

Review

Emerging Roles of Extracellular Vesicles Derived from Bacteria, Mammalian or Plant Cells in the Pathogenesis and Clinical Application of Neurodegenerative Diseases

Yihong Li ^{1,2,3,†} , Chenglong Zhou ^{4,†}, Huina Liu ^{1,2}, Ting Cai ^{1,2} and Huadong Fan ^{1,2,5,*} 

¹ Ningbo No. 2 Hospital, Ningbo 315099, China; yihongli@ucas.ac.cn (Y.L.); liuhuina@ucas.ac.cn (H.L.); caiting@ucas.ac.cn (T.C.)

² Ningbo Institute of Life and Health Industry, University of Chinese Academy of Sciences, Ningbo 315000, China

³ Laboratory of Nanopharmacology Research for Neurodegeneration, Department of Research and Development of Science and Technology, Ningbo Institute of Life and Health Industry, University of Chinese Academy of Sciences, Ningbo 315000, China

⁴ College of Biological & Environmental Sciences, Zhejiang Wanli University, Ningbo 315100, China; 2022881047@zwu.edu.cn

⁵ Laboratory of Dementia and Neurorehabilitation Research, Department of Research and Development of Science and Technology, Ningbo Institute of Life and Health Industry, University of Chinese Academy of Sciences, Ningbo 315000, China

* Correspondence: fanhuadong@ucas.ac.cn; Tel.: +86-0574-87088274

† These authors contributed equally to this work.

Abstract: A growing number of studies have indicated that extracellular vesicles (EVs), such as exosomes, are involved in the development of neurodegenerative diseases. Components of EVs with biological effects like proteins, nucleic acids, or other molecules can be delivered to recipient cells to mediate physio-/pathological processes. For instance, some aggregate-prone proteins, such as β -amyloid and α -synuclein, had been found to propagate through exosomes. Therefore, either an increase of detrimental molecules or a decrease of beneficial molecules enwrapped in EVs may fully or partly indicate disease progression. Numerous studies have demonstrated that dysbiosis of the gut microbiota and neurodegeneration are tightly correlated, well-known as the “gut–brain axis”. Accumulating evidence has revealed that the gut bacteria-derived EVs play a pivotal role in mediating microbe–host interactions and affect the function of the “gut–brain axis”, which subsequently contributes to the pathogenesis of neurodegenerative diseases. In this review, we first briefly discuss the role of EVs from mammalian cells and microbes in mediating the progression of neurodegenerative diseases, and then propose a novel strategy that employs EVs of plants (plant cell-derived exosome-like nanoparticles) for treating neurodegeneration.

Keywords: extracellular vesicles; outer membrane vesicles (OMVs); plant-derived exosome-like nanoparticles (PDELNs); gut dysbiosis; gut–brain axis; microglia; neuroinflammation; neurodegeneration



Citation: Li, Y.; Zhou, C.; Liu, H.; Cai, T.; Fan, H. Emerging Roles of Extracellular Vesicles Derived from Bacteria, Mammalian or Plant Cells in the Pathogenesis and Clinical Application of Neurodegenerative Diseases. *Biomolecules* **2024**, *14*, 312. <https://doi.org/10.3390/biom14030312>

Academic Editors: Mebarek Saida, Leyre Brizuela and Soo-Ho Choi

Received: 1 January 2024

Revised: 21 February 2024

Accepted: 1 March 2024

Published: 6 March 2024



Copyright: © 2024 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

1. Introduction

Neurodegenerative diseases are most commonly characterized by the aggregation of misfolded proteins due to improper post-translational modification of proteins like TDP-43 in amyotrophic lateral sclerosis/frontal temporal lobe dementia (ALS/FTLD), α -synuclein (α -syn) in Parkinson’s disease (PD), and β -amyloid ($A\beta$) in Alzheimer’s disease (AD) [1–3]. Nearly all types of cells, including prokaryotic and eukaryotic cells, secrete extracellular vesicles (EVs) with a diameter ranging approximately from 10 nm to 200 nm. As the mechanism of EV generation may vary among different species depending on parental cells, and because the exact process by which EVs are released from bacterial cells or plant cells remains unclear, here we only introduce an example of how exosomes

are generated by mammalian cells. Generally, the exosomes are produced in a process involving the invagination of a double plasma membrane and the formation of intracellular multivesicular bodies (MVBs) containing intraluminal vesicles (ILVs) [4,5]. Finally, the MVBs undergo a process resembling exocytosis to release mature exosomes [6]. Evidence has shown that exosomes play a key role in propagating disease-associated proteins related to neurodegeneration within the brain [7–9]. Exosomes contain various components of their parental cells, including nucleic acids, lipids, and proteins from the cytoplasm and the surface membrane, as well as cellular metabolites, which can be taken up by their target cells. For instance, exosomes can either carry detrimental factors released from neurons to induce inflammation in glial cells, facilitating the progression of neurodegeneration [10], or engage in neuroprotective signaling transduction [11–14].

2. Extracellular Vesicles from Mammalian Cells

2.1. Behaviors and Functions of Mammalian EVs

The uptake of EVs is mediated in several ways (Table 1), including endocytosis, phagocytosis, and direct fusion with the plasma membrane. It has been demonstrated that the anchor proteins of the surface membrane of EVs can interact with membrane receptors on recipient cells, and this “ligand–receptor” interaction mediates the uptake of EVs by their target cells [15]. To address this mechanism, investigators used specific inhibitors or antibodies to block receptor–ligand interactions, revealing that the uptake of EVs was significantly hampered in a variety of cell types, which demonstrated that receptor-mediated endocytosis contributes to the uptake process of EVs [16–20]. Additionally, another study showed that some EV membranes were able to fuse directly with the plasma membrane of the recipient cells by labelling melanoma cell-derived exosomes with the lipid fluorescent probe Octadecyl Rhodamine B Chloride (R18) [21]. These studies together suggested that there are several known mechanisms underlying EV uptake, and the cells of different types or with different functions may choose a different manner of EV uptake to complete EV-mediated intercellular communication. Below is a table that lists several types of EV uptake.

Table 1. Types of EV uptake.

Types of EVs Uptake	Examples	References
Endocytosis	1: CME. Recipient cells treated with chlorpromazine reduce the uptake of EVs, and chlorpromazine prevents the formation of lattice protein-coated pits in the plasma membrane.	[8,16]
	2: Phagocytosis. EVs were labelled with a fluorescent dye, and dendritic cells were found to have red fluorescence, confirming that they could phagocytose EVs.	[17]
	3: CDE. Endocytosis of EVs by CDE requires activation by dynamin2, which can be blocked by a specific inhibitor, leading to a significant reduction of internalization for EVs.	[18,19]
Membrane Protein interactions	1: Tetraspanins. Treatment of recipient cells with antibodies against tetraspanin reduces EV uptake by dendritic cells.	[20]
	2: Immunoglobulins. Naive T cells have been shown to internalize EVs mediated by the T cell receptor (TCR), CD28, and LFA-1.	[6,21]
	3: Proteoglycan. Acetyl-heparin sulfate proteoglycan (HSPG) acts as a receptor for cancer cell-derived exosomes.	
Cell surface membrane fusion	Purified exosomes from melanoma cells labeled with fluorescent lipid dye showed that some EV membranes were able to fuse directly with the plasma membrane of the recipient cells.	[15]

2.2. Role of EVs of Mammalian Cells in Neurodegenerative Diseases

EVs play a double role in the central nervous system. On the one hand, disease-associated proteins can be propagated by EVs shuttled between different cells. As the

disease develops, these proteins spread from one focal point in the brain to a larger scope of neuronal regions, accelerating the progression of neurodegeneration [22,23]. EVs containing disease-associated proteins involved in Prion disease, Parkinson's disease (PD), Alzheimer's disease (AD), and amyotrophic lateral sclerosis (ALS) have all been found in the cerebral spinal fluid (CSF) and blood of patients affected by these disorders [24]. Prion diseases are a group of rare progressive neurodegenerative diseases, including Creutzfeldt–Jakob disease (CJD), Gerstmann–Straussler–Scheinker disease, and kuru [25,26]. It is now widely accepted that the misfolding of the host-encoded prion protein, PrP^C, into a disease-associated transmissible form, PrP^{SC}, results in the transmission of pathology not only between cells but also from one region to another [27,28]. Both forms of prion proteins were found to be shuttled by exosomes [29]. Exosomal PrP^{SC} was found to transmit protein aggregation in rabbit kidney epithelial cells [30]. Subsequent in vivo experiments showed that exosomes derived from prion-infected mice were able to transmit aggregation to naïve mice [31,32]. For many years, PrP^{SC} involved in prion disease was the only known transmissible protein for the spread of disease, but recent studies using both animal and cellular models have confirmed that other proteins related to neurodegeneration are also transmissible. This includes α -synuclein in PD, and tau and A β in AD [33]. For example, EVs are an efficient carrier of α -synuclein aggregation and propagation between neurons, thus promoting the progression of PD [34]. Furthermore, EVs circulating in the blood and CSF of patients with PD have been found to be highly enriched with α -synuclein and are remarkably correlated with the stage of the disease [35]. For AD, it has been shown that neurotoxic, oligomeric forms of A β protein are wrapped in EVs isolated from brain tissue, and these vesicles can mediate the inter-neuronal propagation of A β [34]. To testify the critical role of EVs in AD development, an in vivo study revealed that injecting 5xFAD mice (AD model mice) with neutral sphingomyelinase 2 (nSMase2), an inhibitor of exosome secretion, significantly reduced amyloid plaque formation in the brain [31]. In addition, another study demonstrated that, as carriers of A β , astrocytes-derived extracellular vesicles (ADEVs) are involved in the pathogenesis of AD [36]. In the brain, astrocytes phagocytose too much fibril A β 42 to digest them, which causes a severe accumulation of intracellular A β . To avoid further intracellular stress, astrocytes release undigested fibrils of A β 42 via EVs, which would, in turn, lead to severe neurotoxicity in neighboring neurons [37]. Also, in ALS patients, astrocytes can generate EVs, which are toxic and lead to adjacent motor neuron death [38]. Furthermore, ADEVs mediate the propagation of neuroinflammation as well as regulate mutual signaling between the brain and the immune system. In a mouse model of inflammatory brain injury, ADEVs rapidly enter the peripheral circulation, inducing an acute peripheral cytokine response to accelerate the migration of peripheral leukocytes to the brain, thereby triggering neuroinflammation [39]. The above experimental data suggested that ADEVs in the peripheral blood might serve as a source of biomarkers for neurological disorders. As the EVs circulating in the blood are likely to be derived from various tissues throughout the body, the isolation of cell type-specific EVs can provide us with information about a certain pathological status. Namely, analyzing the contents of EVs derived from neurons or glial cells in the blood would help to identify novel biomarkers related to neurodegenerative diseases [40].

On the other hand, EVs act as a scavenger that can remove aggregation-prone misfolded proteins of cellular/intercellular space, exerting a neuroprotective effect [41]. As shown by investigators, the correctly folded prion protein (PrP^C) on EVs could trap neurotoxic β -amyloid (A β) to promote its fibrillation. In this case, the role of PrP^C-contained exosomes is to remove A β to diminish its neurotoxicity and prevent the accumulation of misfolded proteins [31]. Additionally, in order to take advantage of the neuroprotective role of mammalian cell-derived EVs, numerous studies have concentrated on the therapeutic effect of stem cell-derived EVs, especially on mesenchymal stromal cell-derived EVs (MSC-EVs) [42–47]. It was initially found that mesenchymal stromal cells (MSCs), isolated from bone marrow or adipose tissues, can significantly mitigate neurodegeneration [46,48]; later, investigators confirmed that even MSC-EVs themselves can strongly alleviate cognitive

impairment caused by brain injury, stroke, or neurodegeneration [14,49,50], accompanied by obvious neuron regeneration throughout the ventricular region, cingulate gyrus, and hippocampus [51–53]. MSCs have the strong ability to migrate and differentiate, interacting with brain parenchyma to release vascular endothelial growth factors (VEGFs), nerve growth factors (NGFs), brain-derived neurotrophic factor (BDNFs), and other bioactive molecules to promote the regeneration of blood vessels and nerves, and the reconstruction of neural synapses, as well as to prevent neuron apoptosis [54–57]. In addition, MSCs can restrict the release of inflammatory molecules like prostaglandins and interleukins to minimize neuroinflammation [58,59]. The above beneficial effects that MSCs display depend on their paracrine function rather than on direct interaction with the diseased site [44,49]. It was later verified that the conditioned medium of cultured MSCs showed a similar therapeutic effect to that of MSCs themselves [60,61]. More interestingly, EVs isolated from an MSCs-cultured medium showed almost the same protective effect as MSCs [59,62].

The exact mechanism underlying the neuroprotective role of MSC-EVs remains ambiguous. Generally, MSC-EVs have bioactive contents that include cytokines, growth factors, signaling lipids, and regulatory microRNAs, which can influence tissue rehabilitation after injury, infection, or disease [59]. For example, over 900 varieties of protein molecules in MSC-EVs have been identified using proteomics technology, including neprilysin, a protease that can degrade A β oligomer [63]. In addition, Egor A. and colleagues found that MSC-EVs exert a neuroprotective role via preventing calcium overload in an PI3K/AKT-dependent manner [14].

2.3. The Potential of MSC-EVs as a Biogenic Drug for Treating AD

In the pathogenesis of AD, a high level of homocysteine in plasma (hyperhomocysteinemia, HHcy) is an independent risk factor [64–67]; HHcy AD mice show an increased A β level in the brain [68]. In homocysteine metabolism, insufficiency of 5-methyltetrahydrofolate (the active form of folate) would result in an accumulation of its upstream substrate, homocysteine [69], which is consistent with another study showing that a folate-deficient diet can also accelerate brain amyloidosis in an AD mouse model [70]. Meanwhile, investigators have indicated that high folate intake decreases the risk of AD [71]. However, sufficient dietary intake of folate does not mean that it is efficiently delivered to the brain; in particular, the blood–brain barrier (BBB) excludes most of the free folate in the plasma. The efficient delivery of folate to the brain parenchyma largely depends on the specific recognition of folate-receptor α (FR α), which is shuttled by EVs derived from choroid plexus epithelial cells [72–74]. Therefore, only with the help of FR α shuttled by exosomes can folate be smoothly transported through the BBB to reach the neurons or glia. Since previous studies have shown that MSCs contain a high level of FR α , it is a strong possibility that FR α might appear in MSC-EVs. In fact, it was independently demonstrated by our lab that there is a high abundance of FR α in MSC-EVs (unpublished data).

In summary, based on the above evidence, as shown in Figure 1, we hypothesize that MSC-EVs might be used for AD treatment through supplementing with FR α (MSC-EVs containing FR α), thus facilitating folate uptake by the brain parenchyma and finally blocking HHcy-facilitated amyloidosis in the brain. In AD patients, one can obtain MSCs from bone marrow, adipose tissue, or umbilical cord blood, and then isolate EVs from MSCs-cultured medium. Upon intranasal administration, the MSC-EVs could easily penetrate the BBB and release folate into the brain parenchyma. The key mechanism of this process is to modulate homocysteine metabolism by affording efficient folate through EVs-mediated folate transportation. In fact, a clinical trial (NCT04388982) on the intranasal administration of MSC-EVs to AD patients is already being conducted by another research group [75].

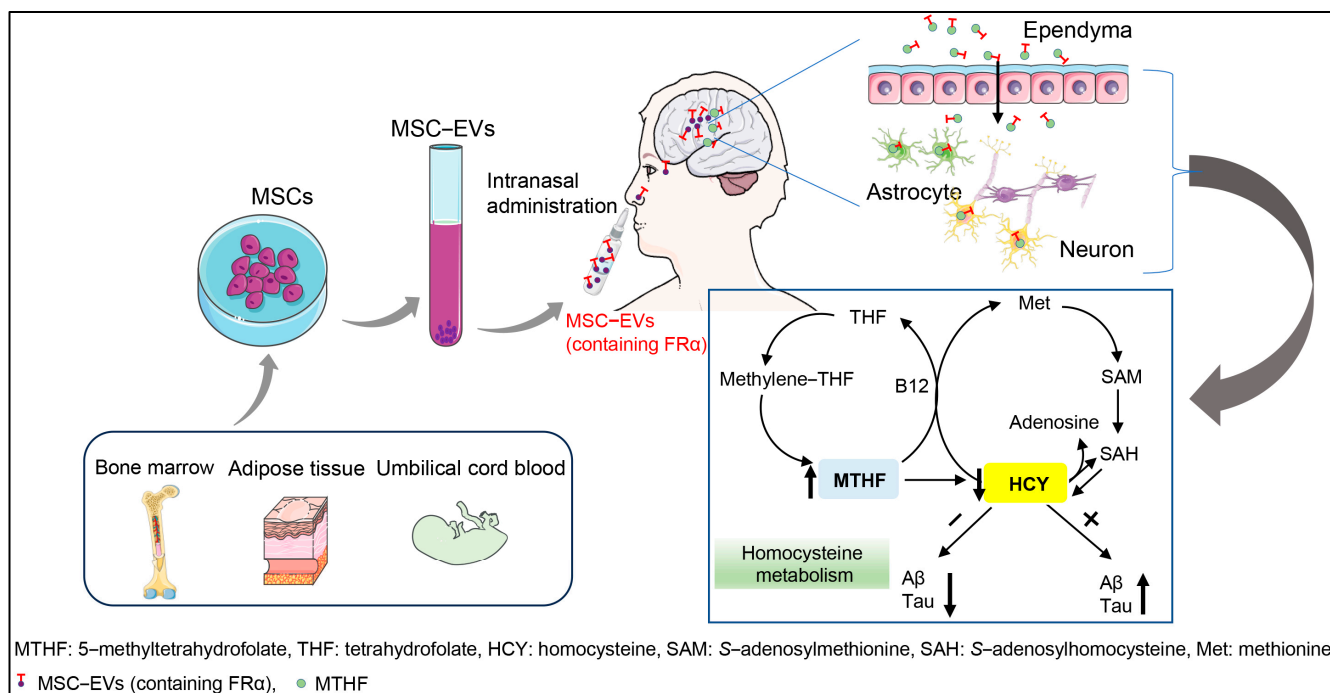


Figure 1. A possible mechanism underlying the neuroprotective role of MSC-EVs in Alzheimer's disease. Abbreviations: MSCs, mesenchymal stromal cells; MSC-EVs, mesenchymal stromal cell-derived extracellular vesicles; MTHF: 5-methyltetrahydrofolate; THF, tetrahydrofolate; HCY, homocysteine; SAM, S-adenosylmethionine; SAH, S-adenosylhomocysteine; Met, methionine.

3. Bacterial EVs

Humans are colonized by multiple commensal organisms. Our gastrointestinal tract provides a residence for both beneficial and pathogenic microorganisms, both of which release bilayer lipid membrane nanovesicles (outer membrane vesicles, OMVs) of spherical morphology with a diameter ranging from 20 to 400 nm [76]. Being different from mammalian EVs, OMVs contain bacteria-specific lipopolysaccharides and peptidoglycans except for regular DNA, RNA, and protein [77,78].

3.1. Behaviors and Functions of Bacterial EVs

The behaviors of OMVs and their biological effects are complicated. Imbalance in the composition of the beneficial and pathogenic bacteria, known as dysbiosis, is considered as a major contributor to inflammatory bowel disease. Also, emerging evidence has indicated that OMVs play a key role in the development of inflammatory diseases. OMVs usually carry immunogenic molecules, including lipopolysaccharides, peptidoglycan or related proteins, that can be recognized by specific receptors expressed in the host cells, which eventually either exacerbate pathological conditions or promote host colonization and confer protective immunity (Table 2) [79]. Generally, there are two methods by which OMVs enter into the host cell as follows: the first can be seen in *Pseudomonas aeruginosa*-secreted OMVs, the plasma of which directly fuses with its host cell's membrane and then releases the carried contents; the second is endocytosis, seen in *Escherichia coli*, through which the contents of OMVs directly enter into the host cytoplasm [80]. For the subsequent biological effects, OMVs from pathogenic bacteria can exacerbate infection by either suppressing the immune response or over-exacerbating it. For example, OMVs from *Pseudomonas aeruginosa* carry a variety of virulent factors, including peptidoglycan hydrolase, phospholipase C, alkaline phosphatase, protease, and hemolysin [81]. These OMV-enwrapped detrimental factors, in combination with Lipopolysaccharide (LPS), elicit an inflammatory response with a bacterial strain-specificity [82]. Furthermore, researchers also found that only OMVs from isolated from *Escherichia coli* cultures can trigger an inflammatory response [83]. In addition

to the directly detrimental effect of OMVs, they can also elicit the pathological function by interacting with the other microorganism. For instance, human *Bacteroides fragilis* can protect the host from colitis by releasing a single microbial molecule polysaccharide (PSA); however, OMVs secreted from *Bacillus subtilis* may disrupt such protection by degrading polysaccharides [84].

Table 2. Varieties of OMV secreted by pathogenic and beneficial bacteria.

Bacteria	Functions of OMVs	References
ETEC	ETEC OMVs can deliver ClyA, a pore-forming cytotoxin expressed by <i>E. coli</i> and some other enterobacteria.	[85]
<i>Pseudomonas aeruginosa</i>	OMVs from <i>Pseudomonas aeruginosa</i> contain multiple virulence factors, resulting a significant increase in the levels of inflammatory factors, triggering inflammation.	[81,82]
<i>H. pylori</i>	<i>H. pylori</i> -derived OMVs exert immunomodulatory effects by inducing the production of pro-inflammatory cytokines and promoting apoptosis of gastric epithelial and immune cells. They also induce apoptosis in human umbilical vein endothelial cells, which may promote atherosclerotic plaque formation.	[85]
<i>V. cholerae</i>	Cholera toxin (CT) is the main virulence factor of <i>Vibrio cholerae</i> , and OMVs may be the important carrier for transporting CT to epithelial cells.	[86]
<i>Bacteroides fragilis</i>	<i>Bacteroides fragilis</i> releases PSA by OMVs, inducing immunomodulatory effects, and prevents experimental colitis.	[87]

3.2. Role of Bacterial EVs in Neurodegenerative Diseases

Substantial evidence has revealed a strong connection between the gut and the brain—referred to as the gut–brain axis—and the composition of the gut microbiota and their derivative OMVs have an important impact on neurological disorders [88,89]. It is reported that, by using mice overexpressing α -synuclein, more α -synuclein aggregates were deposited in the brains of control mice compared to those of germ-free mice, and oral administration of specific bacterial metabolites to germ-free mice enhanced neuroinflammation and motor symptoms, suggesting that the gut microbiota and their secretions in the form of OMVs transmission might be an important contributor to α -synuclein pathology and microglia activation in Parkinson’s disease [90]. Similarly, there appeared to be significant changes of the composition of the gut microbiota in AD model (5xFAD) mice compared to that of control mice at the age of 6 months, which were characterized by a dynamic increase in the abundance of pro-inflammatory molecule-generating bacteria such as *Aspergillus*, *Mimicryptosporium*, and *Terratula* [91]. Consistent with the viewpoint that dysbiosis of gut microbiota might be a risk factor for neurodegeneration, an excellent study conducted by Teng et al. elucidated that isoamylamine (IAA), a metabolite secreted by gut bacteria, promotes age-related cognitive degeneration by inducing microglia death [92]. A reasonable mechanism by which IAA is transported to the brain might be the high permeability of the intestinal mucosal barrier due to exposure to detrimental factors (including OMV content or pro-inflammatory molecules) resulting from the alterations of gut microbiota. Usually, increased permeability of the gut mucosal barrier is accompanied by susceptibility to colitis. Studies have demonstrated that OMVs released by beneficial *Bifidobacterium fragilis* deliver PSA to the dendrite cells (DCs) of the intestine to mediate mutual interactions between the bacteria and the gut immune system, leading to the inhibition of pro-inflammatory cytokine production and the prevention of colitis [87]. In addition, OMVs from *Lactobacillus rhamnosus* GG can increase the expression of antimicrobial peptides and tight junction proteins of the intestine to prevent gut barrier destruction [93]. Based on the current findings detailed above, the health of the gut may strongly influence the functions

of the brain, while the metabolites, particularly the OMVs of microorganism in the gut, probably dominate the pathogenesis of neurodegeneration.

4. Plant Derived Exosome-Like Nanoparticles

Plant-derived exosome-like nanoparticles (PDELNs) are nanosized vesicles usually isolated from edible fruits or vegetables [94], which have huge potential for clinical applications compared to exosomes from mammalian cells or bacteria. The highlighted feature of PDELNs includes non/low toxicity, ideal biodistribution, efficient bioavailability, and high yield [95–97]. Most PDELNs are structurally similar to mammalian exosomes, with an apparent spherical structure of lipid bilayers. In comparison with cholesterol, glycosphingolipids, ceramides, and phosphatidylserine are composed of a mammalian exosome lipid bilayer [98–101], and the membranes of PDELNs are enriched with phosphatidic acid (PA), phosphatidylcholines (PC) digalactosyldiacylglycerol (DGDG), and monogalactosyldiacylglycerol (MGDG), providing inherent mammalian-cell-regulating activities [102,103].

4.1. Biological Functions of PDELNs

It has been reported that plant exosomes can be absorbed by intestinal microorganisms, exemplified by the composition of microorganisms in the feces of C57BL/6 mice that were altered by oral administration with grape exosome-like nanoparticles (GELNs) [103]. GELNs can also regulate the growth of intestinal stem cells, as well as induce IL-22 expression through activation of the AHR pathway, thereby protecting the integrity of the intestinal barrier for the treatment of colitis [103,104]. Furthermore, Li and colleagues found that GELNs can easily pass through the blood–brain barrier in zebrafish and exert a protective effect on neurodevelopment [105]. In addition, grapefruit-derived nanoparticles (GDNs), which are selectively absorbed by intestinal macrophages, can ameliorate dextran sulfate sodium (DSS)-induced colitis by up-regulating heme oxygenase-1 (HO-1) expression and inhibiting the production of inflammatory factors [106]. Sulforaphane (SFN), enwrapped in broccoli-derived nanoparticles (BDNs), protects mice from colitis by inducing the production of anti-inflammatory factors through the AMPK pathway [107]. It has also been shown that, in addition to altering the composition of the intestinal microbiota, lemon exosome-like nanoparticles (LELNs) enhance the pharmaceutical effects of probiotics to inhibit *Clostridioides difficile* infection via AhR-dependent and AhR-independent pathways [108]. In addition to the critical role in regulating the intestinal microbiota, other studies found that the exosome-like nanoparticles from ginger (GELNs) can also inhibit *Porphyromonas gingivalis*—a periodontal pathogen that causes periodontitis—through their interaction with GELN cargo molecules including phosphatidic acid and miRNAs [109]. These current studies suggest that PDELNs may exert their biological function by restoring the gut barrier integrity (colitis), maintaining the normal composition of gut microorganisms, or through direct contact to inhibit the pathogenic bacteria.

4.2. Potential Applications of PDELNs for Treating Neurodegenerative Diseases

Even though there are few studies showing the direct effect of PDELNs on the treatment of neurodegenerative disease, it is now certain that, as a novel therapeutic method, PDELNs have been demonstrated to be highly effective in treating inflammatory diseases like colitis [107], encephalitis [110], periodontitis [109] and so forth, among which the pathogenesis of colitis was considered to be strongly related to the dysfunction of the gut barrier and the dysbiosis of the gut microbiota. The above evidence raises the question of whether PDELNs could be considered a potential drug for the treatment of neurodegenerative disease.

The gut–brain axis is required for transducing detrimental signals from the gut to the brain. For instance, inflammatory factors resulting from leaky gut (with colitis) penetrate the blood–brain barrier (BBB), disrupting BBB integrity. As a result, the activated gut immune cells may be translocated to the brain to amplify neuroinflammation. In addition,

inflammatory factors can also be transmitted through the vagal nerve that connects the gut and the brain, leading to increased neuroinflammation [111]. Regarding such a tight relation between gut and brain, any kinds of PDELNs that are effective for treating inflammatory gut diseases might also be useful for treating neurodegenerative diseases with the hallmark of apparent neuroinflammation. In particular, the efficacy of sulforaphane (SFN) had been systematically studied in neurodegenerative diseases, with an emphasis on its anti-inflammatory role [112]. Since SFN is enriched in broccoli-derived nanoparticles (BDNs) [107], it is highly possible that BDNs would also be effective in treating neurodegenerative diseases.

It is widely accepted that microglia play a pivotal role in the progression of Alzheimer's disease (AD) [113,114]. Usually, normal proliferation, chemotaxis, and phagocytosis are required to remove excessive A β deposition [115]. However, while overactivated, microglia would release inflammatory cytokines to induce neuronal death [116,117]. In the process of microglia development, short-chain fatty acids (SCFA), one of the metabolites of intestinal microorganism, promote microglia maturation as well as their morphological and functional stabilization [118]. Interestingly, Teng and colleagues found that isoamylamine (IAA), a metabolite from pathogenic bacteria in the gut, promotes age-related cognitive dysfunction by inducing microglial cell death [92], which means that PDELNs might be effective for treating the cognitive decline associated with neurodegenerative diseases through restoring the function of microglia.

Taken together, the possible mechanism of a therapeutic role of PDELNs in the treatment of neurodegenerative diseases is through rebalancing the composition of the gut microorganism, preventing the peripheral inflammatory factors entering the brain, thereby diminishing neuroinflammation.

5. Conclusions and Future Perspectives

Although multiple factors (including genetic, environmental, dietary, and metabolic) contribute to the pathogenesis of neurodegenerative diseases such as Alzheimer's disease (AD) and Parkinson's disease (PD), and there exist significant heterogeneities among different individuals, two common factors have been agreed to date as follows: (1) the pathological hallmark is the presence of misfolded protein aggregations/depositions in the brain that are usually defined as proteinopathies; (2) progressive and long lasting neuronal cell death, as well as glia cell reactivation and neuroinflammation. Because emerging evidence has revealed a critical role of tissue-specific exosomes (mammalian EVs) in metabolizing disease-associated proteins, and because bacteria-derived exosomes (OMVs) can modulate gut microbiota thereby influencing gut inflammation and gut barrier integrity, one can postulate that these two varieties of EVs would strongly affect the progression of neurodegeneration.

In the brain, both neurons and microglia generate EVs that carry excessive pro-fibrils of A β . Then, the pro-fibrils of A β are propagated to the extracellular space accompanied by the release of other payloads of EVs, resulting in A β deposition (amyloid plaque) and neuronal cell death. In the gut, dysbiosis of the microbiota leads to an imbalance of beneficial OMVs and pathogenic OMVs, disrupting the integrity of the gut barrier. The leaky gut facilitates more pro-inflammatory molecules and related peripheral immune cells to enter circulation, activating microglia and causing neuroinflammation. As a promising therapeutic approach, plant-derived exosome-like nanoparticles (PDELNs) may protect the brain through re-balancing the composition of the gut microbiota.

One of the possible etiologies involving EVs is illustrated in Figure 2, which concludes that the imbalance of beneficial and pathogenic OMVs enhances the neuroinflammation of neurodegenerative disorders mediated by the gut-brain axis, suggesting a key role of not only mammalian cell-derived EVs but also bacteria-derived EVs in the development of neurodegeneration. However, the latest, more appealing studies conducted by several laboratories have revealed that PDELNs show promising therapeutic effects in terms of re-balancing the composition of the gut microbiota and relieving the inflammation of the

gut and brain. This cross-kingdom EV communication suggests that one may use PDELNs as a potential therapeutic for treating neurodegenerative diseases.

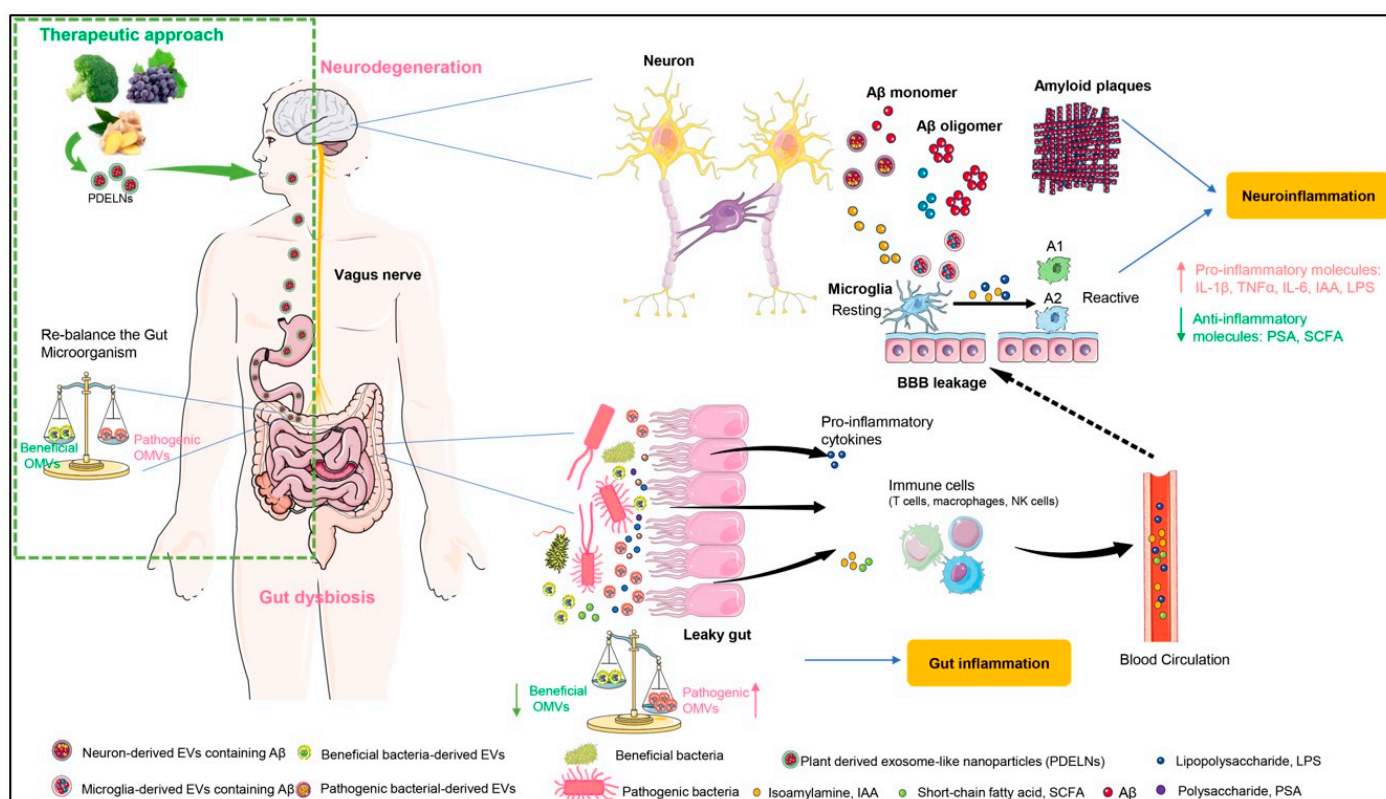


Figure 2. The role of extracellular vesicles from bacteria and their host in the pathogenesis of neurodegeneration, and a possible mechanism of plant-derived exosome-like nanoparticles as a novel approach to treating neurodegeneration. Abbreviations: OMVs, outer membrane vesicles; PDELNs, plant derived exosome-like nanoparticles; BBB, blood brain-barrier; IAA, isoamylamine; LPS, lipopolysaccharide; PSA, polysaccharide; SCFA, short-chain fatty acid.

Although existing studies have provided a solid foundation on the mutual relations between OMVs and mammalian EVs, and between PDELNs and OMVs, respectively, the exact mechanism by which PDELNs reshape the gut microbiota, and whether PDELNs can directly protect the brain through communicating with neuron or glia-derived EVs remain ambiguous. In addition, further studies are needed to establish more effective contents of PDELNs to accelerate their clinical application to treating neurodegenerative disorders.

Author Contributions: Conceptualization, H.F.; Methodology, Y.L. and C.Z.; writing (original draft preparation), Y.L. and C.Z.; writing (review and editing), Y.L., H.F., H.L. and T.C.; supervision, T.C. and H.F. All authors have read and agreed to the published version of the manuscript.

Funding: The study was supported by Young Innovative Talent Project of YongJiang Talent Introduction Programme 2021A-012-G (H.F.) from Ningbo Municipal Government, Ningbo Natural Science Foundation (2021J321 to H.F., and 2021J328 and 2023J365 to Y.L.), Ningbo Public Service Technology Foundation (2022S030 to H.F.), Special Funding for Microfluidic Chip of Biomedicine of Ningbo Institute of Life and Health Industry, University of Chinese Academy of Sciences (2021YJY1006 to H.F., 2021YJY1005 to Y.L.), Medical Scientific Research Foundation of Zhejiang Province (2023KY299 to Y.L., and 2024KY351 to H.F.), and Zhejiang Provincial Natural Science Foundation of China (LQ24H160003 to Y.L.).

Acknowledgments: We apologize to all the researchers whose important works have not been cited due to limited space. Our special thanks to those investigators who contributed, either directly or indirectly, to the field of extracellular vesicles, inflammatory diseases and neurodegeneration. Particularly, the hypothesis of treating AD with MSC-EVs in this review is highly inspired by a research

from Robert Steinfeld from University Medical Center Göttingen (Germany), who demonstrated that exosomes facilitate folate transportation to brain parenchyma. We also appreciate those researchers who revealed certain biological functions of EVs from bacteria and plant cells. In addition, many thanks to Yueyang Fan, the daughter of Fan's, who was very quiet during night, giving her father more time to revise this paper.

Conflicts of Interest: The authors declare no conflict of interest.

References

1. Vaquer-Alicea, J.; Diamond, M.I. Propagation of Protein Aggregation in Neurodegenerative Diseases. *Annu. Rev. Biochem.* **2019**, *88*, 785–810. [\[CrossRef\]](#)
2. Jucker, M.; Walker, L.C. Self-propagation of pathogenic protein aggregates in neurodegenerative diseases. *Nature* **2013**, *501*, 45–51. [\[CrossRef\]](#)
3. Lee, S.J.; Lim, H.S.; Masliah, E.; Lee, H.J. Protein aggregate spreading in neurodegenerative diseases: Problems and perspectives. *Neurosci. Res.* **2011**, *70*, 339–348. [\[CrossRef\]](#)
4. Zhang, Y.; Liu, Y.; Liu, H.; Tang, W.H. Exosomes: Biogenesis, biologic function and clinical potential. *Cell Biosci.* **2019**, *9*, 19. [\[CrossRef\]](#) [\[PubMed\]](#)
5. Wei, H.; Chen, Q.; Lin, L.; Sha, C.; Li, T.; Liu, Y.; Yin, X.; Xu, Y.; Chen, L.; Gao, W.; et al. Regulation of exosome production and cargo sorting. *Int. J. Biol. Sci.* **2021**, *17*, 163–177. [\[CrossRef\]](#) [\[PubMed\]](#)
6. Vella, L.J.; Hill, A.F.; Cheng, L. Focus on Extracellular Vesicles: Exosomes and Their Role in Protein Trafficking and Biomarker Potential in Alzheimer's and Parkinson's Disease. *Int. J. Mol. Sci.* **2016**, *17*, 173. [\[CrossRef\]](#) [\[PubMed\]](#)
7. Coleman, B.M.; Hill, A.F. Extracellular vesicles--Their role in the packaging and spread of misfolded proteins associated with neurodegenerative diseases. *Semin. Cell Dev. Biol.* **2015**, *40*, 89–96. [\[CrossRef\]](#)
8. Peng, C.; Trojanowski, J.Q.; Lee, V.M. Protein transmission in neurodegenerative disease. *Nat. Rev. Neurol.* **2020**, *16*, 199–212. [\[CrossRef\]](#) [\[PubMed\]](#)
9. Huber, C.C.; Wang, H. Pathogenic and therapeutic role of exosomes in neurodegenerative disorders. *Neural Regen. Res.* **2024**, *19*, 75–79. [\[CrossRef\]](#)
10. Gassama, Y.; Favereaux, A. Emerging Roles of Extracellular Vesicles in the Central Nervous System: Physiology, Pathology, and Therapeutic Perspectives. *Front. Cell Neurosci.* **2021**, *15*, 626043. [\[CrossRef\]](#) [\[PubMed\]](#)
11. Kordelas, L.; Rebmann, V.; Ludwig, A.K.; Radtke, S.; Ruesing, J.; Doepfner, T.R.; Epple, M.; Horn, P.A.; Beelen, D.W.; Giebel, B. MSC-derived exosomes: A novel tool to treat therapy-refractory graft-versus-host disease. *Leukemia* **2014**, *28*, 970–973. [\[CrossRef\]](#)
12. Perets, N.; Hertz, S.; London, M.; Offen, D. Intranasal administration of exosomes derived from mesenchymal stem cells ameliorates autistic-like behaviors of BTBR mice. *Mol. Autism.* **2018**, *9*, 57. [\[CrossRef\]](#)
13. Williams, A.M.; Dennahy, I.S.; Bhatti, U.F.; Halaweish, I.; Xiong, Y.; Chang, P.; Nikolian, V.C.; Chtraklin, K.; Brown, J.; Zhang, Y.; et al. Mesenchymal Stem Cell-Derived Exosomes Provide Neuroprotection and Improve Long-Term Neurologic Outcomes in a Swine Model of Traumatic Brain Injury and Hemorrhagic Shock. *J. Neurotrauma* **2019**, *36*, 54–60. [\[CrossRef\]](#) [\[PubMed\]](#)
14. Turovsky, E.A.; Golovicheva, V.V.; Varlamova, E.G.; Danilina, T.I.; Goryunov, K.V.; Shevtsova, Y.A.; Pevzner, I.B.; Zorova, L.D.; Babenko, V.A.; Evtushenko, E.A.; et al. Mesenchymal stromal cell-derived extracellular vesicles afford neuroprotection by modulating PI3K/AKT pathway and calcium oscillations. *Int. J. Biol. Sci.* **2022**, *18*, 5345–5368. [\[CrossRef\]](#)
15. Rana, S.; Zoller, M. Exosome target cell selection and the importance of exosomal tetraspanins: A hypothesis. *Biochem. Soc. Trans.* **2011**, *39*, 559–562. [\[CrossRef\]](#)
16. Christianson, H.C.; Svensson, K.J.; van Kuppevelt, T.H.; Li, J.P.; Belting, M. Cancer cell exosomes depend on cell-surface heparan sulfate proteoglycans for their internalization and functional activity. *Proc. Natl. Acad. Sci. USA* **2013**, *110*, 17380–17385. [\[CrossRef\]](#)
17. Nazarenko, I.; Rana, S.; Baumann, A.; McAlear, J.; Hellwig, A.; Trendelenburg, M.; Lochnit, G.; Preissner, K.T.; Zoller, M. Cell surface tetraspanin Tspan8 contributes to molecular pathways of exosome-induced endothelial cell activation. *Cancer Res.* **2010**, *70*, 1668–1678. [\[CrossRef\]](#) [\[PubMed\]](#)
18. Feng, D.; Zhao, W.L.; Ye, Y.Y.; Bai, X.C.; Liu, R.Q.; Chang, L.F.; Zhou, Q.; Sui, S.F. Cellular internalization of exosomes occurs through phagocytosis. *Traffic* **2010**, *11*, 675–687. [\[CrossRef\]](#) [\[PubMed\]](#)
19. Barres, C.; Blanc, L.; Bette-Bobillo, P.; Andre, S.; Mamoun, R.; Gabius, H.J.; Vidal, M. Galectin-5 is bound onto the surface of rat reticulocyte exosomes and modulates vesicle uptake by macrophages. *Blood* **2010**, *115*, 696–705. [\[CrossRef\]](#)
20. Zech, D.; Rana, S.; Buchler, M.W.; Zoller, M. Tumor-exosomes and leukocyte activation: An ambivalent crosstalk. *Cell Commun. Signal.* **2012**, *10*, 37. [\[CrossRef\]](#)
21. Parolini, I.; Federici, C.; Raggi, C.; Lugini, L.; Palleschi, S.; De Milito, A.; Coscia, C.; Iessi, E.; Logozzi, M.; Molinari, A.; et al. Microenvironmental pH is a key factor for exosome traffic in tumor cells. *J. Biol. Chem.* **2009**, *284*, 34211–34222. [\[CrossRef\]](#) [\[PubMed\]](#)
22. Braak, H.; Rub, U.; Gai, W.P.; Del Tredici, K. Idiopathic Parkinson's disease: Possible routes by which vulnerable neuronal types may be subject to neuroinvasion by an unknown pathogen. *J. Neural. Transm.* **2003**, *110*, 517–536. [\[CrossRef\]](#)
23. Howitt, J.; Hill, A.F. Exosomes in the Pathology of Neurodegenerative Diseases. *J. Biol. Chem.* **2016**, *291*, 26589–26597. [\[CrossRef\]](#) [\[PubMed\]](#)

24. Brettschneider, J.; Del Tredici, K.; Lee, V.M.; Trojanowski, J.Q. Spreading of pathology in neurodegenerative diseases: A focus on human studies. *Nat. Rev. Neurosci.* **2015**, *16*, 109–120. [[CrossRef](#)]
25. Chen, C.; Dong, X.P. Epidemiological characteristics of human prion diseases. *Infect. Dis. Poverty* **2016**, *5*, 47. [[CrossRef](#)] [[PubMed](#)]
26. Collinge, J. Prion diseases of humans and animals: Their causes and molecular basis. *Annu. Rev. Neurosci.* **2001**, *24*, 519–550. [[CrossRef](#)]
27. Noori, L.; Filip, K.; Nazmara, Z.; Mahakizadeh, S.; Hassanzadeh, G.; Caruso Bavisotto, C.; Bucchieri, F.; Marino Gammazza, A.; Cappello, F.; Wnuk, M.; et al. Contribution of Extracellular Vesicles and Molecular Chaperones in Age-Related Neurodegenerative Disorders of the CNS. *Int. J. Mol. Sci.* **2023**, *24*, 927. [[CrossRef](#)]
28. Brandner, S.; Isenmann, S.; Raeber, A.; Fischer, M.; Sailer, A.; Kobayashi, Y.; Marino, S.; Weissmann, C.; Aguzzi, A. Normal host prion protein necessary for scrapie-induced neurotoxicity. *Nature* **1996**, *379*, 339–343. [[CrossRef](#)]
29. Fevrier, B.; Vilette, D.; Archer, F.; Loew, D.; Faigle, W.; Vidal, M.; Laude, H.; Raposo, G. Cells release prions in association with exosomes. *Proc. Natl. Acad. Sci. USA* **2004**, *101*, 9683–9688. [[CrossRef](#)]
30. Vella, L.J.; Sharples, R.A.; Lawson, V.A.; Masters, C.L.; Cappai, R.; Hill, A.F. Packaging of prions into exosomes is associated with a novel pathway of PrP processing. *J. Pathol.* **2007**, *211*, 582–590. [[CrossRef](#)] [[PubMed](#)]
31. Coleman, B.M.; Hanssen, E.; Lawson, V.A.; Hill, A.F. Prion-infected cells regulate the release of exosomes with distinct ultrastructural features. *FASEB J.* **2012**, *26*, 4160–4173. [[CrossRef](#)]
32. Budnik, V.; Ruiz-Canada, C.; Wendler, F. Extracellular vesicles round off communication in the nervous system. *Nat. Rev. Neurosci.* **2016**, *17*, 160–172. [[CrossRef](#)]
33. Quek, C.; Hill, A.F. The role of extracellular vesicles in neurodegenerative diseases. *Biochem. Biophys. Res. Commun.* **2017**, *483*, 1178–1186. [[CrossRef](#)]
34. Emmanouilidou, E.; Melachroinou, K.; Roumeliotis, T.; Garbis, S.D.; Ntzouni, M.; Margaritis, L.H.; Stefanis, L.; Vekrellis, K. Cell-produced alpha-synuclein is secreted in a calcium-dependent manner by exosomes and impacts neuronal survival. *J. Neurosci.* **2010**, *30*, 6838–6851. [[CrossRef](#)]
35. Shi, M.; Liu, C.; Cook, T.J.; Bullock, K.M.; Zhao, Y.; Gingham, C.; Li, Y.; Aro, P.; Dator, R.; He, C.; et al. Plasma exosomal alpha-synuclein is likely CNS-derived and increased in Parkinson's disease. *Acta Neuropathol.* **2014**, *128*, 639–650. [[CrossRef](#)]
36. Rouillard, M.E.; Sutter, P.A.; Durham, O.R.; Willis, C.M.; Crocker, S.J. Astrocyte-Derived Extracellular Vesicles (ADEVs): Deciphering their Influences in Aging. *Aging Dis.* **2021**, *12*, 1462–1475. [[CrossRef](#)] [[PubMed](#)]
37. Dinkins, M.B.; Dasgupta, S.; Wang, G.; Zhu, G.; Bieberich, E. Exosome reduction in vivo is associated with lower amyloid plaque load in the 5XFAD mouse model of Alzheimer's disease. *Neurobiol. Aging* **2014**, *35*, 1792–1800. [[CrossRef](#)]
38. Varcianna, A.; Myszczyńska, M.A.; Castelli, L.M.; O'Neill, B.; Kim, Y.; Talbot, J.; Nyberg, S.; Nyamali, I.; Heath, P.R.; Stopford, M.J.; et al. Micro-RNAs secreted through astrocyte-derived extracellular vesicles cause neuronal network degeneration in C9orf72 ALS. *EBioMedicine* **2019**, *40*, 626–635. [[CrossRef](#)]
39. Dickens, A.M.; Tovar, Y.R.L.B.; Yoo, S.W.; Trout, A.L.; Bae, M.; Kanmogne, M.; Megra, B.; Williams, D.W.; Witwer, K.W.; Gacias, M.; et al. Astrocyte-shed extracellular vesicles regulate the peripheral leukocyte response to inflammatory brain lesions. *Sci. Signal.* **2017**, *10*, eaai7696. [[CrossRef](#)]
40. Tian, Y.; Fu, C.; Wu, Y.; Lu, Y.; Liu, X.; Zhang, Y. Central Nervous System Cell-Derived Exosomes in Neurodegenerative Diseases. *Oxid. Med. Cell Longev.* **2021**, *2021*, 9965564. [[CrossRef](#)]
41. Vinaiphath, A.; Sze, S.K. Proteomics for comprehensive characterization of extracellular vesicles in neurodegenerative disease. *Exp. Neurol.* **2022**, *355*, 114149. [[CrossRef](#)]
42. Chen, S.Y.; Lin, M.C.; Tsai, J.S.; He, P.L.; Luo, W.T.; Chiu, I.M.; Herschman, H.R.; Li, H.J. Exosomal 2',3'-CNP from mesenchymal stem cells promotes hippocampus CA1 neurogenesis/neuritogenesis and contributes to rescue of cognition/learning deficiencies of damaged brain. *Stem. Cells Transl. Med.* **2020**, *9*, 499–517. [[CrossRef](#)]
43. Bodart-Santos, V.; de Carvalho, L.R.P.; de Godoy, M.A.; Batista, A.F.; Saraiva, L.M.; Lima, L.G.; Abreu, C.A.; De Felice, F.G.; Galina, A.; Mendez-Otero, R.; et al. Extracellular vesicles derived from human Wharton's jelly mesenchymal stem cells protect hippocampal neurons from oxidative stress and synapse damage induced by amyloid-beta oligomers. *Stem. Cell Res. Ther.* **2019**, *10*, 332. [[CrossRef](#)]
44. Nakano, M.; Nagaishi, K.; Konari, N.; Saito, Y.; Chikenji, T.; Mizue, Y.; Fujimiya, M. Bone marrow-derived mesenchymal stem cells improve diabetes-induced cognitive impairment by exosome transfer into damaged neurons and astrocytes. *Sci. Rep.* **2016**, *6*, 24805. [[CrossRef](#)]
45. Sharma, P.; Mesci, P.; Carroneu, C.; McClatchy, D.R.; Schiapparelli, L.; Yates, J.R., 3rd; Muotri, A.R.; Cline, H.T. Exosomes regulate neurogenesis and circuit assembly. *Proc. Natl. Acad. Sci. USA* **2019**, *116*, 16086–16094. [[CrossRef](#)] [[PubMed](#)]
46. Reza-Zaldivar, E.E.; Hernandez-Sapiens, M.A.; Minjarez, B.; Gutierrez-Mercado, Y.K.; Marquez-Aguirre, A.L.; Canales-Aguirre, A.A. Potential Effects of MSC-Derived Exosomes in Neuroplasticity in Alzheimer's Disease. *Front. Cell Neurosci.* **2018**, *12*, 317. [[CrossRef](#)] [[PubMed](#)]
47. Forsberg, M.H.; Kink, J.A.; Hematti, P.; Capitini, C.M. Mesenchymal Stromal Cells and Exosomes: Progress and Challenges. *Front. Cell Dev. Biol.* **2020**, *8*, 665. [[CrossRef](#)] [[PubMed](#)]
48. Wei, X.; Yang, X.; Han, Z.P.; Qu, F.F.; Shao, L.; Shi, Y.F. Mesenchymal stem cells: A new trend for cell therapy. *Acta Pharmacol. Sin.* **2013**, *34*, 747–754. [[CrossRef](#)] [[PubMed](#)]

49. Yang, Y.; Ye, Y.; Su, X.; He, J.; Bai, W.; He, X. MSCs-Derived Exosomes and Neuroinflammation, Neurogenesis and Therapy of Traumatic Brain Injury. *Front. Cell Neurosci.* **2017**, *11*, 55. [[CrossRef](#)] [[PubMed](#)]
50. Xiong, Y.; Mahmood, A.; Chopp, M. Emerging potential of exosomes for treatment of traumatic brain injury. *Neural Regen. Res.* **2017**, *12*, 19–22. [[CrossRef](#)] [[PubMed](#)]
51. Xin, H.; Li, Y.; Liu, Z.; Wang, X.; Shang, X.; Cui, Y.; Zhang, Z.G.; Chopp, M. MiR-133b promotes neural plasticity and functional recovery after treatment of stroke with multipotent mesenchymal stromal cells in rats via transfer of exosome-enriched extracellular particles. *Stem. Cells* **2013**, *31*, 2737–2746. [[CrossRef](#)]
52. Doeppner, T.R.; Herz, J.; Gorgens, A.; Schlechter, J.; Ludwig, A.K.; Radtke, S.; de Miroschedji, K.; Horn, P.A.; Giebel, B.; Hermann, D.M. Extracellular Vesicles Improve Post-Stroke Neuroregeneration and Prevent Postischemic Immunosuppression. *Stem. Cells Transl. Med.* **2015**, *4*, 1131–1143. [[CrossRef](#)]
53. Zhang, Y.; Chopp, M.; Zhang, Z.G.; Katakowski, M.; Xin, H.; Qu, C.; Ali, M.; Mahmood, A.; Xiong, Y. Systemic administration of cell-free exosomes generated by human bone marrow derived mesenchymal stem cells cultured under 2D and 3D conditions improves functional recovery in rats after traumatic brain injury. *Neurochem. Int.* **2017**, *111*, 69–81. [[CrossRef](#)]
54. Li, Y.; Chen, J.; Chen, X.G.; Wang, L.; Gautam, S.C.; Xu, Y.X.; Katakowski, M.; Zhang, L.J.; Lu, M.; Janakiraman, N.; et al. Human marrow stromal cell therapy for stroke in rat: Neurotrophins and functional recovery. *Neurology* **2002**, *59*, 514–523. [[CrossRef](#)] [[PubMed](#)]
55. Kurozumi, K.; Nakamura, K.; Tamiya, T.; Kawano, Y.; Kobune, M.; Hirai, S.; Uchida, H.; Sasaki, K.; Ito, Y.; Kato, K.; et al. BDNF gene-modified mesenchymal stem cells promote functional recovery and reduce infarct size in the rat middle cerebral artery occlusion model. *Mol. Ther.* **2004**, *9*, 189–197. [[CrossRef](#)]
56. Kim, H.J.; Lee, J.H.; Kim, S.H. Therapeutic effects of human mesenchymal stem cells on traumatic brain injury in rats: Secretion of neurotrophic factors and inhibition of apoptosis. *J. Neurotrauma* **2010**, *27*, 131–138. [[CrossRef](#)] [[PubMed](#)]
57. Matthay, M.A.; Pati, S.; Lee, J.W. Concise Review: Mesenchymal Stem (Stromal) Cells: Biology and Preclinical Evidence for Therapeutic Potential for Organ Dysfunction Following Trauma or Sepsis. *Stem. Cells* **2017**, *35*, 316–324. [[CrossRef](#)]
58. Nguyen, T.M.; Arthur, A.; Hayball, J.D.; Gronthos, S. EphB and Ephrin-B interactions mediate human mesenchymal stem cell suppression of activated T-cells. *Stem. Cells Dev.* **2013**, *22*, 2751–2764. [[CrossRef](#)]
59. Phinney, D.G.; Pittenger, M.F. Concise Review: MSC-Derived Exosomes for Cell-Free Therapy. *Stem. Cells* **2017**, *35*, 851–858. [[CrossRef](#)]
60. Timmers, L.; Lim, S.K.; Arslan, F.; Armstrong, J.S.; Hofer, I.E.; Doevendans, P.A.; Piek, J.J.; El Oakley, R.M.; Choo, A.; Lee, C.N.; et al. Reduction of myocardial infarct size by human mesenchymal stem cell conditioned medium. *Stem. Cell Res.* **2007**, *1*, 129–137. [[CrossRef](#)] [[PubMed](#)]
61. Mitsialis, S.A.; Kourembanas, S. Stem cell-based therapies for the newborn lung and brain: Possibilities and challenges. *Semin. Perinatol.* **2016**, *40*, 138–151. [[CrossRef](#)]
62. Lai, R.C.; Arslan, F.; Lee, M.M.; Sze, N.S.; Choo, A.; Chen, T.S.; Salto-Tellez, M.; Timmers, L.; Lee, C.N.; El Oakley, R.M.; et al. Exosome secreted by MSC reduces myocardial ischemia/reperfusion injury. *Stem. Cell Res.* **2010**, *4*, 214–222. [[CrossRef](#)]
63. Katsuda, T.; Tsuchiya, R.; Kosaka, N.; Yoshioka, Y.; Takagaki, K.; Oki, K.; Takeshita, F.; Sakai, Y.; Kuroda, M.; Ochiya, T. Human adipose tissue-derived mesenchymal stem cells secrete functional neprilysin-bound exosomes. *Sci. Rep.* **2013**, *3*, 1197. [[CrossRef](#)]
64. Seshadri, S.; Beiser, A.; Selhub, J.; Jacques, P.F.; Rosenberg, I.H.; D’Agostino, R.B.; Wilson, P.W.; Wolf, P.A. Plasma homocysteine as a risk factor for dementia and Alzheimer’s disease. *N. Engl. J. Med.* **2002**, *346*, 476–483. [[CrossRef](#)]
65. Van Dam, F.; Van Gool, W.A. Hyperhomocysteinemia and Alzheimer’s disease: A systematic review. *Arch. Gerontol. Geriatr.* **2009**, *48*, 425–430. [[CrossRef](#)]
66. Seshadri, S. Elevated plasma homocysteine levels: Risk factor or risk marker for the development of dementia and Alzheimer’s disease? *J. Alzheimer’s Dis.* **2006**, *9*, 393–398. [[CrossRef](#)]
67. Miller, J.W. Homocysteine, Alzheimer’s disease, and cognitive function. *Nutrition* **2000**, *16*, 675–677. [[CrossRef](#)] [[PubMed](#)]
68. Pacheco-Quinto, J.; Rodriguez de Turco, E.B.; DeRosa, S.; Howard, A.; Cruz-Sanchez, F.; Sambamurti, K.; Refolo, L.; Petanceska, S.; Pappolla, M.A. Hyperhomocysteinemic Alzheimer’s mouse model of amyloidosis shows increased brain amyloid beta peptide levels. *Neurobiol. Dis.* **2006**, *22*, 651–656. [[CrossRef](#)] [[PubMed](#)]
69. Obeid, R.; Herrmann, W. Mechanisms of homocysteine neurotoxicity in neurodegenerative diseases with special reference to dementia. *FEBS Lett.* **2006**, *580*, 2994–3005. [[CrossRef](#)]
70. Zhuo, J.M.; Pratico, D. Acceleration of brain amyloidosis in an Alzheimer’s disease mouse model by a folate, vitamin B6 and B12-deficient diet. *Exp. Gerontol.* **2010**, *45*, 195–201. [[CrossRef](#)] [[PubMed](#)]
71. Gu, Y.; Nieves, J.W.; Stern, Y.; Luchsinger, J.A.; Scarmeas, N. Food combination and Alzheimer disease risk: A protective diet. *Arch. Neurol.* **2010**, *67*, 699–706. [[CrossRef](#)]
72. Grapp, M.; Wrede, A.; Schweizer, M.; Huwel, S.; Galla, H.J.; Snaidero, N.; Simons, M.; Buckers, J.; Low, P.S.; Urlaub, H.; et al. Choroid plexus transcytosis and exosome shuttling deliver folate into brain parenchyma. *Nat. Commun.* **2013**, *4*, 2123. [[CrossRef](#)]
73. Strazielle, N.; Gherzi-Egea, J.F. Potential Pathways for CNS Drug Delivery Across the Blood-Cerebrospinal Fluid Barrier. *Curr. Pharm. Des.* **2016**, *22*, 5463–5476. [[CrossRef](#)]
74. Weitman, S.D.; Weinberg, A.G.; Coney, L.R.; Zurawski, V.R.; Jennings, D.S.; Kamen, B.A. Cellular localization of the folate receptor: Potential role in drug toxicity and folate homeostasis. *Cancer Res.* **1992**, *52*, 6708–6711.

75. Nikfarjam, S.; Rezaie, J.; Zolbanin, N.M.; Jafari, R. Mesenchymal stem cell derived-exosomes: A modern approach in translational medicine. *J. Transl. Med.* **2020**, *18*, 449. [[CrossRef](#)] [[PubMed](#)]
76. Tarashi, S.; Zamani, M.S.; Omrani, M.D.; Fateh, A.; Moshiri, A.; Saedisomeolia, A.; Siadat, S.D.; Kubow, S. Commensal and Pathogenic Bacterial-Derived Extracellular Vesicles in Host-Bacterial and Interbacterial Dialogues: Two Sides of the Same Coin. *J. Immunol. Res.* **2022**, *2022*, 8092170. [[CrossRef](#)] [[PubMed](#)]
77. Kaparakis-Liaskos, M.; Ferrero, R.L. Immune modulation by bacterial outer membrane vesicles. *Nat. Rev. Immunol.* **2015**, *15*, 375–387. [[CrossRef](#)] [[PubMed](#)]
78. Jones, L.B.; Kumar, S.; Bell, C.R.; Crenshaw, B.J.; Coats, M.T.; Sims, B.; Matthews, Q.L. Lipopolysaccharide Administration Alters Extracellular Vesicles in Cell Lines and Mice. *Curr. Microbiol.* **2021**, *78*, 920–931. [[CrossRef](#)]
79. Bielaszewska, M.; Marejkova, M.; Bauwens, A.; Kunsmann-Prokscha, L.; Mellmann, A.; Karch, H. Enterohemorrhagic Escherichia coli O157 outer membrane vesicles induce interleukin 8 production in human intestinal epithelial cells by signaling via Toll-like receptors TLR4 and TLR5 and activation of the nuclear factor NF-kappaB. *Int. J. Med. Microbiol.* **2018**, *308*, 882–889. [[CrossRef](#)] [[PubMed](#)]
80. Sartorio, M.G.; Pardue, E.J.; Feldman, M.F.; Haurat, M.F. Bacterial Outer Membrane Vesicles: From Discovery to Applications. *Annu. Rev. Microbiol.* **2021**, *75*, 609–630. [[CrossRef](#)] [[PubMed](#)]
81. Kadurugamuwa, J.L.; Beveridge, T.J. Virulence factors are released from Pseudomonas aeruginosa in association with membrane vesicles during normal growth and exposure to gentamicin: A novel mechanism of enzyme secretion. *J. Bacteriol.* **1995**, *177*, 3998–4008. [[CrossRef](#)]
82. Ellis, T.N.; Leiman, S.A.; Kuehn, M.J. Naturally produced outer membrane vesicles from Pseudomonas aeruginosa elicit a potent innate immune response via combined sensing of both lipopolysaccharide and protein components. *Infect. Immun.* **2010**, *78*, 3822–3831. [[CrossRef](#)]
83. Shah, B.; Sullivan, C.J.; Lonergan, N.E.; Stanley, S.; Soult, M.C.; Britt, L.D. Circulating bacterial membrane vesicles cause sepsis in rats. *Shock* **2012**, *37*, 621–628. [[CrossRef](#)]
84. Hickey, C.A.; Kuhn, K.A.; Donermeyer, D.L.; Porter, N.T.; Jin, C.; Cameron, E.A.; Jung, H.; Kaiko, G.E.; Wegorzewska, M.; Malvin, N.P.; et al. Colitogenic Bacteroides thetaiotaomicron Antigens Access Host Immune Cells in a Sulfatase-Dependent Manner via Outer Membrane Vesicles. *Cell. Host Microbe* **2015**, *17*, 672–680. [[CrossRef](#)]
85. Xie, J.; Cools, L.; Van Imschoot, G.; Van Wonterghem, E.; Pauwels, M.J.; Vlaeminck, I.; De Witte, C.; El Andaloussi, S.; Wierda, K.; De Groef, L.; et al. Helicobacter pylori-derived outer membrane vesicles contribute to Alzheimer's disease pathogenesis via C3-C3aR signalling. *J. Extracell. Vesicles.* **2023**, *12*, e12306. [[CrossRef](#)] [[PubMed](#)]
86. Chatterjee, D.; Chaudhuri, K. Association of cholera toxin with Vibrio cholerae outer membrane vesicles which are internalized by human intestinal epithelial cells. *FEBS Lett.* **2011**, *585*, 1357–1362. [[CrossRef](#)] [[PubMed](#)]
87. Shen, Y.; Giardino Torchia, M.L.; Lawson, G.W.; Karp, C.L.; Ashwell, J.D.; Mazmanian, S.K. Outer membrane vesicles of a human commensal mediate immune regulation and disease protection. *Cell Host Microbe* **2012**, *12*, 509–520. [[CrossRef](#)] [[PubMed](#)]
88. Grenham, S.; Clarke, G.; Cryan, J.F.; Dinan, T.G. Brain-gut-microbe communication in health and disease. *Front. Physiol.* **2011**, *2*, 94. [[CrossRef](#)] [[PubMed](#)]
89. Xu, H.; Xu, Z.; Long, S.; Li, Z.; Jiang, J.; Zhou, Q.; Huang, X.; Wu, X.; Wei, W.; Li, X. The role of the gut microbiome and its metabolites in cerebrovascular diseases. *Front. Microbiol.* **2023**, *14*, 1097148. [[CrossRef](#)] [[PubMed](#)]
90. Sampson, T.R.; Debelius, J.W.; Thron, T.; Janssen, S.; Shastri, G.G.; Ilhan, Z.E.; Challis, C.; Schretter, C.E.; Rocha, S.; Gradinaru, V.; et al. Gut Microbiota Regulate Motor Deficits and Neuroinflammation in a Model of Parkinson's Disease. *Cell* **2016**, *167*, 1469–1480.e12. [[CrossRef](#)] [[PubMed](#)]
91. Chen, C.; Ahn, E.H.; Kang, S.S.; Liu, X.; Alam, A.; Ye, K. Gut dysbiosis contributes to amyloid pathology, associated with C/EBPbeta/AEP signaling activation in Alzheimer's disease mouse model. *Sci. Adv.* **2020**, *6*, eaba0466. [[CrossRef](#)]
92. Teng, Y.; Mu, J.; Xu, F.; Zhang, X.; Sriwastva, M.K.; Liu, Q.M.; Li, X.; Lei, C.; Sundaram, K.; Hu, X.; et al. Gut bacterial isoamylamine promotes age-related cognitive dysfunction by promoting microglial cell death. *Cell Host Microbe* **2022**, *30*, 944–960.e8. [[CrossRef](#)]
93. Gu, Z.; Li, F.; Liu, Y.; Jiang, M.; Zhang, L.; He, L.; Wilkey, D.W.; Merchant, M.; Zhang, X.; Deng, Z.B.; et al. Exosome-Like Nanoparticles From Lactobacillus rhamnosusGG Protect Against Alcohol-Associated Liver Disease Through Intestinal Aryl Hydrocarbon Receptor in Mice. *Hepatology. Commun.* **2021**, *5*, 846–864. [[CrossRef](#)]
94. Barzin, M.; Bagheri, A.M.; Ohadi, M.; Abhaji, A.M.; Salarpour, S.; Dehghannoudeh, G. Application of plant-derived exosome-like nanoparticles in drug delivery. *Pharm. Dev. Technol.* **2023**, *28*, 383–402. [[CrossRef](#)]
95. Yang, C.; Zhang, M.; Merlin, D. Advances in Plant-derived Edible Nanoparticle-based lipid Nano-drug Delivery Systems as Therapeutic Nanomedicines. *J. Mater. Chem. B* **2018**, *6*, 1312–1321. [[CrossRef](#)]
96. Kim, J.; Li, S.; Zhang, S.; Wang, J. Plant-derived exosome-like nanoparticles and their therapeutic activities. *Asian J. Pharm. Sci.* **2022**, *17*, 53–69. [[CrossRef](#)] [[PubMed](#)]
97. Zhang, M.; Viennois, E.; Xu, C.; Merlin, D. Plant derived edible nanoparticles as a new therapeutic approach against diseases. *Tissue Barriers* **2016**, *4*, e1134415. [[CrossRef](#)] [[PubMed](#)]
98. Stremersch, S.; De Smedt, S.C.; Raemdonck, K. Therapeutic and diagnostic applications of extracellular vesicles. *J. Control. Release* **2016**, *244*, 167–183. [[CrossRef](#)] [[PubMed](#)]

99. Nishio, M.; Teranishi, Y.; Morioka, K.; Yanagida, A.; Shoji, A. Real-time assay for exosome membrane fusion with an artificial lipid membrane based on enhancement of gramicidin A channel conductance. *Biosens. Bioelectron.* **2020**, *150*, 111918. [[CrossRef](#)] [[PubMed](#)]
100. Mashouri, L.; Yousefi, H.; Aref, A.R.; Ahadi, A.M.; Molaei, F.; Alahari, S.K. Exosomes: Composition, biogenesis, and mechanisms in cancer metastasis and drug resistance. *Mol. Cancer* **2019**, *18*, 75. [[CrossRef](#)] [[PubMed](#)]
101. Waldenstrom, A.; Ronquist, G. Role of exosomes in myocardial remodeling. *Circ. Res.* **2014**, *114*, 315–324. [[CrossRef](#)]
102. Zhang, M.; Viennois, E.; Prasad, M.; Zhang, Y.; Wang, L.; Zhang, Z.; Han, M.K.; Xiao, B.; Xu, C.; Srinivasan, S.; et al. Edible ginger-derived nanoparticles: A novel therapeutic approach for the prevention and treatment of inflammatory bowel disease and colitis-associated cancer. *Biomaterials* **2016**, *101*, 321–340. [[CrossRef](#)]
103. Teng, Y.; Ren, Y.; Sayed, M.; Hu, X.; Lei, C.; Kumar, A.; Hutchins, E.; Mu, J.; Deng, Z.; Luo, C.; et al. Plant-Derived Exosomal MicroRNAs Shape the Gut Microbiota. *Cell Host Microbe* **2018**, *24*, 637–652.e8. [[CrossRef](#)]
104. Ju, S.; Mu, J.; Dokland, T.; Zhuang, X.; Wang, Q.; Jiang, H.; Xiang, X.; Deng, Z.B.; Wang, B.; Zhang, L.; et al. Grape exosome-like nanoparticles induce intestinal stem cells and protect mice from DSS-induced colitis. *Mol. Ther.* **2013**, *21*, 1345–1357. [[CrossRef](#)] [[PubMed](#)]
105. Li, Y.; Cai, T.; Liu, H.; Liu, J.; Chen, S.Y.; Fan, H. Exosome-shuttled miR-126 mediates ethanol-induced disruption of neural crest cell-placode cell interaction by targeting SDF1. *Toxicol. Sci.* **2023**, *195*, 184–201. [[CrossRef](#)]
106. Wang, B.; Zhuang, X.; Deng, Z.B.; Jiang, H.; Mu, J.; Wang, Q.; Xiang, X.; Guo, H.; Zhang, L.; Dryden, G.; et al. Targeted drug delivery to intestinal macrophages by bioactive nanovesicles released from grapefruit. *Mol. Ther.* **2014**, *22*, 522–534. [[CrossRef](#)]
107. Deng, Z.; Rong, Y.; Teng, Y.; Mu, J.; Zhuang, X.; Tseng, M.; Samyuktty, A.; Zhang, L.; Yan, J.; Miller, D.; et al. Broccoli-Derived Nanoparticle Inhibits Mouse Colitis by Activating Dendritic Cell AMP-Activated Protein Kinase. *Mol. Ther.* **2017**, *25*, 1641–1654. [[CrossRef](#)] [[PubMed](#)]
108. Lei, C.; Mu, J.; Teng, Y.; He, L.; Xu, F.; Zhang, X.; Sundaram, K.; Kumar, A.; Sriwastva, M.K.; Lawrenz, M.B.; et al. Lemon Exosome-like Nanoparticles-Manipulated Probiotics Protect Mice from *C. d* iff Infection. *iScience* **2020**, *23*, 101571. [[CrossRef](#)] [[PubMed](#)]
109. Sundaram, K.; Miller, D.P.; Kumar, A.; Teng, Y.; Sayed, M.; Mu, J.; Lei, C.; Sriwastva, M.K.; Zhang, L.; Yan, J.; et al. Plant-Derived Exosomal Nanoparticles Inhibit Pathogenicity of *Porphyromonas gingivalis*. *iScience* **2020**, *23*, 100869. [[CrossRef](#)]
110. Zhuang, X.; Xiang, X.; Grizzle, W.; Sun, D.; Zhang, S.; Axtell, R.C.; Ju, S.; Mu, J.; Zhang, L.; Steinman, L.; et al. Treatment of brain inflammatory diseases by delivering exosome encapsulated anti-inflammatory drugs from the nasal region to the brain. *Mol. Ther.* **2011**, *19*, 1769–1779. [[CrossRef](#)]
111. Agirman, G.; Yu, K.B.; Hsiao, E.Y. Signaling inflammation across the gut-brain axis. *Science* **2021**, *374*, 1087–1092. [[CrossRef](#)]
112. Schepici, G.; Bramanti, P.; Mazzon, E. Efficacy of Sulforaphane in Neurodegenerative Diseases. *Int. J. Mol. Sci.* **2020**, *21*, 8637. [[CrossRef](#)]
113. Thei, L.; Imm, J.; Kaisis, E.; Dallas, M.L.; Kerrigan, T.L. Microglia in Alzheimer’s Disease: A Role for Ion Channels. *Front. Neurosci.* **2018**, *12*, 676. [[CrossRef](#)]
114. Miao, J.; Ma, H.; Yang, Y.; Liao, Y.; Lin, C.; Zheng, J.; Yu, M.; Lan, J. Microglia in Alzheimer’s disease: Pathogenesis, mechanisms, and therapeutic potentials. *Front. Aging Neurosci.* **2023**, *15*, 1201982. [[CrossRef](#)]
115. Kleinberger, G.; Yamanishi, Y.; Suarez-Calvet, M.; Czirr, E.; Lohmann, E.; Cuyvers, E.; Struyfs, H.; Pettkus, N.; Wenninger-Weinzierl, A.; Mazaheri, F.; et al. TREM2 mutations implicated in neurodegeneration impair cell surface transport and phagocytosis. *Sci. Transl. Med.* **2014**, *6*, 243ra286. [[CrossRef](#)]
116. Hansen, D.V.; Hanson, J.E.; Sheng, M. Microglia in Alzheimer’s disease. *J. Cell Biol.* **2018**, *217*, 459–472. [[CrossRef](#)] [[PubMed](#)]
117. Wu, J.; Gao, G.; Shi, F.; Xie, H.; Yang, Q.; Liu, D.; Qu, S.; Qin, H.; Zhang, C.; Xu, G.T.; et al. Activated microglia-induced neuroinflammatory cytokines lead to photoreceptor apoptosis in Abeta-injected mice. *J. Mol. Med.* **2021**, *99*, 713–728. [[CrossRef](#)] [[PubMed](#)]
118. Erny, D.; Hrabé de Angelis, A.L.; Jaitin, D.; Wieghofer, P.; Staszewski, O.; David, E.; Keren-Shaul, H.; Muhlrad, T.; Jakobshagen, K.; Buch, T.; et al. Host microbiota constantly control maturation and function of microglia in the CNS. *Nat. Neurosci.* **2015**, *18*, 965–977. [[CrossRef](#)] [[PubMed](#)]

Disclaimer/Publisher’s Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.