



SFX-01 in hospitalised patients with community-acquired pneumonia during the COVID-19 pandemic: a double-blind, randomised, placebo-controlled trial

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Treatment with stabilised synthetic sulforaphane (SFX-01, 300 mg oral capsule) once daily for 14 days did not modulate key Nrf2 targets or improve clinical outcomes in patients hospitalised with CAP mainly due to SARS-CoV-2 infection <https://bit.ly/4280Tvb>

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Abstract

Introduction Sulforaphane can induce the transcription factor, Nrf2, promoting antioxidant and anti-inflammatory responses. In this study, hospitalised patients with community-acquired pneumonia (CAP) were treated with stabilised synthetic sulforaphane (SFX-01) to evaluate impact on clinical status and inflammation.

Methods Double-blind, randomised, placebo-controlled trial of SFX-01 (300 mg oral capsule, once daily for 14 days) conducted in Dundee, UK, between November 2020 and May 2021. Patients had radiologically confirmed CAP and CURB-65 (confusion, urea >7 mmol·L⁻¹, respiratory rate ≥30 breaths·min⁻¹, blood pressure <90 mmHg (systolic) or ≤60 mmHg (diastolic), age ≥65 years) score ≥1. The primary outcome was the seven-point World Health Organization clinical status scale at day 15. Secondary outcomes included time to clinical improvement, length of stay and mortality. Effects on Nrf2 activity and inflammation were evaluated on days 1, 8 and 15 by measurement of 45 serum cytokines and mRNA sequencing of peripheral blood leukocytes.

Results The trial was terminated prematurely due to futility with 133 patients enrolled. 65 patients were randomised to SFX-01 treatment and 68 patients to placebo. Severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) infection was the cause of CAP in 103 (77%) cases. SFX-01 treatment did not improve clinical status at day 15 (adjusted OR 0.87, 95% CI 0.41–1.83; p=0.71), time to clinical improvement (adjusted hazard ratio (aHR) 1.02, 95% CI 0.70–1.49), length of stay (aHR 0.84, 95% CI 0.56–1.26) or 28-day mortality (aHR 1.45, 95% CI 0.67–3.16). The expression of Nrf2 targets and pro-inflammatory genes, including interleukin (IL)-6, IL-1β and tumour necrosis factor-α, was not significantly changed by SFX-01 treatment. At days 8 and 15, respectively, 310 and 42 significant differentially expressed genes were identified between groups (false discovery rate adjusted p<0.05, log₂FC >1).

Conclusion SFX-01 treatment did not improve clinical status or modulate key Nrf2 targets in patients with CAP primarily due to SARS-CoV-2 infection.



Introduction

Dysregulated inflammatory responses are implicated in the pathogenesis of community-acquired pneumonia (CAP) [1], a leading cause of morbidity and mortality worldwide [2]. In particular, hyperinflammation and cytokine storm are well-established contributors to coronavirus disease 2019 (COVID-19) pneumonia [3], resulting in tissue damage and in the most severe cases, acute respiratory distress syndrome (ARDS) and death [4]. Therapies that reduce mortality in hospitalised COVID-19 patients primarily target overactive inflammatory responses, including corticosteroids, anti-interleukin (IL)-6 receptor monoclonal antibodies and Janus kinase inhibitors [5, 6, 7, 8].

Despite evidence of similar inflammatory involvement in non-COVID-19 pneumonia and ARDS [1], therapeutic advancement in CAP has been relatively neglected since the widespread introduction of antibiotics in the 1950s. Effective therapies and vaccination have dramatically reduced hospitalisation rates and mortality from COVID-19 [9, 10]; however, severe disease is persisting [6], and severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) has become one pathogen among many that can cause CAP [11]. The development of broad-spectrum anti-inflammatory therapies that are effective in CAP remains critical [12].

Oxidant–antioxidant imbalance or oxidative stress can trigger and perpetuate inflammation by activating pro-inflammatory pathways (*e.g.* NF- κ B), inducing metabolic dysfunction and driving tissue damage and cell death [13, 14, 15]. The transcription factor nuclear factor erythroid 2-related factor 2 (Nrf2) is a master regulator of antioxidant and anti-inflammatory responses. Nrf2 targets and upregulates the expression of antioxidant and cytoprotective genes encoding proteins such as NAD(P)H:quinone oxidoreductase 1 (NQO1), haem oxygenase-1 (HO-1) and glutathione-S-synthetase (GSS), which participate in reactive metabolite and oxidant detoxification [16], in addition to directly inhibiting the transcription of inflammatory cytokines including IL-6, IL-1 β and tumour necrosis factor (TNF)- α implicated in cytokine storm [17]. In a murine model of pneumococcal pneumonia, loss of Nrf2 resulted in defective bacterial clearance and increased lung injury [18]. Lung biopsies from COVID-19 patients demonstrated suppression of Nrf2 gene expression, an effect that was abrogated utilising Nrf2 inducers *in vitro*, resulting in beneficial antiviral and anti-inflammatory responses [19]. Pre-clinical research therefore supports Nrf2 activation as a target with potential benefits in both COVID-19 and non-SARS-CoV-2 pneumonia [20].

Sulforaphane is a naturally occurring potent activator of Nrf2 which acts by inhibiting the usually rapid ubiquitination and degradation of Nrf2 triggered by binding to Kelch-like ECH-associated protein 1 (Keap1) [21, 22], and has demonstrated protective effects in animal models of acute lung inflammation [23] as well as potential benefit in chronic respiratory disease [24, 25]. SFX-01 is an α -cyclodextrin-encapsulated, synthetic, stabilised sulforaphane (1-isothiocyanato-4-methyl-sulfinylbutane) formulation utilised to date in two phase I (clinicaltrials.gov identifiers NCT01948362, NCT02055716) and two phase II clinical trials (clinicaltrials.gov identifiers NCT02614742, NCT02970682), with reportedly good safety profile [26].

We hypothesised that treatment with an Nrf2 activator may improve clinical outcomes in CAP by promoting antioxidant and anti-inflammatory responses. We performed a randomised, double-blind, placebo-controlled trial of SFX-01 compared with placebo in patients hospitalised with CAP.

Methods

Trial design and patients

The STAR-COVID-19 trial (SFX-01 Treatment for Acute Respiratory Infections) was a phase II double-blind, randomised, placebo-controlled, trial conducted at Ninewells Hospital, Dundee, UK. Inclusion criteria were age ≥ 18 years, hospitalisation with CAP (defined as new radiographic infiltrate on chest radiograph or computed tomography scan < 48 h after hospitalisation) and an increased risk of mortality (CURB-65 (confusion, urea > 7 mmol·L⁻¹, respiratory rate ≥ 30 breaths·min⁻¹, blood pressure < 90 mmHg (systolic) or ≤ 60 mmHg (diastolic), age ≥ 65 years) score ≥ 1 or bilateral radiographic infiltrates) without requirement for mechanical ventilation. Patients were required to be tested for SARS-CoV-2 infection by reverse transcriptase (RT) quantitative PCR at enrolment.

Key exclusion criteria were inability to provide informed consent, hospital-acquired pneumonia, alanine aminotransferase and/or aspartate aminotransferase more than five times the upper limit of normal and stage 4 chronic kidney disease or requiring dialysis. Complete eligibility criteria are provided in the supplementary material.

Trial oversight

The trial was approved by the Scotland A research ethics committee (20/SS/0092). All patients or legal representatives provided written informed consent. An independent, external data safety monitoring committee reviewed adverse event data. The study was prospectively registered with EudraCT (identifier 2020-003486-19).

Trial procedures

Patients were screened for eligibility up to 24 h prior to randomisation and randomised within 96 h of admission to hospital for CAP. Patients were randomised to treatment with either oral SFX-01 (300-mg capsules) or placebo once daily for 14 days *via* a central web-based randomisation system (TRuST). Randomisation was stratified by pneumonia severity (CURB-65 score 0–2 *versus* 3–5). Justification of the dose used is described in detail in the supplementary material.

Patients' clinical status and safety were evaluated daily while hospitalised and on days 3, 5, 8, 11, 15 and 29 after discharge. Discharged patients continued to receive treatment at home and were invited back to the research unit on day 15 for a follow-up visit including blood sampling.

Primary and secondary outcome measures

The primary study objective was to evaluate the clinical efficacy of SFX-01 compared to placebo, on top of standard care, using the World Health Organization (WHO) seven-point ordinal scale as an outcome measure of clinical status at day 15. Secondary outcome measures included time to an improvement of one category on the WHO scale, clinical status and mean change (WHO scale and National Early Warning Score (NEWS)), time to discharge or NEWS of ≥ 2 (maintained for ≥ 24 h) whichever occurred first, oxygen-free days, duration and incidence of new oxygen use or new mechanical ventilation, and ventilator-free days from day 1 to 29, duration of hospitalisation, and 15- and 28-day mortality. Safety of SFX-01 was evaluated by cumulative incidence of adverse events, serious adverse events (SAEs) and discontinuation of treatment. Patients who discontinued study treatment were asked to remain in the study and attend study visits.

Exploratory objectives and substudy

A pre-specified substudy was performed to evaluate effects of SFX-01 on Nrf2 and the systemic immune response. Peripheral blood was collected at days 1, 8 and 15. Serum cytokines were quantified using the Olink Target 48 cytokine panel and RNA-stabilised whole blood utilised for mRNA sequencing (mRNAseq). Serum cytokine analysis was performed on the intention-to-treat (ITT) population, excluding those receiving tocilizumab due to profound effects of the treatment on cytokine levels, while gene expression analysis was performed on the per-protocol population.

Gene expression changes observed in the present study were compared to publicly available data from isolated peripheral blood mononuclear cells incubated for 24 h with or without 15 μ M L-sulforaphane (L-SFN) followed by RNAseq (Gene Expression Omnibus accession number GSE160353). Detailed methods for exploratory end-points and additional analyses are included in the supplementary material.

Statistical analyses

The study was originally intended to enrol 300 participants, with details of the power calculation shown in the supplementary material. The pre-specified futility analysis was performed by the data monitoring committee on unblinded data for the first 100 subjects. Adjudication on termination for futility used conditional power of detecting odds ratios of 2 and 1.5 given the emerging treatment effect after 100 participants. Conditional power was calculated under two scenarios: 1) the treatment effect after 100 subjects extended for the duration of the trial and 2) odds ratios of 2 and 1.5 for the remainder of the trial. Termination of the trial would be recommended if all analyses showed conditional power <20%.

Primary efficacy analyses were based on the ITT population. Safety analyses were based on a modified ITT population consisting of all participants who were randomised and received at least one dose of randomised therapy. A per-protocol analysis was performed including all participants who completed randomly assigned therapy. The primary end-point, the WHO seven-point ordinal scale, was evaluated using mixed-effects ordinal logistic regression, assuming proportional odds, adjusted for the stratifying factor of CURB-65 score as random effect. Secondary outcomes of time to event were evaluated using Cox proportional hazards regression adjusted for CURB-65 score. Number of days free from oxygen, new oxygen use, days free from ventilation, new ventilation use and adverse events between the SFX-01 and placebo groups were analysed using negative binomial regression.

Pre-specified subgroup analyses were performed for the primary outcome based on age, sex, SARS-CoV-2 positivity, detection of pathogens, and subgroups based on the WHO scale at baseline.

Serum biomarker data were analysed using a mixed-model repeated measures approach (supplementary material). For mRNAseq data, using the Wald test and Benjamini–Hochberg procedure, a false discovery rate adjusted p-value of <0.05 with a \log_2 fold-change ($\log_2\text{FC}$) of >1 or <-1 between treatment groups was considered significant. Full details of exploratory analyses are presented in the supplementary material.

Results

From 20 November 2020 to 5 May 2021, 133 participants were randomised: 65 to the SFX-01 arm and 68 to placebo (figure 1).

Baseline characteristics of the study population were well balanced (table 1). 78.5% of the SFX-01 group and 76.5% of the placebo groups were SARS-CoV-2 positive by RT-PCR. 70.8% of the SFX-01 group and 69.1% of the placebo group also received corticosteroids, predominantly dexamethasone. 18.5% received tocilizumab in the SFX-01 group and 11.8% in the placebo group (table 1).

Primary end-point

At day 15, 70.8% in the SFX-01 group and 69% in the placebo group had been discharged from hospital. One patient withdrew from the study and was excluded from the analysis due to unknown day 15 status. The adjusted odds ratio (aOR) from a proportional odds model in the ITT population was 0.87 (95% CI 0.41–1.83), indicating that SFX-01 treatment did not improve clinical status compared with placebo at day 15 ($p=0.71$) (table 2).

In the per-protocol analysis, the aOR was 0.68 (95% CI 0.29–1.62; $p=0.38$; SFX-01 group $n=43$, placebo group $n=62$). Pre-specified subgroup analyses at day 29 were consistent with the primary result (supplementary tables S2–7 and supplementary figure S1). In particular, SFX-01 treatment did not improve outcomes in patients positive for SARS-CoV-2 (aOR 0.82, 95% CI 0.31–2.20; $p=0.61$).

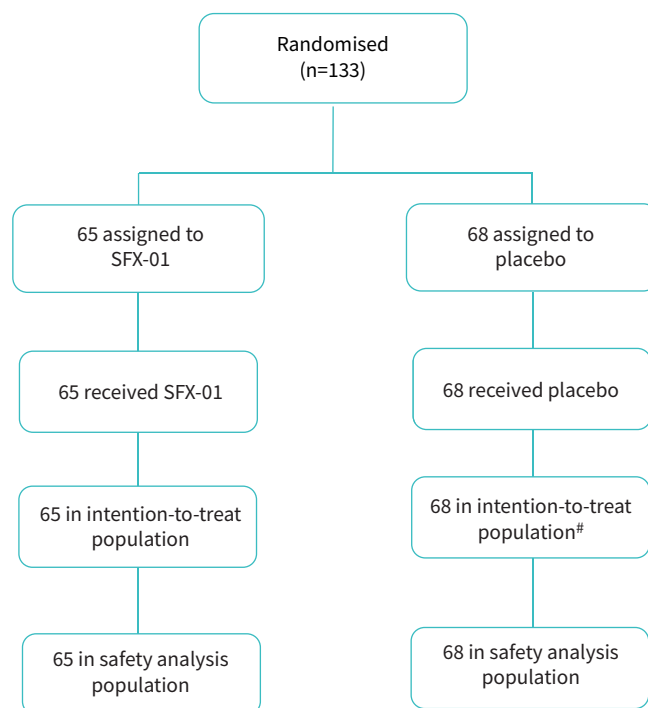


FIGURE 1 Consolidated Standards of Reporting Trials diagram detailing flow of participants in STAR-COVID-19 (SFX-01 Treatment for Acute Respiratory Infections). #: one participant in the placebo group withdrew from the study and day 15 status was unknown.

TABLE 1 Baseline study participant clinical characteristics

	Placebo	SFX-01
Participants	68	65
Gender		
Male	42 (61.8)	36 (55.4)
Female	26 (38.2)	29 (44.6)
Ethnicity		
English/Welsh/Scottish/Northern Irish/British	61 (89.7)	62 (95.4)
Indian	0	1 (1.5)
Pakistani	2 (2.9)	1 (1.5)
Chinese	2 (2.9)	0
Any other Asian background	1 (1.5)	0
African	2 (2.9)	1 (1.5)
Age years	63.6±13.8	61.6±12.7
SARS-CoV-2 PCR status		
Negative	16 (23.5)	14 (21.5)
Positive	52 (76.5)	51 (78.5)
Past medical history		
Chronic cardiac disease	16 (23.5)	10 (15.4)
Hypertension	27 (39.7)	18 (27.7)
COPD	9 (13.2)	6 (9.2)
Chronic pulmonary disease	2 (2.9)	5 (7.7)
Asthma	5 (7.4)	9 (13.8)
Chronic kidney disease (eGFR <44 mL·min ⁻¹ , on dialysis or previous transplant)	2 (2.9)	1 (1.5)
Moderate or severe liver disease	1 (1.5)	1 (1.5)
Mild liver disease	1 (1.5)	0 (0.0)
Chronic neurological disorder	1 (1.5)	3 (4.6)
Malignant neoplasm	9 (13.2)	2 (3.1)
Chronic haematological disease	3 (4.4)	4 (6.2)
Obesity	21 (30.9)	16 (24.6)
Diabetes with complications	12 (17.6)	8 (12.3)
Diabetes without complications	6 (8.8)	2 (3.1)
Rheumatological disorder	5 (7.4)	2 (3.1)
Smoking status		
Current	4 (5.9)	4 (6.2)
Former	34 (50.0)	40 (61.5)
Never	30 (44.1)	21 (32.3)
CURB-65 score		
<3	64 (94.1)	62 (95.4)
3–5	4 (5.9)	3 (4.6)
Seven-point WHO ordinal scale[#]		
3	28 (41.2)	24 (36.9)
4	33 (48.5)	30 (46.2)
5	7 (10.3)	11 (16.9)
NEWS	3.78±2.28	4.74±2.56
Median (interquartile range)	4.0 (2.0–6.0)	5.0 (3.0–7.0)
Range	0.0–8.0	0.0–11.0

Data are presented as n, n (%) or mean±sd, unless otherwise stated. SFX-01: 1-isothiocyanato-4-methylsulfanylbutane; SARS-CoV-2: severe acute respiratory syndrome coronavirus 2; eGFR: estimated glomerular filtration rate; CURB-65: confusion, urea >7 mmol·L⁻¹, respiratory rate ≥30 breaths·min⁻¹, blood pressure <90 mmHg (systolic) or ≤60 mmHg (diastolic), age ≥65 years; WHO: World Health Organization; NEWS: National Early Warning Score. [#]: 3=hospitalised, not requiring supplemental oxygen, 4=hospitalised, requiring supplemental oxygen, 5=hospitalised, on noninvasive ventilation or high-flow oxygen devices.

Secondary end-points

There were 26 deaths during the 28-day study period; 11 in the placebo group and 15 in the SFX-01 group (adjusted hazard ratio (aHR) 1.45, 95% CI 0.67–3.16; p=0.35). 19 deaths occurred on or before day 15; eight in the placebo arm and 11 in the SFX-01 arm (aHR 1.46, 95% CI 0.59–3.62; p=0.42) (table 3).

Improvement in the WHO seven-point scale by at least one category over 29 days was seen in 78.5% of the SFX-01 group and 83.8% of the placebo group. Time to clinical improvement of one category, time to

TABLE 2 Estimates of treatment effect on the primary outcome World Health Organization Clinical Status Scale at day 15

	Placebo	SFX-01	Model	OR (95% CI)	p-value
Participants	68	65			
Clinical status					
Not hospitalised, no limitations on activities	3 (4.4)	0	Unadjusted	0.84 (0.41–1.71)	0.62
Not hospitalised, limitations on activities	44 (64.7)	46 (70.8)	Adjusted [#]	0.87 (0.41–1.83)	0.71
Hospitalised, not requiring supplemental oxygen	5 (7.4)	4 (6.2)			
Hospitalised, requiring supplemental oxygen	4 (5.9)	1 (1.5)			
Hospitalised, on noninvasive ventilation or high-flow oxygen devices	2 (2.9)	3 (4.6)			
Hospitalised, on invasive mechanical ventilation or ECMO	1 (1.5)	0			
Death	8 (11.8)	11 (16.9)			
Missing	1 (1.5)	0			

Data are presented as n or n (%), unless otherwise stated. SFX-01: 1-isothiocyanato-4-methyl-sulfinylbutane; ECMO: extracorporeal membrane oxygenation. [#]: primary outcome: adjusted for CURB-65 (confusion, urea >7 mmol·L⁻¹, respiratory rate ≥30 breaths·min⁻¹, blood pressure <90 mmHg (systolic) or ≤60 mmHg (diastolic), age ≥65 years) score and baseline seven-point ordinal scale; adjusted OR >1.0 indicates a benefit of SFX-01 treatment.

first discharge, duration of hospitalisation, or clinical status or mean change in clinical status, or NEWS from baseline at any of the time points was not different (table 3).

There were no significant differences in secondary outcomes relating to oxygen or ventilator use (table 3).

Safety analysis

Rate of study medication discontinuation was significantly higher in the SFX-01 group (33.8%) than in the placebo group (8.8%) (incidence rate ratio (IRR) 3.79, 95% CI 1.53–9.34; p=0.004) (supplementary table S10). The main reasons for discontinuation were adverse events. Gastrointestinal adverse events were the most common reasons for discontinuation and were documented in 10 participants (nine in the SFX-01 group and one in the placebo group).

TABLE 3 Estimates of treatment effects on secondary end-points

	Placebo	SFX-01	Effect estimate (unadjusted)	p-value	Effect estimate (adjusted [#])	p-value
Participants	68	65				
15-day mortality[¶]	8 (11.8)	11 (16.9)	1.45 (0.58–3.61)	0.91	1.46 (0.59–3.62)	0.92
28-day mortality[¶]	11 (16.2)	15 (23.1)	1.45 (0.66–3.15)	0.35	1.45 (0.67–3.16)	0.35
Clinical improvement by day 29[¶]	57 (83.8)	51 (78.5)	1.02 (0.70–1.49)	0.91	1.02 (0.70–1.49)	0.92
Discharged or NEWS ≤2 at day 29[¶]	61 (89.7)	55 (84.6)	0.83 (0.56–1.24)	0.37	0.80 (0.54–1.20)	0.28
Oxygen-free days⁺	20.3±10.1; n=67 25.0 (18.0–28.0)	19.8±10.6; n=65 25.0 (19.0–28.0)	0.98 (0.71–1.34)	0.89	0.98 (0.72–1.34)	0.91
Duration of new oxygen use⁺	1.8±3.5; n=28 0.0 (0.0–2.0)	1.0±1.7; n=24 0.0 (0.0–1.5)	0.55 (0.16–1.91)	0.35	0.49 (0.15–1.64)	0.25
Ventilation-free days⁺	23.5±9.4; n=67 28.0 (27.0–28.0)	21.7±10.6; n=65 28.0 (22.0–28.0)	0.93 (0.72–1.19)	0.53	0.93 (0.72–1.20)	0.57
Duration of new ventilation use⁺	0.8±2.5; n=60 0.0 (0.0–0.0)	1.7±4.7; n=54 0.0 (0.0–0.0)	2.11 (0.52–8.51)	0.30	2.11 (0.53–8.51)	0.29
Mechanical ventilation-free days⁺	24.6±8.0; n=67 28.0 (28.0–28.0)	23.6±8.6; n=65 28.0 (28.0–28.0)	0.96 (0.82–1.13)	0.63	0.96 (0.82–1.13)	0.63
Duration of new mechanical ventilation use⁺	0.5±2.0; n=39 0.0 (0.0–0.0)	0.6±2.0; n=41 0.0 (0.0–0.0)	1.09 (0.09–13.29)	0.94	2.10 (0.17–25.74)	0.56
Duration of hospitalisation⁺	7.4±7.7; n=57 5.0 (3.0–9.0)	6.2±7.3; n=51 3.0 (1.0–8.0)	0.84 (0.56–1.27)	0.41	0.84 (0.56–1.26)	0.40

Data are presented as n, n (%), mean±SD or median (interquartile range), unless otherwise stated. SFX-01: 1-isothiocyanato-4-methyl-sulfinylbutane; NEWS: National Early Warning Score. [#]: adjusted for CURB-65 (confusion, urea >7 mmol·L⁻¹, respiratory rate ≥30 breaths·min⁻¹, blood pressure <90 mmHg (systolic) or ≤60 mmHg (diastolic), age ≥65 years) score and baseline seven-point ordinal scale; [¶]: effect estimates are presented as hazard ratio (95% CI); ⁺: effect estimates are presented as incidence rate ratio (95% CI).

42 (64.6%) participants in the SFX-01 arm and 30 (44.1%) in the placebo arm reported at least one adverse event (IRR 1.48, 0.92–2.36; $p=0.10$) (table 4). The most common adverse event reported in the SFX-01 group was gastrointestinal disorders affecting 33 (60%) patients, compared to 10 (23.8%) patients in the placebo group. There were 34 SAEs in total; 18 in the placebo arm and 16 in the SFX-01 arm.

Serum cytokines and systemic inflammation

Serum cytokine analysis was performed in all participants for whom serum was available at baseline, day 8 and day 15 ($n=230$ samples). There were no significant differences between the SFX-01 and placebo group at any study time point in key cytokines including IL-6, TNF- α or IL-1 β (figure 2a–c).

Of all the serum proteins measured, only the epidermal growth factor family member, transforming growth factor (TGF)- α ($p=0.003$) (figure 2d) and lymphotoxin- α (LTA) ($p=0.049$) (figure 2e) were significantly different in the SFX-01- and placebo-treated groups by day 15.

Peripheral blood leukocyte gene expression

SFX-01 effects on peripheral blood immune cell gene expression were determined using mRNAseq. At baseline, eight statistically significant differentially expressed genes (DEGs) were identified between SFX-01-treated and placebo-treated individuals; at day 8 there were 310 DEGs (286 of which were upregulated in the SFX-01 group); and at day 15, 42 DEGs were identified (figure 3a–c). The top 10 upregulated and downregulated genes with largest fold change are shown in figure 3d–f. However, mRNA levels of classical Nrf2 targets such as *NQO1*, *GSS* and *HO-1* (table 5), and inflammatory cytokines such as *IL-6*, *TNF- α* or *IL-1 β* (figure 3g–h and supplementary figure S2), were unchanged at all time points. *TGF- α* and *LTA* gene expression was also unchanged (supplementary figure S2).

At day 8, gene ontology analysis suggested relevant differential pathways in the SFX-01 group as transcriptional regulation, B-cell receptor signalling (supplementary figure S5) and proliferation. Both subunits of the B-cell antigen receptor CD79 (*CD79A* and *CD79B*) and downstream signalling molecules *BLK* and *BLNK*, proteins involved in costimulatory regulation *CR2*, *CD19* and *CD22*, and the lymphocyte cytokine receptor *CXCR5*, were upregulated. The antiapoptotic factor *BCL2*, plus proliferation promoters *RRAS2* and *RASGRP3* were also significantly increased in the SFX-01 group.

TABLE 4 Details of total adverse events during the trial period

	Placebo	SFX-01
Participants	68	65
Participants with no adverse events	38 (55.9)	23 (35.4)
Participants with adverse events	30 (44.1)	42 (64.6)
Adverse events	42	55
Severity		
Mild	13 (31.0)	35 (63.6)
Moderate	11 (26.2)	4 (7.3)
Severe	18 (42.9)	16 (29.1)
System organ class level		
Cardiac disorders	2 (4.8)	1 (1.8)
Eye disorders	1 (2.4)	0
Gastrointestinal disorders	10 (23.8)	33 (60.0)
General disorders and administration site conditions	2 (4.8)	1 (1.8)
Infections and infestations	15 (35.7)	15 (27.3)
Metabolism and nutrition disorders	1 (2.4)	0
Musculoskeletal and connective tissue disorders	1 (2.4)	0
Neoplasms benign, malignant and unspecified (including cysts and polyps)	1 (2.4)	1 (1.8)
Nervous system disorders	2 (4.8)	2 (3.6)
Renal and urinary disorders	2 (4.8)	0
Respiratory, thoracic and mediastinal disorders	4 (9.55)	2 (3.6)
Skin and subcutaneous tissue disorders	1 (2.4)	0
Adverse event leading to study drug discontinuation	5 (11.9)	20 (36.4)
Data are presented as n or n (%). SFX-01: 1-isothiocyanato-4-methyl-sulfinylbutane.		

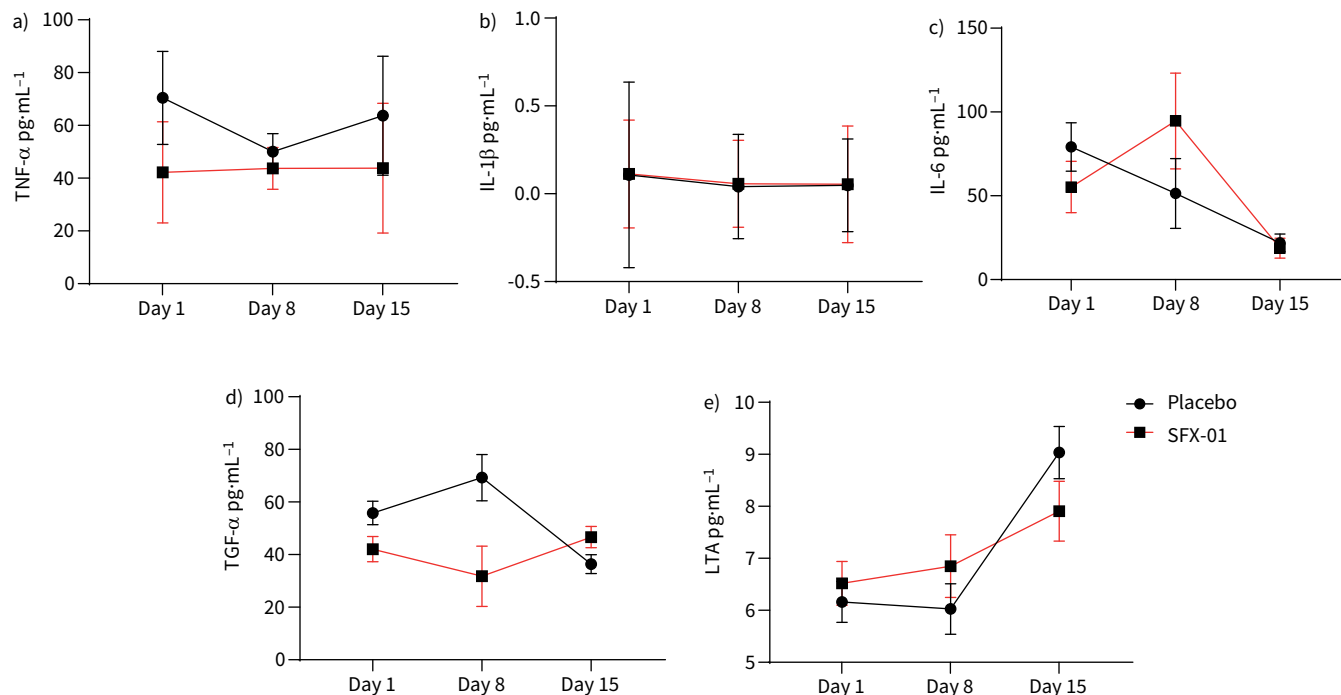


FIGURE 2 Serum cytokine levels and effects of 1-isothiocyanato-4-methyl-sulfinylbutane (SFX-01) treatment. At days 1, 8 and 15, serum was obtained from participants and 45 inflammation-associated cytokines measured. Established Nrf2 targets, **a**) tumour necrosis factor (TNF)- α , **b**) interleukin (IL)-1 β and **c**) IL-6 were not significantly affected by SFX-01 treatment compared with levels in the placebo group. Of the 45 cytokines measured, only the epidermal growth factor family member **d**) transforming growth factor (TGF)- α was significantly higher by day 15 in the SFX-01 group than in the placebo group, and only the apoptosis-inducer **e**) lymphotoxin- α (LTA) was significantly reduced in the SFX-01 group. Data were analysed by a mixed-model repeated measures approach; data represent model-derived mean \pm se. Day 1: SFX-01 n=61, placebo n=63; day 8: SFX-01 n=17, placebo n=21; day 15: SFX-01 n=33, placebo n=38.

Out of the significant DEGs upregulated at day 8 in the SFX-01 group, 13 remained significantly upregulated at day 15, including several B-cell-associated genes (e.g. *CD79A*, *CD19*, *BLK*, *PAX5*, *Fc γ R2*) plus genes regulating B-cell survival (*TCL1A*). Of the DEGs downregulated at day 8, only *SIGLEC1*, an interferon-signalling gene and macrophage/monocyte-associated gene, was still significantly altered at day 15. Furthermore, *CD200*, a macrophage/monocyte suppressor, was significantly upregulated at both time points.

Enrichment analysis identified further pathways associated with SFX-01 treatment at both days 8 and 15 (supplementary figures S3 and S4 and supplementary tables S19 and S20). Although TNF gene expression was not significantly altered, interestingly, pathways relating to both myeloid cell and adaptive responses and also TNF production were downregulated in the SFX-01 group at day 8. At day 15, pathways centred heavily around DNA and RNA processing, but smaller clusters were identified relating to viral responses and interferon signalling, which were suppressed in the SFX-01 group.

Differences in effects of SFX-01 treatment in vivo and L-SFN treatment in vitro

In vivo data in the present study were compared to data from human PBMCs treated for 24 h with or without 15 μ M L-SFN *in vitro*. 1032 genes were significantly differentially expressed in L-SFN-treated PBMCs (false discovery rate (FDR) <0.05, \log_2 FC >1 or <-1) (figure 3i), and 801 when a more stringent FDR of <0.01 was applied. Downregulated pathways include inflammatory cytokine production and also response to oxidative stress, demonstrating significant anti-inflammatory effects. Notably, however, several Nrf2-associated genes unaltered by SFX-01 in our study, including *NQO1*, *GPX*, *HMOX1*, *GSS* and *G6PD*, also showed no significant differences with L-SFN treatment in human PBMCs at this time point (table 5) [27].

Of the DEGs found at day 8 in STAR-COVID-19 trial participants, 250 were identified in PBMCs, and 30 were significantly differentially expressed with L-SFN treatment. 25 of these were upregulated at day 8 *in vivo*; however, all 25 were significantly downregulated in L-SFN-treated PBMCs, including *HLA-DOA*,

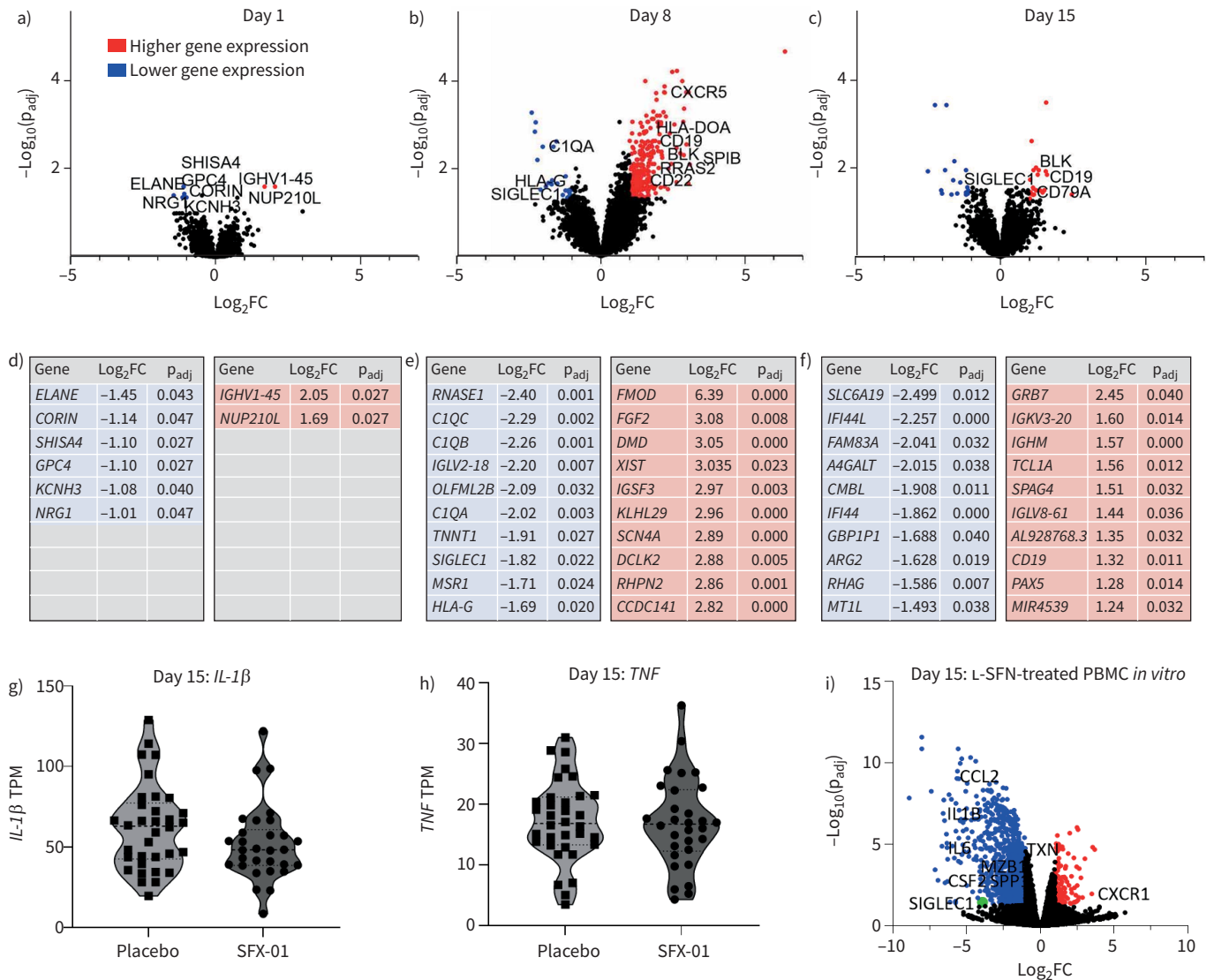


FIGURE 3 Peripheral blood leukocyte gene expression and effects of 1-isothiocyanato-4-methyl-sulfinylbutane (SFX-01) treatment and comparison with *in vitro* sulforaphane treatment. At **a)** day 1, **b)** day 8 and **c)** day 15, peripheral blood leukocyte gene expression analysed by mRNA sequencing was compared between SFX-01- and placebo-treated individuals who completed 14 days of trial treatment and had not discontinued drug at the relevant sampling time point. **a)** Eight significantly differentially expressed genes (DEGs) were identified between the SFX-01 and placebo groups at day 1; **b)** 310 DEGs were identified at day 8; and **c)** 42 at day 15. Higher (red) and lower (blue) gene expression levels in the SFX-01 are shown. **d-f)** The top 10 significant DEGs (adjusted p-value (p_{adj}) <0.05 and \log_2 fold change (FC) >1 or <-1) between SFX-01- and placebo-treated individuals. Red indicates higher expression in the SFX-01 group compared with the placebo group; blue indicates lower expression. Data were analysed using the Wald test and Benjamini-Hochberg procedure. **g, h)** Transcript per million (TPM) values representing relative expression levels of genes of interest: *IL-1β* and *TNFα* at day 15. Day 1: SFX-01 n=59, placebo n=59; day 8: SFX-01 n=14, placebo n=18; day 15: SFX-01 n=30, placebo n=34. **i)** Re-analysis of a published dataset (GSE160353); differential gene expression analysis of isolated human peripheral blood mononuclear cells (PBMCs) treated with L-sulforaphane (L-SFN) (15 μM) or vehicle for 24 h and processed for RNA sequencing (n=4 individuals), 1032 significant DEGs identified (false discovery rate-adjusted p-value <0.05 and \log_2 fold change >1 or <-1).

-*DOB* and -*DQA1*, and *CD22*. The remaining five were downregulated in both the day 8 *in vivo* and PBMC datasets: *OLFML2B*, *RNASE1* *MAFB*, *ODF3B* and *SIGLEC1*.

Of the 13 DEGs consistently upregulated in SFX-01 treatment at both day 8 and day 15, 12 were identified in the PBMCs, and four of these genes were significantly differentially expressed with L-SFN, although these were all downregulated *in vitro*. In particular, *CD19* was reduced in L-SFN-treated PBMCs, with Gene Ontology analysis demonstrating significant changes in B-cell receptor signalling, B-cell surface molecules and regulation of B-cell proliferation.

TABLE 5 Direct Nrf2 gene targets in L-sulforaphane (SFN)-treated human peripheral blood mononuclear cells (PBMCs) *in vitro* and *in vivo* in peripheral blood leukocytes in the STAR-COVID-19 (SFX-01 Treatment for Acute Respiratory Infections) trial of 1-isothiocyanato-4-methylsulfanylbutane (SFX-01)

	Pathway	<i>In vitro</i> L-SFN: PBMCs		<i>In vivo</i> SFX-01: day 8		<i>In vivo</i> SFX-01: day 15	
		Adjusted p-value	Log ₂ FC	Adjusted p-value	Log ₂ FC	Adjusted p-value	Log ₂ FC
<i>FECH</i>	Haem production	0.69	0.16	0.70	0.313	0.98	-0.02
<i>FTL</i>	Haem/iron metabolism	0.05	-0.44	0.98	-0.02	0.78	0.13
<i>G6PD</i>	NADPH regeneration	0.97	-0.03	0.98	-0.02	0.27	0.37
<i>GCLC</i>	GSH production	0.002	0.92	0.67	0.24	0.9	-0.06
<i>GCLM</i>	GSH production	0.003	0.97	0.78	0.15	0.82	-0.1
<i>GSR</i>	GSH production	0.33	0.35	0.84	-0.09	0.8	0.06
<i>GSS</i>	GSH production	0.61	0.18	0.84	0.08	0.72	0.08
<i>GSTM2</i>	ROS detoxification	0.43	-0.42	0.39	0.38	0.26	0.34
<i>GSTM3</i>	ROS detoxification	0.71	0.22	0.12	1.07	0.88	0.1
<i>HMOX1</i>	Haem/iron metabolism	0.62	-0.24	0.33	-0.44	0.67	0.14
<i>ME1</i>	NADPH regeneration	0.73	0.28	0.26	-0.68	0.27	-0.42
<i>NQO1</i>	ROS detoxification	0.24	0.45	0.33	0.45	0.87	0.06
<i>OSGIN1</i>	Various including autophagosome formation	0.34	0.72	0.84	-0.14	0.37	0.35
<i>PGD</i>		NADPH regeneration	<0.001	-1.03	0.88	-0.11	0.52
<i>PRDX1</i>	TXN-based antioxidant system	0.6	-0.16	0.70	0.15	0.66	0.1
<i>RXRA</i>	Lipid metabolism	0.6	-0.2	0.51	0.26	0.49	0.21
<i>SPP1</i>	Various including inflammatory signalling	0.004	-2.75	0.65	0.42	0.93	0.09
<i>SRXN1</i>	TXN-based antioxidant system	0.53	-1.11	0.86	-0.15	0.78	0.18
<i>TALDO1</i>	NADPH regeneration	0.05	-0.43	0.85	-0.12	0.58	0.21
<i>TKT</i>	NADPH regeneration	0.01	-0.62	0.64	-0.24	0.5	0.22
<i>TXN</i>	TXN-based antioxidant system	<0.0001	-1.55	0.98	0.02	0.88	0.09
<i>TXNRD1</i>	TXN-based antioxidant system	0.15	0.42	0.84	-0.1	0.85	0.07

Bold type represents significantly differentially expressed genes between L-SFN-treated and -untreated PBMCs; false discovery rate adjusted p-value <0.05 and log₂ fold change (log₂FC) >1 or <-1. GSH: glutathione; ROS: reactive oxygen species; TXN: thioredoxin.

The only gene to be consistently significantly altered in the same direction at both day 8 and 15 with SFX-01 *in vivo*, and in PBMCs treated with L-SFN *in vitro*, was *SIGLEC1* (supplementary material).

Discussion

In this double-blind randomised trial, SFX-01 did not improve day 15 clinical status in hospitalised patients with CAP. Additionally, we observed an increased rate of treatment discontinuation, mainly due to gastrointestinal adverse effects in the SFX-01 group. The study was terminated early after pre-specified criteria for futility were met. Subgroup analyses including in CAP patients with and without SARS-CoV-2 infection were all consistent with the primary results.

The antioxidant transcription factor Nrf2 is reduced in acute lung infection including in COVID-19 [18, 19]. Animal models as well as human studies have shown significant beneficial effects of Nrf2 activation with sulforaphane or sulforaphane-rich preparations [20, 23, 24, 26, 28, 29, 30, 31]. In view of the negative results we observed, we therefore investigated further whether Nrf2 activation had been achieved in our study. Analyses of serum cytokine levels and peripheral blood leukocyte gene expression evidenced that SFX-01 treatment did not result in Nrf2 activation, shown by a lack of antioxidant gene induction [16] or effects on key inflammatory cytokines such as IL-6 and IL-1 β [17, 32]. Our study is unable to answer why SFX-01 failed to activate Nrf2 targets. Interestingly, a study of sulforaphane in people with asthma showed that Nrf2 activation was highly heterogeneous between individuals [25].

A plausible reason that SFX-01 did not achieve the antioxidant or inflammatory modulation shown in other models may be suboptimal dosing. It should also be noted that Nrf2 activation with SFX-01 has not yet been confirmed in human trials, but was deemed highly likely considering the effects of sulforaphane demonstrated in the literature, and SFX-01 administration resulted in anti-inflammatory effects in animal models [28]. SFX-01 has been utilised in two other phase II trials (clinicaltrials.gov identifiers NCT02614742, NCT02970682) with a twice-daily 300 mg dose for 29 days in subarachnoid haemorrhage or up to 6 months in patients with metastatic breast cancer. These studies did not investigate Nrf2-related gene expression to confirm target engagement. Use of a higher dose in the COVID-19 population is

unlikely to be feasible given the treatment discontinuation rates and prevalence of gastrointestinal-associated adverse events observed in the present study.

COVID-19 patient lung biopsy samples have shown Nrf2 suppression by SARS-CoV-2 infection [19], although Nrf2 status in immune cells still requires further investigation; one further explanation for lack of SFX-01 efficacy could be that infection-related Nrf2 suppression cannot be overcome by the administered therapy. In the present study, we observed no effects of SFX-01 on classical Nrf2 targets; however, we observed effects of treatment on B-cell activation and proliferation. There are contradictory reports of sulforaphane effects on B-cells. Expansion and activation of B-cells was demonstrated in severe COVID-19, although B-cell responses are required for robust adaptive immunity [33]. In both murine arthritis [34] and lupus [35] models, sulforaphane inhibited B-cell proliferation and was associated with reduction in plasma cell numbers, and Nrf2 knockout in mice with chronic airway inflammation showed enhanced plasma cell infiltration and B-cell activation [36]. In contrast, sulforaphane-treated PBMCs demonstrated dose-dependent reductions in numbers of monocytes, consistent with significant reductions in several monocyte-associated genes in our present study, and also increased numbers of dendritic cells, CD19⁺ B-cells and T-lymphocytes [37].

To further understand these changes, gene expression data in the present study were compared to published RNAseq data from *in vitro* sulforaphane-treated human PBMCs [27]. In the additional dataset analysed, several of the B-cell associated genes upregulated in the present dataset showed the opposite trend in PBMCs, in support of results from murine models. Only one gene, *SIGLEC1*, was significantly downregulated in PBMCs and at both trial time points. Interestingly, interferon-inducible *siglec1* (also known as CD169) is capable of direct viral binding at the cell surface [38] and is upregulated in myeloid cells in COVID-19 patients [39]. Nrf2 activation in these cells was indicated by anti-inflammatory pathway induction; however, several Nrf2 targets including *NQO1* were not detectably changed at the sampling time. By contrast, the mRNA levels for the classical Nrf2 targets *NQO1*, *HMOX1* and *AKR1C1* were significantly upregulated by a shorter incubation time (6 h) and lower concentrations of sulforaphane (2 or 5 μ M) in *ex vivo*-treated human PBMCs [32]. Interestingly, there was no concentration dependence in the induction of these genes, with the higher (5 μ M) sulforaphane concentration even showing a slightly diminished effect for the induction of *NQO1* and *AKR1C1* [32], in agreement with results from a human study with topical administration of sulforaphane-rich extracts showing diminished efficacy by the highest sulforaphane dose used [40]. Taken together, while underdosing in our study is a possible explanation, excessive dosing could also theoretically be associated with a reduced effect on Nrf2 targets.

Ours is not the first study to question the effects of sulforaphane administration on Nrf2 activation *in vivo*. Several human studies of sulforaphane found no effects on Nrf2 antioxidant targets [41, 42, 43]. While there are limitations to comparison of the present trial data with the PBMC dataset, including lack of inflammatory stimulus, use of a different sulforaphane formulation, concentration and treatment period, the results highlight the critical need for *in vivo* data on sulforaphane effects in addition to applying caution in extrapolation from *in vitro* findings.

Sulforaphane is reported to have activity which is not dependent on Nrf2 activation. For example, suboptimal dosing without Nrf2 activation demonstrated significant effects on fibrosis [44], sulforaphane improved macrophage phagocytosis *via* Nrf2-independent mechanisms [45] and inhibited interferon- γ and TNF- α -mediated pro-inflammatory responses in both an Nrf2-dependent and Nrf2-independent manner [46], in addition to Nrf2-independent antiviral effects [20]. Further investigation to understand the mechanism of action of sulforaphane utilising appropriate dosing and sampling is required. Importantly, as we are unable to demonstrate Nrf2 activation by SFX-01, our study does not exclude possible clinical benefits with alternative Nrf2-activating drugs in CAP. Other Nrf2 activators, such as the cyanoenone triterpenoids, are currently in various stages of drug development, and our study highlights that a number of challenges still remain [47].

During the COVID-19 pandemic a large number of trials were set up to test repurposed and novel therapeutics particularly in hospitalised patients. It was, and remains, important to rapidly establish the lack of efficacy of drugs so that resources can be invested into other targets. Therefore, while this trial is negative, it provides a clear answer to a relevant question about the potential efficacy of SFX-01 in this population. Key strengths of this study include the gold standard double-blind placebo-controlled design and the inclusion of CAP patients including both COVID-19 and non-COVID-19 CAP, recognising that in future SARS-CoV-2 will be just one of several circulating pathogens responsible for CAP in hospitalised patients; excessive systemic inflammation in pneumonia is a driver of high mortality rates in CAP, and further human studies of anti-inflammatory agents are critical.

In conclusion, 300 mg SFX-01 once-daily treatment for 14 days in hospitalised patients with CAP did not result in Nrf2 activation or improved clinical status.

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This study is registered at <https://eudract.ema.europa.eu/> with identifier number 2020-003486-19. Deidentified patient data are available from the corresponding author following publication along with the study protocol and data dictionary.

Ethics statement: The trial was approved by the Scotland A research ethics committee (20/SS/0092). All patients or legal representatives provided written informed consent. An independent, external data safety monitoring committee reviewed adverse event data.

Author contributions: Conception and design of study: M.B. Long, A.T. Dinkova-Kostova and J.D. Chalmers. Acquisition of data: M.B. Long, H. Abo-Leyah, Y.H. Giam, R.C. Hull, H.R. Keir, T. Pembridge, D.A. De Lima, L. Delgado, S.K. Inglis, C. Hughes, A. Gilmore, B.J.M. New and J.D. Chalmers. Analysis and/or interpretation of data: M.B. Long, H. Abo-Leyah, Y.H. Giam, T. Vadeloo, R.C. Hull, G. MacLennan, A.T. Dinkova-Kostova and J.D. Chalmers. Drafting the manuscript: M.B. Long, H. Abo-Leyah, Y.H. Giam, A.T. Dinkova-Kostova and J.D. Chalmers. Revising the manuscript critically for important intellectual content: all authors. Approval of the version of the manuscript to be published: all authors.

Conflict of interest: H.R. Keir reports receiving personal fees for educational lecture from Insmad Inc., outside the submitted work. A.T. Dinkova-Kostova participates on the Evgen Pharma Scientific Advisory Board, outside the submitted work. J.D. Chalmers reports support for the present manuscript from LifeArc; grants or contracts from AstraZeneca, Genentech, Gilead Sciences, GlaxoSmithKline, Insmad, Grifols, Novartis and Boehringer Ingelheim, outside the submitted work; consulting fees from AstraZeneca, Chiesi, GlaxoSmithKline, Insmad, Grifols, Novartis, Boehringer Ingelheim, Pfizer, Janssen, Antabio and Zambon, outside the submitted work; and is an associate editor of this journal. The remaining authors have nothing to disclose.

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