root Crops), IITA (International Institute of Tropical Agriculture) and Government of Malawi, pp 163–166
- 615. Xiang N, Lawrence KS, Donald PA (2018) Biological control potential of plant growth-promoting rhizobacteria suppression of Meloidogyne incognita on cotton and Heterodera glycines on soybean: A review. J Phytopathol 166:449–458. https://doi.org/ 10.1111/jph.12712
- 616. Xie X, Li Y, Sun H, Liu L (2009) Exhaled droplets due to talking and coughing. J R Soc Interface 6:703–714. https://doi.org/10. 1098/rsif.2009.0388.focus
- 617. Xu D, Adkar-Purushothama CR, Lemoyne P et al (2021) First report of Grapevine Yellow Speckle Viroid 1 infecting grapevine

(Vitis vinifera) in Canada. Plant Dis 12. https://doi.org/10.1094/ PDIS-04-21-0863-PDN

- 618. Xu J, Jiang J, Dong X et al (2012) Introgression of bacterial blight (BB) resistance genes Xa7 and Xa21 into popular restorer line and their hybrids by molecular marker-assisted backcross (MABC) selection scheme. African J Biotechnol 11:8225–8233. https://doi.org/10.5897/ajb12.341
- 619. Xu L, Wang JW, Zhu DZ et al (2017) First report of hop stunt viroid from sweet cherry with dapple fruit symptoms in China. Plant Dis 101:394
- 620. Xu X-M, Pettitt T (2004) Overwintering of rose downy mildew (Peronospora sparsa). In: Spencer-Phillips P, Jeger M (eds) Advances in Downy Mildew Research, vol 2. Kluwer Academic Publishers. Dordrecht, TheNetherlands, pp 99–106
- 621. Xue AG, Cober E, Morrison MJ et al (2007) Effect of seed treatments on emergence, yield, and root rot severity of soybean under Rhizoctonia solani inoculated field conditions in Ontario. Can J Plant Sci 87:167–173
- 622. Yadav RKP, Halley JM, Karamanoli K et al (2004) Bacterial populations on the leaves of Mediterranean plants: Quantitative features and testing of distribution models. Environ Exp Bot 52:63–77. https://doi.org/10.1016/j.envexpbot.2004.01.004
- 623. Yadav RKP, Karamanoli K, Vokou D (2005) Bacterial colonization of the phyllosphere of mediterranean perennial species as influenced by leaf structural and chemical features. Microb Ecol 50:185–196. https://doi.org/10.1007/s00248-004-0171-y
- 624. Yakop F, Taha H, Shivanand P (2019) Isolation of fungi from various habitats and their possible bioremediation. Curr Sci 116:733–740. https://doi.org/10.18520/cs/v116/i5/733-740
- 625. Yan H, Li Y, Zhang Y et al (2021) Deciphering of microbial diversity and antibiotic resistome of bioaerosols in swine confinement buildings. Sci Total Environ 781:147056. https://doi.org/10.1016/j.scitotenv.2021.147056
- 626. Yan X, Talbot NJ (2016) Investigating the cell biology of plant infection by the rice blast fungus Magnaporthe oryzae. Curr Opin Microbiol 34:147–153. https://doi.org/10.1016/j.mib.2016.10.001
- 627. Yang W, Elankumaran S, Marr LC (2011) Concentrations and size distributions of airborne influenza A viruses measured indoors at a health centre, a day-care centre and on aeroplanes. J R Soc Interface 8:1176–1184. https://doi.org/10.1098/rsif.2010. 0686

- 628. Yasuda F, Kobayashi T, Watanabe H, Izawa H (2003) Addition of Pestalotiopsis spp. to leaf spot pathogens of Japanese persimmon. J Gen Plant Pathol 69:29–32. https://doi.org/10.1007/ s10327-002-0011-1
- Yousif MT, Kheyr-Pour A, Gronenborn B et al (2007) Sources of resistance to watermelon Chlorotic Stunt Virus in Melon. 126:422–427.https://doi.org/10.1111/j.1439-0523.2007.01366.x
- 630. Zhai Y, Li X, Wang T et al (2018) A review on airborne microorganisms in particulate matters: Composition, characteristics and influence factors. Environ Int 113:74–90. https://doi.org/10. 1016/j.envint.2018.01.007
- 631. Zhang H, Mao R, Wang Y et al (2019) Transcriptome-wide alternative splicing modulation during plant-pathogen interactions in wheat. Plant Sci 288:110160. https://doi.org/10.1016/j.plantsci. 2019.05.023
- 632. Zhao J, Li L, Liu Q et al (2019) A MIF-like effector suppresses plant immunity and facilitates nematode parasitism by interacting with plant annexins. J Exp Bot 70:5943–5958. https://doi.org/10. 1093/asj/sjy179/5059000
- 633. Zhao P, Sun X, Li P et al (2019) Infection characteristics of rice stripe mosaic virus in the body of the vector leafhoppers. Front Microbiol 9:3258. https://doi.org/10.3389/fmicb.2018. 03258
- 634. Zhdanov VM, Gaudamovich SY (1982) Special Virology: Handbook. Meditsina, Moscow
- 635. Zheng A, Lin R, Zhang D et al (2013) The evolution and pathogenic mechanisms of the rice sheath blight pathogen. Nat Commun 4:1424. https://doi.org/10.1038/ncomms2427

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