

# SPECIAL ISSUE: MACROALGAE IN A CHANGING WORLD

# Projecting kelp (*Ecklonia radiata*) gametophyte thermal adaptation and persistence under climate change

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• **Background and aims** Kelp forests underpin temperate marine ecosystems but are declining due to ocean warming, causing loss of associated ecosystem services. Projections suggest significant future decline but often only consider the persistence of adult sporophytes. Kelps have a biphasic life cycle, and the haploid gametophyte can be more thermally tolerant than the sporophyte. Therefore, projections may be altered when considering the thermal tolerance of gametophytes.

• **Methods** We undertook thermal tolerance experiments to quantify the effect of temperature on gametophyte survival, relative growth rate (RGR) and sex ratio for three genetically distinct populations of *Ecklonia radiata* gametophytes from comparatively high, mid- and low latitudes (43°, 33° and 30°S). We then used these data to project the likely consequences of climate-induced thermal change on gametophyte persistence and performance across its eastern Australian range, using generalized additive and linear models.

• **Key results** All populations were adapted to local temperatures and their thermal maximum was 2–3 °C above current maximum *in situ* temperatures. The lowest latitude population was most thermally tolerant (~70 % survival up to 27 °C), while survival and RGR decreased beyond 25.5 and 20.5 °C for the mid- and low-latitude populations, respectively. Sex ratios were skewed towards females with increased temperature in the low- and high-latitude populations. Spatially explicit model projections under future ocean warming (2050-centred) revealed a minimal decline in survival (0–30 %) across populations, relative to present-day predictions. RGRs were also projected to decline minimally (0–2 % d<sup>-1</sup>).

• **Conclusions** Our results contrast with projections for the sporophyte stage of *E. radiata*, which suggest a 257-km range contraction concurrent with loss of the low-latitude population by 2100. Thermal adaptation in *E. radiata* gametophytes suggests this life stage is likely resilient to future ocean warming and is unlikely to be a bottleneck for the future persistence of kelp.

**Key words:** Climate change, *Ecklonia radiata*, gametophytes, generalized additive modelling, genetic clusters, kelp forests, thermal adaptation, ocean warming, spatial projections, range edge, thermal resilience.

# INTRODUCTION

Global climate change is increasingly affecting marine ecosystems (Harley et al., 2006; Doney et al., 2011; Gissi et al., 2021) with rising ocean temperatures driving profound changes to the distribution, ecological structure and reproductive phenology of subtidal marine communities (Sorte et al., 2010; Poloczanska et al., 2013; Boyce et al., 2015). As all marine species have a thermal niche in which they thrive (Tittensor et al., 2010; Stuart-Smith et al., 2017), increasing temperatures are shifting the distribution of species poleward (Sorte et al., 2010; Bates et al., 2014; Chaudhary et al., 2021) and causing local extinctions at trailing range edges (Nicastro et al., 2013; Fredston-Hermann et al., 2020; Pinsky et al., 2020). The impacts of increased temperature may vary across species' life stages and spatial distribution (Pankhurst and Munday, 2011; de Bettignies et al., 2018), making it vital to capture the thermal response of different life stages and populations to project distributional changes (Foden et al., 2019; Twiname et al., 2020).

Kelp forests are among the coastal marine ecosystems most impacted by global climate change (Bartsch et al., 2012; Harley et al., 2012; Wernberg et al., 2019b) with losses of biomass, abundance and range as well as local extinctions documented in North America and Canada (Filbee-Dexter et al., 2016; Berry et al., 2021), California and South America (Schiel et al., 2004; Arafeh-Dalmau et al., 2019), Australia and New Zealand (Johnson et al., 2011; Wernberg et al., 2016; Thomsen et al., 2019) and Europe (Moy and Christie, 2012; Voerman et al., 2013). Kelps are foundation species that provide habitat and vast ecosystem services in temperate marine environments (Dayton, 1985; Steneck et al., 2002), estimated to value up to 684 billion USD annually (Eger et al., 2023). The loss of these foundation species signifies a loss in overall marine biodiversity (Castorani et al., 2018; Gabara et al., 2021; Pessarrodona et al., 2021) and fragmentation of kelp habitat reduces recruitment, triggering further kelp decline (Layton et al., 2020; Reeves et al., 2022).

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To enable proactive management and conservation of kelp forests globally, predictive modelling has been used to elucidate where kelp is likely to be lost (Assis et al., 2018; Martínez et al., 2018; Sudo et al., 2020) and to identify areas of persistence in the future (Bekkby and Moy, 2011; Franco et al., 2018; Davis et al., 2021, 2022). Range contractions are common along coastlines affected by ocean warming and can be especially severe where there is no suitable habitat for poleward range expansion. For example, the distribution of sporophyte populations of the main canopy-forming kelp in Australia, Ecklonia radiata, are projected to decline by 71 and 49 % under RPC 6.0 and 2.6 emission scenarios, respectively (Martínez et al., 2018). More nuanced estimates show a 257-km range contraction, a shift to deep cooler water and 30 % loss of biomass on Australia's east coast (Davis et al., 2022). Similarly, suitable habitat for kelp forests in Japan is projected to decline between 50 and 100 % resulting in extinction of six out of 11 native kelp species (Sudo et al., 2020). A common shortcoming of such models used to predict future kelp forest distributions, however, is the consideration of only the macroscopic sporophyte stage of kelp. Despite its importance in kelp forest persistence, the survival and performance of the microscopic gametophyte stage is rarely considered.

Kelps are characterized by a biphasic life history, where the adult diploid sporophyte alternates with the microscopic haploid gametophyte (Pedersen, 1981). The gametophyte stage has been considerably less researched than the sporophyte stage, but is integral for kelp forest persistence (Veenhof et al., 2022a) as the stage through which regeneration occurs is pivotal in understanding species reactions to climate change (Parmesan and Hanley, 2015). The complex life history of kelps makes predicting distributional changes convoluted as two separate life stages are to be considered (Murphy et al., 2016, 2017). Most projections of distributional change are based on records of sporophyte presence in the field (i.e. Martínez et al., 2018). Some authors have included theoretical model parameters accounting for dispersal and delayed development of microscopic stages (Assis et al., 2017) or physiological thresholds of the microscopic phase based on the literature (Murphy et al., 2016, 2017). Few authors have tried to experimentally test limits of microscopic life stages and integrate them in models derived from mechanistic relationships (but see Capdevila et al., 2019). Mechanistic relationships derived from experimental data on species' physiological thresholds have the potential for highly accurate projections and the microscopic life stage of kelp is well suited for this method (Kearney and Porter, 2009; Twiname et al., 2020).

Predictive models based on gametophyte physiology could supplement existing models projecting sporophyte decline, together presenting a holistic view of kelp distributional changes and where critical bottlenecks might lie. Gametophyte temperature thresholds can be higher than those of sporophytes (Bartsch *et al.*, 2013; Mohring *et al.*, 2014; Martins *et al.*, 2017; Becheler *et al.*, 2022), allowing gametophytes to persist through stressful conditions (Barradas *et al.*, 2011; Carney *et al.*, 2013). For example, the known thermal tolerance of *E. radiata* gametophytes ranges between 1 and 28 °C (tom Dieck, 1993; Mabin *et al.*, 2013; Mohring *et al.*, 2014), compared to temperature thresholds of 8 and 25 °C for sporophyte populations (Wernberg *et al.*, 2019*a*). Gametophyte models are likely to present altered projections of decline if gametophytes can survive more extreme temperatures during periods of thermal stress. Kelps can also display local thermal adaptation throughout their range which, if incorporated into model projections, would likely alter outcomes. Gametophyte populations from warmer edges of their distribution can have a higher thermal tolerance (Oppliger *et al.*, 2012; Mohring *et al.*, 2014; Martins *et al.*, 2020; Liesner *et al.*, 2022), which may be underpinned by both plasticity and heritable genetic variation across a species' range (King *et al.*, 2019; Vranken *et al.*, 2021; Wood *et al.*, 2021) and so the warmer, trailing edge may respond differently to climate change (Hampe and Petit, 2005; Mota *et al.*, 2018). To accurately predict changes at trailing edges, rather than use a single thermal envelope as a predictor, spatial variation in a species' thermal tolerance underlaid by genetic variation should be captured in species distribution models (Russell *et al.*, 2012).

Models that explicitly quantify the drivers of early kelp life stages using laboratory-derived data are vital to better understand how kelp forests will cope in the face of climate change (Assis et al., 2017; Twiname et al., 2020). Here we use thermal experiments to develop such models for gametophytes of the main canopy-forming species in Australia, E. radiata, sampled across ~1500 km of latitude. First, we identify the specific thermal tolerance of genetically distinct populations of E. radiata gametophytes across its range along eastern Australia. Second, we use these data to construct spatially explicit models of gametophyte survival, relative growth rate (RGR) and sex ratio that account for differential responses of thermally adjusted gametophytes. Third, we project these responses under historical and future ocean conditions and compare the change in gametophyte response, and fourth, we compare outcomes of the gametophyte projections to projected changes to sporophyte distributions to create a holistic view of future persistence of E. radiata in eastern Australia.

# MATERIALS AND METHODS

#### Study species

Ecklonia radiata gametophytes were sourced at three different populations (two or three sites within each population) off the east coast of Australia based on distinct genetic clusters (AJP Minne et al., in prep.; Coleman et al., 2011b). Approximately ten fertile plants were collected from each site and used to create stock cultures from (1) Charlesworth Bay (30°27'S, 153°14"E), Diggers Beach (30°28'S, 153°15'E) and Sawtell (30°37'S, 153°08'E) on 16 July 2021 (population Coffs Harbour), (2) in Bateau Bay (33°23'S, 151°29'E) and Toowoon Point (33°21'S, 151°30'E) on the 5 June 2022 (population Sydney) and (3) in Fortescue Bay (43°13'S, 147°97'E), Apex Point (43°06'S, 147°43'E) and Coal Point (43°33'S, 147°32'E) on 13, 11 and 15 August 2020, respectively (population Tasmania). These populations were chosen to represent a comparatively low-latitude (Coffs Harbour), mid-latitude (Sydney) and high-latitude (Tasmania) ranges of genetically distinct clusters within the east coast distribution of E. radiata (Minne et al., in prep.; Coleman et al., 2011b). Spores were released and gametophytes grown in culture as in Veenhof et al. (2022b). Cultures were then refreshed monthly with 1 um filtered. UV-sterilized seawater (FSW) containing quarter strength F growing media (AlgaBoost<sup>™</sup> 2000×, AusAqua

Pty Ltd, Wallaroo, SA, Australia) under an irradiation of 20  $\mu$ mol m<sup>-2</sup> s<sup>-1</sup> at a 12:12-h cycle and 20, 16 and 12 °C for Coffs Harbour, Sydney and Tasmania, respectively. Cultures were maintained in climatic chambers fitted with LED lamps. These temperatures were based on the approximate average local *in situ* winter temperatures for each population at the time of collection (Wijffels *et al.*, 2018). Coffs Harbour cultures were ~1 year old, Sydney cultures ~2 months old and Tasmania cultures 2 years old before use in this experiment. Before use in this experiment, for each population one culture flask was gently scraped clean with a sterile scraper and blended up to single fragment filaments (~185 ± 15 µm, mean ± s.e.) and diluted at a 1:12 ratio for the final gametophyte stock solution (~27 ± 5 gametophytes mL<sup>-1</sup>). Gametophytes from each population were kept separately and used in independent experiments as below.

#### Experimental design

To create a continuous and stable thermal gradient to measure gametophyte responses, we used a purpose-built aluminium thermal gradient block. This aluminium block had 50 wells (41 mm  $\emptyset$ , 25 mm deep) in a 10  $\times$  5 grid. At the far ends of the block were bores through which recirculating hot and cold water flowed continuously. This water was supplied by baths that were cooled using a cooler/heater unit (Teco TK 500) and heated by aquarium heaters (Aqualogic DC series). This system established a stabile thermal gradient resulting in ten temperature treatments along the length of the block that were replicated five times along the breadth of the block. Fifty glass beakers (38 mm  $\emptyset$ ) were placed in the temperature block. Each beaker contained 40 mL diluted gametophyte stock solution and two round coverslips (12 mm Ø) as a surface for gametophyte settlement. The opening of each beaker was covered with parafilm to prevent evaporation while allowing sufficient light to penetrate the bottom of the glass beakers (27  $\mu$ mol m<sup>-2</sup> s<sup>-1</sup>). Light was supplied via TL fluorescent lamps wrapped in shade cloth, and set at ~27  $\mu$ mol m<sup>-2</sup> s<sup>-1</sup> as measured in the wells covered with parafilm, which is within the light optimum for E. radiata gametophytes (Novaczek, 1984a). For each population, a different starting temperature was chosen based on the cultures holding temperatures, which were related to average in situ winter temperatures at the time of collection to minimize thermal shock (Supplementary Data Fig. S1). The cultures were left for 24 h before starting thermal treatments to allow settlement and attachment to the coverslips and bottom of the glass beakers. One set of replicates (n = 5) was kept outside of the temperature block at the respective holding temperatures of 19, 16 and 12 °C and under the experimental light conditions for each population. Abundance, length and sex ratio of these replicates were used as a baseline to calculate survivorship and RGR and sex ratio relative to the starting values for each population (see below). Temperatures in the water baths were altered at a rate of 1–2.5 °C each day (24-h period) depending on the starting temperature, until the final temperatures ranging between 15 and 30.5 °C were reached across the thermal gradient block (Fig. S1). This acclimation period lasted for 7 d for each experiment. Temperatures were checked daily, and light levels monitored weekly throughout the experiment to ensure stability (Light:  $26.6 \pm 0.14 \mu mol m^{-2} s^{-1}$ , Temperature: see Fig. S1). In addition, a wireless high-resolution temperature logger was used to measure the lowest (15  $^{\circ}$ C) and highest (30.5  $^{\circ}$ C) temperature treatments every 10 min for 4 d, to ensure stability in the temperature block (see Fig. S2). For each population, gametophyte cultures were exposed to experimental temperatures for 14 d.

#### Sampling procedure and measurements

At the start of the experiment (baseline measurement) and after 14 d, the response of gametophytes to the thermal gradient was recorded. Cultures were sampled by taking a coverslip, placing it under a glass microscope slide and taking three photos randomly using a MIchrome 20 Color Microscope camera mounted on a stereo microscope (Olympus BX53) at 100× magnification. For each photo (three for each replicate), gametophyte length [averaged over ten individuals per field of view (FOV, 3.2 mm<sup>2</sup>), um] was measured and the number of individuals (both male and female), as well as number of males, females and juvenile sporophytes per FOV (three for each replicate) was counted. As starting densities and size of gametophytes in each experiment varied slightly, we calculated change relative to the starting density and size, to allow for comparison among experiments and have ecological relevance for spatial modelling. The metrics used were gametophyte relative survival, RGR and male/female ratio.

Survivorship was calculated as number of viable gametophytes (defined as containing at least one pigmented cell) compared to initial number of gametophytes established directly after settlement. RGR was calculated using the following formula (Alsuwaiyan *et al.*, 2021):

RGR 
$$(\% d^{-1}) = [(\ln L_2 - \ln L_1) / t_2 - t_1] * 100$$

where  $L_1$  and  $L_2$  represent gametophyte length (µm) directly after settlement ( $L_1$ ) and at the end of the experiment ( $L_2$ ), and t represents the number of days at the start ( $t_1$ ) and end ( $t_2$ ) of the experiment. Branch length was measured using ImageJ 1.53e software for ten haphazardly selected individuals.

Due to the destructive nature of sampling, baseline measurements  $(t_1)$  were independent of final  $(t_2)$  samples. Therefore, RGR values were capped at 0, and survival values were capped at 100.

The male:female ratio was calculated from the total numbers of male and female gametophytes surveyed from three FOV from each replicate coverslip.

#### Gametophyte model development

We applied a generalized linear modelling (GLM) framework to assess for the effects of the fixed factor treatment temperature on the survival, RGR and sex ratio of gametophytes of each population. Treatment temperatures as used in the GLM analysis were based on the measured averages (Supplementary Data Fig. S1) but were rounded up at 0.5 °C intervals. GLMs used a logistic link function to model the survival and sex ratio (i.e. binomially distributed response variables), and an identity link function for RGR (i.e. Gaussian distributed response variable). Under-dispersion was detected in the binomial GLMs, so a corrected standard error using a quasi-GLM model was implemented (Zuur *et al.*, 2009). Residual plots were assessed visually to confirm the GLMs satisfied assumptions of homogeneity of variance. Significant effects of temperature were followed by pairwise comparison between sites for every temperature point using estimated marginal means and at a significance level of alpha = 0.05.

For each population, generalized additive models (GAMs) were used to assess the effect of temperature as a continuous variable on gametophyte survival and RGR using the same link functions and distributions as described above. Smoothing functions were removed if the effective degrees of freedom were approximately equal to 1, which is indicative of approximately linear responses (Zuur *et al.*, 2009). All analyses were undertaken in R (R Core Team, 2022) using the *mgcv* package (Wood *et al.*, 2016) to fit all GAMs and GLMs. Pairwise comparisons were done using the package *emmeans* (Length, 2022), and plots were generated with the packages *ggplot2* (Wickham, 2016) and *visreg* (Breheny and Burchett, 2017).

# Spatial analyses

Historical spatial predictions and future spatial projections were made for gametophyte survival and RGR using the population-specific GAMs and GLMs that related these response variables to temperature. Sex ratio was not included in spatial analyses as no significant effect of temperature on sex ratio was identified for the Sydney population. Spatial analyses were restricted to depths shallower than 50 m, using depth data from the General Bathymetric Chart of the Oceans (GEBCO 2020) dataset. Historical predictions and future projections were created for three distinct regions using unique models fitted to data obtained from the genetically distinct E. radiata populations that occur within each region. Specifically, models fitted to thermal response data from the low-latitude Coffs Harbour population were projected to 28°S-31°S, models fitted to data from the mid-latitude Sydney population were projected to 34.5°S-31°S and models fitted to data from the high-latitude Tasmania population were projected to 41°S-44°S.

Historical predictions were made for each season and population and were averaged over a 28-year period encompassing 1994–2022. Temperature data used in the historical predictions of gametophyte survival and RGR were obtained from the reprocessed (L4) Operational SST and Ice Analysis (OSTIA) system (Good *et al.*, 2020), downloaded from the Copernicus Marine Environment Monitoring Service (https://marine.copernicus.eu; product #010\_011) and had a 0.05° spatial resolution. Daily measures of sea surface temperature (SST) were seasonally aggregated and averaged over a 28-year period (encompassing 1994–2022).

Future projections for each season and population were centred on a near-future 2050 (2040–2059 period). Temperature data used in future projections of gametophyte survival and RGR were obtained from six global climate models (GCMs) forced under RCP4.5 and 8.5 emissions scenarios from the Coupled Model Intercomparison Project (CMIP5; Table S1). Given existing variability among GCMs (Drenkard *et al.*, 2021), we used a multi-model ensemble of SST data consisting of the average of all six models to provide a robust estimate of future ocean conditions off eastern Australia. To match the spatial resolution of our historical predictions (0.05°), we used

the delta 'change-factor' method (e.g. Morley *et al.*, 2018; Navarro-Racines *et al.*, 2020; von Hammerstein *et al.*, 2022) to downscale SST from ~1° to a common 0.05° spatial resolution. Delta downscaling has the advantage of providing highresolution temperature data over decadal time periods while being relatively simple in application (Navarro-Racines *et al.*, 2020; Drenkard *et al.*, 2021). In addition, this method reduces model bias by use of high-resolution observed data to generate future temperature data, thus including empirical data on small-scale variations into the final model (Pourmokhtarian *et al.*, 2016).

The delta downscaling process involved (1) remapping the curvilinear source GCM temperature data to a global 1° rectilinear grid using the second-order conservative algorithm (remapcon2) in Climate Data Operators (Schulzweida, 2021); (2) infilling missing data adjacent to the continental coast for datasets describing zonal (U) and meridional (V) flows using thin plated splines interpolation in R (R Core Team, 2022); (3) calculating the difference (i.e. delta value) between seasonally aggregated temperature data for the period 2040-2059 and a modelled historical baseline period encompassing 1993-2012 for each GCM and RCP scenario; (4) disaggregating delta value matrices from their native model resolution  $(\sim 1^{\circ})$  to the finer resolution of observed ocean data (i.e. 0.05°) using bilinear interpolation; and (5) adding delta values to the observed seasonal temperature data encompassing the period 1993-2012. This method produced future ocean temperature data, seasonally aggregated and downscaled to a common 0.05° resolution for the period 2040-2059 which was used to generate future projections of gametophyte survival and RGR.

Finally, the historical predictions were used to calculate the projected change between historical and future periods for gametophyte survival and RGR for the three genetically distinct populations. Future projections under RCP4.5 are presented in the Supplementary Data, while projections under RCP8.5 are presented in the main text as this scenario most closely aligns with the current trajectory of climate change (Schwalm *et al.*, 2020). Spatial analyses were undertaken using the *raster* package (Hijmans and Etten, 2012) and maps were generated using the *tmap* package (Tennekes, 2018) in R (R CoreTeam, 2022).

#### RESULTS

# Effects of temperature on gametophyte survival, RGR and sex ratio on the populations

Survival varied among temperatures for each population in *E. radiata* gametophytes (Table 1; Fig. 1A). Pairwise comparison showed that the low-latitude gametophytes had a lower overall survival rate between 15 and 20.5 °C (13–37 %) compared to the survival between 22 and 27 °C (45–82 % *P* < 0.05, Fig. 1A). In contrast, mid- and high-latitude gametophytes had highest survival from 15 to 20.5 °C (79–100 %) compared to higher temperatures (*P* < 0.05, Fig. 1A). Survival was significantly lowered for low-latitude gametophytes from 29 °C (<10 %, *P* < 0.05, Fig. 1A). Mid-latitude gametophytes had a significantly lower survival from 24 °C (49 %), which was significantly lower again from 27 °C (<10 %, *P* < 0.05, Fig. 1A).

TABLE I. Results of one-way ANOVAs on the survival, RGR and sex ratio of E. radiata gametophytes using fixed factor Temperature (levels: 15, 17, 19, 20.5, 22, 24, 25.5, 27, 29, 30.5 °C) for each population (Coffs Harbour, Sydney, Tasmania). Bold printed p-values signify significance at the  $\alpha \leq 0.05$  level.

ANOVAs for each population of the effect of treatment temperature					
Population	Response variable	Factor	d.f.	F	<b>P</b> -value
Coffs Harbour	Survival	Temperature	9	11.25	<0.001
		Residuals	40		
	RGR	Temperature	9	10.97	< 0.001
		Residuals	39		
	Sex ratio	Temperature	9	22.66	< 0.001
		Residuals	37		
Sydney	Survival	Temperature	9	67.97	< 0.001
		Residuals	40		
	RGR	Temperature	9	37.55	< 0.001
		Residuals	38		
	Sex ratio	Temperature	9	0.15	0.997
		Residuals	37		
Tasmania	Survival	Temperature	9	292.69	< 0.001
		Residuals	40		
	RGR	Temperature	6	7.57	< 0.001
		Residuals	20		
	Sex ratio	Temperature	5	31.30	< 0.001
		Residuals	18		

Survival was significantly lower at temperatures above 22 °C for high-latitude gametophytes (<2.5 %, P < 0.05, Fig. 1A). At 29 and 30.5 °C, there was no survival for high-latitude gametophytes, while 1–6 % of gametophytes from low and midlatitudes survived in the two highest temperature treatments (Fig. 1A).

Differential survival among gametophyte populations resulted in unique survival curves for each population (Fig. 1B–D). For the low-latitude population, a GAM with temperature as a significant (P = 0.03) explanatory variable resulted in a bell-shaped curve with maximum survival around 23 °C, and lowest survival at either extreme of 15 and 30 °C (Fig. 1B). In contrast, smoothers were dropped in favour of a linear model with a temperature as a significant predictor for both mid-(P < 0.001, coefficient estimate: -0.7687, Fig. 1C) and highlatitude populations (P = 0.01, coefficient estimate: -2.19, Fig. 1D). Mid-latitude gametophytes had a 100 % survival rate up to 20 °C, where survival decreased gradually with temperature until 30 °C (Fig. 1C). High-latitude gametophytes survived up to 20 °C, and then survival decreased steeply to 0 % at 23 °C (Fig. 1D).

RGR of kelp gametophytes was found to vary with temperature for each population (Table 1; Fig. 2A). Low-latitude gametophytes maintained a relatively high RGR (4.5–5.5 % d<sup>-1</sup>) up to 27 °C. Pairwise comparison showed that low-latitude gametophytes had a significantly lower RGR between 29 and 30.5 °C compared to lower temperatures (P < 0.05, Fig. 2A). Despite this, low-latitude populations maintained positive RGR (0.5–1.5 % d<sup>-1</sup>) until 30.5 °C (Fig. 2A). Mid-latitude gametophytes had a lower RGR from 22 °C (2.7 % d<sup>-1</sup>) and RGR was lowest from 27 °C onward (<1 % d<sup>-1</sup>, P < 0.05, Fig. 2A).



FIG. 1. Survival of *E. radiata* gametophytes of genetically separate populations. (A) Mean percentage survival of each population over ten temperatures ranging from 15 to 30.5 °C. Error bars represent standard error (n = 5). Letters denote significant differences (one-way ANOVA for each population, post-hoc pairwise test). (B) Effect of temperature on the fitted values of the optimal model for the survival of Coffs Harbour (low-latitude) gametophytes, (C) Sydney (mid-latitude) gametophytes and (D) Tasmania (high-latitude) gametophytes. For all three model plots (B–D), grey areas denote the 95 % confidence interval and rugs (short stripes) represent the observed data.



FIG. 2. Relative growth rate (RGR,  $\% d^{-1}$ ) of *E. radiata* gametophytes of genetically separate populations. (A) Mean RGR of each population over ten temperatures ranging from 15 to 30.5 °C. Error bars represent standard error (n = 5, but from 22 °C onward the Tasmanian gametophytes are associated with fewer replicates due to death of gametophytes, as were the Coffs Harbour and Sydney gametophytes at 30.5 °C). Letters denote significant differences (one-way ANOVA for each population, post-hoc pairwise test). (B) Effect of temperature on the fitted values of the optimal model for the survival of Coffs Harbour (low-latitude) gametophytes, (C) Sydney (mid-latitude) gametophytes and (D) Tasmania (high-latitude) gametophytes. For all three model plots (B–D), grey areas denote the 95 % confidence interval and rugs (short stripes) represent the observed data.

High-latitude populations had a significantly lower RGR from 20.5 °C onward (<1 % d<sup>-1</sup>, P < 0.05, Fig. 2A). High-latitude gametophytes did not survive past 25.5 °C and therefore no RGR is shown at values higher than this.

Differential RGRs among gametophyte populations resulted in unique RGR curves for each population (Fig. 2B–D). For low-latitude gametophytes, a GAM with temperature as a significant (P < 0.001) explanatory variable revealed gametophytes had an RGR of ~5 % d<sup>-1</sup> between 15 and 22 °C, peaking with an RGR of 6 % d<sup>-1</sup> at 23 °C and then gradually declining to ~1 % d<sup>-1</sup> at 30 °C (Fig. 2B). For mid-latitude gametophytes, a GAM with temperature as a significant (P < 0.001) explanatory variable showed the highest RGR of 5 % d<sup>-1</sup> occurred at 15 °C, gradually declining to an RGR of 0 % d<sup>-1</sup> at 30 °C (Fig. 2C). For high-latitude gametophytes, a GAM with temperature as a significant (P < 0.001) explanatory variable resulted in an RGR of 4 % d<sup>-1</sup> between 15 and 17 °C and declining to 0 % d<sup>-1</sup> between 20 and 25 °C, after which no gametophytes survived (Fig. 2D).

Sex ratios of kelp gametophytes were influenced by temperature in each population (Table 1; Fig. 3A). Pairwise comparison showed a significant decrease in male gametophytes in the lowlatitude population at 29 and 30.5 °C compared to lower temperatures (P < 0.05, Fig. 3A). Mid-latitude gametophytes stayed at an approximate equal sex ratio (Fig. 3A). High-latitude gametophytes had a skew towards females at 15 °C (P < 0.05) and 19 °C (P < 0.05, Fig. 3A). High-latitude gametophyte sex ratio was significantly lower compared to to higher temperatures of 20.5–24 °C (P < 0.05, Fig. 3A), resulting in significantly fewer males occurring within high-latitude gametophytes.

Differential sex ratios resulted in separate sex ratio curves for each population (Fig. 3B–D). For all populations, smoothers

were dropped in favour of linear models with temperature as a significant linear predictor for low- (P = 0.003, coefficient estimate: -0.069, Fig. 3B) and high-latitude gametophyte sex ratio (P < 0.001, coefficient estimate: -0.264, Fig. 3D), while temperature was not a significant predictor of sex ratio for midlatitude gametophytes (Fig. 3C). Both low- and high-latitude gametophytes have an equal (0.5) sex ratio at relatively low temperatures, and the sex ratio (male:female) decreased with increasing temperatures (Fig. 3B, D). The sex ratio of 0.5 in mid-latitude gametophytes is unaffected by temperature (Fig. 3C).

#### Projected survival and RGR under future ocean warming

Projections of gametophyte survival and RGR varied seasonally and latitudinally under ocean warming. Relative to present-day conditions, survival of gametophytes is projected to decline by 20–30 % in the mid-latitude population by 2050 under RCP8.5 (Fig. 4A). Relatively lower declines in survival (0–10 %) are projected for the low-latitude populations, and no change in survival for the high-latitude populations by 2050 under this climate change scenario (Fig. 4A).

Generally, projections under RCP8.5 and RCP4.5 were consistent, but the magnitude and direction of changes were intensified under RCP8.5 (Figs. 5 and 6; Supplementary Data Figs S3 and S4 present projected survival and RGR under RCP4.5, respectively). Overall survival rates of the lower latitude population (28°S–31°S) were projected to range between 70 and 40 % in 2050 under RCP8.5 (Fig. 5A), which was lower than for the mid- and high-latitude populations (Fig. 5B, C). Lowest



FIG. 3. Sex ratio (male:female) of *E. radiata* gametophytes of genetically separate populations. (A) Mean sex ratio of each population over ten temperatures ranging from 15 to  $30.5^{\circ}$ . Error bars represent standard error (n = 5, but from 22 °C onward the Tasmanian gametophytes are associated with fewer replicates due to death of gametophytes, as were the Coffs Harbour and Sydney gametophytes at  $30.5^{\circ}$ C). Letters denote significant differences (one-way ANOVA for each population, post-hoc pairwise test). (B) Effect of temperature on the fitted values of the optimal model for the sex ratio of Coffs Harbour (low-latitude) gametophytes and (D) Tasmania (high-latitude) gametophytes. The red dotted line denotes a 0.5 sex ratio, where there is an equal number of males and females. For all three model plots (B–D), grey areas denote the 95 % confidence interval and rugs (short stripes) represent the observed data.

survival rates in the low-latitude population were projected in summer (~40 %), and this was intensified under RCP8.5 (Fig. 5A; Fig. S3). The highest survival rates in the low-latitude population were projected in spring under RCP8.5, with up to 70 % survival (Fig. 5A). The mid-latitude populations ( $31^{\circ}S-34.5^{\circ}S$ ) had a broader projected range of survival rates under RCP8.5, between 20 and 100 % (Fig. 5B), as well as more pronounced latitudinal differences in projected survival compared to the other two populations (Fig. 5). Stark seasonal differences in projected survival became apparent in the mid-lattitude population under RCP8.5, where summer survival was projected to be as low as 20 %, while survival was projected between 80 and 100 % in spring under RCP8.5 (Fig. 5B). The high-latitude populations ( $41.5^{\circ}S-44^{\circ}S$ ) had a projected survival rate of 100 % across seasons and latitude under RCP8.5 (Fig. 5C).

RGR was projected to be higher within the lower latitude population (28°S–31°S; 4–6 % d<sup>-1</sup>) over all seasons, compared to the other populations by 2050 under RCP8.5 (Fig. 6). Relative to present day levels, RGR was projected to decline between 1.8 and 1.2 % d<sup>-1</sup> in the mid-latitude population by 2050 under RCP8.5 (Fig. 4B). Relatively lower declines in RGR (0–0.5 % d<sup>-1</sup>) are projected for the low-latitude populations (Fig. 4B). An increase in RGR of up to 1.2 % d<sup>-1</sup> was projected for the high-latitude population under RCP8.5 in the southern regions, while a small decrease in RGR (>0.5 % d<sup>-1</sup>) was projected in the northern regions of that population under the same ocean warming scenario (Fig. 4B).

The low-latitude population had the lowest projected RGR in summer (4 % d<sup>-1</sup>), which was slightly more pronounced under RCP8.5 (Fig. 6A; Supplementary Data Fig. S4). The midlatitude population (31°S–34.5°S) also had lower projected RGRs in summer under RCP8.5 with RGR of near 0 % d<sup>-1</sup> in the northmost regions (Fig. 6B). RGRs were projected to be highest in spring for the mid-latitude population under RCP8.5, between 4 and 5 % d<sup>-1</sup> (Fig. 6B). The high-latitude population (41.5°S–44°S) had the lowest projected RGRs under RCP8.5 compared to the other two populations (Fig. 6), ranging between 1 and 3 % d<sup>-1</sup> (Fig. 6C). The starkest contrast between RCPs was projected in this population for autumn, where a relatively high RGR was projected under RCP4.5 (3 % d<sup>-1</sup>, Fig. S3), but under RCP8.5, RGR was projected to be nearly 0 % d<sup>-1</sup> (Fig. 6C). Winter had high projected RGRs under RCP8.5 for the high-latitude population (4 % d<sup>-1</sup>), while a latitudinal difference in projected RGR became apparent in summer, with lower projected RGRs in the northern ranges of the population (Fig. 6C).

# DISCUSSION

Kelp forests are projected to decline as ocean warming increases (Wernberg *et al.*, 2019*b*). However, these projections fail to consider the microscopic gametophyte stage of kelps, which often has a higher thermal tolerance than adult plants (tom Dieck, 1993; Veenhof *et al.*, 2022*a*), or spatial variation in kelp thermal tolerance. Here, for the first time, we used laboratory-derived thermal performance data for *E. radiata* gametophytes to model and project changes in the survival and RGR of *E. radiata* gametophytes along eastern Australia.

Many kelp gametophyte populations show variability in their thermal response, which can suggest adaptation to local temperatures (Mabin *et al.*, 2019; Muth *et al.*, 2019; Liesner *et al.*, 2020; Alsuwaiyan *et al.*, 2021). This ability to adapt to



FIG. 4. Spatial projections of the change in (A) survival (left panels) and (B) RGR (% d<sup>-1</sup>, right panels) between the historical baseline (encompassing monthly averaged aggregated to annual averages from 1994 to 2022) and future climate scenario RCP8.5 centred around 2050-future (2040–2059 period). The three populations are low latitude (Coffs Harbour, 28°S–31°S), mid-latitude (Sydney, 31°S–34.5°S) and high latitude (Tasmania, 41.5°S–44°S).

local temperatures ensures kelps can thrive in a broad range of temperature conditions (Müller *et al.*, 2008; King *et al.*, 2018). We found evidence for thermal plasticity among populations of *E. radiata* gametophytes, which was related to local temperatures. Survival was higher for the low-latitude gametophytes in the warmer temperature treatments (22.5–27 °C), reflecting

the temperatures this population experiences in the field (17–27 °C; Wijffels *et al.*, 2018). Similarly, the upper survival and RGR limit for mid- and high-latitude gametophytes was 25.5 and 20.5 °C, respectively, which was congruent with local temperature ranges of 15–23 °C for the mid-latitudes and 11–18 °C for high latitudes (Wijffels *et al.*, 2018). Thermal plasticity



Fig. 5. Survival of *E. radiata* gametophytes among three separate populations for a 2050-centred future period (2040–2059) under climate scenario RCP8.5. The three populations are (A) low latitude (Coffs Harbour, 28°S–31°S), (B) mid-latitude (Sydney, 31°S–34.5°S) and (C) high latitude (Tasmania, 41.5°S–44°S). Monthly projections have been seasonally aggregated (Winter = June to August, Spring = September to November, Summer = December to February, Autumn = March to May).



FIG. 6. Relative growth rate (RGR, % d<sup>-1</sup>) of *E. radiata* gametophytes among three separate populations for a 2050-centred future period (2040–2059) under climate scenario RCP8.5. The three populations are (A) low latitude (Coffs Harbour, 28°S–31°S), (B) mid-latitude (Sydney, 31°S-34.5°S) and (C) high latitude (Tasmania, 41.5°S–44°S). Monthly projections have been seasonally aggregated (Winter = June to August, Spring = September to November, Summer = December to February, Autumn = March to May).

in accordance with local temperatures among geographically distinct populations of gametophytes has also been found on the west coast of Australia in *E. radiata* (Mohring *et al.*, 2014), and among populations in *Macrocystis pyrifera* (Hollarsmith *et al.*, 2020) and *Laminaria digitata* (Liesner *et al.*, 2020, Martins *et al.*, 2020). As we used different holding temperatures (*in situ* winter temperatures) for each gametophyte population to have ecologically realistic data for modelling purposes, we cannot untangle the influence of recent environmental history from an underlying genetic basis in the thermal tolerance expressed in this experiment. However, genotype × environment interactions have been found in *E. radiata* (Mabin *et al.*, 2019, Alsuwaiyan *et al.*, 2021) and hence our results are likely due to both phenotypic plasticity and underlying genetic variation related to thermal traits.

Ecklonia radiata gametophytes from the low-latitude population had the highest thermal tolerance recorded for kelp gametophytes in any study (maintaining positive RGRs up to 30.5 °C), akin to warm-water-adapted kelp species (monthly means above 15 °C in summer; Veenhof et al., 2022a) Eisenia bicyclis and Undaria pinnatifida (tom Dieck, 1993). Notably, each population had a thermal tolerance range that exceeds their maximum local temperature by ~2-3 °C, meaning none of the populations are nearing their thermal maxima within their current distributions. Thermally tolerant gametophytes are common in warm-adapted kelp species, such as Ecklonia radicosa (Komazawa et al., 2015), where maximum survival temperatures are higher than current in situ temperatures (tom Dieck, 1993, Veenhof et al., 2022a). This thermal tolerance may play an important role in resilience to future warming, though some southern hemisphere gametophytes are at their current thermal limit (Paine et al., 2021). Despite the thermal resilience of gametophyte survival and growth, the transition from gametophyte to sporophyte stages can be more sensitive to temperature (Martins et al., 2020; Veenhof et al., 2022a).

The proportion of males to females in gametophyte populations can vary depending on environmental conditions (Bolton and Lüning, 1982; Oppliger et al., 2011; Martins et al., 2020; Veenhof et al., 2022a). A skewed sex ratio is often caused by post-germination mortality of either males or females in response to environmental stress such as light and temperature (Xu et al., 2015; Martins et al., 2020). Ecklonia radiata gametophytes in this study skewed sex ratios to favour more females when temperatures approached the population's thermal maximum, but only for the two range-edge populations. Increased survival of females under increased temperature at range edges has also been observed in Lessonia nigrescens (Oppliger et al., 2011) and Macrocystis pyrifera (Rodriguez et al., 2019). This contrasts with other, non-range-edge populations, where survival of males is higher under temperature stress (e.g. Bolton and Lüning, 1982; Komazawa et al., 2015; Martins et al., 2020). An increased number of females at lower latitudes may help increase fecundity, as the number and size of eggs per female can decrease in low-latitude gametophyte populations (Camus et al., 2021) as well as with increased temperature stress (Müller et al., 2008; Martins et al., 2017; Hollarsmith et al., 2020). An increased number of females may also result in parthenogenesis, asexual reproduction through female gametophytes, which can occur in range edges in response to marginal conditions (Oppliger et al., 2014).

Incorporating thermal tolerances of different populations can considerably improve projections of future survival for kelps (King et al., 2018). While the projected survival for the lowlatitude gametophyte population ranged between 30 and 70 %, their projected RGRs were the highest among populations. Conversely, high-latitude populations had the lowest projected RGRs, down to 0 in autumn. This may indicate separate tradeoffs between survival and growth for lower and higher latitude populations. Low survival but high RGRs at higher temperatures cause fewer, but larger gametophytes in the low-latitude populations, and high survival but low RGRs at higher temperatures cause a greater number of smaller gametophytes for the high-latitude population. A similar trade-off to that of the low-latitude populations under increased temperatures (lower survival but increased growth) has been observed for E. radiata gametophytes across Western Australia (Mohring et al., 2013). Larger female gametophytes produce more eggs per female (Muñoz et al., 2004; Camus et al., 2021), and this may compensate for reduced fertility resulting from higher mortality and lowered fecundity in the low-latitude population (Camus et al., 2021). The timing of spore supply may also compensate for the lowered projected survival in summer for mid- and low-latitude populations. Spore supply peaks in autumn in mid-latitude populations (Mabin et al., 2013) and spore supply is available year-round in high-latitude populations (Veenhof et al., 2023). Any gametophyte die-off during hotter summer months can thus be replenished with fresh spores in autumn. Spore supply also peaks in autumn for high-latitude populations (Tatsumi et al., 2021), meaning under an RCP8.5 2050 future RGRs will be lowest when the highest spore release occurs. However, fertility peaks may also shift due to ocean warming and may result in a mismatch between peak spore supply and optimum gametophyte survival and RGR (Martins et al., 2017).

Models using data on adult E. radiata sporophytes have projected a range contraction of ~275 km in the area that corresponds with our low- and mid-latitude population by 2100 under RCP8.5 (Davis et al., 2022). Projections based on gametophyte models revealed minimal changes in survival compared to historical predictions. In addition, thermal adaptation of gametophytes may lead to higher survival than the projected survival rates, making even these minimal projections conservative (Mabin et al., 2019; Becheler et al., 2022). The midlatitude population was identified as the most vulnerable, with a projected 30 % decrease in survival. This is due to a relatively lower thermal tolerance of that population compared to the low-latitude population, while some parts of the mid-latitude population would experience temperature increases similar to the low-latitude population. However, the low-latitude population had a projected 0 % change in survival, and high connectivity among E. radiata populations along eastern Australia (Coleman *et al.*, 2011*a*, *b*, 2017) may result in more thermally tolerant gametophytes migrating to higher latitudes, potentially increasing projected survival rates of the mid-latitude population.

While sporophyte populations are projected to decline, gametophyte populations may act as a buffer during periods of high temperature stress such as summers and marine heatwaves and facilitate rapid recovery. Gametophytes can persist in the form of vegetatively growing, mixed-origin seedbanks persisting through temperature extremes of up to 27–30 °C in the lower latitudes (*sensu* Carney *et al.*, 2013). The exact temperature window for transition to sporophytes is not known for the separate populations, though it is likely between 16 and 20 °C in high-latitude populations (from 34°S and higher; Novaczek, 1984b; Mabin *et al.*, 2013; Mohring *et al.*, 2014; Alsuwaiyan *et al.*, 2021). Recruitment patterns may shift in *Ecklonia* during the hot summer months when sporophyte loss is more likely to occur, but surviving gametophytes may facilitate recruitment and repopulation in winter and spring. Integrating the exact temperature thresholds of recruitment (transition from gametophyte into sporophyte) across populations into kelp models may more accurately project future distributions of kelp populations (Assis *et al.*, 2017; Paine *et al.*, 2021), which is an important avenue of future research.

# CONCLUSION

Kelp forest persistence is threatened by ocean warming globally. Here we reveal, via population-specific gametophyte models, that E. radiata gametophytes have thermal tolerances in accordance with their local temperatures across 13° of latitude along the east coast of Australia. They also have a thermal tolerance of ~2-3 °C above their local thermal maxima, indicating that the gametophyte stage of E. radiata is likely to be resilient to near future ocean warming and may provide population resilience as a thermally tolerant seedbank. However, mismatches between peak spore supply and optimum survival may occur under future warming scenarios. It will also be crucial to pinpoint the exact temperatures of stage transitions from gametophyte to sporophyte to fully understand the effects of ocean warming on kelp persistence. Projections of future kelp loss should consider local thermal adaptation, as well as the thermal resilience of the gametophyte stage to provide more accurate outcomes.

# SUPPLEMENTARY DATA

Supplementary data are available online at https://academic. oup.com/aob and consist of the following.

Figure S1. Treatment temperatures during the thermal experiment for populations Coffs Harbour, Sydney and Tasmania. Figure S2. Temperature logger data recording temperature every 10 min for the 15 and 30.5 °C treatment for 4 d. Figure S3. Survival of *E. radiata* gametophytes among three separate populations for a 2050-centred future period under climate scenario RCP4.5. Figure S4. Relative growth rate of *E. radiata* gametophytes among three separate populations for a 2050-centred future period under climate scenario RCP4.5. Table S1. Details of GCMs (CMIP5) downscaled (0.05°) to support projections gametophyte survival and RGR off eastern Australia. The downscaled variable from each model include was sea surface temperature under RCP4.5 and 8.5 emissions scenarios

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#### LITERATURE CITED

- Alsuwaiyan NA, Vranken S, Filbee-Dexter K, Cambridge M, Coleman MA, Wernberg T. 2021. Genotypic variation in response to extreme events may facilitate kelp adaptation under future climates. *Marine Ecology Progress Series* 672: 111–121. doi:10.3354/meps13802.
- Arafeh-Dalmau N, Montaño-Moctezuma G, Martínez JA, Beas-Luna R, Schoeman DS, Torres-Moye G. 2019. Extreme marine heatwaves alter kelp forest community near its equatorward distribution limit. Frontiers in Marine Science 6: 20190801.
- Assis J, Berecibar E, Claro B, et al. 2017. Major shifts at the range edge of marine forests: the combined effects of climate changes and limited dispersal. *Scientific Reports* 7: 44348. doi:10.1038/srep44348.
- Assis J, Araújo MB, Serrão EA. 2018. Projected climate changes threaten ancient refugia of kelp forests in the North Atlantic. *Global Change Biology* 24: e55–e66. doi:10.1111/gcb.13818.
- Barradas A, Alberto F, Engelen AH, Serrao EA. 2011. Fast sporophyte replacement after removal suggests banks of latent microscopic stages of *Laminaria ochroleuca* (Phaeophyceae) in tide pools in northern Portugal. *Cahiers de Biologie Marine* 52: 435–439.
- Bartsch I, Wiencke C, Laepple T. 2012. Global seaweed biogeography under a changing climate: the prospected effects of temperature. In: *Seaweed biology*. Berlin: Springer, 383–406.
- Bartsch I, Vogt J, Pehke C, Hanelt D. 2013. Prevailing sea surface temperatures inhibit summer reproduction of the kelp *Laminaria digitata* at Helgoland (North Sea). *Journal of Phycology* 49: 1061–1073. doi:10.1111/jpy.12125.
- Bates AE, Pecl GT, Frusher S, et al. 2014. Defining and observing stages of climate-mediated range shifts in marine systems. *Global Environmental Change* 26: 27–38. doi:10.1016/j.gloenvcha.2014.03.009.
- Becheler R, Haverbeck D, Clerc C, *et al.* 2022. Variation in thermal tolerance of the giant kelp's gametophytes: suitability of habitat, population quality or local adaptation? *Frontiers in Marine Science* 9: 802535.
- Bekkby T, Moy FE. 2011. Developing spatial models of sugar kelp (Saccharina latissima) potential distribution under natural conditions and areas of its disappearance in Skagerrak. Estuarine, Coastal and Shelf Science 95: 477–483. doi:10.1016/j.ecss.2011.10.029.
- Berry HD, Mumford TF, Christiaen B, *et al.* 2021. Long-term changes in kelp forests in an inner basin of the Salish Sea. *PLoS One* 16: e0229703. doi:10.1371/journal.pone.0229703.
- Bolton JJ, Lüning K. 1982. Optimal growth and maximal survival temperatures of Atlantic Laminaria species (Phaeophyta) in culture. *Marine Biology* 66: 89–94.
- Boyce DG, Frank KT, Worm B, Leggett WC. 2015. Spatial patterns and predictors of trophic control in marine ecosystems. *Ecology Letters* 18: 1001–1011. doi:10.1111/ele.12481.
- Breheny P, Burchett W. 2017. Visualization of regression models using visreg. *The R Journal* 9: 56–71. doi:10.32614/rj-2017-046.
- Camus C, Solas M, Martínez C, et al. 2021. Mates matter: gametophyte kinship recognition and inbreeding in the giant kelp, *Macrocystis pyrifera* (Laminariales, Phaeophyceae). Journal of Phycology 57: 711–725. doi:10.1111/jpy.13146.
- Capdevila P, Hereu B, Salguero-Gómez R, et al. 2019. Warming impacts on early life stages increase the vulnerability and delay the population

recovery of a long-lived habitat-forming macroalga. *Journal of Ecology* **107**: 1129–1140. doi:10.1111/1365-2745.13090.

- Carney LT, Bohonak AJ, Edwards MS, Alberto F. 2013. Genetic and experimental evidence for a mixed-age, mixed-origin bank of kelp microscopic stages in southern California. *Ecology* 94: 1955–1965. doi:10.1890/13-0250.1.
- Castorani MCN, Reed DC, Miller RJ. 2018. Loss of foundation species: disturbance frequency outweighs severity in structuring kelp forest communities. *Ecology* 99: 2442–2454. doi:10.1002/ecy.2485.
- Chaudhary C, Richardson AJ, Schoeman DS, Costello MJ. 2021. Global warming is causing a more pronounced dip in marine species richness around the equator. *Proceedings of the National Academy of Sciences* 118: e2015094118.
- Coleman MA, Chambers J, Knott NA, et al. 2011a. Connectivity within and among a network of temperate marine reserves. PLoS One 6: e20168. doi:10.1371/journal.pone.0020168.
- Coleman MA, Roughan M, Macdonald HS, et al. 2011b. Variation in the strength of continental boundary currents determines continentwide connectivity in kelp. Journal of Ecology 99: 1026–1032. doi:10.1111/j.1365-2745.2011.01822.x.
- Coleman MA, Cetina-Heredia P, Roughan M, Feng M, van Sebille E, Kelaher BP. 2017. Anticipating changes to future connectivity within a network of marine protected areas. *Global Change Biology* 23: 3533– 3542. doi:10.1111/gcb.13634.
- Davis TR, Champion C, Coleman MA. 2021. Climate refugia for kelp within an ocean warming hotspot revealed by stacked species distribution modelling. *Marine Environmental Research* 166: 105267. doi:10.1016/j. marenvres.2021.105267.
- Davis TR, Champion C, Coleman MA. 2022. Ecological interactions mediate projected loss of kelp biomass under climate change. *Diversity and Distributions* 28: 306–317. doi:10.1111/ddi.13462.
- Dayton PK. 1985. Ecology of kelp communities. Annual Review of Ecology and Systematics 16: 215–245. doi:10.1146/annurev. es.16.110185.001243.
- de Bettignies T, Wernberg T, Gurgel CFD. 2018. Exploring the influence of temperature on aspects of the reproductive phenology of temperate seaweeds. *Frontiers in Marine Science* 5: 218.
- Doney SC, Ruckelshaus M, Emmett Duffy J, et al. 2011. Climate change impacts on marine ecosystems. Annual Review of Marine Science 4: 11–37. doi:10.1146/annurev-marine-041911-111611.
- Drenkard EJ, Stock C, Ross AC, et al. 2021. Next-generation regional ocean projections for living marine resource management in a changing climate. *ICES Journal of Marine Science* 78: 1969–1987. doi:10.1093/icesjms/ fsab100.
- Eger A, Marzinelli E, Baes R, et al. 2023. The value of ecosystem services in global marine kelp forests. *Nature communications* 14: 1894. doi:10.1038/ s41467-023-37385-0.
- Filbee-Dexter K, Feehan CJ, Scheibling RE. 2016. Large-scale degradation of a kelp ecosystem in an ocean warming hotspot. *Marine Ecology Progress Series* 543: 141–152. doi:10.3354/meps11554.
- Foden WB, Young BE, Akçakaya HR, et al. 2019. Climate change vulnerability assessment of species. WIREs Climate Change 10: e551.
- Franco JN, Tuya F, Bertocci I, et al. 2018. The 'golden kelp' Laminaria ochroleuca under global change: Integrating multiple eco-physiological responses with species distribution models. Journal of Ecology 106: 47–58.
- Fredston-Hermann A, Selden R, Pinsky M, Gaines SD, Halpern BS. 2020. Cold range edges of marine fishes track climate change better than warm edges. *Global Change Biology* 26: 2908–2922. doi:10.1111/gcb.15035.
- Gabara SS, Konar BH, Edwards MS. 2021. Biodiversity loss leads to reductions in community-wide trophic complexity. *Ecosphere* 12: e03361.
- Gissi E, Manea E, Mazaris AD, et al. 2021. A review of the combined effects of climate change and other local human stressors on the marine environment. Science of the Total Environment 755: 142564. doi:10.1016/j. scitotenv.2020.142564.
- Good S, Fiedler E, Mao C, et al. 2020. The current configuration of the OSTIA system for operational production of foundation sea surface temperature and ice concentration analyses. *Remote Sensing* 12: 720. doi:10.3390/ rs12040720.
- Hampe A, Petit RJ. 2005. Conserving biodiversity under climate change: the rear edge matters. *Ecology Letters* 8: 461–467. doi:10.1111/j.1461-0248.2005.00739.x.

- Harley CDG, Randall Hughes A, Hultgren KM, et al. 2006. The impacts of climate change in coastal marine systems. *Ecology Letters* 9: 228–241. doi:10.1111/j.1461-0248.2005.00871.x.
- Harley CDG, Anderson KM, Demes KW, et al. 2012. Effects of climate change on global seaweed communities. Journal of Phycology 48: 1064– 1078. doi:10.1111/j.1529-8817.2012.01224.x.
- Hijmans RJ, Etten JV. 2012. raster: Geographic Analysis and Modeling with Raster Data. R package version 2.0-12.
- Hollarsmith JA, Buschmann AH, Camus C, Grosholz ED. 2020. Varying reproductive success under ocean warming and acidification across giant kelp (*Macrocystis pyrifera*) populations. *Journal of Experimental Marine Biology and Ecology* 522: 151247. doi:10.1016/j. jembe.2019.151247.
- Johnson CR, Banks SC, Barrett NS, et al. 2011. Climate change cascades: shifts in oceanography, species' ranges and subtidal marine community dynamics in eastern Tasmania. *Journal of Experimental Marine Biology* and Ecology 400: 17–32. doi:10.1016/j.jembe.2011.02.032.
- Kearney M, Porter W. 2009. Mechanistic niche modelling: combining physiological and spatial data to predict species' ranges. *Ecology Letters* 12: 334–350. doi:10.1111/j.1461-0248.2008.01277.x.
- King NG, McKeown NJ, Smale DA, Moore PJ. 2018. The importance of phenotypic plasticity and local adaptation in driving intraspecific variability in thermal niches of marine macrophytes. *Ecography* 41: 1469– 1484. doi:10.1111/ecog.03186.
- King NG, McKeown NJ, Smale DA, et al. 2019. Evidence for different thermal ecotypes in range centre and trailing edge kelp populations. *Journal of Experimental Marine Biology and Ecology* 514-515: 10–17. doi:10.1016/j.jembe.2019.03.004.
- Komazawa I, Sakanishi Y, Tanaka J. 2015. Temperature requirements for growth and maturation of the warm temperate kelp *Eckloniopsis radicosa* (Laminariales, Phaeophyta). *Phycological Research* 63: 64–71.
- Layton C, Cameron MJ, Tatsumi M, Shelamoff V, Wright JT, Johnson CR. 2020. Habitat fragmentation causes collapse of kelp recruitment. *Marine Ecology Progress Series* 648: 111–123. doi:10.3354/meps13422.
- Length RV. 2022. emmeans: Estimated Marginal Means, aka Least-Squares Means. R package version 1.7.2.
- Liesner D, Shama LN, Diehl N, Valentin K, Bartsch I. 2020. Thermal plasticity of the kelp *Laminaria digitata* (Phaeophyceae) across life cycle stages reveals the importance of cold seasons for marine forests. *Frontiers* in Marine Science 7: 456.
- Liesner D, Pearson GA, Bartsch I, et al. 2022. Increased heat resilience of intraspecific outbred compared to inbred lineages in the kelp Laminaria digitata: physiology and transcriptomics. Frontiers in Marine Science 9: 838793.
- Mabin CJ, Gribben PE, Fischer A, Wright JT. 2013. Variation in the morphology, reproduction and development of the habitat-forming kelp *Ecklonia* radiata with changing temperature and nutrients. *Marine Ecology* Progress Series 483: 117–131. doi:10.3354/meps10261.
- Mabin CJT, Johnson CR, Wright JT. 2019. Family-level variation in early life-cycle traits of kelp. *Journal of Phycology* 55: 380–392. doi:10.1111/ jpy.12820.
- Martínez B, Radford B, Thomsen MS, et al. 2018. Distribution models predict large contractions of habitat-forming seaweeds in response to ocean warming. Diversity and Distributions 24: 1350–1366.
- Martins N, Tanttu H, Pearson GA, Serrão EA, Bartsch I. 2017. Interactions of daylength, temperature and nutrients affect thresholds for life stage transitions in the kelp *Laminaria digitata* (Phaeophyceae). *Botanica Marina* 60: 109–121.
- Martins N, Pearson GA, Bernard J, Serrão EA, Bartsch I. 2020. Thermal traits for reproduction and recruitment differ between Arctic and Atlantic kelp *Laminaria digitata*. *PLoS One* 15: e0235388. doi:10.1371/journal. pone.0235388.
- Mohring MB, Kendrick GA, Wernberg T, Rule MJ, Vanderklift MA. 2013. Environmental influences on kelp performance across the reproductive period: an ecological trade-off between gametophyte survival and growth? *PLoS One* 8: e65310. doi:10.1371/journal.pone.0065310.
- Mohring MB, Wernberg T, Wright JT, Connell SD, Russell BD. 2014. Biogeographic variation in temperature drives performance of kelp gametophytes during warming. *Marine Ecology Progress Series* 513: 85–96. doi:10.3354/meps10916.
- Morley JW, Selden RL, Latour RJ, Frölicher TL, Seagraves RJ, Pinsky ML. 2018. Projecting shifts in thermal habitat for 686 species on the

North American continental shelf. *PLoS One* **13**: e0196127. doi:10.1371/journal.pone.0196127.

- Mota CF, Engelen AH, Serrao EA, et al. 2018. Differentiation in fitnessrelated traits in response to elevated temperatures between leading and trailing edge populations of marine macrophytes. PLoS One 13: e0203666. doi:10.1371/journal.pone.0203666.
- Moy FE, Christie H. 2012. Large-scale shift from sugar kelp (Saccharina latissima) to ephemeral algae along the south and west coast of Norway. Marine Biology Research 8: 309–321. doi:10.1080/17451000.2011.6375 61.
- Müller R, Wiencke C, Bischof K. 2008. Interactive effects of UV radiation and temperature on microstages of Laminariales (Phaeophyceae) from the Arctic and North Sea. *Climate Research* 37: 203–213. doi:10.3354/ cr00762.
- Muñoz V, Hernandez-Gonzalez MC, Buschmann AH, Graham MH, Vasquez JA. 2004. Variability in per capita oogonia and sporophyte production from giant kelp gametophytes (*Macrocystis pyrifera*, Phaeophyceae). *Revista Chilena de Historia Natural* 77: 639–647.
- Murphy JT, Johnson MP, Viard F. 2016. A modelling approach to explore the critical environmental parameters influencing the growth and establishment of the invasive seaweed Undaria pinnatifida in Europe. Journal of Theoretical Biology 396: 105–115. doi:10.1016/j.jtbi.2016.01.038.
- Murphy JT, Johnson MP, Viard F. 2017. A theoretical examination of environmental effects on the life cycle schedule and range limits of the invasive seaweed Undaria pinnatifida. Biological Invasions 19: 691–702. doi:10.1007/s10530-016-1357-1.
- Muth AF, Graham MH, Lane CE, Harley CDG. 2019. Recruitment tolerance to increased temperature present across multiple kelp clades. *Ecology* 100: e02594. doi:10.1002/ecy.2594.
- Navarro-Racines C, Tarapues J, Thornton P, Jarvis A, Ramirez-Villegas J. 2020. High-resolution and bias-corrected CMIP5 projections for climate change impact assessments. *Scientific Data* 7: 1–14.
- Nicastro KR, Zardi GI, Teixeira S, Neiva J, Serrão EA, Pearson GA. 2013. Shift happens: trailing edge contraction associated with recent warming trends threatens a distinct genetic lineage in the marine macroalga *Fucus vesiculosus*. *BMC Biology* **11**: 6. doi:10.1186/1741-7007-11-6.
- **Novaczek I. 1984***a*. Response of *Ecklonia radiata* (Laminariales) to light at 15°C with reference to the field light budget at Goat Island Bay, New Zealand. *Marine Biology* **80**: 263–272. doi:10.1007/bf00392821.
- Novaczek I. 1984b. Response of gametophytes of *Ecklonia radiata* (Laminariales) to temperature in saturating light. *Marine Biology* 82: 241–245. doi:10.1007/bf00392405.
- Oppliger LV, Correa JA, Faugeron S, et al. 2011. Sex ratio variation in the Lessonia nigrescens complex (Laminariales, Phaeophyceae): effect of latitude, temperature, and marginality. Journal of Phycology 47: 5–12.
- **Oppliger LV, Correa JA, Engelen AH**, *et al.* **2012**. Temperature effects on gametophyte life-history traits and geographic distribution of two cryptic kelp species. *PLoS One* **7**: e39289. doi:10.1371/journal. pone.0039289.
- Oppliger LV, von Dassow P, Bouchemousse S, et al. 2014. Alteration of sexual reproduction and genetic diversity in the kelp species Laminaria digitata at the southern limit of its range. PLoS One 9: e102518. doi:10.1371/journal.pone.0102518.
- Paine ER, Schmid M, Gaitán-Espitia JD, et al. 2021. Narrow range of temperature and irradiance supports optimal development of *Lessonia* corrugata (Ochrophyta) gametophytes: implications for kelp aquaculture and responses to climate change. *Journal of Applied Phycology* 33: 1721–1730. doi:10.1007/s10811-021-02382-7.
- Pankhurst NW, Munday PL. 2011. Effects of climate change on fish reproduction and early life history stages. *Marine and Freshwater Research* 62: 1015–1026. doi:10.1071/mf10269.
- Parmesan C, Hanley ME. 2015. Plants and climate change: complexities and surprises. Annals of Botany 116: 849–864. doi:10.1093/aob/ mcv169.
- Pedersen P. 1981. Phaeophyta: life histories. In: Lobban CS, Wynne M. eds. *The biology of seaweeds*. Berkeley: University of California Press, 194–217.
- **Pessarrodona A, Filbee-Dexter K, Alcoverro T**, *et al.* **2021**. Homogenization and miniaturization of habitat structure in temperate marine forests. *Global Change Biology* **00**: 1–14.
- Pinsky ML, Selden RL, Kitchel ZJ. 2020. Climate-driven shifts in marine species ranges: scaling from organisms to communities. *Annual Review of Marine Science* 12: 153–179. doi:10.1146/annurev-marine-010419-010916.

- Poloczanska ES, Brown CJ, Sydeman WJ, et al. 2013. Global imprint of climate change on marine life. Nature Climate Change 3: 919–925. doi:10.1038/nclimate1958.
- Pourmokhtarian A, Driscoll CT, Campbell JL, Hayhoe K, Stoner AM. 2016. The effects of climate downscaling technique and observational data set on modeled ecological responses. *Ecological Applications* 26: 1321–1337. doi:10.1890/15-0745.
- Reeves SE, Kriegisch N, Johnson CR, Ling SD. 2022. Kelp habitat fragmentation reduces resistance to overgrazing, invasion and collapse to turf dominance. *Journal of Applied Ecology* 59: 1619–1631. doi:10.1111/1365-2664.14171.
- Rodriguez JP, Terrados J, Rosenfeld S, Mendez F, Ojeda J, Mansilla A. 2019. Effects of temperature and salinity on the reproductive phases of *Macrocystis pyrifera* (L.) C. Agardh (Phaeophyceae) in the Magellan region. *Journal of Applied Phycology* 31: 915–928.
- Russell BD, Harley CD, Wernberg T, et al. 2012. Predicting ecosystem shifts requires new approaches that integrate the effects of climate change across entire systems. *Biology Letters* 8: 20120423.
- Schiel DR, Steinbeck JR, Foster MS. 2004. Ten years of induced ocean warming causes comprehensive changes in marine benthic communities. *Ecology* 85: 1833–1839. doi:10.1890/03-3107.
- Schulzweida U. 2021. *Climate data operators user's guide*. 1.1. 9th edn. Hamburg: Max-Planck Institute for Meteorology.
- Schwalm CR, Glendon S, Duffy PB. 2020. RCP8.5 tracks cumulative CO<sub>2</sub> emissions. Proceedings of the National Academy of Sciences 117: 19656– 19657. doi:10.1073/pnas.2007117117.
- Sorte CJB, Williams SL, Carlton JT. 2010. Marine range shifts and species introductions: comparative spread rates and community impacts. *Global Ecology and Biogeography* 19: 303–316. doi:10.1111/j.1466-8238.2009.00519.x.
- Steneck RS, Graham MH, Bourque BJ, et al. 2002. Kelp forest ecosystems: biodiversity, stability, resilience and future. Environmental Conservation 29: 436–459. doi:10.1017/s0376892902000322.
- Stuart-Smith RD, Edgar GJ, Bates AE. 2017. Thermal limits to the geographic distributions of shallow-water marine species. *Nature Ecology & Evolution* 1: 1846–1852. doi:10.1038/s41559-017-0353-x.
- Sudo K, Watanabe K, Yotsukura N, Nakaoka M. 2020. Predictions of kelp distribution shifts along the northern coast of Japan. *Ecological Research* 35: 47–60. doi:10.1111/1440-1703.12053.
- Tatsumi M, Mabin CJT, Layton C, et al. 2021. Density-dependence and seasonal variation in reproductive output and sporophyte production in the kelp, Ecklonia radiata. Journal of Phycology 58: 92–104. doi:10.1111/ jpy.13214.
- R Core Team. 2022. R: a language and environment for statistical computing. Vienna: R Foundation for Statistical Computing.
- Tennekes M. 2018. tmap: thematic maps in R. Journal of Statistical Software 84: 1–39. doi:10.18637/jss.v084.i06.
- Thomsen MS, Mondardini L, Alestra T, et al. 2019. Local extinction of bull kelp (Durvillaea spp.) due to a marine heatwave. Frontiers in Marine Science 6: 20190301.
- Tittensor DP, Mora C, Jetz W, et al. 2010. Global patterns and predictors of marine biodiversity across taxa. Nature 466: 1098–1101. doi:10.1038/ nature09329.
- tom Dieck I. 1993. Temperature tolerance and survival in darkness of kelp gametophytes (Laminariales, Phaeophyta) - ecological and biogeographical implications. *Marine Ecology Progress Series* 100: 253–264.
- Twiname S, Audzijonyte A, Blanchard JL, et al. 2020. A cross-scale framework to support a mechanistic understanding and modelling of marine climate-driven species redistribution, from individuals to communities. *Ecography* 43: 1764–1778. doi:10.1111/ecog.04996.
- Veenhof RJ, Champion C, Dworjanyn SA, et al. 2022a. Kelp gametophytes in changing oceans. Oceanography and Marine Biology: An Annual Review 60: 335–372.
- Veenhof RJ, Dworjanyn SA, Champion C, Coleman MA. 2022b. Grazing and recovery of kelp gametophytes under ocean warming. *Frontiers in Marine Science* 9: 866136.
- Veenhof RJ, Champion C, Dworjanyn SA, Shalders TC, Coleman MA. 2023. Reproductive phenology of the kelp *Ecklonia radiata* at its Australian warm range edge and the influence of environmental factors. *Marine and Freshwater Research* 74: 928–940. doi:10.1071/mf22259.
- Voerman SE, Llera E, Rico JM. 2013. Climate driven changes in subtidal kelp forest communities in NW Spain. *Marine Environmental Research* 90: 119–127. doi:10.1016/j.marenvres.2013.06.006.

- von Hammerstein H, Setter RO, van Aswegen M, Currie JJ, Stack SH. 2022. High-resolution projections of global sea surface temperatures reveal critical warming in humpback whale breeding grounds. *Frontiers in Marine Science* 9: 20220501.
- Vranken S, Wernberg T, Scheben A, et al. 2021. Genotype–environment mismatch of kelp forests under climate change. *Molecular Ecology* 30: 3730–3746. doi:10.1111/mec.15993.
- Wernberg T, Bennett S, Babcock RC, et al. 2016. Climate-driven regime shift of a temperate marine ecosystem. *Science* 353: 169–172. doi:10.1126/science.aad8745.
- Wernberg T, Coleman MA, Babcock RC, et al. 2019a. Biology and ecology of the globally significant kelp Ecklonia radiata. Oceanography and Marine Biology: An Annual Review 57: 265–324.
- Wernberg T, Krumhansl K, Filbee-Dexter K, Pedersen MF. 2019b. Status and trends for the world's kelp forests. In: Sheppard C. ed. World seas: an environmental evaluation. London: Elsevier, 57–78.
- Wickham H. 2016. ggplot2: elegant graphics for data analysis. New York: Springer.

- Wijffels SE, Beggs H, Griffin C, et al. 2018. A fine spatial-scale sea surface temperature atlas of the Australian regional seas (SSTAARS): Seasonal variability and trends around Australasia and New Zealand revisited. Journal of Marine Systems 187: 156–196. doi:10.1016/j. jmarsys.2018.07.005.
- Wood SN, Pya N, Säfken B. 2016. Smoothing parameter and model selection for general smooth models. *Journal of the American Statistical Association* 111: 1548–1563. doi:10.1080/01621459.2016 .1180986.
- Wood G, Marzinelli EM, Campbell AH, Steinberg PD, Vergés A, Coleman MA. 2021. Genomic vulnerability of a dominant seaweed points to futureproofing pathways for Australia's underwater forests. *Global Change Biology* 27: 2200–2212. doi:10.1111/gcb.15534.
- Xu D, Wang DS, Li B, et al. 2015. Effects of CO<sub>2</sub> and seawater acidification on the early stages of Saccharina japonica development. Environmental Science & Technology 49: 3548–3556. doi:10.1021/es5058924.
- Zuur AF, Ieno EN, Walker NJ, Saveliev AA, Smith GM. 2009. Mixed effects models and extensions in ecology with R. New York: Springer.